**HIGH RESOLUTION MASS SPECTROMETRY PROVIDES NOVEL INSIGHTS INTO PRODUCTS OF HUMAN METABOLISM OF ORGANOPHOSPHATE AND BROMINATED FLAME RETARDANTS** 

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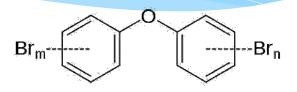




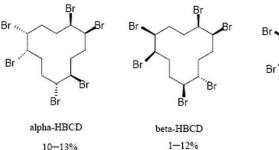
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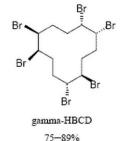
# **Organic Flame Retardants**

- \* Diverse group of halogenated chemicals.
- \* Widely applied in building materials and consumer products.
- Persistent, Bioaccumulative and Toxic (PBT) properties.
- \* Some are banned (e.g. Penta-BDEs) and others undergoing risk assessment.





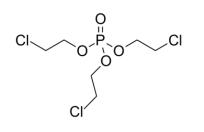




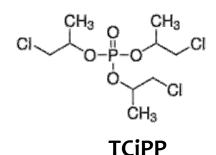






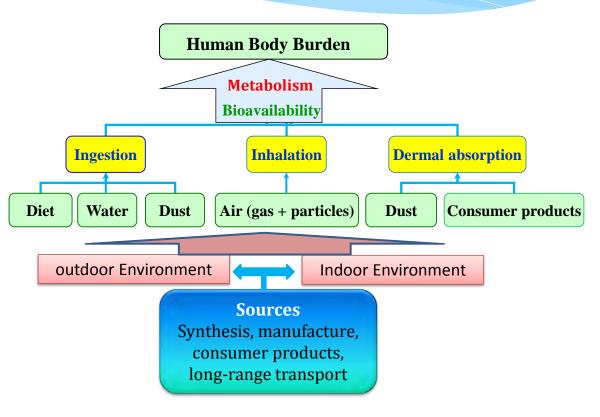


TCEP



# **Organic Flame Retardants**

- \* Most FRs are physically blended with rather than chemically bonded to polymers.
- \* Indoor dust has been consistently identified as a major pathway of human exposure to FRs.
- \* Higher BFRs can be biotransformed to more toxic and bioaccumulative metabolites.
- \* Further understanding of the metabolic process is essential for accurate risk assessment of these hazardous chemicals





#### **Research Gap**

- \* Few studies have investigated the metabolic pathways of different flame retardants present in indoor dust.
- Most of these studies have focused on PBDEs using animal or human LME, hepatic S9 fractions and rarely, human hepatocytes.

# PBDEs are banned

Very little is known about the metabolic pathways of alternative flame retardants in humans.



#### **Research Gap**

- \* Most *in vitro* biotransformation studies focus on exposing the metabolising system (LME, S9 or hepatocytes) to a single xenobiotic at a time which doesn't mimic the *in vivo* situation.
- \* No studies of HBCD metabolism in humans.
- \* No studies of TCEP, TCIPP and TDCPP in human hepatocytes which contain both Phase I and Phase II metabolic enzymes.
- \* The analytical capabilities and performance of the Orbitrap<sup>™</sup>-MS have not been fully evaluated in the field of POPs analysis.



# The Orbitrap-MS (Exactive plus)

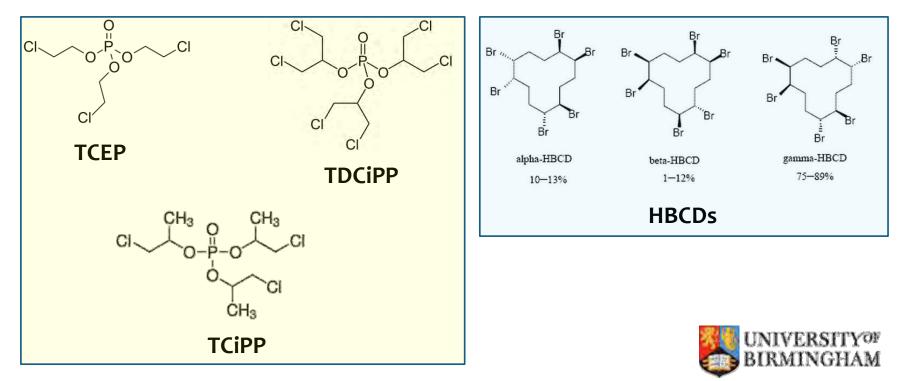
- \* High resolution (up to 140,000 FWHM).
- \* High mass accuracy (up to 1 ppm)-improved selectivity.
- \* High sensitivity- Adjustable AGC
- \* High scan rate.
- \* Rapid polarity switching of the ion source.
- \* Optional HCD cell- AIF spectra for structural confirmation.
- \* Optional quadropole for MS/MS analysis-we didn't have that.







\* Study the metabolic profiles of HBCDs, TCEP, TCIPP and TDCPP in indoor dust (NIST SRM 2585), applied concomitantly to human hepatocyte cultures using UPLC-Orbitrap-MS.



#### **Experimental**

- \* <u>Cell cultures:</u> Human HEPG2/C3A cell lines were seeded and cultured in 6-well plates at 2 x 10<sup>6</sup> cells/well in modified William's E medium (containing 5% FBS).
- \* **Dosing Solutions:** 
  - \* D1- SRM 2585 dust extract (using Dionex ASE 350)
  - D2-Synthetic mixture of the target compounds (HBCDs and 3 PFRs) with the same concentrations as in D1.
  - \* *Exposure Scenario*: 2 million cells exposed to the equivalent of 12 mg dust based on a 12.3 Kg toddler ingesting 200 mg dust/day.





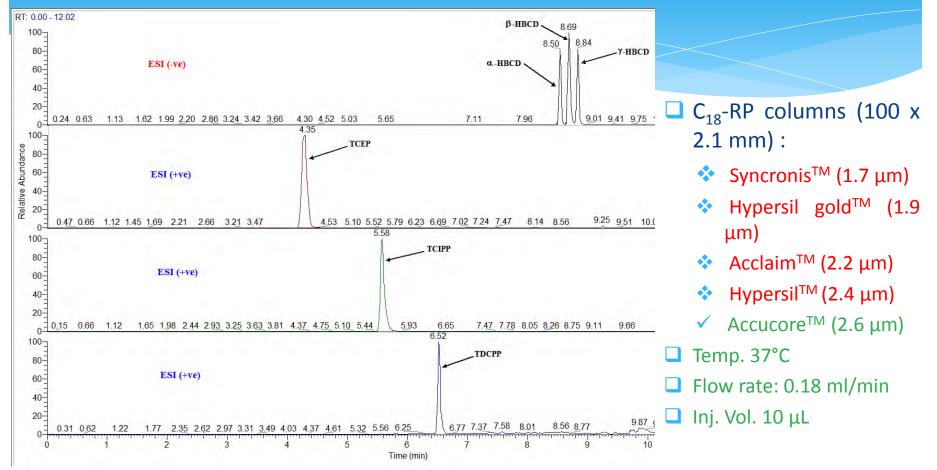
# Incubation at 37°C with humidified air containing 5% CO<sub>2</sub> for 24 hours

**Extraction with methanol - QUESHERS** 

Instrumental Analysis- UPLC-ESI-Orbitrap MS

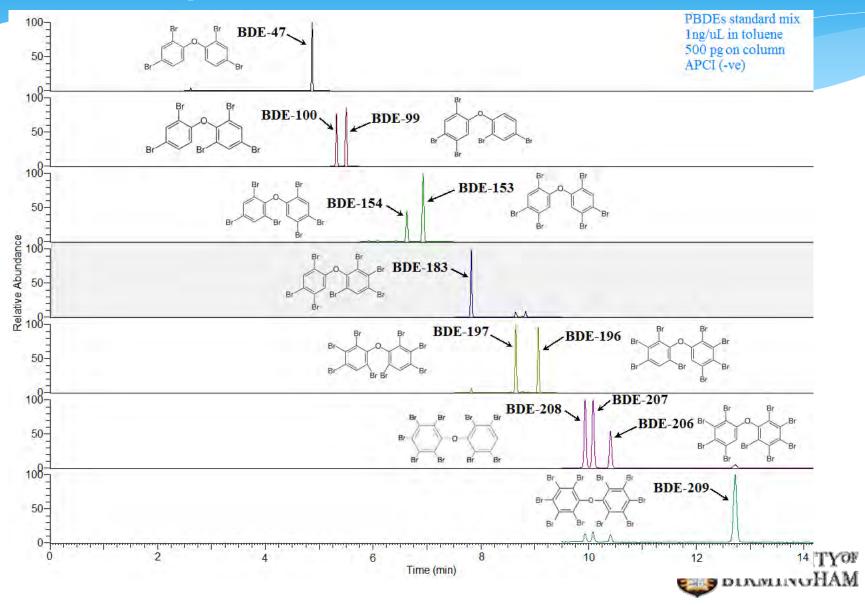


# **Results-Separation**



**Mobile phase:** 1 mM ammonium acetate (mobile phase A) and Methanol (mobile phase B), each modified with 0.1% formic acid. The elution programme commenced with 25% B ramped up to 50% B over 0.5 min, then increased linearly to 100% B over 6 minutes. This was held for 4 minutes, then decreased to 50% B over 0.5 min and kept at this composition (to equilibriate the column) for a further 1 minute.

#### **Results-Separation**



# **Results-Optimisation**

Capillary temperature (°C)	300
Source heater temperature (°C)	300
Electrospray voltage (V)	4500
Sheath gas flow (a.u.)*	15
Auxiliary gas flow (a.u.)*	10
S-lens frequence (Hz)	50
Maximum injection time (ms)	80
Automatic gain control (ions)	3 x 10 <sup>6</sup>
HCG energy (ev)	35
MS resolution (FWHM)	35000

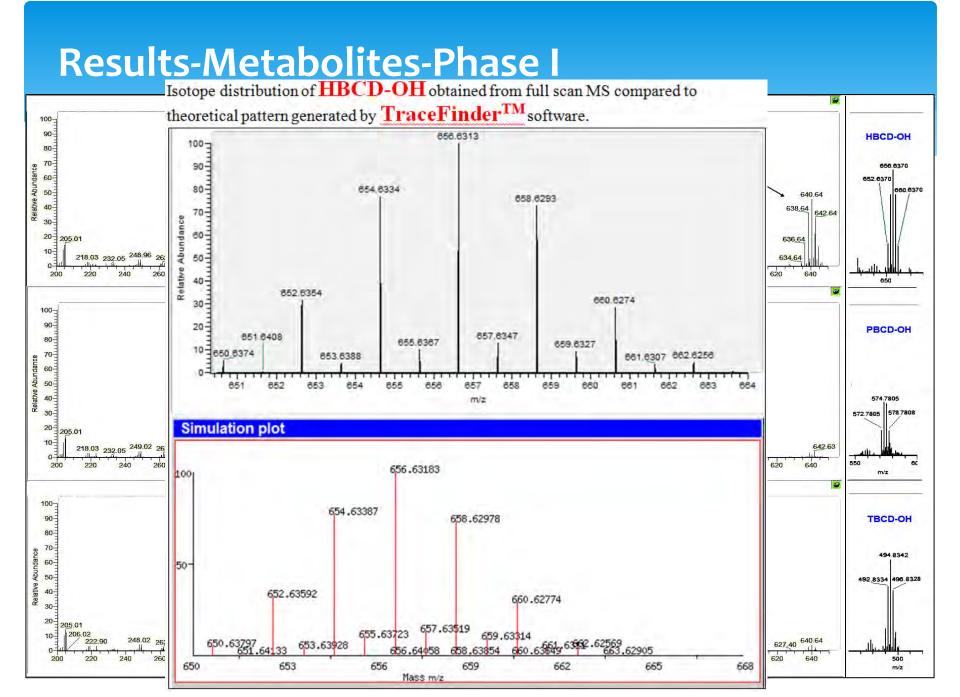


#### **Results-Metabolites**

#### \* Metabolite identification:

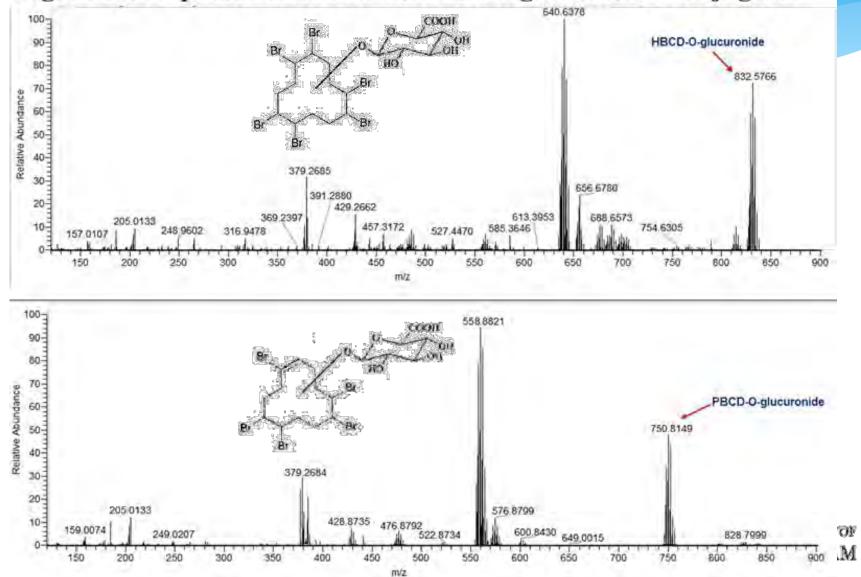
- \* MS full scan, accurate mass (4 digits)-retention times.
- \* Software (Analyst-Trace finder)
- \* AIF spectra-useful for conjugates.
- \* Confirmatory MS/MS analysis.





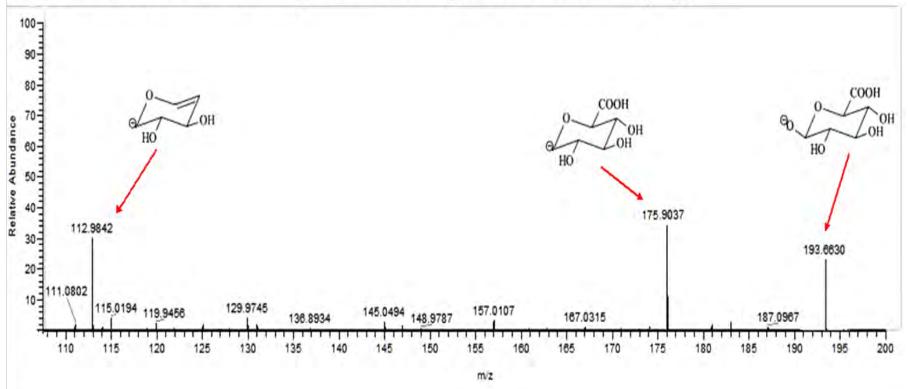
#### **Results-Metabolites-Phase II**

Fig. 3: Mass spectra of HBCD and PBCD glucuronide conjugates



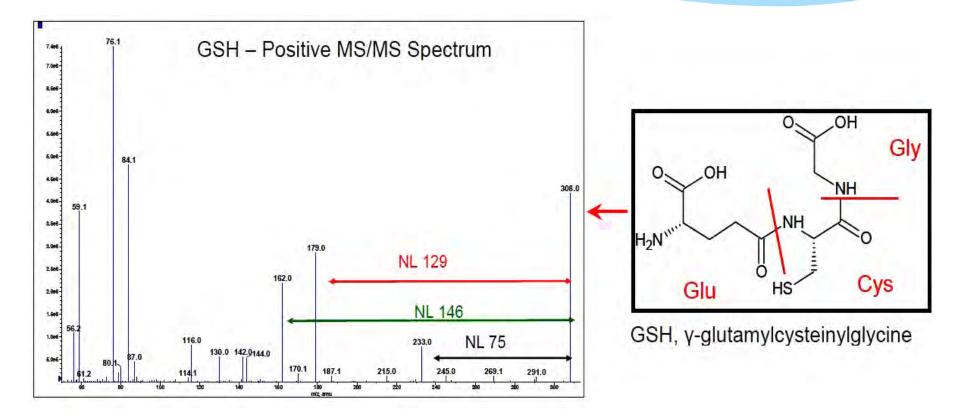
## **Metabolites-Phase II-confirmation**

Fig. 5: All ion fragmentation (AIF) spectrum showing the characteristic mass fragments of a glucuronide conjugate





#### **Metabolites-Phase II-confirmation**



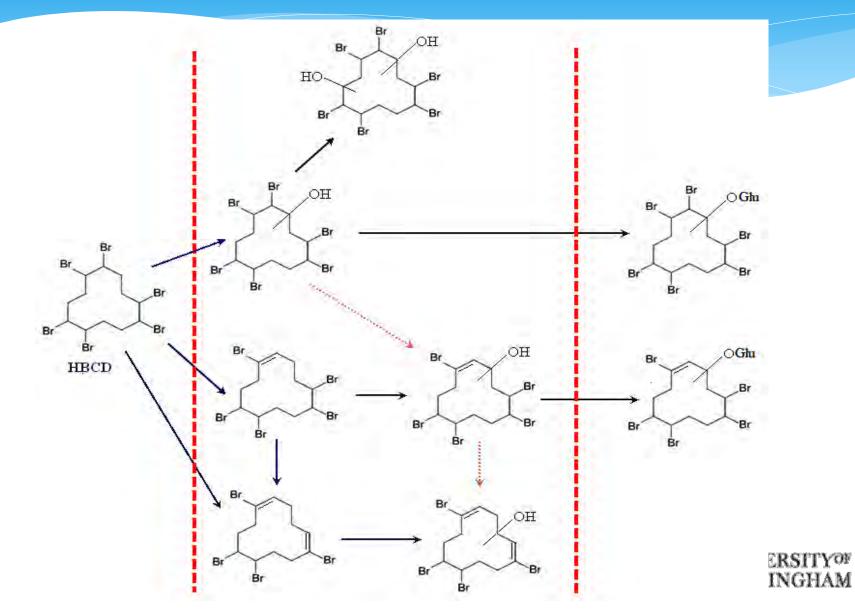


# **Metabolites-HBCDs**

Reaction	Abbreviation	Chemical	Mol. ion	Main	Ret. time
		formula	[M-H] <sup>-</sup>	fragment	(min)
Phase I					
Reductive	PBCD	$C_{6}H_{17}Br_{5}$	560.6388	80.9153	7.19, 7.58
debromination	(2 isomers)				
Reductive	TBCD	$C_6 H_{16} Br_4$	480.6618	80.9152	6.63
debromination					
Hydroxylation	HBCD-OH	$C_6 H_{17} Br_6 O$	656.6376	80.9152	5.89, 6.09,
	(5 isomers)				6.38, 6.72,
					7.11
Hydroxylation	Di-hydroxyl	$C_{6} H_{17} Br_{6} O_{2}$	672.6412	80.9152	5.08
	HBCD				
Hydroxylation	PBCD-OH	$C_6 H_{16} Br_5 O$	576.6780	80.9154	5.48, 5.71
	(2 isomers)				
Hydroxylationn	TBCD-OH	$C_6 H_{15} Br_4 O$	496.6778	80.9153	5.29
Phase II					
Glucuronidation	HBCD-O-Glu	${\rm C}_{12}{\rm H}_{26}{\rm Br}_6{\rm O}_6$	832.5766	80.9153	4.68
Glucuronidation	PBCD-O-Glu	$C_{12}H_{25}Br_5O_6$	750.8149	80.9153	3.22
		12 23 3 0			



# **Metabolic Profile-HBCD**



# **Metabolites-chlorinated PFRs**

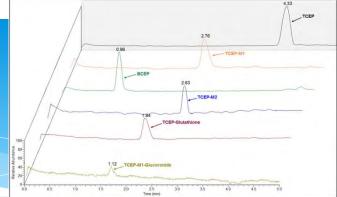
- \* Phase I:
  - \* Dealkylation: Formation of the Di-phosphate ester.
  - \* Oxidative dehalogenation: replacement of Cl with OH

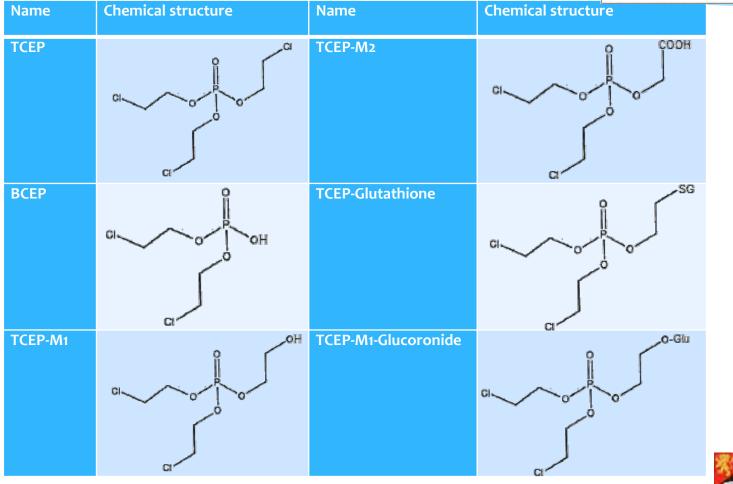
#### \* Phase II:

- \* Glucuronide conjugates
- \* Glutathione conjugates.



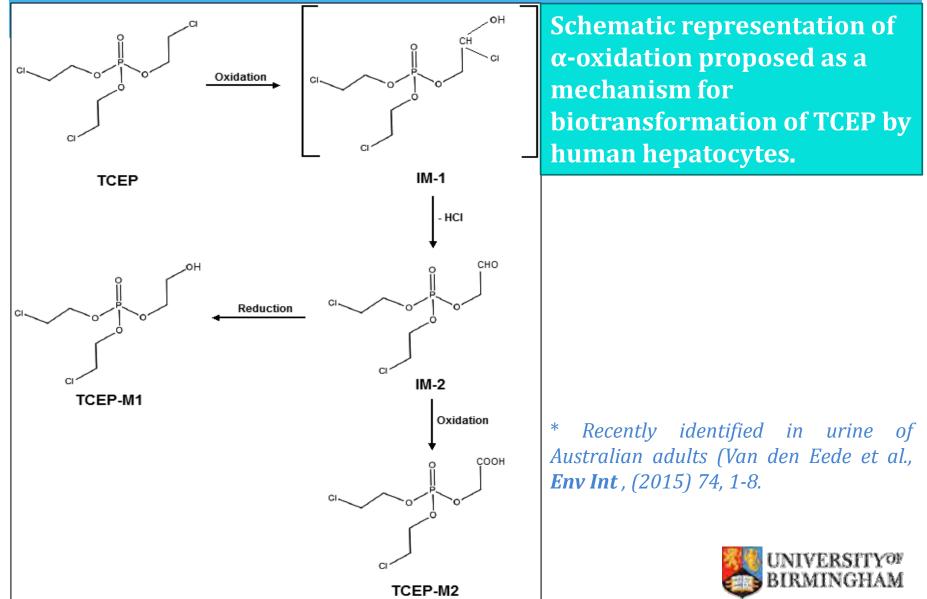
# **Metabolic Profile-TCEP**







# **Metabolic Profile-TCEP**



# **Metabolic Profile of TCiPP**

the second s		Molecular ion [M+H]+	Theoretical mass	
C9H18Cl3O4P	a do for	327.0081	327.0009	
C6 H13 Cl2 O4 P	a do de de	250.9929	251.0002	
C9 H19 Cl2 O5 P	C C C C C C C C C C C C C C C C C C C	309.0402	309.0348	
C6 H12 Cl2 O6 P	CI COOH	322.9892	323.0140	
C19 H34 Cl2 N3 O10 P S	a si	598.1295	598.1080	
	formula C9H18Cl3O4P C6 H13 Cl2 O4 P C9 H19 Cl2 O5 P C6 H12 Cl2 O6 P C19 H34 Cl2 N3	formula $C_9H_{18}Cl_3O_4P$ $4$ <	formulaion [M+H]+C9H18Cl3O4P $\int_{G} \int_{G} \int_$	



Name	Molecular formula	Chemical structure	Molecular ion [M+H]+	Theoretical mass
TDCIPP	C9 H15 Cl6 O4 P		430.8882	430.8809
BDCIPP	C6 H11 Cl4 O4 P		320.9192	320.9120
DCIPP	C3 H7 Cl2 O4 P		208.9533	208.9459
TDCIPP-M1	C9 H16 Cl5 O5 P		412.9062	412.9149
TDCIPP-M2	C9 H14 CI5 O6 P		426.8787	426.8942

COOH

,CI

0

702.0208

701.9982

C<sub>19</sub> H<sub>31</sub> Cl<sub>5</sub> N<sub>3</sub> O<sub>10</sub> P S

a.

c

TDCIPP-

Glutathione



## **Conclusions**

- \* Human HepG2 cell lines can metabolise HBCDs, TCEP, TCIPP and TDCPP present in indoor dust.
- \* HBCDs undergoes oxidative hydroxylation and reductive debromination during phase I metabolism. Penta- and Tetrabrominated derivatives were detected together with their hydroxylated metabolites. Phase II glucuronidation was observed for both HBCDs and PBCDs.
- \* The biphosphate ester was the major metabolite observed for TCEP, TCIPP and TDCPP followed by the oxidative dechlorinated metabolite. Both glucuronide and glutathione conjugates were detected as a result of Phase II metabolism.





- \* α-oxidation was proposed as a mechanism for biotransformation of PFRs by human hepatocytes.
- \* In total, 6 different brominated and chlorinated FRs and their 37 metabolites were *simultaneously* separated and identified in one run.



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