Electronic supplementary information

Imaging and spectroscopic comparison of multi-step methods to form DNA arrays based on the biotin-streptavidin system

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Figure ESI-1. The cross sectional analysis of topographic AFM micrographs (*cf.* Fig.2), recorded from epoxy-silanized SiO₂ surfaces after the successive steps of the different approaches followed for the immobilization of capture oligonucleotides in spots (*cf.* Fig.1 for used notation), and acquired from the bare SiO₂ substrate as the control (characterized by positive surface skewness, RMS = 0.22 nm and <h> = 0.43 nm).



Figure ESI-2. High resolution XPS C1s core-level surface spectra recorded after characteristic steps of the three immobilization approaches (*cf.* Fig. 1). Contributions to the C1s envelope are shown, referred to various carbon environments (see legend). Note, that the intensities of signals from carbon in carbonyl O=C (and amide O=CN) groups, that are specific for biomolecules, and of the epoxy-groups of the silane layer (non-reacted) are anti-correlated.



Figure ESI-3. TOF-SIMS microanalysis of streptavidin immobilized at the surface after the successive steps of deposition/reaction involved in the three immobilization approaches (see Fig.1 for the used notation). Secondary ion intensities (normalized per total ion intensity) characteristic for streptavidin ($C_{11}H_8NO^+$ from tryptophan) determined for uniform surfaces (samples A-C, E and G-H) as well as inside (s - spot) and outside (b - background) oligonucleotide spots (samples D, F and I). Error bars are standard deviation values determined from repetitive (3-8) measurements of the same surface.

Approach I			Approach II				Approach III			
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Mean spot intensity (RFU) of 64 spots										
26680±1541			15406±1054				6797±601			

Figure ESI-4. Fluorescence microscopy images of the spots of fluorescently-labelled oligonucleotides on glass slides, prepared according to protocols I, II, and III and the respective mean fluorescence intensity values \pm SD obtained from 64 spots.

Cumulative material distribution and complementary molecular composition maps within DNA spots, obtained with TOF-SIMS from 1.0 mm x 1.0 mm areas



Figure ESI-5. Molecular distribution of the whole material (the row 'total (-)') and composition maps of oligonucleotide (the rows 'oligo', corresponding to signal of PO_3^- and nucleotide bases fragments SUM DNA⁻) obtained by TOF-SIMS imaging of the oligonucleotide spots created following each one of the three immobilization protocols (columns D, F and I, respectively, *cf.* Fig. 1) on epoxy-silanized SiO₂ surfaces. The TOF-SIMS intensity maps of all negative ion fragments (corresponding to row 'total (-)') were used to normalize for each pixel the intensities of DNA-derived negative ions (shown after normalization as the rows 'oligo').

Cumulative material distribution and complementary molecular composition maps within DNA spots, obtained with TOF-SIMS from 0.5 mm x 0.5 mm areas



Figure ESI-6. Molecular distribution of the whole material (row 'total (-)') and composition maps (*cf.* Fig. 8) of oligonucleotide (rows 'oligo', corresponding to signal of ion PO_3^- and nucleotide bases fragments SUM DNA⁻), obtained by TOF-SIMS imaging of the DNA spots created following each one of the three immobilization protocols (columns D, F and I, respectively, *cf.* Fig. 1) on epoxy-silanized SiO₂ surfaces. The TOF-SIMS intensity maps of all positive ion fragments (row 'total (+)') were used to normalize for each pixel the intensities of positive fragment ions characteristic for the different molecules (shown after normalization as the rows 'oligo').



Figure ESI-7. Molecular distribution of the whole material (row 'total (+)') and complementary composition maps (*cf.* Fig. 9) of streptavidin (row 'Str', corresponding to signal of ion $C_9H_8N^+$ characteristic for tryptophan), BSA (row 'BSA', corresponding to signal of ion $C_5H_{10}N^+$ specific for lysine), epoxy-terminated silane GOPS (row 'epoxy', corresponding to signal of ion CH_3O^+) and

silicon substrate (row 'SiO₂', corresponding to signal of ion Si⁺), obtained by TOF-SIMS imaging of the DNA spots created following each one of the three immobilization protocols (columns D, F and I, respectively, *cf.* Fig. 1). The TOF-SIMS intensity maps of all positive ion fragments (row 'total (+)') were used to normalize for each pixel the intensities of positive fragment ions characteristic for the different molecules (shown after normalization at the rest of the rows).