

Supporting Informations

Oxidant controlled regioselective mono- and di-functionalizations of coumarins

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Instrumentation and Chemicals:

All the reagents were commercial grade and purified according to the established procedures. Organic extracts were dried over anhydrous sodium sulphate. Solvents were removed in a rotary evaporator under reduced pressure. Silica gel (60-120 mesh size) was used for the column chromatography. Reactions were monitored by TLC on silica gel 60 F₂₅₄ (0.25 mm). NMR spectra were recorded in CDCl₃ with tetramethylsilane as the internal standard for ¹H NMR (400 and 600 MHz), CDCl₃ solvent as the internal standard for ¹³C NMR (100 and 150 MHz). MS spectra were recorded using ESI mode. IR spectra were recorded in KBr or neat. Starting materials (3-substituted coumarins) (**1-15**) are prepared by reacting salicylaldehydes with active methylene compounds (diethyl malonate / ethylcyanoacetate / ethyl acetoacetate etc.) in presence of catalytic amount of piperidine using Knoevenagel condensation.¹

1. M. Ghandi, A.-T. Ghomi, and M. Kubicki, *J. Org. Chem.*, 2013, **78**, 2611.

Experimental procedure:

(A) Synthesis of 3-cyclohexyl-2H-chromen-2-one (**1a**) from 3-acetyl-2H-chromen-2-one (**1**) and cyclohexane (**a**)

To an oven dried 25 mL round bottom flask fitted with reflux condenser, 3-acetylcoumarin (**1**) (0.047g, 0.25 mmol), di-*tert*-butylperoxide DTBP (0.146g, 1.0 mmol), Fe(acac)₃ (0.004g, 0.013 mmol) and cyclohexane (**a**) (0.5 mL, 4.6 mmol) were added together in chlorobenzene (1.0 mL) solvent. The reaction mixture was then heated in an oil bath at 110 °C. The progress of the reaction was monitored by TLC and after completion of reaction (12 h) solvents were evaporated under reduced pressure. The reaction mixture was then cooled to room

temperature, admixed with water (2 mL). It was then extracted with ethyl acetate (3 x 10 mL), dried over anhydrous sodium sulphate Na_2SO_4 , and evaporated under reduced pressure. The crude product obtained here was further purified over a short column of silica gel (hexane / ethyl acetate, 10:0.1) to give pure 3-cyclohexyl-2*H*-chromen-2-one (**1a**) (0.030g, yield 52%). The identity and purity of the product was confirmed by spectroscopic analysis.

(B) *Synthesis of 3-Acetyl-3-(tert-butylperoxy)-4-cyclohexylchroman-2-one (**1a'**) from 3-acetyl-2*H*-chromen-2-one (**1**), tert-butyl hydroperoxide and cyclohexane (**a**)*

To an oven-dried 25 mL round bottom flask fitted with a reflux condenser was added 3-acetylcoumarin (**1**) (0.047g, 0.25 mmol), decane solution of TBHP (5–6 M) (200 μL , 1.0 mmol), AcOH (30 μL , 0.50 mmol), and cyclohexane (**a**) (0.5 mL, 4.6 mmol) in chlorobenzene (1.0 mL) solvent. The reaction mixture was heated in an oil bath at 110 °C. After completion (3 h) of the reaction, solvents were evaporated under reduced pressure. The reaction mixture was then cooled to room temperature and admixed with water (2 mL). The product was extracted with ethyl acetate (2 x 10 mL), dried over anhydrous sodium sulphate (Na_2SO_4), and evaporated under reduced pressure. The crude product so obtained was purified over a column of silica gel (hexane / ethyl acetate, 10:0.1) to give pure 3-acetyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (**1a'**) (0.058g, yield 65%). The identity and purity of the product was confirmed by spectroscopic analysis.

Crystallographic Description

Crystal data were collected with Bruker Smart Apex-II CCD diffractometer using graphite monochromated MoK α radiation ($\lambda = 0.71073 \text{ \AA}$) at 298 K. Cell parameters were retrieved using SMART^a software and refined with SAINT^a on all observed reflections. Data reduction was performed with the SAINT software and corrected for Lorentz and polarization effects. Absorption corrections were applied with the program SADABS^b. The structure was solved by direct methods implemented in SHELX-97^c program and refined by full-matrix least-squares methods on F2. All non-hydrogen atomic positions were located in difference Fourier maps and refined anisotropically. The hydrogen atoms were placed in their geometrically generated positions. colourless crystals were isolated in rectangular shape from acetonitrile at room temperature.

- a. SMART V 4.043 Software for the CCD Detector System; Siemens Analytical Instruments Division: Madison, WI, 1995.
- b. SAINT V 4.035 Software for the CCD Detector System; Siemens Analytical Instruments Division: Madison, WI, 1995.
- c. Sheldrick, G. M. SHELXL-97, Program for the Refinement of Crystal Structures; University of Göttingen: Göttingen (Germany), 1997.

CCDC number for compound 1c: CCDC 1412809. This data can be obtained free of charge from the Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/datarequest/cif.

Crystallographic description of **1c:** Crystal dimension (mm): 0.38 x 0.20 x 0.16. C₁₇H₂₀O₂, Mr = 256.33. triclinic, space group p -1; a = 6.7321 (3) Å, b = 9.3720 (4) Å, c = 11.4758 (5) Å; α = 95.498 (3)°, β = 99.490 (3)°, γ = 104.725 (3)°, V = 683.45 (5) Å³; Z = 2; ρ_{cal} = 1.246 g/cm³; μ (mm⁻¹) = 0.080; F (000) = 276.0; Reflection collected / unique = 2348 / 1909; Refinement method = Full-matrix least-squares on F²; Final R indices [I>2σ_I] R1 = 0.0389, wR2 = 0.0985, R indices (all data) R1 = 0.0480, wR2 = 0.1132; goodness of fit = 1.079.

Table S1. Optimization of reaction parameters

Entry	Catalyst (mol %)	Additive (equiv.)	Oxidant ^d (equiv.)	Yield (%) ^{a,b}
1	—	—	DTBP (3.0)	20 00
2	—	—	DTBP (4.0)	40 00
3	Cu(OAc) ₂ (10.0)	—	DTBP (4.0)	34 00
4	CuCl ₂ (10.0)	—	DTBP (4.0)	38 00
5	FeCl ₂ (10.0)	—	DTBP (4.0)	26 00
6	Fe ₃ O ₄ (10.0)	—	DTBP (4.0)	Trace 00
7	Fe(acac) ₃ (10.0)	—	DTBP (4.0)	53 00
8	Fe(acac)₃ (5.0)	—	DTBP (4.0)	52 00
9	Fe(acac) ₃ (5.0)	AcOH (1.0)	DTBP (4.0)	47 00
10	—	AcOH (1.0)	DTBP (4.0)	00 00
11	—	PTSA (1.0)	DTBP (4.0)	00 00
12 ^c	—	AcOH (1.0)	TBHP (4.0)	00 54
13 ^c	—	PTSA (1.0)	TBHP (4.0)	00 00
14 ^c	—	TfOH (1.0)	TBHP (4.0)	00 00
15 ^c	—	PhCOOH (1.0)	TBHP (4.0)	00 30
16 ^c	—	PivOH (1.0)	TBHP (4.0)	00 27
17^c	—	AcOH (2.0)	TBHP (4.0)	00 65
18 ^c	—	—	TBHP (4.0)	00 08

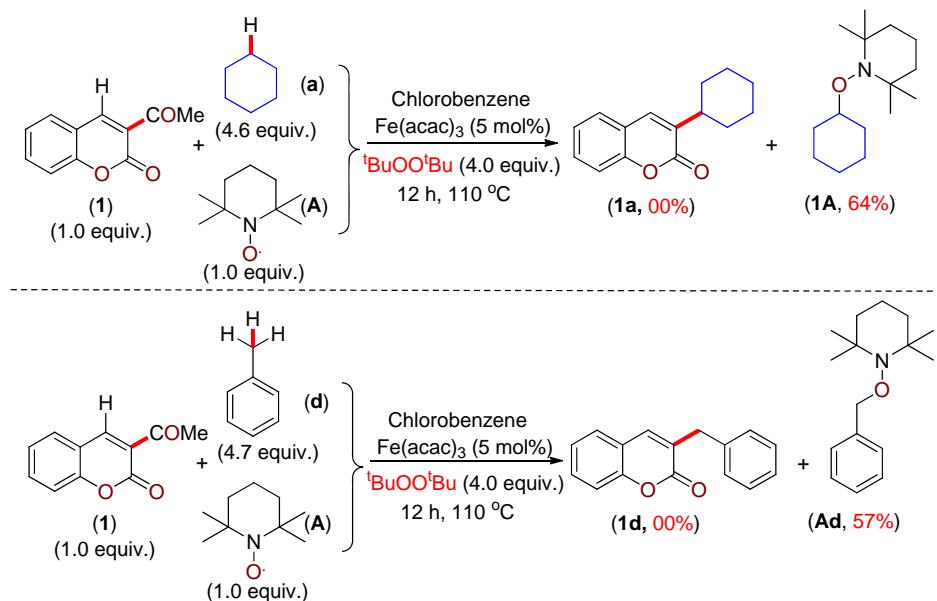
^aReaction conditions: **1** (0.25 mmol), cyclohexane (**a**) (0.5 mL, 4.6 mmol) in chlorobenzene (1.0 mL) at 110 °C for 12 h. ^bIsolated yield. ^cReaction performed for 3 h. ^dOxidants were used portion wise (four times for both DTBP and TBHP).

Mechanistic investigation:

Trapping of cyclohexyl and benzyl intermediates with TEMPO during mono-functionalization:

An oven-dried 25 mL round bottom flask was charged with 3-acetylcoumarin (**1**) (0.047g, 0.25 mmol), di-*tert*-butylperoxide DTBP (0.146g, 1.0 mmol), Fe(acac)₃ (0.004g, 0.013 mmol), TEMPO (**A**) (0.039 g, 0.25 mmol) and cyclohexane (**a**) (0.5 mL, 4.6 mmol) in chlorobenzene (1.0 mL). The flask was fitted with a reflux condenser and the reaction mixture was stirred in a preheated oil bath at 110 °C for 12 h. After 12 h of the reaction and usual work up the cyclohexyl-TEMPO adduct 1-(cyclohexyloxy)-2,2,6,6-tetramethylpiperidine (**1A**) was obtained in 64% isolated yield with no traces of the desired product (**1a**).

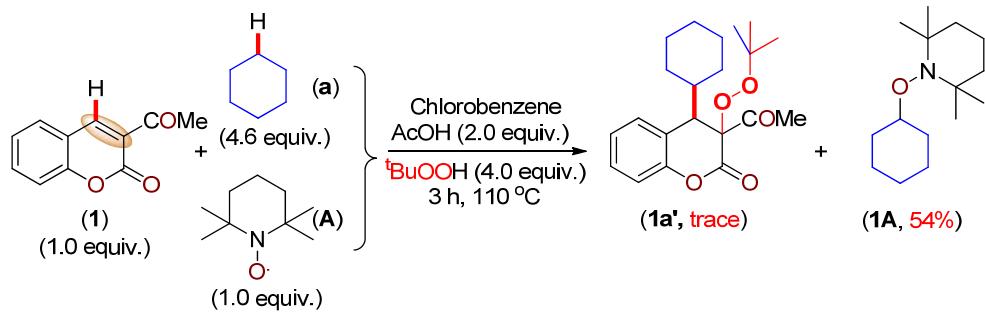
Similar reaction between 3-acetylcoumarin (**1**) (0.047g, 0.25 mmol) and toluene (**a**) (0.5 mL, 4.7 mmol) in the presence of radical scavenger TEMPO (**A**) (0.039 g, 0.25 mmol) under otherwise identical reaction conditions to that of above provided benzyl-TEMPO adduct 1-(benzyloxy)-2,2,6,6-tetramethylpiperidine (**1B**) in 57% isolated yield with no trace of desired product (**1a**).



Scheme S 2: Control experiments with radical scavenger TEMPO

Trapping of cyclohexyl intermediate with TEMPO during di-functionalization:

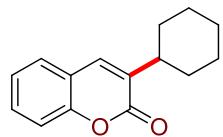
An oven-dried 25 mL round bottom flask was charged with 3-acetylcoumarin (**1**) (0.047g, 0.25 mmol), decane solution of TBHP (5–6 M) (200 μ L, 1.0 mmol), AcOH (30 μ L, 0.50 mmol), TEMPO (**A**) (0.039 g, 0.25 mmol) and cyclohexane (**a**) (0.5 mL, 4.6 mmol) in chlorobenzene (1.0 mL). The flask was fitted to a reflux condenser and the reaction mixture was stirred in a preheated oil bath at 110 °C. After 3 h of the reaction and usual work up, the cyclohexyl-TEMPO adduct **1A'** (cyclohexyloxy)-2,2,6,6-tetramethylpiperidine was obtained in 54% isolated yield along with a trace of the desired product (**1a'**).



Scheme S3: control experiment with radical scavenger TEMPO

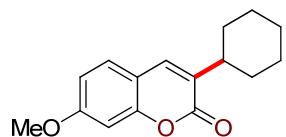
Spectral Data:

3-Cyclohexyl-2*H*-chromen-2-one (1a):



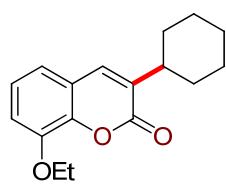
Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.47–7.44 (m, 3H), 7.31 (d, 1H, J = 8.4 Hz), 7.24 (t, 1H, J = 7.2 Hz), 2.81–2.76 (m, 1H), 1.98 (dd, 2H, J = 15.6, 4.2 Hz), 1.87–1.84 (m, 2H), 1.80–1.77 (m, 1H), 1.49–1.41 (m, 2H), 1.34–1.24 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 161.8, 152.9, 136.5, 135.2, 130.2, 127.5, 124.4, 119.9, 116.5, 38.4, 32.3, 26.7, 26.4; IR (KBr, cm^{-1}): 3058, 2928, 2850, 1710, 1652, 1632, 1609, 1488, 1455, 1387, 1277, 1255, 1234, 1184, 1173, 1136, 1064, 1042, 986, 956, 924, 890, 781, 754; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{16}\text{O}_2$ ($\text{M} + \text{H}^+$) 229.1229, found 229.1224.

3-Cyclohexyl-7-methoxy-2*H*-chromen-2-one (2a):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.38 (s, 1H), 7.33 (d, 1H, J = 8.4 Hz), 6.83–6.79 (m, 2H), 3.85 (s, 3H), 2.75–2.71 (m, 1H), 1.96 (d, 2H, J = 12.0 Hz), 1.86–1.82 (m, 2H), 1.78–1.75 (m, 1H), 1.45–1.40 (m, 2H), 1.30–1.26 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 162.1, 161.9, 154.6, 136.6, 131.2, 128.4, 113.5, 112.5, 100.6, 55.9, 38.2, 32.4, 26.8, 26.4; IR (KBr, cm^{-1}): 2959, 2924, 2850, 1701, 1652, 1628, 1593, 1572, 1464, 1433, 1403, 1297, 1265, 1158, 1138, 1034, 964, 937, 774; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{18}\text{O}_3$ ($\text{M} + \text{H}^+$) 259.1335, found 259.1327.

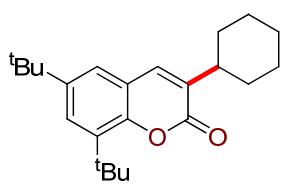
3-Cyclohexyl-8-ethoxy-2*H*-chromen-2-one (3a):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.41 (s, 1H), 7.14 (t, 1H, J = 8.4 Hz), 6.99 (d, 2H, J = 7.2 Hz), 4.19–4.16 (q, 2H, J = 7.2 Hz), 2.81–2.76 (m, 1H), 1.97 (d, 2H, J = 12.6 Hz), 1.86–1.82 (m, 2H), 1.78–1.76 (m, 1H), 1.48 (t, 3H, J = 10.0 Hz), 1.46–1.41 (m,

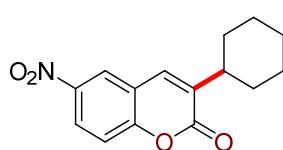
2H), 1.32–1.23 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 161.3, 146.5, 142.8, 136.6, 135.3, 124.2, 120.7, 118.9, 113.9, 65.1, 38.3, 32.3, 26.7, 26.4, 14.9; IR (KBr, cm^{-1}): 2977, 2926, 2852, 1719, 1608, 1579, 1471, 1449, 1392, 1355, 1276, 1181, 1114, 1098, 1064, 1044, 982, 770, 731; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{20}\text{O}_3$ ($\text{M} + \text{H}^+$) 273.1492, found 273.1485.

6,8-Di-*tert*-butyl-3-cyclohexyl-2*H*-chromen-2-one (4a):



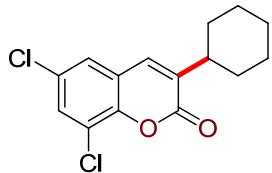
Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.49 (d, 1H, $J = 2.4$ Hz), 7.43 (s, 1H), 7.26 (s, 1H), 2.79–2.76 (m, 1H), 1.97 (d, 2H, $J = 12.0$ Hz), 1.86–1.83 (m, 2H), 1.78–1.76 (m, 1H), 1.51 (s, 9H), 1.46–1.42 (m, 3H), 1.35 (s, 9H), 1.33–1.29 (m, 2H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 161.6, 149.6, 146.5, 137.8, 137.0, 133.8, 125.8, 122.2, 119.7, 38.1, 35.2, 34.8, 32.4, 31.6, 30.1, 26.7, 26.4; IR (KBr, cm^{-1}): 2961, 2929, 2853, 1711, 1586, 1477, 1445, 1393, 1363, 1243, 1217, 1170, 1135, 1068, 1032, 1004, 983, 939, 888, 783; HRMS (ESI) calcd for $\text{C}_{23}\text{H}_{32}\text{O}_2$ ($\text{M} + \text{H}^+$) 341.2482, found 341.2470.

3-Cyclohexyl-6-nitro-2*H*-chromen-2-one (5a):



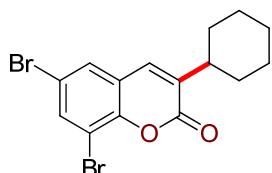
Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 8.39 (d, 1H, $J = 2.4$ Hz), 8.33 (d, 1H, $J = 9.6$ Hz), 7.51 (s, 1H), 7.42 (d, 1H, $J = 9.0$ Hz), 2.82–2.77 (m, 1H), 1.99 (d, 2H, $J = 12.0, 4.2$ Hz), 1.88 (dt, 2H, $J = 13.2, 3.0$ Hz), 1.80 (d, 1H, $J = 13.2$ Hz), 1.49–1.42 (m, 2H), 1.35–1.27 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 160.1, 156.5, 144.2, 137.7, 135.2, 125.5, 123.4, 119.9, 117.6, 38.8, 32.2, 26.6, 26.2; IR (KBr, cm^{-1}): 2927, 2855, 1728, 1652, 1632, 1617, 1529, 1486, 1342, 1270, 1169, 1090, 1060, 1041, 983, 935, 844, 836, 749, 668; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{15}\text{NO}_4$ ($\text{M} + \text{H}^+$) 274.1081, found 274.1074.

6,8-Dichloro-3-cyclohexyl-2*H*-chromen-2-one (6a):



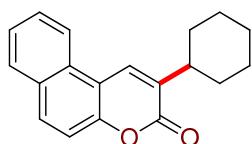
Semi-solid; ^1H NMR (CDCl_3 , 400 MHz): δ 7.50 (d, 1H, $J = 2.4$ Hz), 7.35–7.34 (m, 2H), 2.81–2.75 (m, 1H), 1.98–1.95 (m, 2H), 1.88–1.77 (m, 3H), 1.50–1.39 (m, 2H), 1.33–1.19 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 159.9, 147.3, 137.4, 134.9, 130.6, 129.4, 125.4, 122.3, 121.7, 38.6, 32.2, 26.6, 26.3; IR (KBr, cm^{-1}): 2927, 2853, 1733, 1660, 1635, 1564, 1452, 1352, 1252, 1172, 1135, 1063, 992, 888, 859, 830, 778, 736; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{14}\text{Cl}_2\text{O}_2$ ($\text{M} + \text{H}^+$) 297.0450, found 297.0442.

3-Cyclohexyl-6-nitro-2*H*-chromen-2-one (7a):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.79 (s, 1H), 7.53 (s, 1H), 7.31 (s, 1H), 2.80–2.76 (m, 1H), 1.96 (d, 2H, $J = 12.0$ Hz), 1.86–1.84 (m, 2H), 1.79–1.76 (m, 1H), 1.45–1.41 (m, 2H), 1.35–1.27 (m, 3H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 160.0, 148.9, 137.3, 136.1, 134.8, 129.1, 122.2, 116.8, 111.0, 38.6, 32.2, 26.6, 26.3; IR (KBr, cm^{-1}): 2964, 2922, 2846, 1735, 1658, 1641, 1635, 1448, 1260, 1020, 796; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{14}\text{Br}_2\text{O}_2$ ($\text{M} + \text{H}^+$) 384.9433, found 384.9424.

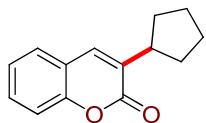
2-Cyclohexyl-3*H*-benzo[*f*]chromen-3-one (8a):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 8.28 (d, 1H, $J = 8.4$ Hz), 8.24 (s, 1H), 7.91 (t, 2H, $J = 8.4$ Hz), 7.67 (t, 1H, , $J = 7.8$ Hz), 7.55 (t, 1H, $J = 7.8$ Hz), 7.45 (d, 1H, $J = 9.0$ Hz), 2.88–2.86 (m, 1H), 2.06 (d, 2H, $J = 12.0$ Hz), 1.91–1.89 (m, 2H), 1.81 (d, 1H, $J = 13.2$ Hz), 1.51–1.42 (m, 4H), 1.34–1.27 (m, 1H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 161.8, 152.2, 134.3, 132.2, 131.8, 130.5, 129.2, 128.0, 125.9, 121.7, 116.9, 113.8, 38.8, 32.5, 26.8, 26.4; IR (KBr, cm^{-1}): 3066, 2921, 2849, 1705, 1635, 1591, 1517, 1448, 1438, 1236, 1213, 1171, 1135, 1088, 1075, 1044, 986, 882, 813,

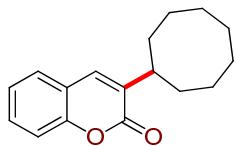
782, 738; HRMS (ESI) calcd for C₁₉H₁₈O₂ (M + H⁺) 279.1386, found 279.1389

3-Cyclopentyl-2*H*-chromen-2-one (1b):



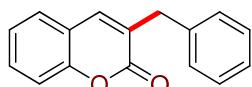
Semi-solid; ¹H NMR (CDCl₃, 400 MHz): δ 7.50 (s, 1H), 7.45 (m, 2H), 7.31 (m, 1H), 7.26–7.23 (m, 1H), 3.20–3.11 (m, 1H), 2.14–2.06 (m, 2H), 1.85–1.76 (m, 4H), 1.73–1.53 (m, 2H); ¹³C NMR (CDCl₃, 150 MHz): δ 161.9, 153.1, 136.2, 133.7, 130.6, 127.4, 124.4, 119.8, 116.5, 40.9, 31.9, 25.3; IR (KBr, cm⁻¹): 2955, 2872, 1722, 1630, 1609, 1454, 1276, 1224, 1173, 1121, 1057, 1022, 948, 922, 755; HRMS (ESI) calcd for C₁₄H₁₄O₂ (M + H⁺) 215.1073, found 215.1064.

3-Cyclooctyl-2*H*-chromen-2-one (1c):



Crystalline needles; m.p. 85–86 °C; ¹H NMR (CDCl₃, 600 MHz): δ 7.46 (s, 1H), 7.44 (d, 2H, *J* = 6.0 Hz), 7.29 (d, 1H, *J* = 8.4 Hz), 7.24 (t, 1H, *J* = 7.5 Hz), 3.10–3.03 (m, 1H), 1.84–1.82 (m, 2H), 1.80–1.73 (m, 4H), 1.72–1.64 (m, 8H); ¹³C NMR (CDCl₃, 150 MHz): δ 161.7, 152.9, 139.8, 136.5, 130.5, 127.4, 124.3, 119.8, 116.4, 38.2, 31.8, 26.9, 26.4, 25.8; IR (KBr, cm⁻¹): 3057, 2963, 2922, 2856, 1717, 1627, 1608, 1489, 1473, 1462, 1454, 1388, 1360, 1281, 1274, 1258, 1230, 1182, 1274, 1258, 1230, 1181, 1174, 1146, 1124, 1058, 1011, 1027, 1000, 971, 956, 932, 923, 872, 853, 830, 777, 754, 733; HRMS (ESI) calcd for C₁₇H₂₀O₂ (M + H⁺) 257.1543, found 257.1539.

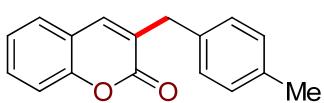
3-Benzyl-2*H*-chromen-2-one (1d):



Semi-solid; ¹H NMR (CDCl₃, 600 MHz): δ 7.44 (t, 1H, *J* = 7.2 Hz), 7.34–7.31 (m, 2H), 7.29–7.24 (m, 5H), 7.19 (t, 2H, *J* = 7.2 Hz), 3.86 (s, 2H); ¹³C NMR (CDCl₃, 150 MHz): δ 161.9, 153.3, 139.5, 137.8, 130.9, 130.0, 129.6, 128.9, 127.6, 127.0, 124.5,

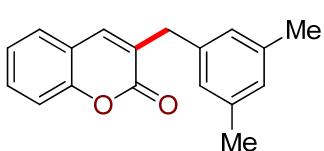
119.6, 116.6, 36.8; IR (KBr, cm^{-1}): 3022, 2988, 2917, 2841, 1712, 1631, 1608, 1489, 1455, 1434, 1387, 1224, 147, 1123, 1075, 1051, 1028, 956, 930, 825, 753, 731, 726, 700; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{12}\text{O}_2$ ($\text{M} + \text{H}^+$) 237.0917, found 237.0920.

3-(4-Methylbenzyl)-2*H*-chromen-2-one (1e):



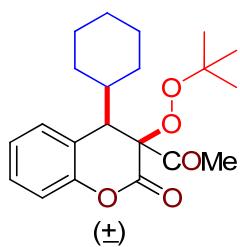
Semi-solid; ^1H NMR (CDCl_3 , 400 MHz): δ 7.44 (t, 1H, $J = 7.4$ Hz), 7.35–7.29 (m, 2H), 7.25–7.23 (m, 1H), 7.21–7.14 (m, 5H), 3.82 (s, 2H), 2.34 (s, 3H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 161.2, 153.2, 139.3, 136.6, 134.7, 130.9, 129.9, 129.7, 129.5, 127.6, 124.4, 119.7, 116.6, 36.3, 21.3; IR (KBr, cm^{-1}): 2956, 2923, 2846, 1709, 1631, 1608, 1533, 1513, 1487, 1455, 1388, 1287, 1230, 1125, 1050, 956, 786, 752, 722; HRMS (ESI) calcd for $\text{C}_{17}\text{H}_{14}\text{O}_2$ ($\text{M} + \text{H}^+$) 251.1073, found 251.1086.

3-(3,5-Dimethylbenzyl)-2*H*-chromen-2-one (1f):



Semi-solid; ^1H NMR (CDCl_3 , 400 MHz): δ 7.39 (t, 1H, $J = 7.4$ Hz), 7.30 (d, 1H, $J = 7.6$ Hz), 7.25–7.19 (m, 2H), 7.15 (t, 1H, $J = 7.2$ Hz), 6.83 (s, 3H), 3.75 (s, 2H), 2.24 (s, 6H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 161.9, 153.3, 139.4, 138.5, 137.7, 130.9, 129.8, 128.7, 127.6, 127.4, 124.4, 119.7, 116.6, 36.5, 21.5; IR (KBr, cm^{-1}): 3043, 3014, 2995, 2919, 2849, 1714, 1632, 1606, 487, 1456, 1429, 1386, 1257, 1232, 1196, 1163, 1124, 1055, 958, 923, 854, 837, 753, 704; HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{16}\text{O}_2$ ($\text{M} + \text{H}^+$) 265.1229, found 265.1220.

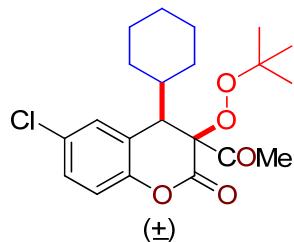
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (1a'):



Solid; m.p. 180–181 °C; ^1H NMR (CDCl_3 , 400 MHz): δ 7.24–7.19 (m, 1H), 7.06–6.99 (m, 2H), 6.96 (d, 1H, $J = 8.0$ Hz), 2.90 (d, 1H, $J = 4.2$ Hz), 2.40 (s, 3H), 1.69–1.44 (m, 6H), 1.24–1.04 (m, 3H), 0.94 (s, 9H), 0.84–0.73 (m, 1H), 0.56–0.46 (m, 1H); ^{13}C NMR

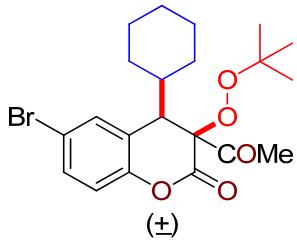
(CDCl₃, 150 MHz): δ 203.4, 161.2, 151.1, 130.2, 128.6, 123.9, 121.1, 116.3, 86.0, 82.3, 48.9, 38.4, 32.3, 27.1, 26.9, 26.7, 26.2, 25.9, 25.8; IR (KBr, cm⁻¹): 2990, 2972, 2935, 2952, 1770, 1714, 1611, 1587, 1499, 1462, 1442, 1420, 1390, 1380, 1365, 1352, 1301, 1260, 1247, 1198, 1174, 1124, 1068, 1015, 918, 876, 776, 768; HRMS (ESI) calcd for C₂₁H₂₈O₅ (M + Na⁺) 383.1834, found 383.1842.

3-Acetyl-3-(*tert*-butylperoxy)-6-chloro-4-cyclohexylchroman-2-one (13a'):



Solid; m.p. 168–170 °C; ¹H NMR (CDCl₃, 400 MHz): δ 7.22–7.18 (m, 1H), 7.00 (d, 1H, J = 2.4 Hz), 6.91 (d, 1H, J = 8.8 Hz), 2.87 (d, 1H, J = 2.8 Hz), 2.40 (s, 3H), 1.66–1.63 (m, 2H), 1.54–1.47 (m, 4H), 1.22–1.04 (m, 3H), 0.96 (s, 9H), 0.92–0.78 (m, 1H), 0.55–0.45 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 203.0, 160.5, 149.7, 129.8, 128.9, 128.6, 122.8, 117.5, 85.5, 82.5, 48.7, 38.2, 32.2, 27.0, 26.8, 26.5, 26.2, 25.8, 25.7; IR (KBr, cm⁻¹): 2984, 2942, 2921, 2853, 1790, 1725, 1482, 1455, 1415, 1367, 1351, 1273, 1260, 1226, 1217, 1194, 1180, 1148, 1120, 1094, 1083, 1066, 1041, 1034, 930, 911, 896, 891, 882, 870, 848, 812, 787, 759, 745, 707; HRMS (ESI) calcd for C₂₁H₂₇ClO₅ (M + Na⁺) 417.1445, found 417.1436.

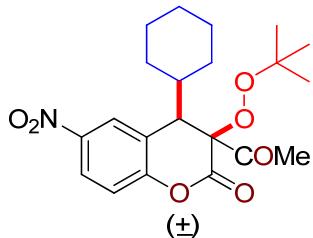
3-Acetyl-6-bromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (14a'):



Solid; m.p. 155–158 °C; ¹H NMR (CDCl₃, 400 MHz): δ 7.40 (dd, 1H, J_1 = 8.0 Hz, J_2 = 2.4 Hz), 7.21 (d, 1H, J = 2.4 Hz), 6.92 (d, 1H, J = Hz), 2.93 (d, 1H, J = 2.8 Hz), 2.46 (s, 3H), 1.73–1.69 (m, 2H), 1.61–1.53 (m, 4H), 1.25–1.11 (m, 3H), 1.02 (s, 9H), 0.91–0.86 (m, 1H), 0.58–0.55 (m, 1H); ¹³C NMR (CDCl₃, 150 MHz): δ 202.2, 160.6, 150.3, 132.7, 131.7, 131.2, 123.4, 118.9, 117.9, 85.6, 82.6, 48.7, 38.3, 32.3, 27.1, 26.9, 26.6, 26.3, 25.9, 25.7; IR (KBr, cm⁻¹): 2940, 2926, 2850, 1791, 1700, 1697, 1681, 1650, 1633, 1560,

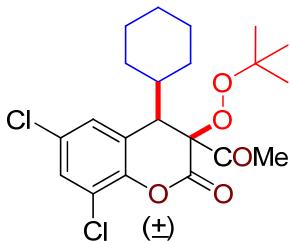
1478, 1450, 1412, 1389, 1364, 1260, 1245, 1224, 1175, 1153, 1092, 1026, 912, 891, 873, 858, 810, 775; HRMS (ESI) calcd for $C_{21}H_{27}BrO_5$ ($M + Na^+$) 461.0940, found 461.0932.

3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-6-nitrochroman-2-one (5a'):



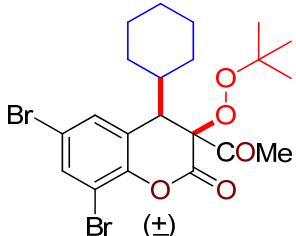
Semi-solid; 1H NMR ($CDCl_3$, 400 MHz): δ 8.19 (dd, 1H, $J_1 = 8.8$ Hz, $J_2 = 2.4$ Hz), 7.99 (d, 1H, $J = 2.8$ Hz), 7.15 (d, 1H, $J = 8.8$ Hz), 3.07 (d, 1H, $J = 2.8$ Hz), 2.47 (s, 3H), 1.71–1.51 (m, 6H), 1.27–1.05 (m, 3H), 0.99 (s, 9H), 0.88–0.79 (m, 1H), 0.54–0.44 (m, 1H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 202.4, 159.6, 155.5, 143.8, 125.7, 124.7, 122.6, 117.1, 85.2, 82.8, 48.8, 38.2, 32.1, 27.0, 26.9, 26.4, 26.2, 25.8, 25.6; IR (KBr, cm^{-1}): 3092, 2984, 2924, 2850, 1798, 1720, 1627, 1586, 1559, 1525, 1483, 1450, 1433, 1392, 1367, 1341, 1308, 1260, 1233, 1219, 1184, 1150, 1112, 1087, 1066, 1044, 1029, 1010, 967, 939, 915, 902, 868, 841, 812, 777, 753, 752, 740; HRMS (ESI) calcd for $C_{21}H_{27}NO_7$ ($M + Na^+$) 428.1685, found 428.1676.

3-Acetyl-3-(*tert*-butylperoxy)-6,8-dichloro-4-cyclohexylchroman-2-one (6a'):



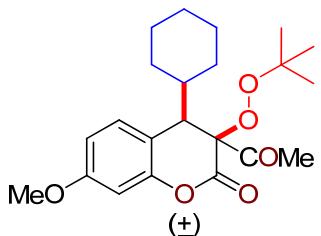
Semi-solid; 1H NMR ($CDCl_3$, 400 MHz): δ 7.38 (s, 1H), 6.98 (s, 1H), 2.95 (d, 1H, $J = 2.4$ Hz), 2.47 (s, 3H), 1.73–1.70 (m, 2H), 1.63–1.55 (m, 4H), 1.25–1.12 (m, 3H), 1.04 (s, 9H), 0.95–0.88 (m, 1H), 0.62–0.54 (m, 1H); ^{13}C NMR ($CDCl_3$, 100 MHz): δ 202.5, 159.4, 145.7, 129.0, 128.8, 128.2, 124.3, 122.1, 85.2, 82.6, 49.0, 38.0, 32.2, 27.0, 26.9, 26.4, 26.0, 25.7, 25.6; IR (KBr, cm^{-1}): 3084, 2979, 2928, 2852, 1795, 1721, 1460, 1420, 1363, 1262, 1249, 1212, 1182, 1154, 1123, 1100, 1070, 1044, 1014, 973, 937, 917, 900, 881, 870, 855, 825, 786, 754, 746; HRMS (ESI) calcd for $C_{21}H_{26}Cl_2O_5$ ($M + Na^+$) 451.1055, found 451.1049.

3-Acetyl-6,8-dibromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (7a'):



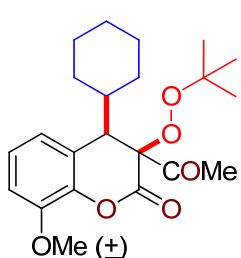
Semi-solid; ^1H NMR (CDCl_3 , 400 MHz): δ 7.64 (d, 1H, J = 2.4 Hz), 7.13 (d, 1H, J = 2.0 Hz), 2.90 (d, 1H, J = 3.2 Hz), 2.44 (s, 3H), 1.70–1.48 (m, 6H), 1.24–1.09 (m, 3H), 1.01 (s, 9H), 0.92–0.86 (m, 1H), 0.55–0.51 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 202.6, 159.5, 147.4, 134.7, 131.7, 124.7, 116.4, 111.0, 85.3, 82.7, 49.2, 38.1, 32.3, 27.2, 27.1, 26.8, 26.5, 26.2, 25.8, 25.6; IR (KBr, cm^{-1}): 2960, 2926, 2856, 1800, 1725, 1683, 1643, 1565, 1560, 1449, 1413, 1365, 1254, 1190, 1148, 1095, 1026, 931, 865, 800; HRMS (ESI) calcd for $\text{C}_{21}\text{H}_{26}\text{Br}_2\text{O}_5$ ($\text{M} + \text{Na}^+$) 539.0039, found 539.0027.

3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-7-methoxychroman-2-one (2a'):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 6.94 (d, 1H, J = 8.4 Hz), 6.64 (dd, 1H, J_1 = 9.0 Hz, J_2 = 3.0 Hz), 6.56 (d, 1H, J = 2.4 Hz), 3.78 (s, 3H), 2.89 (d, 1H, J = 3.0 Hz), 2.42 (s, 3H), 1.70–1.64 (m, 2H), 1.58–1.47 (m, 4H), 1.24–1.10 (m, 3H), 1.00 (s, 9H), 0.86–0.84 (m, 1H), 0.57–0.56 (m, 1H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 203.4, 161.2, 159.9, 151.9, 130.7, 112.9, 109.9, 101.9, 86.1, 82.3, 55.7, 48.3, 38.5, 32.3, 27.0, 26.8, 26.6, 26.3, 25.9, 25.8; IR (KBr, cm^{-1}): 2990, 2970, 2928, 2850, 1776, 1722, 1624, 1585, 1559, 1507, 1453, 1432, 1364, 1321, 1285, 1277, 1238, 1210, 1189, 1156, 1124, 1090, 1045, 1030, 922, 902, 886, 870, 859, 838, 806, 792, 763, 747, 712; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{30}\text{O}_6$ ($\text{M} + \text{Na}^+$) 413.1940, found 413.1944.

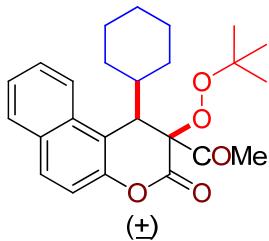
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-8-methoxychroman-2-one (15a'):



Semi-solid; ^1H NMR (CDCl_3 , 600 MHz): δ 7.00 (d, 1H, J = 7.8 Hz), 6.85 (d, 1H, J = 7.8 Hz), 6.61 (d, 1H, J = 7.2 Hz), 3.83 (s, 3H), 2.92 (d, 1H, J = 3.0 Hz), 2.42 (s, 3H), 1.71–1.47 (m, 6H), 1.23–1.08 (m, 3H), 0.96 (s, 9H), 0.86–0.80 (m, 1H), 0.63–0.58 (m,

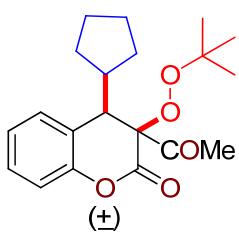
1H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 203.5, 160.6, 147.1, 140.4, 123.7, 122.2, 121.7, 111.4, 85.8, 82.2, 56.2, 48.9, 38.2, 32.4, 27.2, 27.0, 26.6, 26.0, 25.9, 25.8; IR (KBr, cm^{-1}): 2982, 2930, 2856, 1780, 1724, 1620, 1590, 1554, 1504, 1485, 1460, 1366, 1320, 1304, 1273, 1250, 1182, 1160, 1107, 1062, 1016, 971, 913, 866, 804, 785, 732; HRMS (ESI) calcd for $\text{C}_{22}\text{H}_{30}\text{O}_6$ ($\text{M} + \text{Na}^+$) 413.1940, found 413.1931.

2-Acetyl-2-(*tert*-butylperoxy)-1-cyclohexyl-1*H*-benzo[*f*]chromen-3(2*H*)-one (8a'):



Solid; m.p. 156–160 °C; ^1H NMR (CDCl_3 , 400 MHz): δ 7.90 (d, 1H, $J = 8.8$ Hz), 7.86 (d, 1H, $J = 8.0$ Hz), 7.78 (d, 1H, $J = 8.8$ Hz), 7.56 (d, 1H, $J = 7.2$ Hz), 7.47 (t, 1H, $J = 7.0$ Hz), 7.22 (t, 1H, $J = 9.2$ Hz), 3.77 (d, 1H, $J = 3.6$ Hz), 2.56 (s, 3H), 1.86–1.43 (m, 6H), 1.20–1.06 (m, 3H), 0.90 (s, 9H), 0.85–0.81 (m, 1H), 0.77–0.68 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 203.6, 161.3, 148.7, 132.2, 131.2, 129.4, 129.1, 127.1, 125.0, 123.8, 116.7, 116.1, 85.5, 82.3, 44.3, 39.1, 33.3, 28.5, 27.4, 26.8, 26.4, 26.1, 25.6; IR (KBr, cm^{-1}): 2942, 2927, 2850, 1780, 1745, 1720, 1628, 1602, 1558, 1516, 1507, 1463, 1438, 1395, 1367, 1264, 1220, 1186, 1162, 1115, 1080, 1063, 1045, 1028, 975, 864, 820, 789, 752; HRMS (ESI) calcd for $\text{C}_{25}\text{H}_{30}\text{O}_5$ ($\text{M} + \text{Na}^+$) 433.1991, found 433.1998.

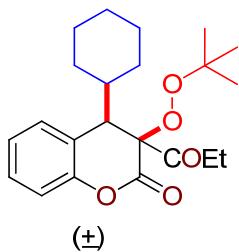
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclopentylchroman-2-one (1b'):



Solid; m.p. 160–162 °C; ^1H NMR (CDCl_3 , 400 MHz): δ 7.24–7.19 (m, 1H), 7.07–7.01 (m, 2H), 6.96 (d, 1H, $J = 8.0$ Hz), 3.20 (d, 1H, $J = 4.0$ Hz), 2.41 (s, 3H), 1.87–1.79 (m, 1H), 1.63–1.58 (m, 2H), 1.38–1.32 (m, 4H), 1.26–1.15 (m, 1H), 0.94 (s, 9H), 0.73–0.63 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 203.3, 161.2, 151.2, 130.3, 128.6, 123.9, 121.7, 116.2, 86.5, 82.3, 45.6, 39.7, 31.1, 27.1, 26.5, 26.2, 24.3, 23.5; IR (KBr, cm^{-1}): 2990, 2958, 2872, 1784, 1715, 1615, 1587, 1489, 1467, 1423, 1373, 1366, 1358, 1226, 1198,

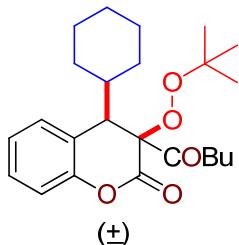
1169, 1136, 1116, 1093, 1056, 1017, 986, 920, 904, 774, 755, 720;
 HRMS (ESI) calcd for C₂₀H₂₆O₅ (M + Na⁺) 369.1678, found 369.1672.

3-(*tert*-Butylperoxy)-4-cyclohexyl-3-propionylchroman-2-one (9a'**):**



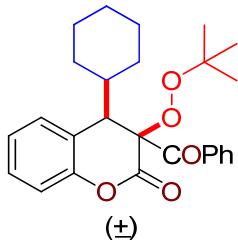
Solid; m.p. 162–164 °C; ¹H NMR (CDCl₃, 400 MHz): δ 7.21 (t, 1H, J = 8.0 Hz), 7.05–6.99 (m, 2H), 6.95 (d, 1H, J = 8.4 Hz), 2.95–2.90 (m, 1H), 2.88 (d, 1H, J = 2.4 Hz), 2.76–2.66 (m, 1H), 1.71–1.60 (m, 2H), 1.52–1.43 (m, 4H), 1.25–1.18 (m, 2H), 1.09 (t, 3H, J = 7.0 Hz), 0.92 (s, 9H), 0.87–0.73 (m, 2H), 0.56–0.46 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 205.9, 161.3, 151.1, 130.1, 128.5, 123.8, 121.2, 116.2, 86.2, 82.2, 49.2, 38.4, 32.2, 31.9, 26.9, 26.6, 26.2, 25.8, 25.7, 7.5; IR (KBr, cm^{−1}): 2974, 2930, 2857, 1786, 1716, 1614, 1588, 1490, 1458, 1405, 1390, 1376, 1370, 1364, 1352, 1325, 1260, 1225, 1179, 1155, 1119, 1082, 1066, 1020, 965, 940, 907, 898, 875, 842, 788, 757, 749, 703; HRMS (ESI) calcd for C₂₂H₃₀O₅ (M + Na⁺) 397.1991, found 397.1980.

3-(*tert*-Butylperoxy)-4-cyclohexyl-3-pentanoylchroman-2-one (10a'**):**



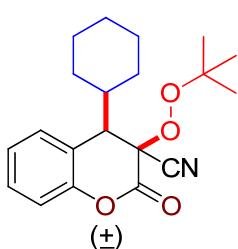
Solid; m.p. 132–136 °C; ¹H NMR (CDCl₃, 400 MHz): δ 7.21 (t, 1H, J = 7.4 Hz), 7.05–6.99 (m, 2H), 6.95 (d, 1H, J = 8.4 Hz), 2.88 (s, 1H), 2.84–2.81 (m, 1H), 2.71–2.62 (m, 1H), 1.71–1.60 (m, 4H), 1.51–1.40 (m, 4H), 1.16–1.04 (m, 4H), 0.92 (s, 9H), 0.91–0.86 (m, 3H), 0.82–0.73 (m, 2H), 0.54–0.45 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz): δ 205.3, 161.3, 151.1, 130.1, 128.5, 123.8, 121.1, 116.2, 86.0, 82.1, 49.1, 40.4, 38.3, 32.2, 26.8, 26.6, 26.2, 25.8, 25.7, 16.7, 13.9; IR (KBr, cm^{−1}): 2980, 2927, 2850, 1787, 1721, 1615, 1586, 1490, 1464, 1366, 1260, 1227, 1182, 1158, 1148, 1112, 1084, 1021, 966, 948, 911, 898, 877, 790, 765, 704; HRMS (ESI) calcd for C₂₄H₃₄O₅ (M + Na⁺) 425.2304, found 425.2310.

3-Benzoyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (11a'):



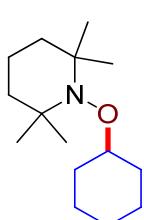
Solid; m.p. 158–160 °C; ^1H NMR (CDCl_3 , 400 MHz): δ 8.13 (d, 2H, $J = 7.2$ Hz), 7.49 (t, 1H, $J = 7.0$ Hz), 7.37 (t, 2H, $J = 7.4$ Hz), 7.23–7.21 (m, 1H), 7.07–7.05 (m, 2H), 7.80 (d, 1H, $J = 8.4$ Hz), 3.27 (s, 1H), 1.82–1.78 (m, 1H), 1.63–1.50 (m, 4H), 1.45–1.41 (m, 2H), 1.18–1.10 (m, 3H), 0.81 (s, 9H), 0.60–0.53 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 194.2, 160.1, 151.2, 136.4, 132.9, 130.2, 129.5, 128.6, 128.2, 123.8, 120.9, 116.2, 87.8, 82.2, 50.6, 38.5, 32.3, 26.7, 26.5, 26.1, 25.9, 25.8; IR (KBr, cm^{-1}): 2970, 2928, 2854, 1777, 1734, 1692, 1612, 1600, 1490, 1458, 1448, 1375, 1276, 1245, 1226, 1198, 1170, 1133, 1116, 1066, 1019, 952, 923, 880, 812, 770, 752; HRMS (ESI) calcd for $\text{C}_{26}\text{H}_{30}\text{O}_5$ ($M + \text{Na}^+$) 445.1991, found 445.1984.

3-(*tert*-Butylperoxy)-4-cyclohexyl-2-oxochroman-3-carbonitrile (12a'):



Solid; m.p. 154–156 °C; ^1H NMR (CDCl_3 , 400 MHz): δ 7.32 (t, 1H, $J = 8.0$ Hz), 7.16 (t, 1H, $J = 7.4$ Hz), 7.10–7.04 (m, 2H), 3.22 (d, 1H, $J = 3.2$ Hz), 2.15–2.08 (m, 1H), 1.77–1.73 (m, 2H), 1.63–1.53 (m, 3H), 1.35–1.18 (m, 3H), 1.06 (s, 9H), 0.95–0.85 (m, 1H), 0.70–0.60 (m, 1H); ^{13}C NMR (CDCl_3 , 100 MHz): δ 159.5, 150.5, 130.2, 129.3, 124.8, 119.1, 116.5, 114.5, 83.3, 80.7, 50.1, 39.6, 32.1, 27.1, 26.4, 26.1, 25.8, 25.6; IR (KBr, cm^{-1}): 2980, 2927, 2855, 2212, 1768, 1620, 1558, 1486, 1459, 1367, 1361, 1260, 1239, 1150, 1135, 1064, 1050, 1030, 1008, 910, 896, 856, 798, 767, 751, 704; HRMS (ESI) calcd for $\text{C}_{20}\text{H}_{25}\text{NO}_4$ ($M + \text{Na}^+$) 366.1681, found 366.1678.

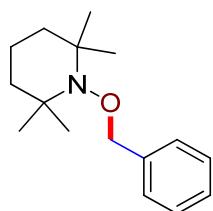
1-(Cyclohexyloxy)-2,2,6,6-tetramethylpiperidine (1A):



Gummy; ^1H NMR (CDCl_3 , 400 MHz): δ 3.58 (bs, 1H), 2.04 (bs, 2H), 1.74 (bs, 2H), 1.54–1.45 (m, 7H), 1.31–1.19 (m, 5H), 1.14 (s,

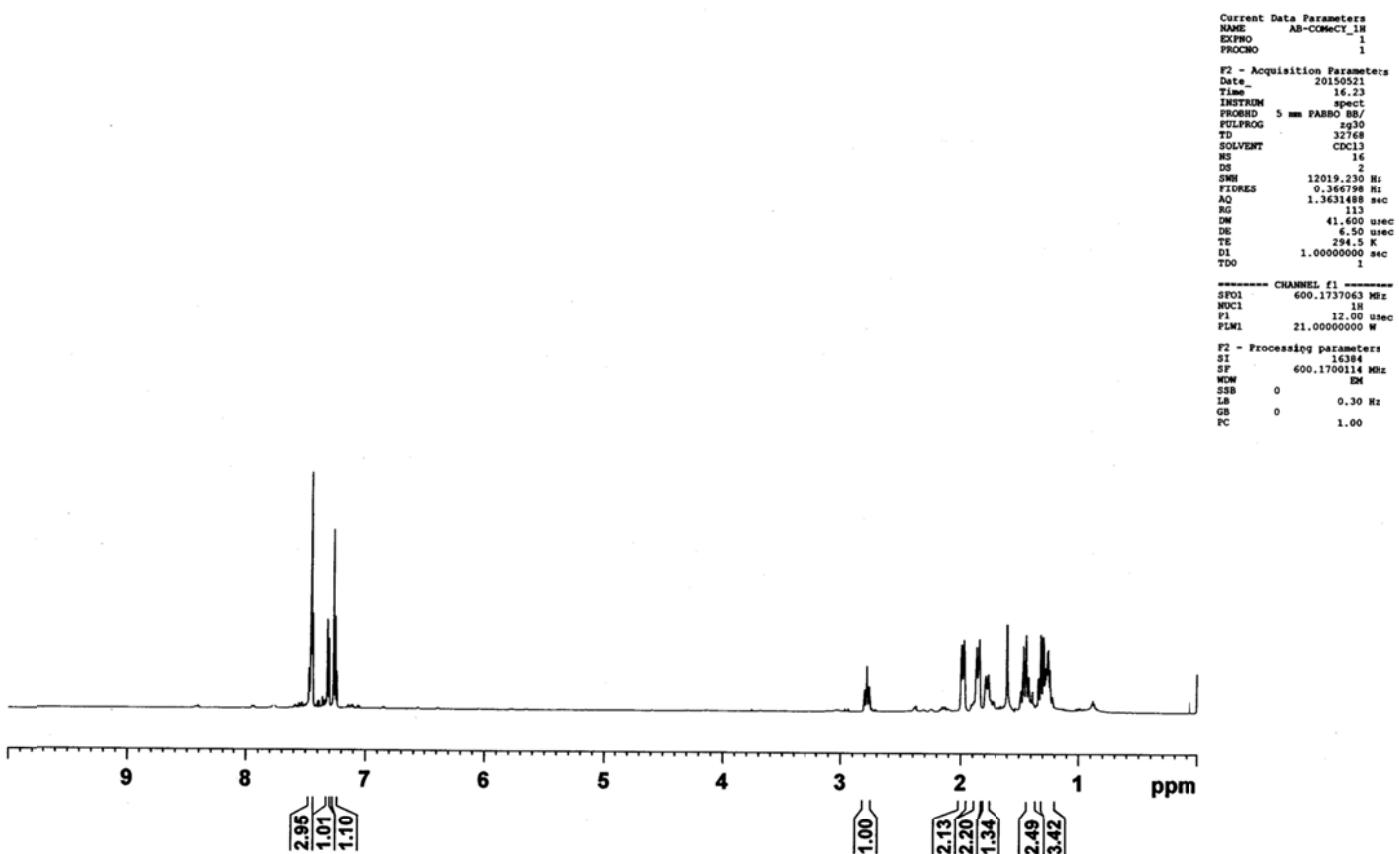
6H), 1.11 (s, 6H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 81.9, 59.8, 40.5, 34.7, 33.1, 26.2, 25.3, 17.6; IR (KBr, cm^{-1}): 2972, 2931, 2855, 1467, 1452, 1374, 1359, 1257, 1242, 1208, 1132, 1058, 1044, 1021, 966, 913, 785, 710; HRMS (ESI) calcd for $\text{C}_{15}\text{H}_{29}\text{NO}$ ($\text{M} + \text{H}^+$) 240.2329, found 240.2335.

1-(BenzylOxy)-2,2,6,6-tetramethylpiperidine (1B):

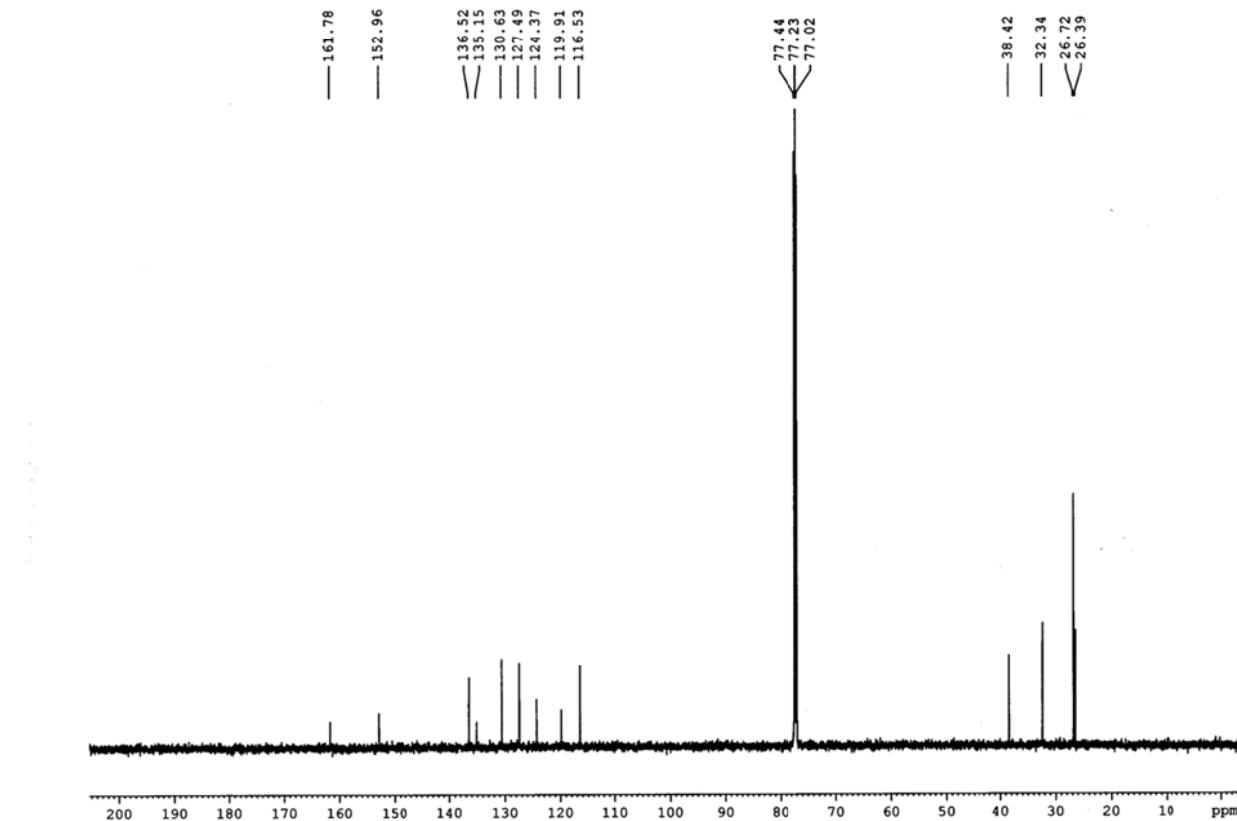


Gummy; ^1H NMR (CDCl_3 , 400 MHz): δ 7.38–7.25 (m, 5H), 4.82 (s, 2H), 1.58–1.48 (m, 5H), 1.37–1.33 (m, 1H), 1.26 (s, 6H), 1.15 (s, 6H); ^{13}C NMR (CDCl_3 , 150 MHz): δ 138.5, 128.4, 127.7, 127.5, 78.9, 60.2, 39.9, 33.3, 20.5, 17.3; IR (KBr, cm^{-1}): 2973, 2929, 2871, 1496, 1469, 1452, 1373, 1359, 1262, 1243, 1207, 1183, 1133, 1081, 1045, 1028, 992, 955, 926, 732; HRMS (ESI) calcd for $\text{C}_{16}\text{H}_{25}\text{NO}$ ($\text{M} + \text{H}^+$) 248.2016, found 248.2011.

3-Cyclohexyl-2*H*-chromen-2-one (1a**): ^1H NMR (CDCl_3 , 600 MHz)**



3-Cyclohexyl-2H-chromen-2-one (1a): ^{13}C NMR (CDCl_3 , 150 MHz)



Current Data Parameters
NAME AB-COMeDTF- ^{13}C
EXPNO 1
PROCNO 1

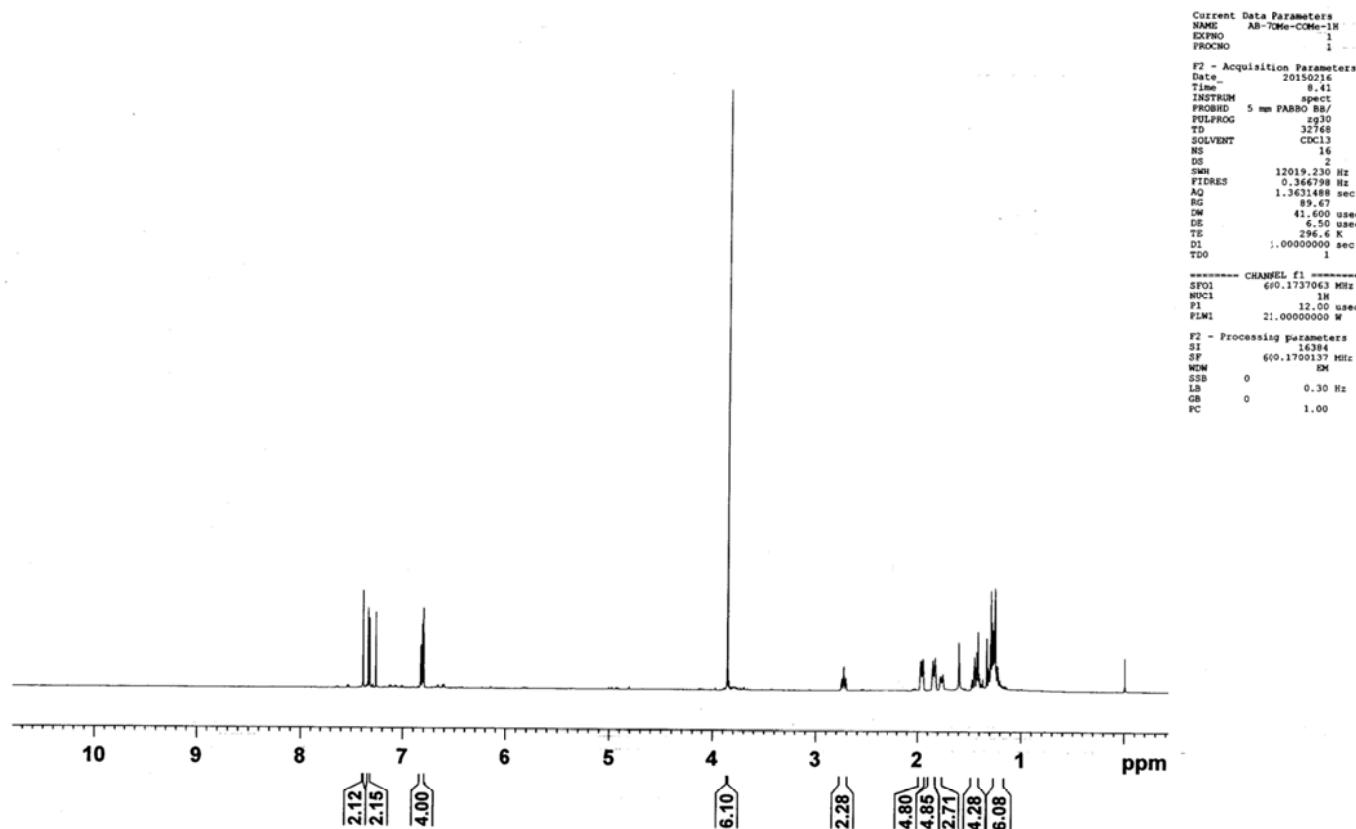
F2 - Acquisition Parameters
Date 20150302
Time 10:28
INSTRUM spect
PROBID 5 mm PABBO BB/
PULPROG zppg30
TD 32768
SOLVENT CDCl3
NS 201
DS 2
SW0 36057.62 Hz
FIDRES 1.10493 Hz
AQ 0.4543829 sec
RG 65.24
DW 13.867 usec
DE 6.50 usec
TE 298.4 K
D1 1.0000000 sec
D11 0.03000000 sec
TDO 1

CHANNEL f1
SF01 150.9279571 MHz
NUC1 ^{13}C
P1 10.50 usec
PLW1 95.00000000 W

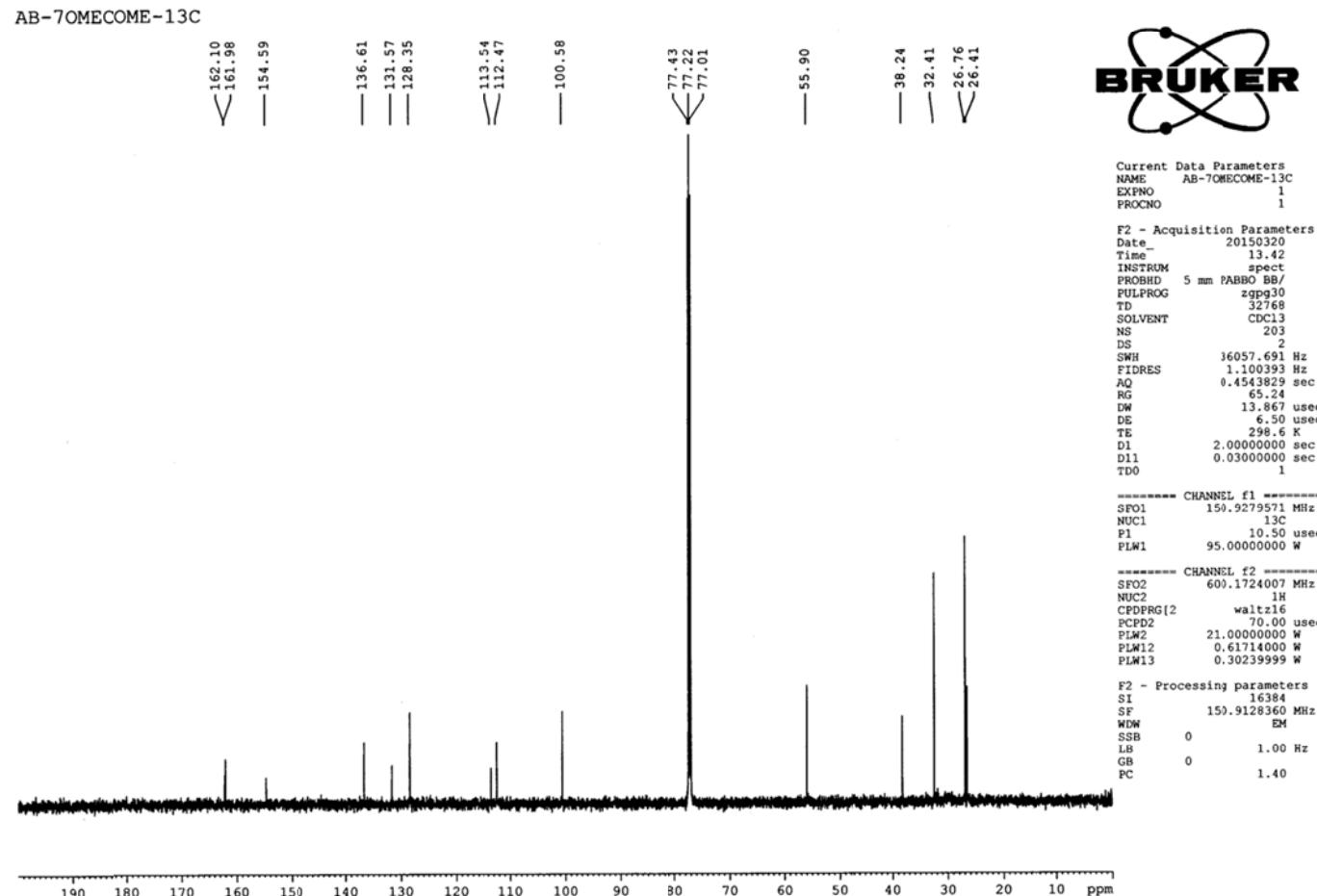
CHANNEL f2
SF02 60.1724007 MHz
NUC2 ^1H
CPDPG[2 waltz16
PCPD2 70.00 usec
PLW2 21.00000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

F2 - Processing parameters
SI 16384
SF 150.9128355 MHz
WDW EM
SSB 0 1.00 Hz
LB 0 1.40
GB 0
PC

3-Cyclohexyl-7-methoxy-2H-chromen-2-one (2a): ^1H NMR (CDCl_3 , 600 MHz)

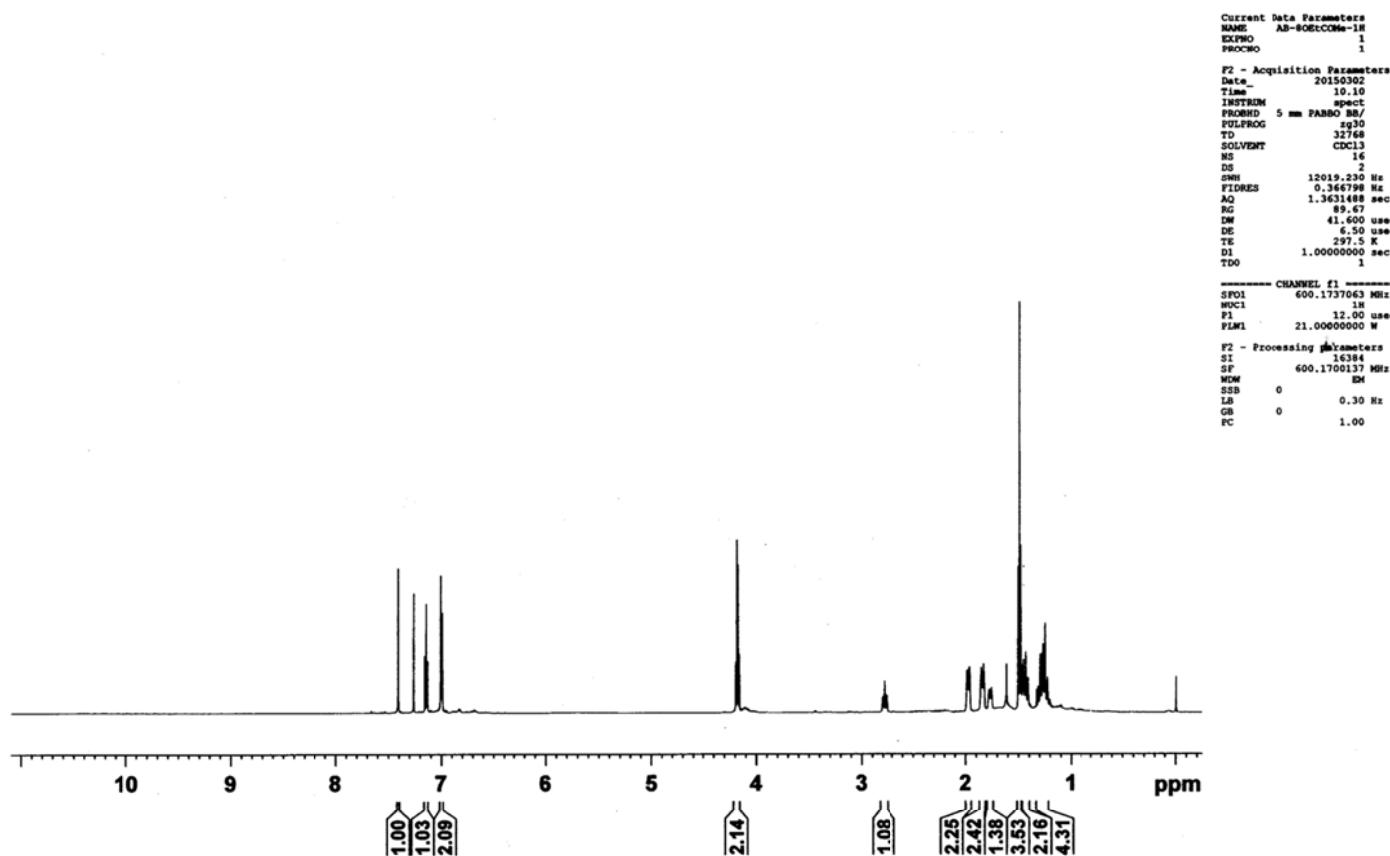


3-Cyclohexyl-7-methoxy-2H-chromen-2-one (2a): ^{13}C NMR (CDCl_3 , 150 MHz)

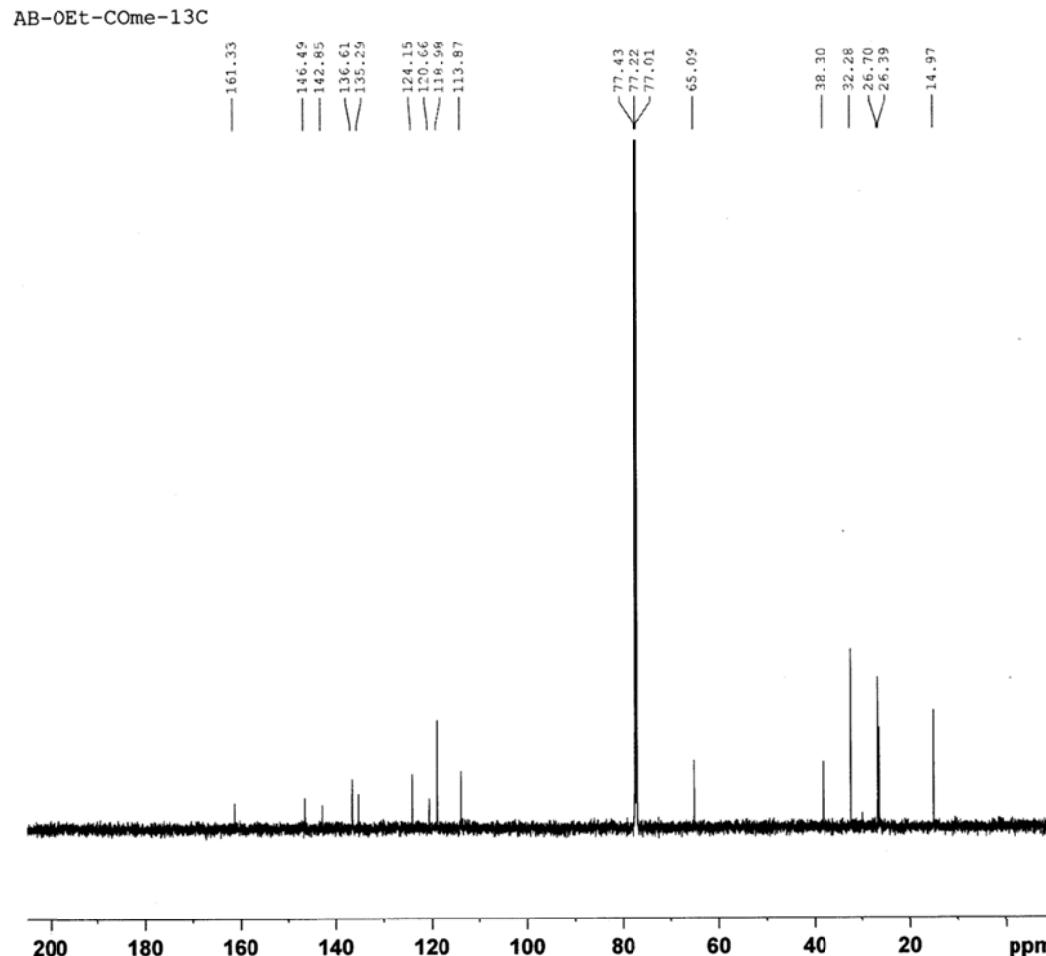


3-Cyclohexyl-8-ethoxy-2H-chromen-2-one (3a): ^1H NMR (CDCl_3 , 600 MHz)

AB-8OEtCOMe-1H



3-Cyclohexyl-8-ethoxy-2H-chromen-2-one (3a): ^{13}C NMR (CDCl_3 , 150 MHz)



Current Data Parameters
 NAME AB-OEt-COme- ^{13}C
 EXPNO 1
 PROCNO 1

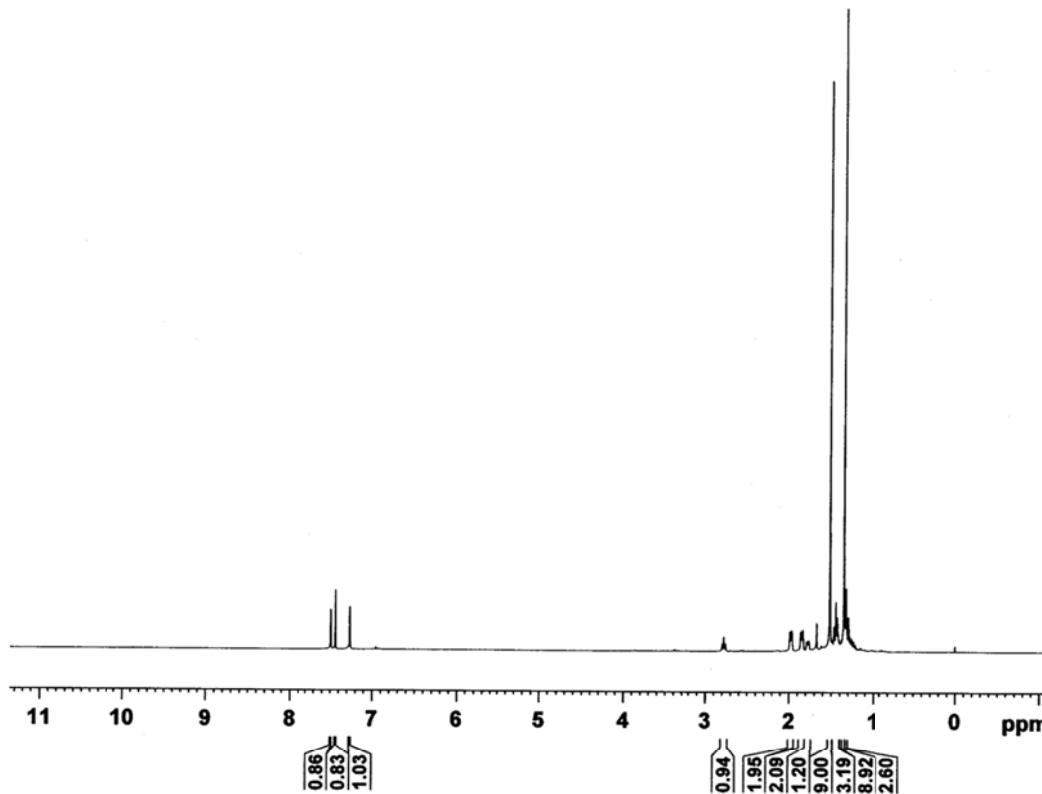
F2 - Acquisition Parameters
 Date 20150305
 Time 12.26
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 201
 DS 2
 SWH 36057.691 Hz
 FIDRES 1.100393 Hz
 AQ 0.4543829 sec
 RG 200.18
 DW 13.867 usec
 DE 6.50 usec
 TE 298.0 K
 D1 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

===== CHANNEL f1 ======
 SFO1 150.9279571 MHz
 NUC1 ^{13}C
 P1 10.50 usec
 PLW1 95.0000000 W

===== CHANNEL f2 ======
 SFO2 600.1724007 MHz
 NUC2 ^1H
 CPDPRG[2] waltz16
 PCPD2 70.00 usec
 PLW2 21.00000000 W
 PLW12 0.61714000 W
 PLW13 0.30239999 W

F2 - Processing parameters
 SI 16384
 SF 150.9128372 MHz
 WDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

6,8-Di-*tert*-butyl-3-cyclohexyl-2*H*-chromen-2-one (4a): ^1H NMR (CDCl_3 , 600 MHz)



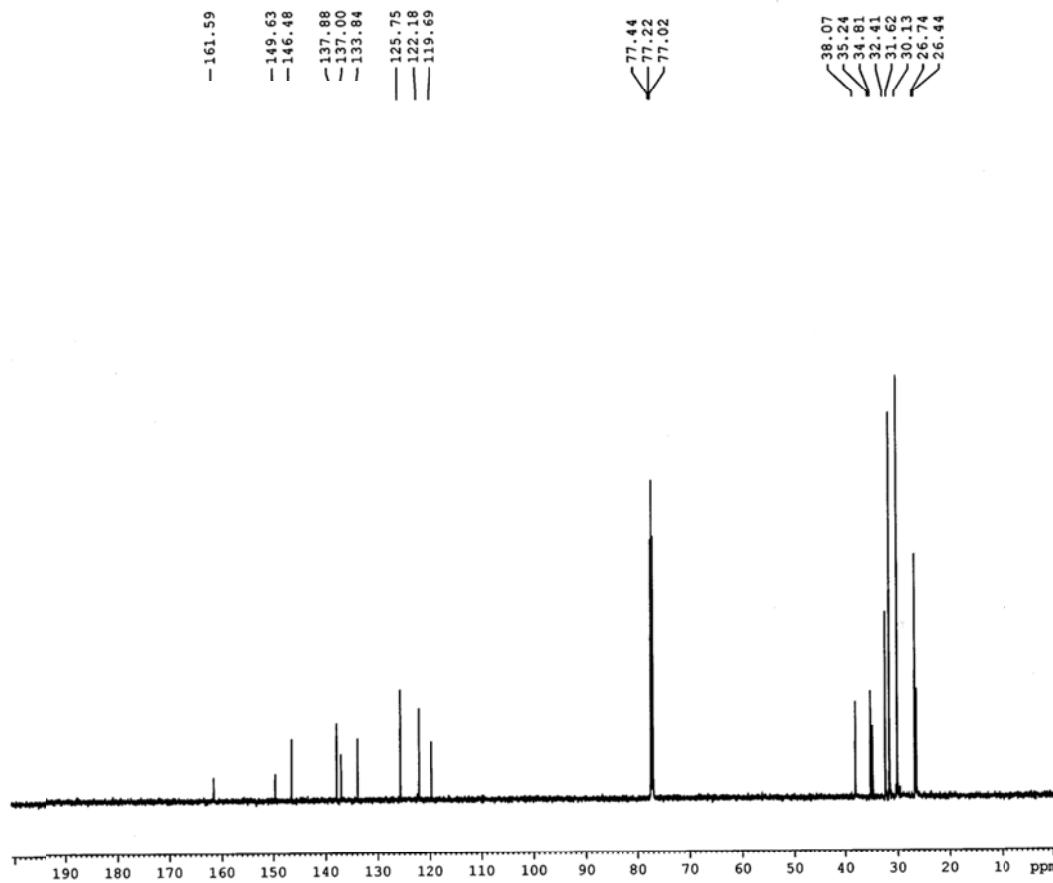
Current Data Parameters
NAME SS-6,8-tBu-DTBP-1H
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20150318
Time 10.33
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 2
SWH 12019.230 Hz
FIDRES 0.183399 Hz
AQ 2.7262976 sec
RG 31.5
DW 41.600 usec
DE 6.50 usec
TE 297.7 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 600.1737063 MHz
NUC1 1H
P1 12.00 usec
PLW1 21.0000000 W

F2 - Processing parameters
SI 65536
SF 600.1700142 MHz
WDW EM
SSB 0 0.30 Hz
LB 0
GB 0
PC 1.00

6,8-Di-*tert*-butyl-3-cyclohexyl-2*H*-chromen-2-one (4a): ^{13}C NMR (CDCl_3 , 150 MHz)



Current Data Parameters
 NAME SS-6,8-tBu-DTBP-13C
 EXPNO 1
 PROCNO 1

F2 - Acquisition Parameters
 Date 20150318
 Time 10.25
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 120
 DS 2
 SWH 36057.691 Hz
 FIDRES 1.100393 Hz
 AQ 0.4543829 sec
 RG 65.24
 DW 13.867 usec
 DE 6.50 usec
 TE 298.0 K
 DI 2.0000000 sec
 D11 0.03000000 sec
 TDO 1

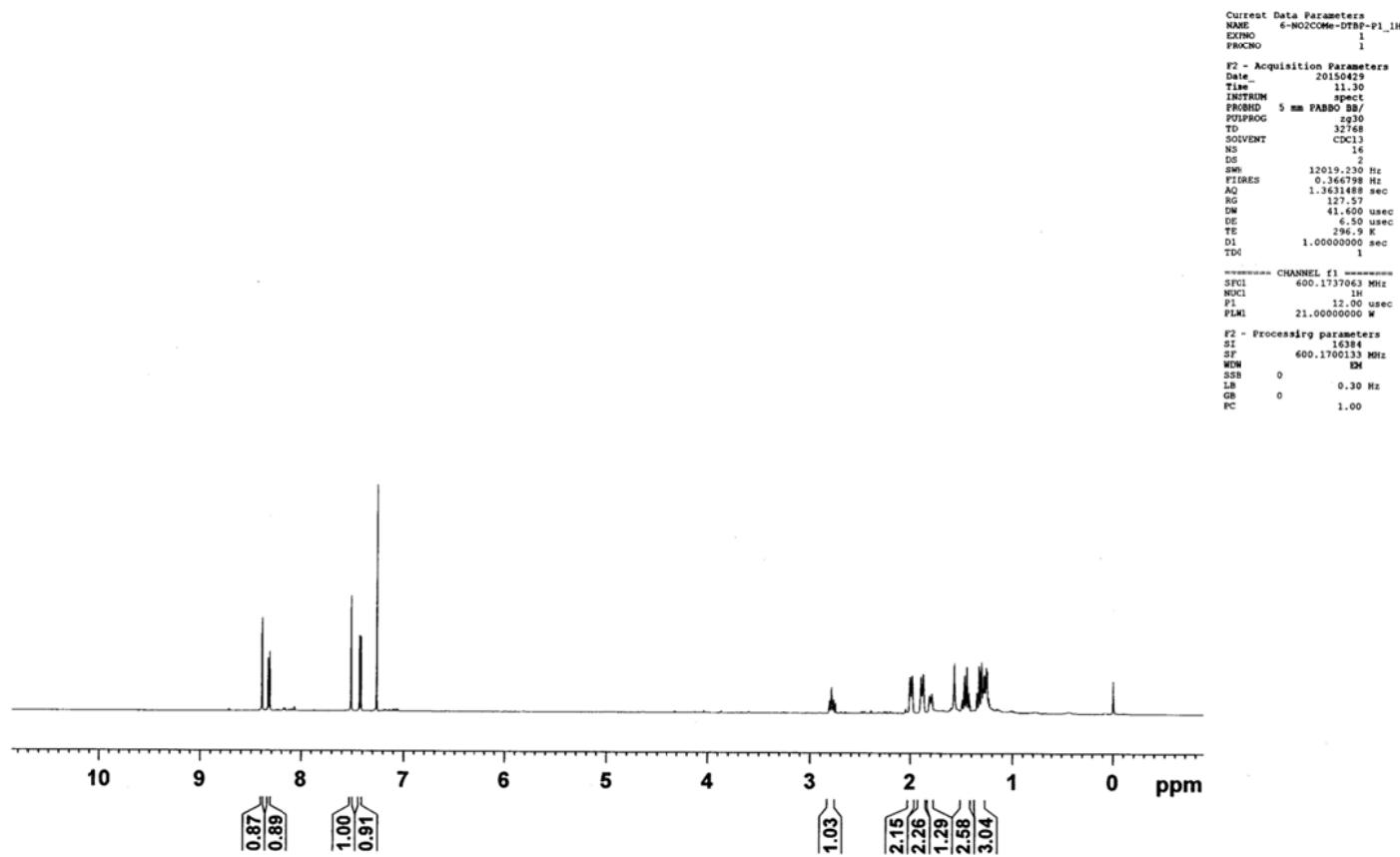
----- CHANNEL f1 -----
 SFO1 150.9279571 MHz
 NUC1 ^{13}C
 P1 10.50 usec
 PLW1 95.0000000 W

----- CHANNEL f2 -----
 SFO2 600.1724007 MHz
 NUC2 ^1H
 CPDPRG[2 waltz16
 PCPD2 70.00 usec
 PLW2 21.00000000 W
 PLW12 0.61714000 W
 PLW13 0.30239999 W

F2 - Processing parameters
 SI 16384
 SF 150.9128381 MHz
 WDW EM
 SSB 0 1.00 Hz
 LB 0
 GB 0 1.40
 PC

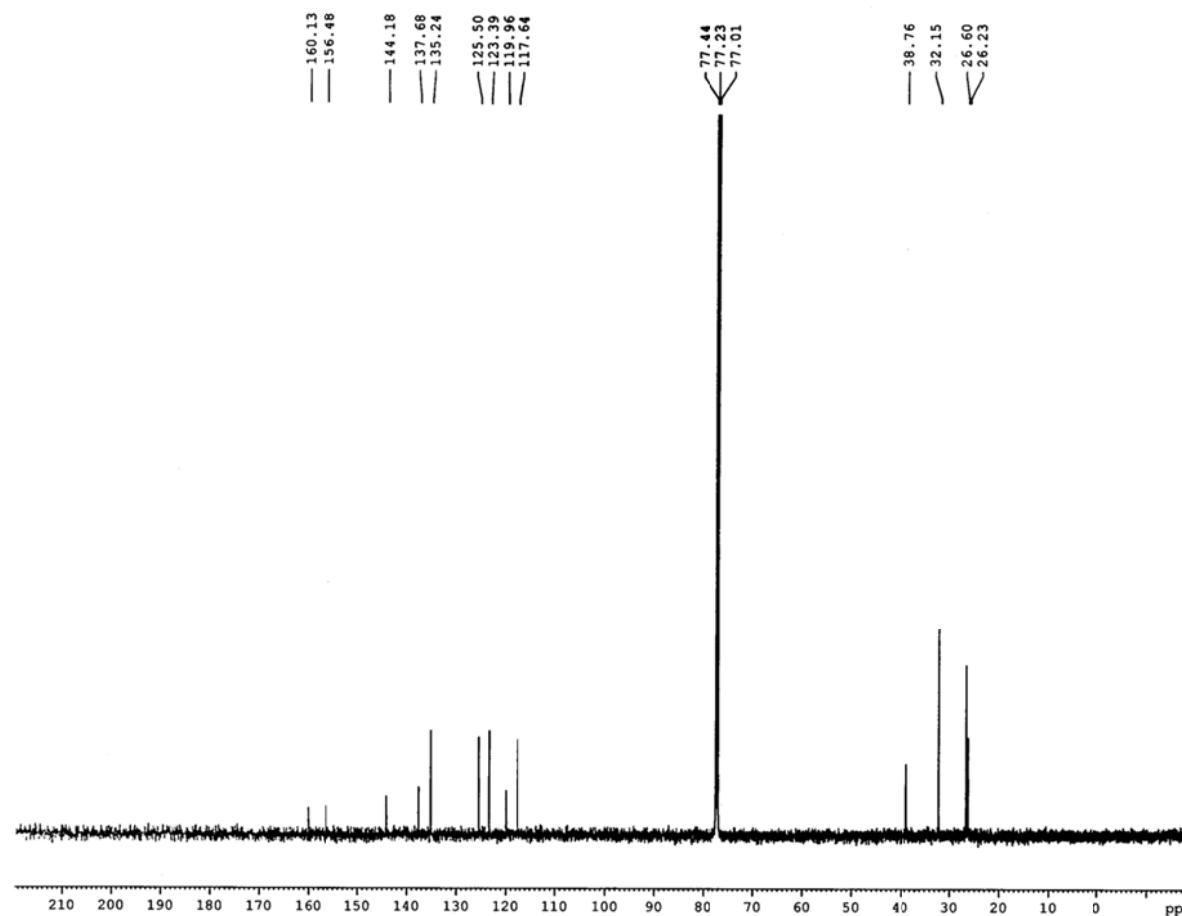
3-Cyclohexyl-6-nitro-2H-chromen-2-one (5a): ^1H NMR (CDCl_3 , 600 MHz)

6-NO₂COMe-DTBP- 1H



3-Cyclohexyl-6-nitro-2H-chromen-2-one (5a): ^{13}C NMR (CDCl_3 , 150 MHz)

AB-6NO2-BTBP-13C



Current Data Parameters
NAME AB-6NO2-RTRP-13C
EXPNO 1
PROCNO 1

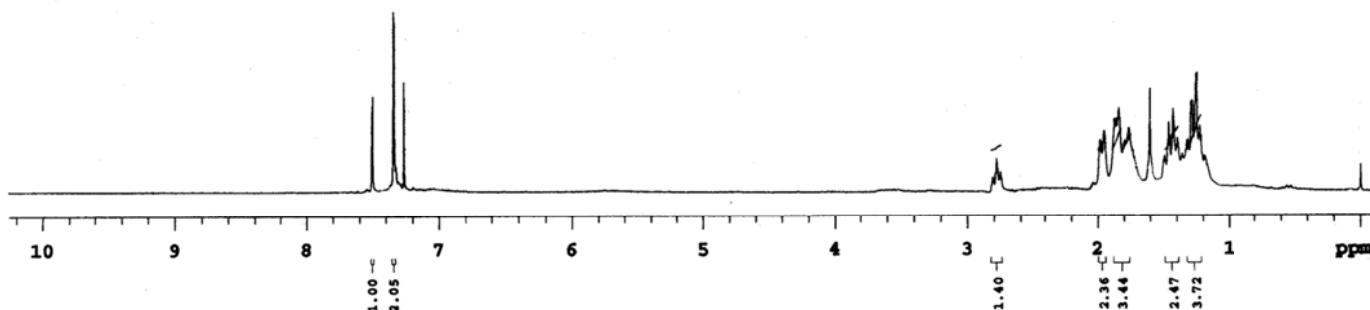
F2 - Acquisition Parameters
Date 20150430
Time 12:34
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpg30
TD 32768
SOLVENT CDCl3
NS 733
DS 2
SWH 36057.691 Hz
FIDRES 1.100393 Hz
AQ 0.4543829 sec
RG 200.18
DW 13.867 usec
DE 6.50 usec
TE 29.6 K
D1 2.0000000 sec
D11 0.0300000 sec
TDO 1

----- CHANNEL f1 -----
SF01 150.9279571 MHz
NUC1 ^{13}C
P1 10.50 usec
PLW1 95.00000000 W

----- CHANNEL f2 -----
SF02 600.1724007 MHz
NUC2 ^1H
CPDPG12 waltz16
PCPD2 70.00 usec
PLW2 21.00000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

F2 - Processing parameters
SI 16384
SF 150.9128347 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

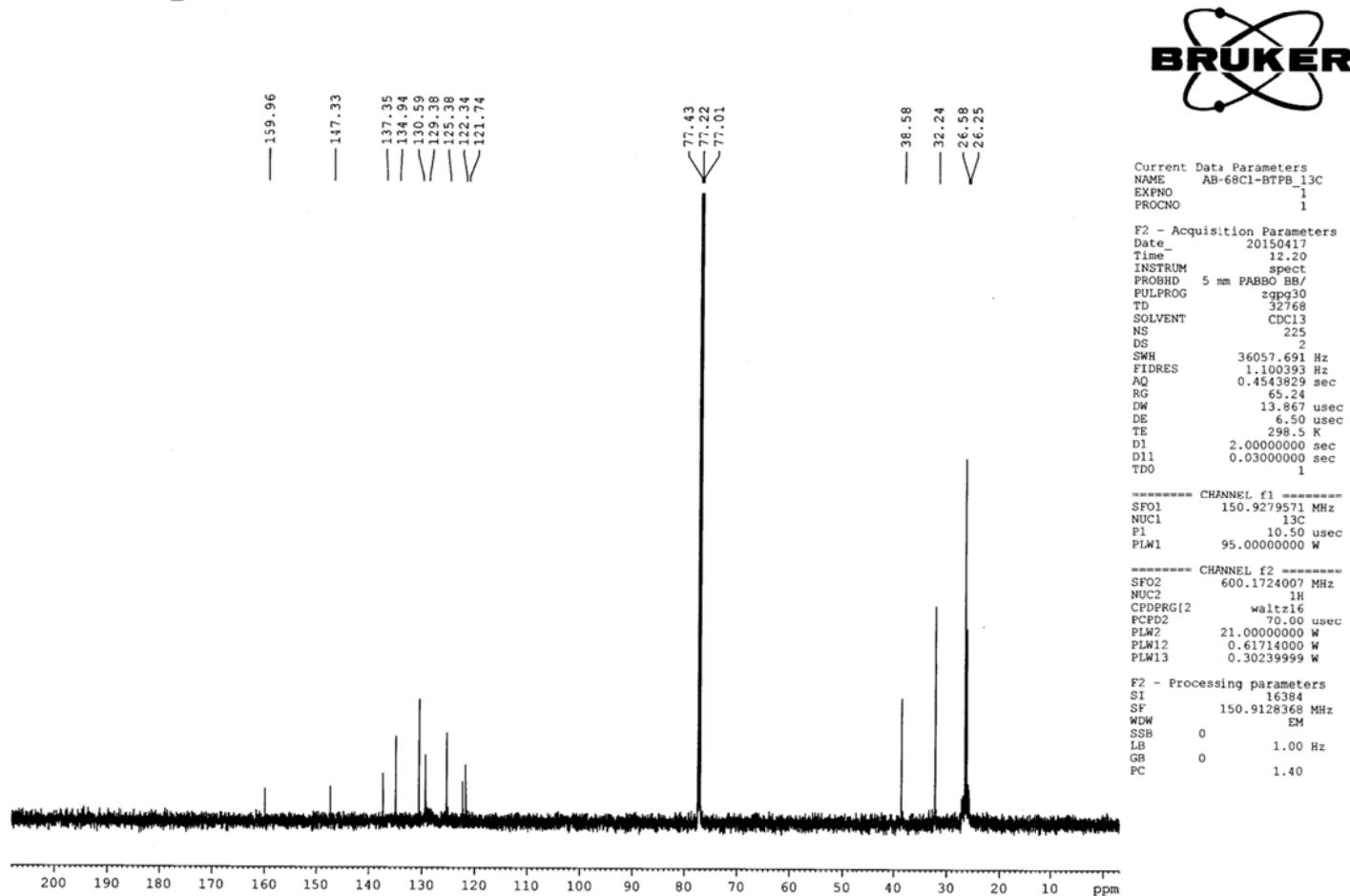
6,8-Dichloro-3-cyclohexyl-2H-chromen-2-one (6a): ^1H NMR (CDCl_3 , 600 MHz)



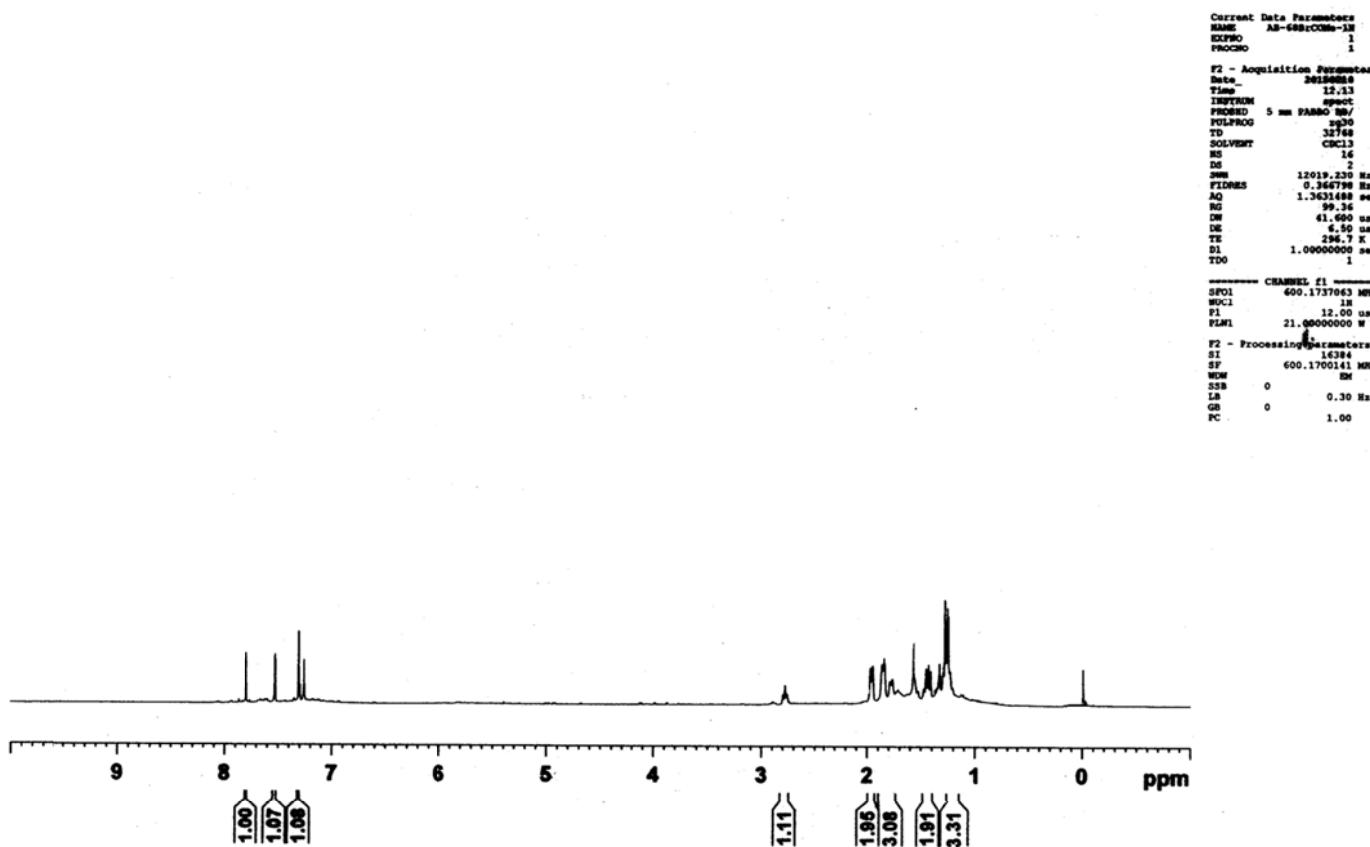
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 2.561 sec Width 6398.0 Hz 32 repetitions	OBSERVE H1, 399.8509609	DATA PROCESSING FT size 32768 Total time 1 minutes	AB-68C1C0Me-DTBP-P1-1H Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: chem Mercury-400 "IITG-NMR"
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6,8-Dichloro-3-cyclohexyl-2H-chromen-2-one (6a): ^{13}C NMR (CDCl_3 , 150 MHz)

AB-68Cl-BTPB_13C

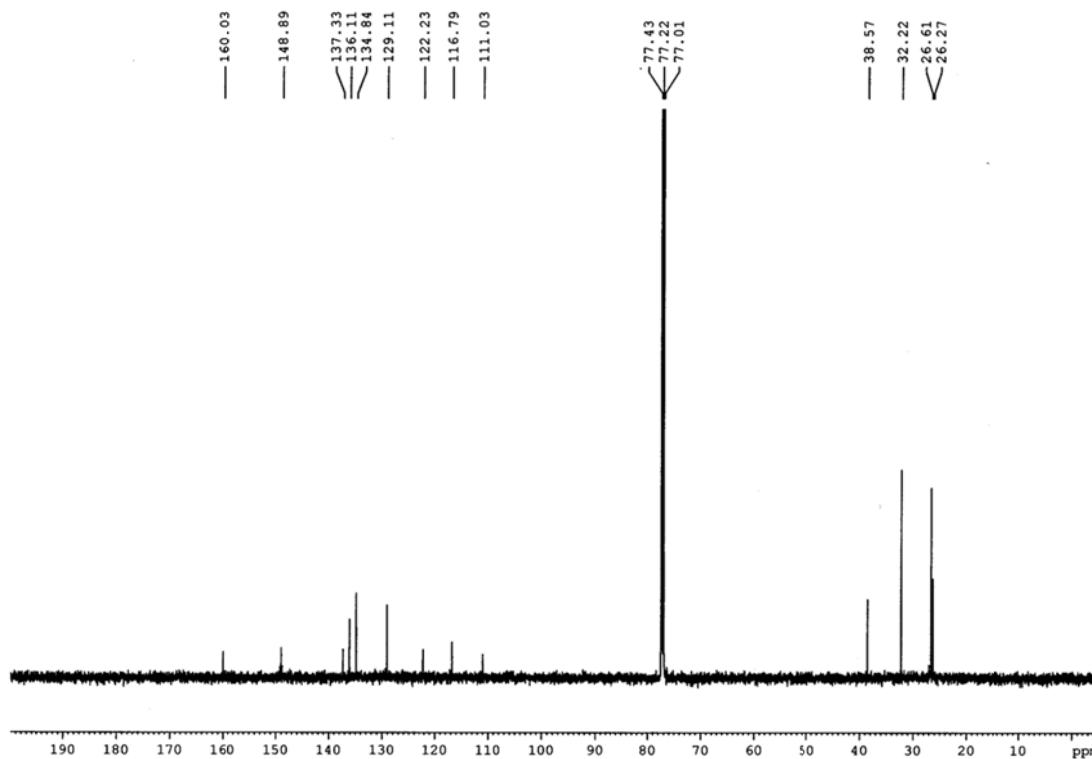


6,8-Bromo-3-cyclohexyl-2H-chromen-2-one (7a): ^1H NMR (CDCl_3 , 600 MHz)



6,8-Bromo-3-cyclohexyl-2H-chromen-2-one (7a):¹³C NMR (CDCl₃, 150 MHz)

AB-68BrDp-13C



Current Data Parameters
NAME AB-68BrDp-13C
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20150511
Time 13.32
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg3d30
TD 32768
SOLVENT CDCl3
NS 765
DS 2
SWH 36057.69 Hz
FIDRES 1.100393 Hz
AQ 0.4543829 sec
RG 200.18
DW 13.88 usec
DE 50.00 usec
TE 299.9 K
D1 2.00000000 sec
D11 0.03000000 sec
TDO 1

CHANNEL f1
SFO1 150.9279571 MHz
NUC1 13C
PI 10.50 usec
PLW1 95.00000000 W

CHANNEL f2
SFO2 600.1724007 MHz
NUC2 1H
CPDPRG12 waltz16
CPD2 70.00 usec
PLW2 21.00000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

F2 - Processing parameters
SI 16384
SF 150.9128332 MHz
WM EM
TDW 0
LB 1.00 Hz
GB 0
PC 1.40

2-Cyclohexyl-3*H*-benzo[*f*]chromen-3-one (8a): ^1H NMR (CDCl_3 , 600 MHz)

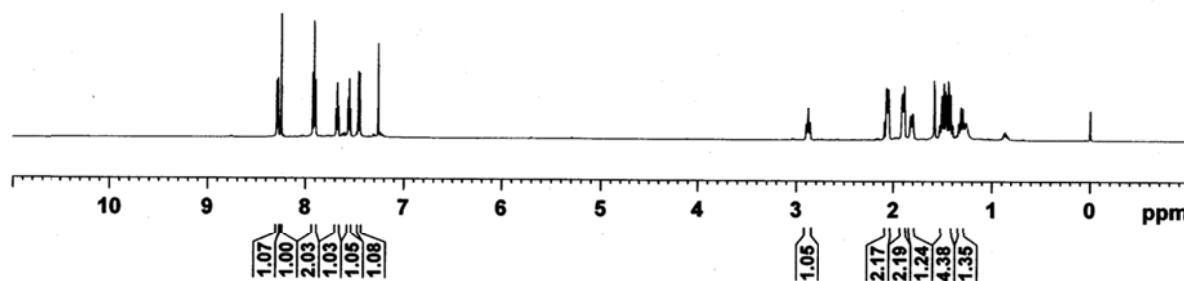
AB_NAPCOME-1H

Current Data Parameters
NAME AB_NAPCOME-1H
EXPNO 1
PROCNO 1

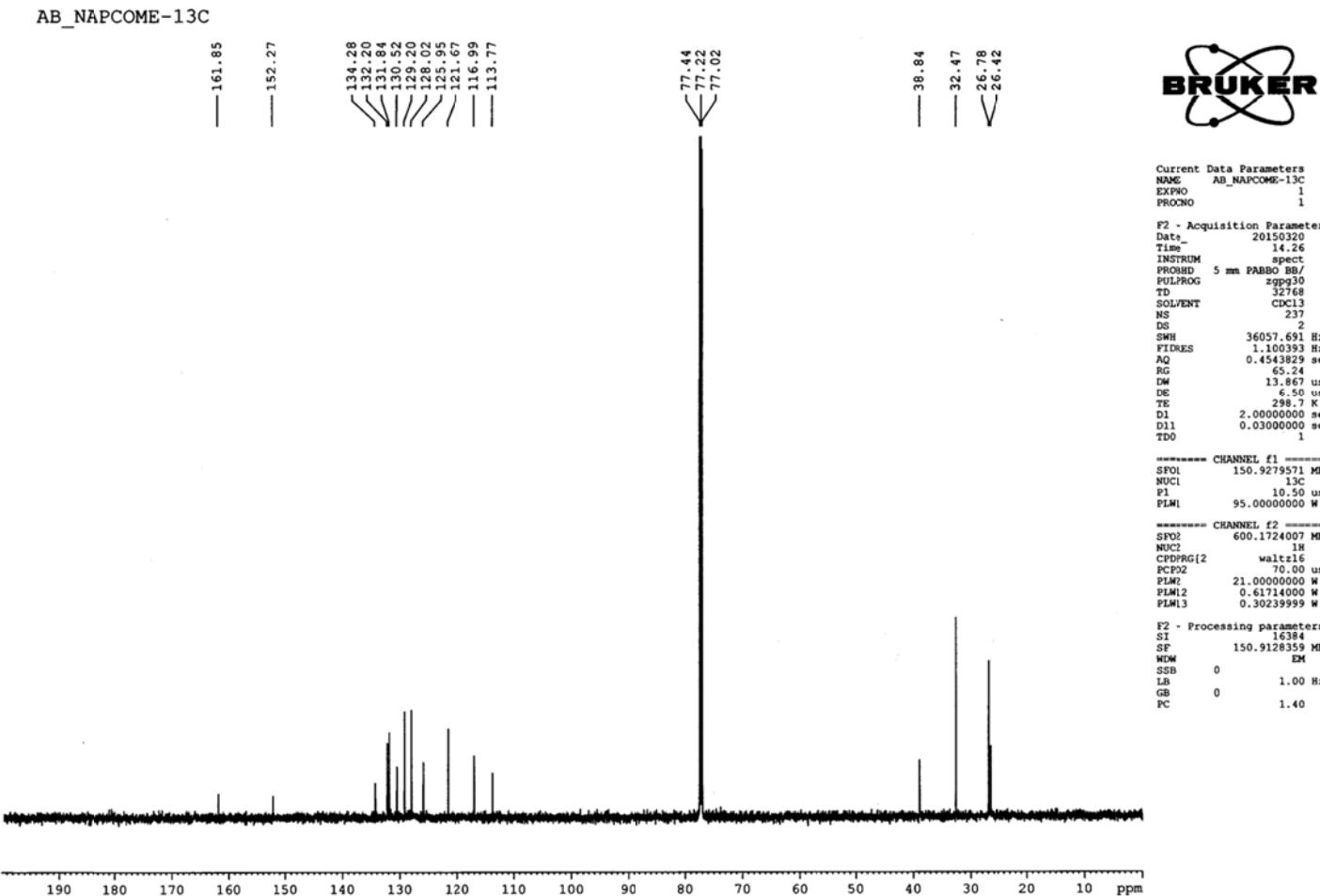
F2 - Acquisition Parameters
Date 20150320
Time 14:21
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zg30
TD 32768
SOLVENT CDCl3
NS 16
DS 2
SW0 12019.22 Hz
FIDRES 0.366798 Hz
AQ 1.3631488 sec
RG 89.67
DM 41.6 usec
DE 6.50 usec
TE 297.6 K
D1 1.0000000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 600.1737063 MHz
NUC1 13C
P1 12.00 usec
PLW1 21.00000000 W

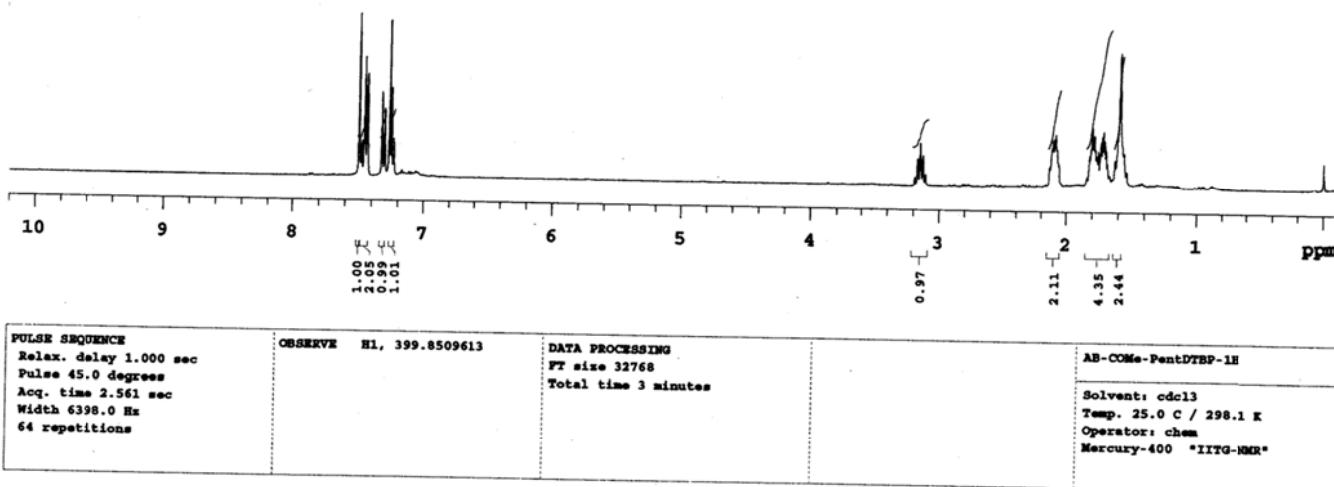
F2 - Processing parameters
SI 16384
SF 600.1700144 MHz
WDW EM
SSB 0
LB 0.30 Hz
GB 0
PC 1.00



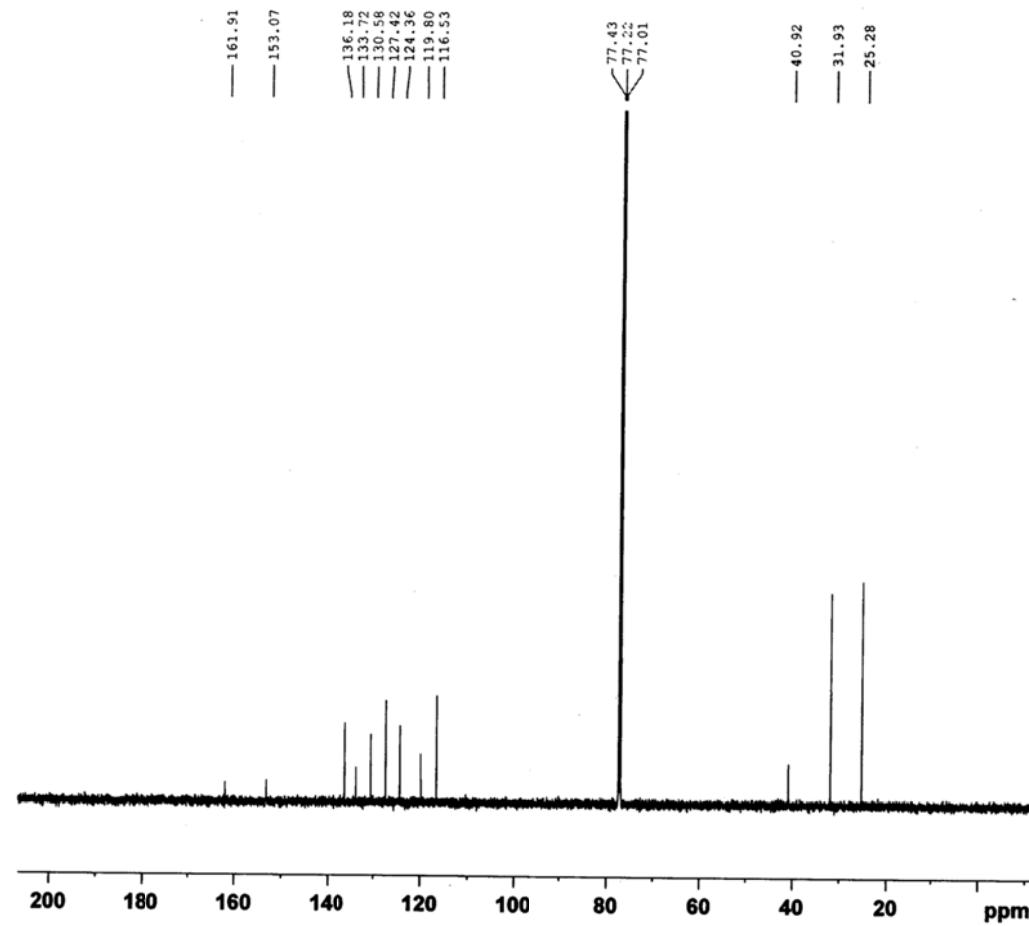
2-Cyclohexyl-3H-benzo[f]chromen-3-one (8a): ^{13}C NMR (CDCl_3 , 150 MHz)



3-Cyclopentyl-2H-chromen-2-one (1b): ^1H NMR (CDCl_3 , 600 MHz)



3-Cyclopentyl-2H-chromen-2-one (1b): ^{13}C NMR (CDCl_3 , 150 MHz)



Current Data Parameters
 NAME AB-COMe-pent-DTBP-13C
 EXPNO 1
 PROCNO 1

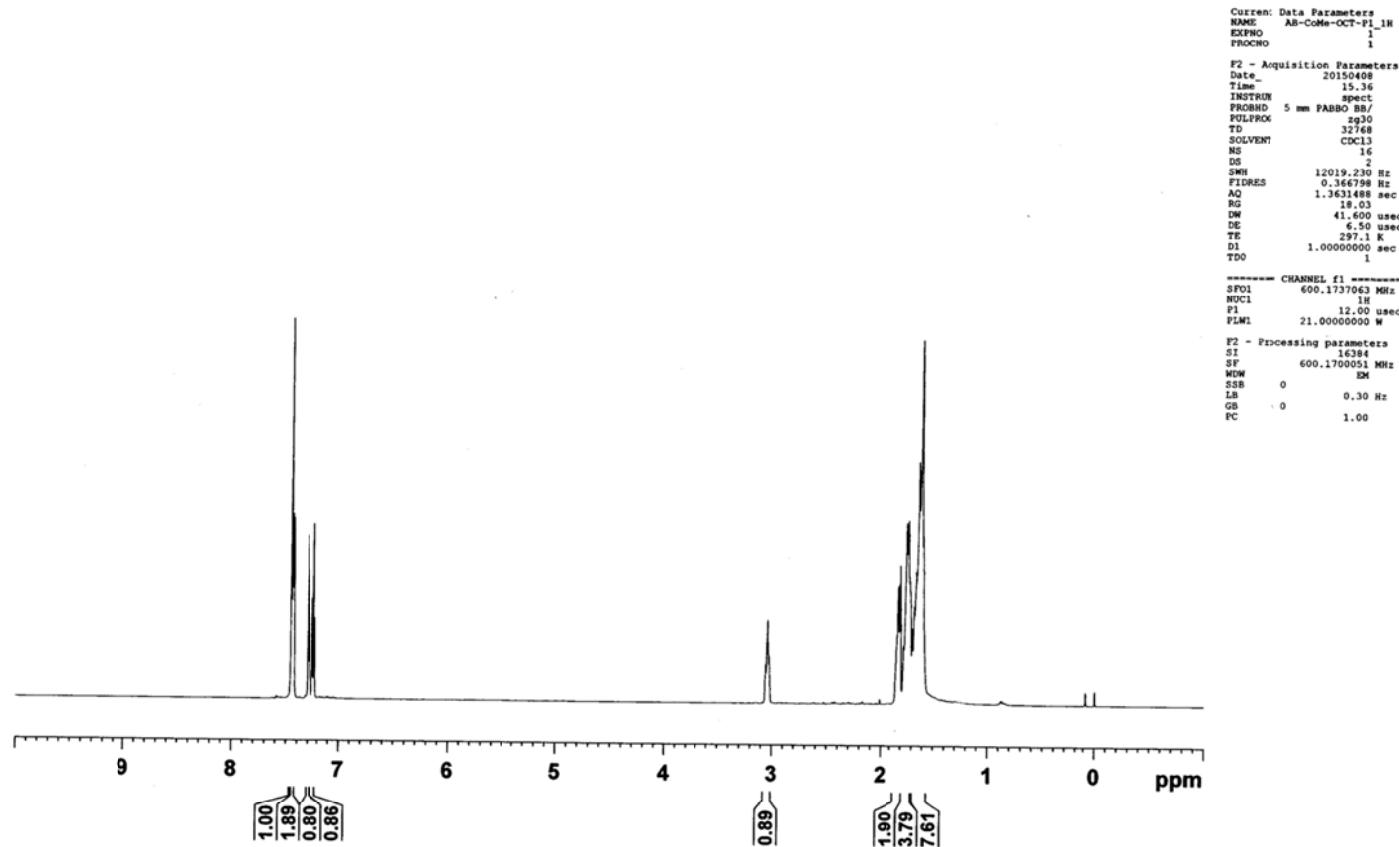
F2 - Acquisition Parameters
 Date 20150416
 Time 11.19
 INSTRUM spect
 PROBHD 5 mm PABBO B8/
 PULPROG zgpp30
 TD 32768
 SOLVENT CDCl3
 NS 214
 DS 2
 SWH 36057.691 Hz
 FIDRES 1.100393 Hz
 AQ 0.4543829 sec
 RG 200.10°
 DW 13.867 usec
 DE 6.50 usec
 TE 298.8 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 TDO 1

CHANNEL f1
 SFO1 150.9279571 MHz
 NUC1 ^{13}C
 P1 10.50 usec
 PLW1 95.0000000 W

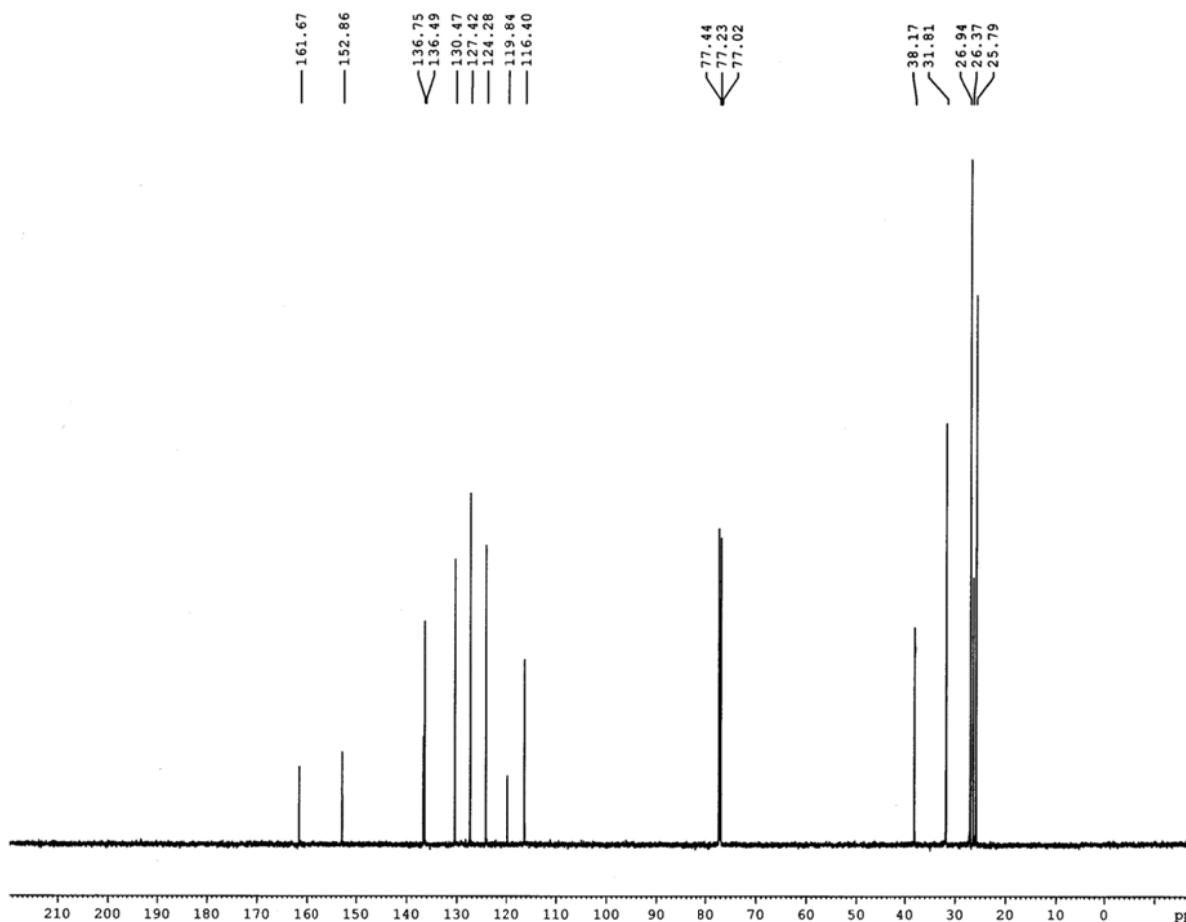
CHANNEL f2
 SFO2 600.1724007 MHz
 NUC2 ^1H
 CPDPRG[2] waltz16
 PCPD2 70.000 usec
 PLW2 21.00000000 W
 PLW12 0.61714000 W
 PLW13 0.30239999 W

F2 - Processing parameters
 SI 16384
 SF 150.9128358 MHz
 DW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

3-Cyclooctyl-2H-chromen-2-one (1c): ^1H NMR (CDCl_3 , 600 MHz)



3-Cyclooctyl-2H-chromen-2-one (1c): ^{13}C NMR (CDCl_3 , 150 MHz)



Current Data Parameters
 NAME AB-CoMe-OCT-P1_13C
 EXPNO 1
 PROCN0 1

F2 - Acquisition Parameters
 Date 20150408
 Time 15.29
 INSTRUM spect
 PROBHD 5 mm PABBO BB/
 PULPROG zgpg30
 TD 32768
 SOLVENT CDCl3
 NS 101
 DS 2
 SWH 36057.691 Hz
 FIDRES 1.100393 Hz
 AQ 0.4543829 sec
 RG 65.24
 DW 13.867 usec
 DE 6.50 usec
 TB 297.2 K
 D1 2.0000000 sec
 D11 0.0300000 sec
 T00 1

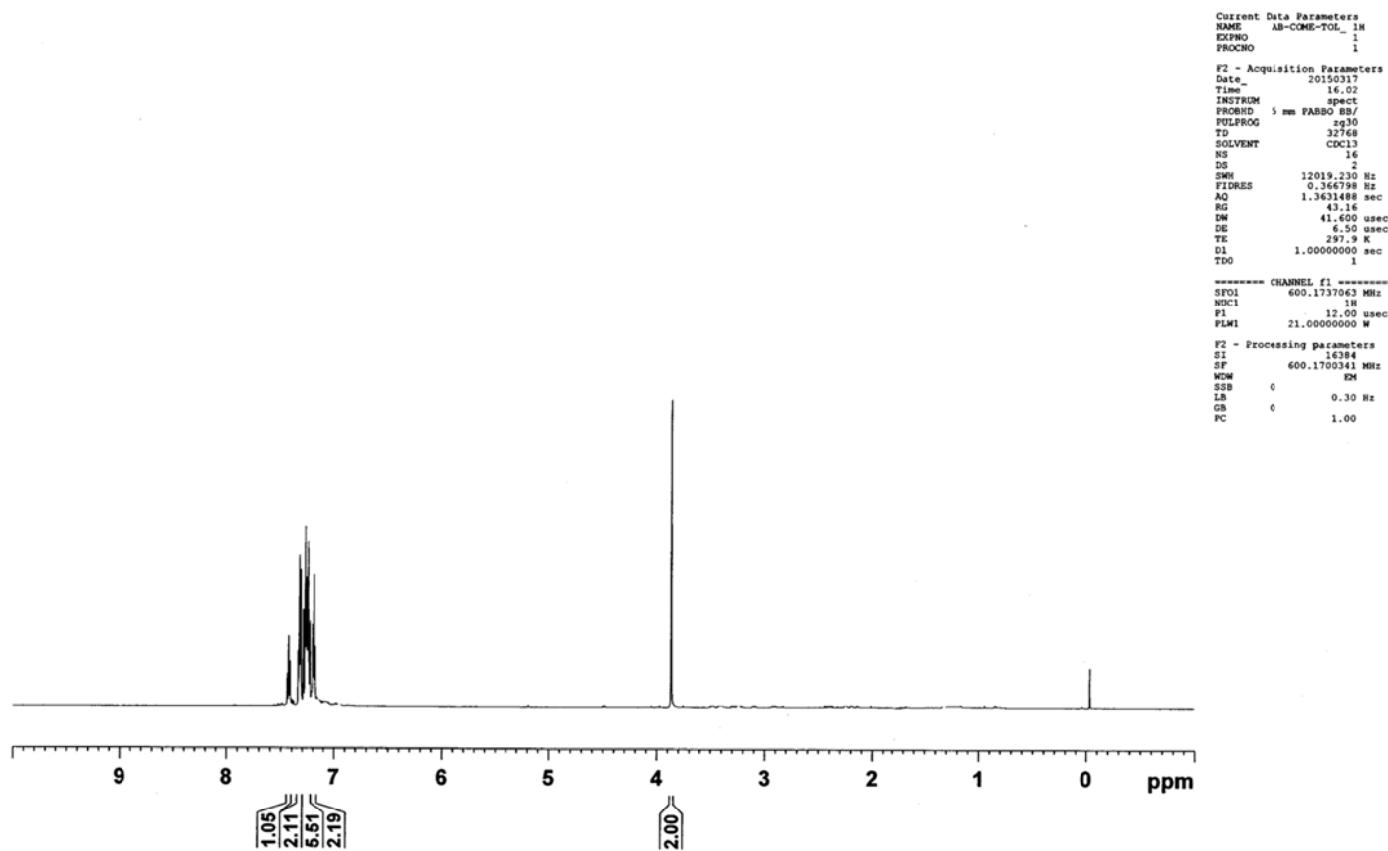
----- CHANNEL f1 -----
 SF01 150.9279571 MHz
 NUC1 ^{13}C
 P1 10.50 usec
 PLW1 95.00000000 W

----- CHANNEL f2 -----
 SF02 600.1724007 MHz
 NUC2 ^1H
 CPDPRG[2] waltz16
 PCPDR2 70.00 usec
 PLW2 21.00000000 W
 PLW12 0.61714000 W
 PLW13 0.30239999 W

F2 - Processing parameters
 SI 16384
 SF 150.9128447 MHz
 MDW EM
 SSB 0
 LB 1.00 Hz
 GB 0
 PC 1.40

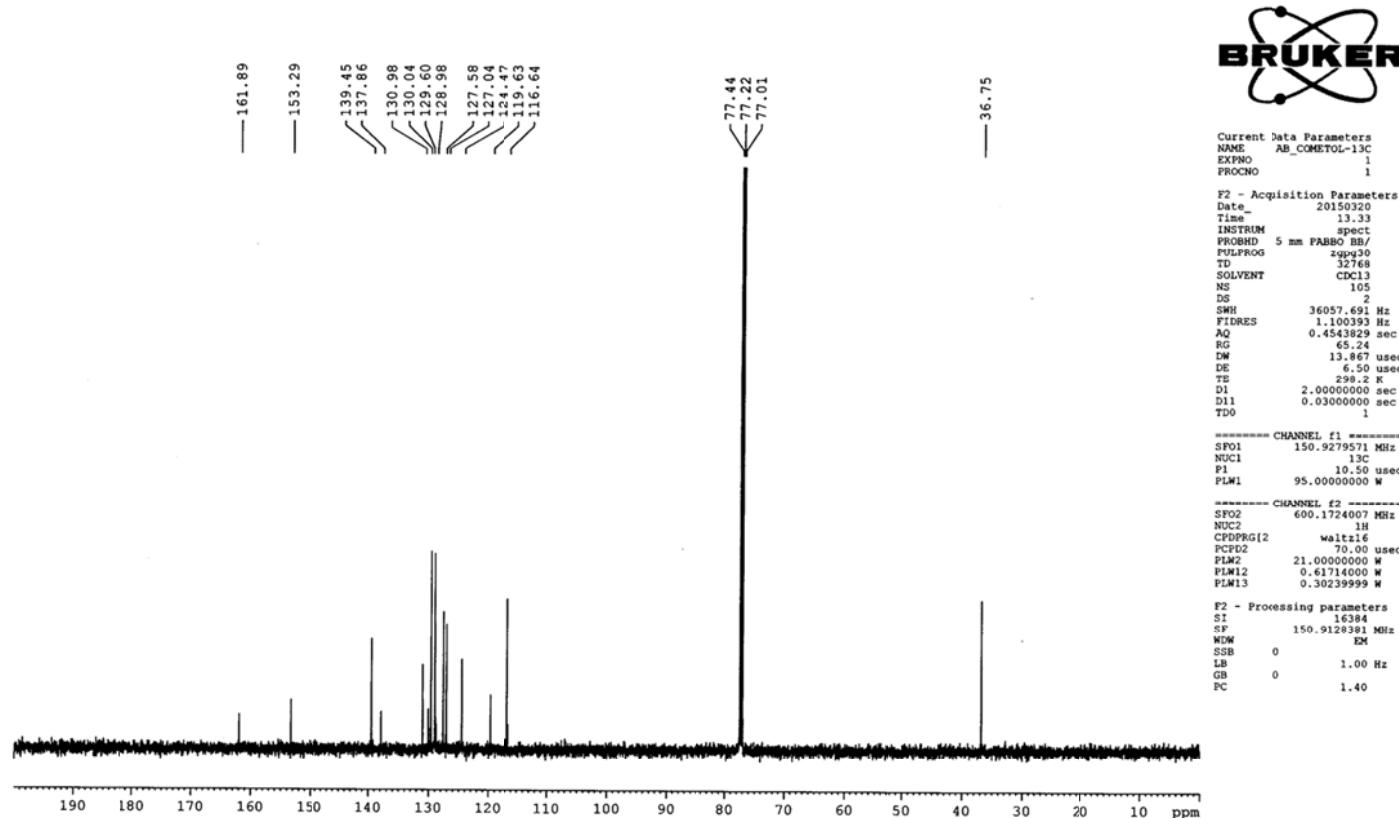
3-Benzyl-2H-chromen-2-one (1d): ^1H NMR (CDCl_3 , 600 MHz)

AB-COME-TOL_ 1H

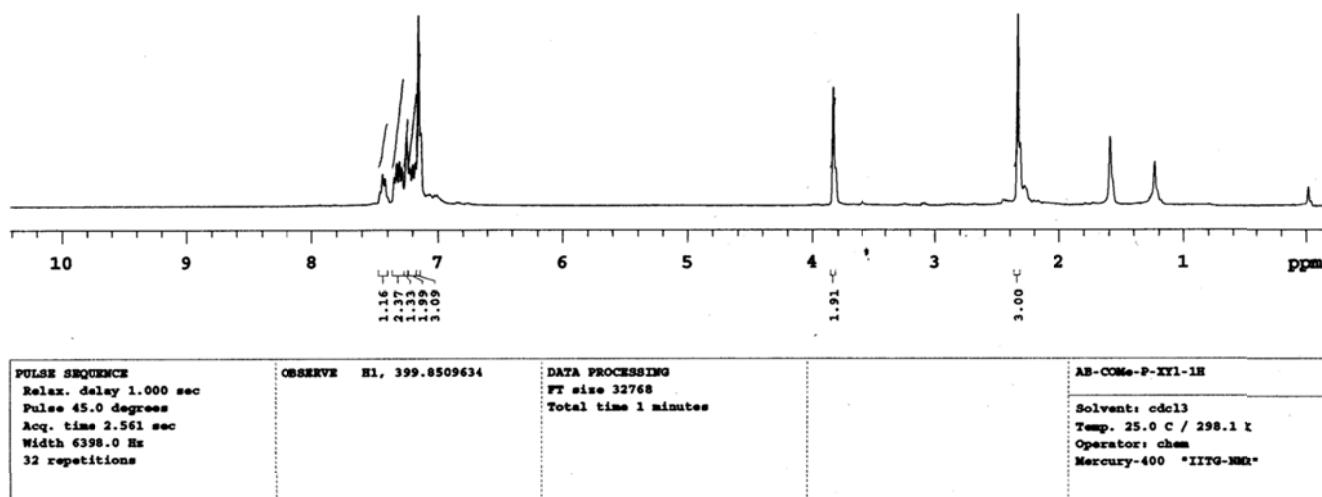


3-Benzyl-2H-chromen-2-one (1d): ^{13}C NMR (CDCl_3 , 150 MHz)

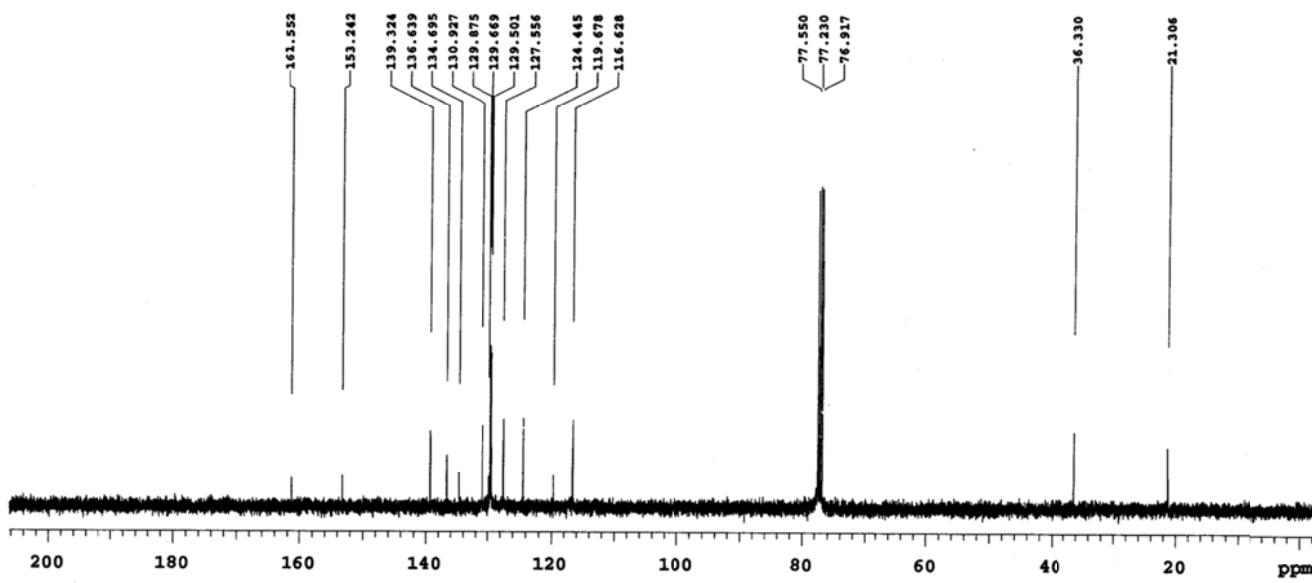
AB_COMETOL-13C



3-(4-Methylbenzyl)-2H-chromen-2-one (1e): ^1H NMR (CDCl_3 , 400 MHz)

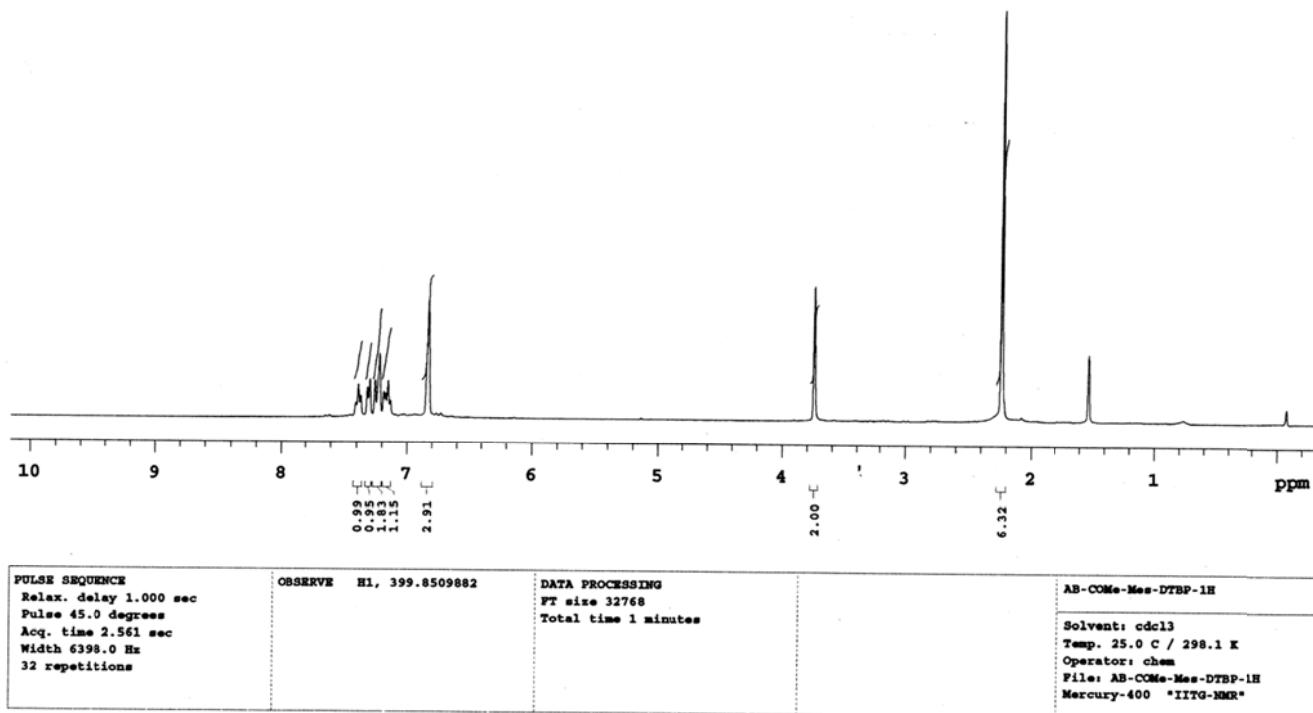


3-(4-Methylbenzyl)-2H-chromen-2-one (1e): ^{13}C NMR (CDCl_3 , 100 MHz)



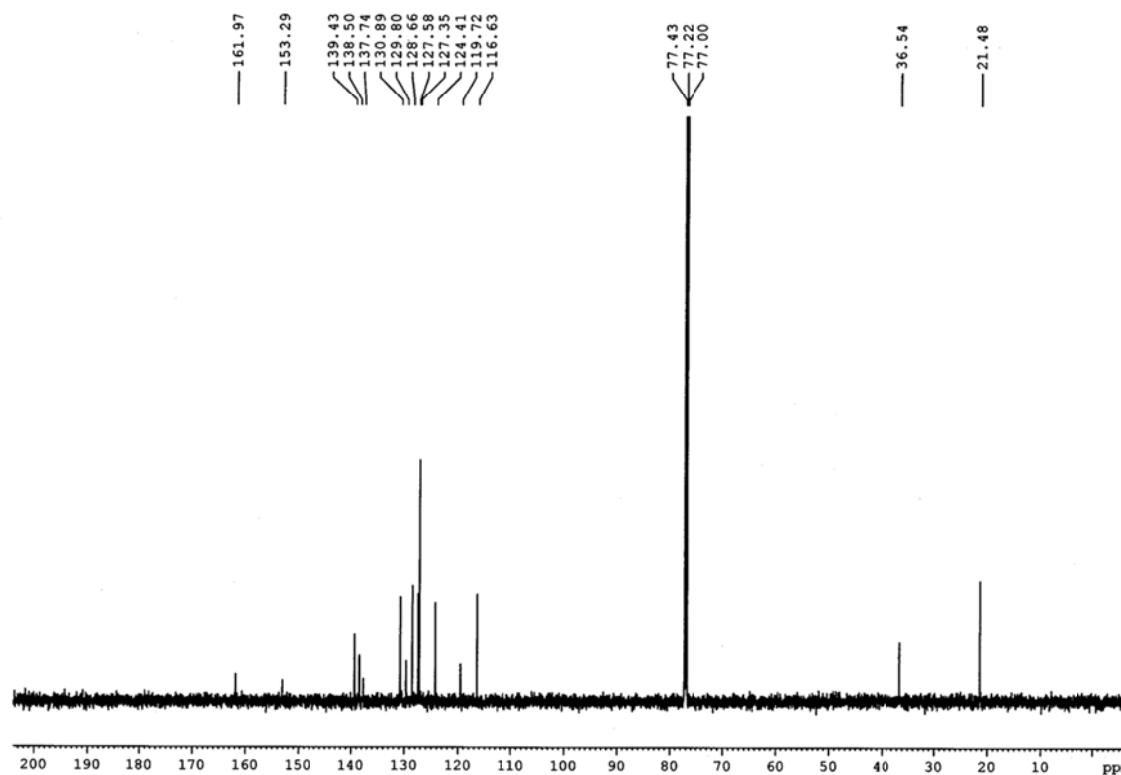
PULSE SEQUENCE	OBSERVE C13, 100.5425824	DATA PROCESSING	AB-COMeCou-P-XYL-13C
Relax. delay 1.000 sec	DECOUPLE B1, 399.8529994	Line broadening 0.5 Hz	Solvent: cdcl_3
Pulse 45.0 degrees	Power 42 dB	FT size 65536	Temp. 25.0 °C / 298.1 K
Acq. time 1.304 sec	continuously on	Total time 49 minutes	Operator: chem
Width 25125.6 Hz	MALTZ-16 modulated		Mercury-400 "IITG-NMR"
1290 repetitions			

3-(3,5-Dimethylbenzyl)-2H-chromen-2-one (1f): ^1H NMR (CDCl_3 , 400 MHz)



3-(3,5-Dimethylbenzyl)-2H-chromen-2-one (1f): ^{13}C NMR (CDCl_3 , 150 MHz)

AB_COME_MeS_ 13C



Current Data Parameters
NAME AB_COME_MeS_ 13C
EXPNO 1
PROCNO 1

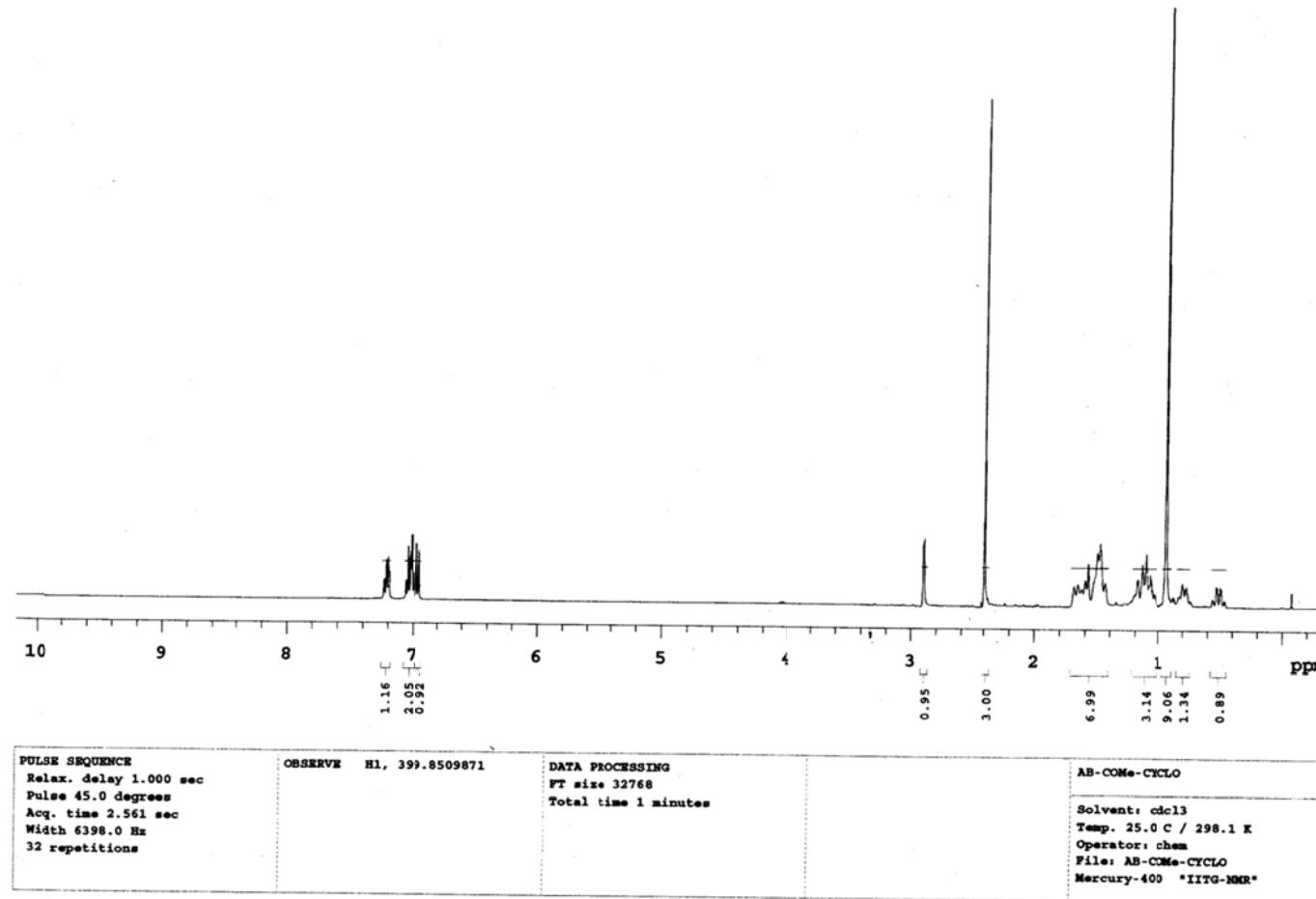
F2 - Acquisition Parameters
Date 20150327
Time 14.59
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgppg30
TD 32768
SOLVENT CDCl3
NS 114
DS 2
SWH 36057.691 Hz
FIDRES 1.100393 Hz
AQ 0.4543929 sec
RG 65.24
DW 13.867 usec
DE 6.50 usec
TE 298.2 K
D1 0.0000000 sec
D11 0.0300000 sec
TDO 1

===== CHANNEL f1 =====
SFO1 150.9279571 MHz
NUC1 13C
P1 10.50 usec
PLW1 95.00000000 W

===== CHANNEL f2 =====
SFO2 60.1724007 MHz
NUC2 1H
CPDPRG[2] waltz16
PCPD2 70.00 usec
PLW2 21.00000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

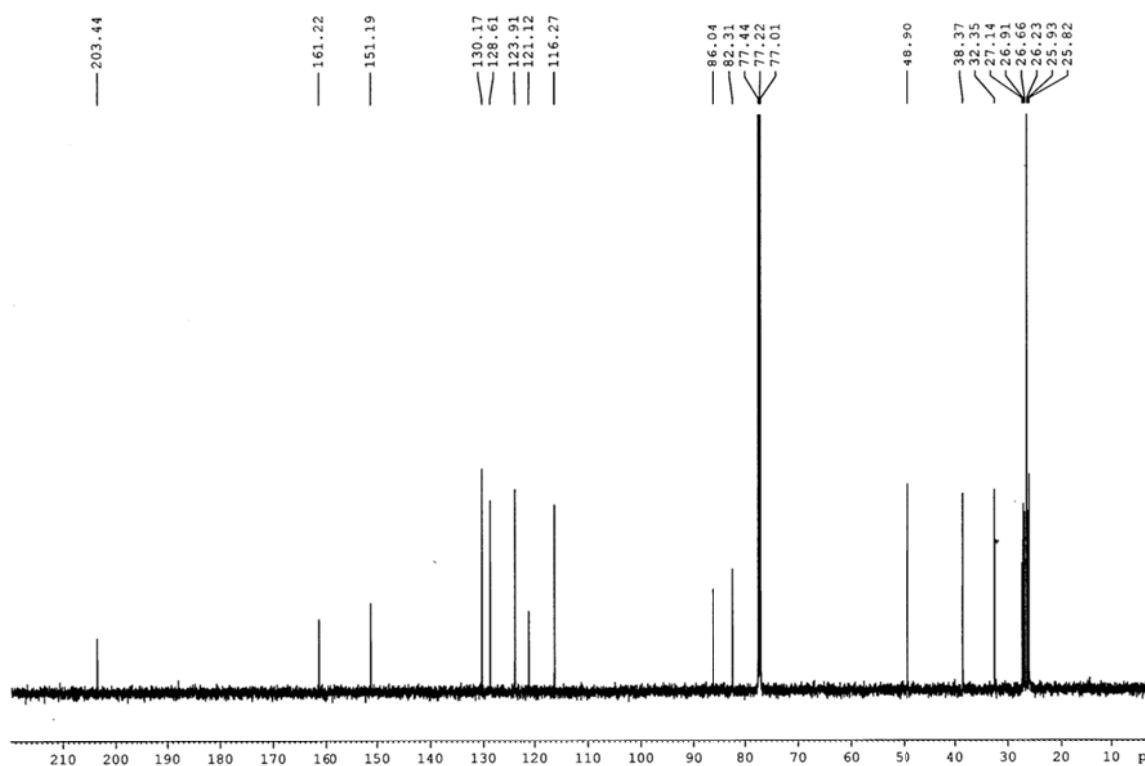
F2 - Processing parameters
SI 16384
SF 150.9128380 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (1a'): ^1H NMR (CDCl_3 , 400 MHz)



3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (1a'): ^{13}C NMR (CDCl_3 , 150 MHz)

AB-COME-CYCLO-13C



Current Data Parameters
NAME AB-COME-CYCLO-13C
EXPNO 1
PROCNO 1

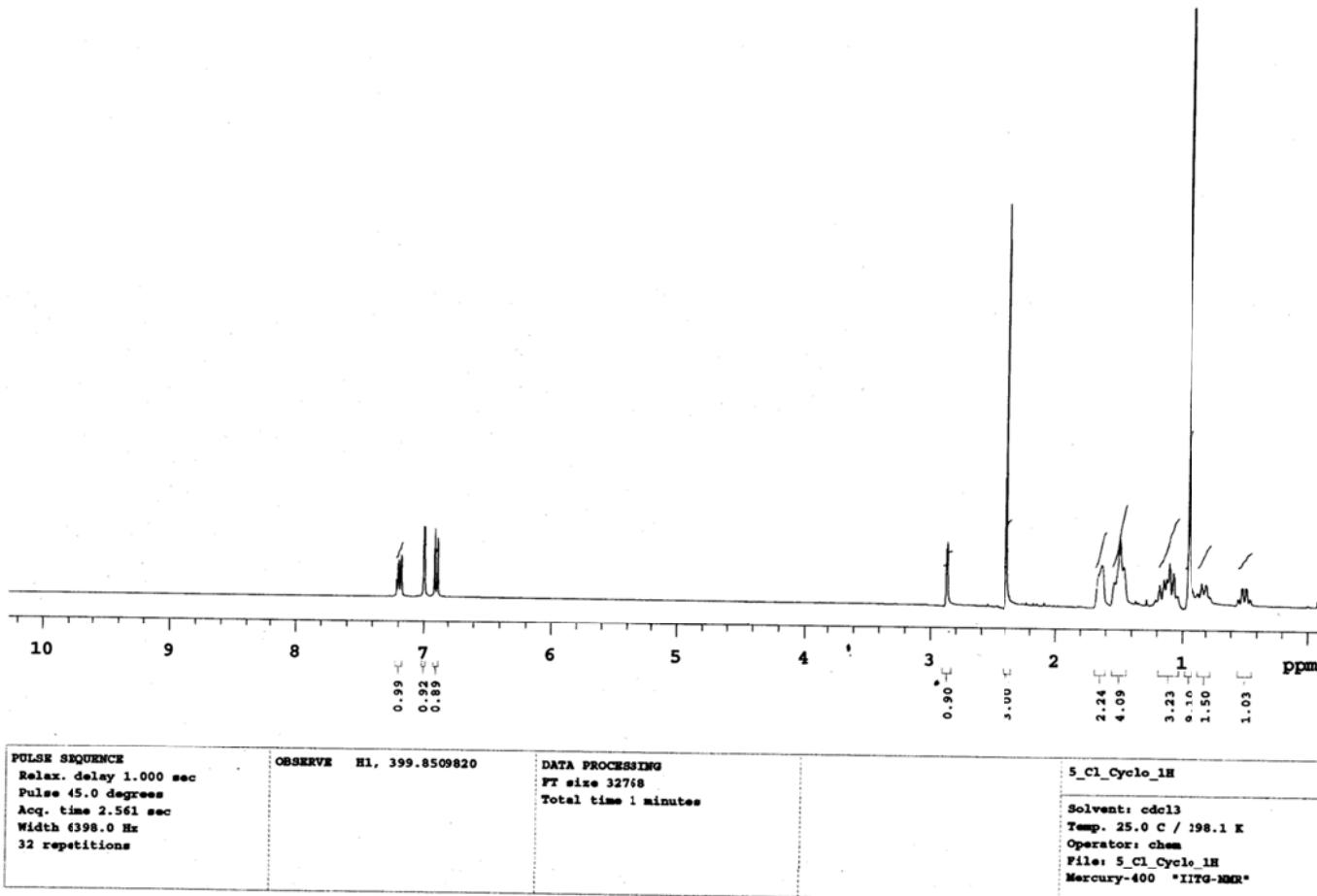
F2 - Acquisition Parameters
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Time 13.59
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PULPROG zgpp30
TD 32768
SOLVENT CDCl3
NS 206
DS 2
SWH 36057.491 Hz
FIDRES 1.100000 Hz
AQ 0.1543829 sec
RG 55.24
DW 13.867 usec
DE 6.50 usec
TE 300.1 K
D1 2.0000000 sec
D11 0.0000000 sec
TD0 1

===== CHANNEL f1 =====
SFO1 150.9279571 MHz
NUC1 ^{13}C
P1 10.50 usec
PLW1 95.0000000 W

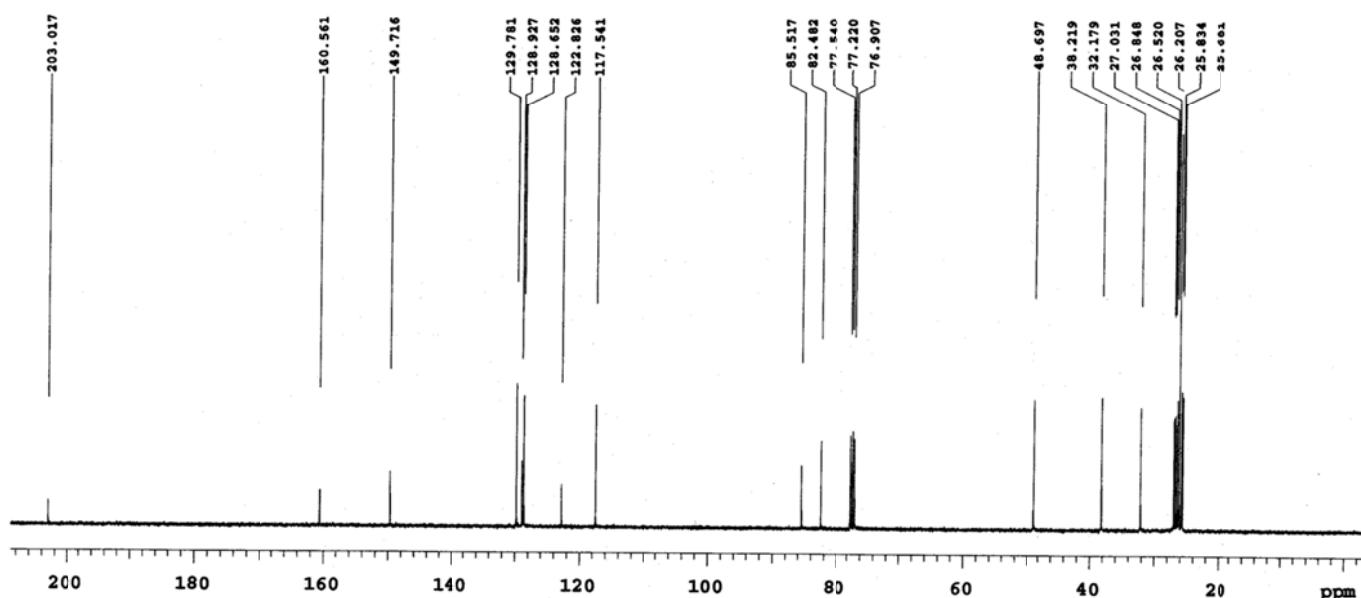
===== CHANNEL f2 =====
SFO2 600.1724007 MHz
NUC2 ^1H
CPDPRG[2] walkin
PCPDG2 70.00 usec
PLW2 21.0000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

F2 - Processing parameters
SI 16384
SF 150.9128363 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40

3-Acetyl-3-(*tert*-butylperoxy)-6-chloro-4-cyclohexylchroman-2-one (13a'): ^1H NMR (CDCl_3 , 400 MHz)

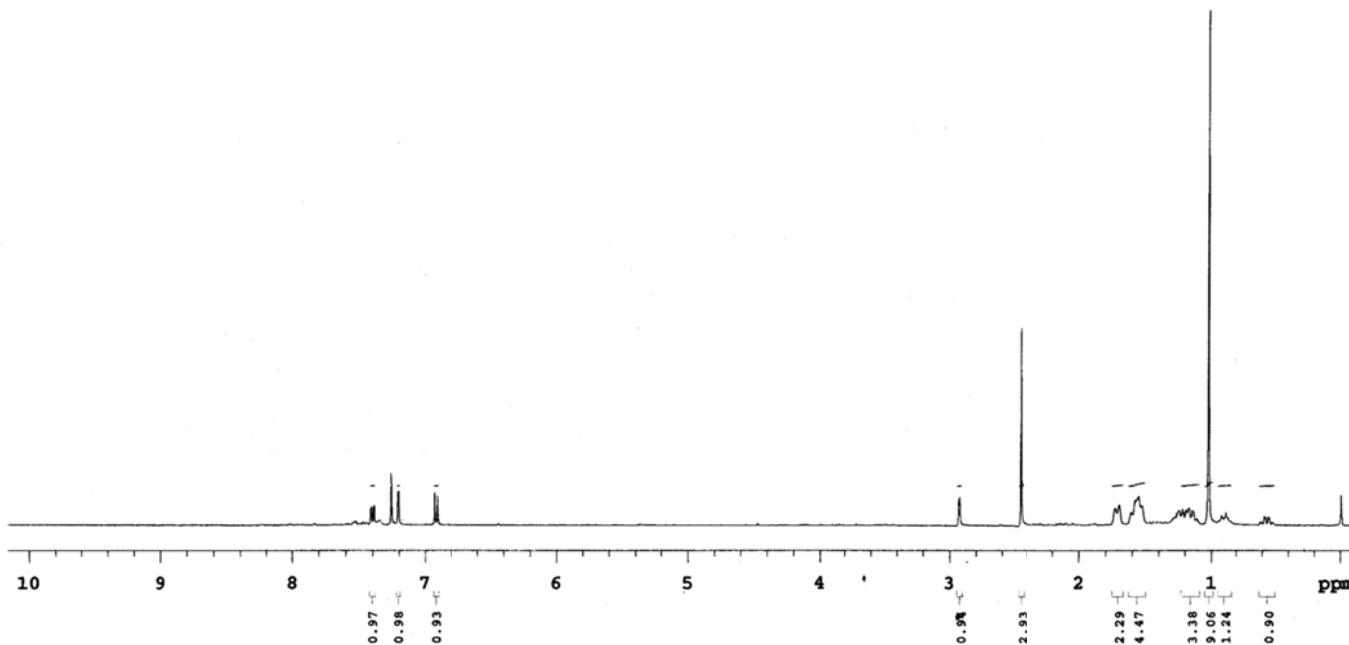


3-Acetyl-3-(*tert*-butylperoxy)-6-chloro-4-cyclohexylchroman-2-one (13a'): ^{13}C NMR (CDCl_3 , 100 MHz)



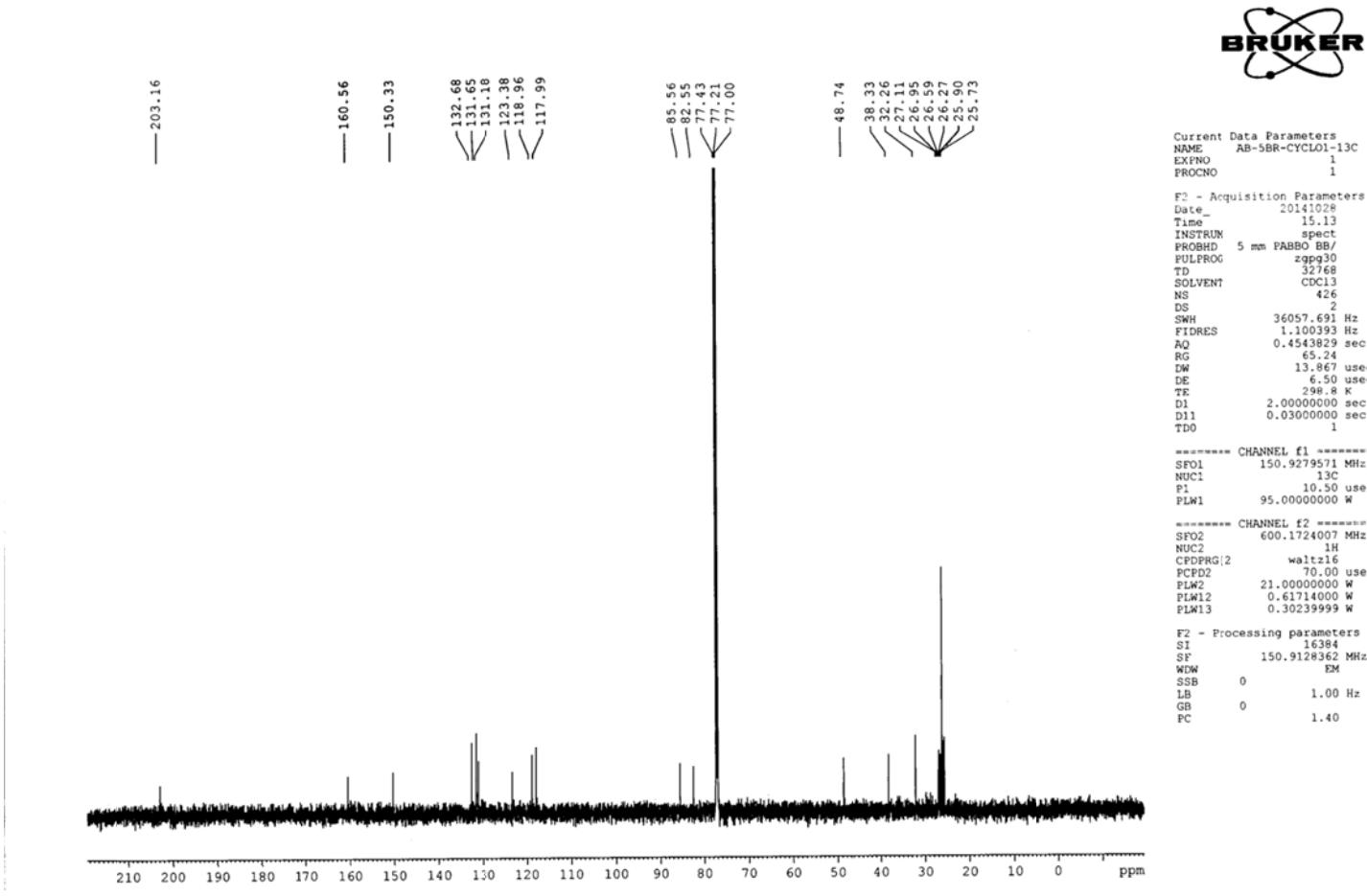
PULSE SEQUENCE	OBSERVE C13, 100.5425873 DECOUPLE H1, 399.8529994	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 26 minutes	5_Cl_Cyclo_13C
Relax. delay 1.000 sec Pulse 45.0 degrees Acc. time 1.304 sec Width 25125.6 Hz 700 repetitions	continuously on MALTZ-16 modulated		Solvent: cdcl_3 Temp. 25.0 °C / 298.1 K Operator: chem File: 5_Cl_Cyclo_13C Mercury-400 "ITG-NMR"

3-Acetyl-6-bromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (14a'): ^1H NMR (CDCl_3 , 400 MHz)

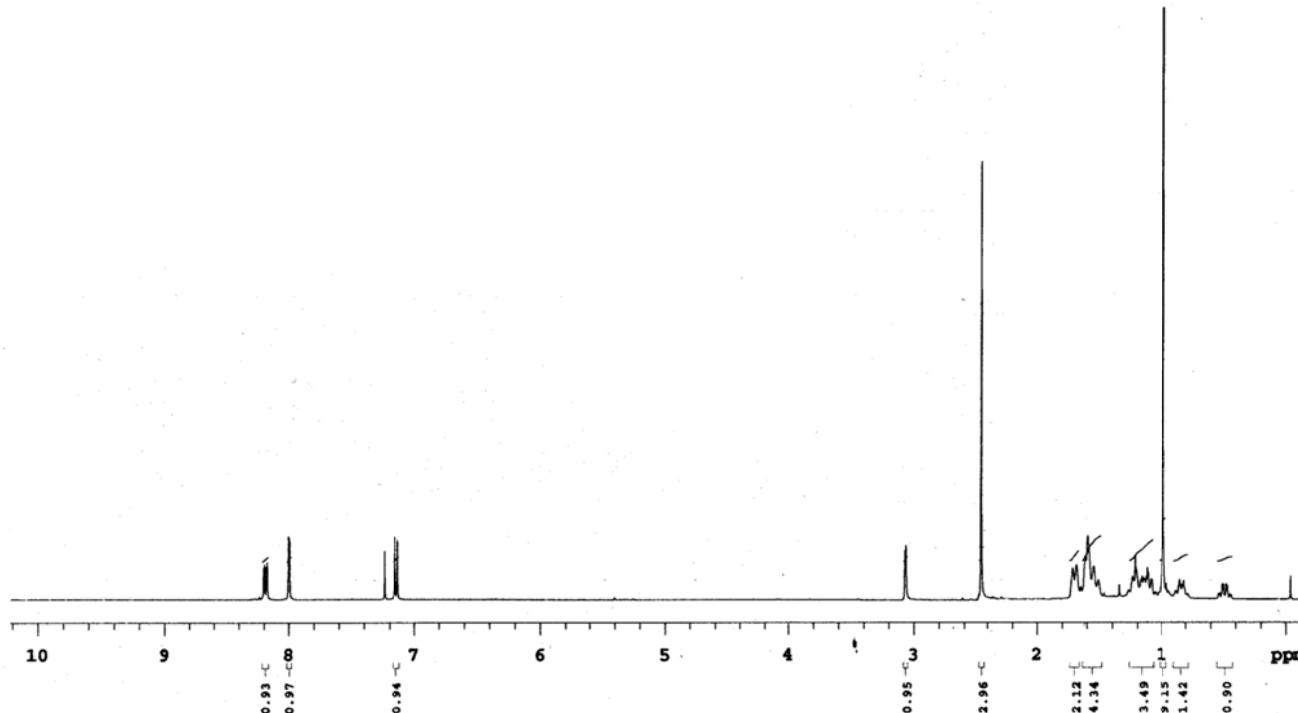


PULSE SEQUENCE	OBSERVE H1, 399.8509641	DATA PROCESSING	AB_5_Br_COMe_Cyclo_1H
Relax. delay 1.000 sec			Solvent: cdcl_3
Pulse 45.0 degrees			Temp. 25.0 C / 298.1 K
Acq. time 2.561 sec			Operator: chem
Width 6398.0 Hz			File: AB_5_Br_COMe_Cyclo_1H
32 repetitions			Mercury-400 "ITG-NMR"

3-Acetyl-6-bromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (14a'): ^{13}C NMR (CDCl_3 , 150 MHz)

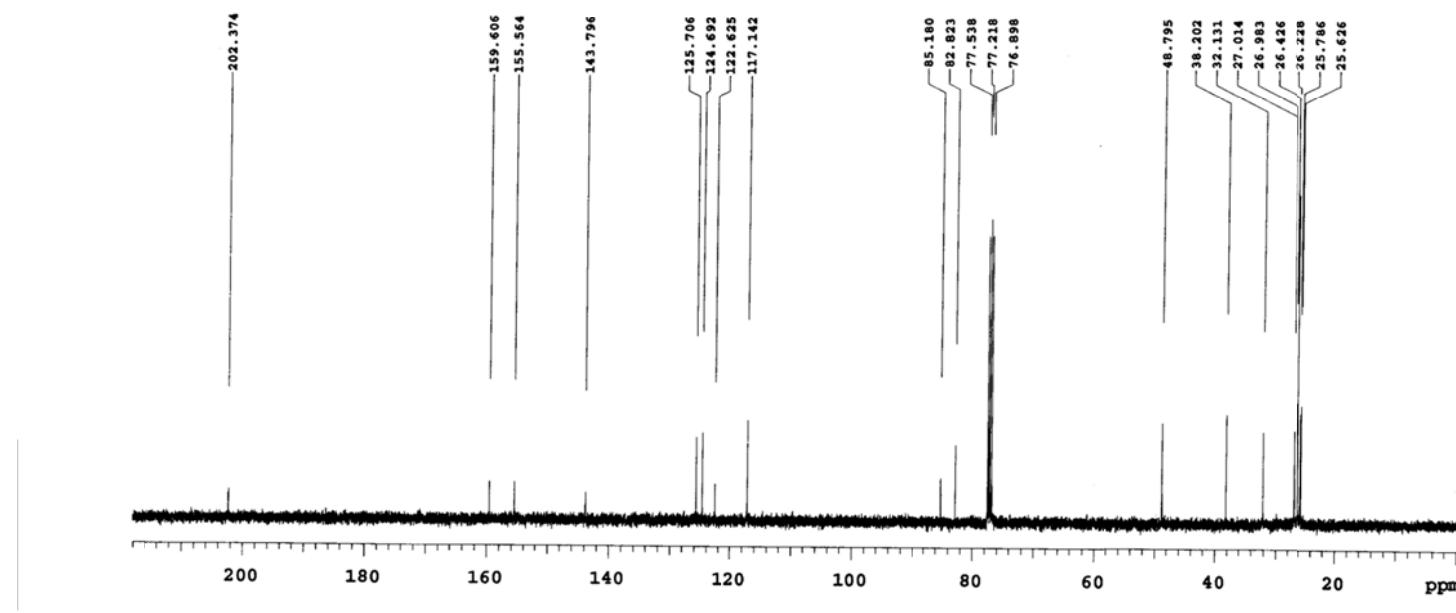


3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-6-nitrochroman-2-one (5a'): ^1H NMR (CDCl_3 , 400 MHz)



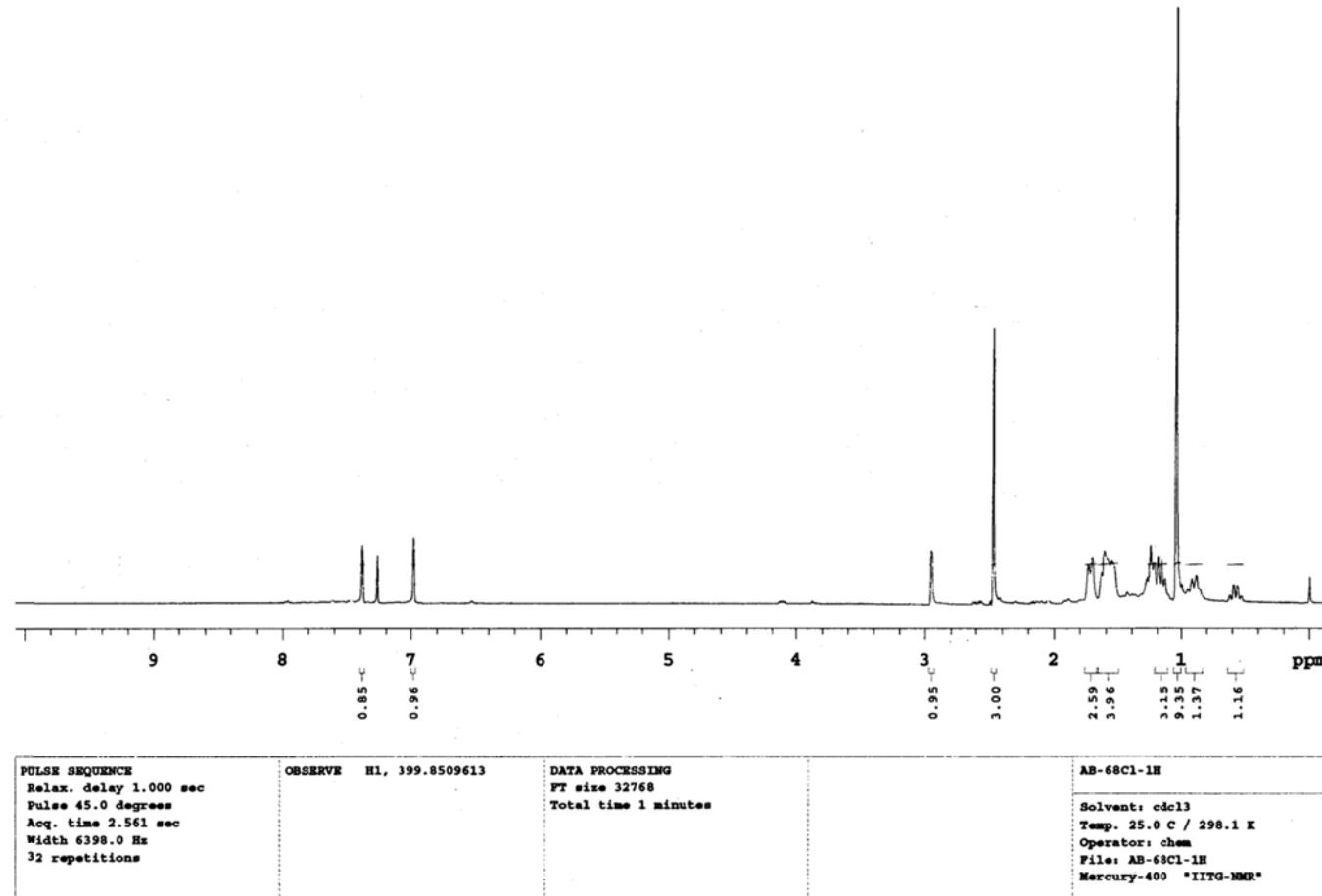
PULSE SEQUENCE Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 2.561 sec Width 6396.0 Hz 32 repetitions	OBSERVE H1, 399.8509721	DATA PROCESSING FT size 32768 Total time 1 minutes	AB-5-Me2COMe-Cyclo Solvent: cdcl3 Temp. 25.0 C / 298.1 K Operator: chem File: AB-5-Me2COMe-Cyclo Mercury-400 "ITG-NMR"
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3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-6-nitrochroman-2-one (5a'): ^{13}C NMR (CDCl_3 , 100 MHz)

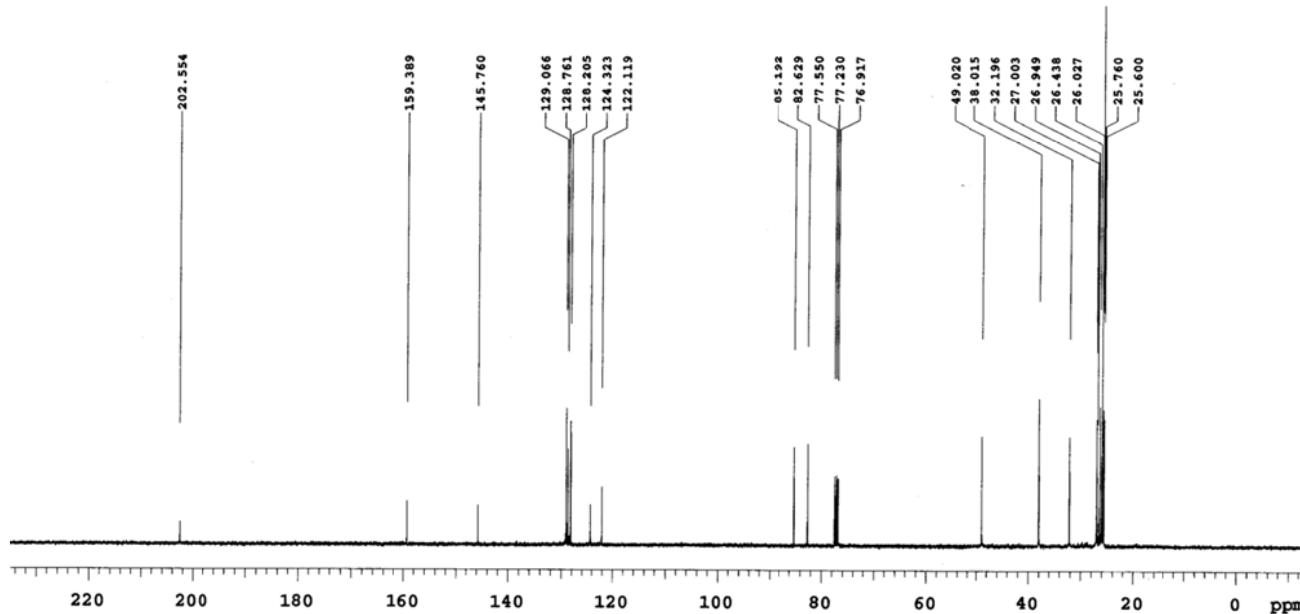


PULSE SEQUENCE	OBSERVE C13, 100.5425852	DATA PROCESSING	SS-AB-SNO2-C-13C
Relax. delay 1.000 sec	DECOUPLE H1, 399.8529994	Line broadening 0.5 Hz	Solvent: cdcl_3
Pulse 45.0 degrees	Power 42 dB	FT size 65536	Temp. 25.0 C / 298.1 K
Acq. time 1.304 sec	continuously on	Total time 17 minutes	Operator: chem
Width 25125.6 Hz	WALTZ-16 modulated		File: SS-AB-SNO2-C-13C
160 repetitions			Mercury-400 "IITG-NMR"

3-Acetyl-3-(*tert*-butylperoxy)-6,8-dichloro-4-cyclohexylchroman-2-one (6a'): ^1H NMR (CDCl_3 , 400 MHz)

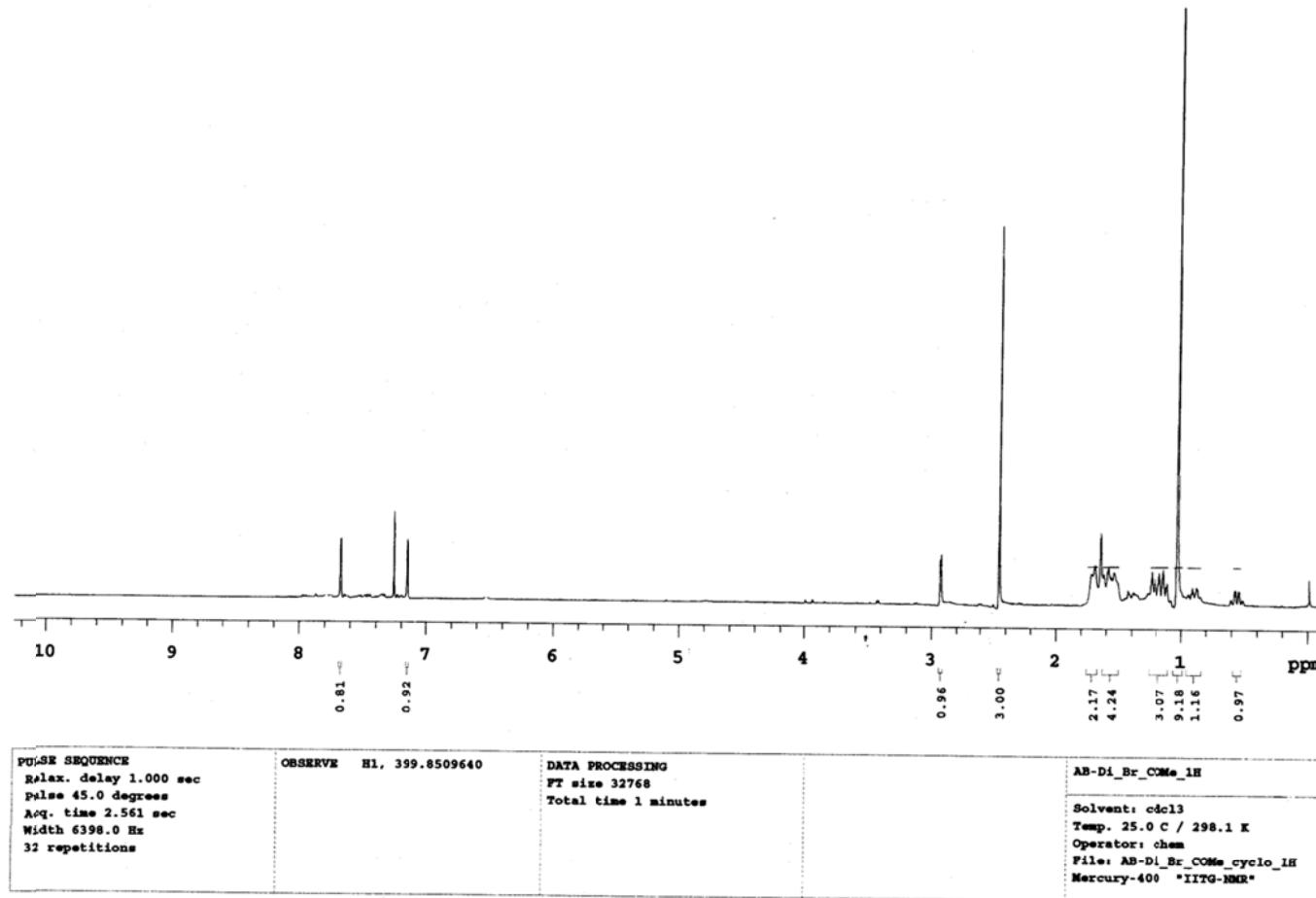


3-Acetyl-3-(*tert*-butylperoxy)-6,8-dichloro-4-cyclohexylchroman-2-one (6a'): ^{13}C NMR (CDCl_3 , 100 MHz)

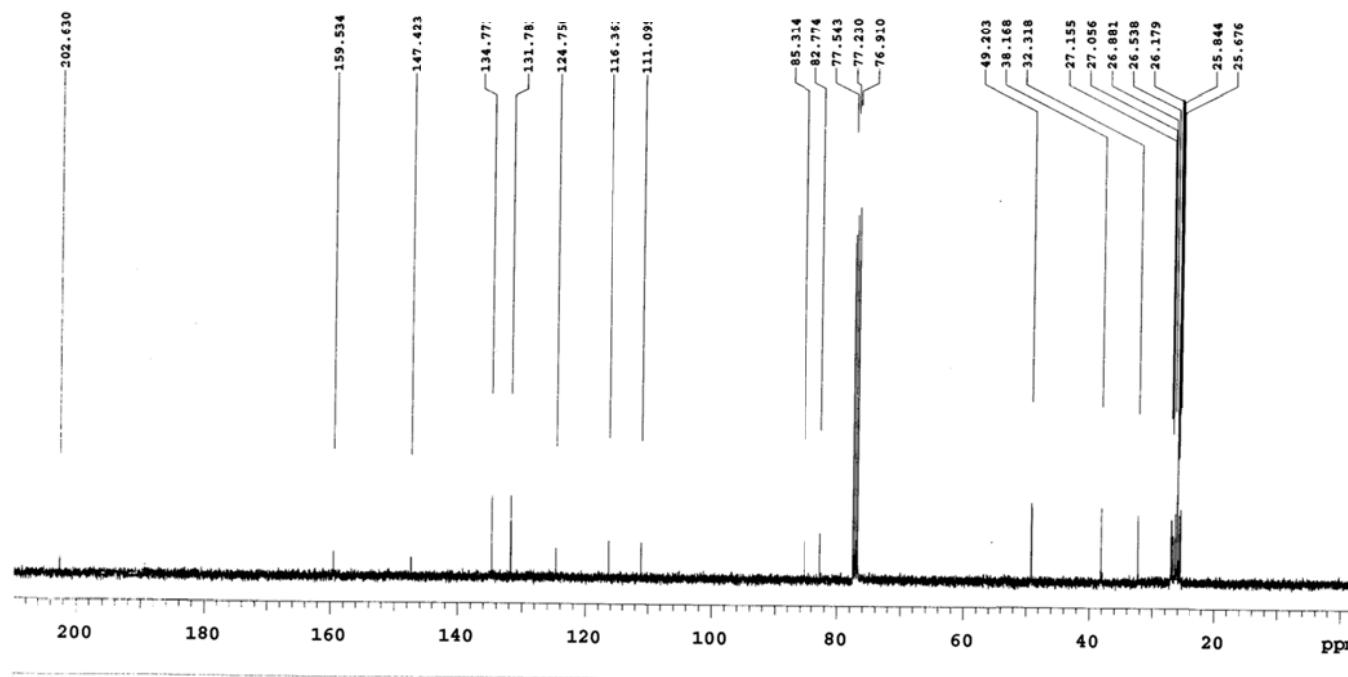


PULSE SEQUENCE	OBSERVE C13, 100.5425909 DECOUPLE H1, 399.8529994	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 12 minutes	AB-35C1Cyclo_U2_13C
Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 1.304 sec Width 25125.6 Hz 320 repetitions	WALTZ-16 modulated		Solvent: cdcl_3 Temp. 25.0 C / 298.1 K Operator: chem File: AB-35C1Cyclo_U2_13C Mercury-400 "IITG-NMR"

3-Acetyl-6,8-dibromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (7a'): ^1H NMR (CDCl_3 , 400 MHz)



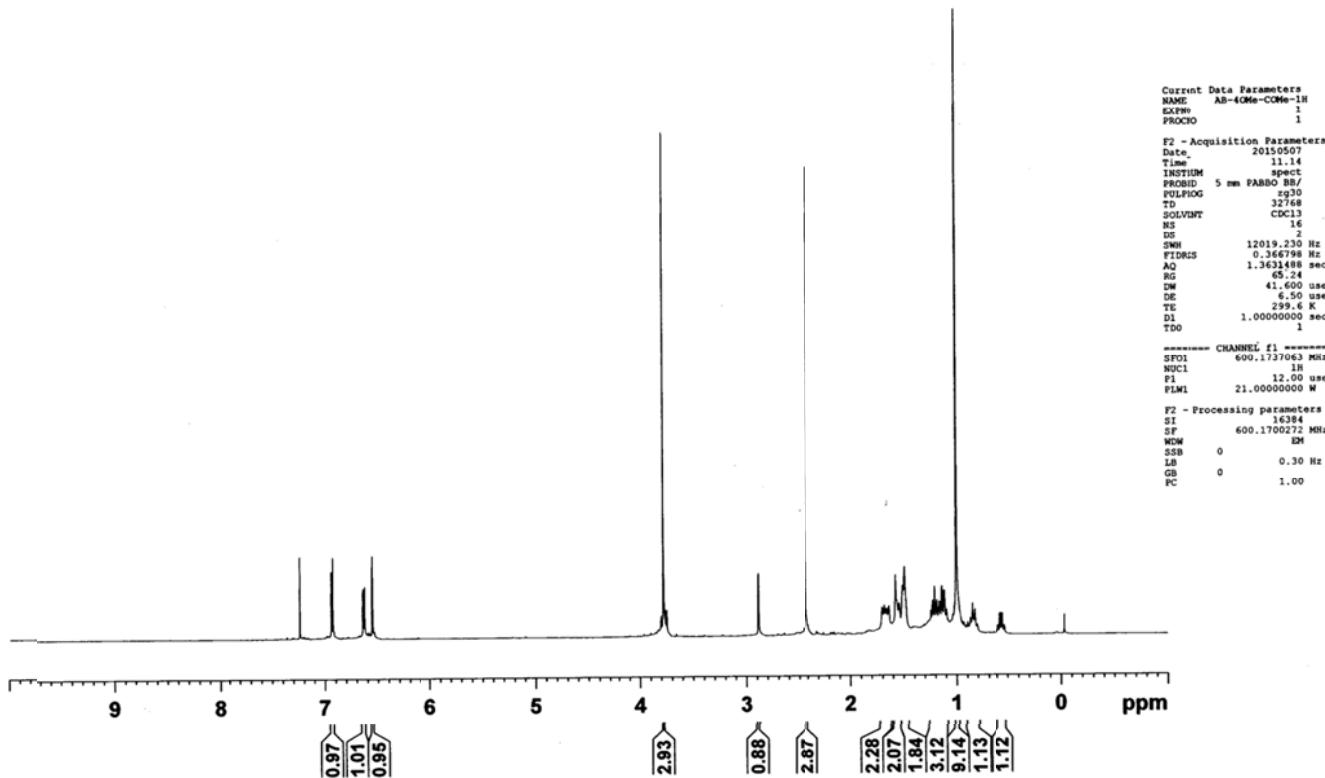
3-Acetyl-6,8-dibromo-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (7a'): ^{13}C NMR (CDCl_3 , 100 MHz)



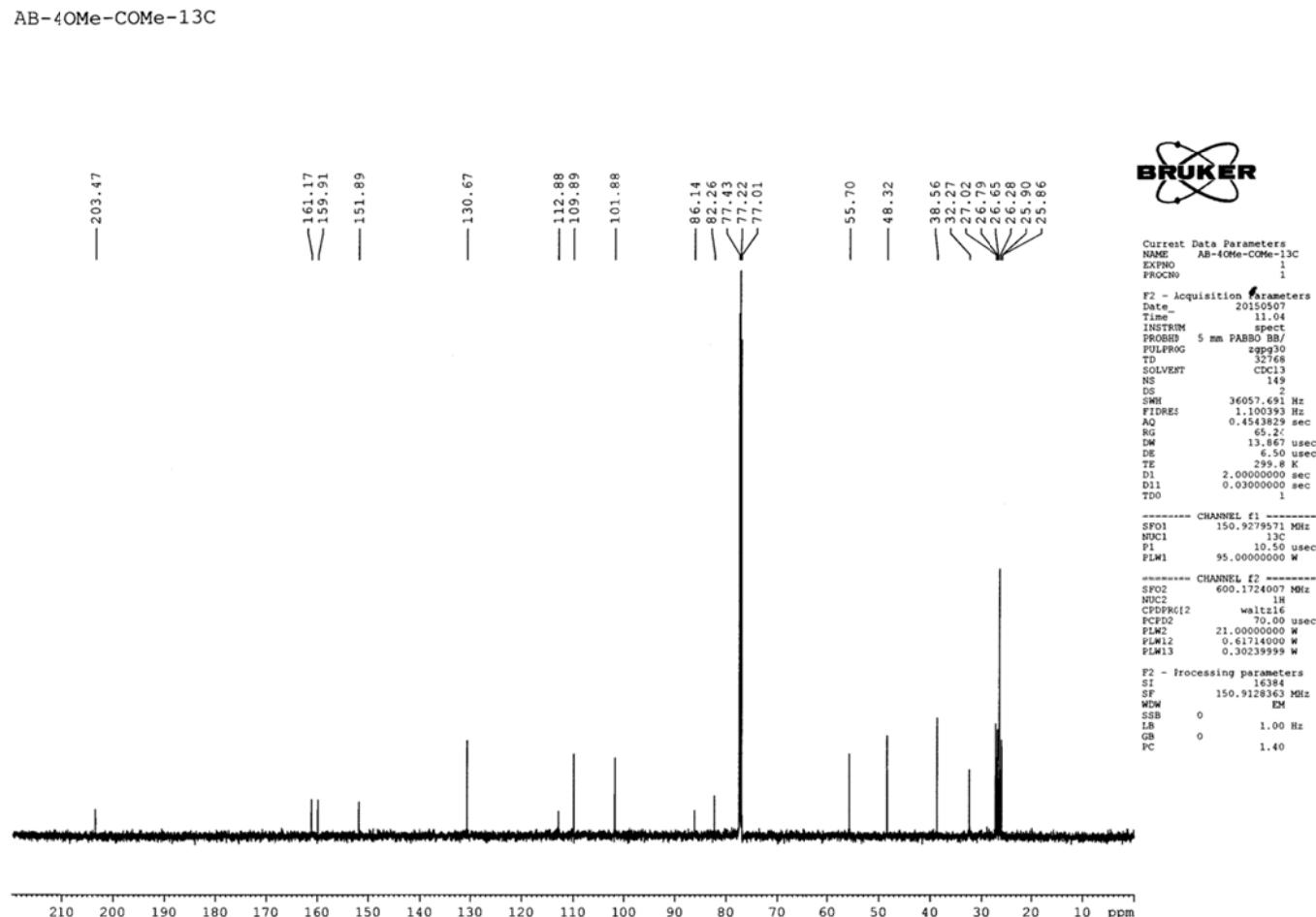
PULSE SEQUENCE	OBSERVE C13, 100.5425840 DECOUPLE H1, 399.8529994	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 36 minutes	AB-Di_Br_COMe_Cyclo_13C
Relax. delay 1.000 sec Pulse 45.0 degrees Aqg. time 1.304 sec Width 25125.6 Hz 940 repetitions	WALTZ-16 modulated		Solvent: cdcl_3 Temp. 25.0 C / 298.1 K Operator: chem File: AB-Di_Br_COMe_cyclo_13C Mercury-400 "IITG-NMR"

3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-7-methoxychroman-2-one (2a'): ^1H NMR (CDCl_3 , 600 MHz)

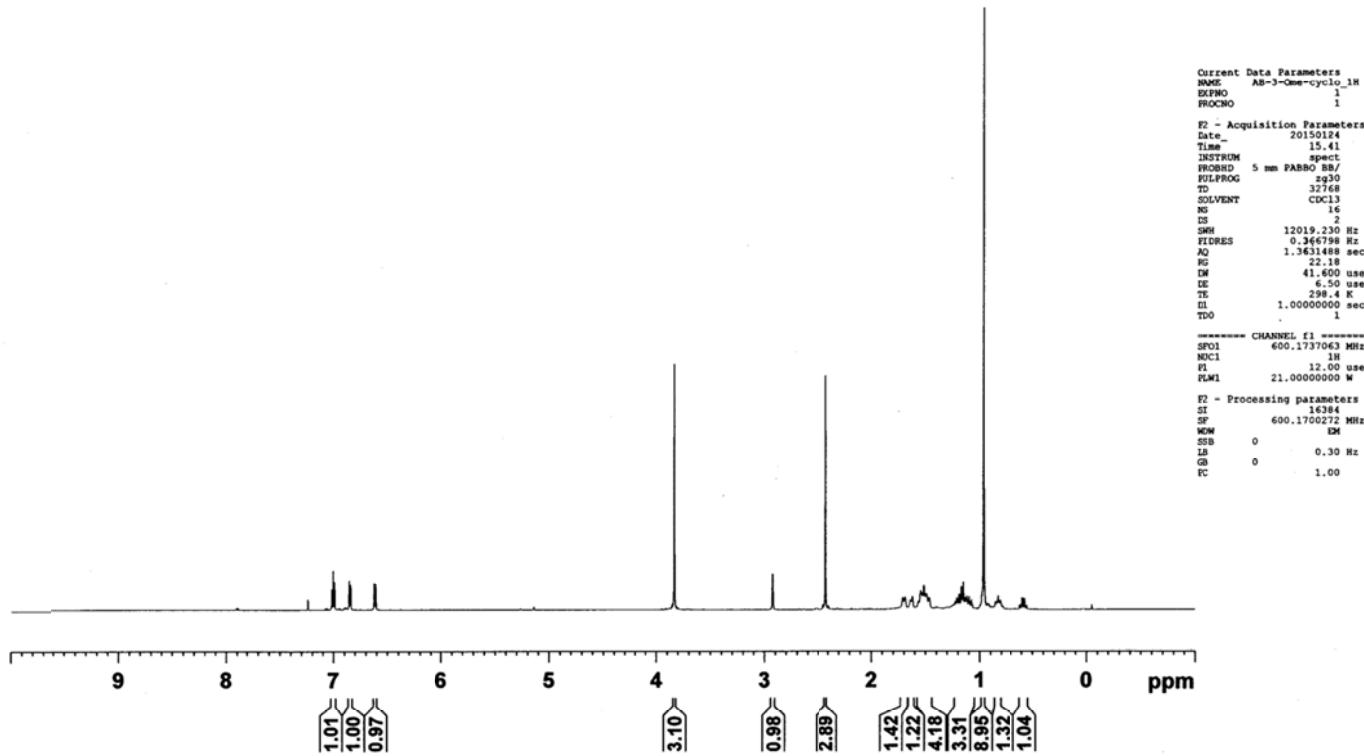
AB-4OMe-COMe-1H



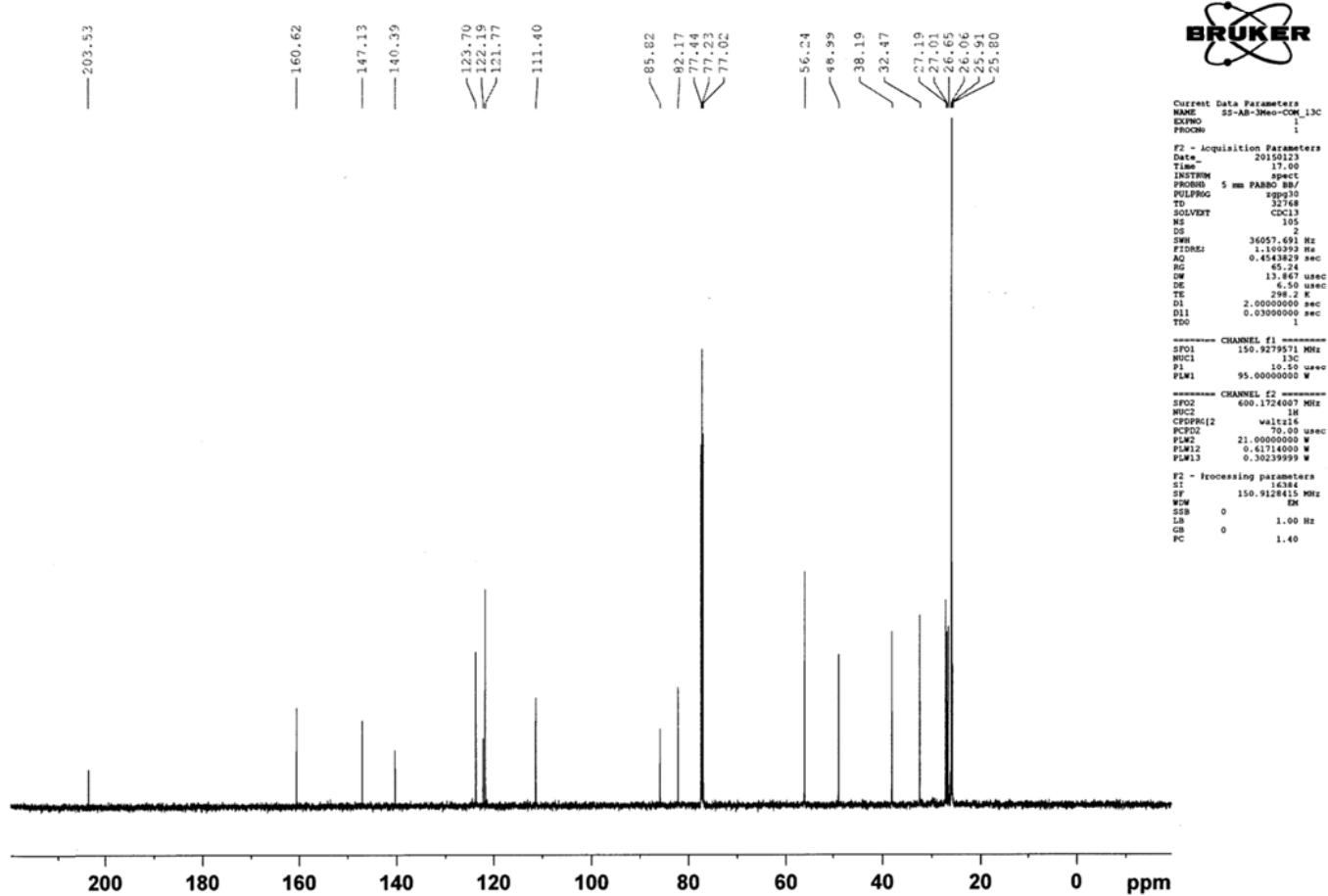
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-7-methoxychroman-2-one (2a'): ^{13}C NMR (CDCl_3 , 150 MHz)



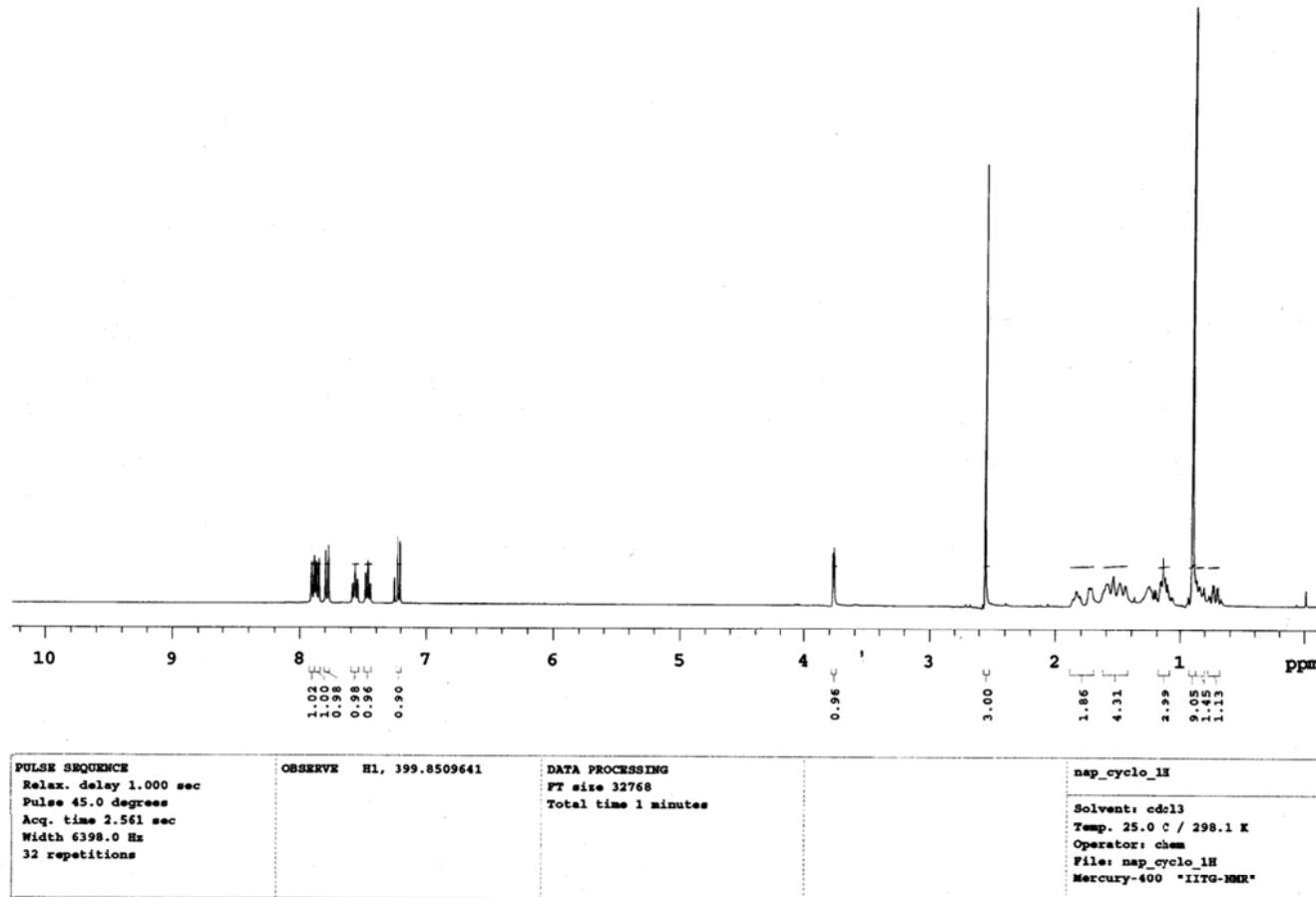
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-8-methoxychroman-2-one (15a'): ^1H NMR (CDCl_3 , 600 MHz)



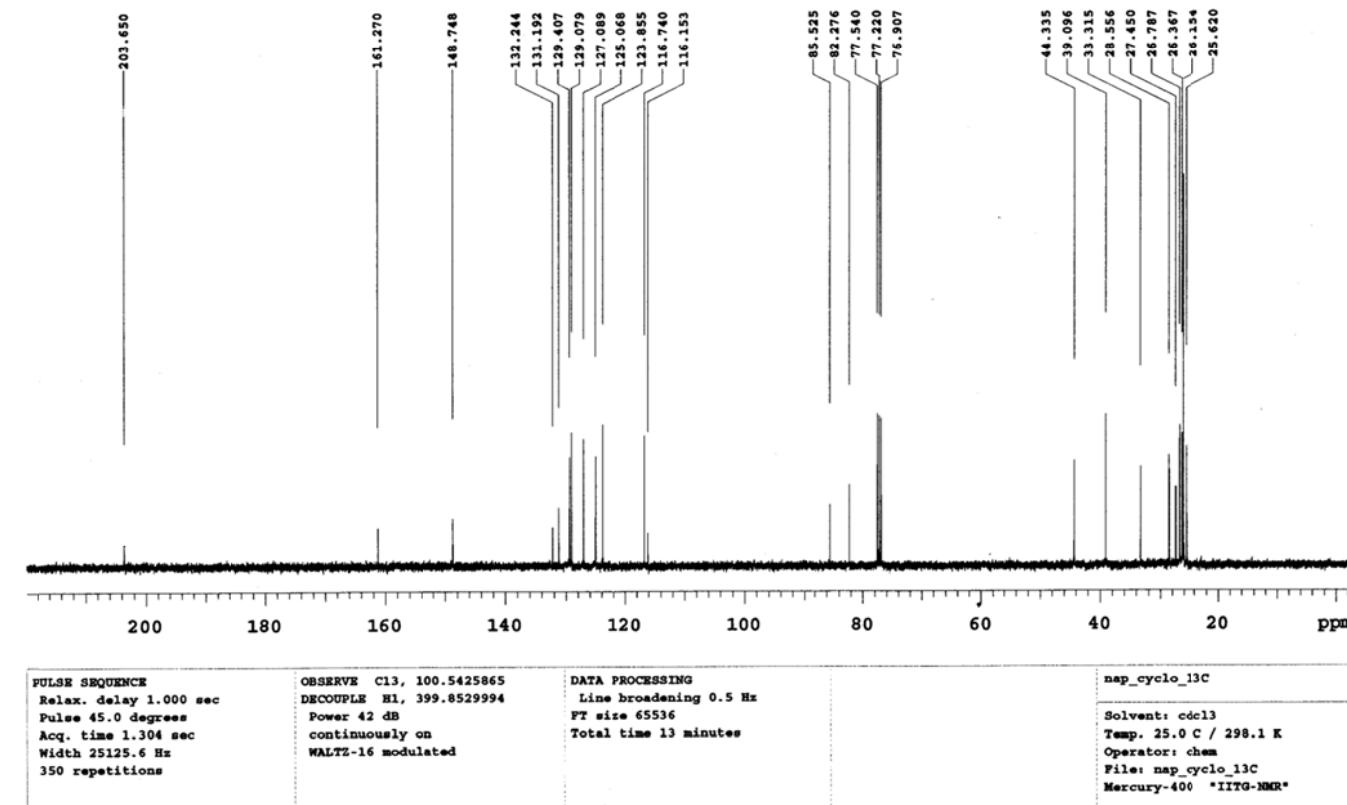
3-Acetyl-3-(*tert*-butylperoxy)-4-cyclohexyl-8-methoxychoman-2-one (15a'): ^{13}C NMR (CDCl_3 , 150 MHz)



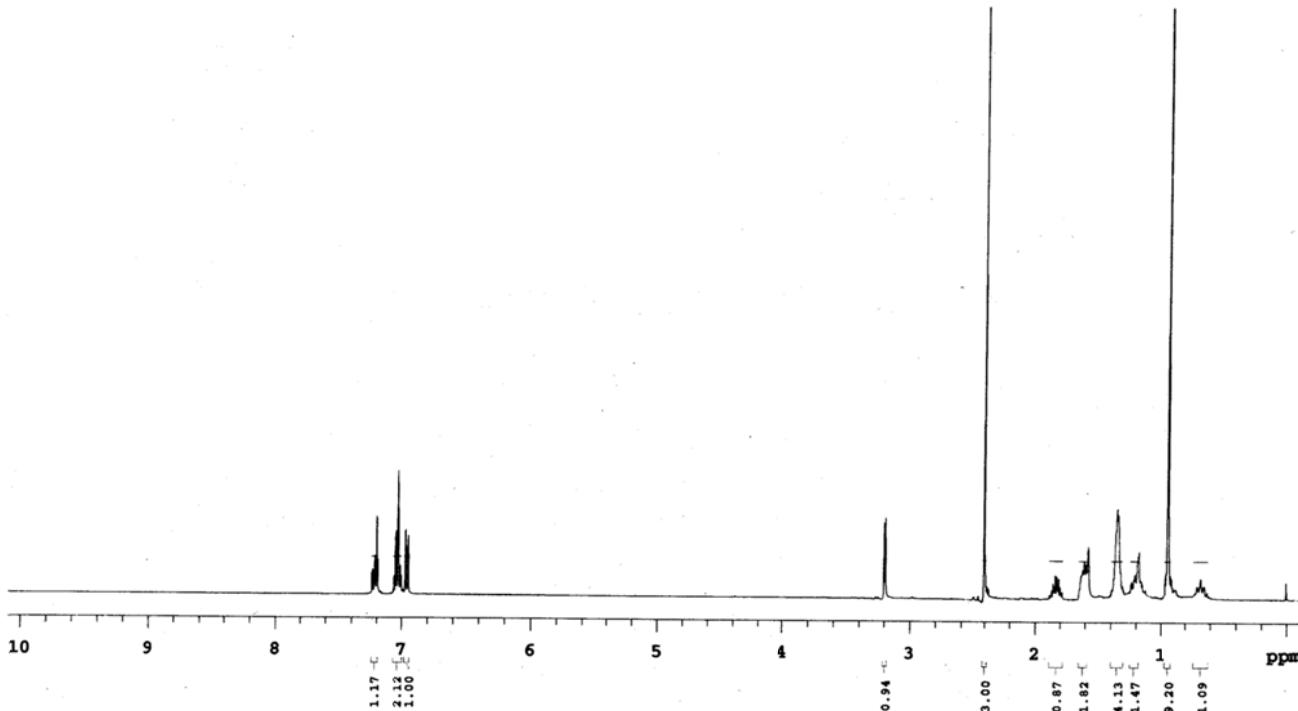
2-Acetyl-2-(*tert*-butylperoxy)-1-cyclohexyl-1H-benzo[*f*]chromen-3(2H)-one (8a'): ^1H NMR (CDCl_3 , 400 MHz)



2-Acetyl-2-(*tert*-butylperoxy)-1-cyclohexyl-1H-benzo[*f*]chromen-3(2H)-one (8a'): ^{13}C NMR (CDCl_3 , 100 MHz)

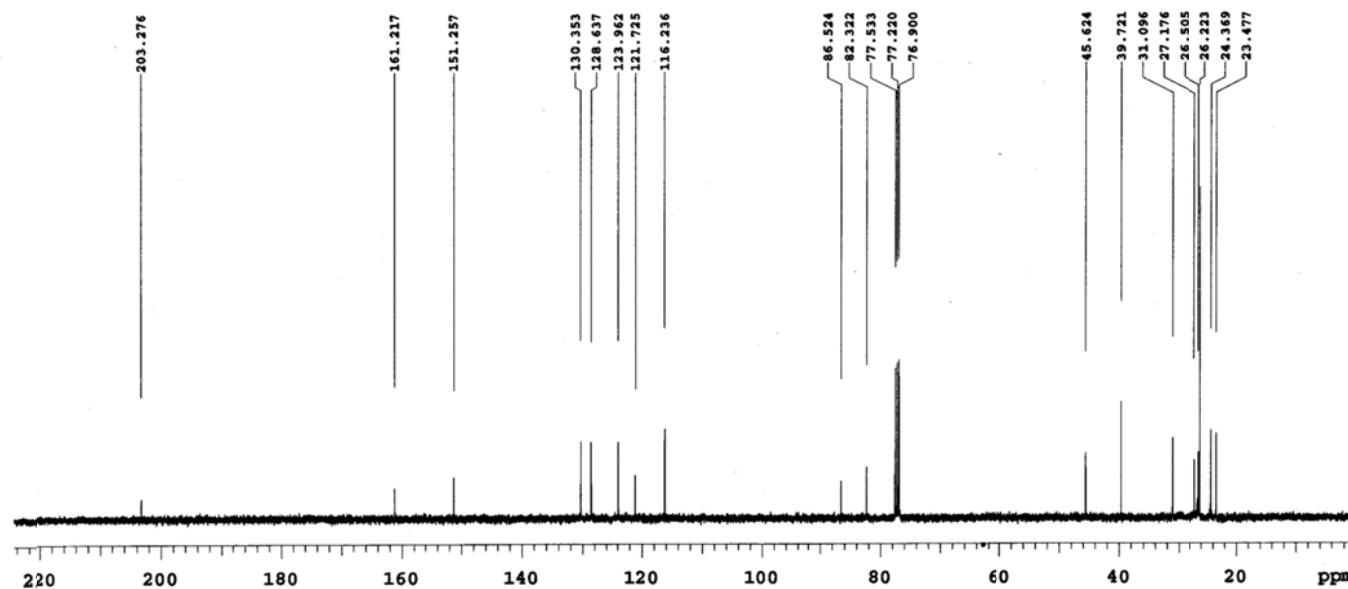


3-Acetyl-3-(*tert*-butylperoxy)-4-cyclopentylchroman-2-one (1b'): ^1H NMR (CDCl_3 , 400 MHz)



PULSE SEQUENCE	OBSERVE	DATA PROCESSING	
Relax. delay 1.000 sec	H1, 399.8509847	FT size 32768	AB_COMe_Cyclopentane_1H
Pulse 45.0 degrees		Total time 1 minutes	Solvent: cdcl3
Acq. time 2.561 sec			Temp. 25.0 C / 298.1 K
Width 6398.0 Hz			Operator: chem
32 repetitions			File: AB_COMe_Cyclopentane_1H
			Mercury-400 "IITG-NMR"

3-Acetyl-3-(*tert*-butylperoxy)-4-cyclopentylchroman-2-one (1b'): ^{13}C NMR (CDCl_3 , 100 MHz)



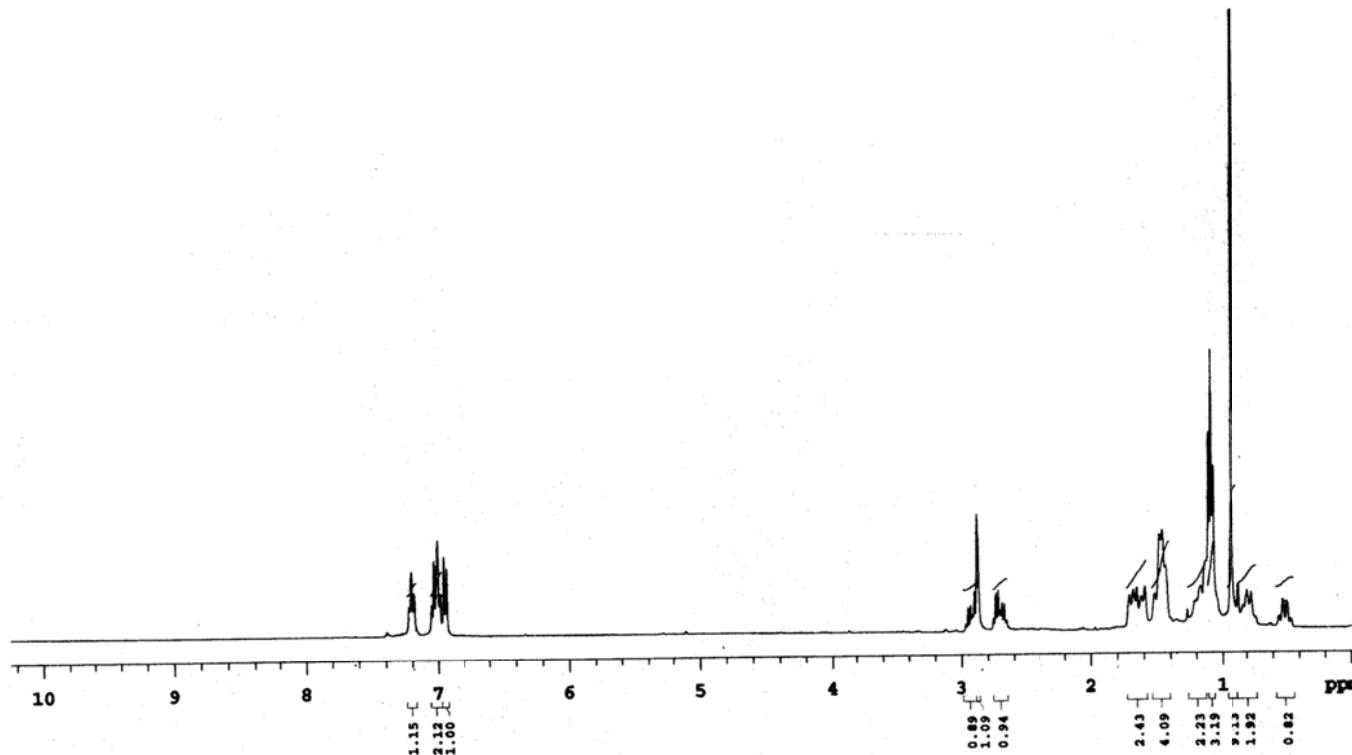
PULSE SEQUENCE
Relax. delay 1.000 sec
pulse 45.0 degrees
Acq. time 1.304 sec
Width 25125.6 Hz
520 repetitions

OBSERVE C13, 100.5425842
DECOUPLE H1, 399.8529994
Power 42 dB
continuously on
WALTZ-16 modulated

DATA PROCESSING
Line broadening 0.5 Hz
FT size 65536
Total time 19 minutes

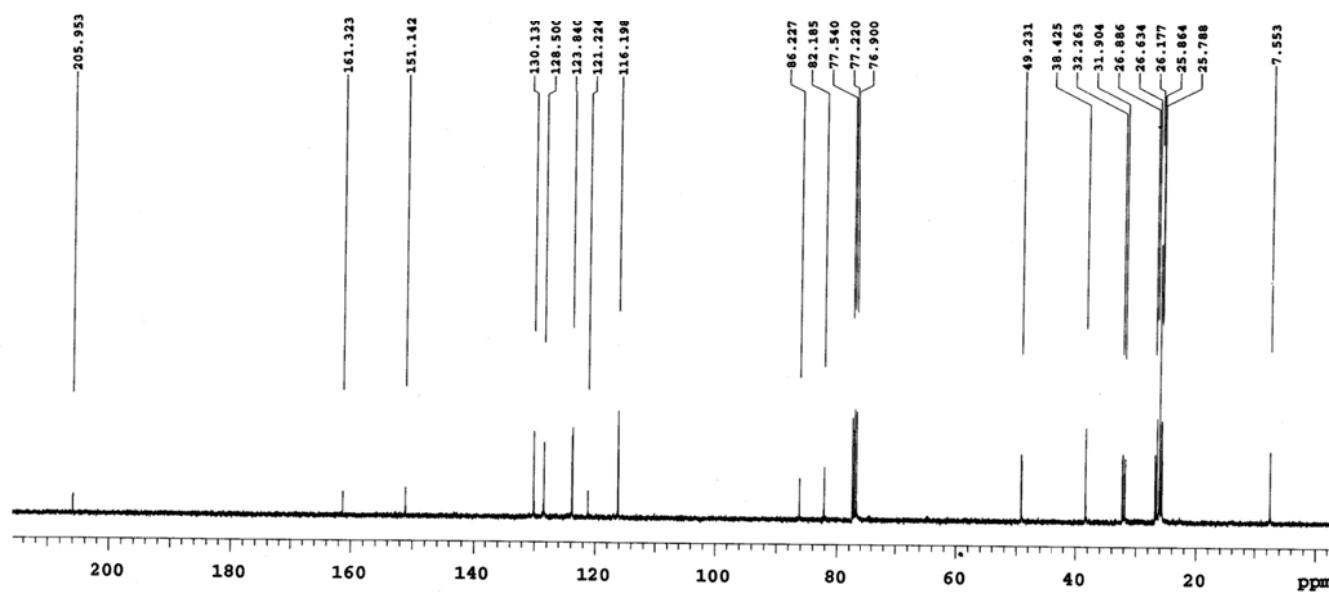
AB_COMe_Cyclopentane_13C
Solvent: cdcl_3
Temp. 25.0 C / 298.1 K
Operator: chem
File: AB_COMe_Cyclopentane_13C
Mercury-401 "IIIG-NMR"

3-(*tert*-Butylperoxy)-4-cyclohexyl-3-propionylchroman-2-one (9a'): ^1H NMR (CDCl_3 , 400 MHz)



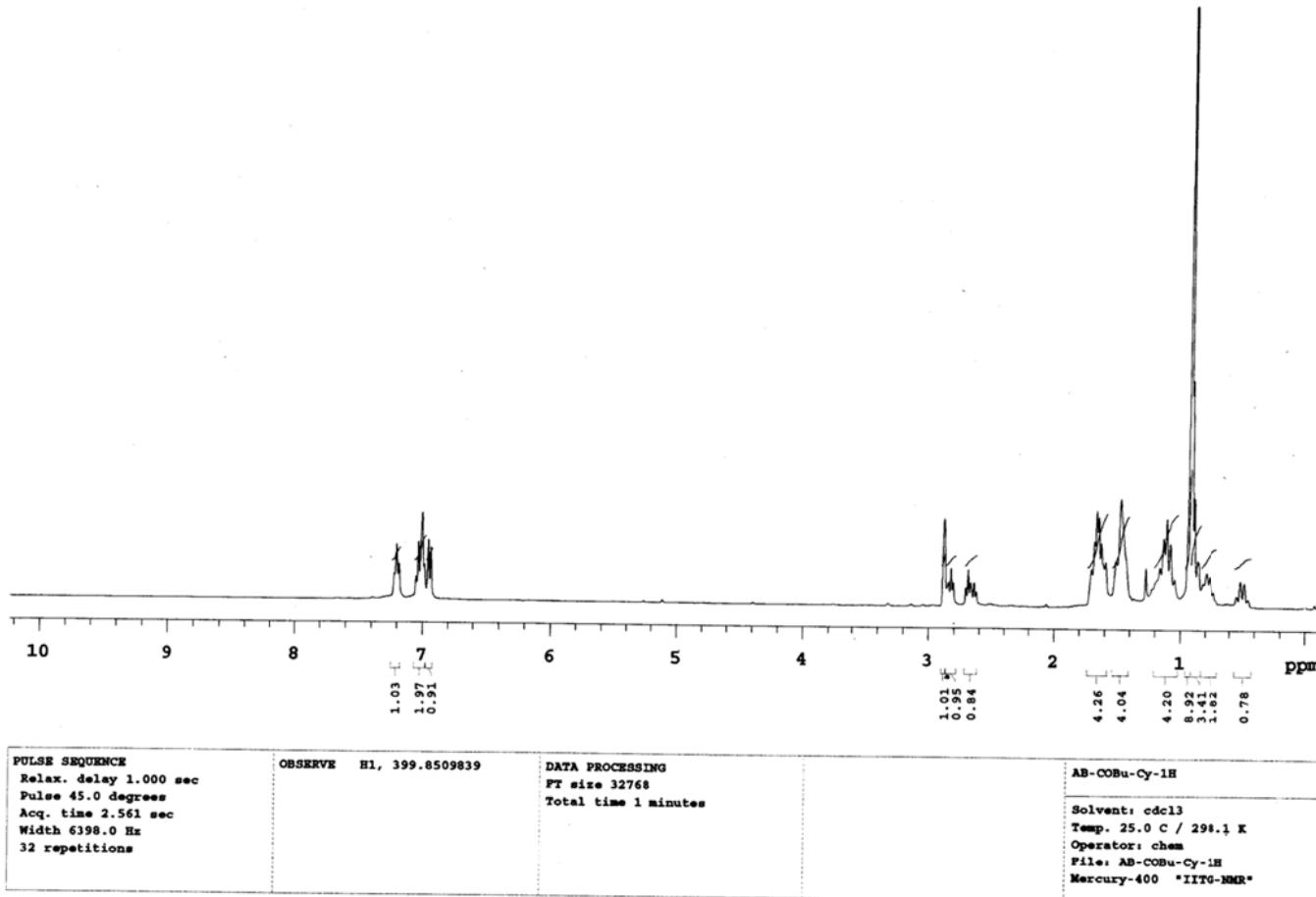
PULSE SEQUENCE	OBSERVE	DATA PROCESSING	AB-CORT-Cy-1H
Relax. delay 1.000 sec Pulse 45.0 degrees Aq. time 2.561 sec Width 4398.0 Hz 32 repetitions	H1, 399.8509836	FT size 32768 Total time 1 minutes	Solvent: cdcl_3 Temp. 25.0 C / 298.1 K Operator: chem File: AB-CORT-Cy-1H Mercury-400 "ITIG-NMR"

3-(*tert*-Butyperoxy)-4-cyclohexyl-3-propionylchroman-2-one (9a'): ^{13}C NMR (CDCl_3 , 100 MHz)

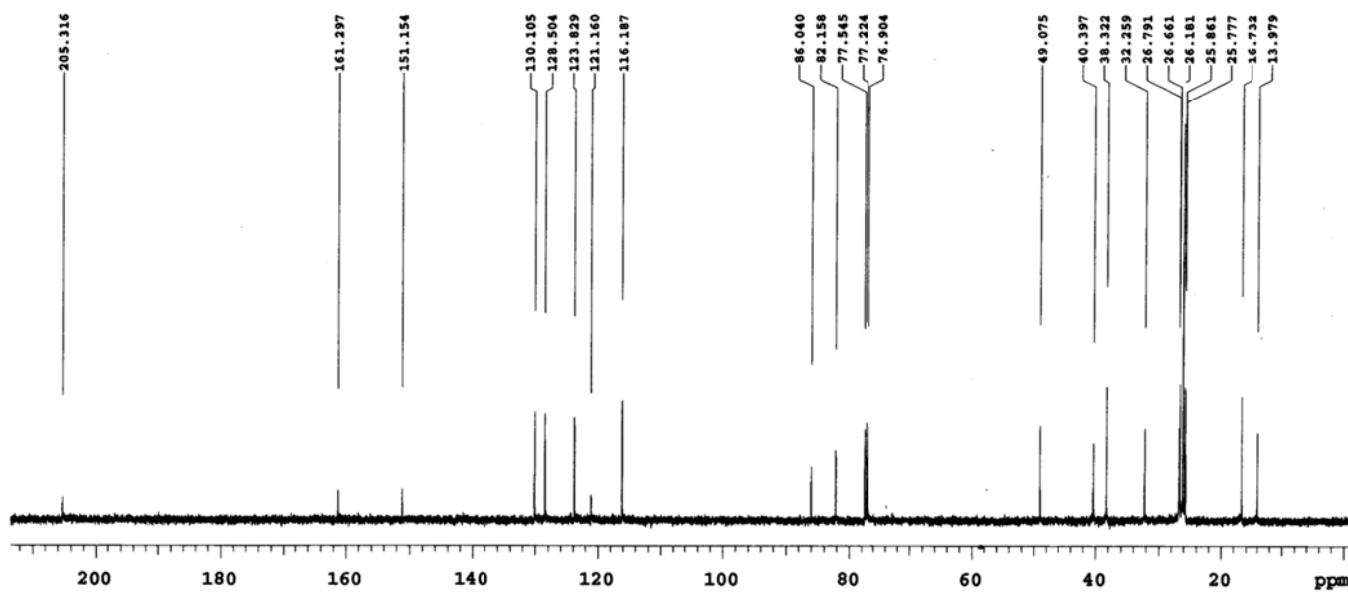


PULSE SEQUENCE	OBSERVE C13, 100.5425865 DECOUPLE H1, 399.8529994	DATA PROCESSING	AB-CORT-Cy-13C
Relax. delay 1.000 sec		Line broadening 0.5 Hz	Solvent: cdcl_3
Pulse 45.0 degrees		FT size 65536	Temp. 25.0 C / 298.1 K
Acq. time 1.304 sec	continuously on	Total time 19 minutes	Operator: chem
Width 25125.6 Hz	MALTZ-16 modulated		File: AB-CORT-Cy-13C
500 repetitions			Mercury-400 "ITIG-NMR"

3-(*tert*-Butylperoxy)-4-cyclohexyl-3-pentanoylchroman-2-one (10a'): ^1H NMR (CDCl_3 , 400 MHz)

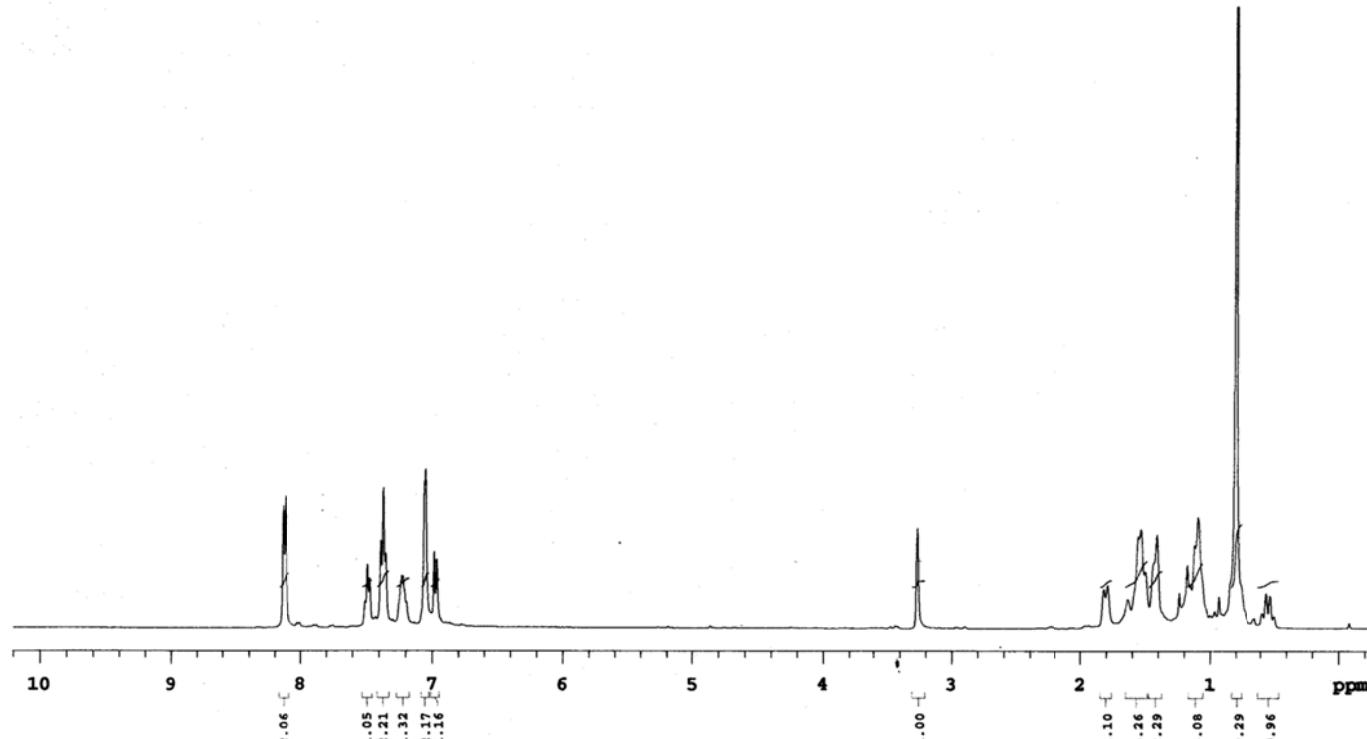


3-(*tert*-Butylperoxy)-4-cyclohexyl-3-pentanoylchroman-2-one (10a'): ^{13}C NMR (CDCl_3 , 100 MHz)



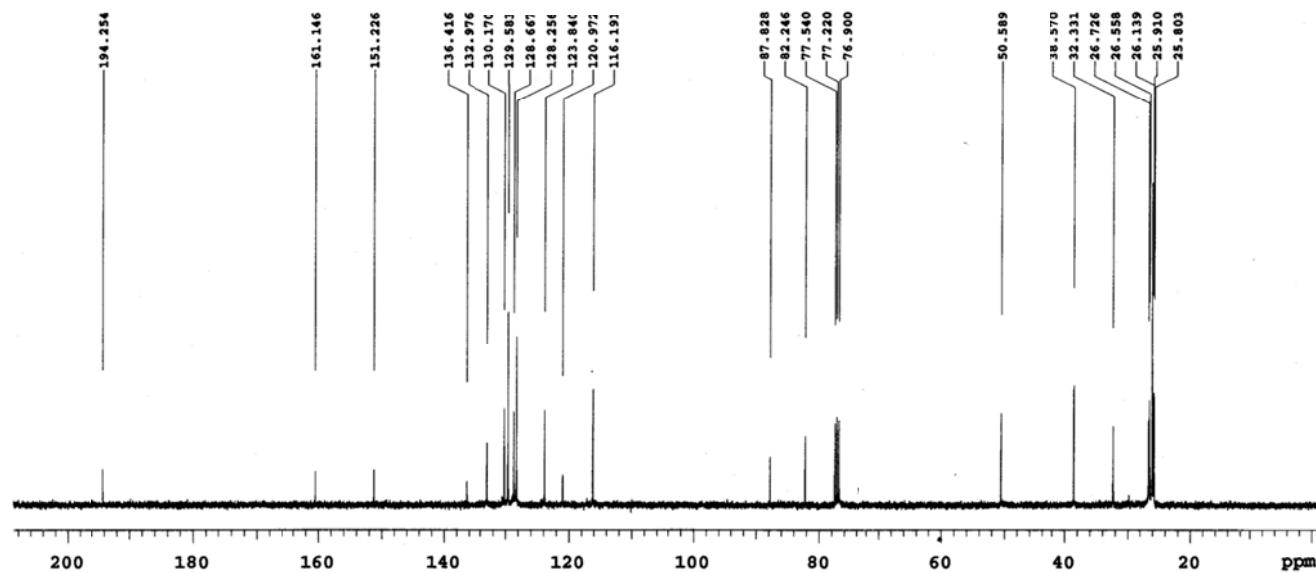
PULSE SEQUENCE	OBSERVE C13, 100.5425868 DECOUPLE H1, 399.8529994	DATA PROCESSING Line broadening 0.5 Hz FT size 65536 Total time 10 minutes	AB-COBu-Cy-13C Solvent: cdcl_3 Temp. 25.0 °C / 298.1 K Operator: chem File: AB-COBu-Cy-13C Mercury-400 "IITG-NMR"
Relax. delay 1.000 sec Pulse 45.0 degrees Acc. time 1.204 sec Width 25125.6 Hz 270 repetitions	WALTZ-16 modulated		

3-Benzoyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (11a'): ^1H NMR (CDCl_3 , 400 MHz)



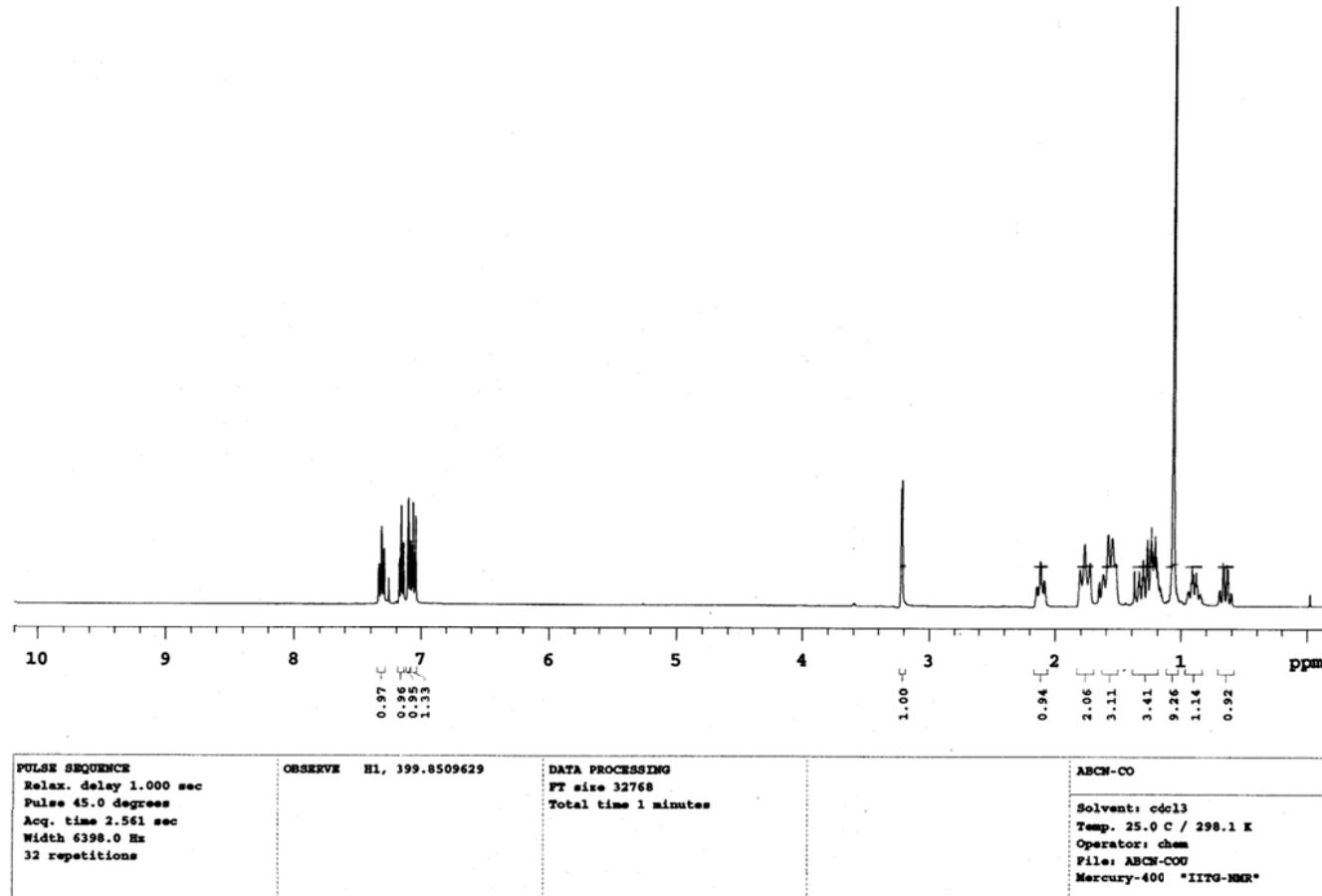
PULSE SEQUENCE	OBSERVE H1, 399.8509902	DATA PROCESSING	AB-COPh-Cy-1H
Relax. delay 1.000 sec			Solvent: cdcl_3
Pulse 45.0 degrees			Temp. 25.0 C / 298.1 K
Acq. time 2.561 sec			Operator: chem
Width 6398.0 Hz			File: AB-COPh-Cy-1H
32 repetitions		Total time 1 minutes	Mercury-400 "IITG-NMR"

3-Benzoyl-3-(*tert*-butylperoxy)-4-cyclohexylchroman-2-one (11a'): ^{13}C NMR (CDCl_3 , 100 MHz)

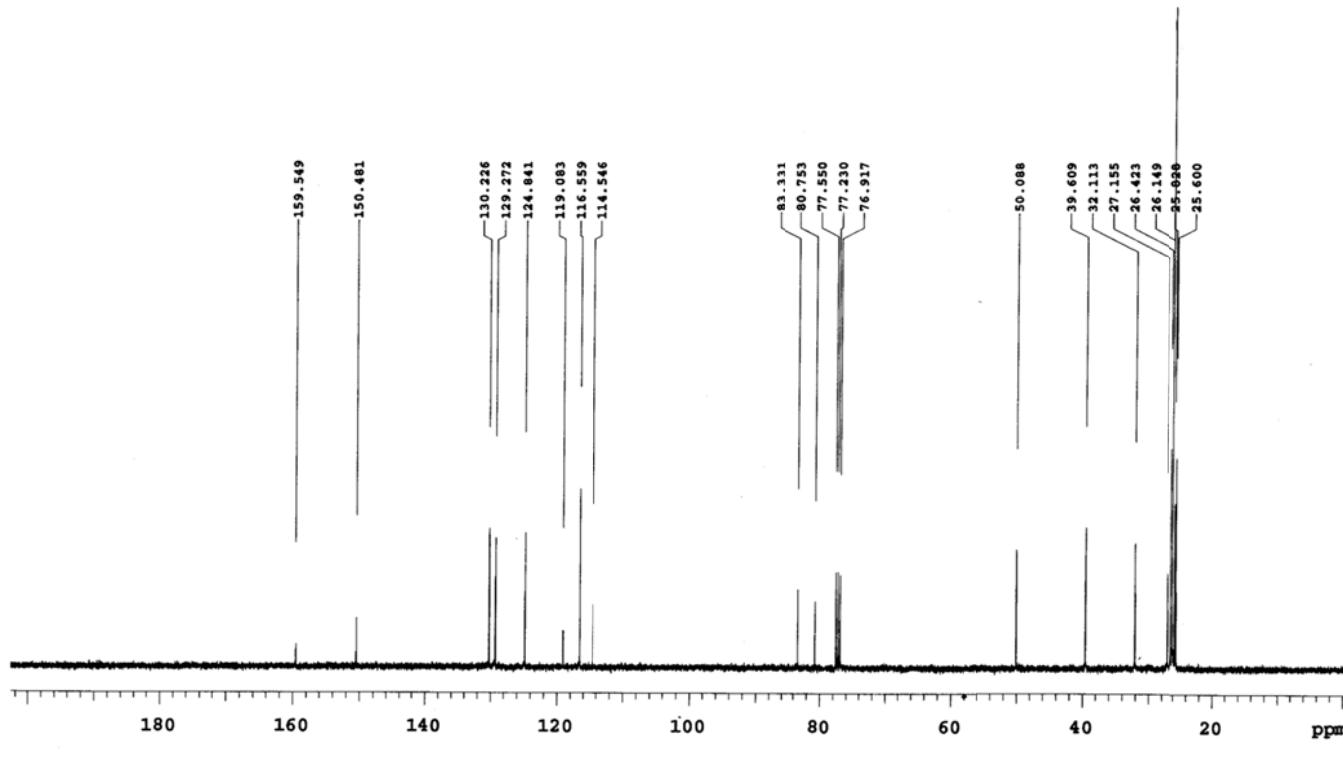


PULSE SEQUENCE	OBSERVE C13, 100.5425880 DECOUPLE H1, 399.8529994	DATA PROCESSING	AB-COPh-Cy-13C
Relax. delay 1.000 sec		Line broadening 0.5 Hz	Solvent: cdcl_3
Pulse 45.0 degrees		FT size 65536	Temp. 25.0 C / 298.1 K
Acq. time 1.304 sec	continuously on	Total time 15 minutes	Operator: chem
Width 25125.6 Hz	WALTZ-16 modulated		File: AB-COPh-Cy-13C
410 repetitions			Mercury-400 "IITG-NMR"

3-(*tert*-Butyperoxy)-4-cyclohexyl-2-oxochroman-3-carbonitrile (12a'): ^1H NMR (CDCl_3 , 400 MHz)

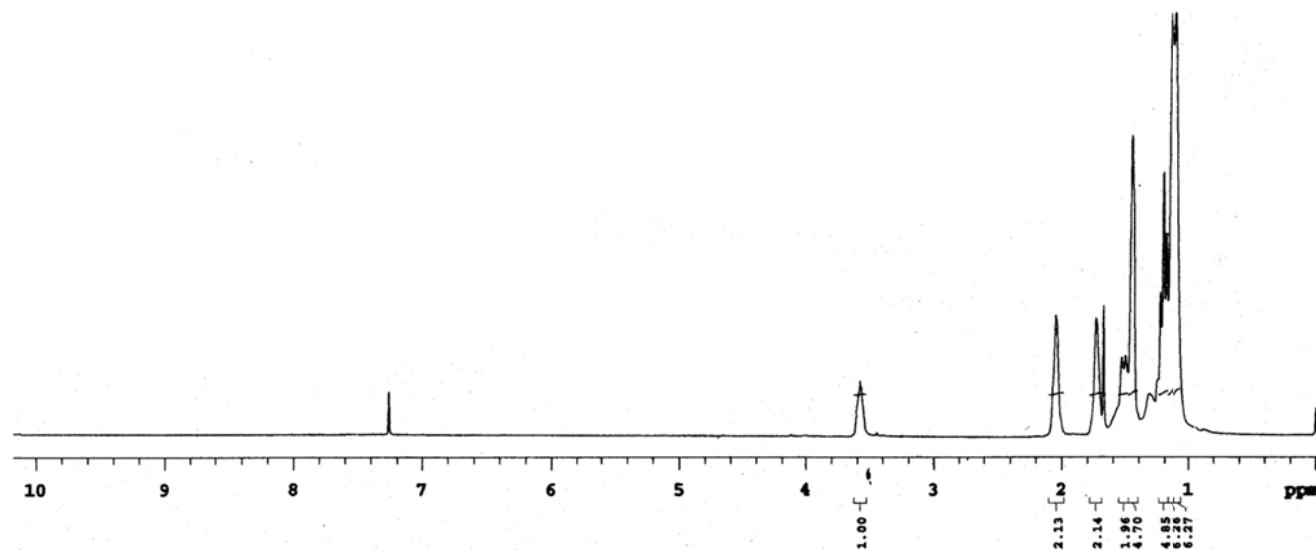


3-(*tert*-Butyperoxy)-4-cyclohexyl-2-oxochroman-3-carbonitrile (12a'): ^{13}C NMR (CDCl_3 , 100 MHz)



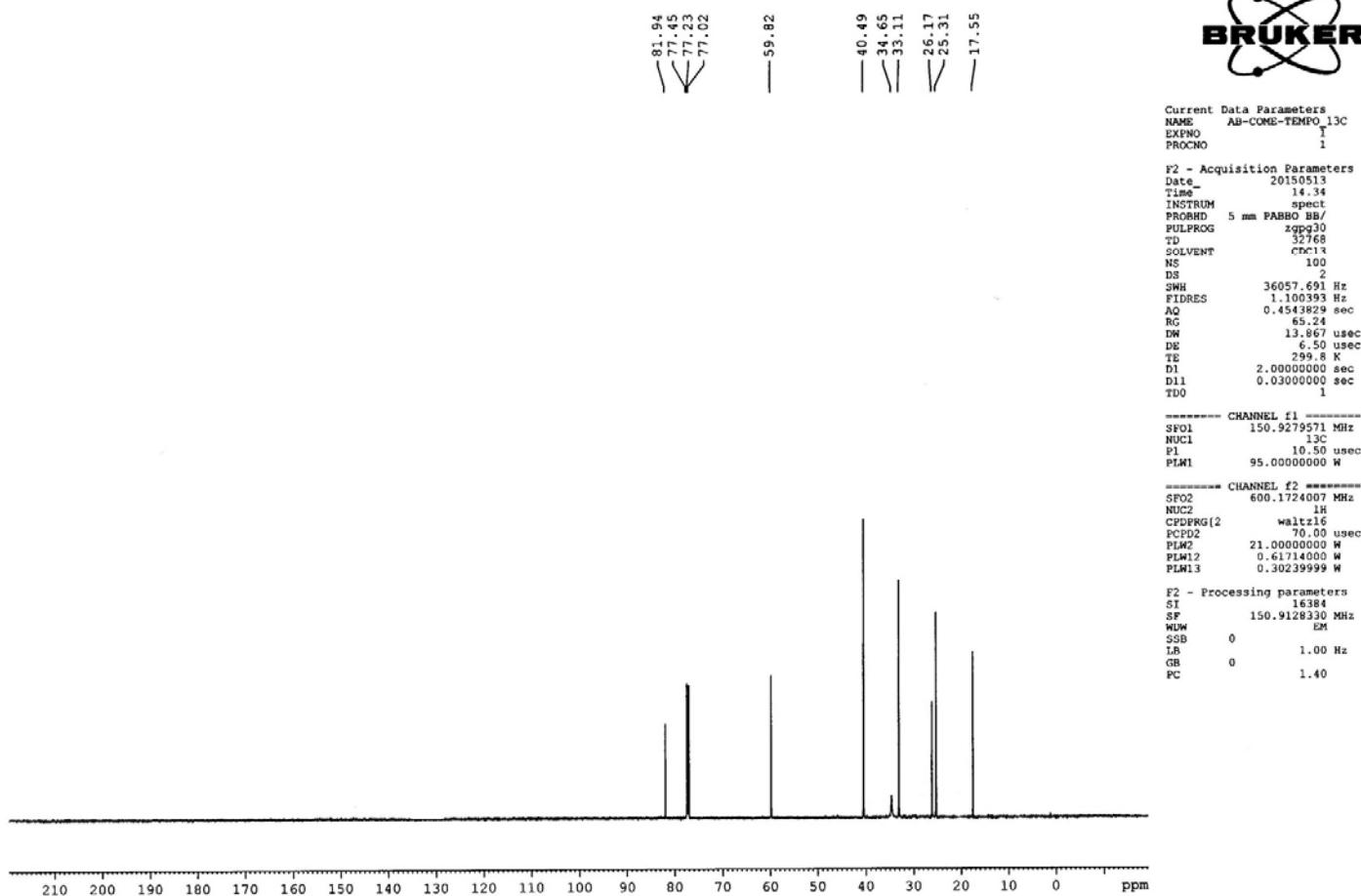
PULSE SEQUENCE	OBSERVE C13, 100.5425870 DECOUPLE H1, 399.8529994	DATA PROCESSING	AB_CH_COU_Cy_13C
Relax. delay 1.000 sec		Line broadening 0.5 Hz	
Pulse 45.0 degrees		FT size 65536	
Acq. time 1.304 sec		Total time 9 minutes	
Width 25125.6 Hz	continuously on		Solvent: cdcl_3
260 repetitions	WALTZ-16 modulated		Temp. 25.0 C / 298.1 K
			Operator: chem
			File: AB_CH_COU_Cy_13C
			Mercury-400 "IITG-NMR"

1-(Cyclohexyloxy)-2,2,6,6-tetramethylpiperidine (1A): ^1H NMR (CDCl_3 , 400 MHz)

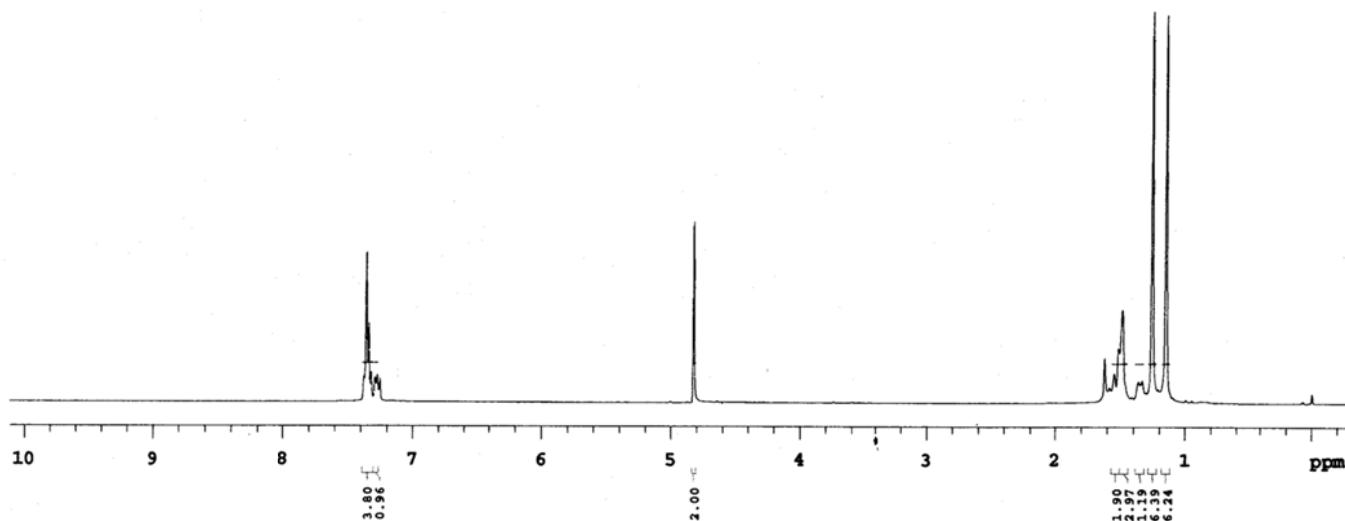


PULSE SEQUENCE	OBSERVE H1, 399.8509625	DATA PROCESSING	AB-Cy-TEMPO-1H
Relax. delay 1.000 sec Pulse 45.0 degrees Acq. time 2.561 sec Width 6398.0 Hz 36 repetitions		FT size 32768 Total time 2 minutes	Solvent: cdcl_3 Temp. 25.0 C / 298.1 K Operator: chem Mercury-400 "ITIG-MER"

1-(Cyclohexyloxy)-2,2,6,6-tetramethylpiperidine (1A): ^{13}C NMR (CDCl_3 , 150 MHz)



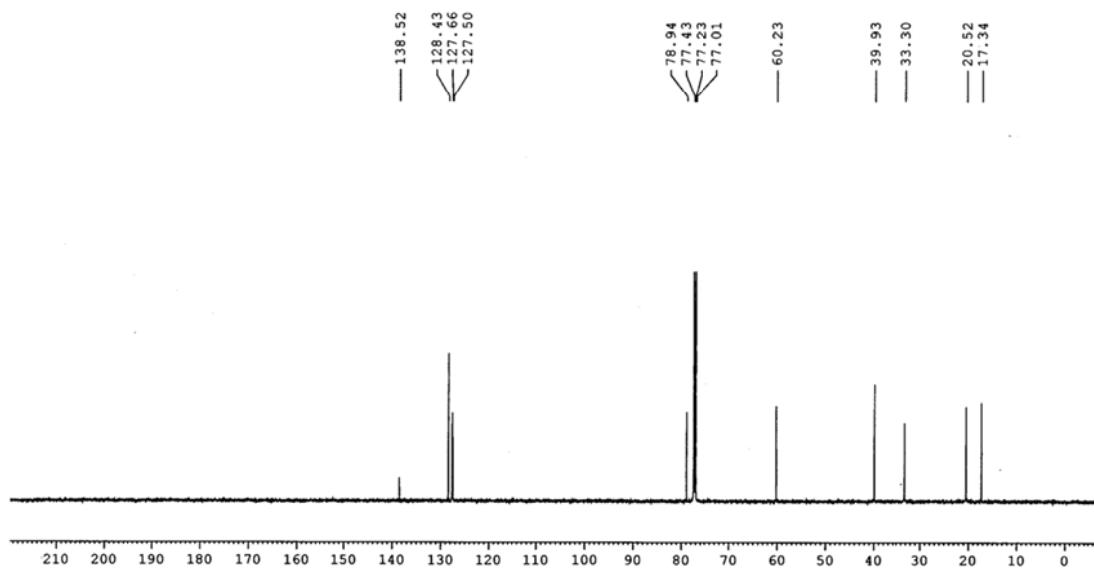
1-(Benzylxy)-2,2,6,6-tetramethylpiperidine (1B): ^1H NMR (CDCl_3 , 400 MHz)



PULSE SEQUENCE	OBSERVE H1, 399.8509672	DATA PROCESSING	AB-Tol-TEMPO-1H
Relax. delay 1.000 sec		FT size 32768	Solvent: cdcl_3
Pulse 45.0 degrees		Total time 1 minutes	Temp. 25.0 C / 298.1 K
Acq. time 2.561 sec			Operator: chem
Width 6398.0 Hz			Mercury-400 "IITG-NMR"
32 repetitions			

1-(Benzylxy)-2,2,6,6-tetramethylpiperidine (1B): ^{13}C NMR (CDCl_3 , 150 MHz)

AB-TOi-TPO_13C



Current Data Parameters
NAME AB-TOi-TPO_13C
EXPNO 1
PROCNO 1

F2 - Acquisition Parameters
Date 20150430
Time 16.12
INSTRUM spect
PROBHD 5 mm PABBO BB/
PULPROG zgpp30
TD 32768
SOLVENT CDCl3
NS 131
DS 2
SWH 36057.691 Hz
FIDRES 1.100393 Hz
AQ 0.4543829 sec
RG 65.6
DW 13.867 usec
DE 6.50 usec
TE 296.6 K
D1 2.0000000 sec
D11 0.0300000 sec
TDO 1

CHANNEL f1
SF01 154.9279571 MHz
NUC1 13C
P1 10.50 usec
PLW1 95.00000000 W

CHANNEL f2
SF02 600.1724007 MHz
NUC2 1H
CPDPGR2[2 waltz16
PCPD2 70.00 usec
PLW2 21.00000000 W
PLW12 0.61714000 W
PLW13 0.30239999 W

F2 - Processing parameters
SI 16384
SF 150.9128370 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.40