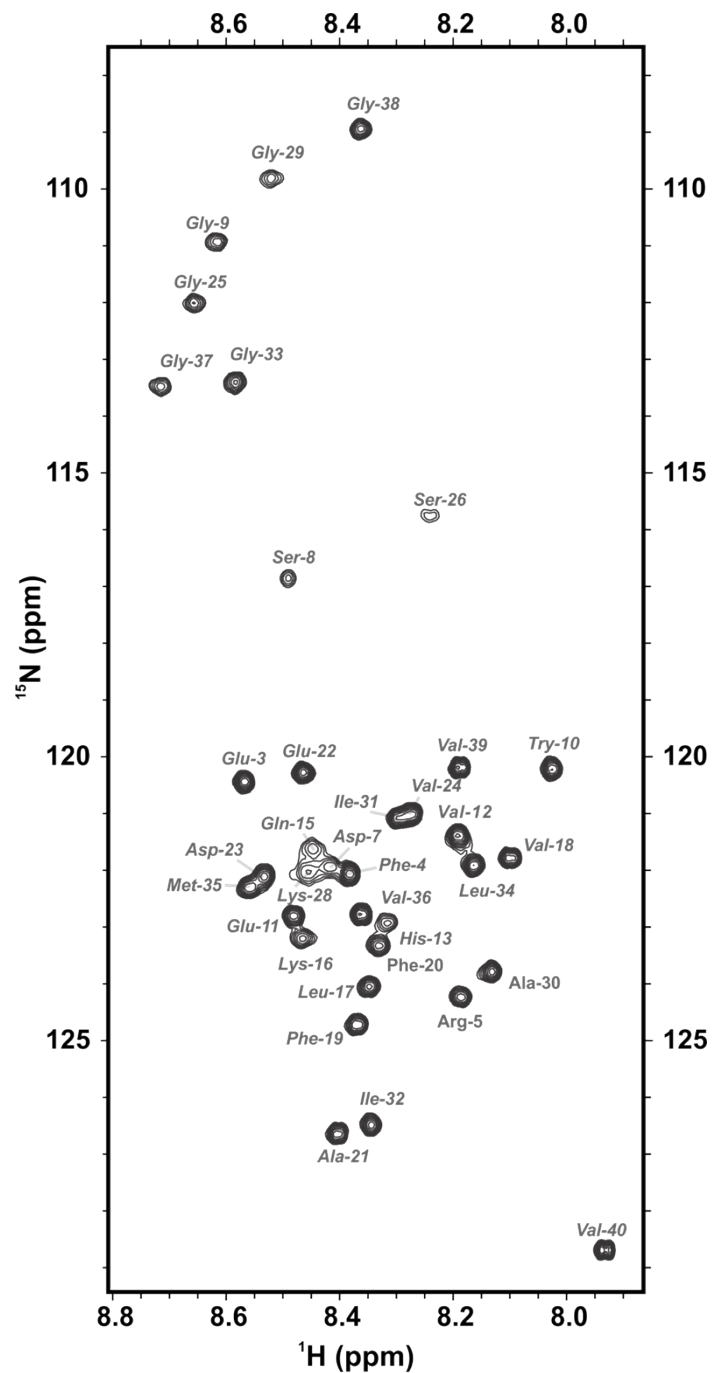


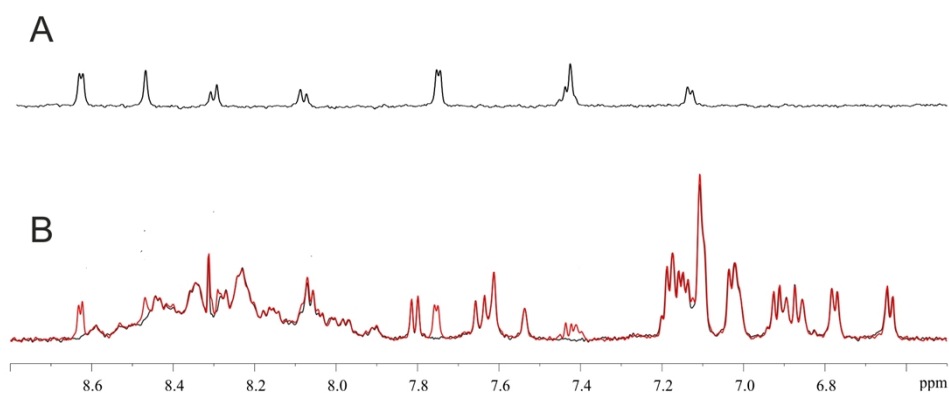
## Supplementary information

### Preparation of A $\beta$ samples

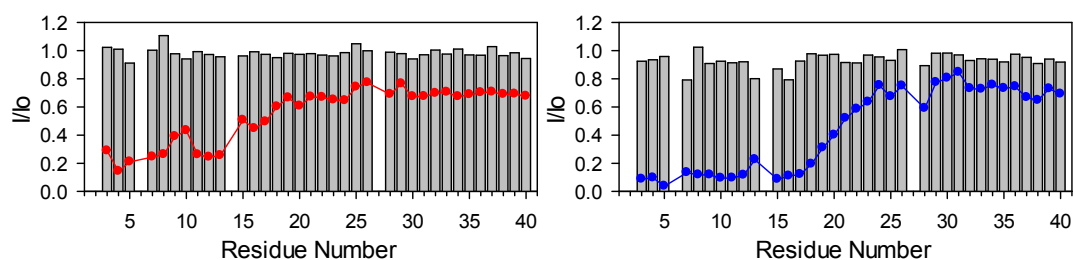
Non-labeled and  $^{15}\text{N}$  isotopically enriched A $\beta$ (1-40) samples were purchased from EZBiolab and Alexotech, respectively. Both companies dispose the samples in vials containing 1 mg of lyophilized A $\beta$ . The peptide was prepared as suggested by the suppliers and described previously, using an alkaline dissolution protocol<sup>1</sup>, which reduces the formation of pre-aggregated species during the solvation of the peptide. The lyophilized peptide was dissolved in 400  $\mu\text{L}$  of NaOH 10 mM, incubated during 30 min in ice, aliquoted, and stored at  $-80\text{ }^\circ\text{C}$ . This NaOH A $\beta$  peptide solution constituted the starting material for all subsequent structural experiments. Immediately before recording the NMR experiments, one or more A $\beta$  aliquots were diluted in TRIS 20 mM. The pH was then adjusted to 7.4 and the samples were centrifuged at 20,000  $\times g$  to eliminate potentially pre-formed aggregates. All procedures were conducted at 4  $^\circ\text{C}$ . The peptide concentration was measured by UV spectroscopy using a molar extinction coefficient  $\epsilon_{280\text{nm}} = 1490\text{ M}^{-1}\text{ cm}^{-1}$ .



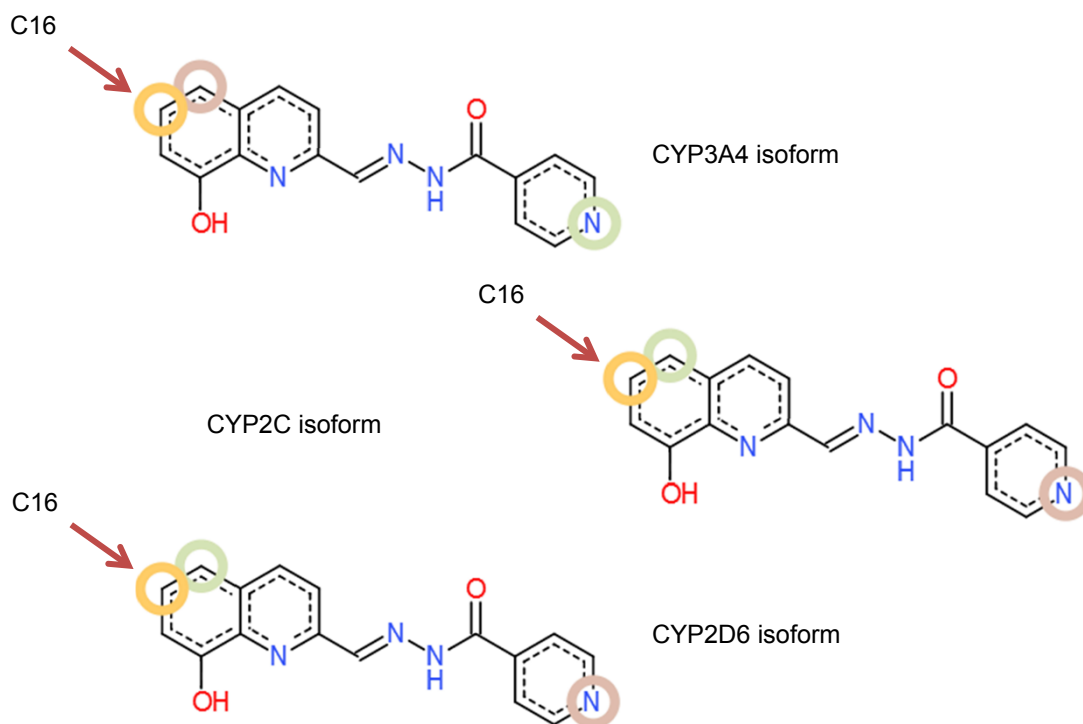
**Figure S1.** Assignment of backbone amide A $\beta$ (1-40) peptide resonances. The  $^1\text{H}$ - $^{15}\text{N}$  HSQC spectrum was acquired at 5  $^\circ\text{C}$  on 50  $\mu\text{M}$  peptide samples dissolved in TRIS buffer 20 mM, pH 7.4. Resonance assignments were based on the literature<sup>1, 2</sup> and further confirmed by 2D  $^1\text{H}$ - $^1\text{H}$  TOCSY and 2D  $^1\text{H}$ - $^1\text{H}$  NOESY experiments following standard strategies<sup>3</sup>.



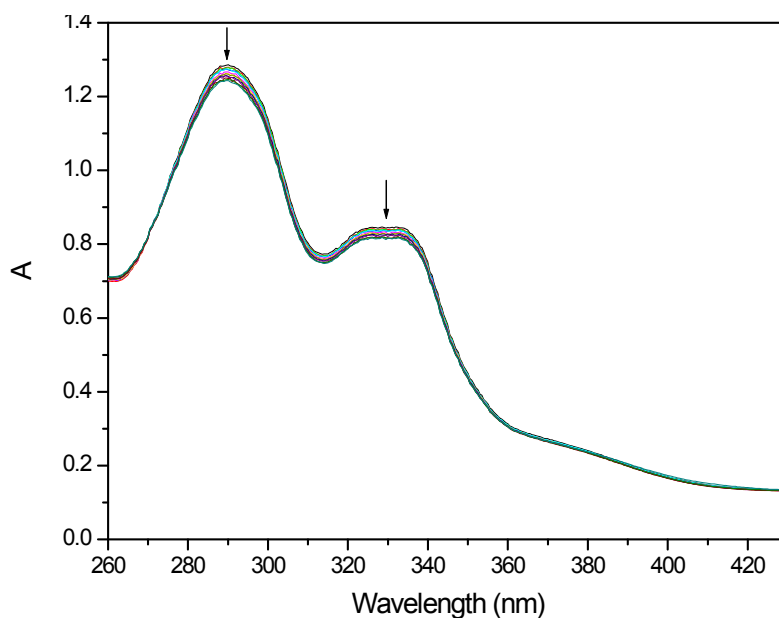
**Figure S2.** **A.** 1D  $^1\text{H-NMR}$  spectrum of 100  $\mu\text{M}$  INHHQ registered at 5  $^\circ\text{C}$  on samples dissolved in buffer TRIS 20 mM, pH 7.4. **B.** 1D  $^1\text{H-NMR}$  spectrum of 50  $\mu\text{M}$  A $\beta$  samples in the absence (black) and presence (red) of 100  $\mu\text{M}$  INHHQ in TRIS 20 mM, pH 7.4, at 5  $^\circ\text{C}$ . Addition of INHHQ did not affect A $\beta$ (1-40) peptide amide and aromatic resonances.



**Figure S3.** Effect of EDTA on the A $\beta$ -Zn $^{2+}$  (left) and A $\beta$ -Cu $^{2+}$  (right) interactions. I/I $_0$  intensity profiles of 50  $\mu\text{M}$  A $\beta$  samples measured in the presence of 1 equivalent of Zn $^{2+}$  (red circles) and Cu $^{2+}$  (blue circles) ions. The grey bars correspond to data measured after the addition of 1 equivalent of EDTA to the solutions containing the A $\beta$  metal-complexes.



**Figure S4.** INHHQ atoms most susceptible to oxidation by different isoforms of the CYP superfamily of enzymes, circled, as calculated by the SMARTCyp V. 2.4.2 software package. Yellow circles show the sites with highest oxidizing probability, followed by salmon and, finally, green circles. C16 is the most reactive site for all the studied isoforms.



**Figure S5.** UV-Vis spectra of a  $3.0 \times 10^{-7}$  M INHHQ solution in 10% DMSO/saline solution vehicle. Readings were taken every hour, for 12 h. The arrows stand for decreases in the intensity of the two absorption bands over time, related to the partial hydrolysis of INHHQ.

### Additional references

1. L. M. Hou, H. Y. Shao, Y. B. Zhang, H. Li, N. K. Menon, E. B. Neuhaus, J. M. Brewer, I. J. L. Byeon, D. G. Ray, M. P. Vitek, T. Iwashita, R. A. Makula, A. B. Przybyla, M. G. Zagorski, Solution NMR studies of the A beta(1-40) and A beta(1-42) peptides establish that the met35 oxidation state affects the mechanism of amyloid formation. *Journal of the American Chemical Society* 2004, *126*. 1992-2005.
2. J. Danielsson, J. Jarvet, P. Damberg, A. Graslund, Two-site binding of beta-cyclodextrin to the Alzheimer Abeta(1-40) peptide measured with combined PFG-NMR diffusion and induced chemical shifts. *Biochemistry* 2004, *43*. 6261-9; J. Danielsson, R. Pierattelli, L. Banci, A. Gräslund, High-resolution NMR studies of the zinc-binding site of the Alzheimer's amyloid b-peptide. *FEBS Journal* 2007, *274*. 46-59; N. L. Fawzi, J. Ying, D. A. Torchia, G. M. Clore, Kinetics of amyloid beta monomer-to-oligomer exchange by NMR relaxation. *J Am Chem Soc* 2010, *132*. 9948-51; C. D. Syme, J. H. Viles, Solution <sup>1</sup>H NMR investigation of Zn<sup>2+</sup> and Cd<sup>2+</sup> binding to amyloid-beta peptide (Abeta) of Alzheimer's disease. *Biochimica et biophysica acta* 2006, *1764*. 246-56; F. Bousejra-ElGarah, C. Bijani, Y. Coppel, P. Faller, C. Hureau, Iron(II) binding to amyloid-beta, the Alzheimer's peptide. *Inorganic chemistry* 2011, *50*. 9024-30; S. Sinha, Z. Du, P. Maiti, F. G. Klarner, T. Schrader, C. Wang, G. Bitan, Comparison of three amyloid assembly inhibitors: the sugar scyllo-inositol, the polyphenol epigallocatechin gallate, and the molecular tweezer CLR01. *ACS chemical neuroscience* 2012, *3*. 451-8.
3. K. Wüthrich, *NMR of proteins and nucleic acids*. Wiley: New York, 1986; p 292.