

## Supporting Information

### Chemical Synthesis of the Tumor-Associated Globo H Antigen

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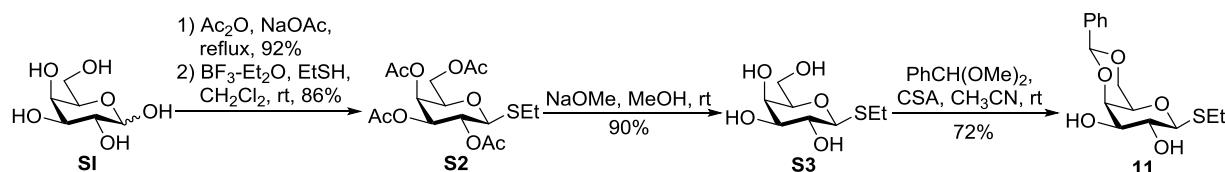
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### Supplementary Experimental Section

**General Methods.** Chemicals and materials were obtained from commercial sources and were used as received without further purification unless otherwise noted. 4Å molecular sieves (MS) were flame-dried under high vacuum and used immediately after cooling under a N<sub>2</sub> atmosphere. Analytical TLC was carried out on silica gel 60Å F<sub>254</sub> plates with detection by a UV detector and/or by charring with 15% (v/v) H<sub>2</sub>SO<sub>4</sub> in EtOH. NMR spectra were recorded on a 400, 500, or 600 MHz machine with chemical shifts reported in ppm ( $\delta$ ) downfield from tetramethylsilane (TMS) that was used as an internal reference.

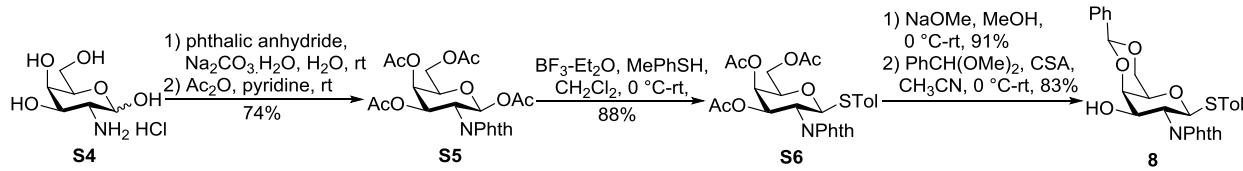


**Ethyl 2,3,4,6-tetra-O-Acetyl-1-thio-β-D-galactopyranoside S2.**<sup>[1]</sup> After a mixture of D-galactose **S1** (10.0 g, 55.5 mmol), anhydrous NaOAc (5.9 g, 72.2 mmol) and Ac<sub>2</sub>O (50 ml) was stirred at 80 °C for 4 h, with the reaction progress monitored by TLC, it was cooled to rt and then poured into cold saturated aq. NaHCO<sub>3</sub> solution. The mixture was stirred at 0 °C for 1.5 h and extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was separated and dried over Na<sub>2</sub>SO<sub>4</sub>. The solvent was removed

in vacuum and the residue was purified by silica gel column chromatography (AcOEt/petroleum ether. 1:4, v/v) to give the peracetylated galactose (19.8 g, 92%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ: 5.69 (d, *J* = 8.1 Hz, 1 H, H-1), 5.42 (d, *J* = 3.2 Hz, 1 H, H-4), 5.33 (t, *J* = 8.1 Hz, 1 H, H-2), 5.07 (dd, *J* = 10.5, 4.0 Hz, 1 H, H-3), 4.19-4.08 (m, 2 H, H-6, H-6'), 4.07-4.02 (m, 1 H, H-5), 2.10 (s, 3 H, OAc), 2.06 (s, 6 H, OAc), 2.01 (s, 3 H, OAc), 1.97 (s, 3 H, OAc). The product (19.0 g, 48.7 mmol) and ethanethiol (5.41 mL, 73.0 mmol) were dissolved in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (120 mL) at 0 °C, and then BF<sub>3</sub>·Et<sub>2</sub>O (12.0 mL, 97.4 mmol) was added dropwise. The solution was stirred at 0 °C until when TLC showed the completion of the reaction. The reaction mixture was washed with saturated aq. NaHCO<sub>3</sub> solution and brine and dried over Na<sub>2</sub>SO<sub>4</sub>, and the organic phase was concentrated under reduced pressure. The residue was purified by flash silica gel column chromatography (AcOEt/petroleum ether. 1:4, v/v) to give **S2** (16.4 g, 86%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 5.39 (d, *J* = 3.2 Hz, 1 H, H-4), 5.19 (t, *J* = 9.7 Hz, 1 H, H-2), 5.01 (dd, *J* = 9.7, 3.2 Hz, 1 H, H-3), 4.46 (d, *J* = 11.3 Hz, 1 H, H-1), 4.16-4.03 (m, 2 H, H-6, H-6'), 3.90 (t, *J* = 6.5 Hz, 1 H, H-5), 2.77-2.60 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 2.11 (s, 3 H, OAc), 2.03 (s, 3 H, OAc), 2.03 (s, 3 H, OAc), 1.94 (s, 3 H, OAc), 1.25 (t, *J* = 6.5 Hz, 3 H, SCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 170.4, 170.2, 170.0, 169.5, 84.0, 74.3, 71.6, 67.2, 67.1, 61.4, 24.3, 20.8, 20.6, 20.5, 20.4, 14.8.

**Ethyl 1-Thio-β-D-galactopyranoside S3.**<sup>[2]</sup> After **S2** (16.0 g, 40.8 mmol) was dissolved in MeOH (70 mL), a CH<sub>3</sub>ONa/CH<sub>3</sub>OH solution (0.4 M) was added until pH reached 9. The reaction was stirred at rt for 3 h. When TLC showed the completion of reaction, the solution was neutralized with Amberlyst (H<sup>+</sup>) resin to pH 6-7, and then concentrated in vacuum. The residue was purified by flash column chromatography (MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 1:5, v/v) to afford **S3** (8.2 g, 90%) as a white solid. <sup>1</sup>H NMR (400 MHz, CD<sub>3</sub>OD): δ 4.32 (d, *J* = 9.7 Hz, 1 H, H-1), 3.88 (d, *J* = 3.2 Hz, 1 H, H-4), 3.77-3.64 (m, 2 H, H-6, H-6'), 3.52 (t, *J* = 8.1 Hz, 1 H, H-2), 3.46 (dd, *J* = 9.7, 3.2 Hz, 1 H, H-3), 3.33-3.29 (m, 1 H, H-5), 2.83-2.65 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 1.28 (t, *J* = 6.5 Hz, 3 H SCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CD<sub>3</sub>OD): δ 86.0, 79.2, 74.8, 69.9, 69.1, 61.2, 23.5, 14.1.

**Ethyl 4:6-O-Phenylmethylen-1-thio-β-D-galactopyranoside 11.**<sup>[3]</sup> To the solution of **S3** (8.0 g, 35.7 mmol) in anhydrous CH<sub>3</sub>CN (30 mL) were added benzaldehyde dimethyl acetal (6.45 mL, 42.8 mmol) and CSA (2.0 g, 8.9 mmol) at rt, and the reaction was stirred at rt with occasional vacuum application until TLC showed that the reaction was completed. Then the reaction was quenched with Et<sub>3</sub>N (2.47 mL, 17.8 mmol) and diluted with CH<sub>2</sub>Cl<sub>2</sub> (60 mL). The solution was washed with brine, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuum. The residue was purified by flash column chromatography (MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 1:10, v/v) to give **11** (8.02 g, 72%) as a white solid. <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.56-7.49 (m, 2 H, ArH), 7.42-7.38 (m, 3 H, ArH), 5.53 (s, 1 H, CHPh), 4.34 (d, *J* = 8.8 Hz, 1 H, H-1), 4.33 (d, *J* = 12.8 Hz, 1 H, H-6), 4.25 (d, *J* = 3.2 Hz, 1 H, H-4), 4.03 (d, *J* = 12.8 Hz, 1 H, H-6'), 3.81 (t, *J* = 8.8 Hz, 1 H, H-2), 3.68 (dd, *J* = 10.0, 4.4 Hz, 1 H, H-3), 3.53-3.50 (m, 1 H, H-5), 2.88-2.69 (m, 2 H, SCH<sub>2</sub>CH<sub>3</sub>), 2.51 (bs, 2 -OH), 1.34 (t, *J* = 6.5 Hz, 3 H, SCH<sub>2</sub>CH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 137.5, 129.3, 128.3, 126.4, 101.5, 85.3, 75.6, 73.9, 70.1, 69.7, 69.3, 23.5, 14.1.

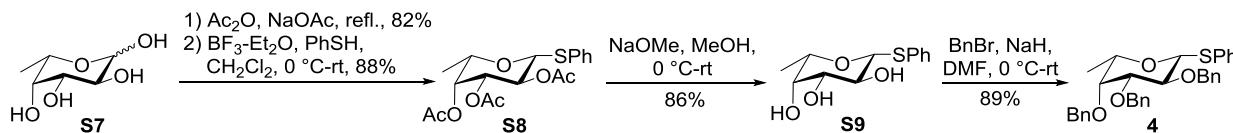


**2-Deoxy-2-phthalimido- $\beta$ -D-galactopyranose tetraacetate S5.**<sup>[4]</sup> After the suspension of D-galactosamine hydrochloride **S4** (5.0 g, 23.3 mmol),  $\text{Na}_2\text{CO}_3 \cdot \text{H}_2\text{O}$  (2.5 g 23.3 mmol) and phthalic anhydride (3.5 g, 23.3 mmol) in  $\text{H}_2\text{O}$  (50 ml) was stirred at rt for 3 h, the resultant clear solution (pH 3.0; initial pH 8.5) was cooled to -78 °C and lyophilized to offer a pale yellow solid residue (9.16 g). This residue was suspended in a mixture of pyridine (200 ml) and acetic anhydride (100 ml) at 0 °C and was stirred for 20 h. Acetic anhydride and pyridine were evaporated in vacuum, and the residue was co-evaporated with toluene (10 ml) twice, and finally subjected to silica gel chromatography ( $\text{EtOAc/toluene}$  1:9, v/v) to obtain **S5** (8.18 g, 74%) as a white solid along.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.90-7.80 (m, 2 H, ArH), 7.78-7.71 (m, 2 H, ArH), 6.52 (dd,  $J$  = 11.3, 3.2 Hz, 1 H, H-3), 6.34 (d,  $J$  = 6.8 Hz, 1 H, H-1), 5.66 (d,  $J$  = 3.2 Hz, 1 H, H-4), 4.90 (dd,  $J$  = 11.3, 3.2 Hz, H-2), 4.49 (t,  $J$  = 8.1 Hz, 1 H, H-5), 4.17-4.11 (m, 2 H, H-6, H-6'), 2.18 (s, 3 H, OAc), 2.05 (s, 6 H, 2×OAc), 1.88 (s, 3 H, OAc);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  169.9, 169.83, 169.81, 169.29, 167.5, 166.8, 134.0, 133.9, 129.2, 123.3, 123.2, 83.9, 74.1, 68.4, 66.5, 61.2, 49.7, 20.7, 20.3, 20.2, 20.1.

**para-Tolyl 3,4,6-tri-O-Acetyl-2-deoxy-2-phthalimido-1-thio- $\beta$ -D-galactopyranoside S6.**<sup>[5]</sup> To the stirred solution of **S5** (5.0 g, 10.4 mmol) and *p*-toluenethiol (1.95 g, 15.7 mmol) in anhydrous  $\text{CH}_2\text{Cl}_2$  (30 mL) at 0 °C was added  $\text{BF}_3 \cdot \text{Et}_2\text{O}$  (1.6 mL, 12.6 mmol) dropwise. When TLC showed the completion of reaction, the reaction mixture was washed with saturated aq.  $\text{NaHCO}_3$  solution and brine, dried over anhydrous  $\text{Na}_2\text{SO}_4$ , and concentrated in vacuum. The residue was purified by flash column chromatography ( $\text{EtOAc/hexane}$  1:4, v/v) to give **S6** (4.96 g, 88%) as a white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.89-7.80 (m, 2 H, ArH), 7.79-7.70 (m, 2 H, ArH), 7.33-7.26 (m, 2 H, ArH), 7.09-7.01 (m, 2 H, ArH), 5.79 (dd,  $J$  = 11.3, 3.2 Hz, 1 H, H-3), 5.63 (d,  $J$  = 9.7 Hz, 1 H, H-1), 5.47 (d,  $J$  = 3.2 Hz, 1 H, H-4), 4.49 (t,  $J$  = 11.3 Hz, 1 H, H-2), 4.25-4.18 (m, 1 H, H-6), 4.17-4.1 (m, 1 H, H-6'), 4.10-4.05 (m, 1 H, H-5), 2.29 (s, 3 H,  $\text{SPhCH}_3$ ), 2.16 (s, 3 H, OAc), 2.03 (s, 6 H, 2×OAc), 1.81 (s, 3 H, OAc);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ):  $\delta$  170.3, 170.2, 169.7, 167.9, 167.2, 138.4, 134.4, 134.3, 133.4, 133.2, 131.5, 131.2, 129.7, 129.6, 128.0, 123.7, 123.6, 84.3, 74.5, 68.8, 66.9, 61.5, 50.1, 20.1, 20.7, 20.6, 20.5.

**para-Tolyl 2-Deoxy-4:6-O-phenylmethylene-2-phthalimido-1-thio- $\beta$ -D-galactopyranoside 8.** To a solution of **S6** (4.9 g, 9.1 mmol) in  $\text{MeOH}$  (25 mL) was added a  $\text{CH}_3\text{ONa}/\text{CH}_3\text{OH}$  solution (0.4 M) until pH reached 9.5. The reaction mixture was stirred at rt 4 h. When TLC showed the completion of reaction, it was neutralized to pH 6-7 with Amberlyst ( $\text{H}^+$ ) resin and concentrated in vacuum. The residue was purified by flash column chromatography ( $\text{MeOH}/\text{CH}_2\text{Cl}_2$ , 1:8, v/v) to give a solid compound (3.43 g, 91%), which is directly used in next reaction. After the product was dissolved in anhydrous  $\text{CH}_3\text{CN}$  (40 mL), benzaldehyde dimethyl acetal (1.5 mL, 9.8 mmol) and CSA (0.48 g, 2.05 mmol) were added. The reaction was stirred at rt with occasional vacuum

application until TLC showed the completion reaction. It was quenched with Et<sub>3</sub>N (0.57 mL, 4.1 mmol) and diluted with CH<sub>2</sub>Cl<sub>2</sub> (30 mL). The solution was then washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated in vacuum. The residue was purified by flash column chromatography (MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 1:13, v/v) to give **8** (3.4 g, 83%) as a light yellow solid. [α]<sub>D</sub><sup>25</sup> = -28.72° (c 3.33, CHCl<sub>3</sub>). <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>): δ 7.87-7.81 (m, 2 H, ArH), 7.75-7.71 (m, 2 H, ArH), 7.50-7.46 (m, 2 H, ArH), 7.45-7.38 (m, 5 H, ArH), 7.08 (d, J = 8.1 Hz, 1 H, ArH), 5.62 (d, J = 9.7 Hz, 1 H, H-1), 5.56 (s, 1 H, CHPh), 4.49 (dd, J = 9.7, 3.2 Hz, 1 H, H-3), 4.43 (t, J = 9.7 Hz, 1 H, H-2), 4.41 (d, J = 12.9 Hz, 1 H, H-6), 4.28 (d, J = 3.2 Hz, 1 H, H-4), 4.06 (d, J = 12.9 Hz, 1 H, H-6'), 3.68 (s, 1 H, H-5), 2.41 (bs, 1 H, -OH), 2.35 (s, 3 H, SPhCH<sub>3</sub>); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>): δ 168.6, 167.8, 138.3, 137.4, 134.1, 131.9, 131.7, 129.5, 129.4, 128.3, 127.3, 126.7, 123.6, 123.2, 101.6, 82.7, 75.0, 70.1, 69.4, 68.7, 53.2, 21.3; HRMS (ESI TOF): calcd. for C<sub>28</sub>H<sub>25</sub>NNaO<sub>6</sub> [M+Na]<sup>+</sup> m/z, 526.1300; found, 526.1302.

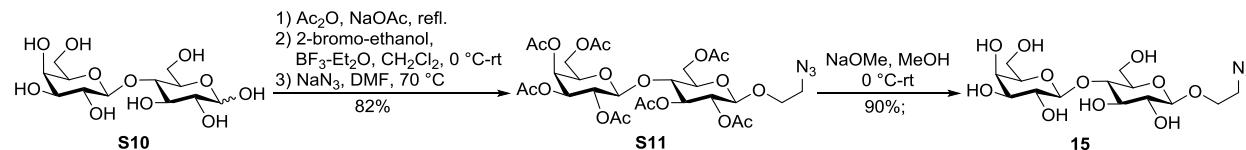


**Phenyl 2,3,6-tri-O-Acetyl-β-L-fucopyranoside S8.**<sup>[6]</sup> A mixture of L-fucose (4.0 g, 24.36 mmol) anhydrous NaOAc (2.6 g, 31.37 mmol) and Ac<sub>2</sub>O (30 ml) was stirred at 70 °C for 8 h with the reaction progress monitored by TLC. After the reaction was completed, it was cooled to rt and then poured into cold saturated aq. NaHCO<sub>3</sub> solution. After the mixture was stirred for 1.5 h, it was extracted with CH<sub>2</sub>Cl<sub>2</sub>, and the organic phase was dried over Na<sub>2</sub>SO<sub>4</sub>. After the solvent was removed in vacuum, the residue was purified by flash silica gel column chromatography (AcOEt/toluene, 1:4, v/v) to give the peracetylated fucose (6.63 g, 81.9%) as sticky syrup. This product (1.3 g, 3.9 mmol) and benzentethiol (1.6 mL, 15.6 mmol) were dissolved in anhydrous CH<sub>2</sub>Cl<sub>2</sub> (15 mL), and to this solution was then added BF<sub>3</sub>·Et<sub>2</sub>O (0.73 mL, 5.87 mmol) at 0 °C. When TLC showed the completion of reaction, it was quenched and washed with saturated aq. NaHCO<sub>3</sub> solution and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and then concentrated. The residue was purified by flash column chromatography (EtOAc/Hexane 1:5, v/v) to afford **S8** (1.38 g, 88%) as a white solid. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.53-7.48 (m, 2 H, ArH), 7.34-7.26 (m, 3 H, ArH), 5.26 (d, J = 3.0 Hz, 1 H, H-4), 5.22 (t, J = 9.8 Hz, 1 H, H-2), 5.05 (dd, J = 9.8, 3.0 Hz, 1 H, H-3), 4.70 (d, J = 10.4 Hz, 1 H, H-1), 3.83 (q, J = 6.7 Hz, 1 H, H-5), 2.13 (s, 3 H, OAc), 2.07 (s, 3 H, OAc), 1.96 (s, 3 H, OAc), 1.23 (d, J = 6.7 Hz, 3 H, H-6); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>): δ 170.8, 170.1, 169.5, 132.9, 132.3, 128.9, 127.9, 86.4, 73.1, 72.4, 70.3, 67.3, 20.9, 20.7, 20.6, 16.5.

**Phenyl β-L-Fucopyranoside S9.**<sup>[7]</sup> To the solution of **S8** (1.35 g, 3.37 mmol) dissolved in MeOH (10 mL) was added a CH<sub>3</sub>ONa/CH<sub>3</sub>OH solution (0.4 M) until pH reached 9.5. The solution was stirred at rt for 2 h. When TLC showed the completion reaction, it was neutralized to pH 6-7 with Amberlyst (H<sup>+</sup>) resin and then concentrated in vacuum. The residue was finally purified by flash column chromatography (MeOH/CH<sub>2</sub>Cl<sub>2</sub>, 1:9, v/v) to give **S9** (0.73 g, 86%) as a yellowish solid. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.54-7.48 (m, 2 H, ArH), 7.34-7.26 (m, 3 H, ArH), 4.56 (d, J =

9.8 Hz, 1 H, H-1), 4.04 (bs, 3 -OH), 3.82 (d,  $J$  = 2.9 Hz, 1 H, H-3), 3.77 (t,  $J$  = 9.8 Hz, 1 H, H-2), 3.66 (dd,  $J$  = 9.8, 2.9 Hz, 1 H, H-4), 3.59 (q,  $J$  = 6.8 Hz, 1 H, H-5), 1.30 (d,  $J$  = 6.8 Hz, 3 H, H-6);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  133.4, 131.84, 131.8, 128.9, 127.5, 88.6, 75.2, 74.7, 71.7, 70.0, 16.7.

**Phenyl 2,3,6-tri-O-Phenylmethyl- $\beta$ -L-fucopyranoside 4.**<sup>[6]</sup> After a mixture of **S9** (0.70 g, 2.73 mmol) and NaH (230 mg, 9.6 mmol) in DMF (10 mL) was stirred at 0 °C for 45 min, BnBr (1.62 mL, 13.67 mmol) was added at 0 °C. Thereafter, the mixture was stirred for another 6 h. When TLC showed the completion of reaction, it was quenched with  $\text{H}_2\text{O}$  at 0 °C, and the mixture was diluted with EtOAc. The aq. layer was washed with EtOAc (5 × 5 mL) followed by drying over  $\text{Na}_2\text{SO}_4$ . The solvent was evaporated under reduced pressure, and the crude product was purified by flash column chromatography (acetone/hexane 1:10, v/v) to give **4** (1.28 g, 89%) as colorless syrup.  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.66-7.62 (m, 2 H, ArH), 7.46-7.31 (m, 15 H, ArH), 7.28-7.23 (m, 3 H, ArH), 5.06 (d,  $J$  = 11.6 Hz, 1 H,  $\text{CH}_2\text{Ph}$ ), 4.85 (d,  $J$  = 10.4 Hz, 1 H,  $\text{CH}_2\text{Ph}$ ), 4.79 (s, 2 H,  $\text{CH}_2\text{Ph}$ ), 4.78 (d,  $J$  = 10.4 Hz, 1 H,  $\text{CH}_2\text{Ph}$ ), 4.73 (d,  $J$  = 11.6 Hz, 1 H,  $\text{CH}_2\text{Ph}$ ), 4.66 (d,  $J$  = 9.8 Hz, 1 H, H-1), 3.99 (t,  $J$  = 9.8 Hz, 1 H, H-2), 3.69 (d,  $J$  = 3.1 Hz, 1 H, H-3), 3.65 (dd,  $J$  = 9.2, 3.1 Hz, 1 H, H-4), 3.60-3.56 (m, 1 H, H-5), 1.32 (d,  $J$  = 6.1 Hz, 3 H, H-6);  $^{13}\text{C}$  NMR (125 MHz,  $\text{CDCl}_3$ ):  $\delta$  138.8, 138.5, 138.4, 134.4, 131.6, 128.8, 128.5, 128.4, 128.3, 128.0, 127.8, 127.7, 127.6, 127.0, 87.6, 84.6, 77.4, 77.2, 76.7, 75.6, 74.7, 72.9, 17.4; HRMS (ESI TOF): calcd. for  $\text{C}_{33}\text{H}_{34}\text{NaO}_4\text{S} [\text{M}+\text{Na}]^+$   $m/z$ , 549.2076; found, 549.2063.



**2-Azidoethyl 2,3,4,6-tetra-O-Acetyl- $\beta$ -D-galactopyranosyl-(1→4)-2,3,6-tri- $\beta$ -D-glucopyranoside S11.**<sup>[8]</sup> Lactose **S10** was converted into **S11** according to a literature procedure in three steps in an 82% (8.01 g) overall yield.

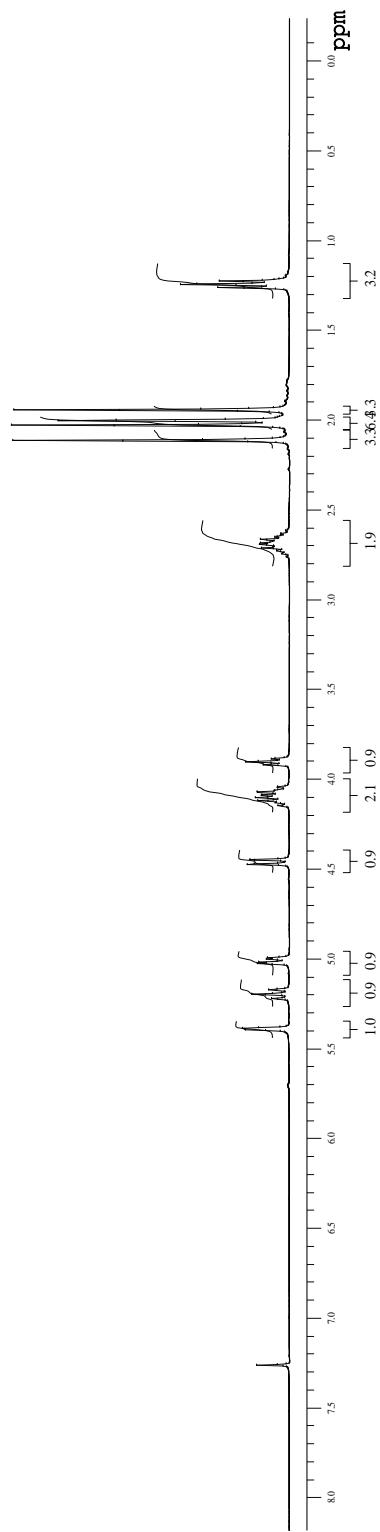
**2-Azidoethyl  $\beta$ -D-Galactopyranosyl-(1→4)- $\beta$ -D-glucopyranoside 15.**<sup>[9]</sup> To the solution of **S11** (8.0 g, 11.33 mmol) dissolved in MeOH (60 mL) was added a  $\text{CH}_3\text{ONa}/\text{CH}_3\text{OH}$  solution (0.4 M) until pH reached 10. The solution was stirred at rt for 12 h. When TLC showed the completion reaction, it was neutralized to pH 6-7 with Amberlyst ( $\text{H}^+$ ) resin and concentrated in vacuum. The residue was finally purified by flash column chromatography ( $\text{MeOH}/\text{CH}_2\text{Cl}_2$ , 1:4, v/v) to give **15** (4.18 g, 90%) as a white solid.  $^1\text{H}$  NMR (400 MHz,  $\text{CD}_3\text{OD}$ ):  $\delta$  4.37 (d,  $J$  = 8.1 Hz, 1 H, H-1'), 4.35 (d,  $J$  = 7.8 Hz, 1 H, H-1), 4.05-3.98 (m, 1 H,  $\text{OCH}_2\text{CH}_2\text{N}_3$ ), 3.95-3.66 (m, 6 H,  $\text{OCH}_2\text{CH}_2\text{N}_3$ , H-6a', H-6b', H-4, H-4', H-6b), 3.62-3.39 (m, 8 H, H-5, H-2, H-3, H-6a, H-3', H-5',  $\text{OCH}_2\text{CH}_2\text{N}_3$ ), 3.32-3.24 (m, 1 H, H-2');  $^{13}\text{C}$  NMR (100 MHz,  $\text{CD}_3\text{OD}$ ):  $\delta$  103.7, 102.9, 79.2, 75.7, 75.1, 75.0, 73.4, 73.3, 71.1, 68.9, 68.0, 61.1, 60.5, 50.6.

## **Supplementary References**

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- [8] G. Milkereit, V. Vill, *J. Carbohydr. Chem.* **2006**, *25*, 615-632.
- [9] W. Wang, H. Wang, C. Ren, J. Wang, M. Tan, J. Shen, Z. Yang, P. G. Wang, L. Wang, *Carbohydr. Res.* **2011**, *346*, 1013-1017.

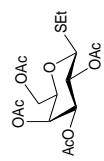
## Compound NMR and MS Spectra

Mercury 400 spectrometer

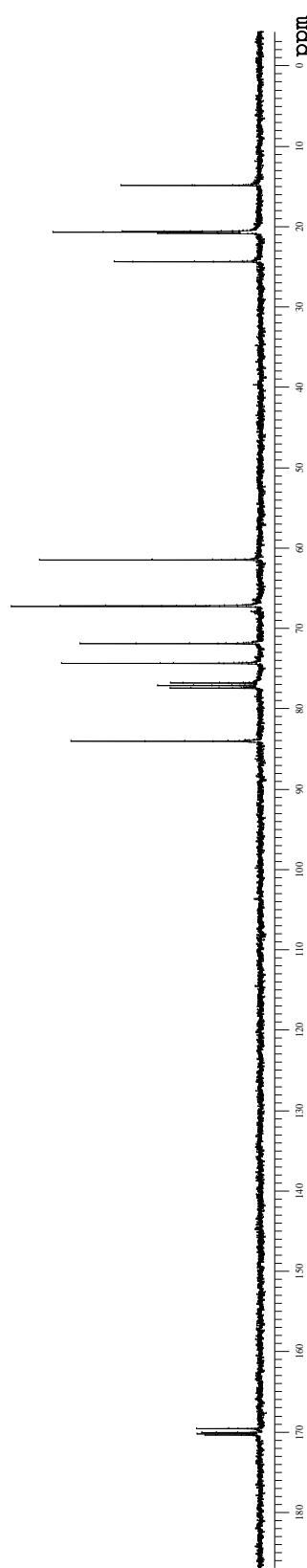


<sup>1</sup>H NMR Spectrum of compound **S2** (CDCl<sub>3</sub>, 400 MHz)

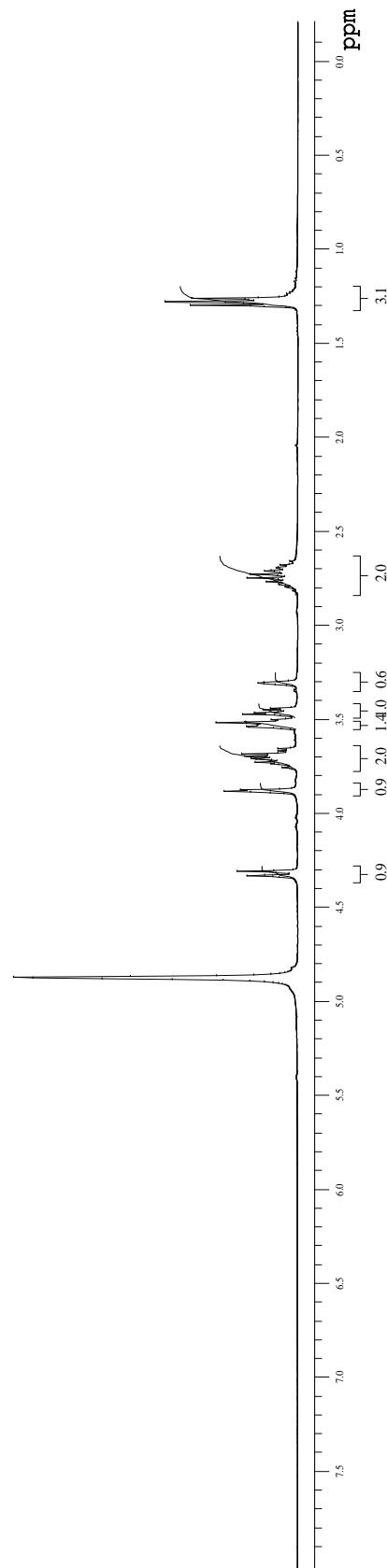
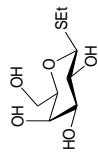
Mercury 400 spectrometer



<sup>13</sup>C NMR Spectrum of compound **S2** (CDCl<sub>3</sub>, 400 MHz)

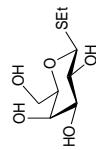


Mercury 400 spectrometer

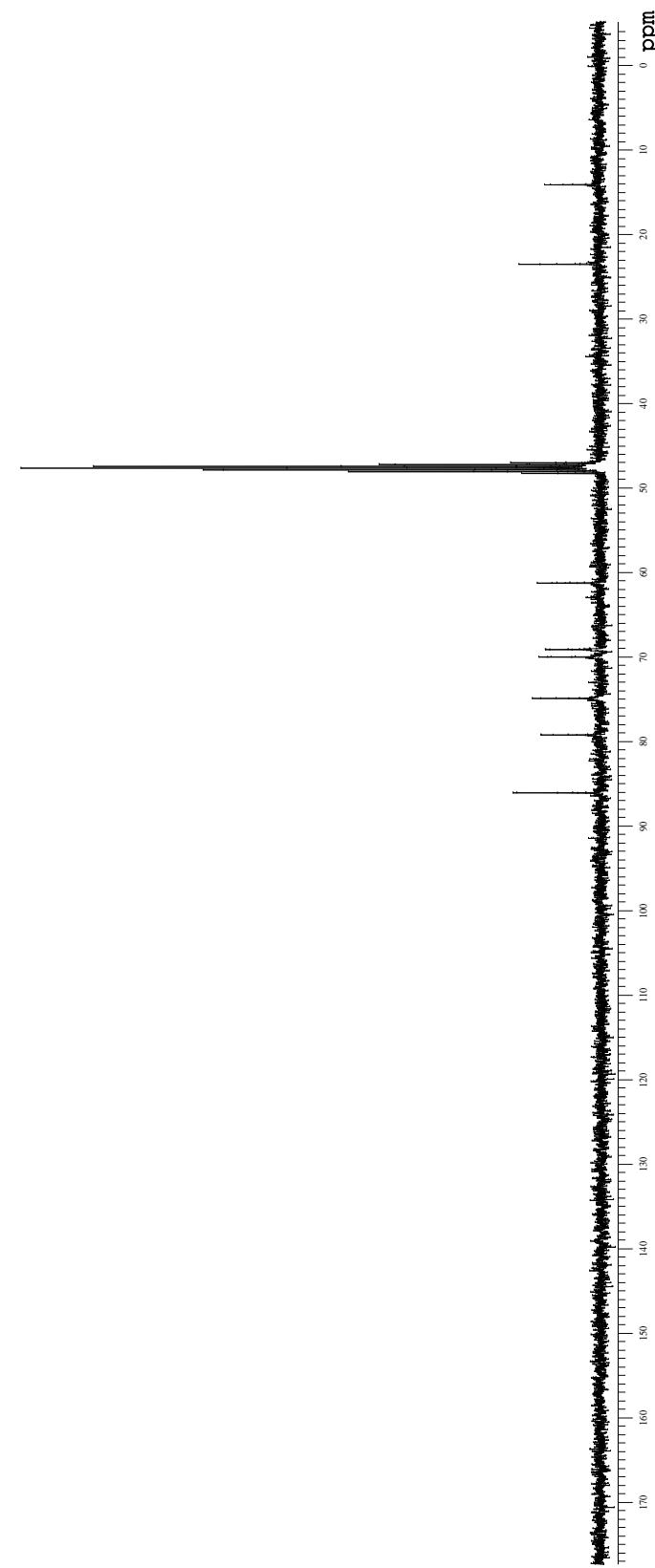


<sup>1</sup>H NMR Spectrum of compound S3 (CD<sub>3</sub>OD, 400 MHz)

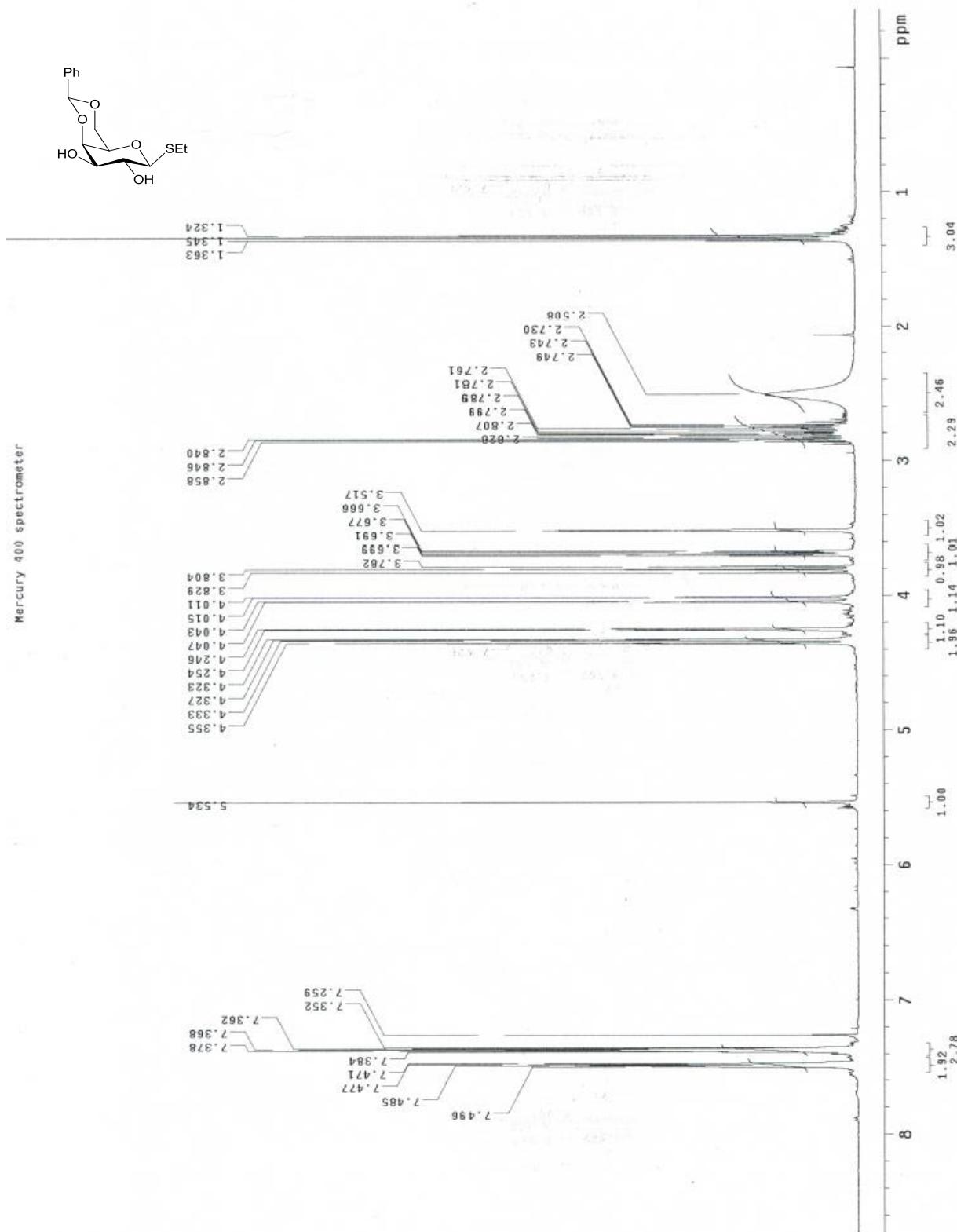
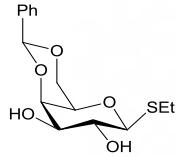
Mercury 400 spectrometer



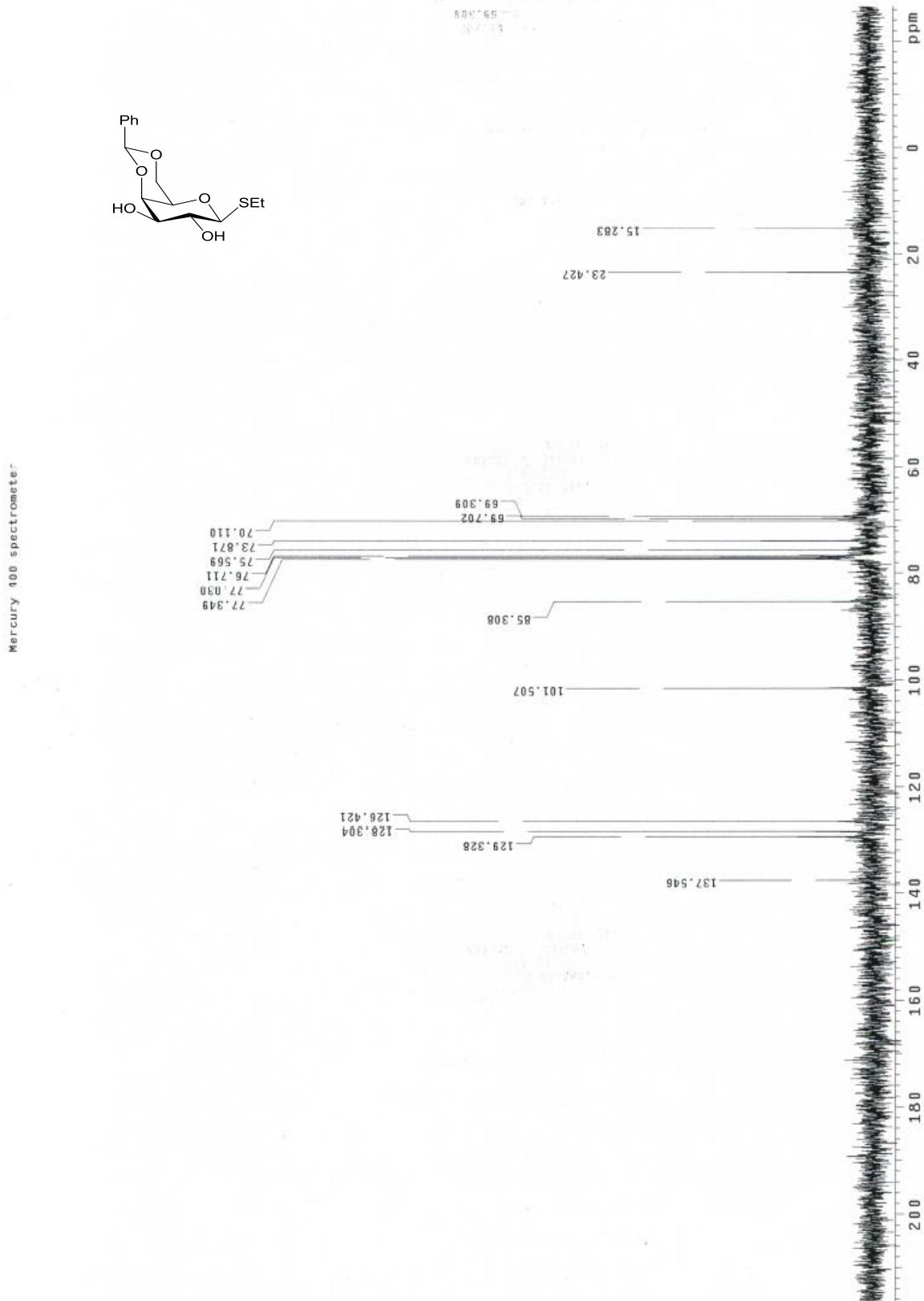
<sup>13</sup>C NMR Spectrum of compound S3 (CD<sub>3</sub>OD, 400 MHz)



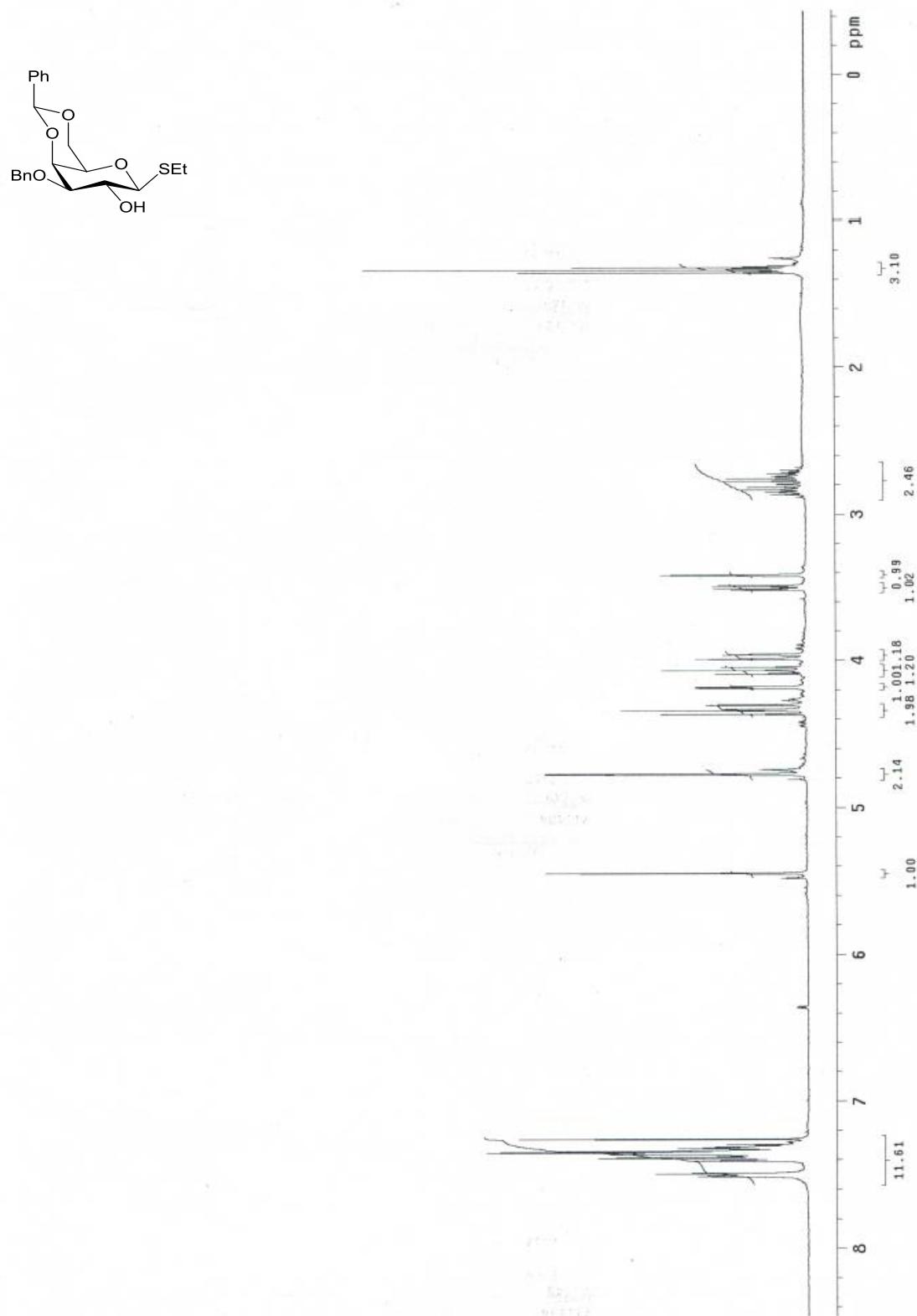
#### Mercury 400 spectrometer



<sup>1</sup>H NMR Spectrum of compound **11** (CDCl<sub>3</sub>, 400 MHz)

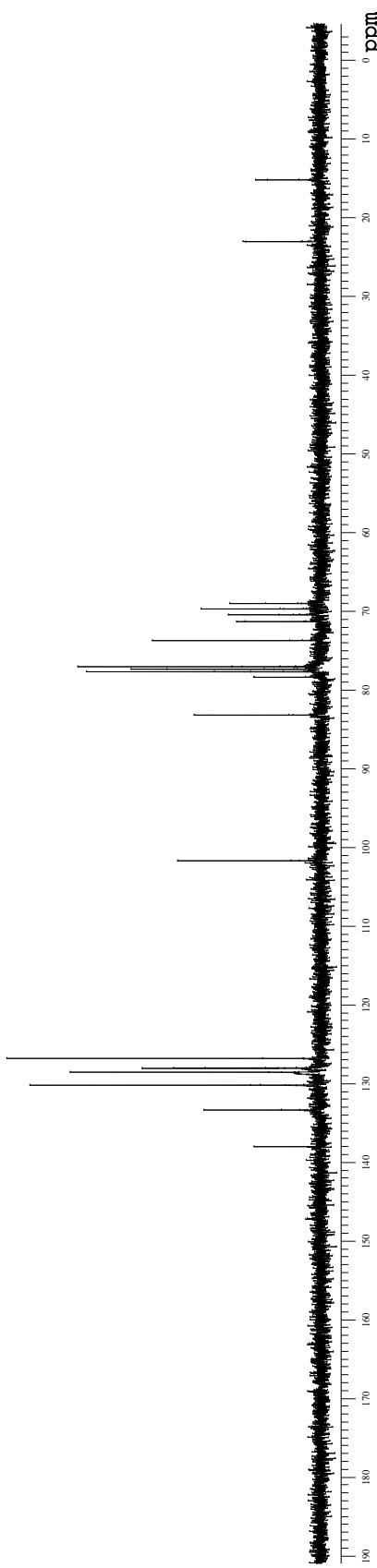
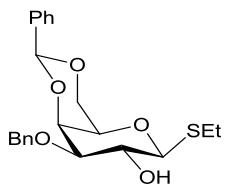


$^{13}\text{C}$  NMR Spectrum of compound **11** ( $\text{CDCl}_3$ , 400 MHz)



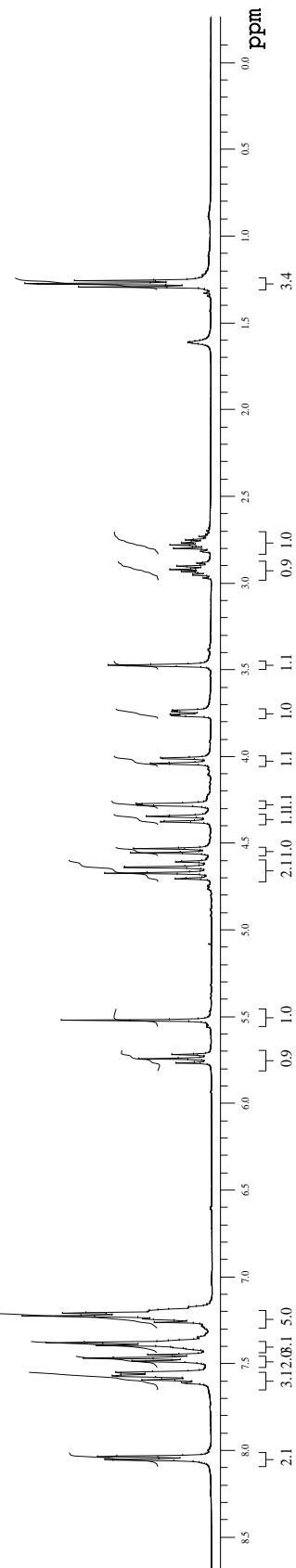
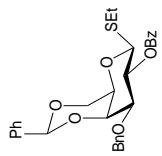
<sup>1</sup>H NMR Spectrum of compound 12 (CDCl<sub>3</sub>, 400 MHz)

Mercury 400 spectrometer



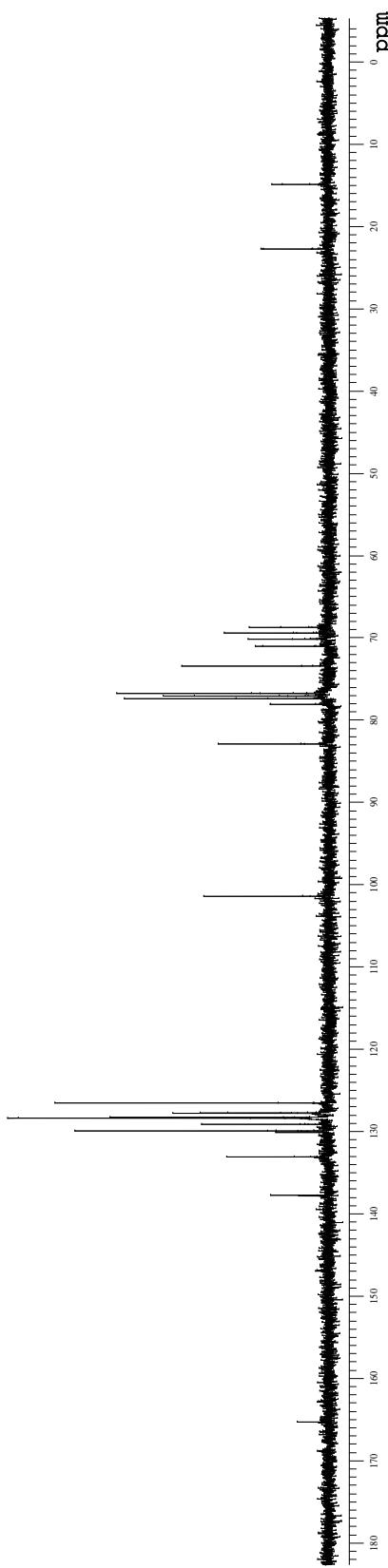
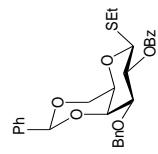
<sup>13</sup>C NMR Spectrum of compound **12** (CDCl<sub>3</sub>, 400 MHz)

## Mercury 400 spectrometer



<sup>1</sup>H NMR Spectrum of compound **7** (CDCl<sub>3</sub>, 400 MHz)

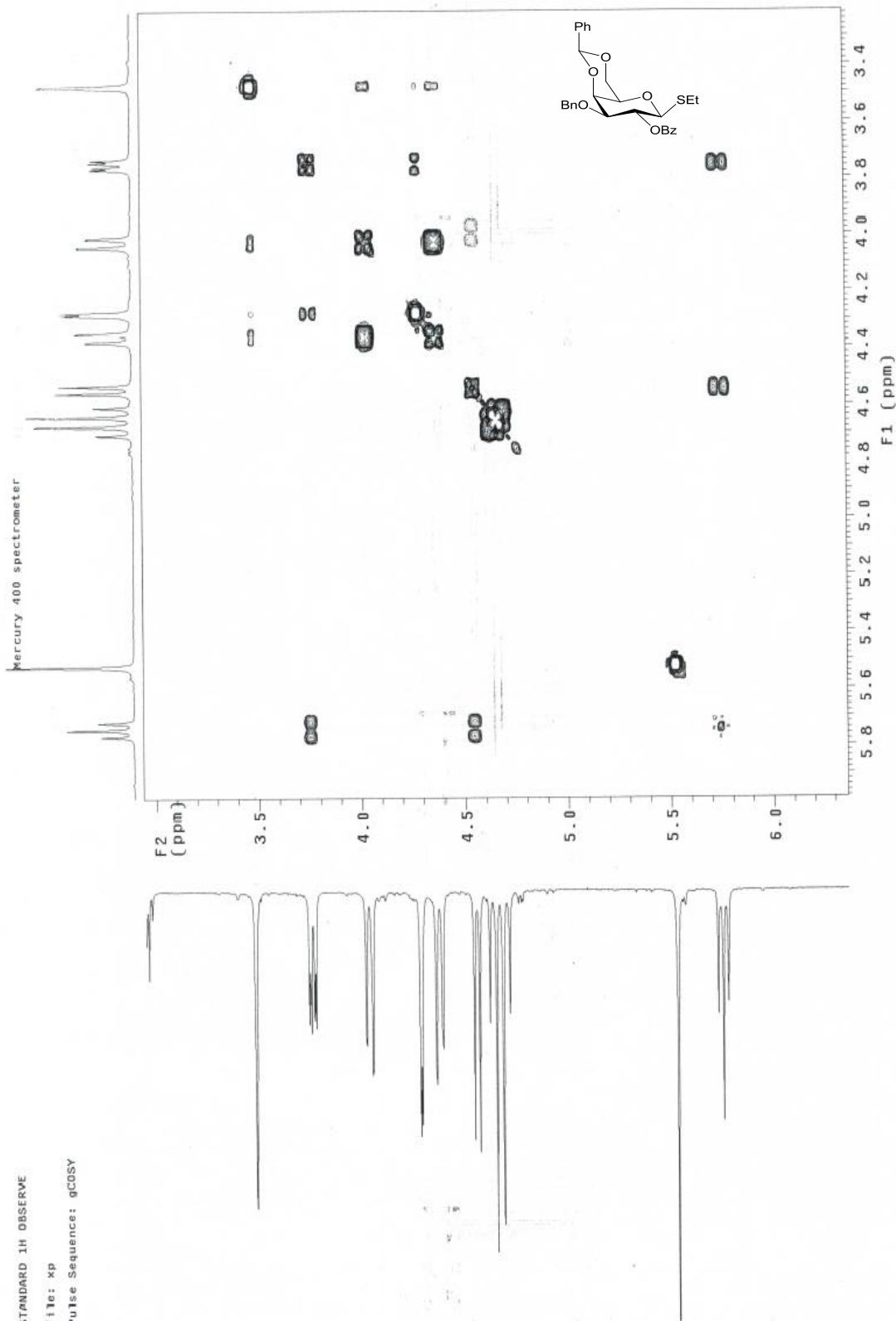
Mercury 400 spectrometer



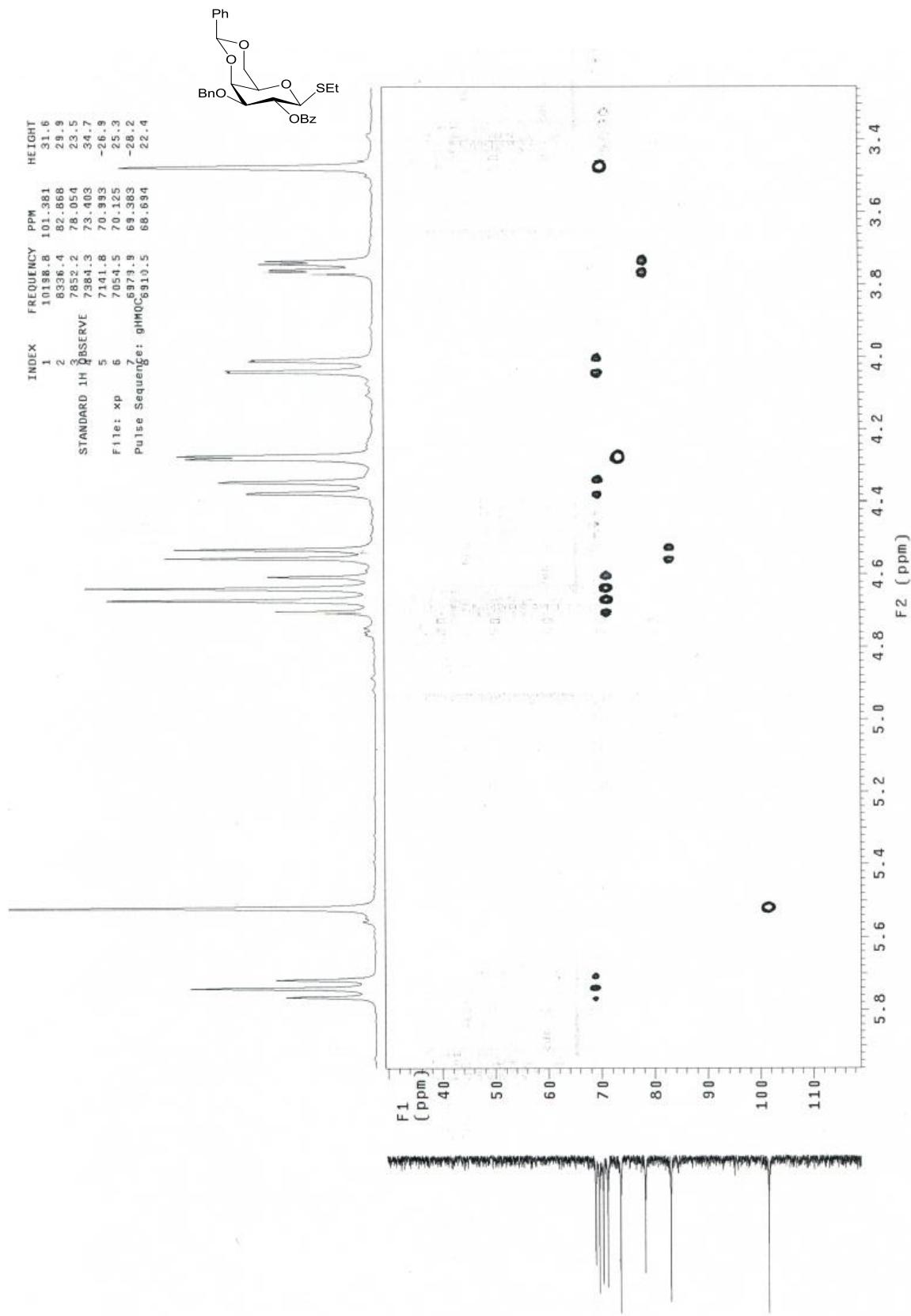
$^{13}\text{C}$  NMR Spectrum of compound 7 ( $\text{CDCl}_3$ , 400 MHz)

STANDARD 1H OBSERVE  
File: xp  
Pulse Sequence: gCOSY

Mercury 400 spectrometer



$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound 7 ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound 7 ( $\text{CDCl}_3$ , 400/100 MHz)

## Elemental Composition Report

Page 1

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

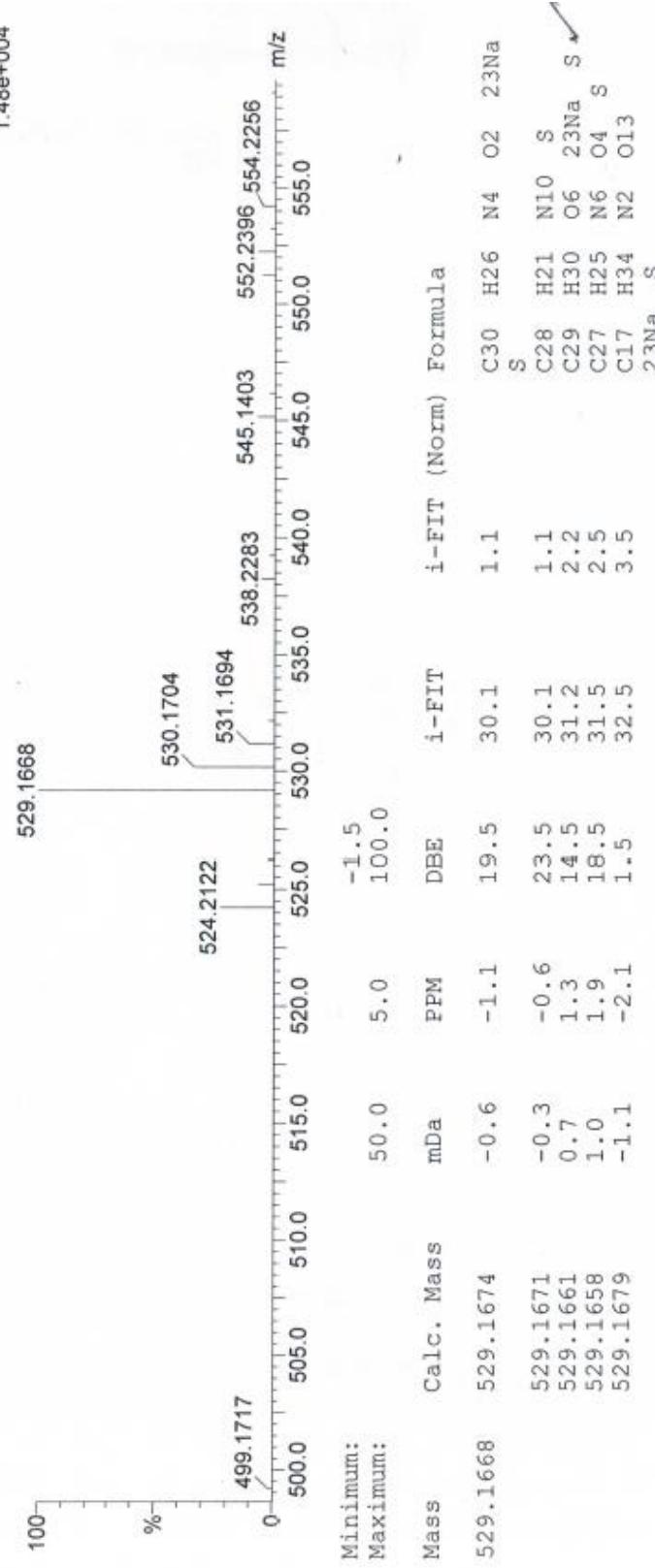
### Monoisotopic Mass, Even Electron Ions

4185 formula(e) evaluated with 19 results within limits (all results (up to 1000) for each mass)

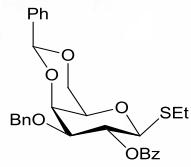
Elements Used:

C: 0-100 H: 0-100 N: 0-10 O: 0-30 23Na: 0-1 S: 1-2  
SSM:**4<sub>al</sub>-OBz(SEt)**

SATADRUMANDAL  
2013\_0129\_3051\_2 14 (0.283) Cm (11:18-1:9x2.000)

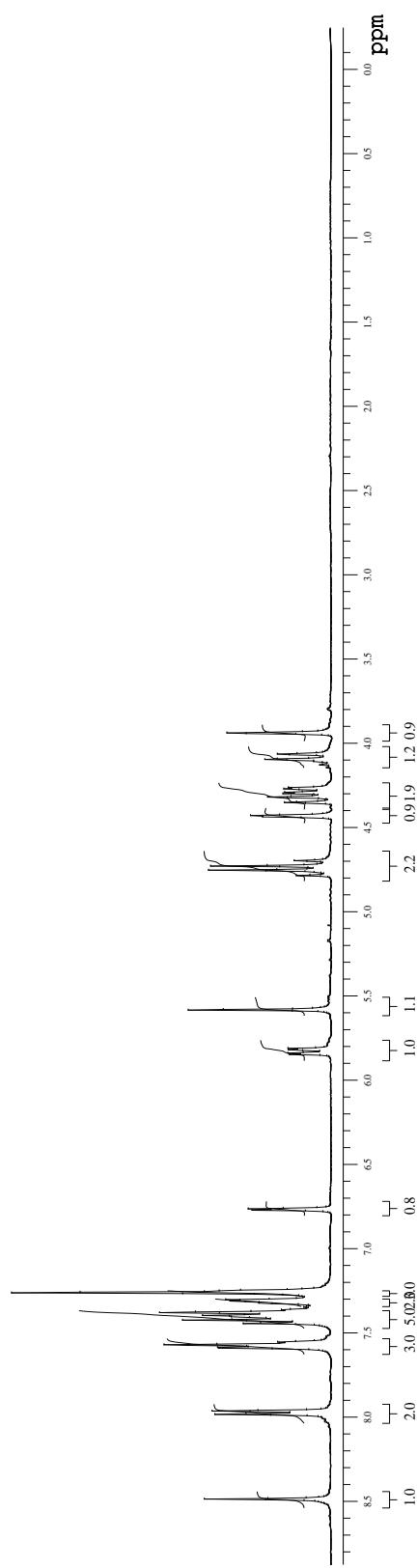
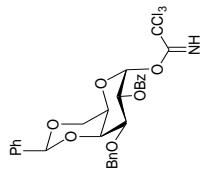


HRMS (ESI MS) spectrum of compound 7



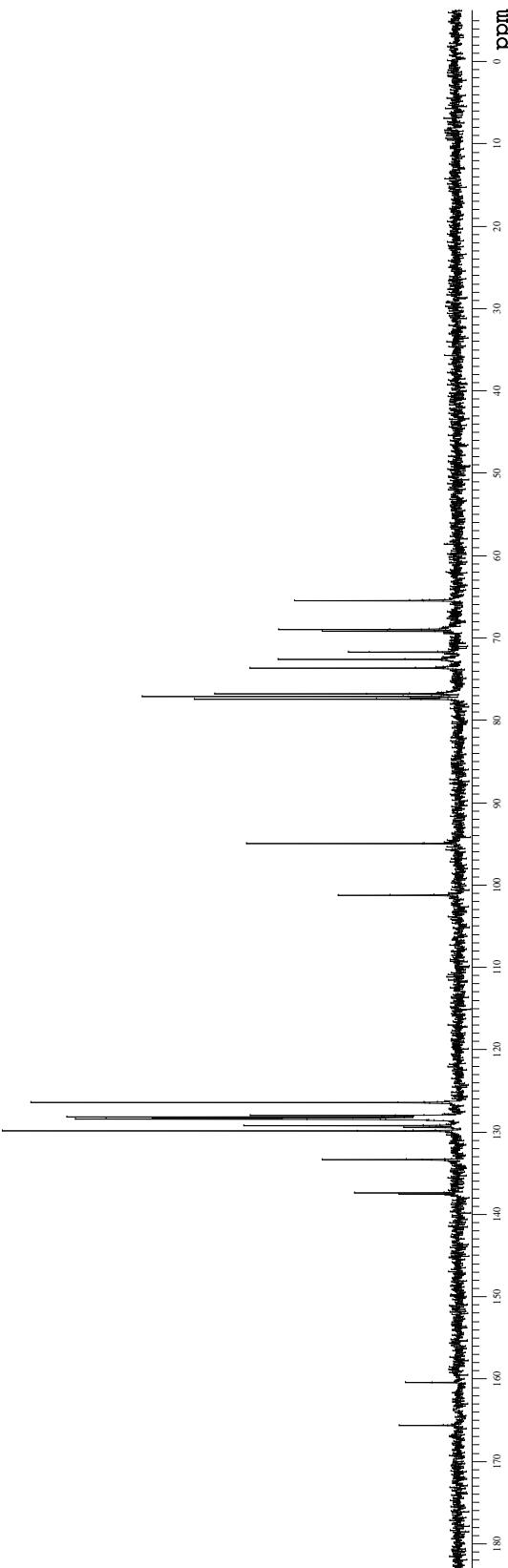
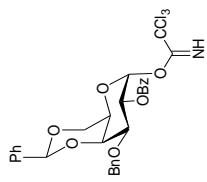
LCT2008-07b.pro 2010-cif.spillCT Premier  
1: TOF MS ES+  
1.48e+004

Mercury 400 spectrometer

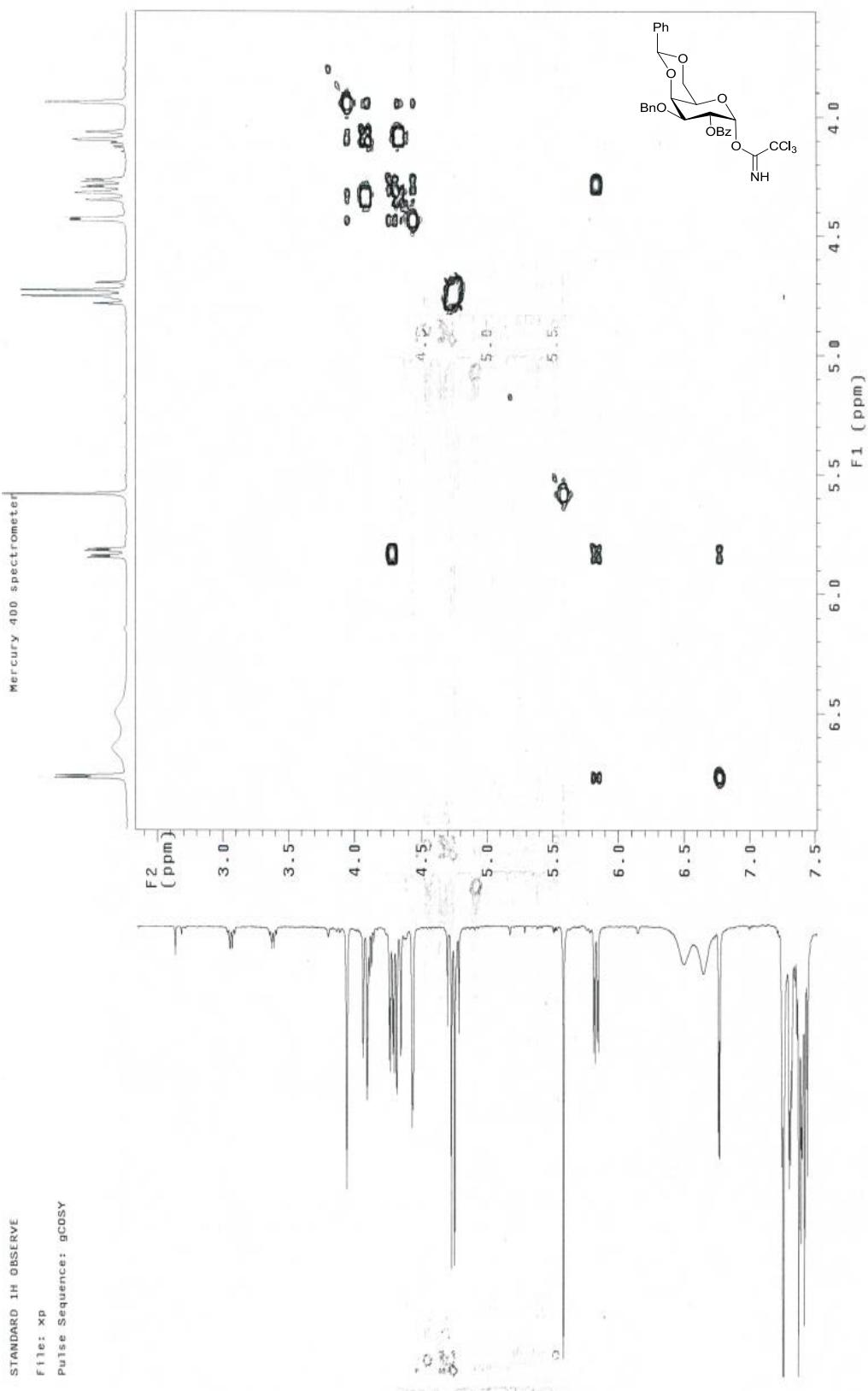


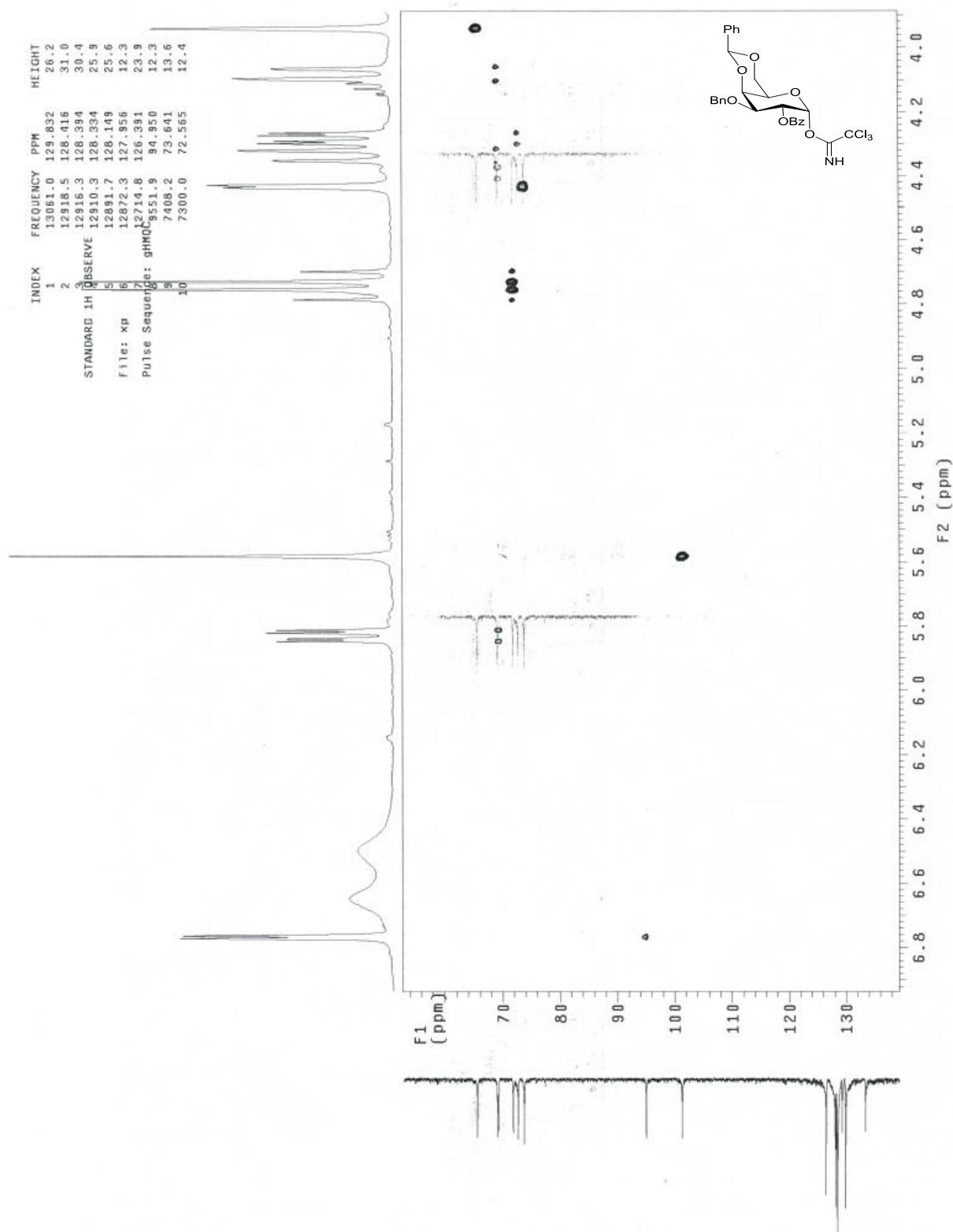
<sup>1</sup>H NMR Spectrum of compound 19 (CDCl<sub>3</sub>, 400 MHz)

Mercury 400 spectrometer



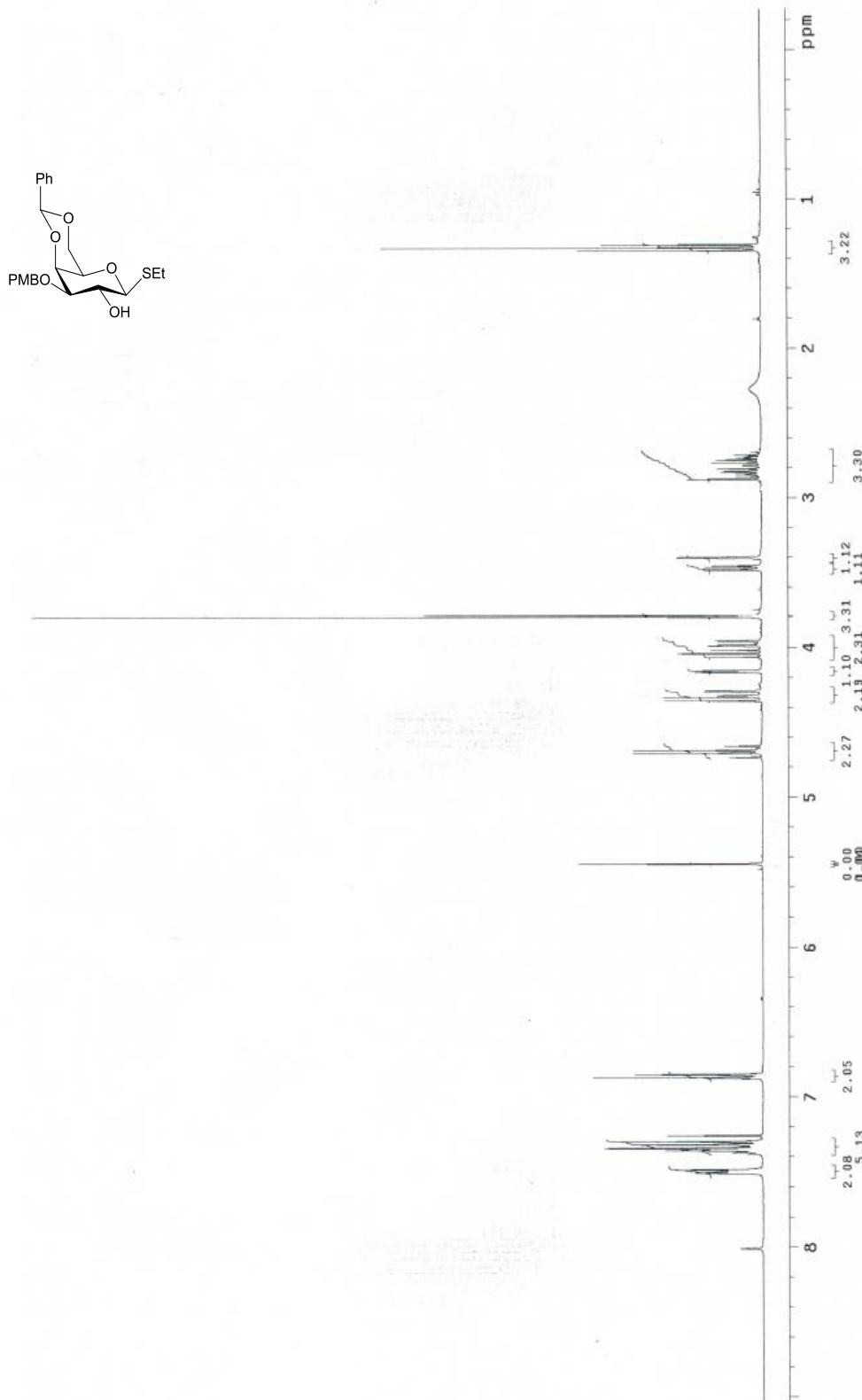
<sup>13</sup>C NMR Spectrum of compound **19** (CDCl<sub>3</sub>, 400 MHz)

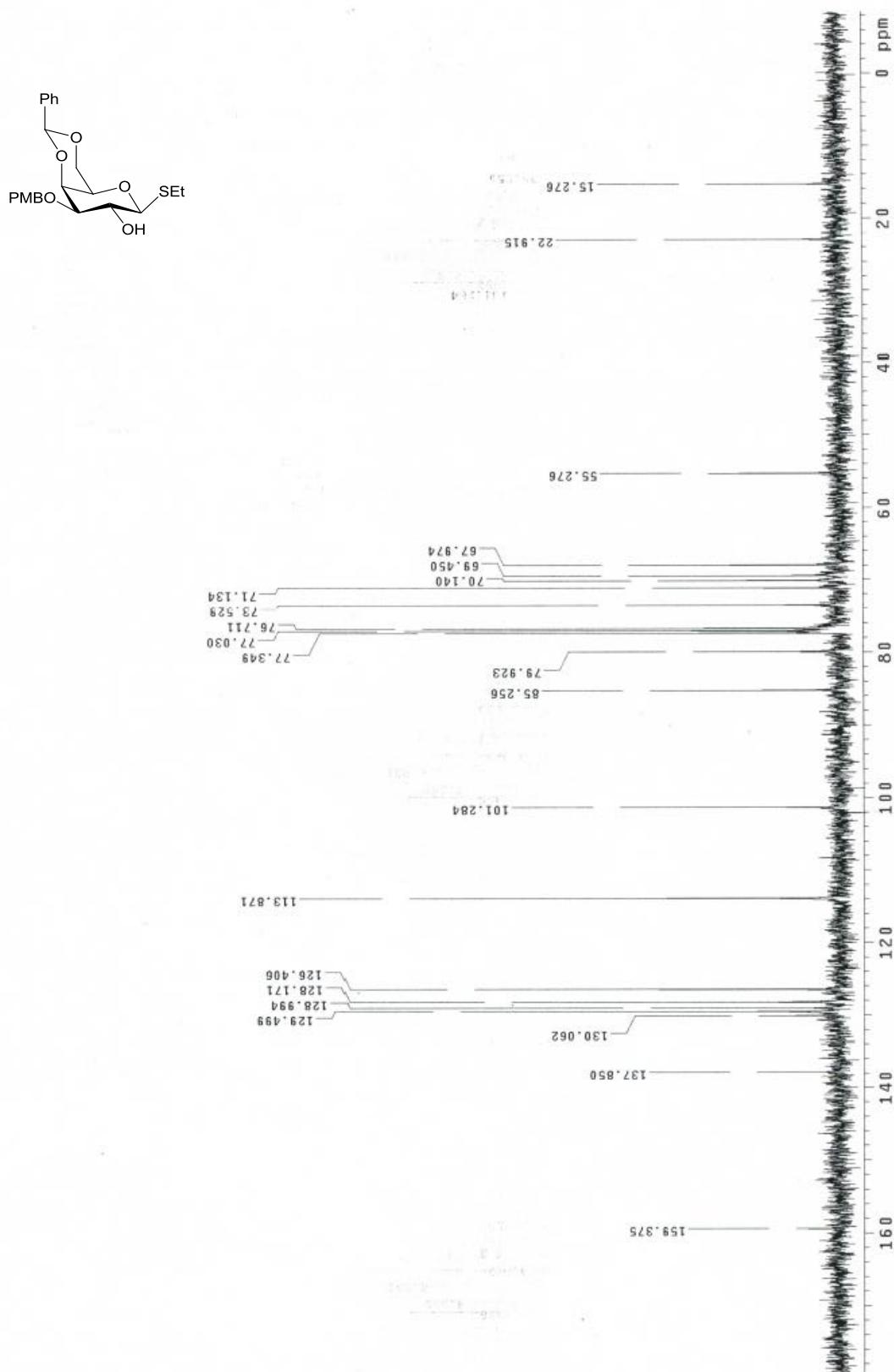




<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **19** (CDCl<sub>3</sub>, 400/100 MHz)

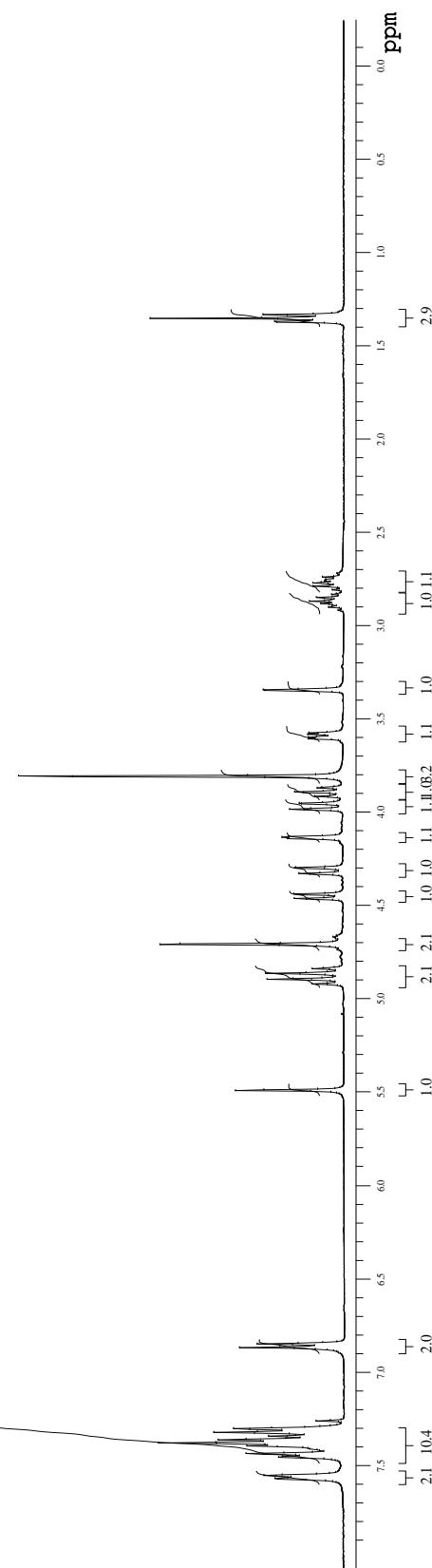
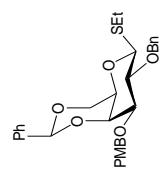
Mercury 400 spectrometer





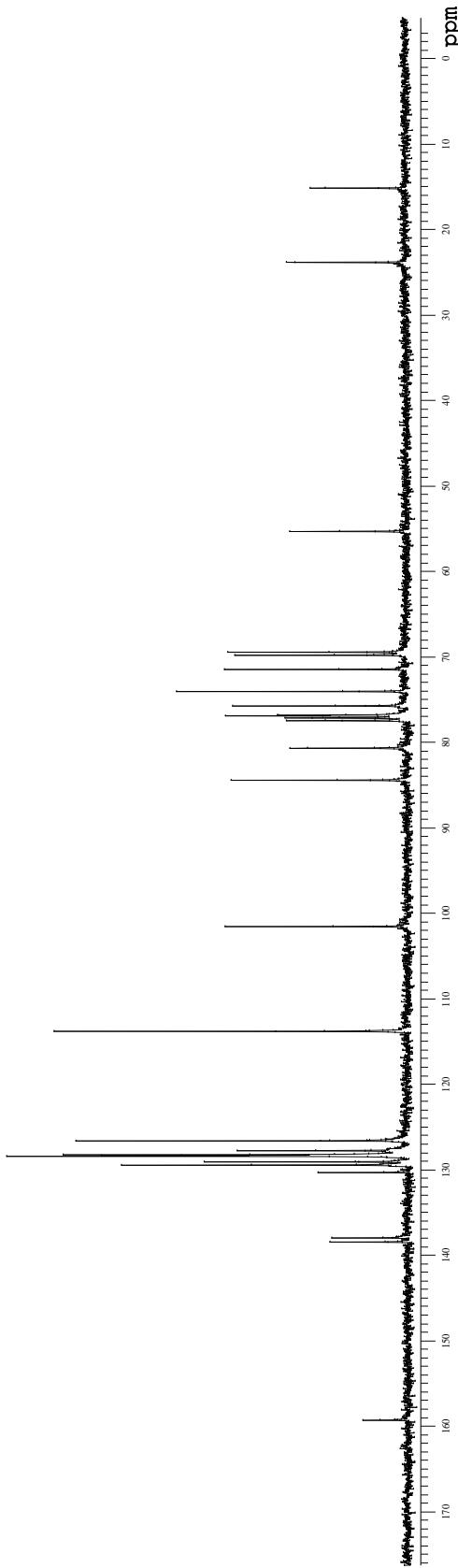
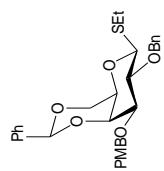
<sup>13</sup>C NMR Spectrum of compound 13 (CDCl<sub>3</sub>, 400 MHz)

Mercury 400 spectrometer



<sup>1</sup>H NMR Spectrum of compound **14** (CDCl<sub>3</sub>, 400 MHz)

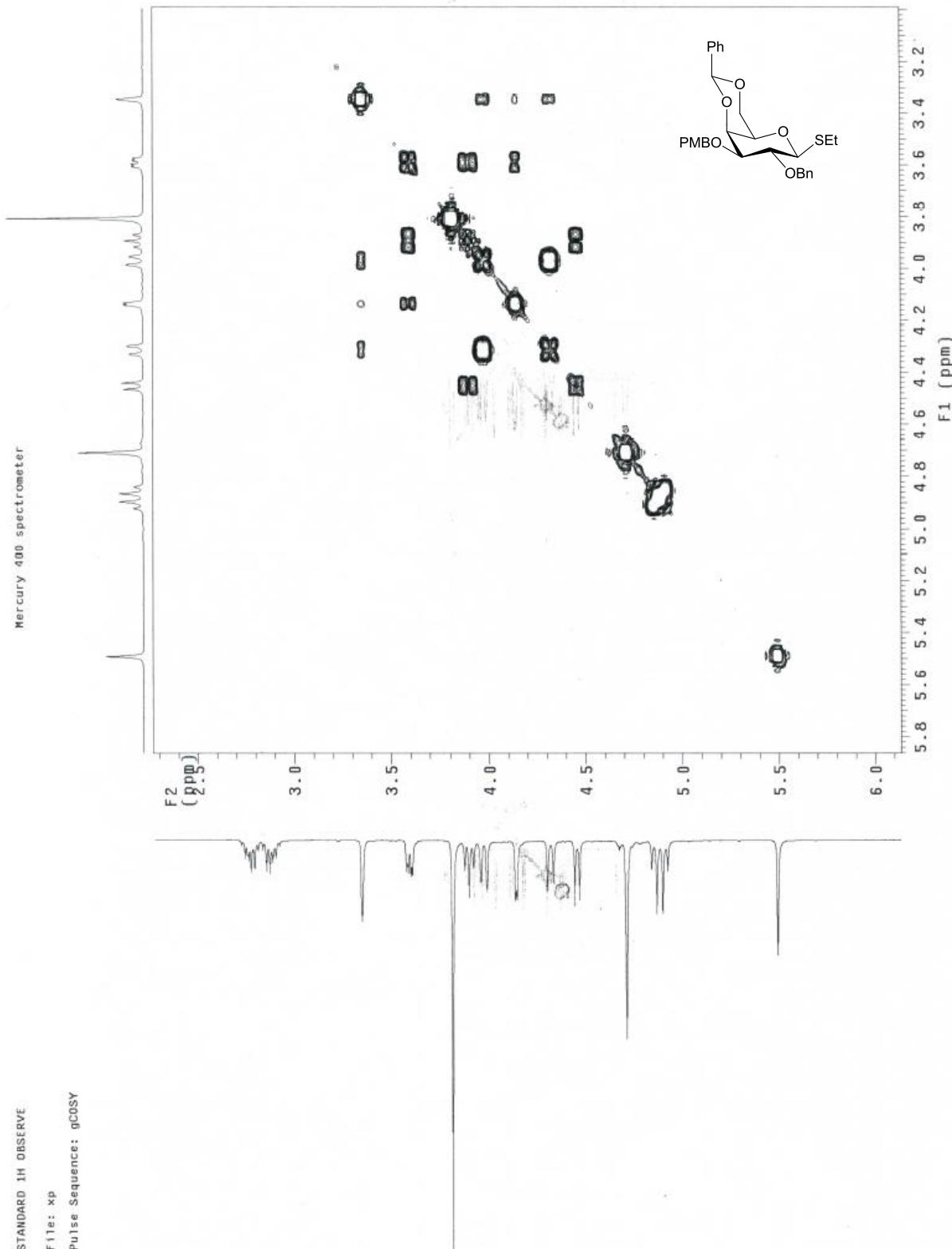
Mercury 400 spectrometer



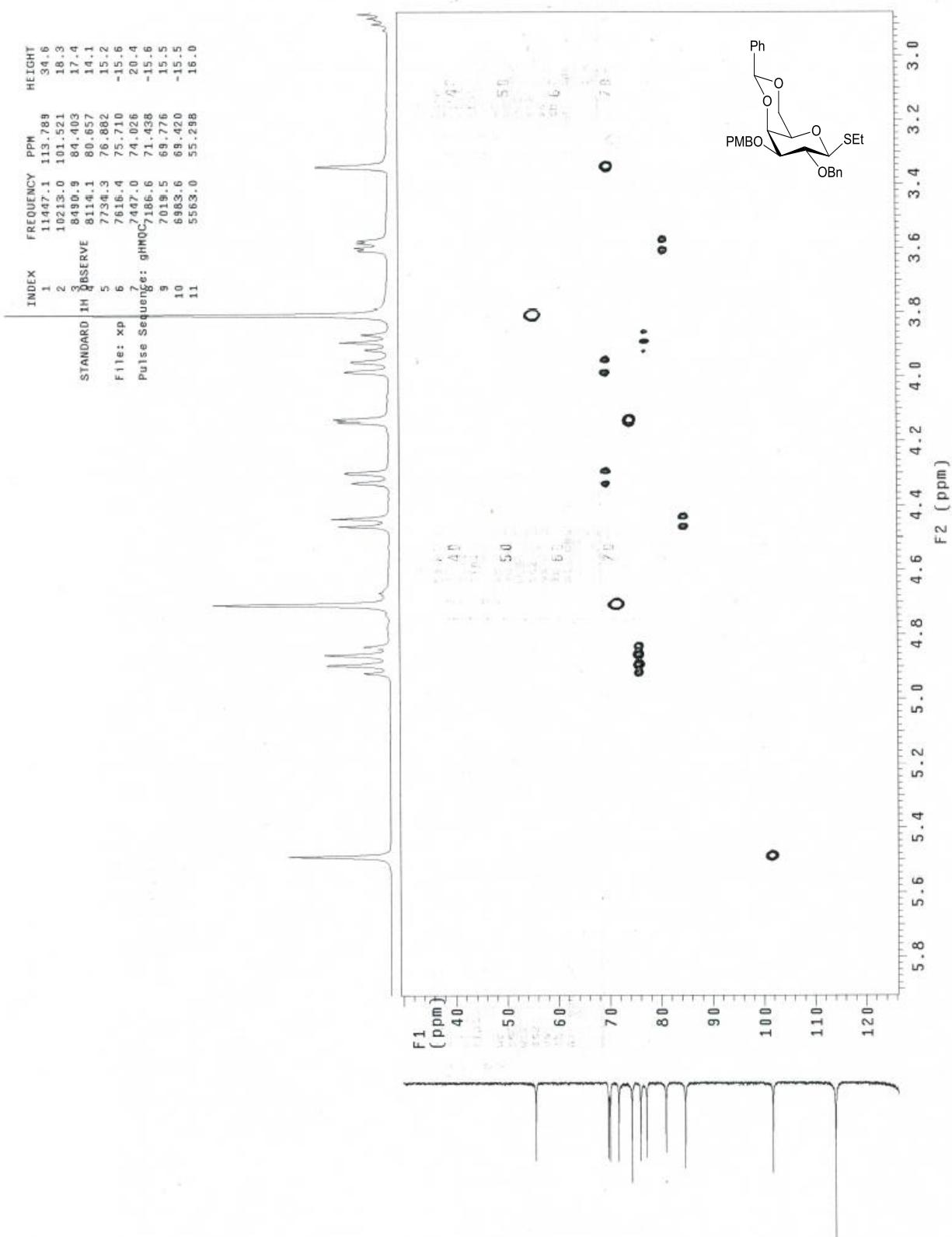
$^{13}\text{C}$  NMR Spectrum of compound **14** ( $\text{CDCl}_3$ , 400 MHz)

STANDARD 1H OBSERVE  
File: xp  
Pulse Sequence: gCOSY

Mercury 400 spectrometer



$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **14** ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **14** ( $\text{CDCl}_3$ , 400/100 MHz)

## Elemental Composition Report

### Single Mass Analysis

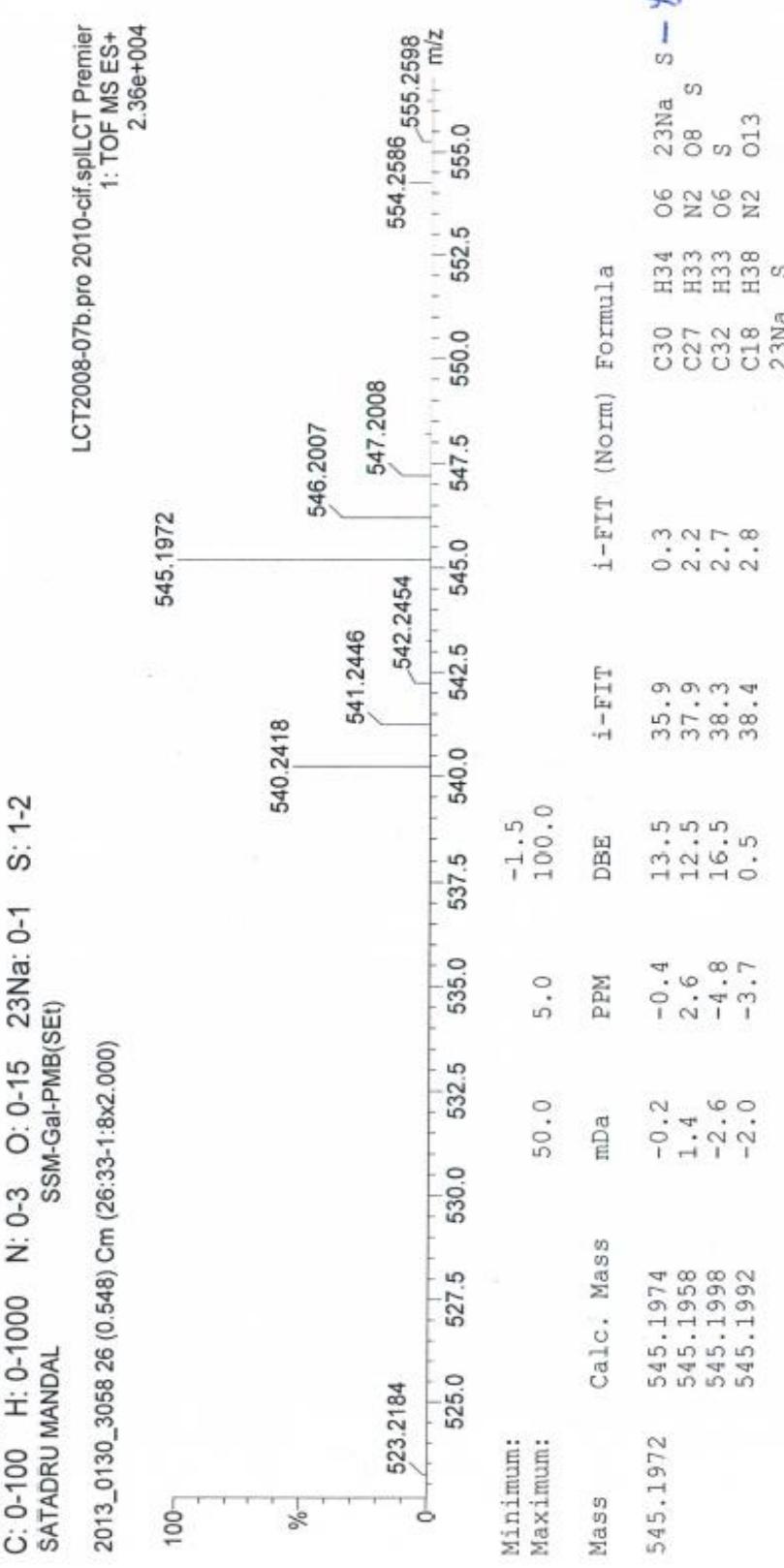
Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

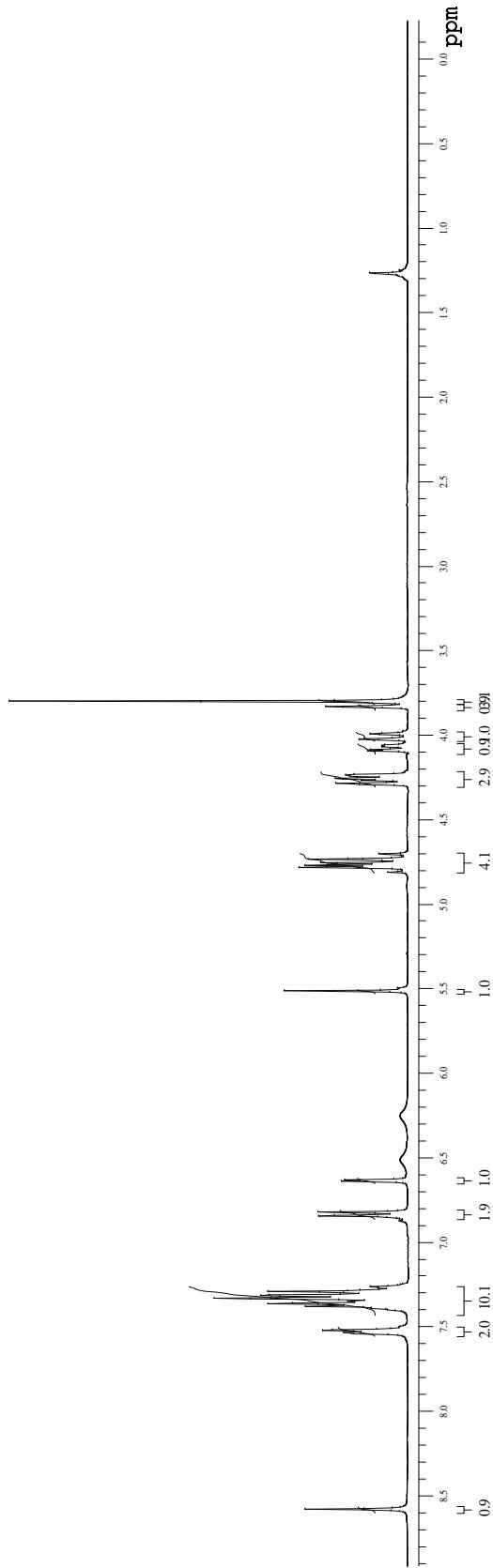
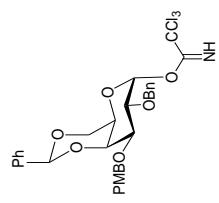
Monoisotopic Mass, Even Electron Ions  
1325 formula(e) evaluated with 8 results within limits (all results (up to 1000) for each mass)  
Elements Used:  
C: 0-100 H: 0-1000 N: 0-3 O: 0-15 23Na: 0-1 S: 1-2  
SSM-GaI-PMB(SEt)

2013\_0130\_3058 26 (0.548) Cm (26:33:1.8x2.000)



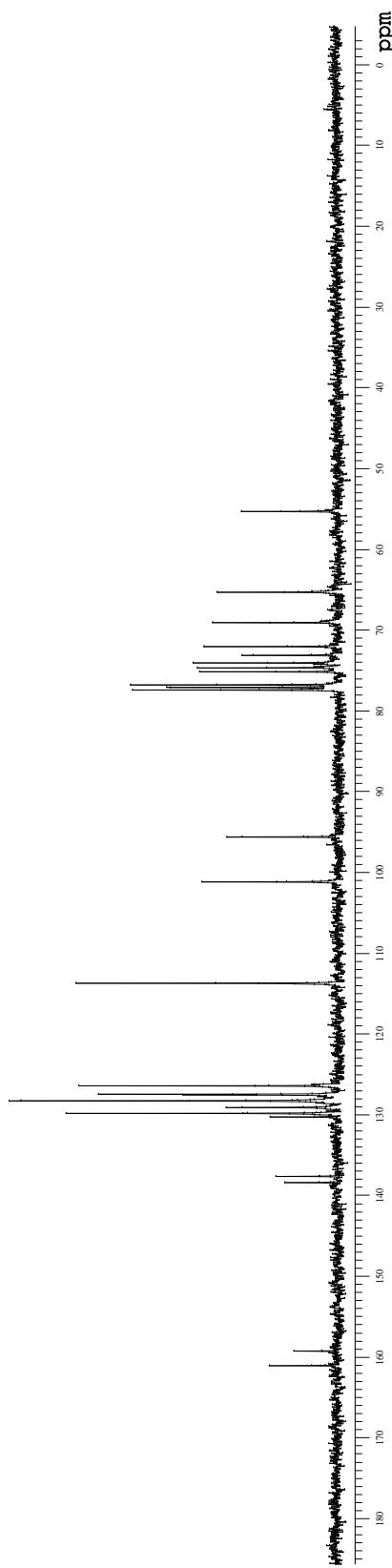
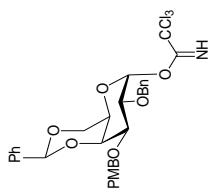
HRMS (ESI MS) spectrum of compound 14

Mercury 400 spectrometer

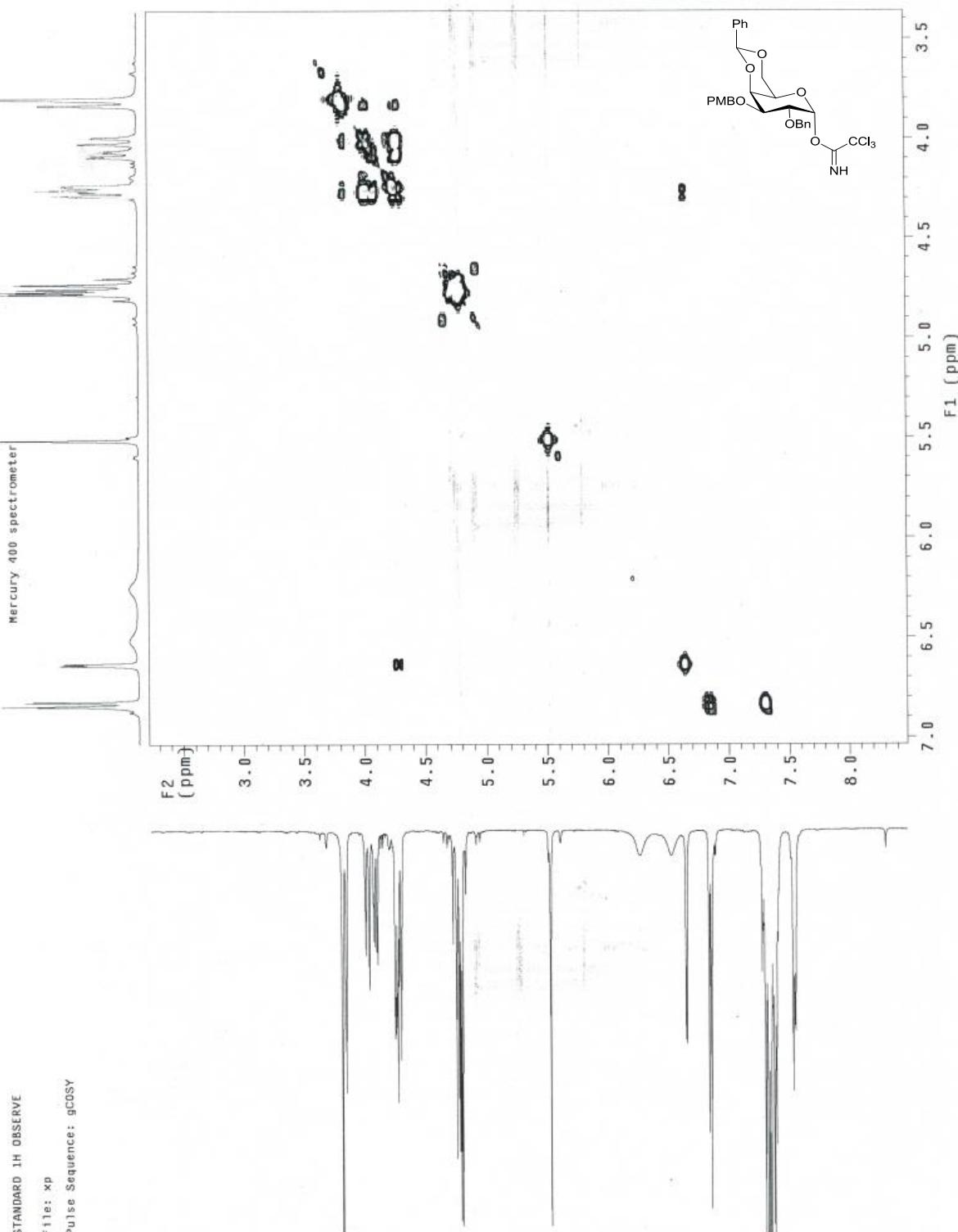


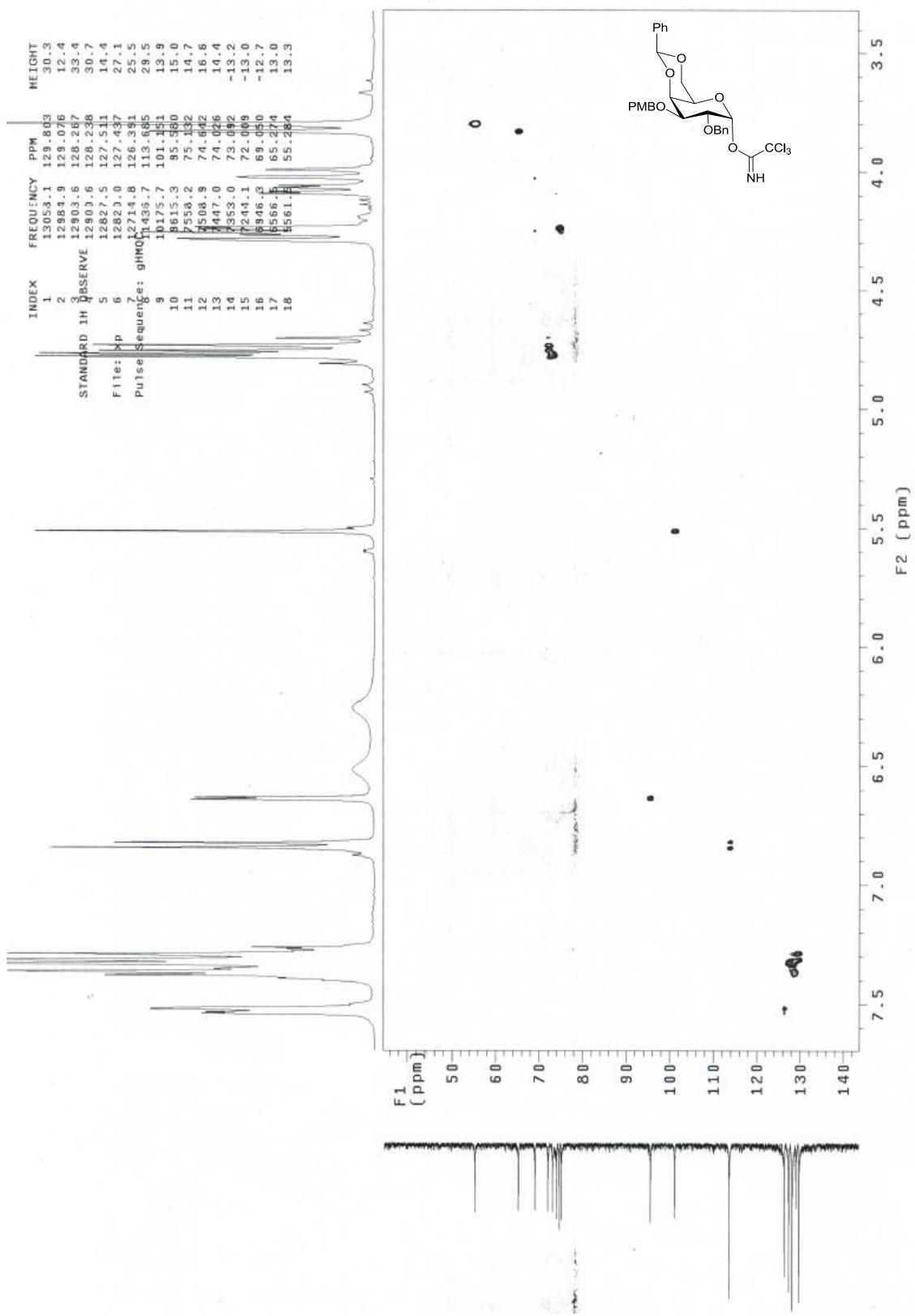
<sup>1</sup>H NMR Spectrum of compound 9 (CDCl<sub>3</sub>, 400 MHz)

Mercury 400 spectrometer



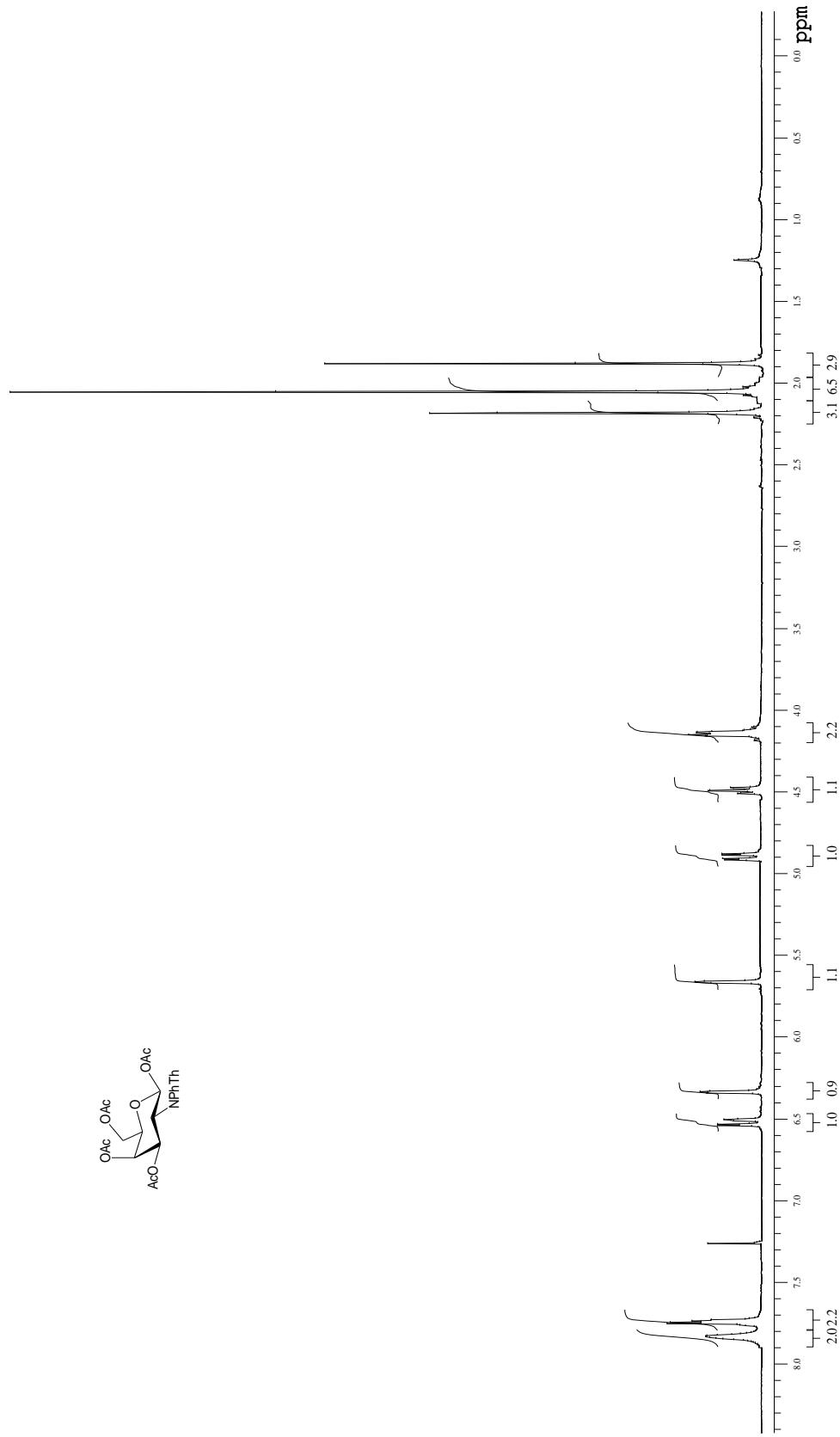
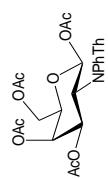
<sup>13</sup>C NMR Spectrum of compound 9 (CDCl<sub>3</sub>, 400 MHz)





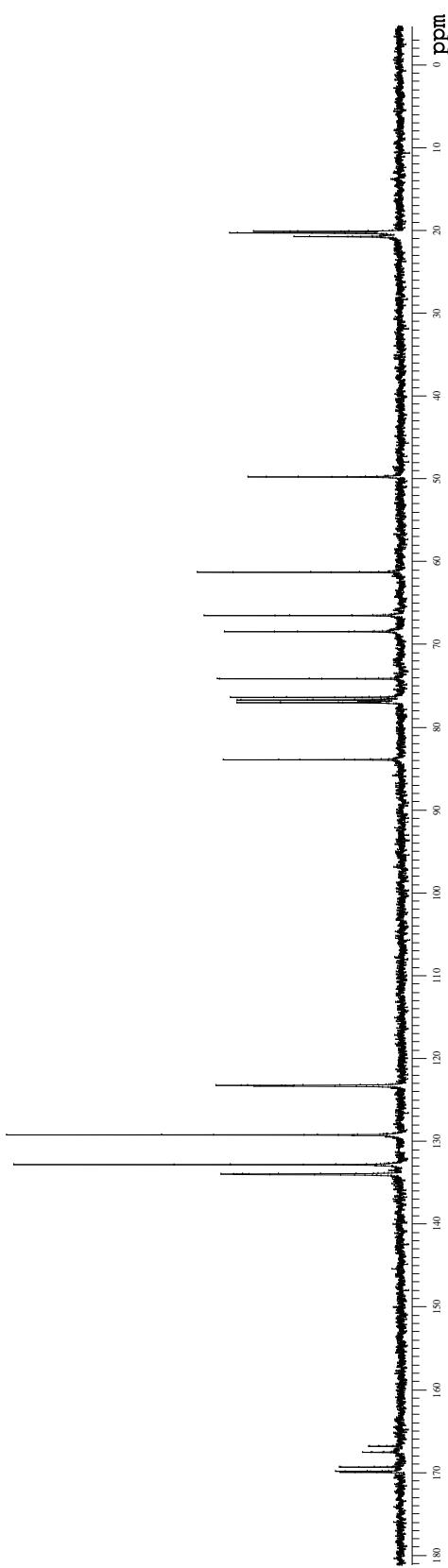
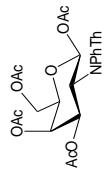
<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **9** (CDCl<sub>3</sub>, 400/100 MHz)

Mercury 400 spectrometer



<sup>1</sup>H NMR Spectrum of compound **S5** (CDCl<sub>3</sub>, 400 MHz)

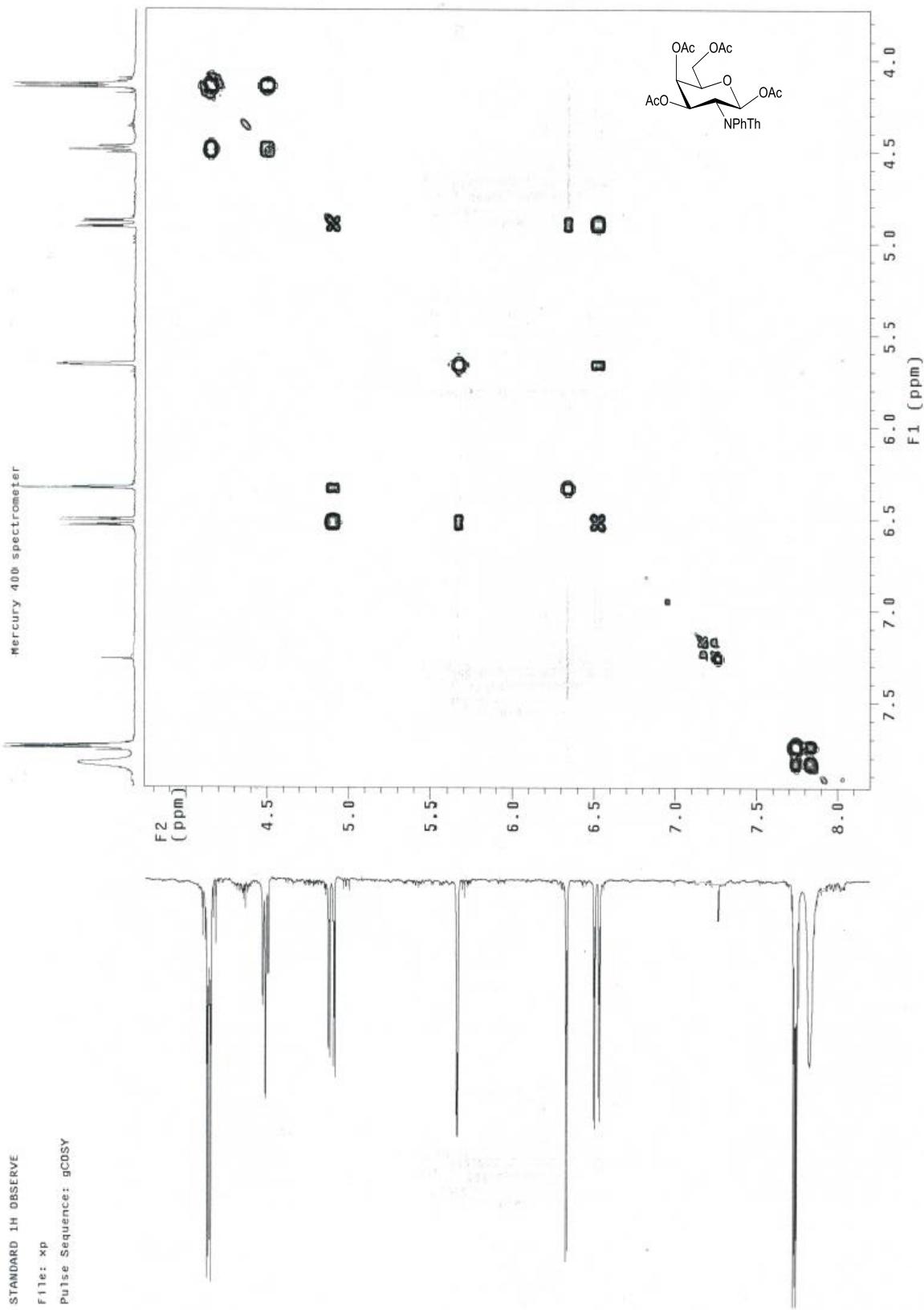
Mercury 400 spectrometer



<sup>13</sup>C NMR Spectrum of compound S5 (CDCl<sub>3</sub>, 400 MHz)

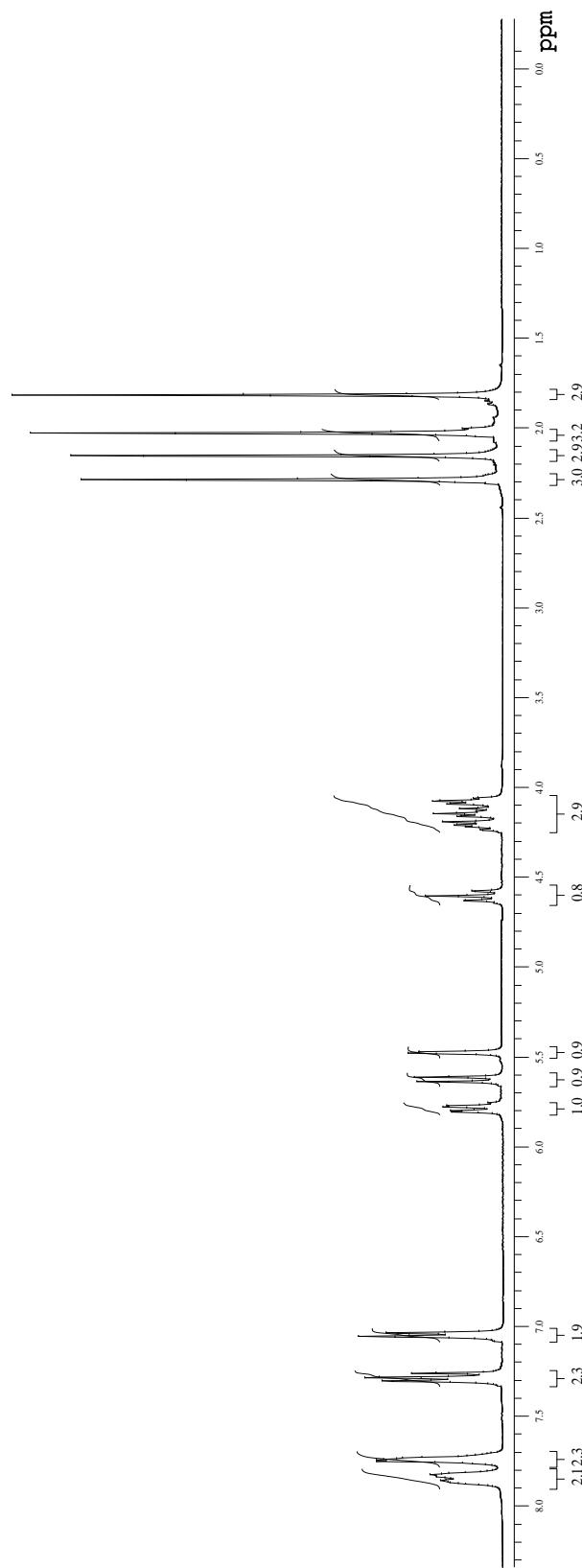
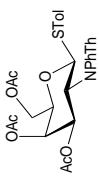
STANDARD 1H OBSERVE  
F1le: xp  
Pulse Sequence: gCOSY

Mercury 400 spectrometer



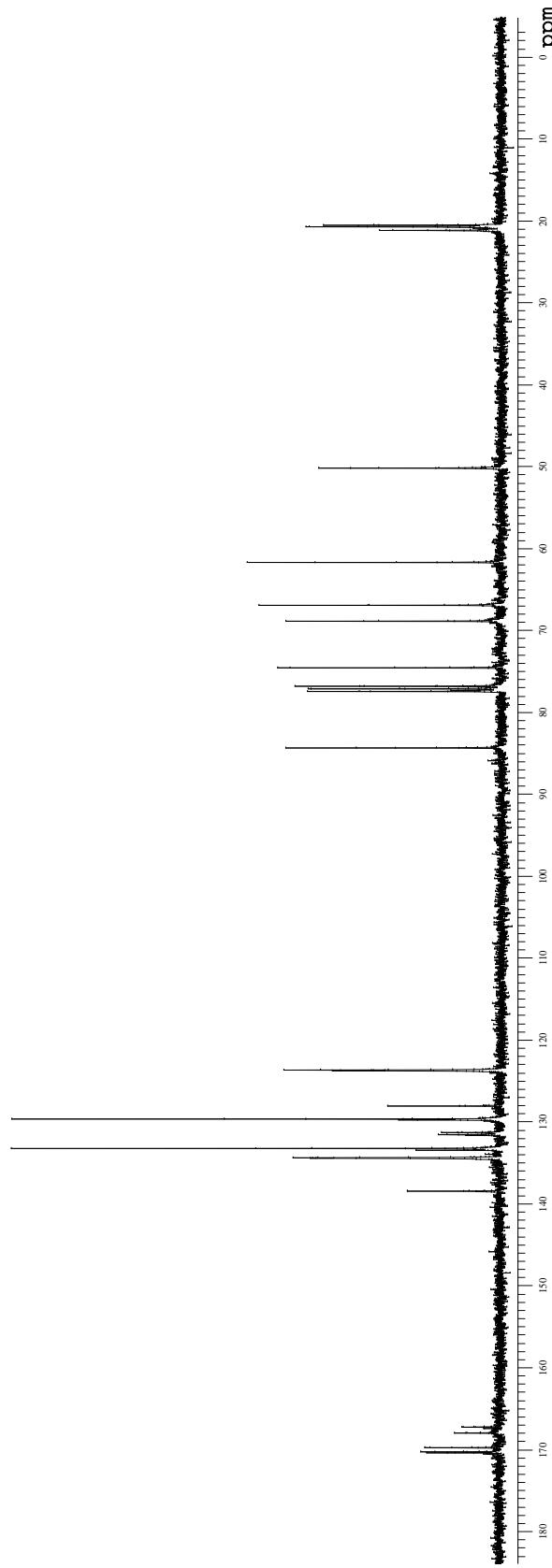
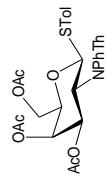
$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound S5 ( $\text{CDCl}_3$ , 400 MHz)

Mercury 400 spectrometer



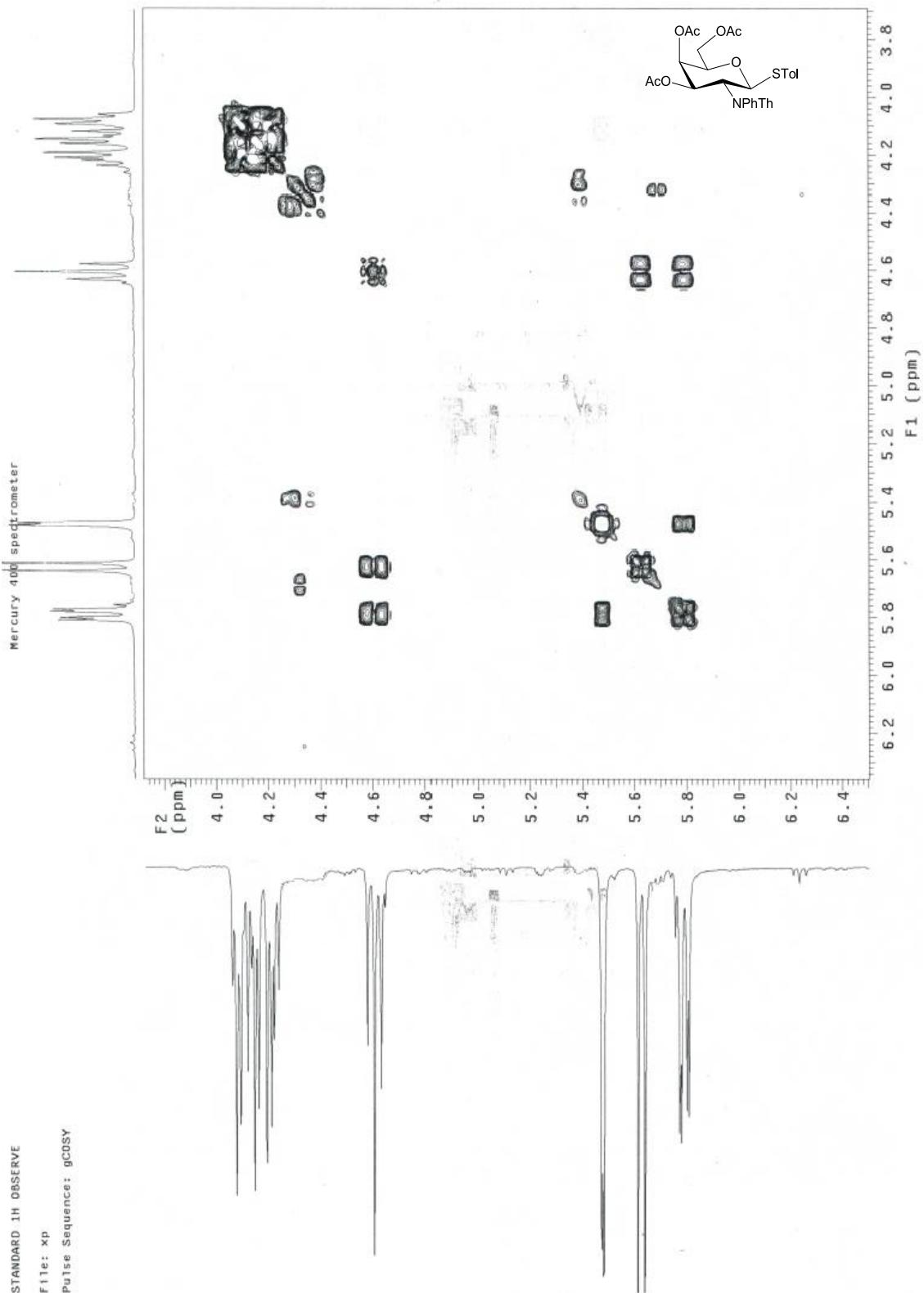
<sup>1</sup>H NMR Spectrum of compound S6 (CDCl<sub>3</sub>, 400 MHz)

Mercury 400 spectrometer



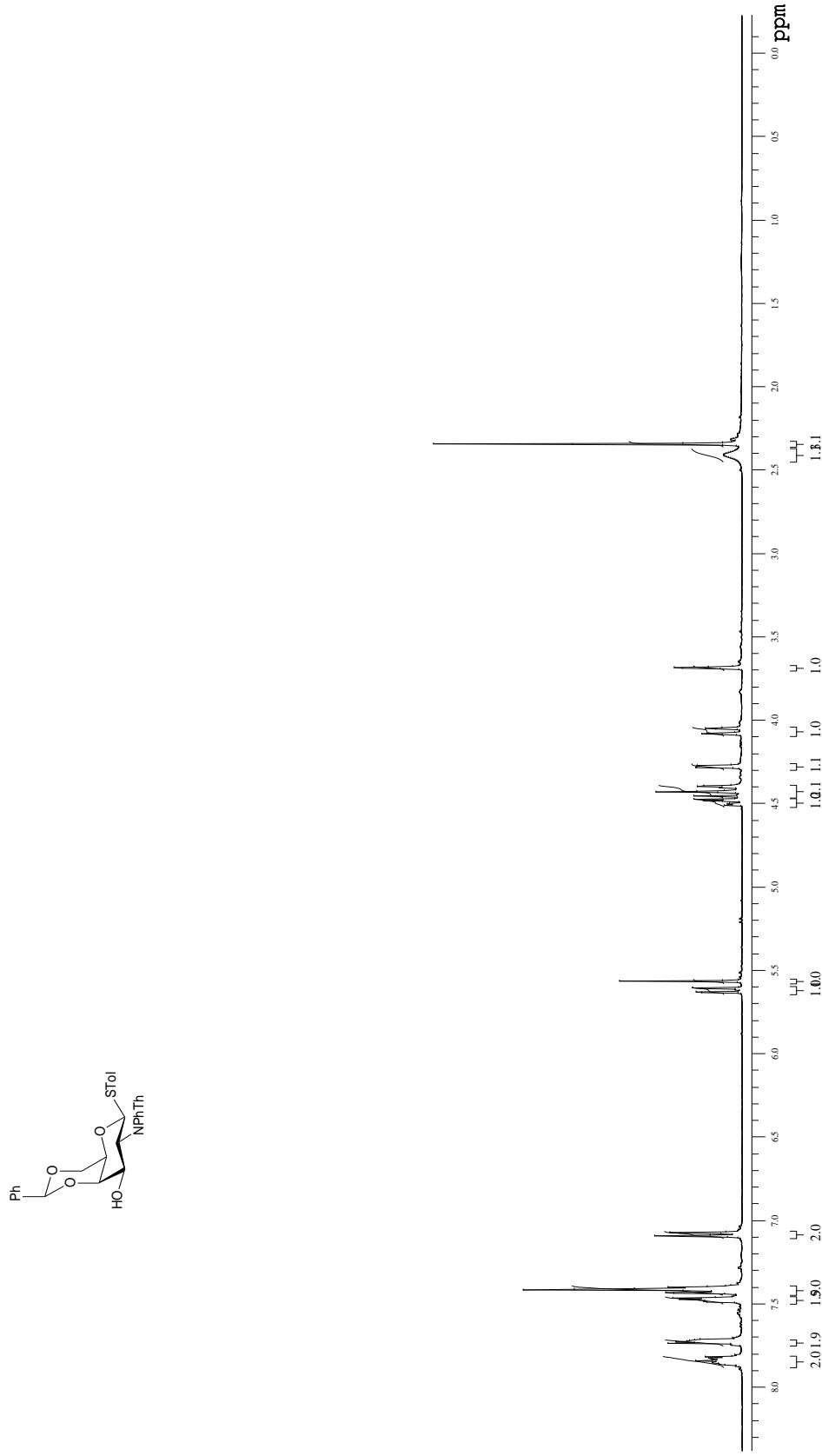
$^{13}\text{C}$  NMR Spectrum of compound **S6** ( $\text{CDCl}_3$ , 400 MHz)

STANDARD 1H OBSERVE  
F1le: xp  
Pulse Sequence: gCOSY



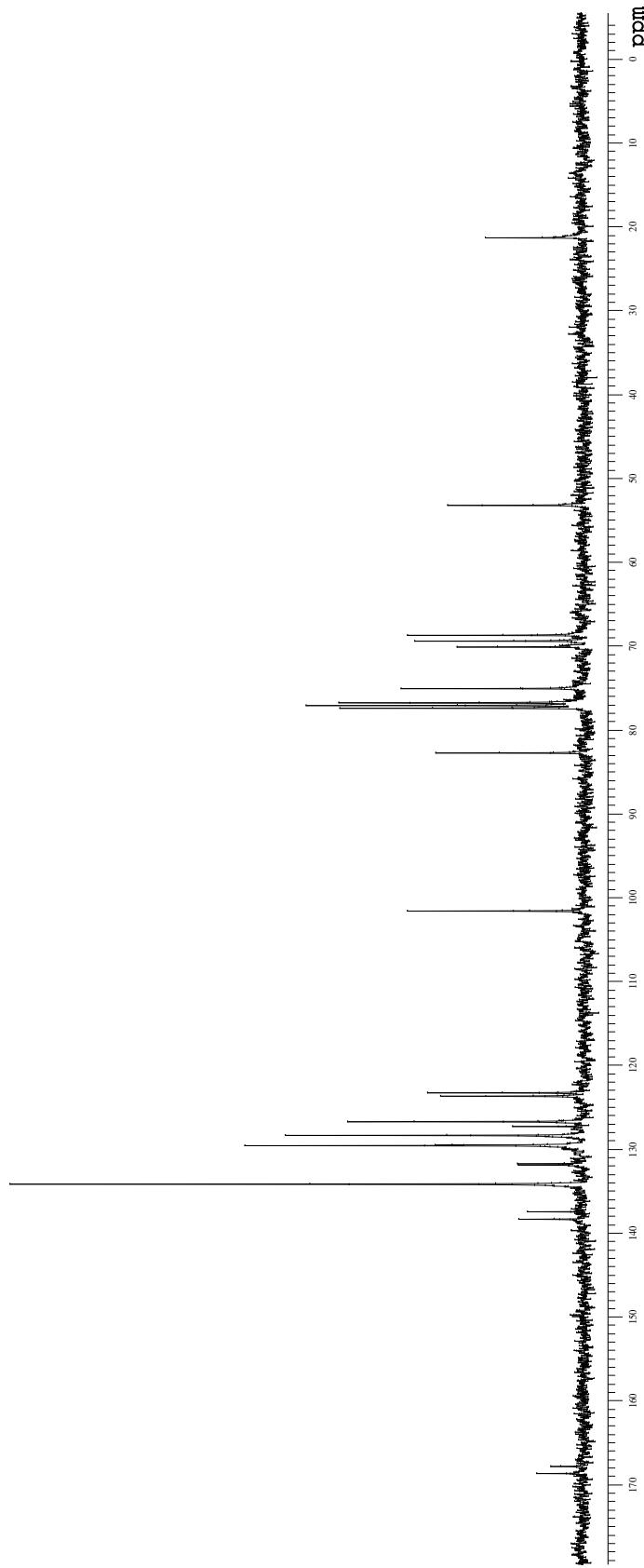
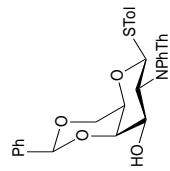
$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **S6** ( $\text{CDCl}_3$ , 400 MHz)

Mercury 400 spectrometer

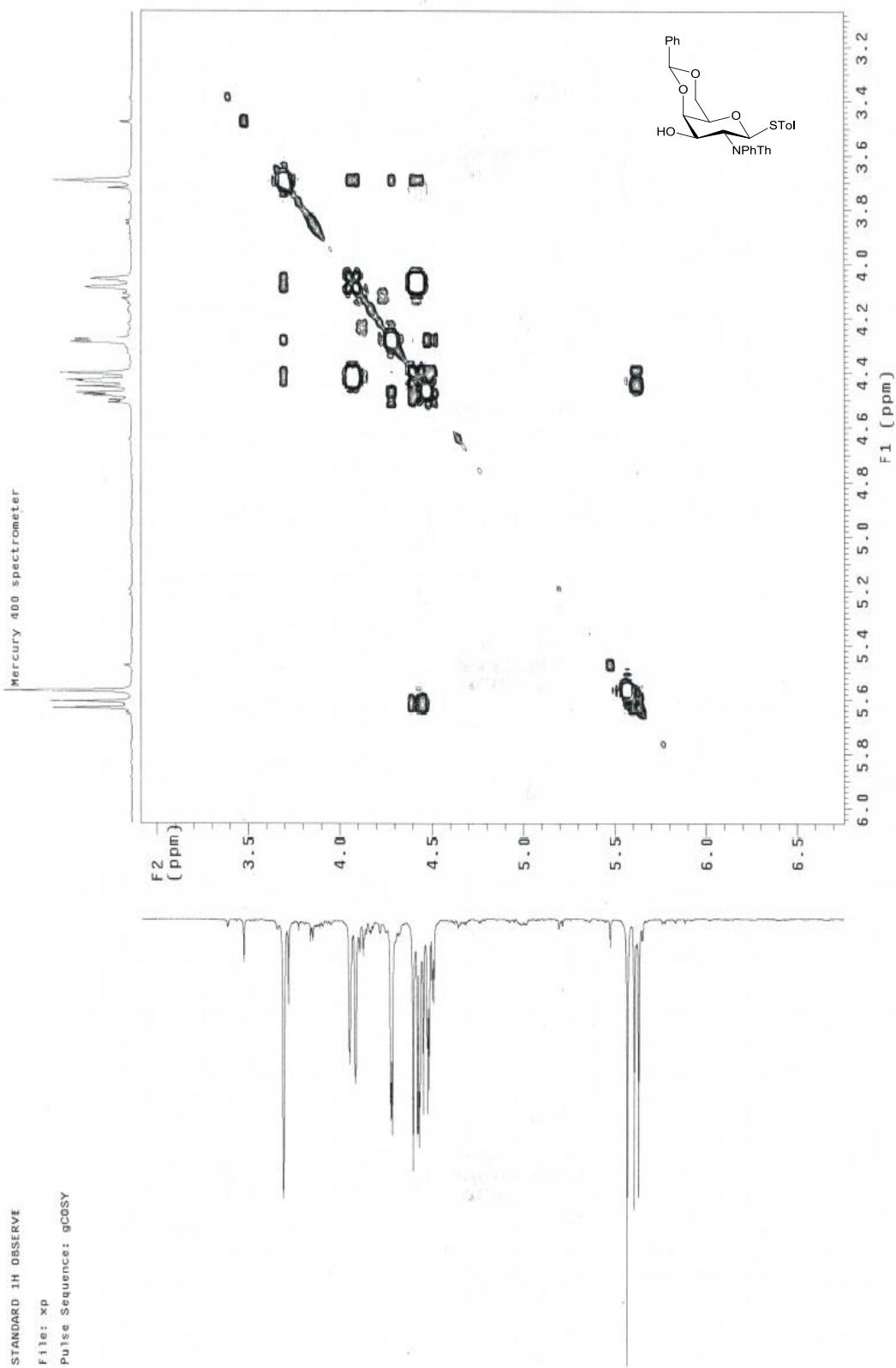


<sup>1</sup>H NMR Spectrum of compound 8 (CDCl<sub>3</sub>, 400 MHz)

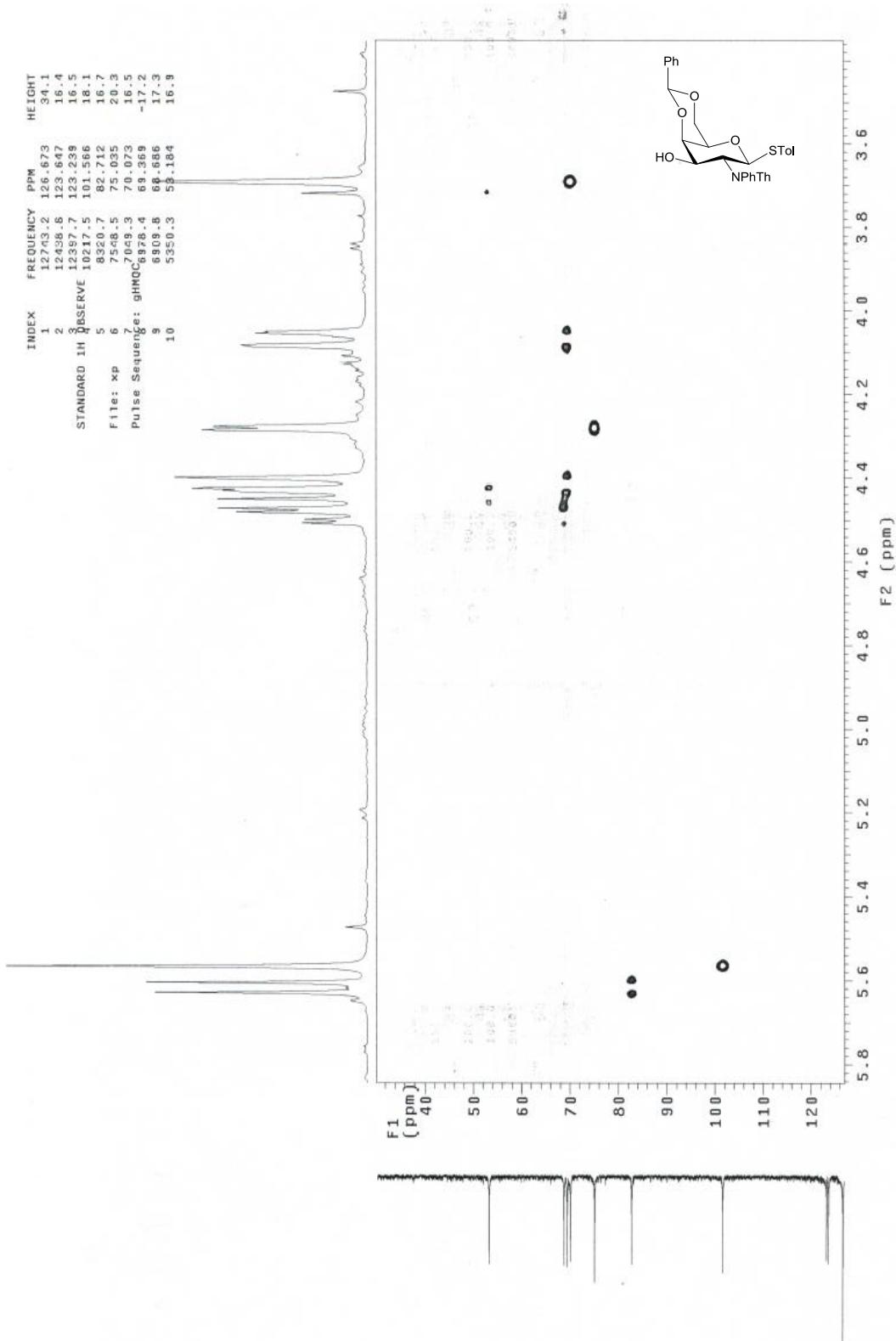
Mercury 400 spectrometer



$^{13}\text{C}$  NMR Spectrum of compound 8 ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **8** ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **8** ( $\text{CDCl}_3$ , 400/100 MHz)

## Elemental Composition Report

Page 1

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

### Monoisotopic Mass, Even Electron Ions

1271 formula(e) evaluated with 7 results within limits (all results (up to 1000) for each mass)

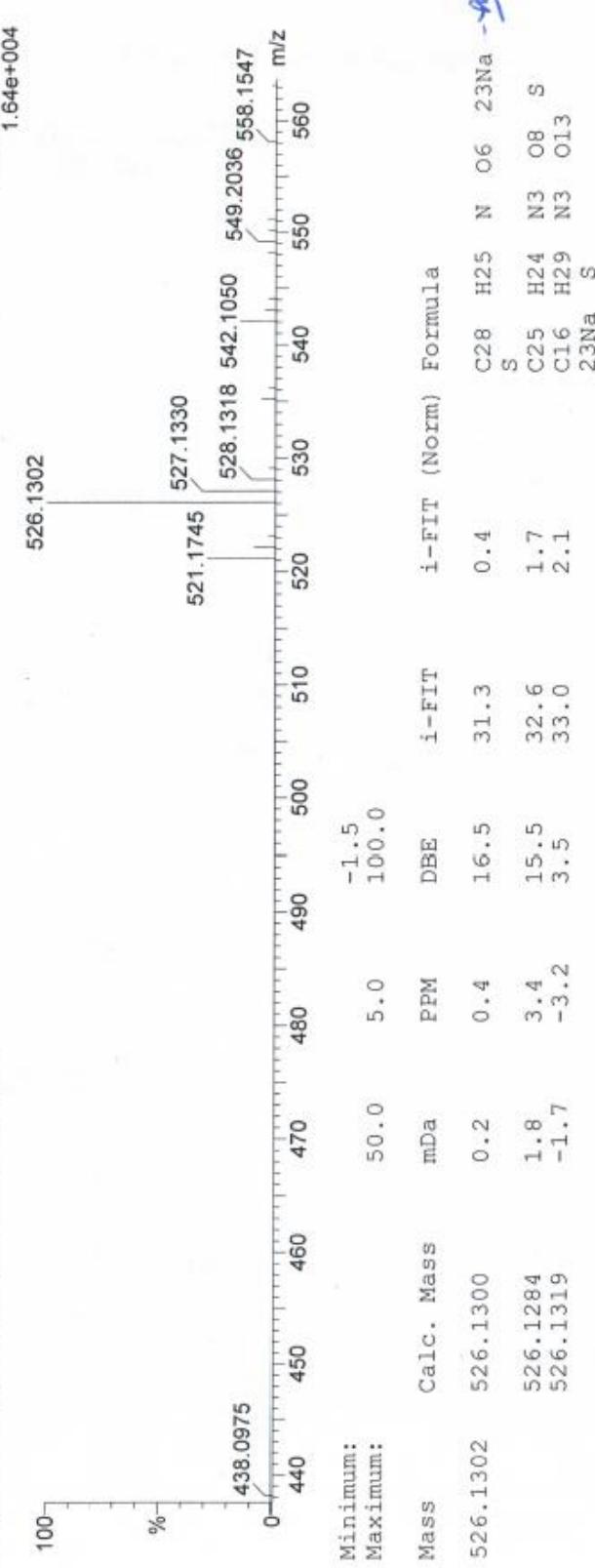
Elements Used:

C: 0-100 H: 0-1000 N: 0-3 O: 0-15 23Na: 0-1 S: 1-2

SATADRUMANDAL

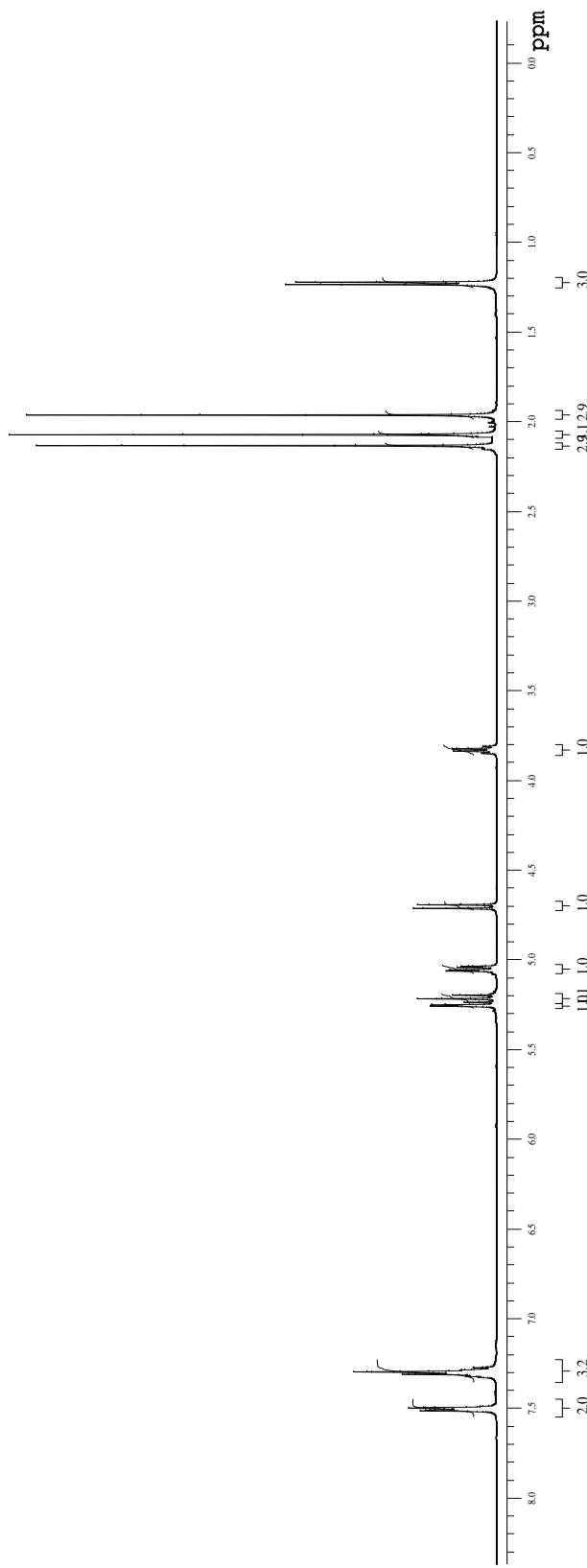
SSM-Gal-Amine-Acceptor

2013\_0130\_3056 20 (0.405) Cm (20:25-1:8x2,000)



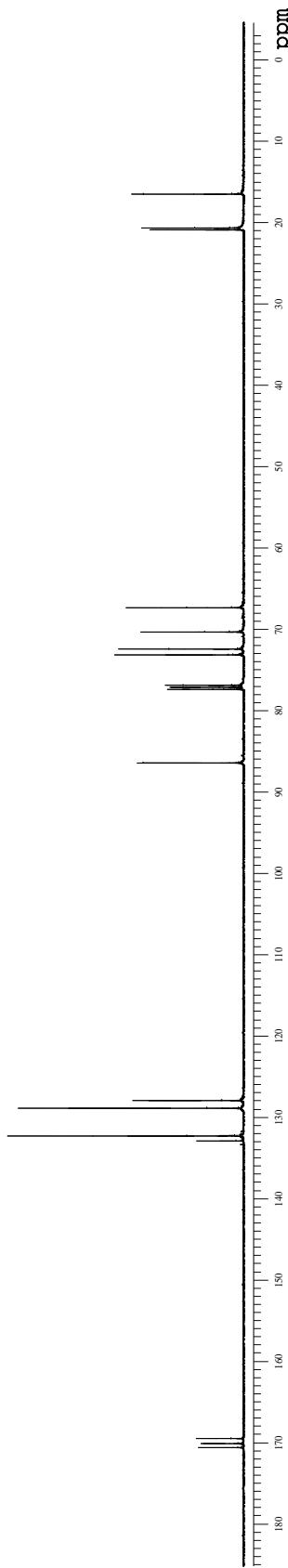
HRMS (ESI MS) spectrum of compound 8

Varian 500 NMR spectrometer

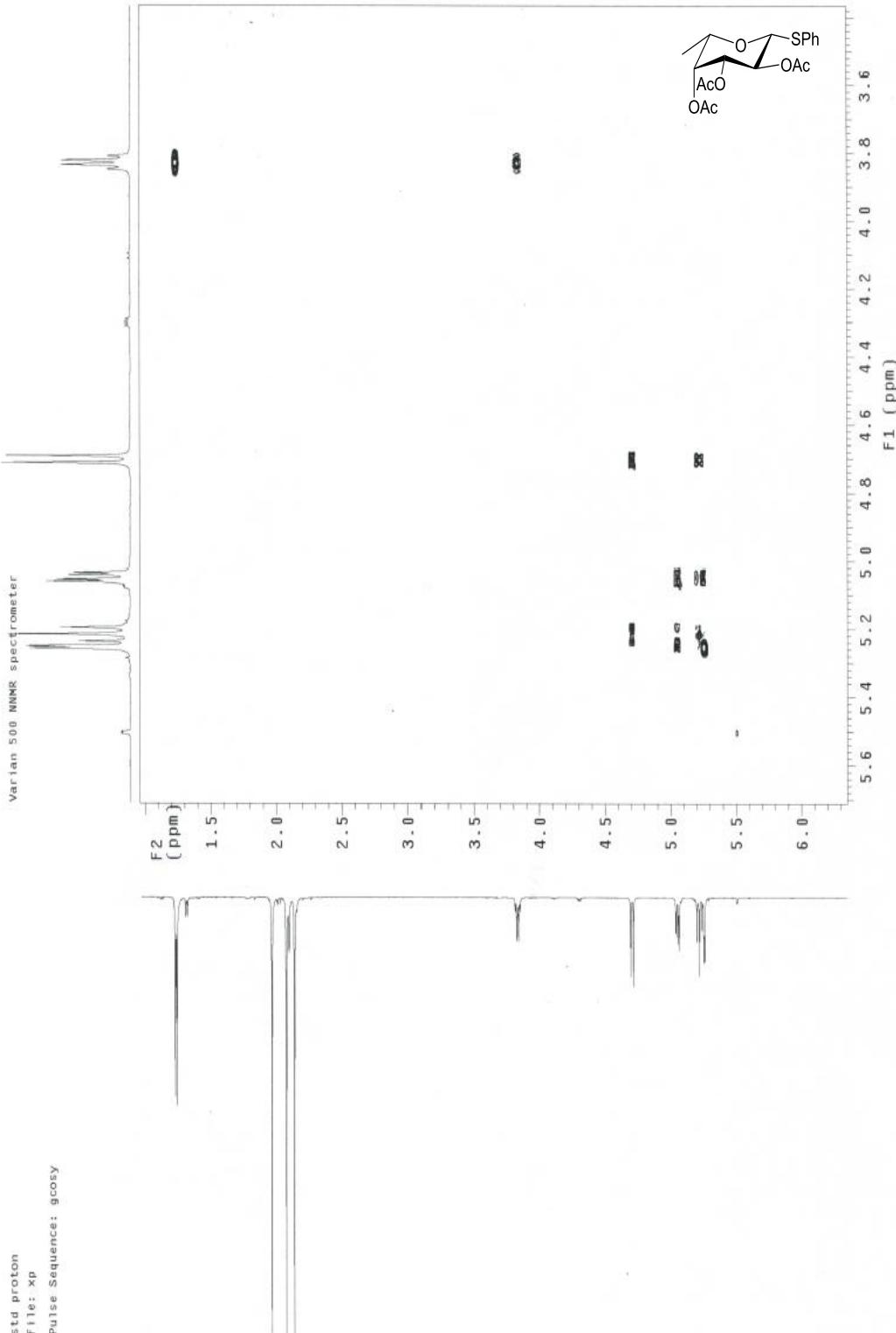


<sup>1</sup>H NMR Spectrum of compound **S8** (CDCl<sub>3</sub>, 500 MHz)

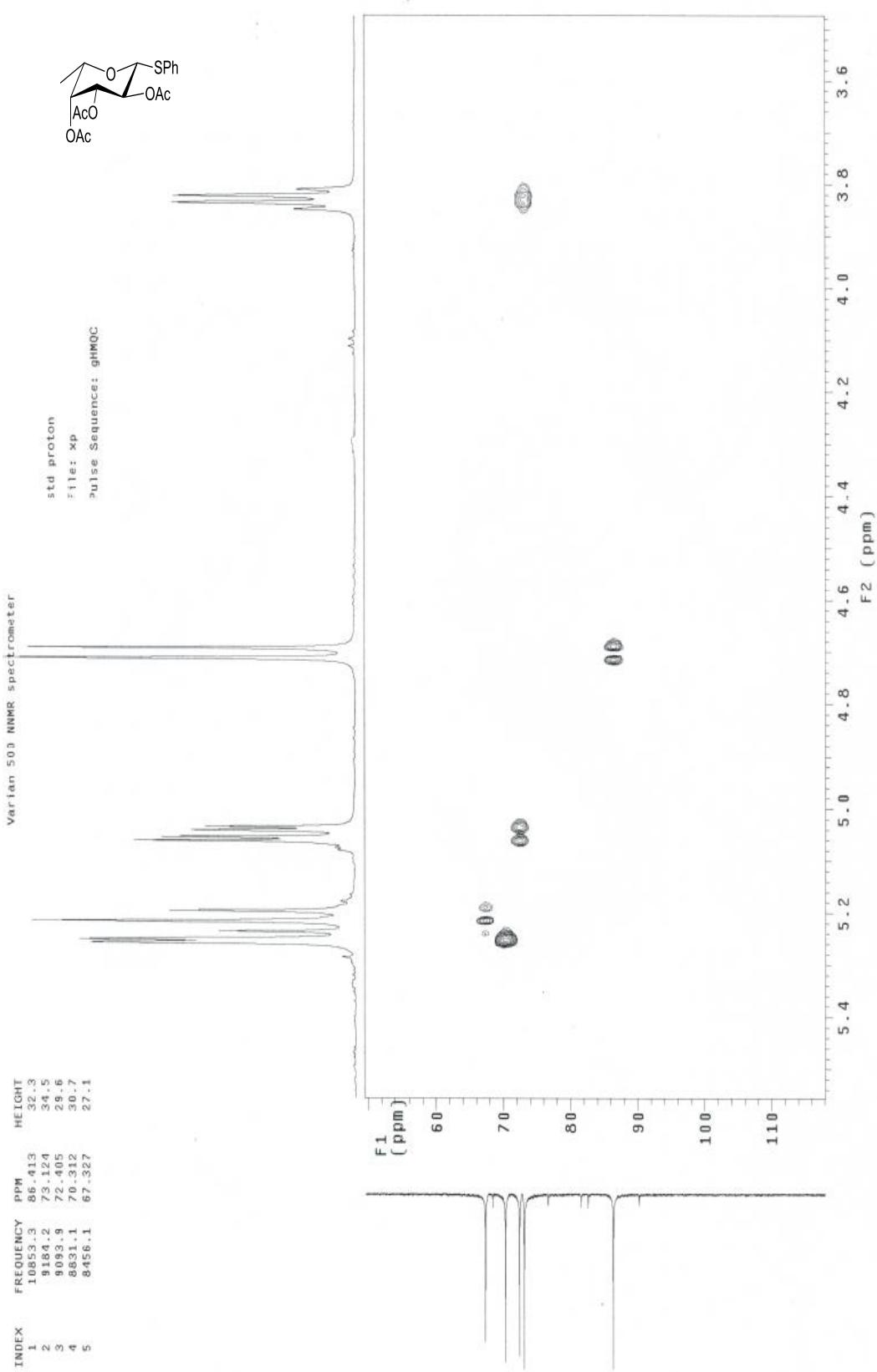
Variian 500 NMR spectrometer



<sup>13</sup>C NMR Spectrum of compound **S8** (CDCl<sub>3</sub>, 500 MHz)

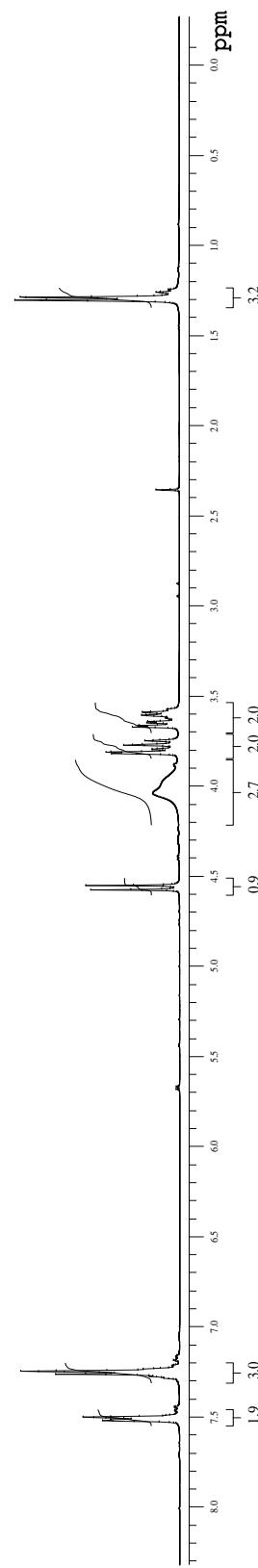


$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **S8** ( $\text{CDCl}_3$ , 500 MHz)



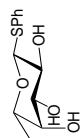
$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **S8** ( $\text{CDCl}_3$ , 500/125 MHz)

Varian MR-400 NMR spectrometer

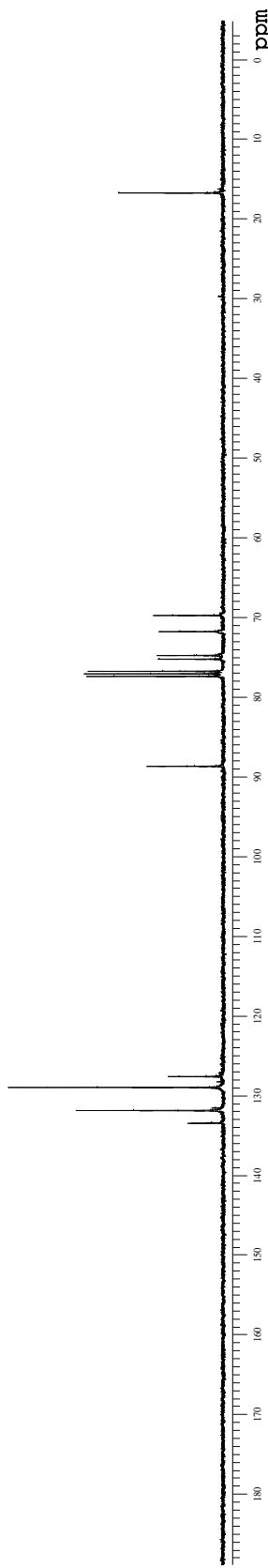


<sup>1</sup>H NMR Spectrum of compound **S9** (CDCl<sub>3</sub>, 400 MHz)

Varian MR-400 NMR spectrometer



$^{13}\text{C}$  NMR Spectrum of compound **S9** ( $\text{CDCl}_3$ , 400 MHz)

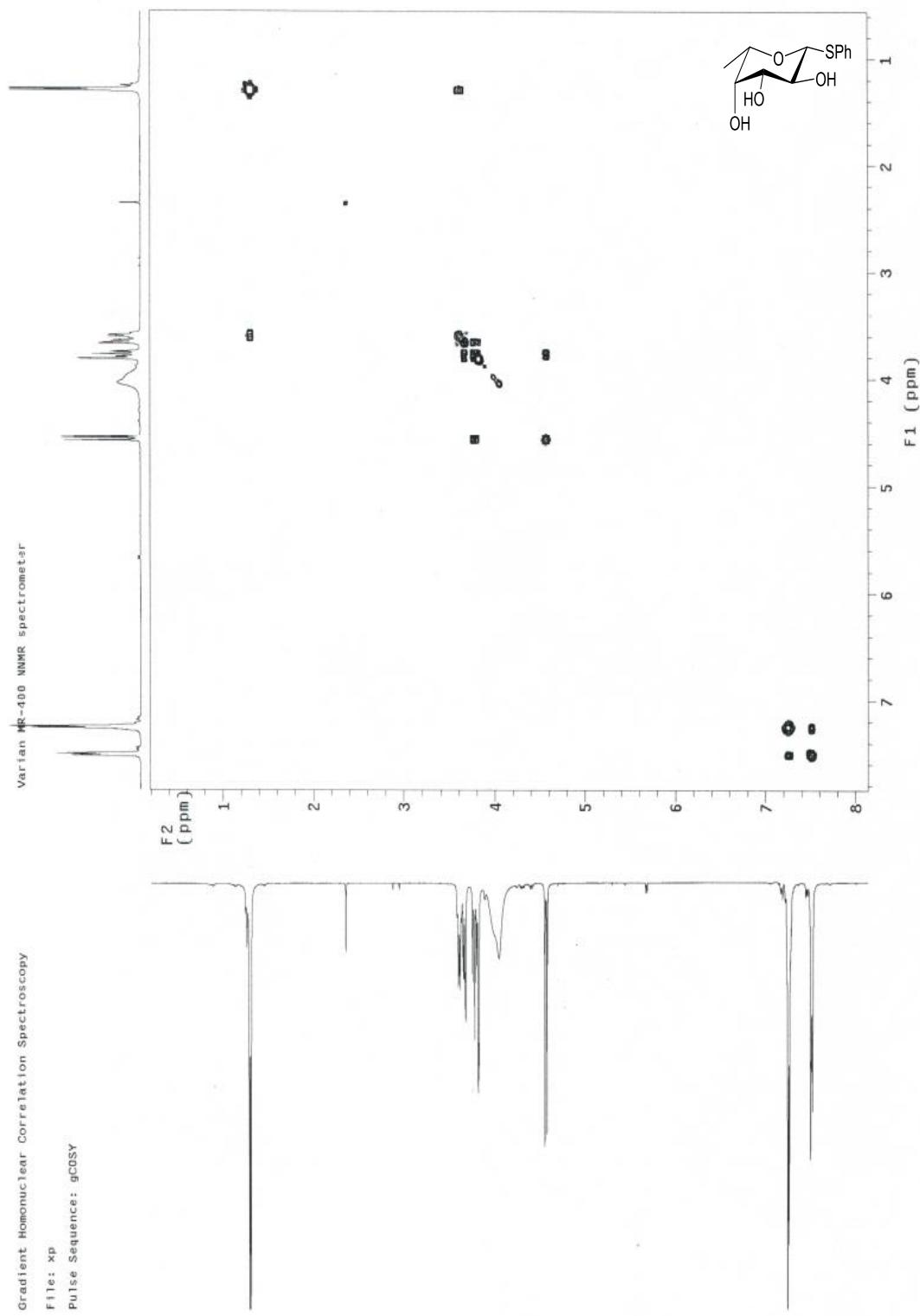


Gradient Homonuclear Correlation Spectroscopy

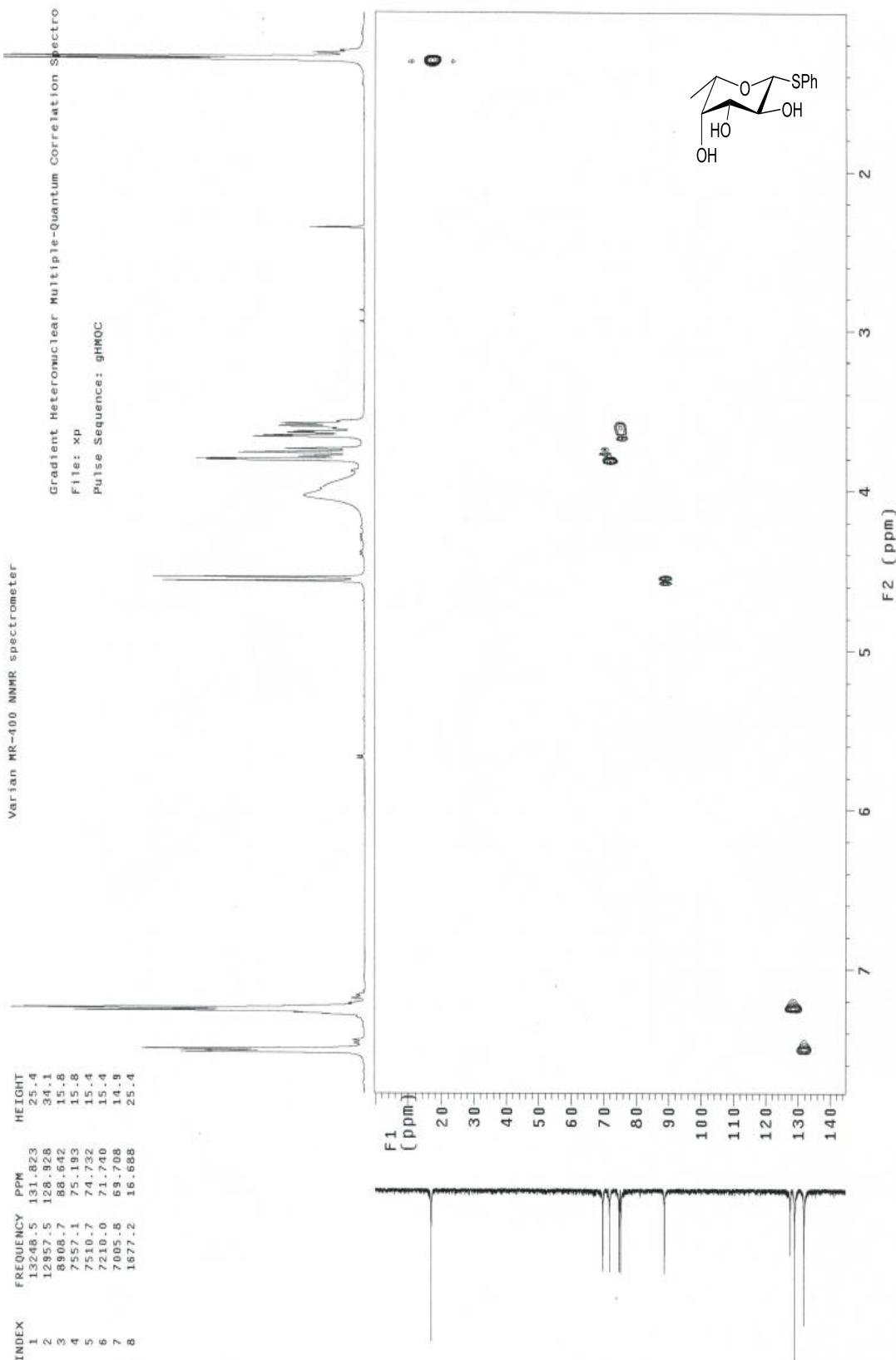
File: xp

Pulse Sequence: gCOSY

Varian MR-400 NMR spectrometer

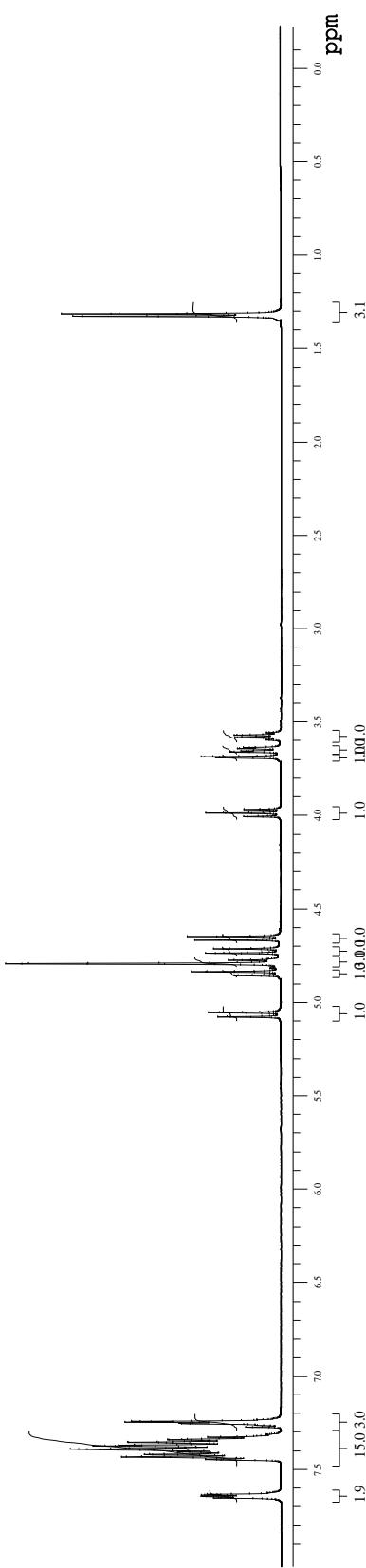
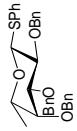


$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **S9** ( $\text{CDCl}_3$ , 400 MHz)



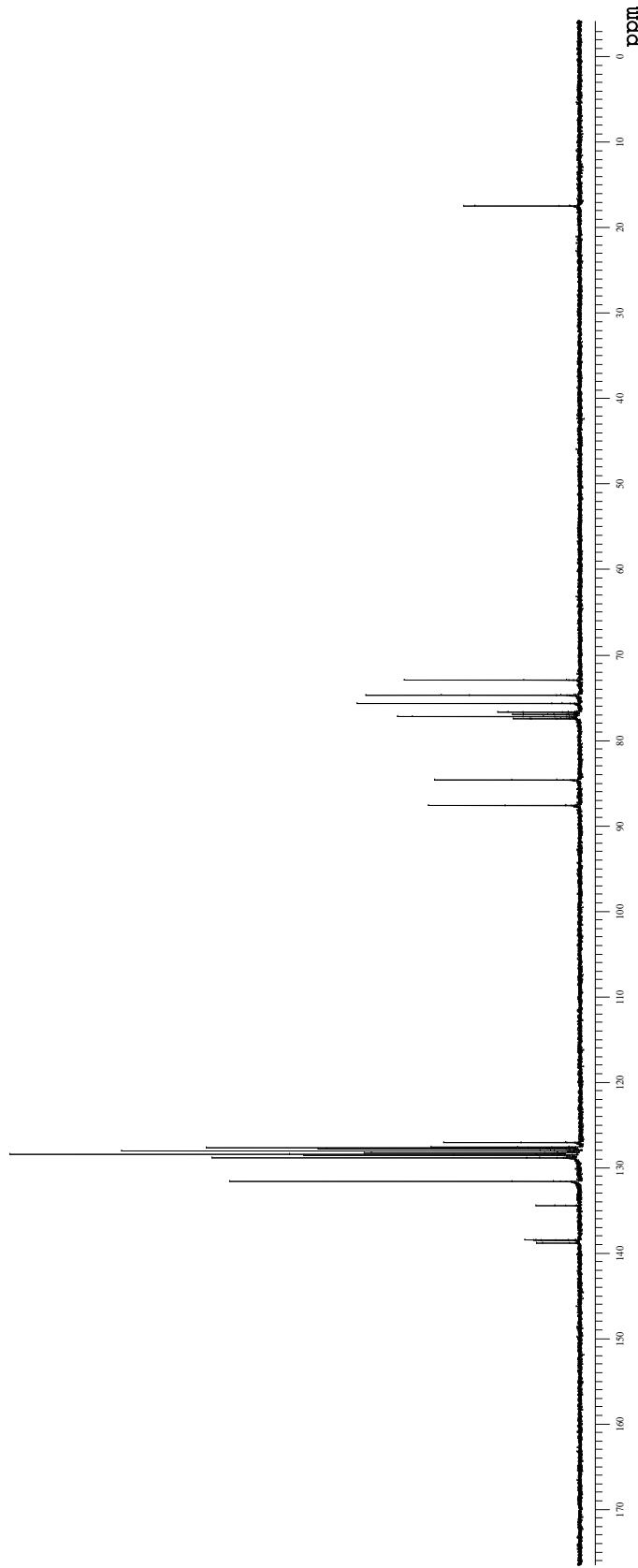
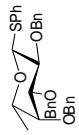
$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **S9** ( $\text{CDCl}_3$ , 400/100 MHz)

Varian 500 NMR spectrometer

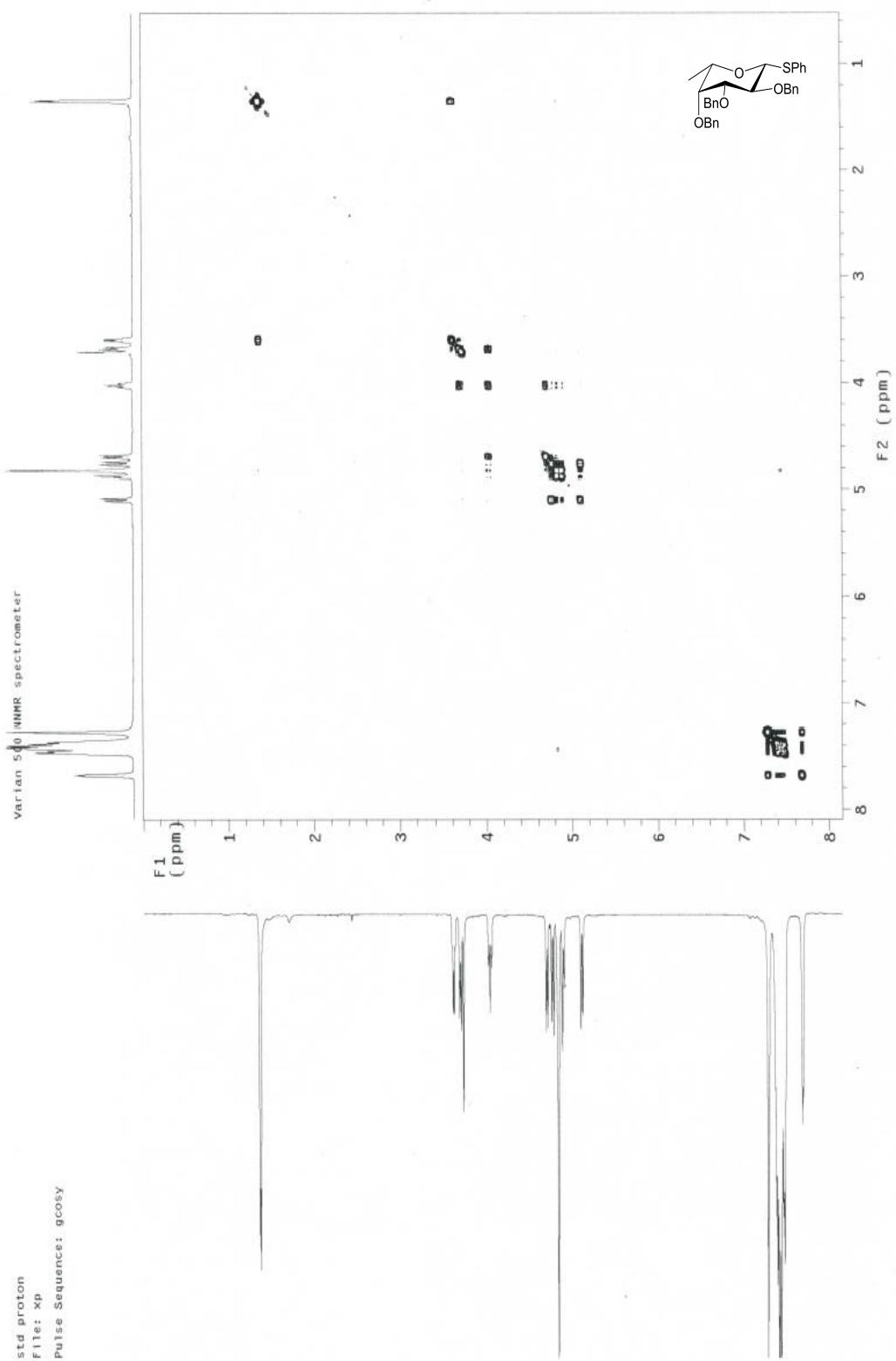


<sup>1</sup>H NMR Spectrum of compound **4** (CDCl<sub>3</sub>, 500 MHz)

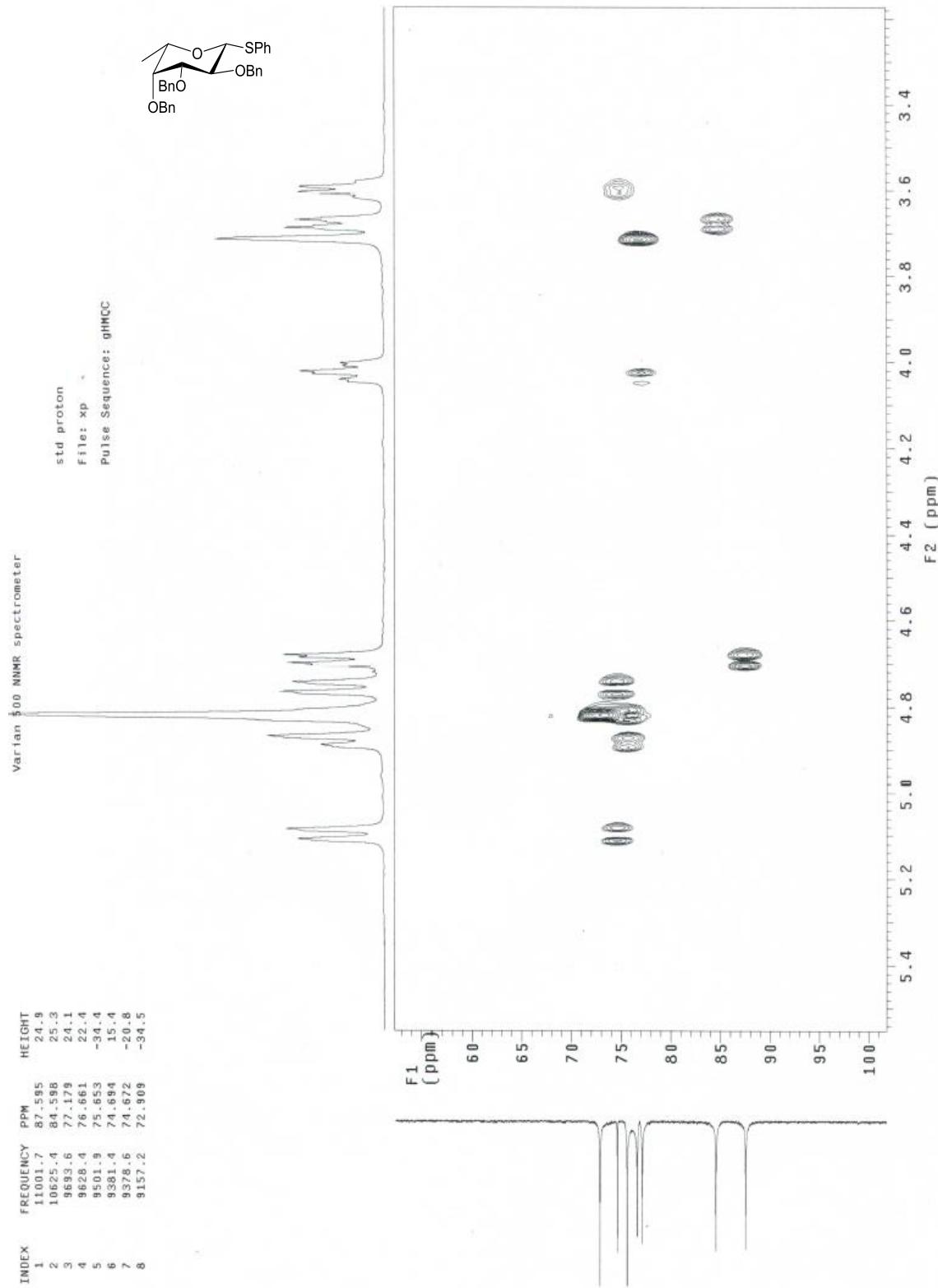
Varian 500 NMR spectrometer



$^{13}\text{C}$  NMR Spectrum of compound 4 ( $\text{CDCl}_3$ , 500 MHz)



$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **4** ( $\text{CDCl}_3$ , 500 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound 4 ( $\text{CDCl}_3$ , 500/125 MHz)

## Elemental Composition Report

Page 1

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

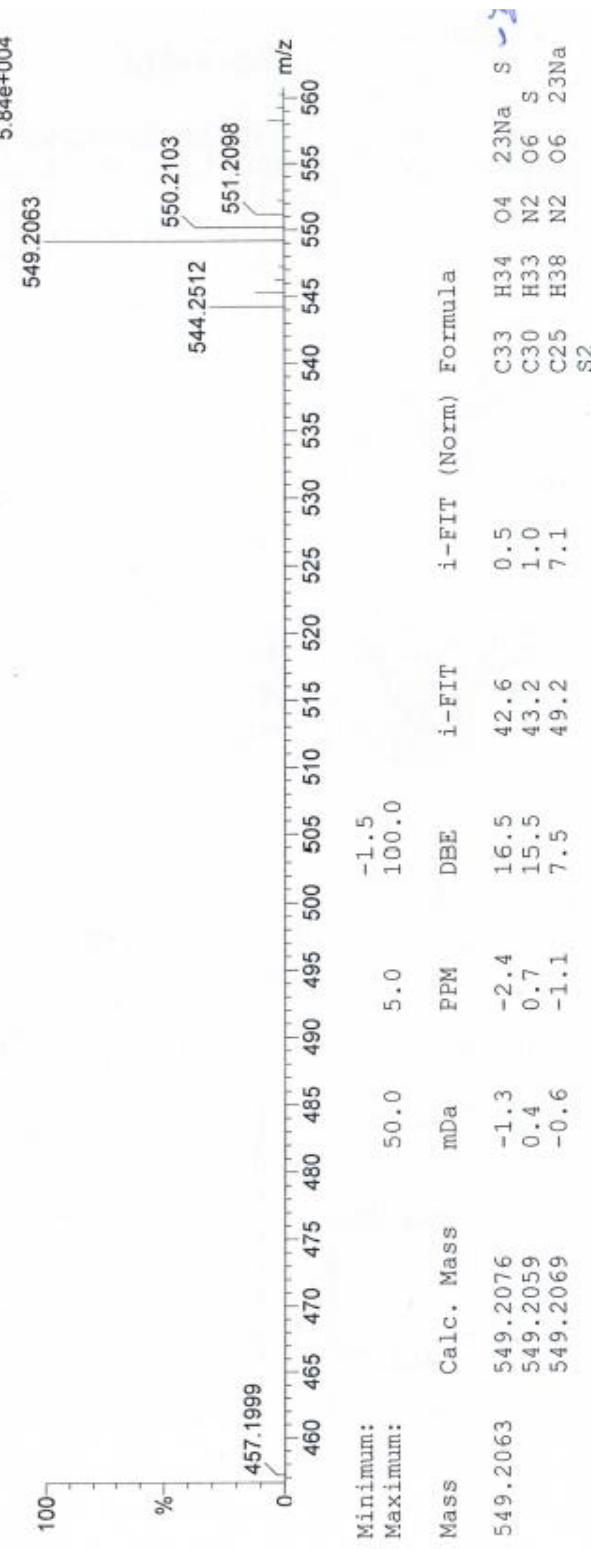
Monoisotopic Mass, Even Electron Ions

1007 formula(e) evaluated with 3 results within limits (all results (up to 1000) for each mass)

Elements Used:

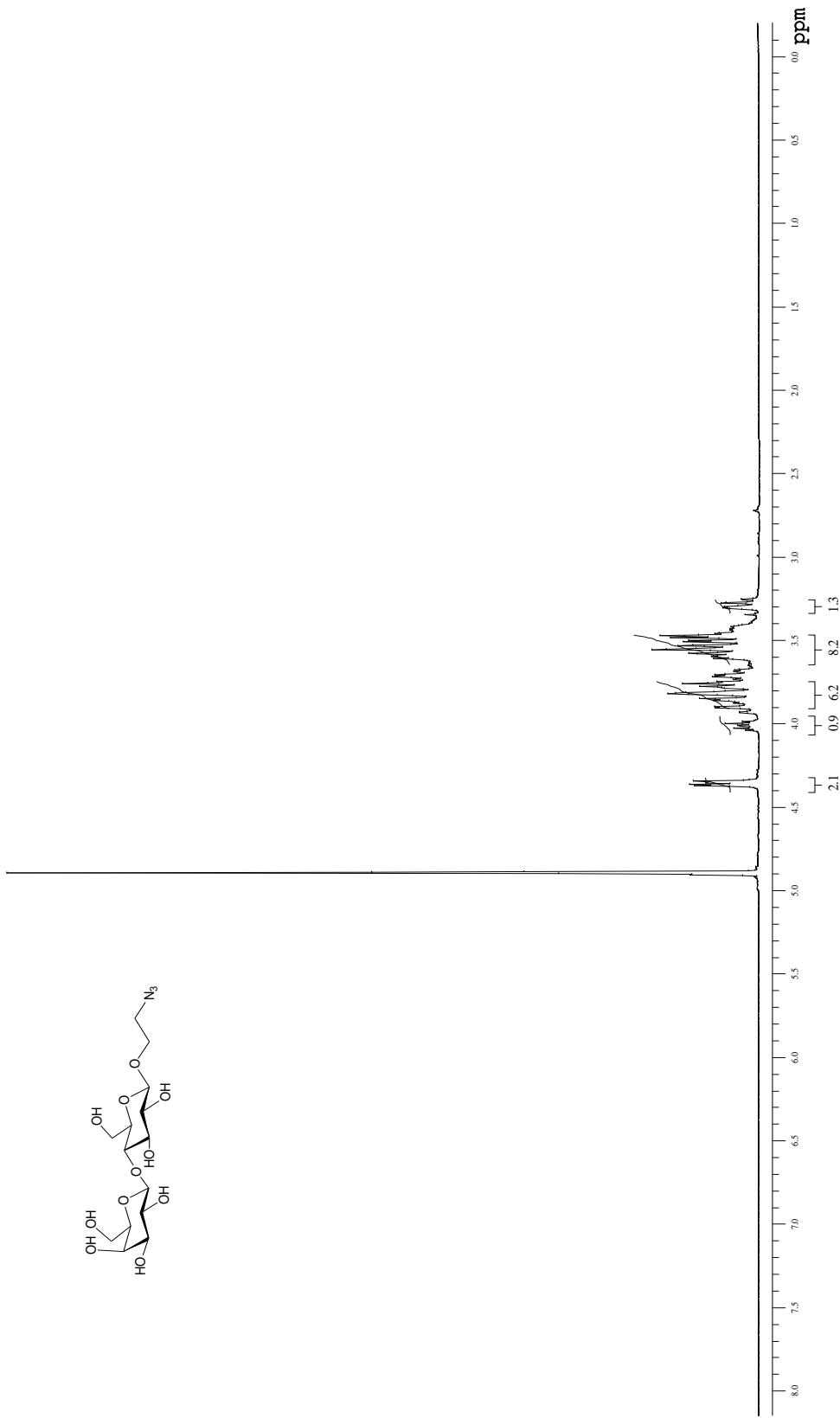
C: 0-100 H: 0-1000 N: 0-3 O: 0-10 23Na: 0-1 S: 1-2  
SSM-F-SPh  
SATADRU MANDAL

2013\_0131\_3060 22 (0.460) Cm (22:29:1.8x2.000)



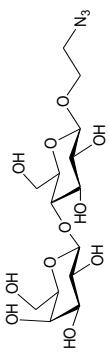
HRMS (ESI MS) spectrum of compound 4

Varian 500 NMR spectrometer

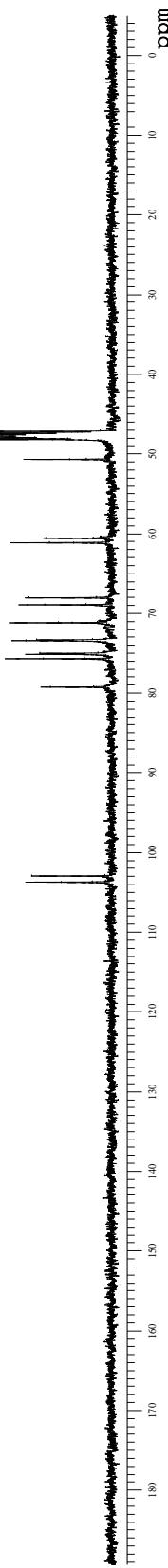


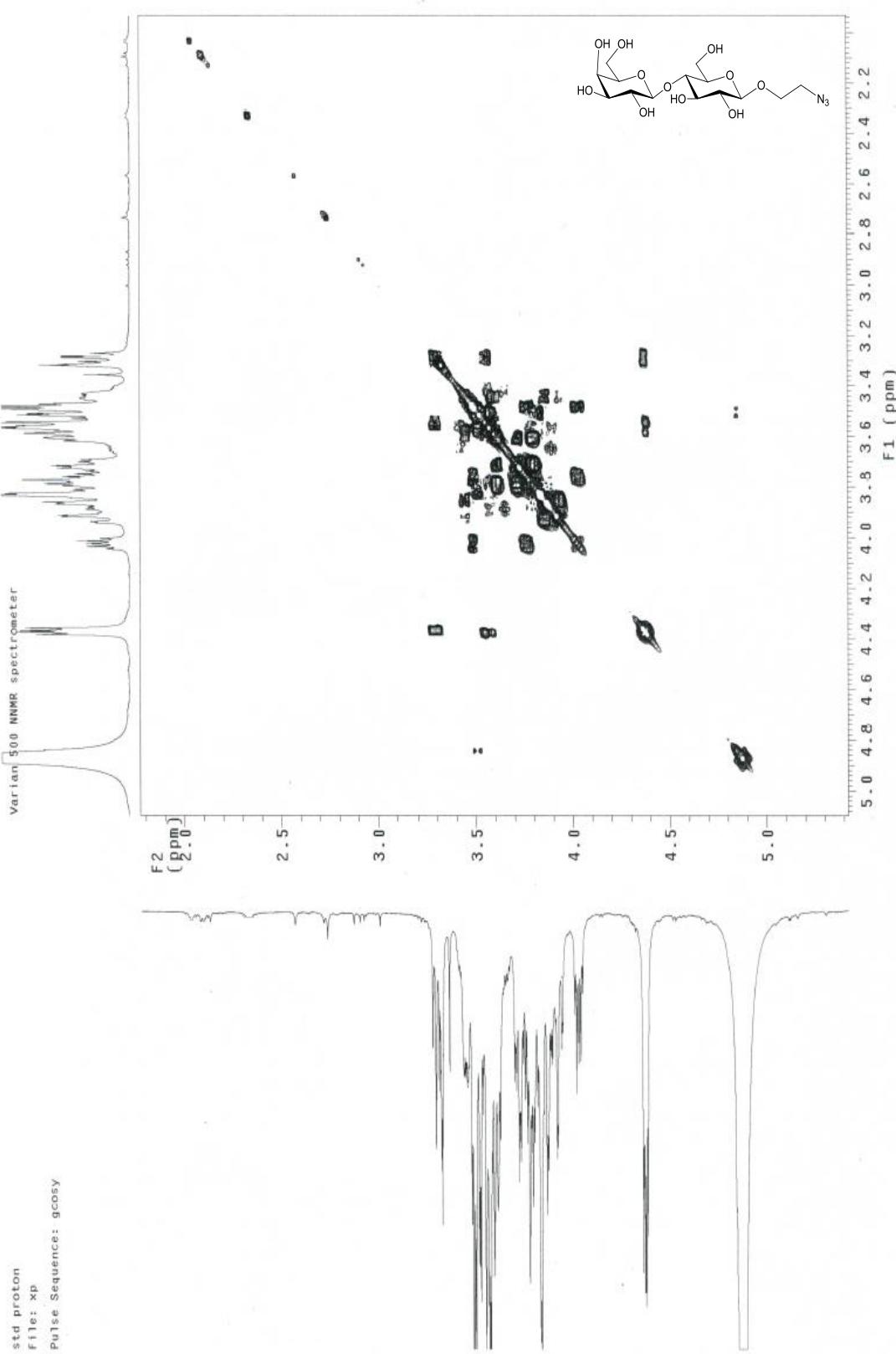
<sup>1</sup>H NMR Spectrum of compound **15** (CD<sub>3</sub>OD, 500 MHz)

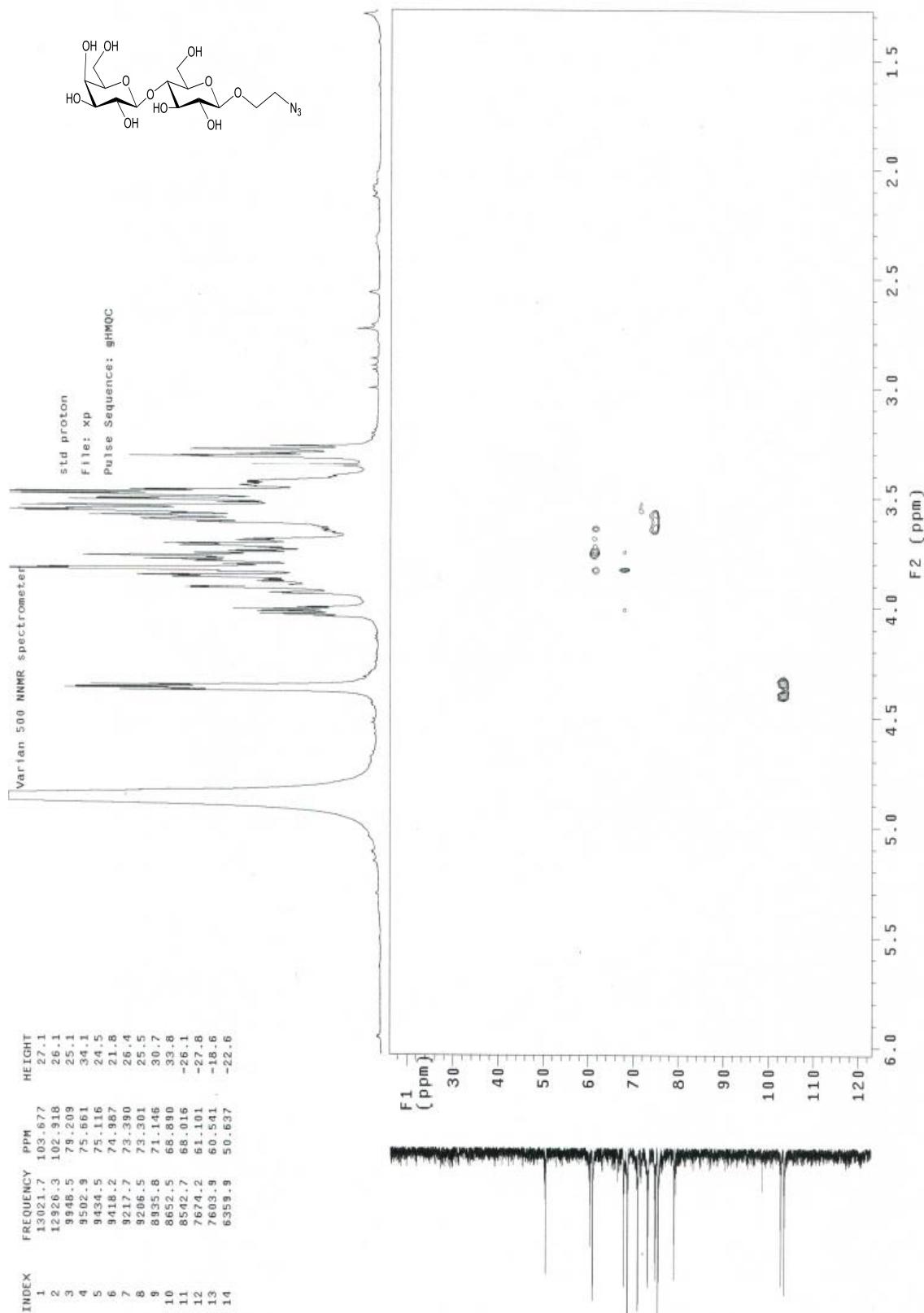
Varian 500 NMR spectrometer



<sup>13</sup>C NMR Spectrum of compound **15** (CD<sub>3</sub>OD, 500 MHz)

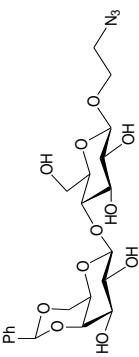






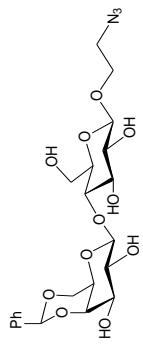
$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **15** ( $\text{CD}_3\text{OD}$ , 500/125 MHz)

Varian MR-400 NMR spectrometer

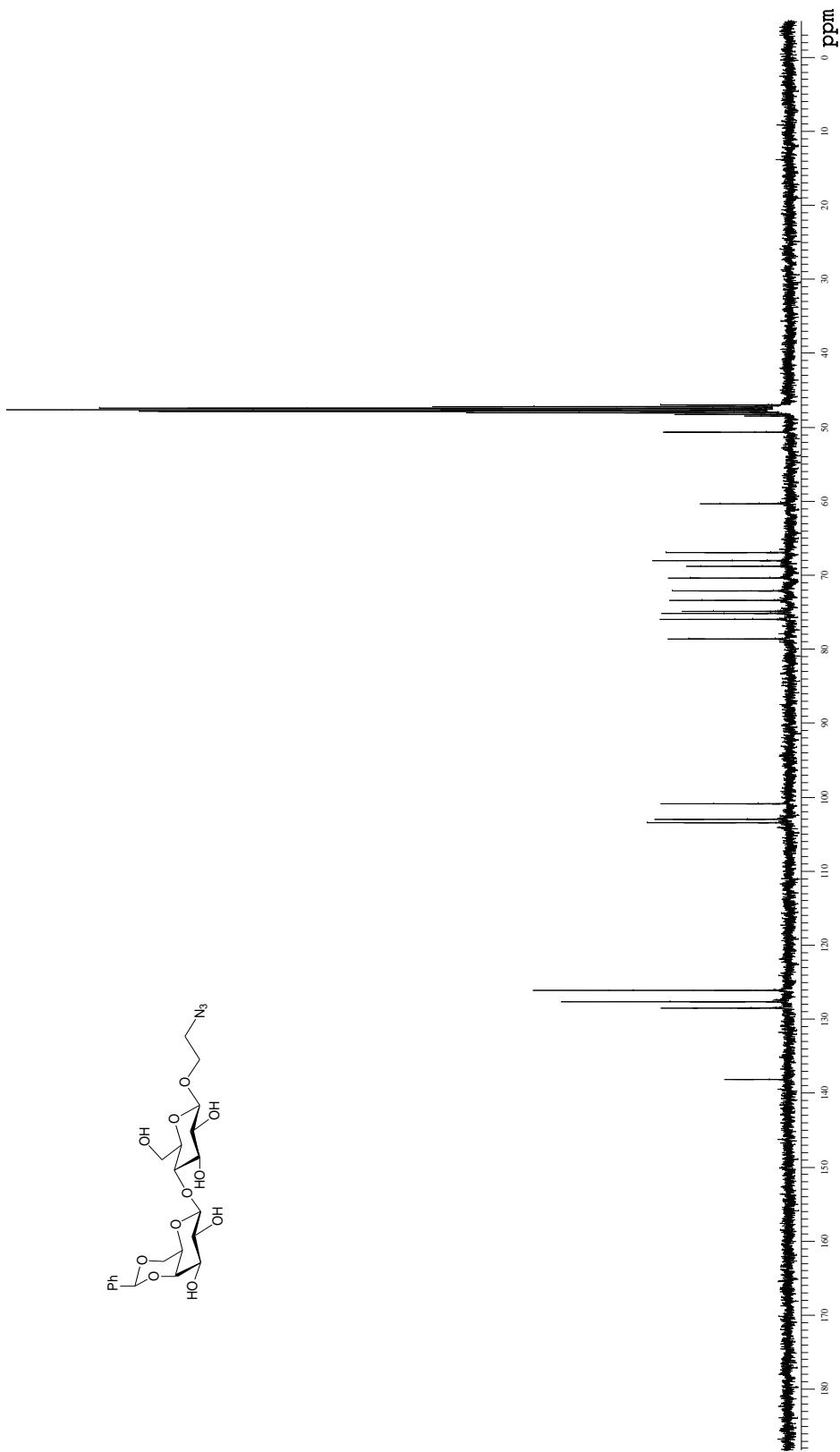


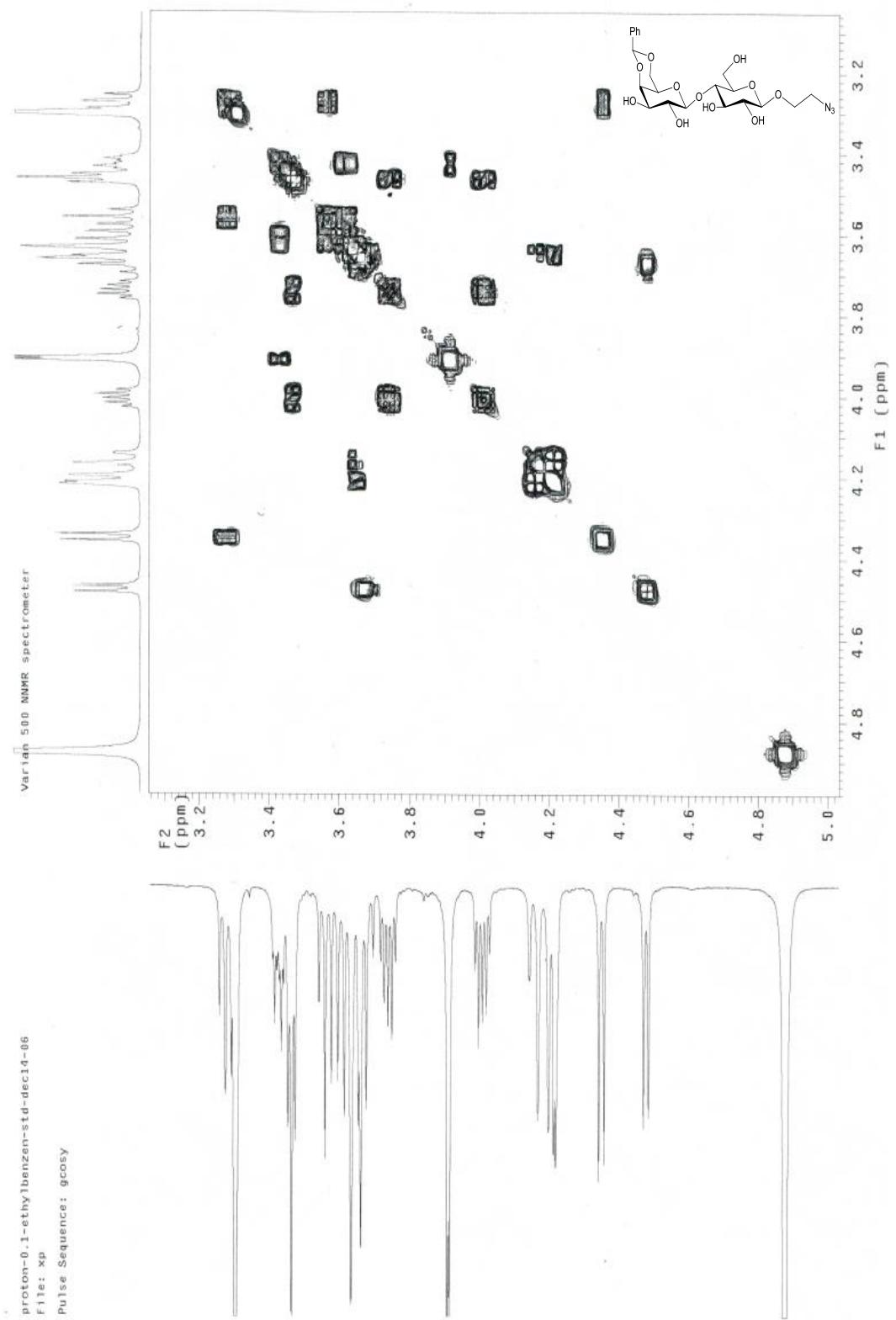
<sup>1</sup>H NMR Spectrum of compound **16** (CD<sub>3</sub>OD, 400 MHz)

Varian MR-400 NMR spectrometer

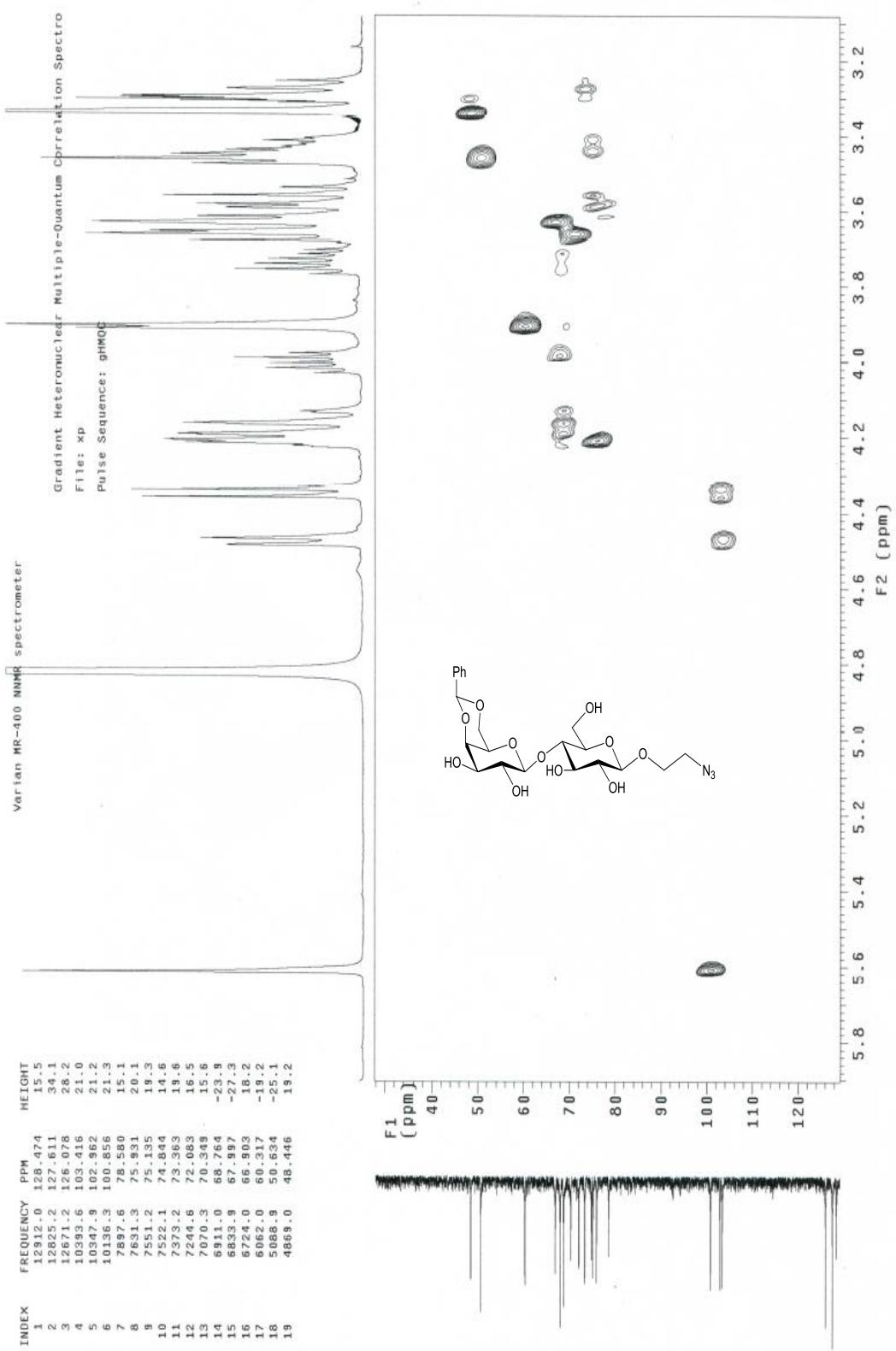


$^{13}\text{C}$  NMR Spectrum of compound **16** ( $\text{CD}_3\text{OD}$ , 400 MHz)

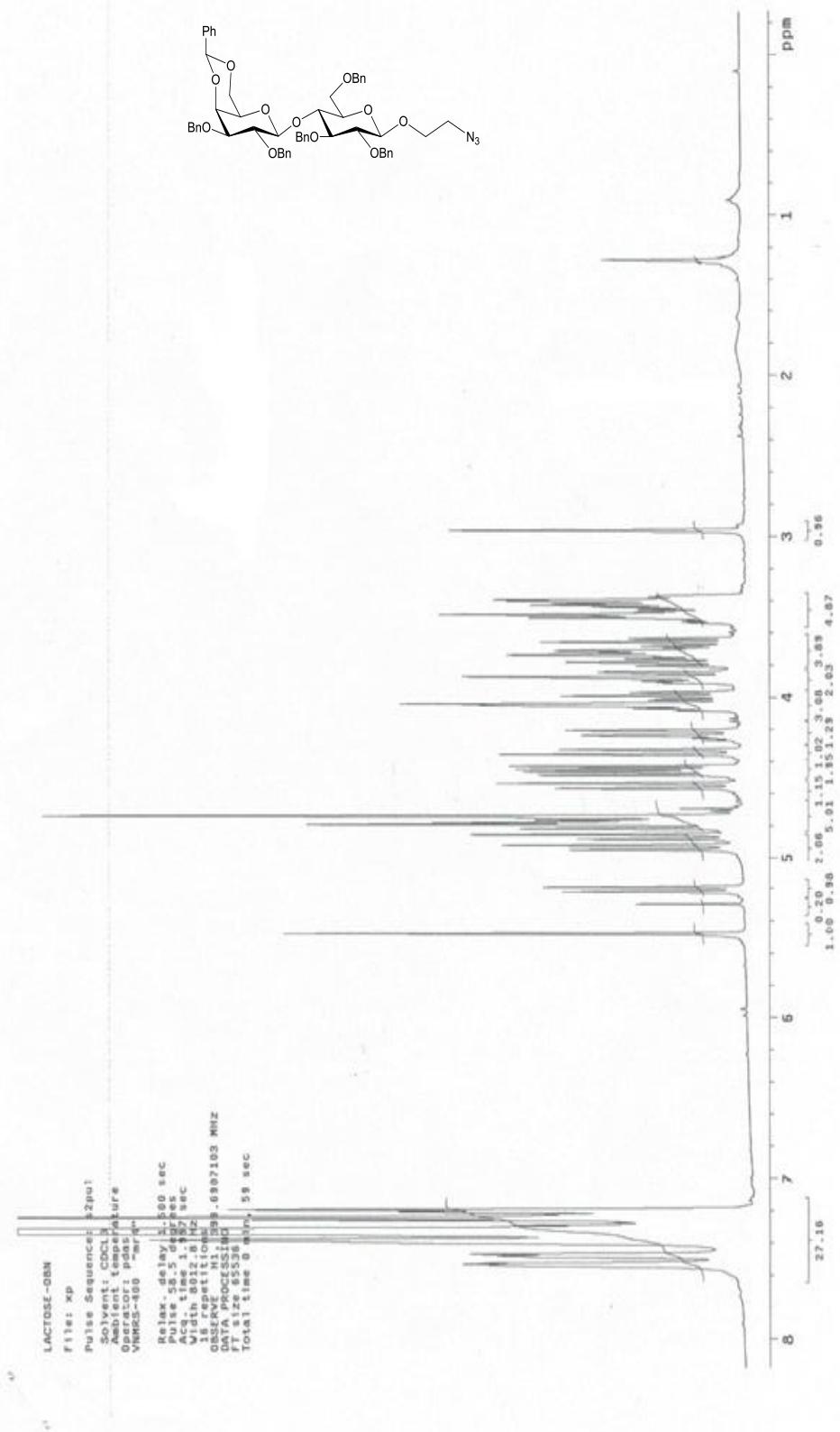




$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **16** ( $\text{CD}_3\text{OD}$ , 500 MHz)



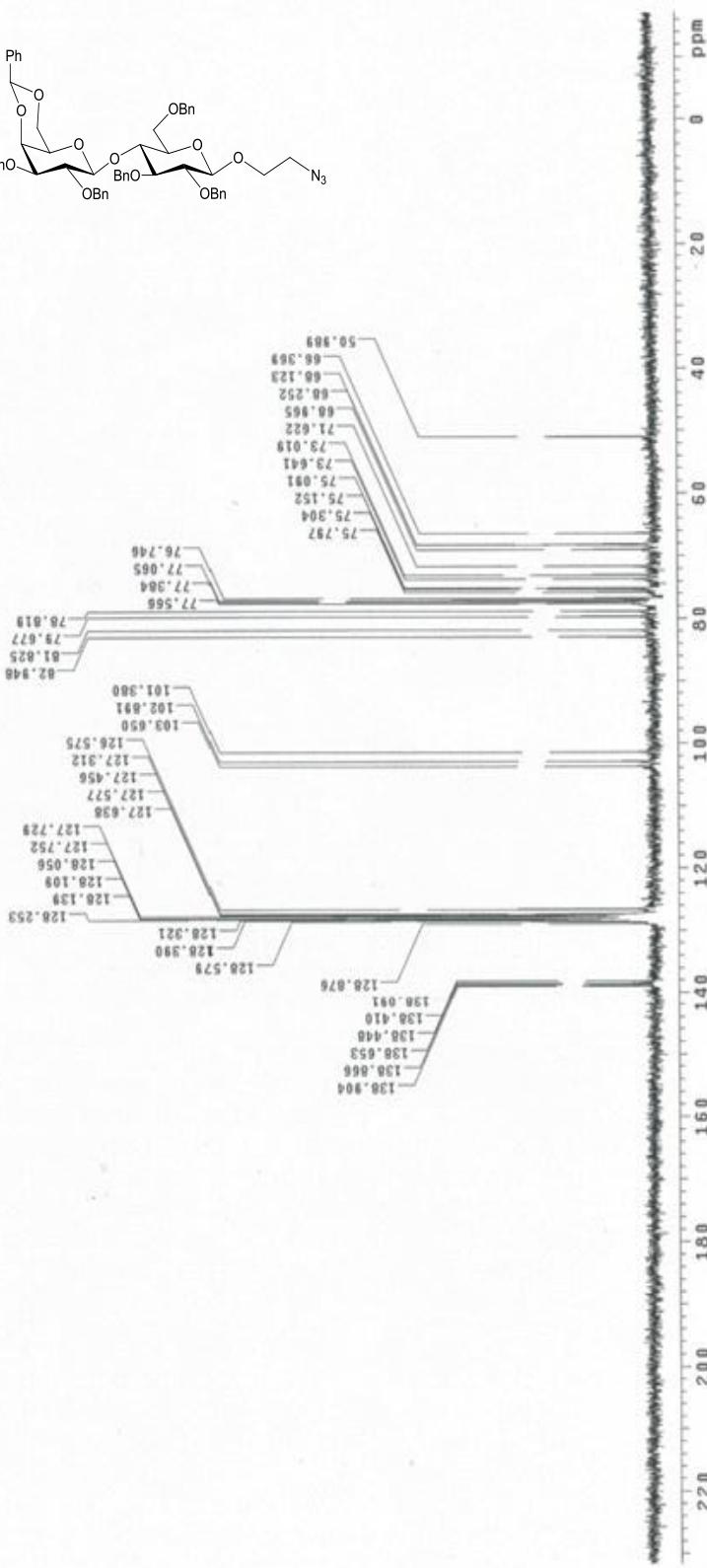
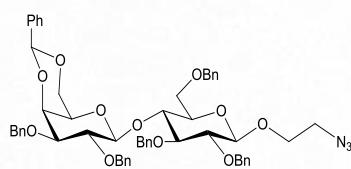
<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **16** (CD<sub>3</sub>OD, 400/100 MHz)



<sup>1</sup>H NMR Spectrum of compound 17 (CDCl<sub>3</sub>, 400 MHz)

Varian NMR-400 NMR spectrometer

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<sup>13</sup>C NMR Spectrum of compound **17** (CDCl<sub>3</sub>, 400 MHz)

Gradient Homonuclear Correlation Spectroscopy

File: xp

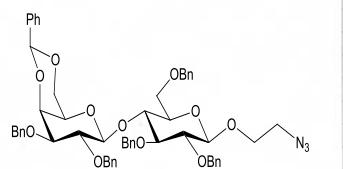
Pulse Sequence: gCOSY

Variian MR-400 NMR spectrometer

F2 (ppm)

2.5  
3.0  
3.5  
4.0  
4.5  
5.0  
5.5  
6.0

6.0  
5.5  
5.0  
4.5  
4.0  
3.5  
3.0  
2.5



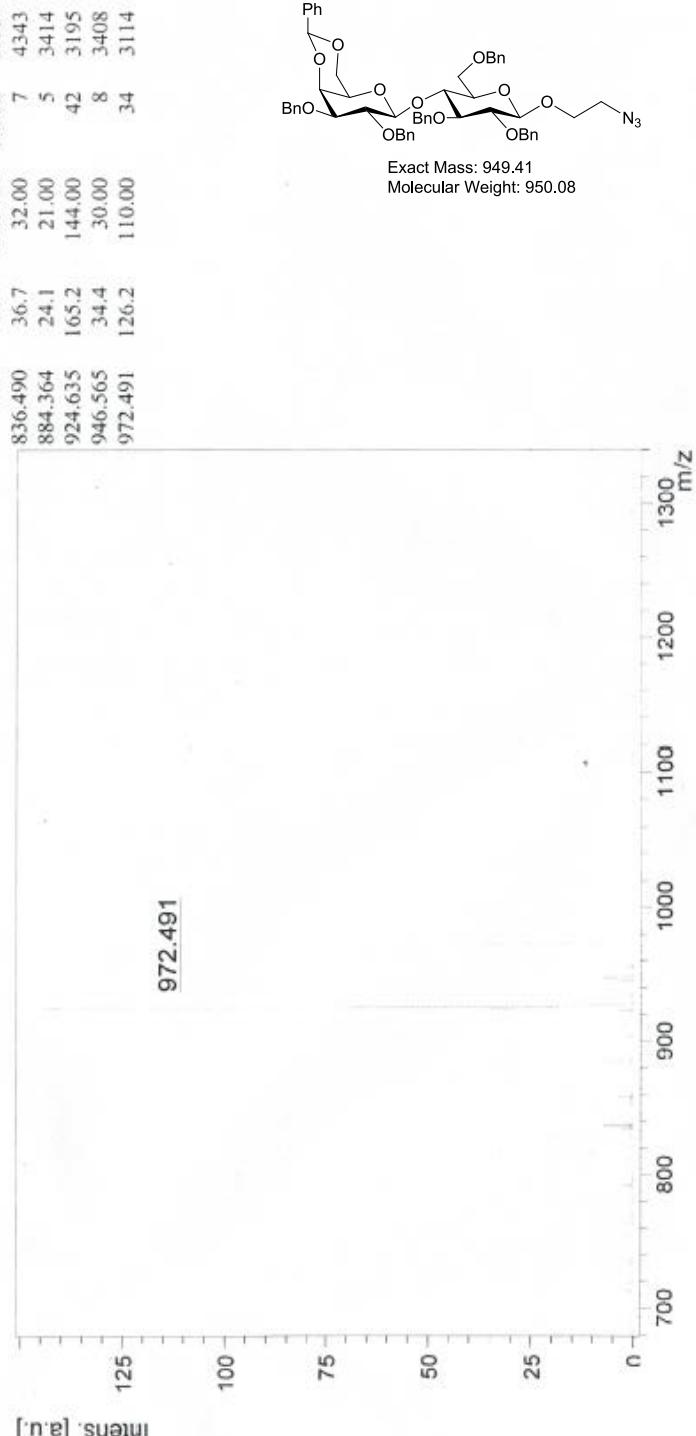
6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5

F1 (ppm)

$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **17** ( $\text{CDCl}_3$ , 400 MHz)

D:\Data\Guo\_lab\SS Mandal\L-OBn\L-OBn

Comment 1                    DHB-Positive-Method  
Comment 2                    L-OBn

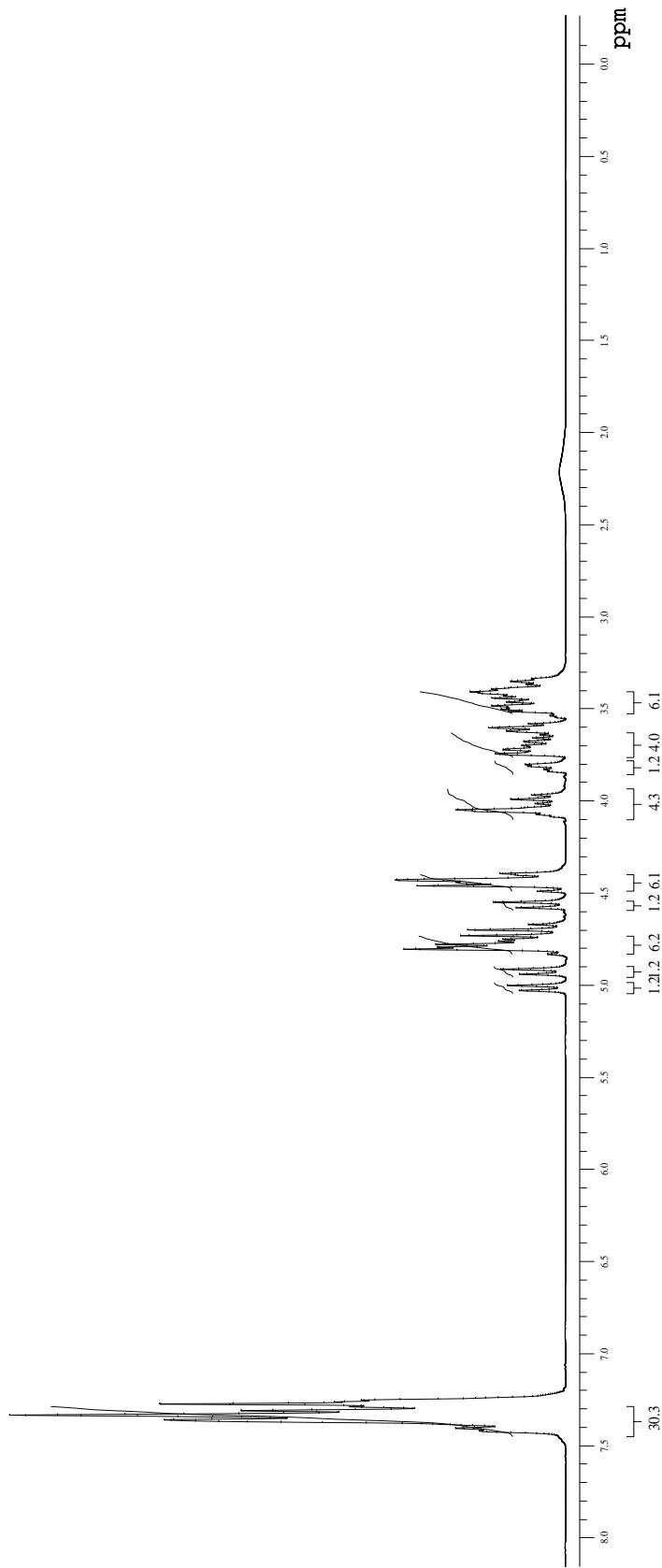
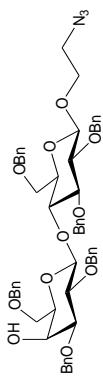


#### Acquisition Parameter

D:\Methods\1User\_Methods\Guo\RP\_3147\_PepMixDHB\_May-2012-BSR.par

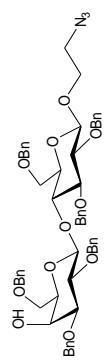
MALDI-TOF MS spectrum of compound **17**

## Mercury 400 spectrometer

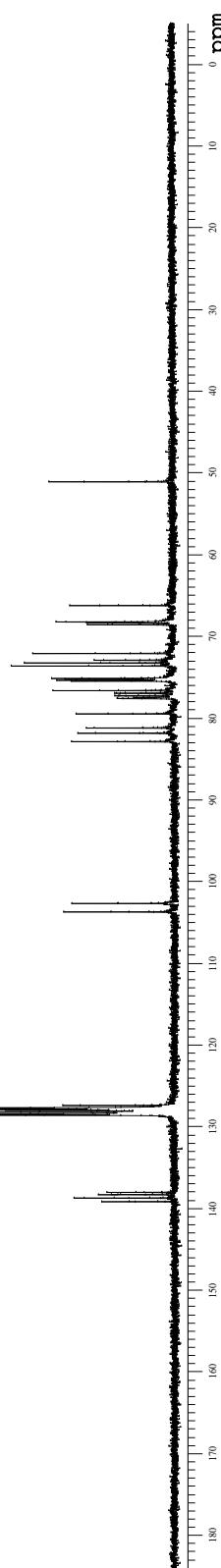


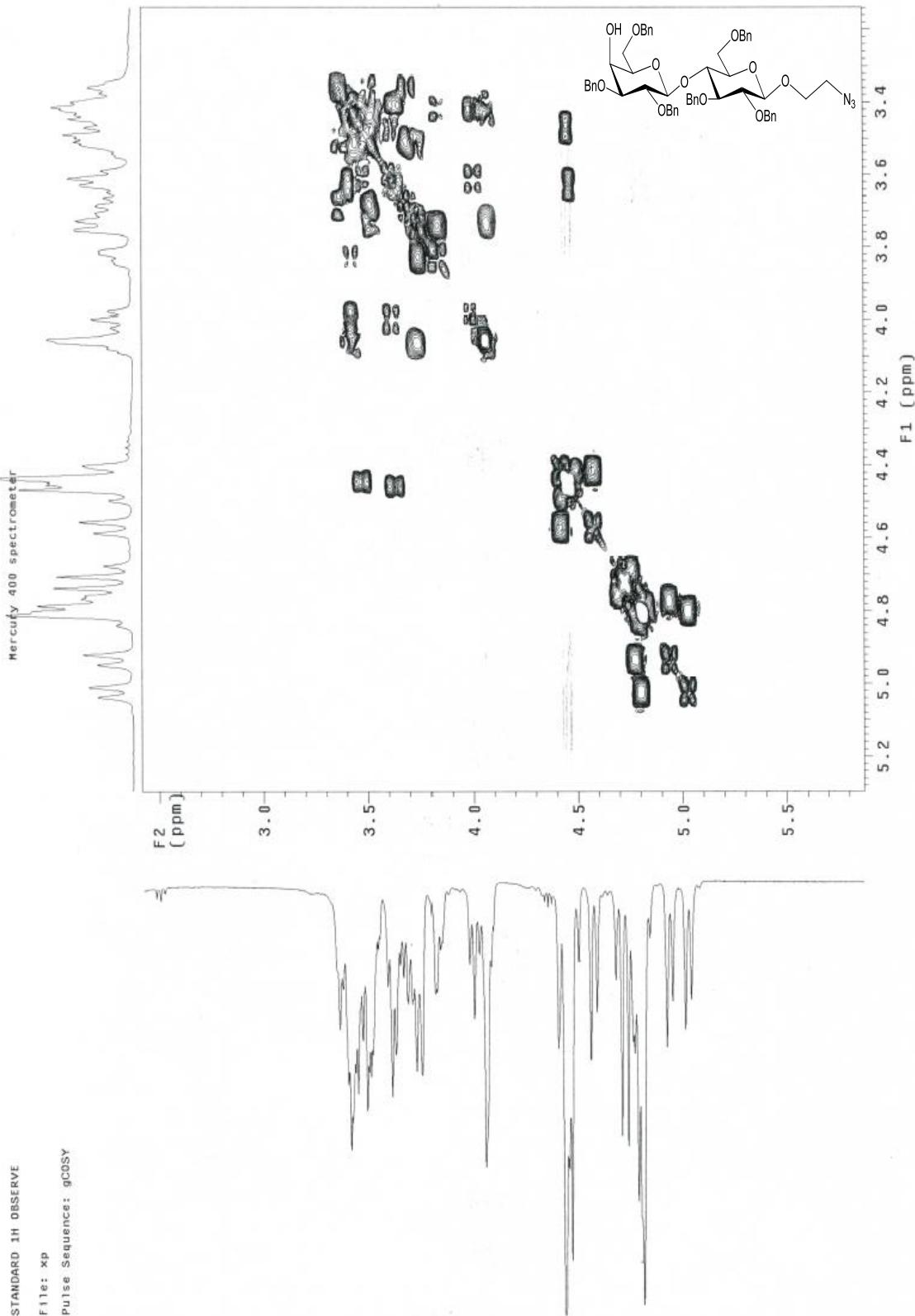
<sup>1</sup>H NMR Spectrum of compound **10** (CDCl<sub>3</sub>, 400 MHz)

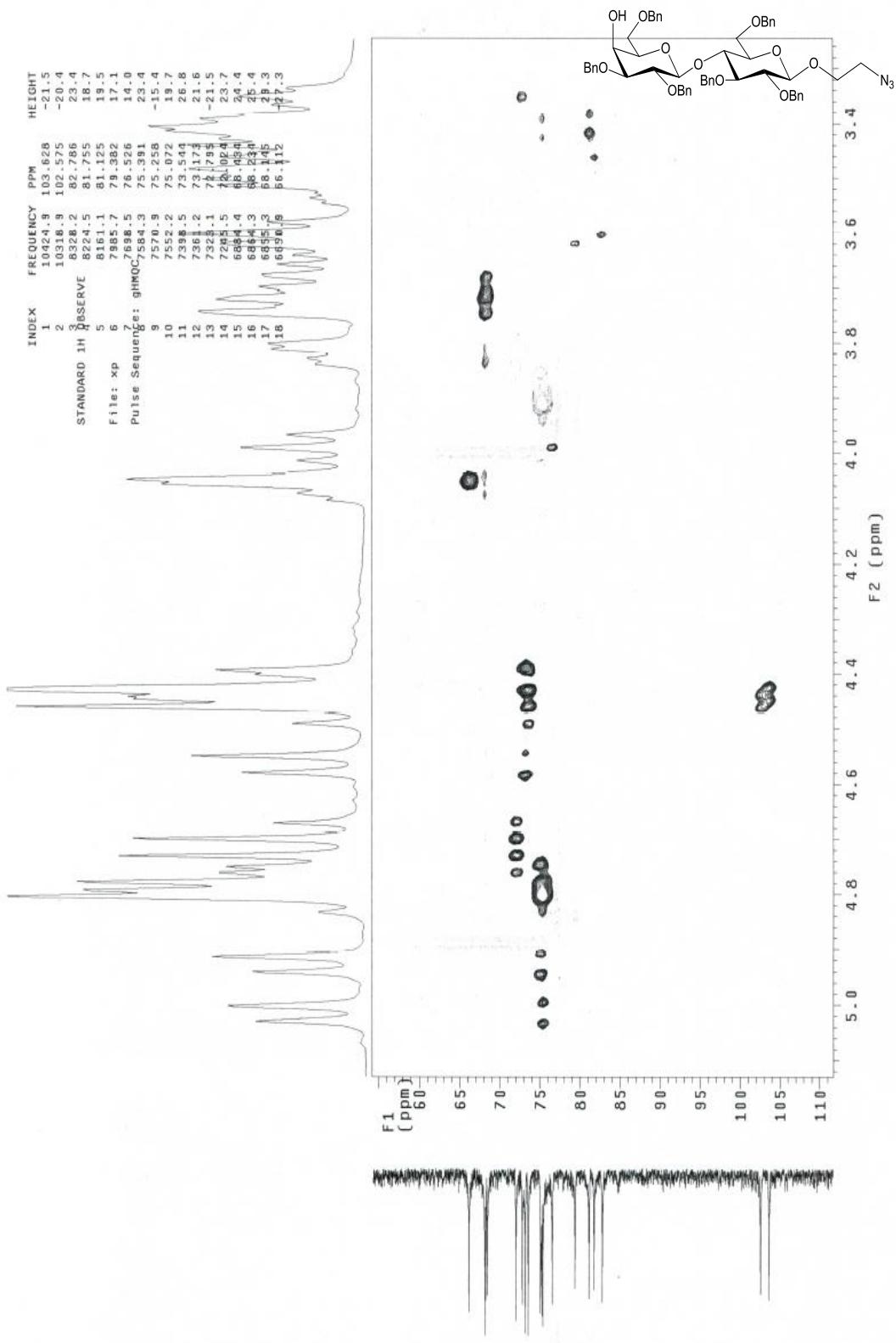
Mercury 400 spectrometer



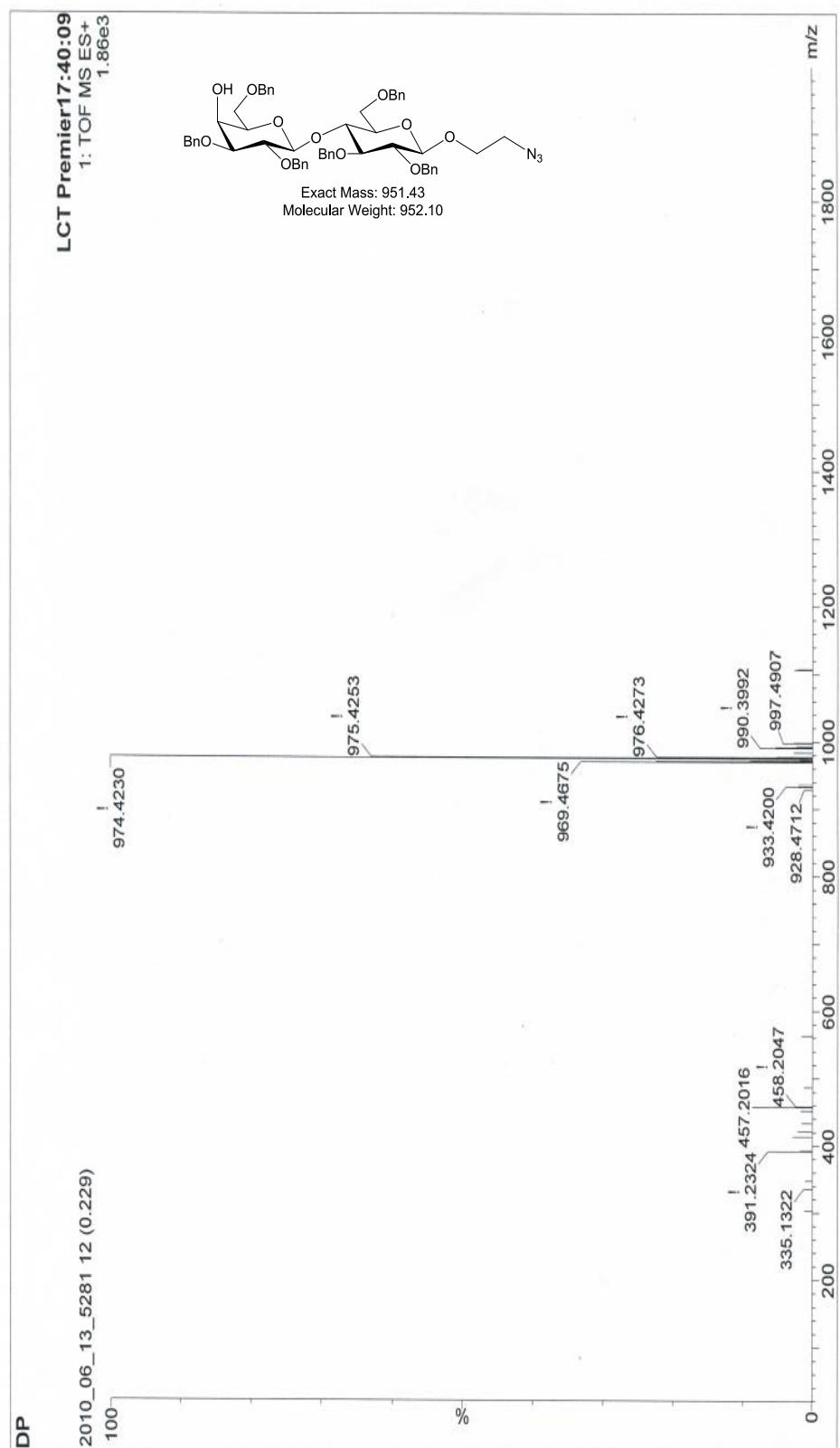
<sup>13</sup>C NMR Spectrum of compound **10** (CDCl<sub>3</sub>, 400 MHz)







<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **10** (CDCl<sub>3</sub>, 400/100 MHz)



ESI MS spectrum of compound **10**

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

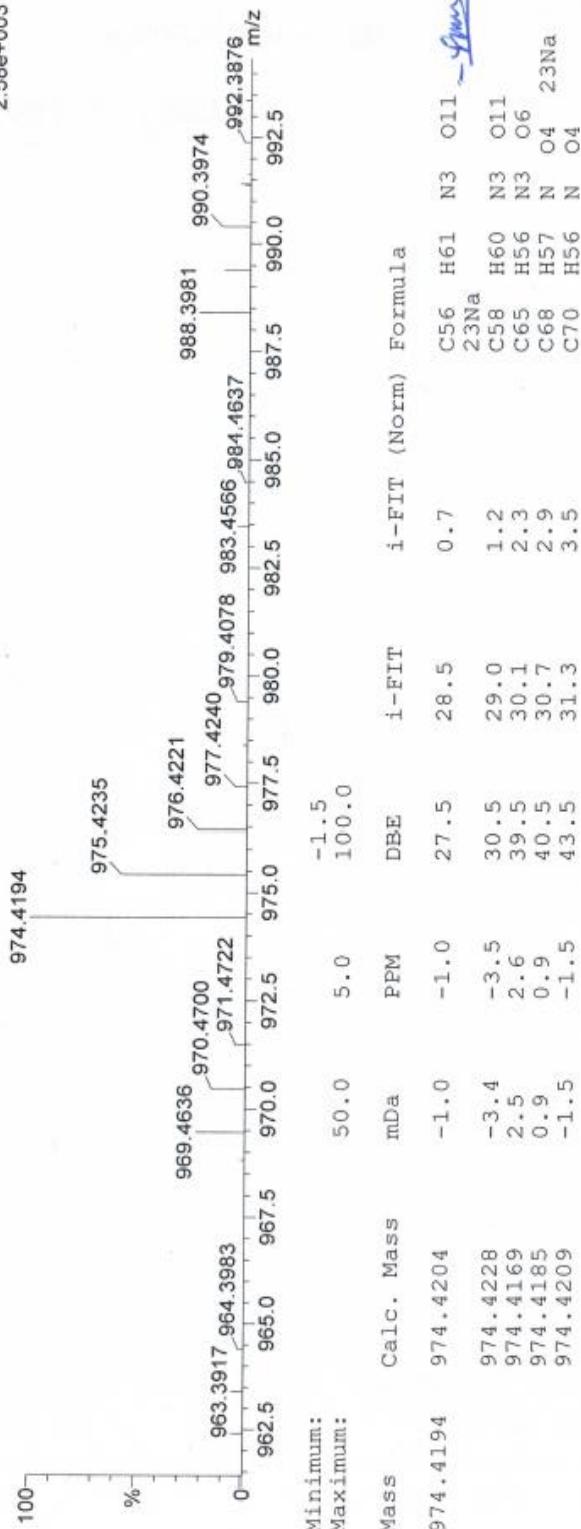
### Monoisotopic Mass, Even Electron Ions

1393 formula(e) evaluated with 5 results within limits (all results (up to 1000) for each mass)

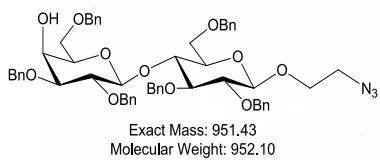
Elements Used:

C: 0-100 H: 0-1000 N: 0-3 O: 0-15 23Na: 0-1  
SATADRU MANDAL SSM-Lactose-4'-OH

2013\_0130\_3057\_2\_31 (0.653) Cm (31:37:1:8x2.000)

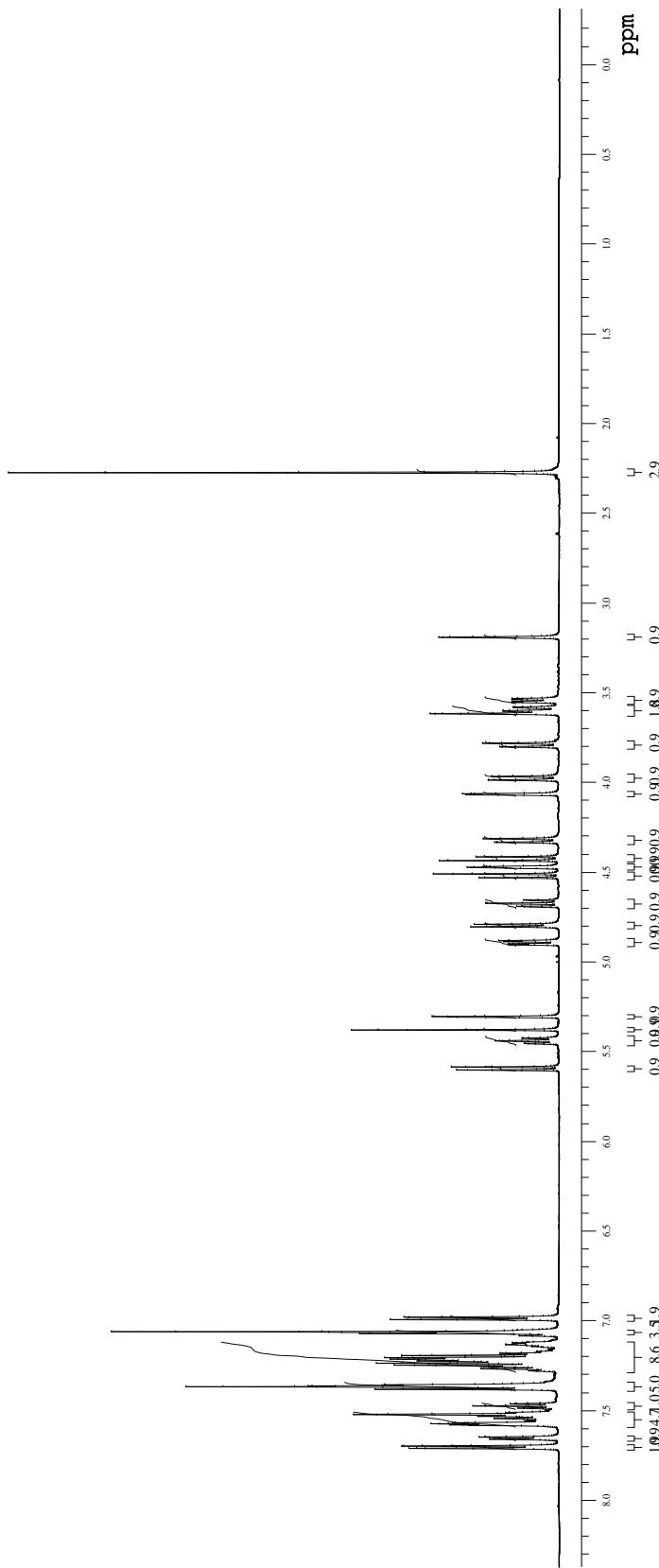
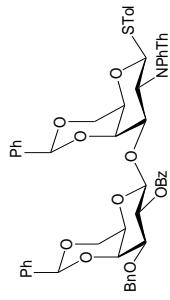


HRMS (ESI MS) spectrum of compound **10**



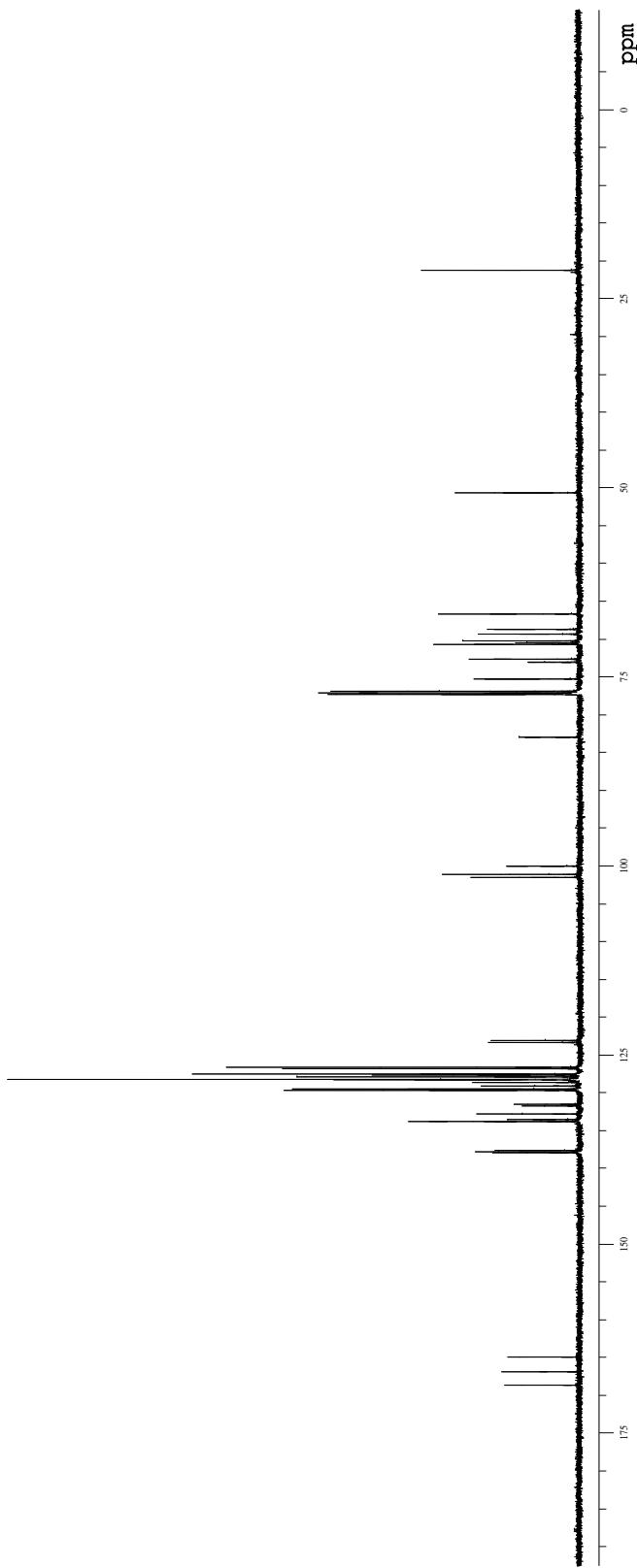
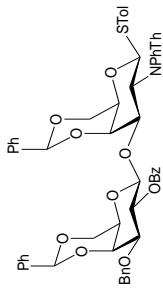
LCT2008-07b.pro 2010-cif spILCT Premier  
1: TOF MS ES+  
2.58e+003

Agilent 600 NMR spectrometer

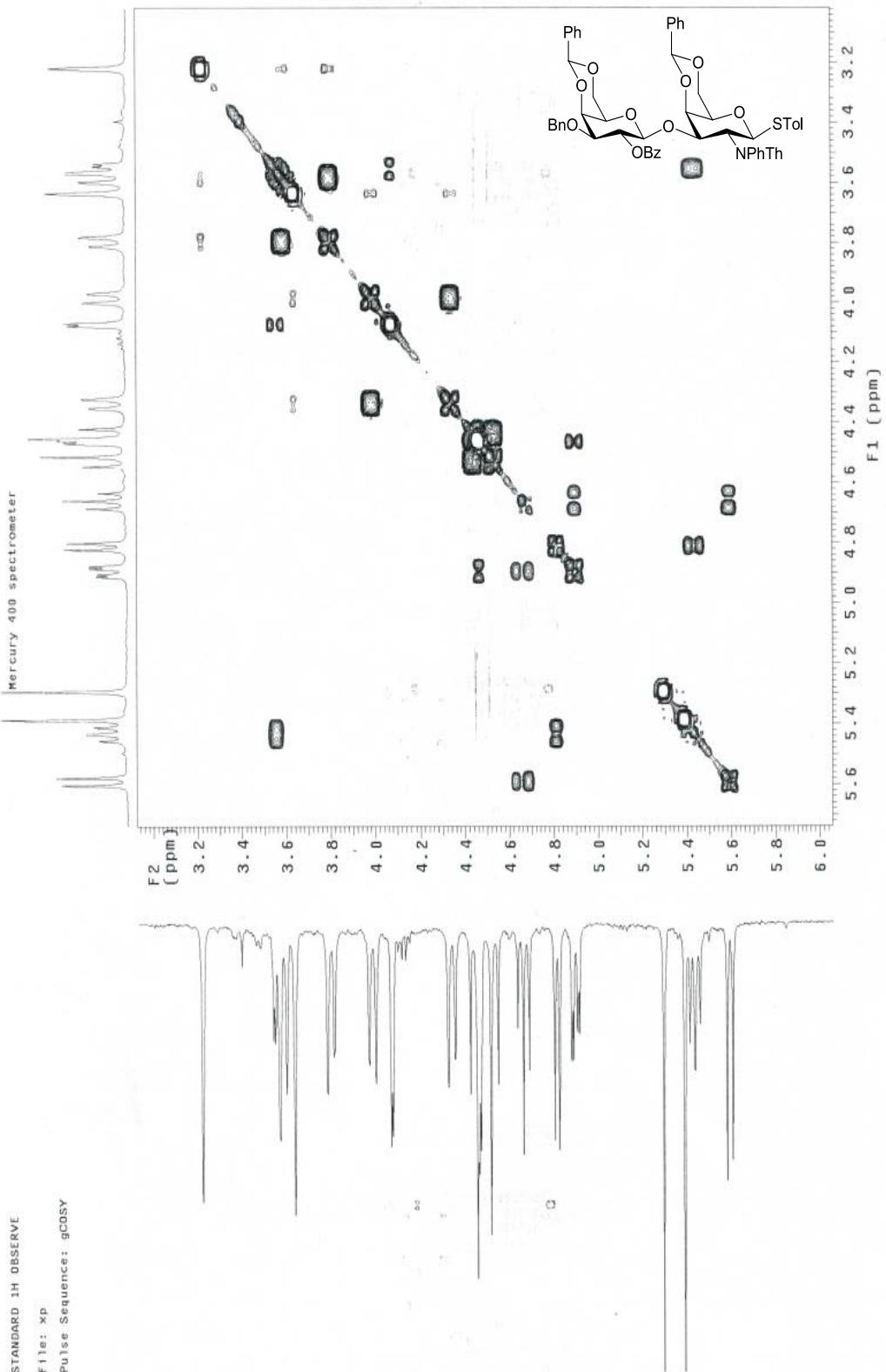


<sup>1</sup>H NMR Spectrum of compound **5** (CDCl<sub>3</sub>, 600 MHz)

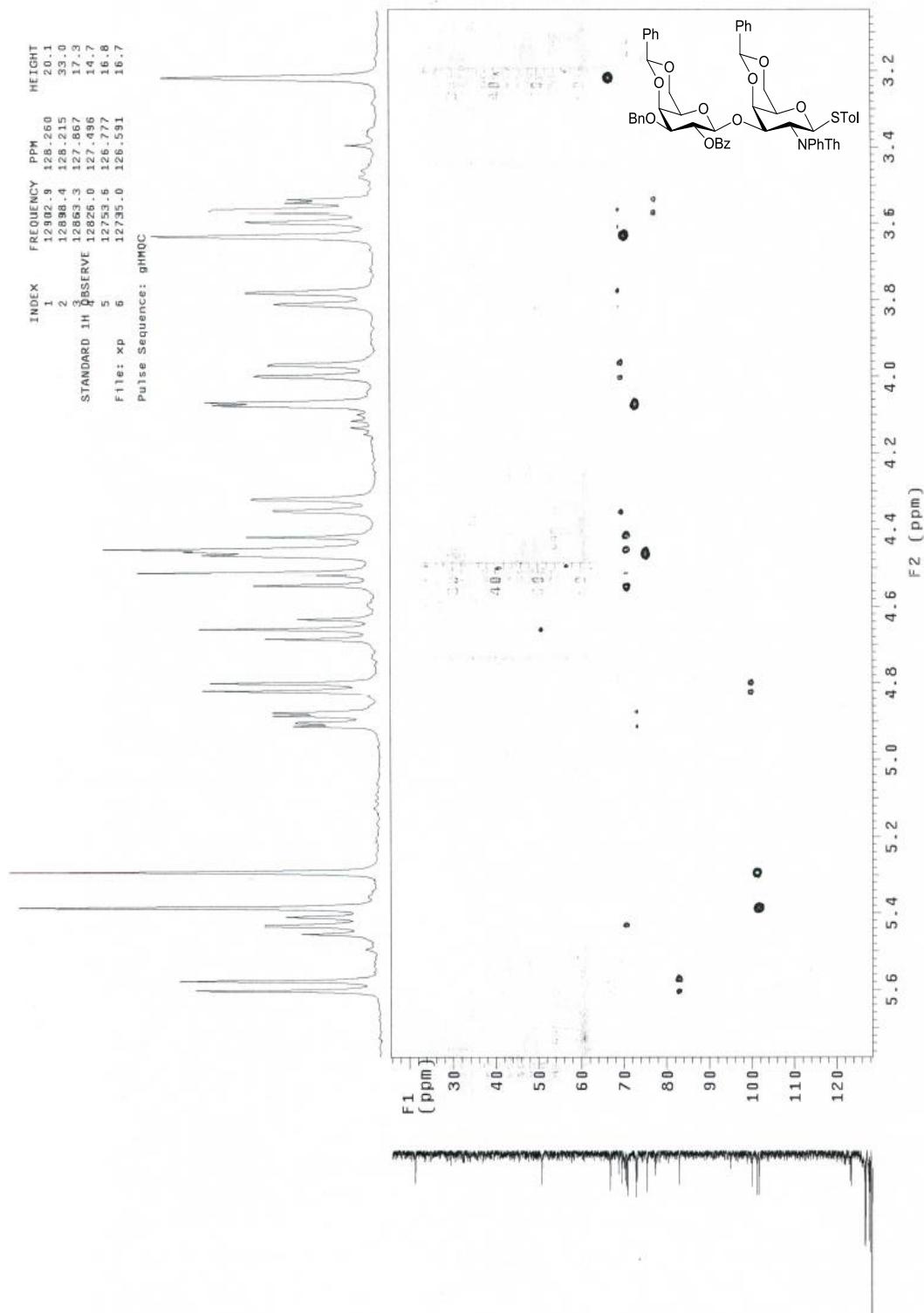
Agilent 600 NMR spectrometer



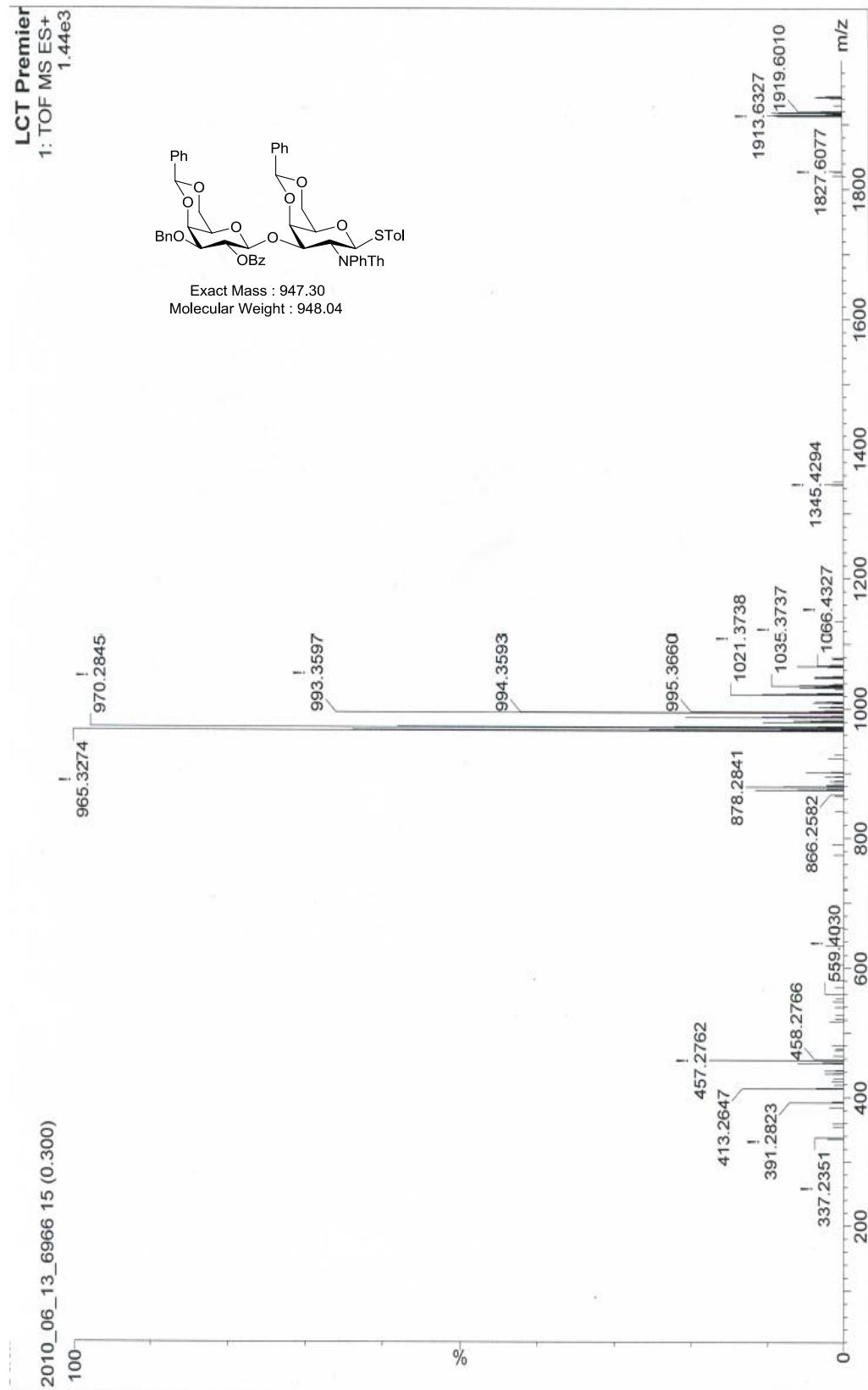
$^{13}\text{C}$  NMR Spectrum of compound 5 ( $\text{CDCl}_3$ , 600 MHz)



$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **5** ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound 5 ( $\text{CDCl}_3$ , 400/100 MHz)



ESI MS spectrum of compound 5

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions

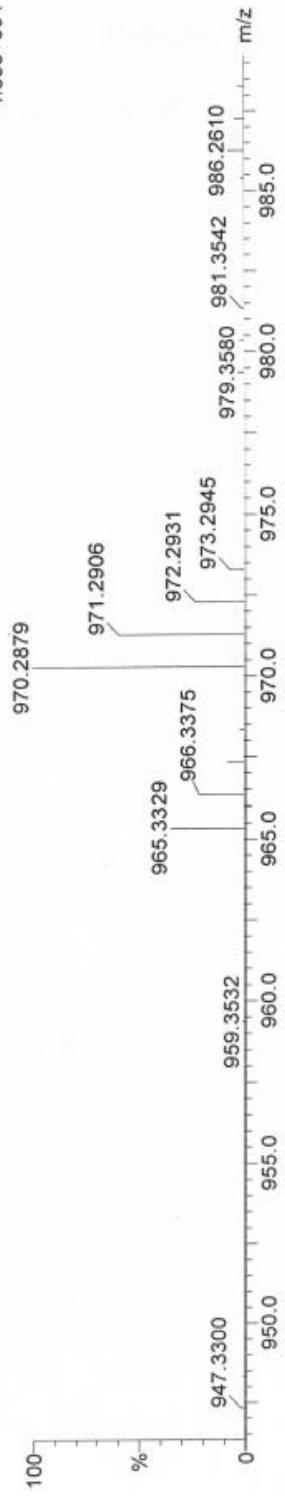
2625 formula(e) evaluated with 16 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-100 H: 0-1000 N: 0-3 O: 0-15 S: 1-2

SATADRU MANDAL  
SSM-Disaccharide

2013\_0130\_3055\_12 (0.263) Cm (8:20-1:7x2,000)

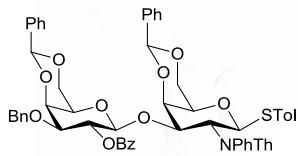


Minimum: 50.0  
Maximum: 100.0

Mass	Calc. Mass	mDa	PPM	DBE	i-FIT	(Norm)	Formula
970.2879	970.2868	1.1	1.1	49.5	47.3	1.3	C68 H41 N3 O 23Na
970.2873	970.2873	0.6	0.6	31.5	47.5	1.5	S C55 H49 N O12 23Na

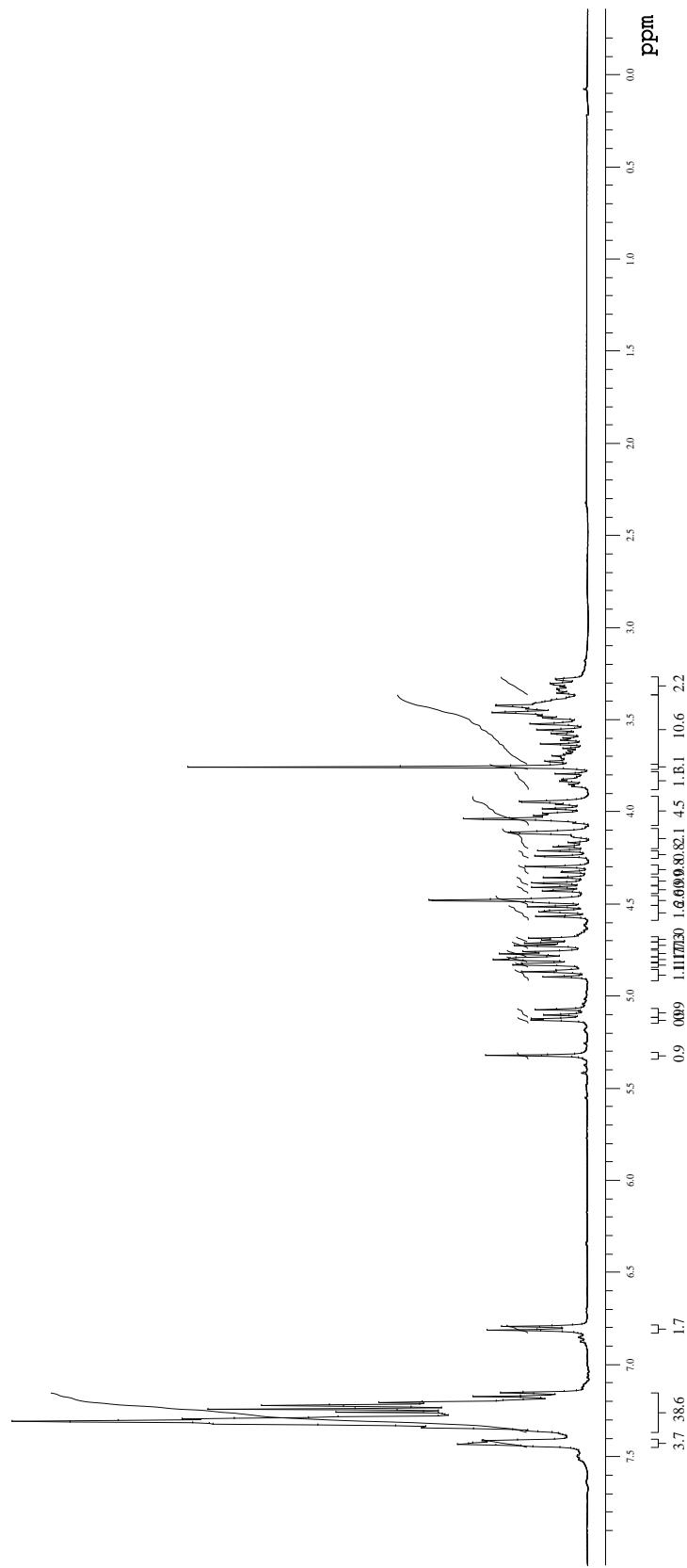
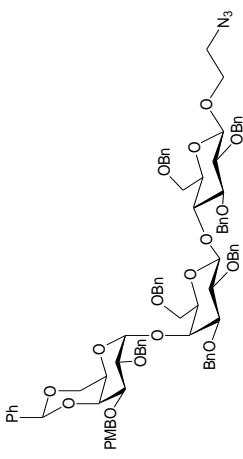
HRMS (ESI MS) spectrum of compound 5

Page 1



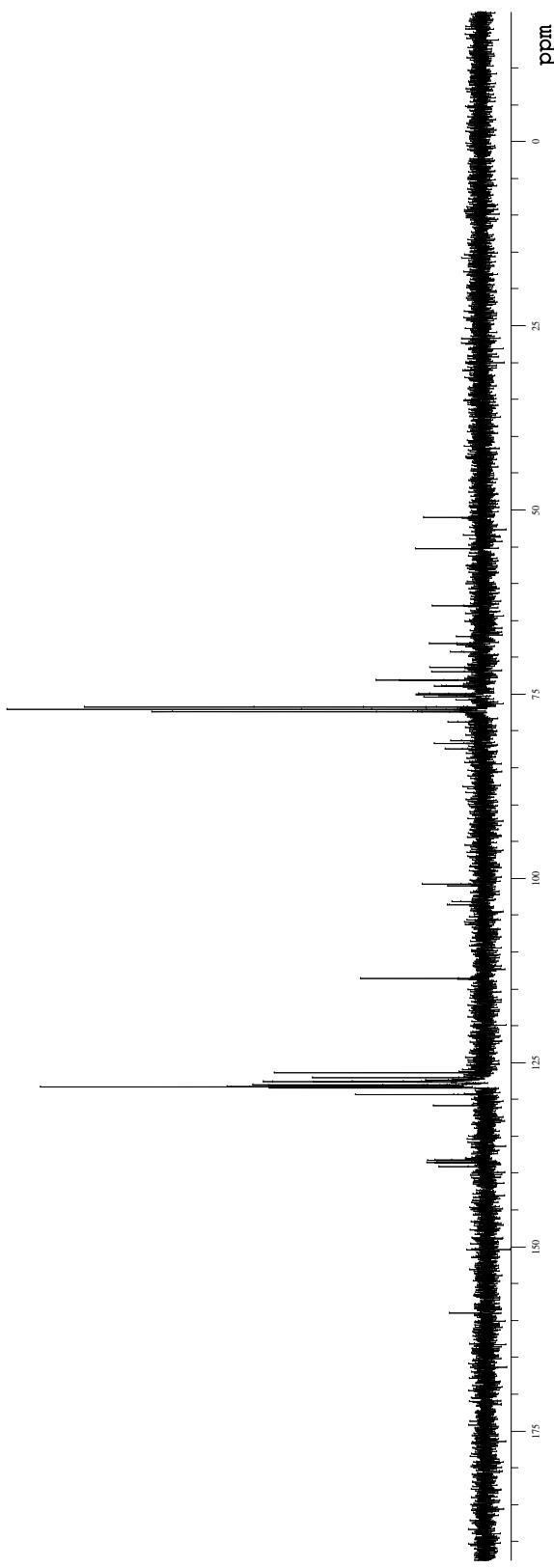
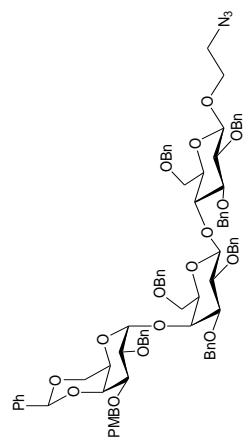
Exact Mass : 947.30  
Molecular Weight : 948.04

## Mercury 400 spectrometer



<sup>1</sup>H NMR Spectrum of compound **6** (CDCl<sub>3</sub>, 400 MHz)

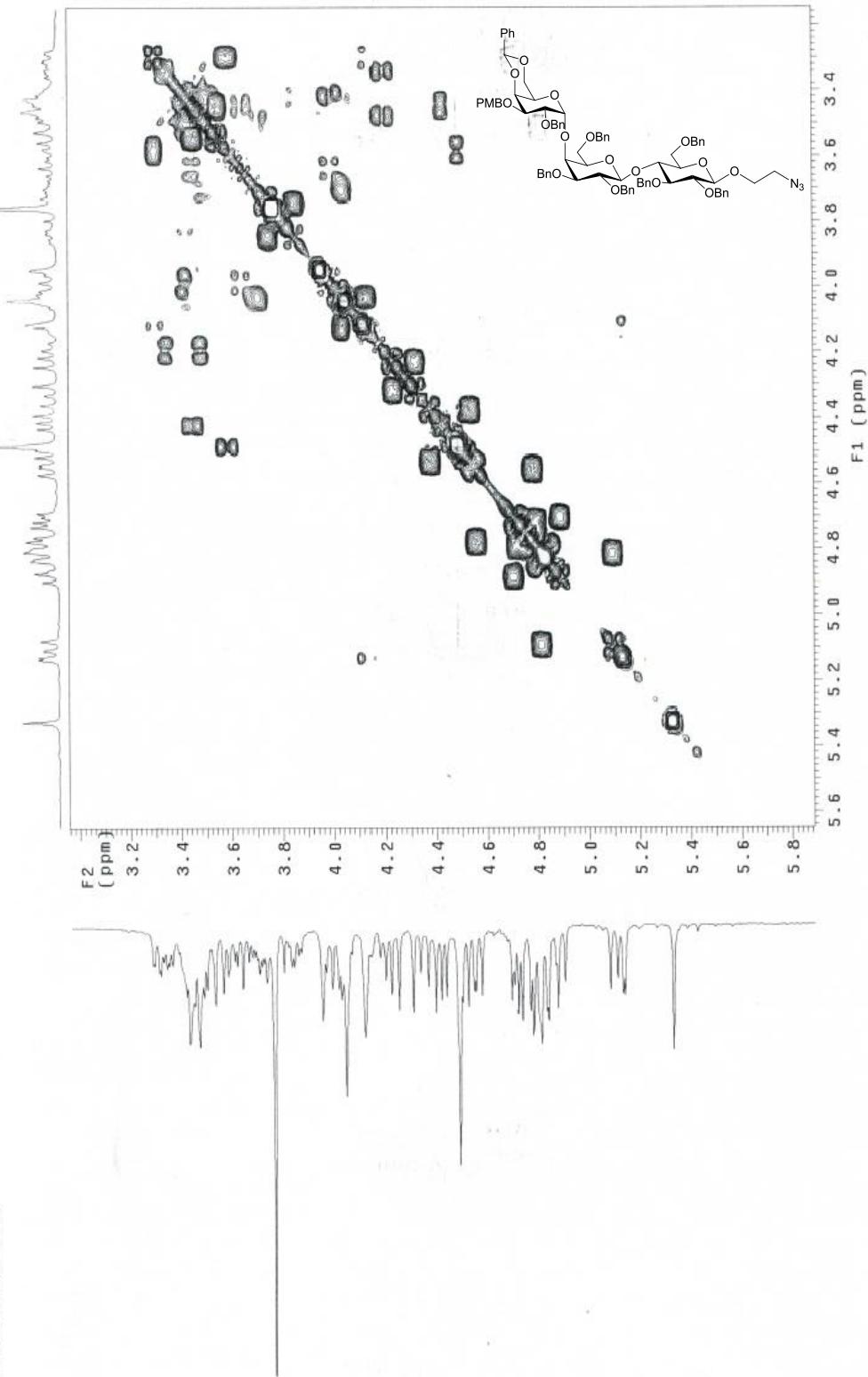
Mercury 400 spectrometer



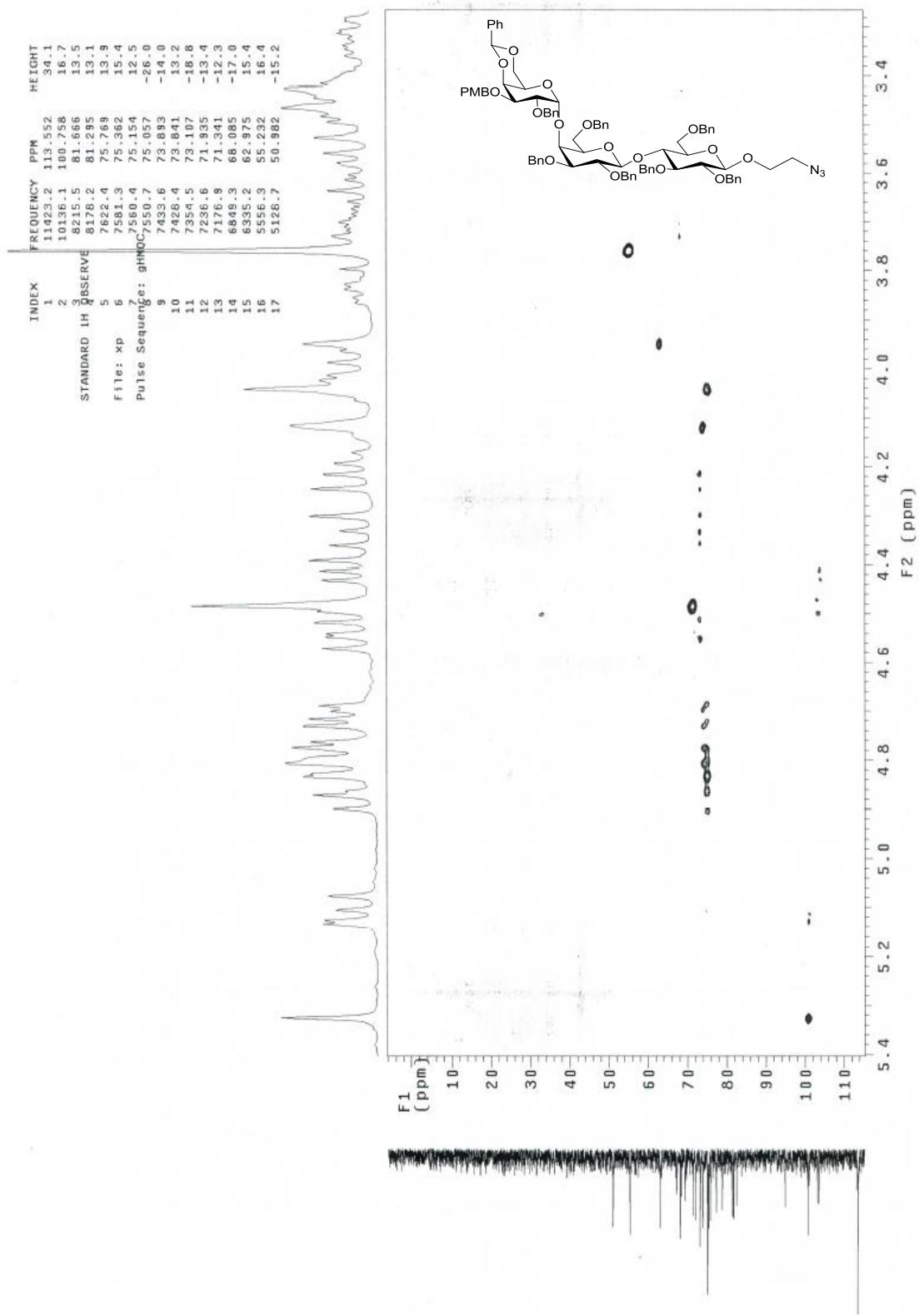
<sup>13</sup>C NMR Spectrum of compound 6 (CDCl<sub>3</sub>, 400 MHz)

STANDARD 1H OBSERVE  
File: xp  
Pulse Sequence: gCOSY

Mercury 400 spectrometer

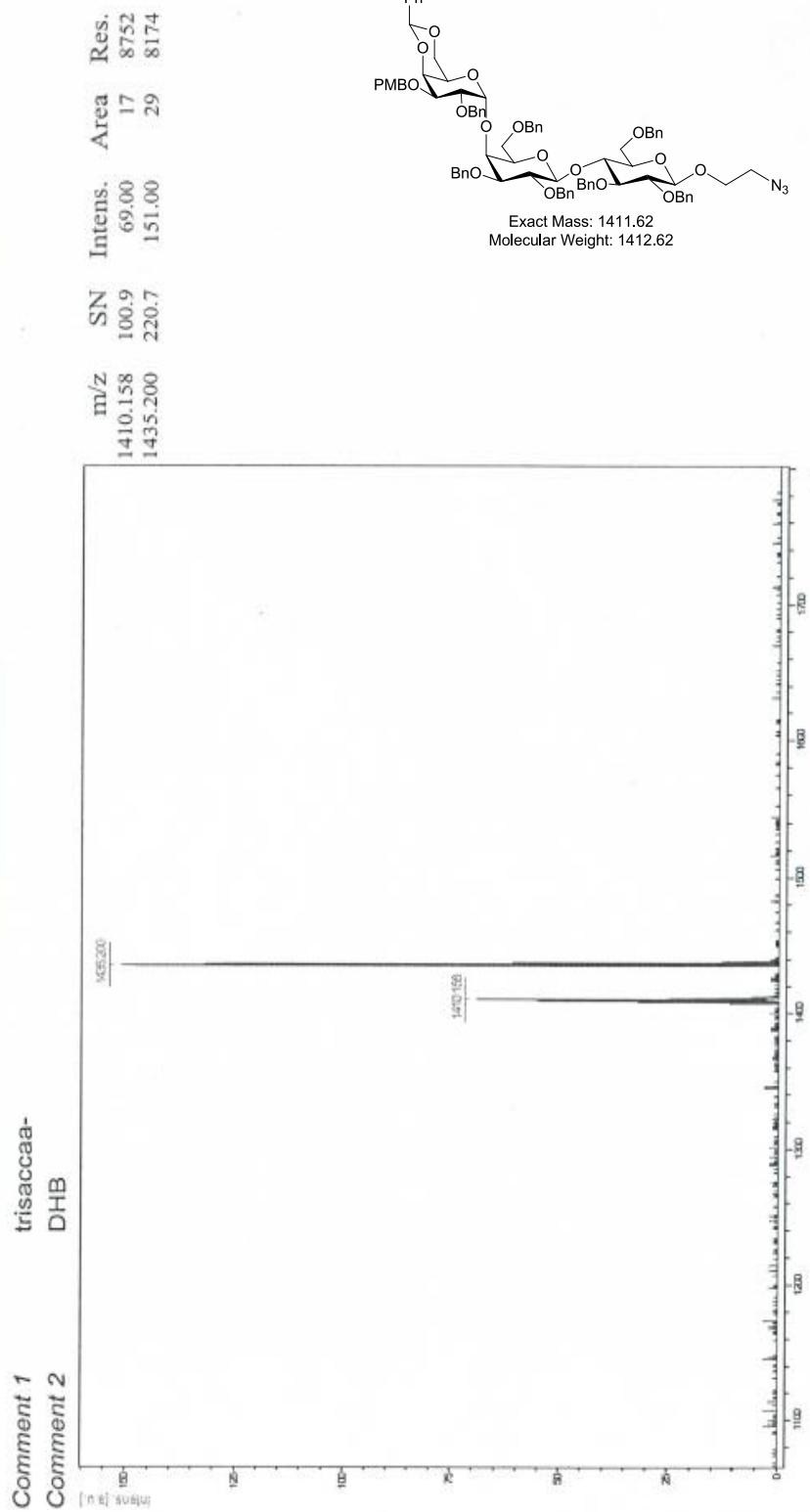


$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound 6 ( $\text{CDCl}_3$ , 400 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **6** ( $\text{CDCl}_3$ , 400/100 MHz)

D:\Data\Guo\_Lab\S S Mandal\Giobo-Hitrisaccaa\0\_I19\1



Acquisition Parameter

D:\Methods\1User\_Methods\Guo\RP\_3147\_PepMixDHB\_June 1-2011.par

MALTI-TOF MS spectrum of compound 6

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

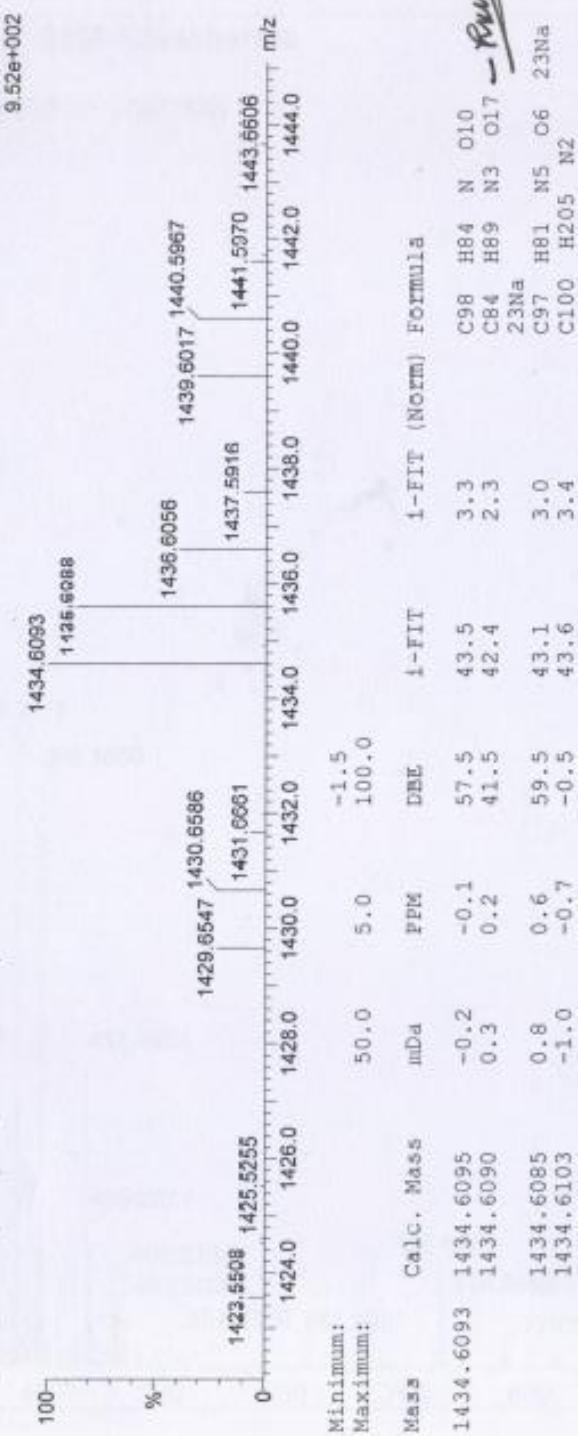
Monoisotopic Mass, Even Electron Ions  
2826 formula(e) evaluated with 14 results within limits (all results (up to 1000) for each mass)

Elements Used:

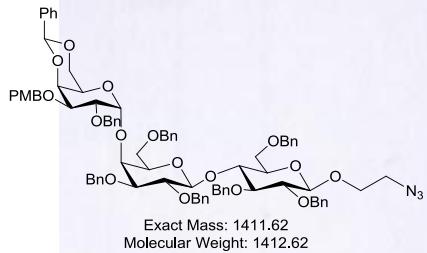
C: 0-100 H: 0-1000 N: 0-5 O: 0-20 23Na: 0-1

SATARDU S MANDAL SSM-Trisacharide

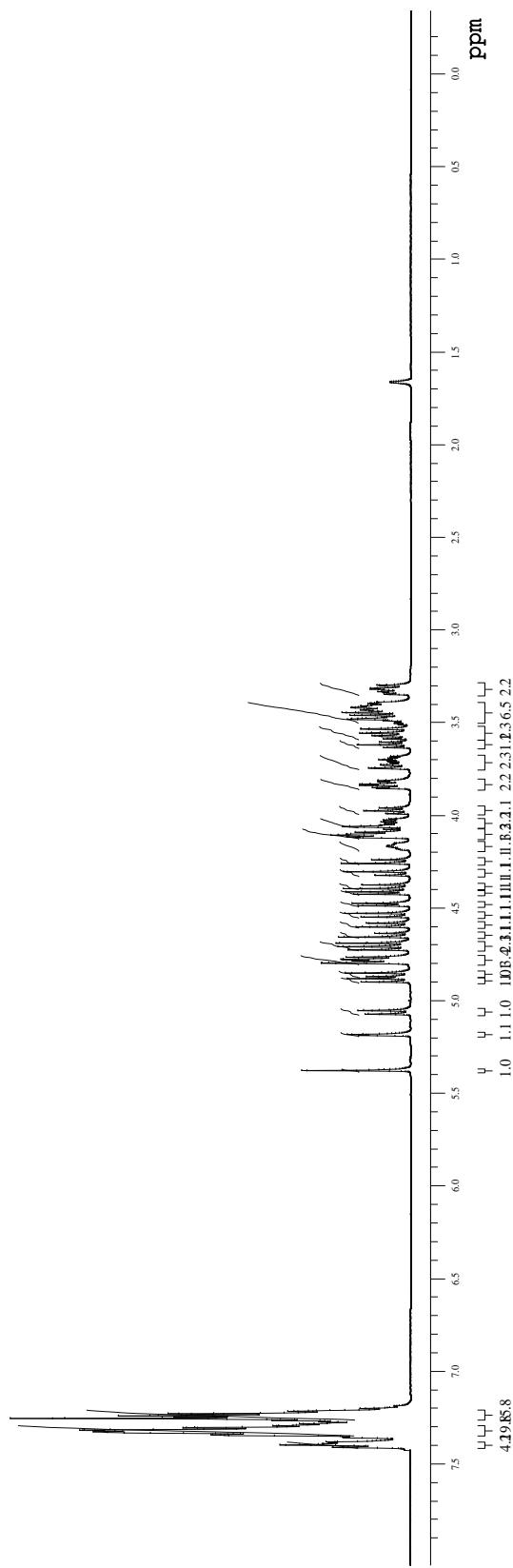
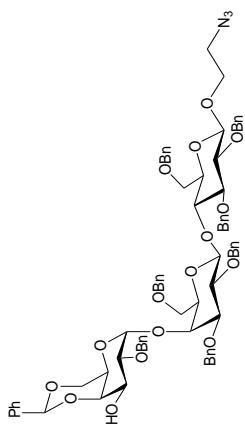
2013\_0215\_3095 16 (0.317) Cm (16.17-1.7x2.000)



HRMS (ESI MS) spectrum of compound 6

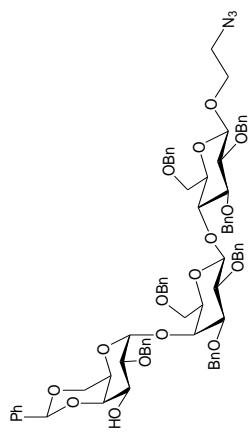


Varian 500 NMR spectrometer

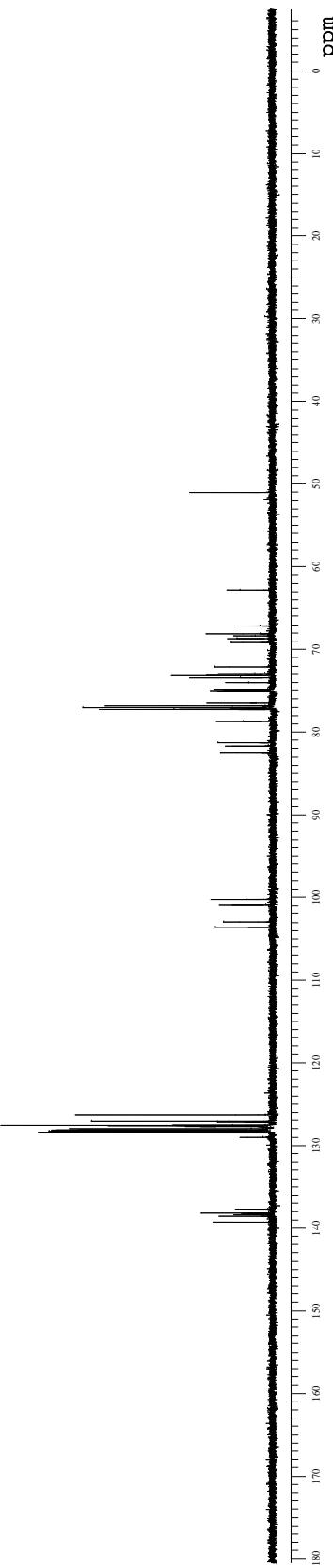


<sup>1</sup>H NMR Spectrum of compound **20** (CDCl<sub>3</sub>, 500 MHz)

Varian 500 NMR spectrometer

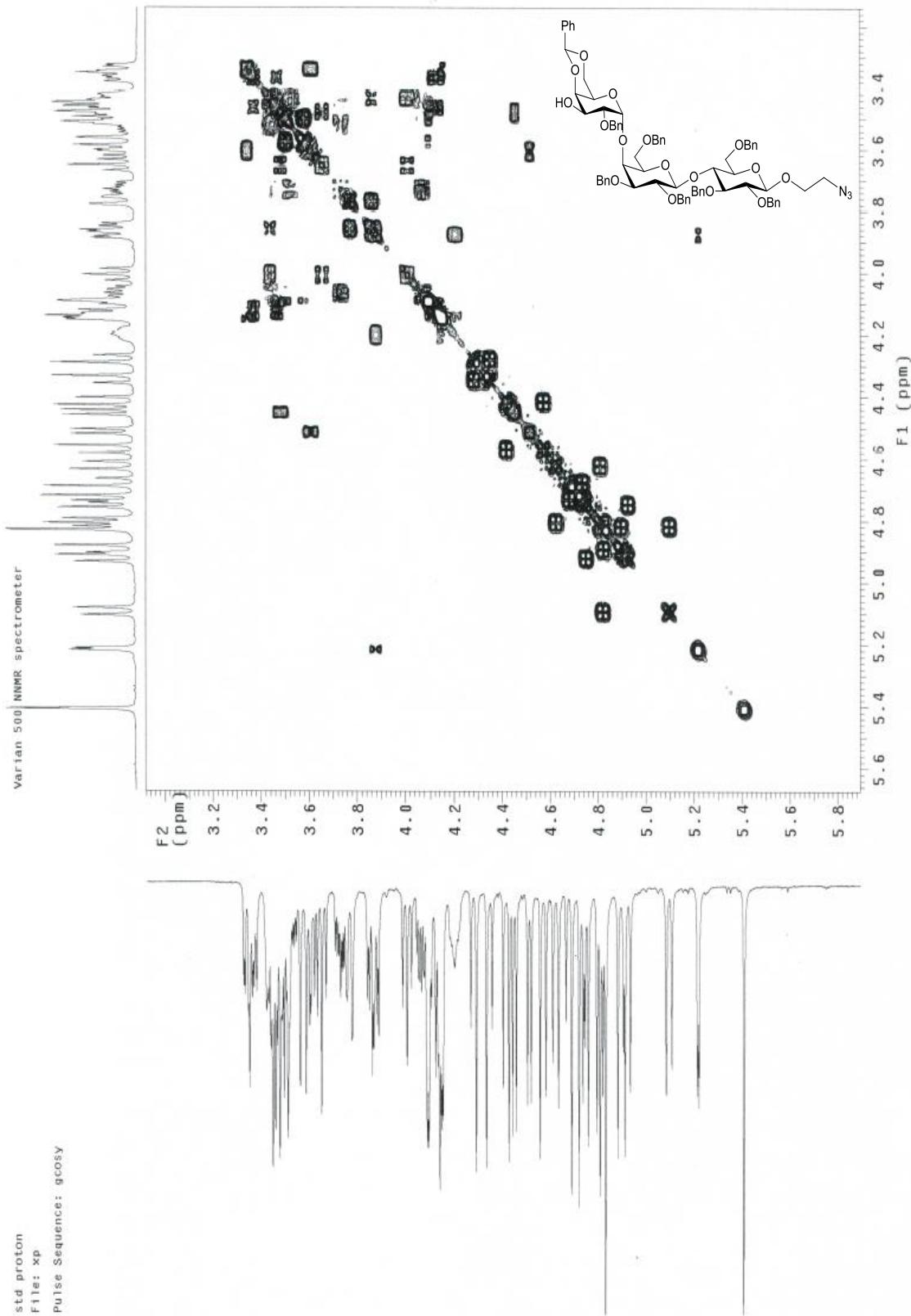


$^{13}\text{C}$  NMR Spectrum of compound **20** ( $\text{CDCl}_3$ , 500 MHz)

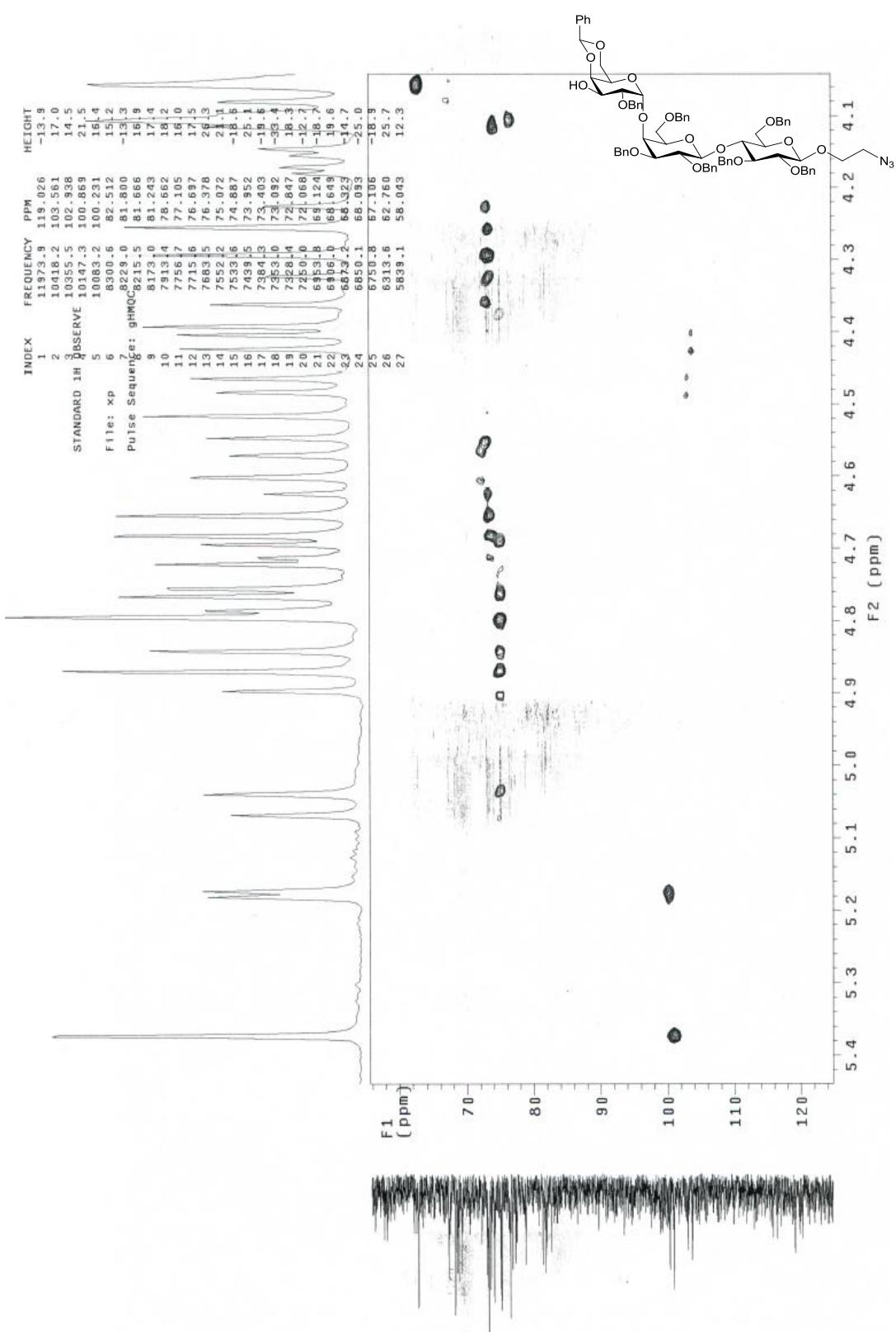


std proton  
File: xp  
Pulse Sequence: gcosy

Varian 500 NMR spectrometer



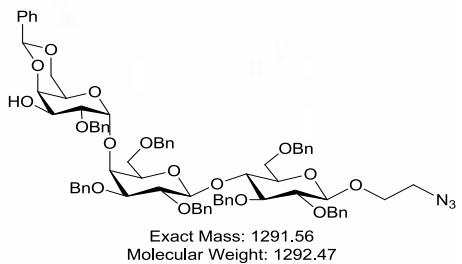
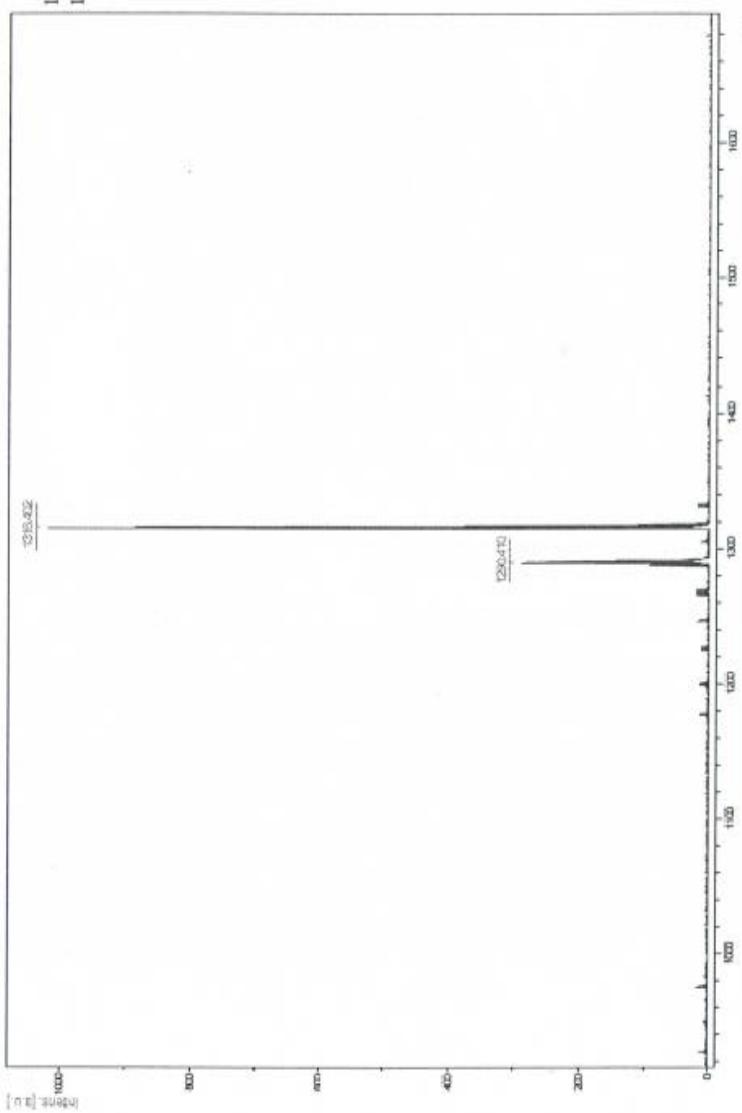
<sup>1</sup>H-<sup>1</sup>H COSY Spectrum of compound **20** (CDCl<sub>3</sub>, 500 MHz)



<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **20** (CDCl<sub>3</sub>, 500/125 MHz)

D:\Data\Guo\_lab\SS Mandal\Globo-H\trisaccaa-Depmb\0\_F22\1

Comment 1 trisaccaa-Depmb  
Comment 2 DHB



Acquisition Parameter  
D:\Methods\1User\_Methods\Guo\RP\_3147\_PepMixDHB\_June 1\2011.par

MALDI-TOF MS spectrum of compound 20

## Elemental Composition Report

### **Single Mass Analysis**

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0  
Element prediction: Off

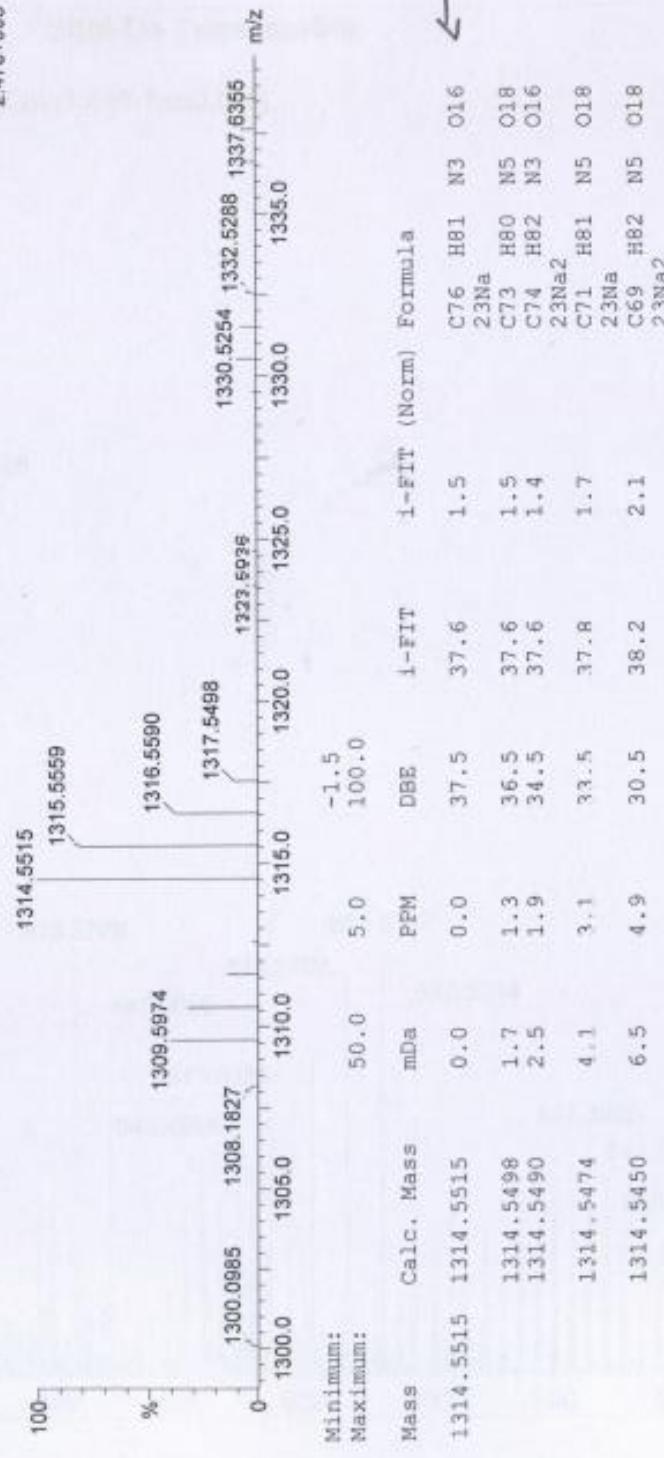
Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions  
1228 formula(e) evaluated with 5 results within limits (all results (up to 1000) for each mass)

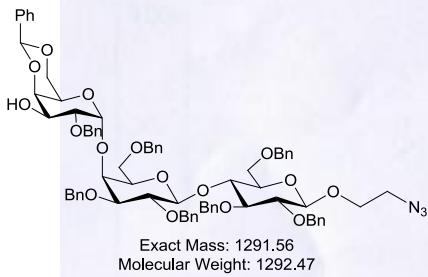
Elements Used:

C: 0-76 H: 0-1000 N: 0-5 O: 0-20 23Na: 0-2  
SATARU S MANDAL

SSM-De Trisachande  
2013\_0215\_3091\_2\_14 (0.283) Gm (11:18-1:9x2.000)

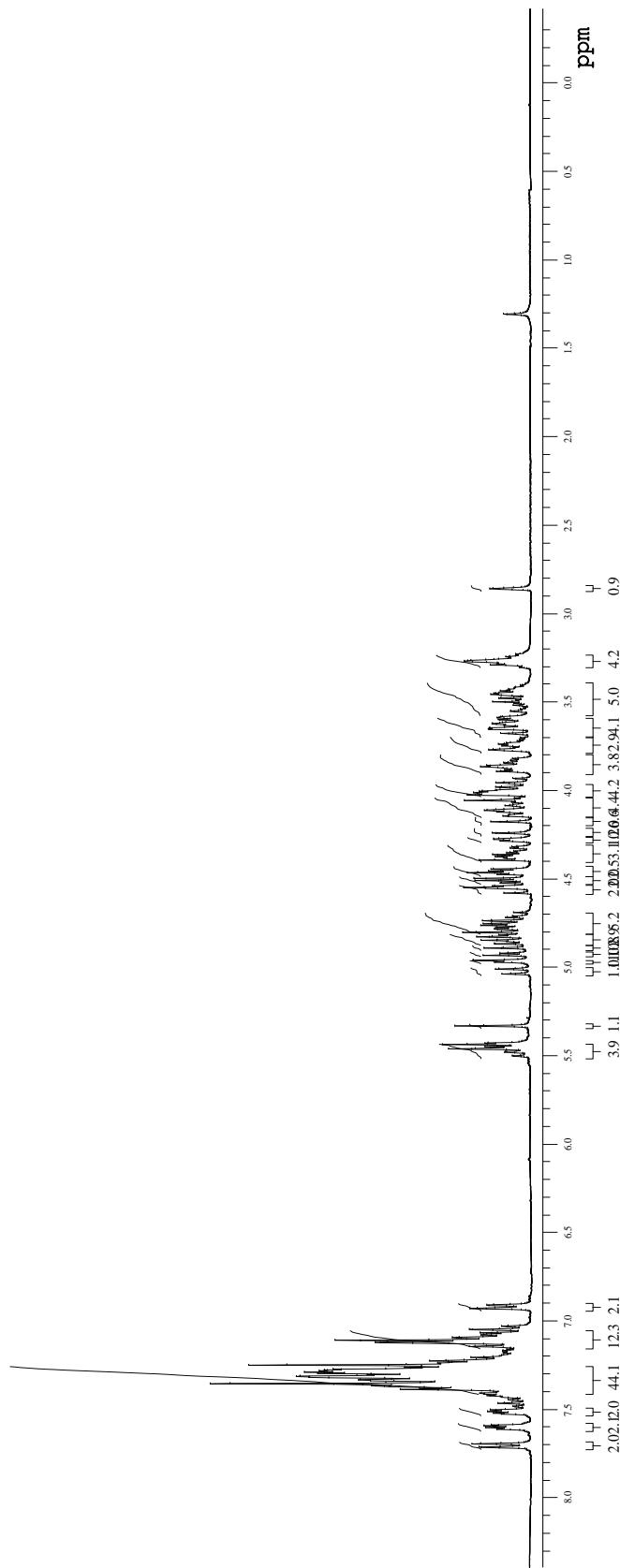
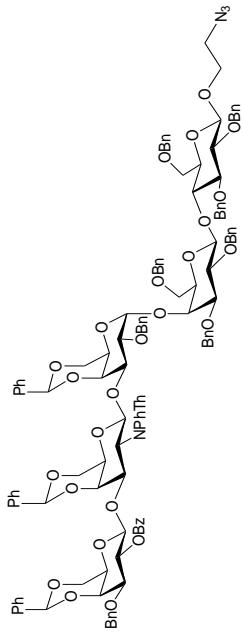


**Page 1**



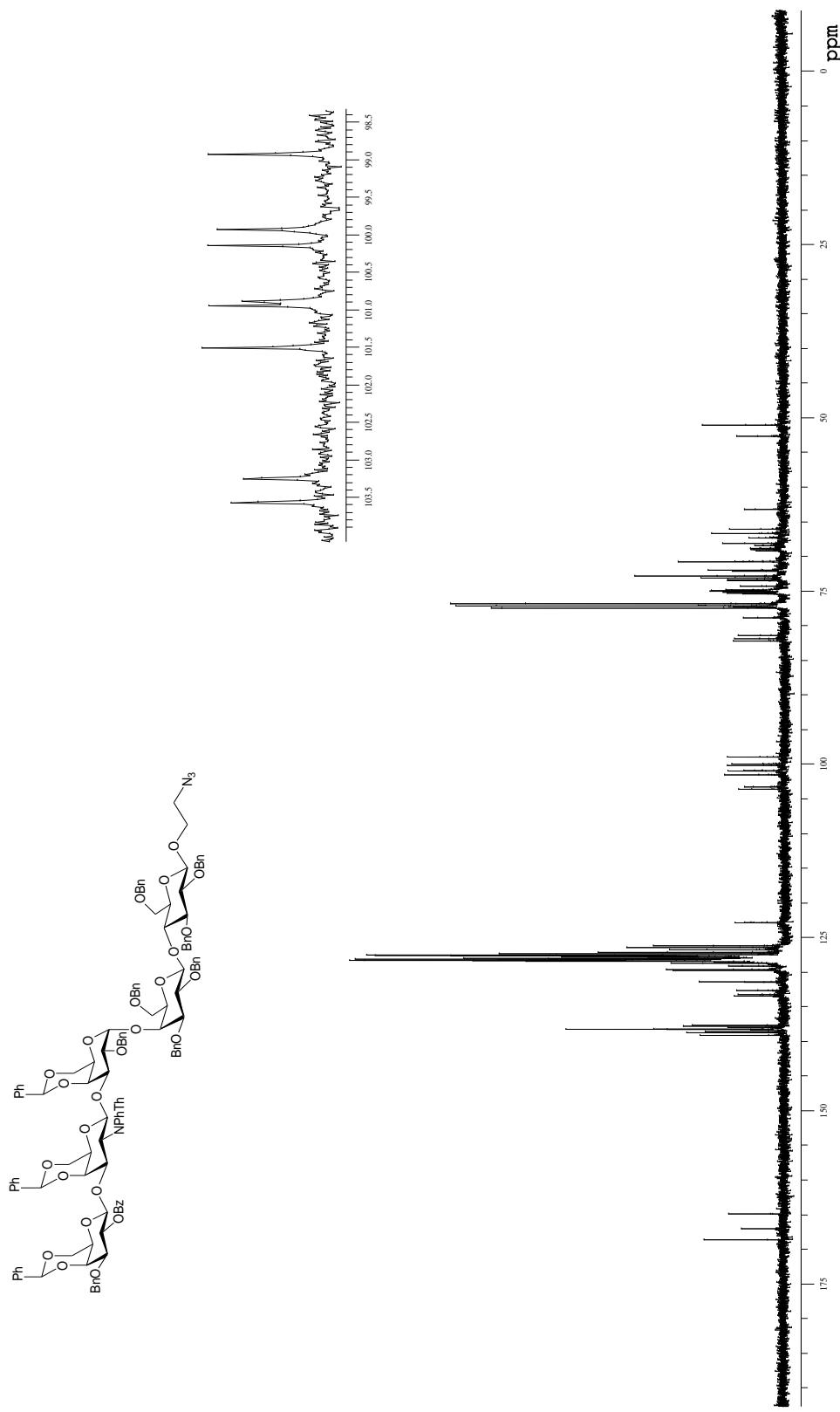
HRMS (ESI MS) spectrum of compound 20.

Varian 500 NMR spectrometer



<sup>1</sup>H NMR Spectrum of compound 3 (CDCl<sub>3</sub>, 500 MHz)

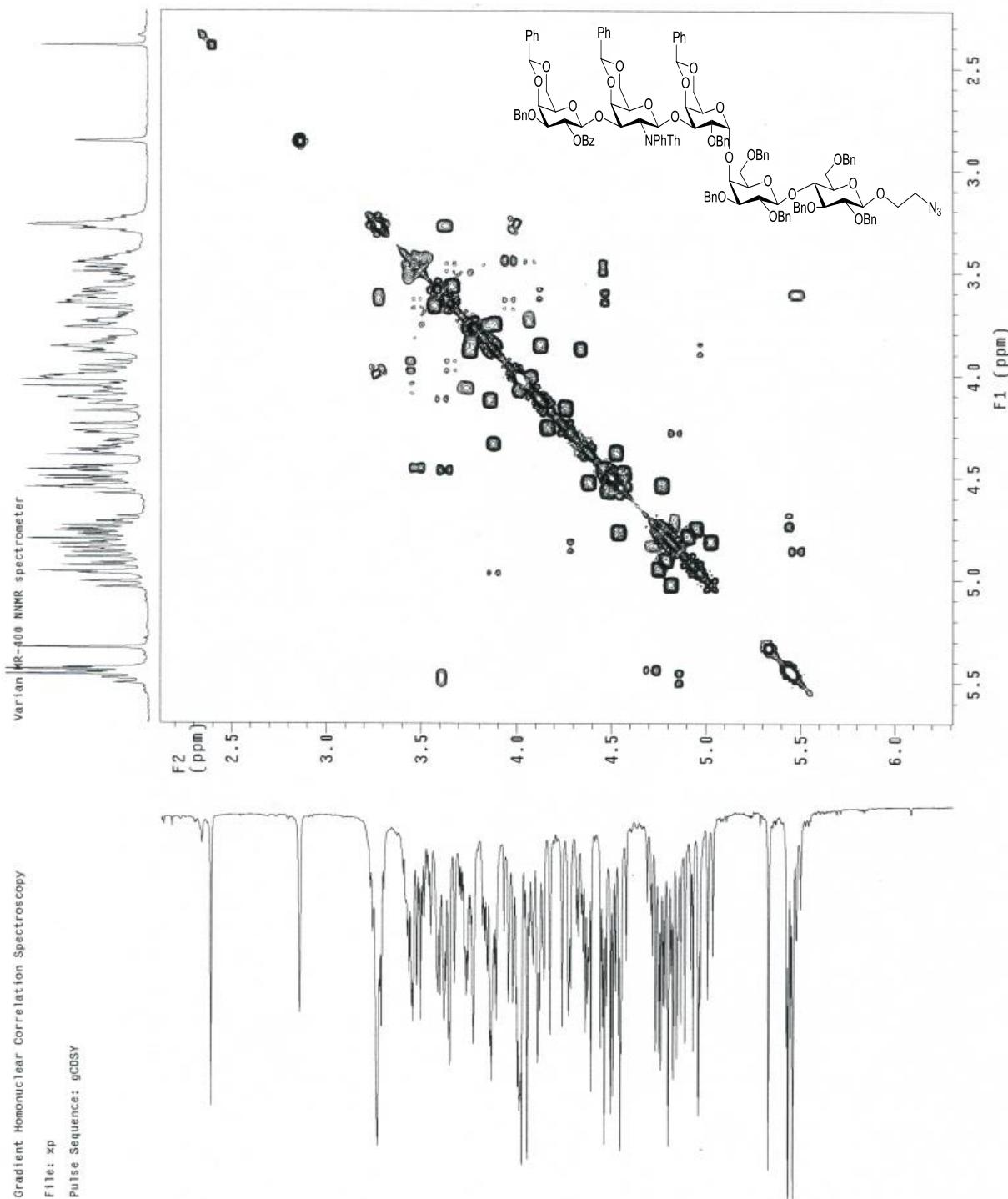
Varian 500 NMR spectrometer



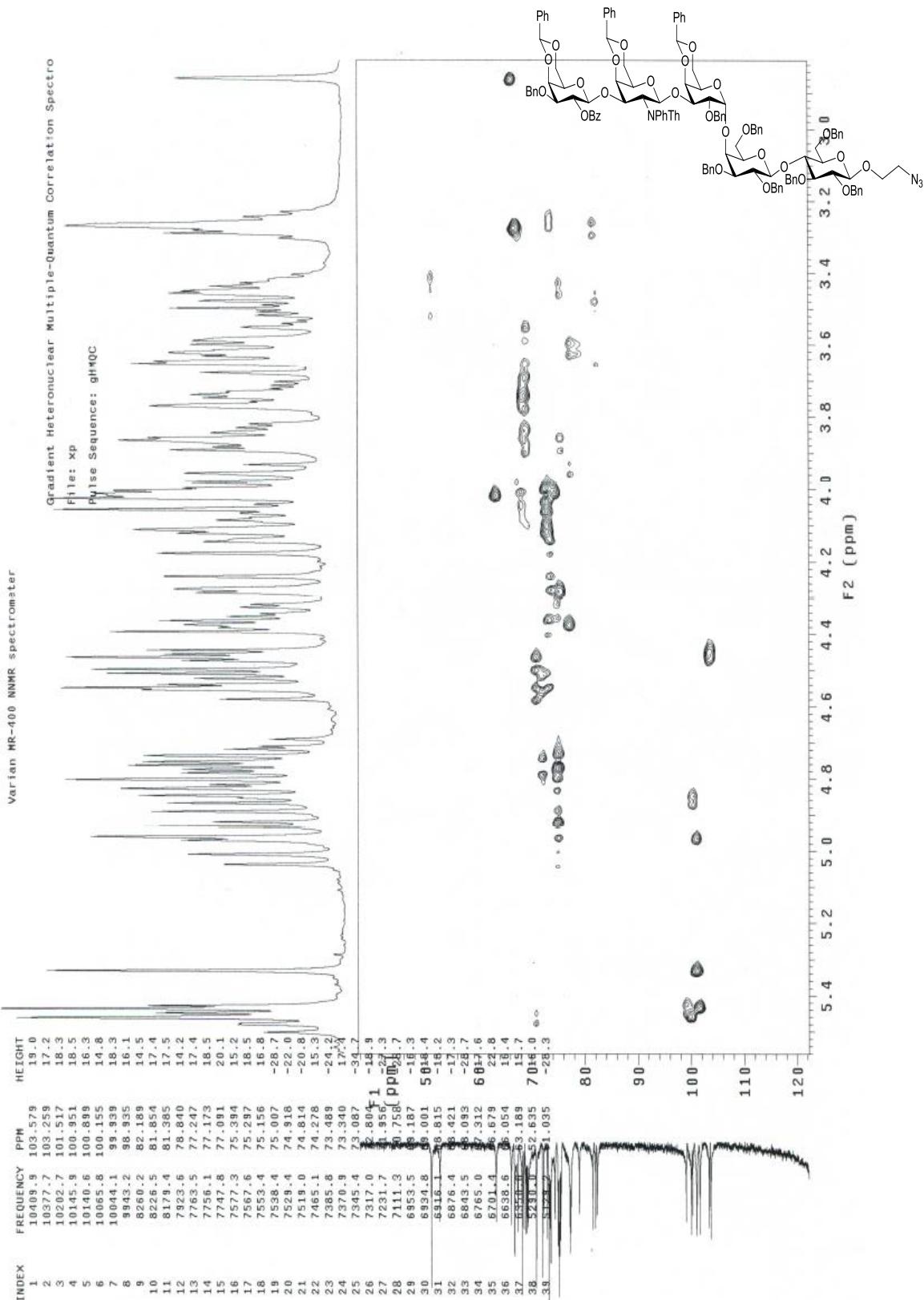
$^{13}\text{C}$  NMR Spectrum of compound 3 ( $\text{CDCl}_3$ , 500 MHz)

Gradient Heteronuclear Correlation Spectroscopy  
FILE: xp  
Pulse Sequence: gCOSY

Varian INE-400 NMR spectrometer



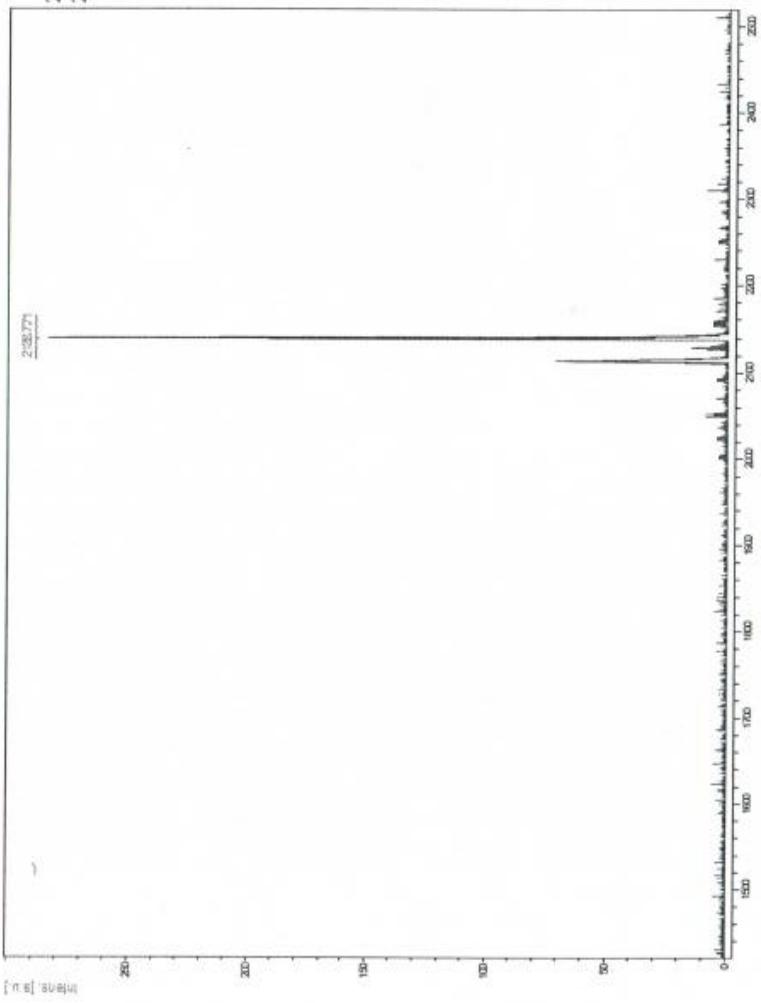
$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound 3 ( $\text{CDCl}_3$ , 400 MHz)



<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound 3 (CDCl<sub>3</sub>, 400/100 MHz)

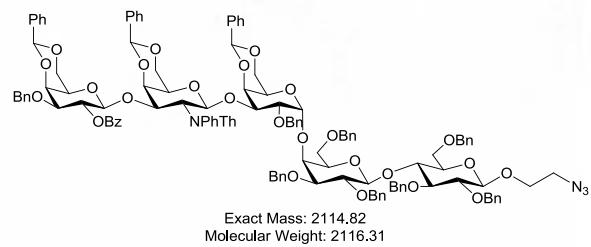
D:\Data\Guo\_lab\S S Mandal\Globo-H\pentasaccaa-0\_E2411

Comment 1      pentasaccaa-  
Comment 2      DHB



MALTI-TOF MS spectrum of compound 3

S-99



Acquisition Parameter  
D:\Methods\1User\_Methods\Guo\RP\_3147\epMixDHD\_June 1-2011.par

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

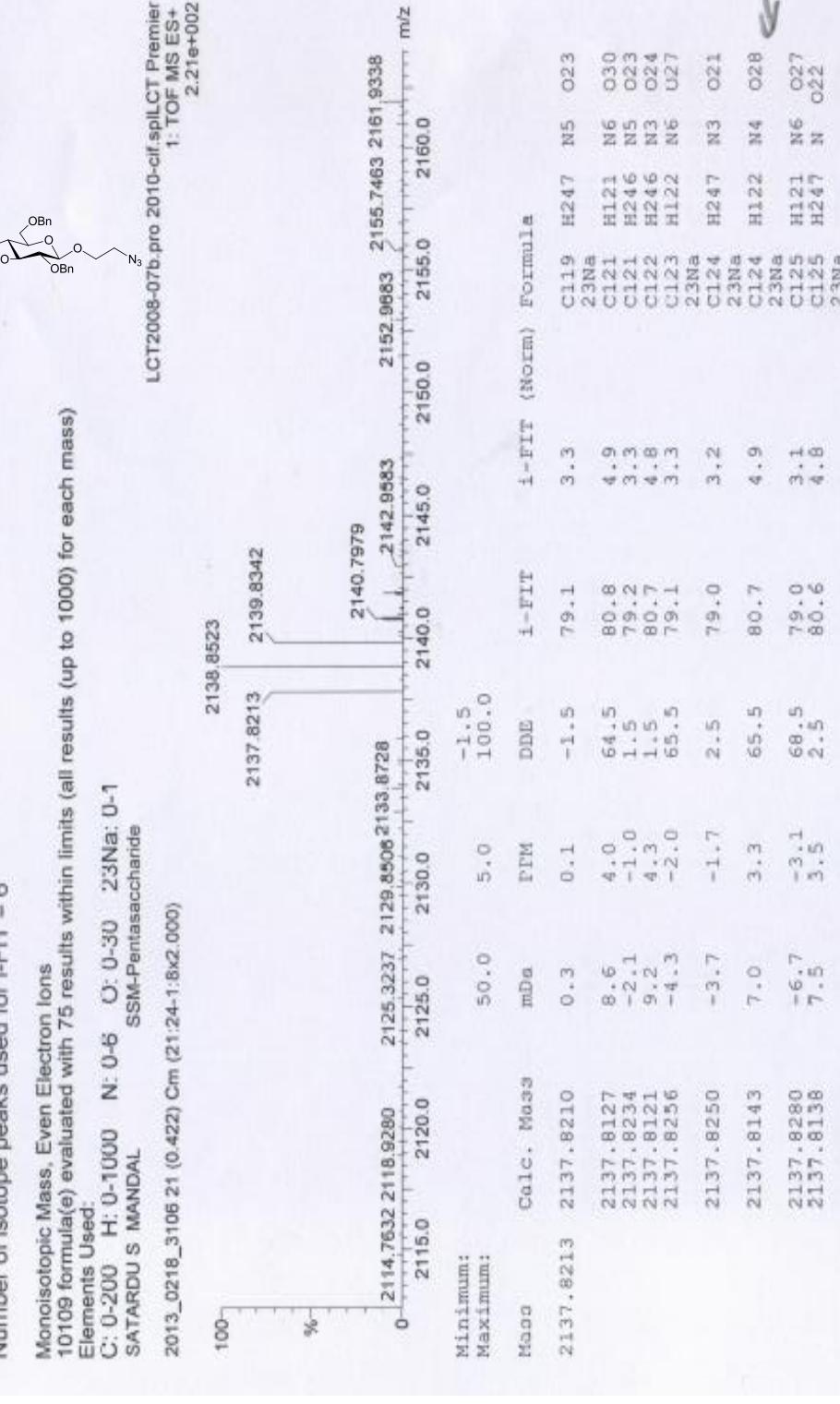
Number of isotope peaks used for i-FIT = 6

Monoisotopic Mass, Even Electron Ions  
100109 formula(e) evaluated with 75 results within limits (all results (up to 1000) for each mass)

Elements Used:  
C: U-200 H: U-1UU N: U-6 O: U-3U 23Na: U-1

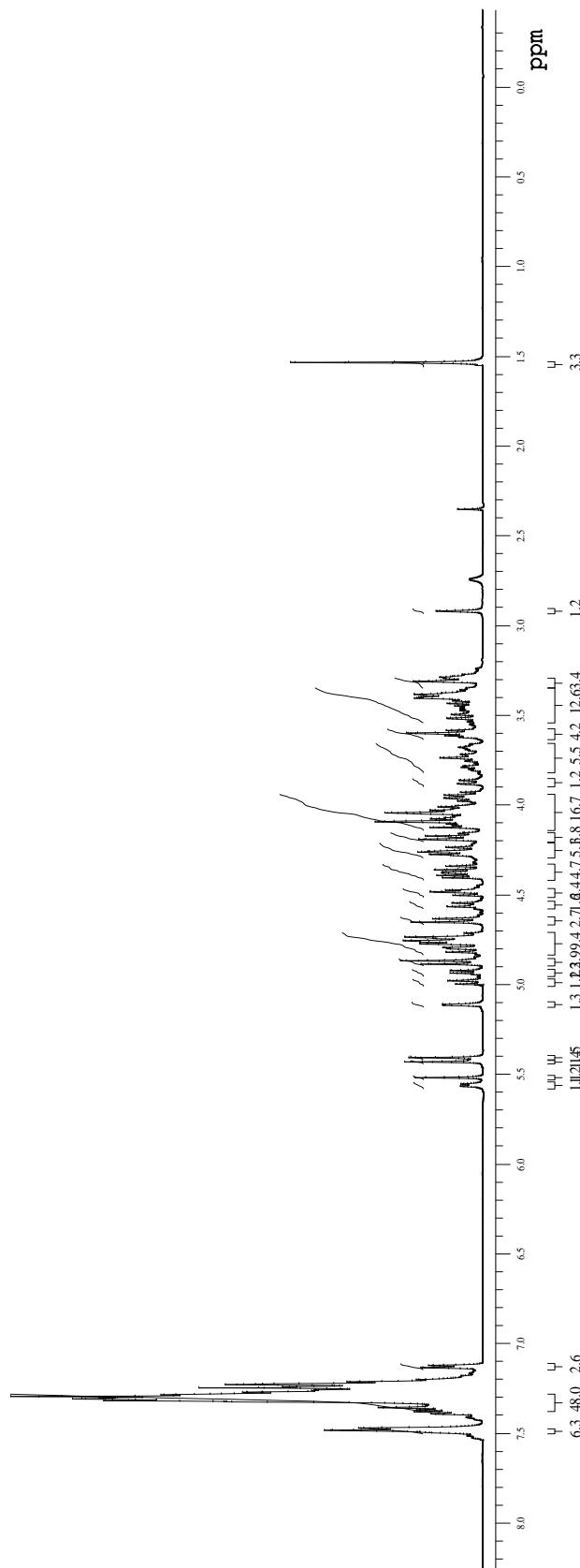
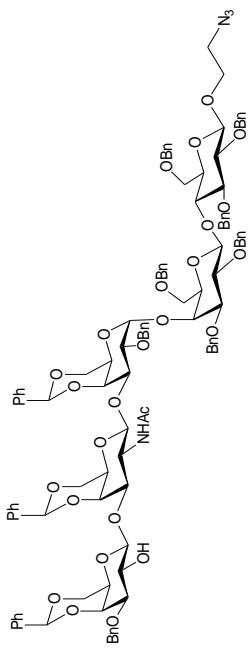
SATARDU S MANDAL

2013\_0218\_3106\_21 (0.422) Cm (21:24:1:6x2.000)



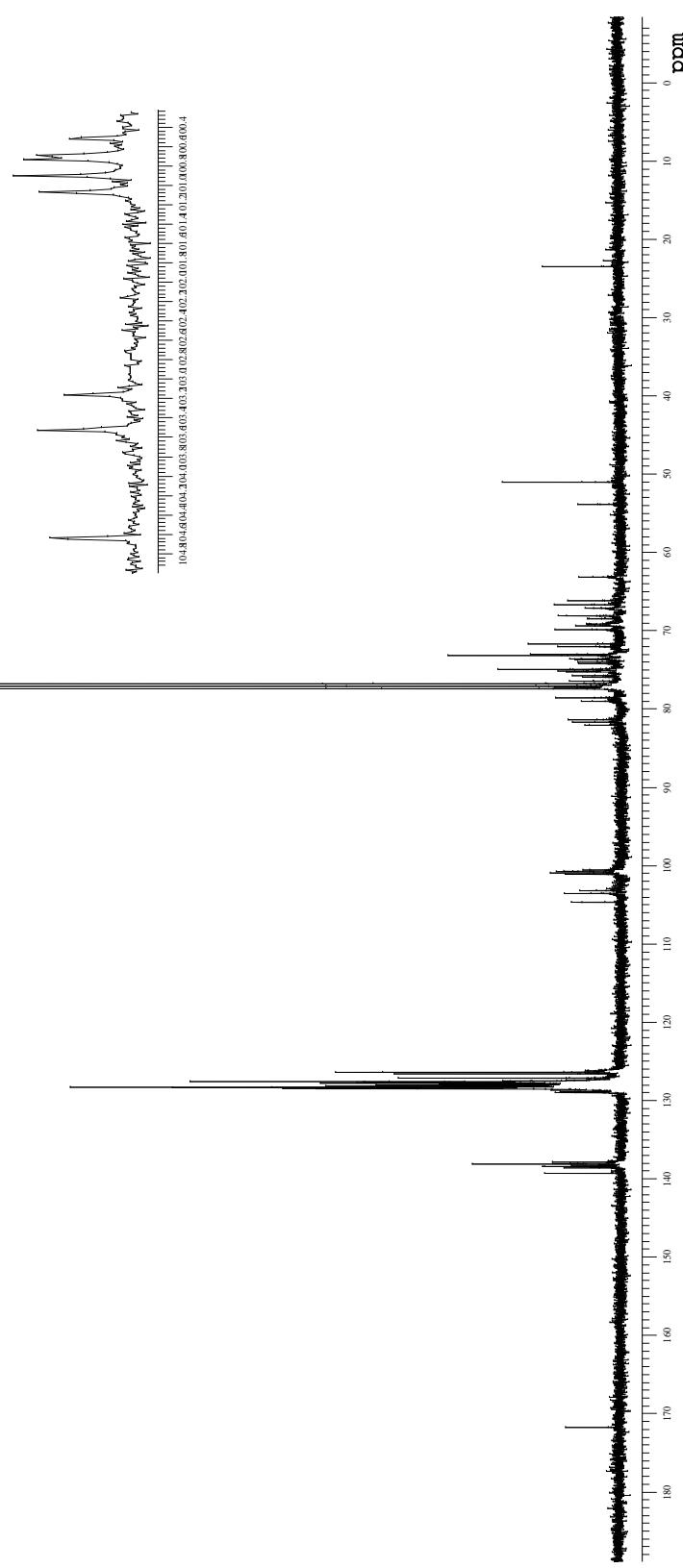
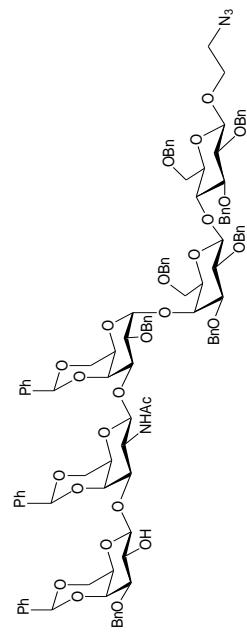
HRMS (ESI MS) spectrum of compound 3.

Agilent 600 NMR spectrometer



<sup>1</sup>H NMR Spectrum of compound **21** (CDCl<sub>3</sub>, 600 MHz)

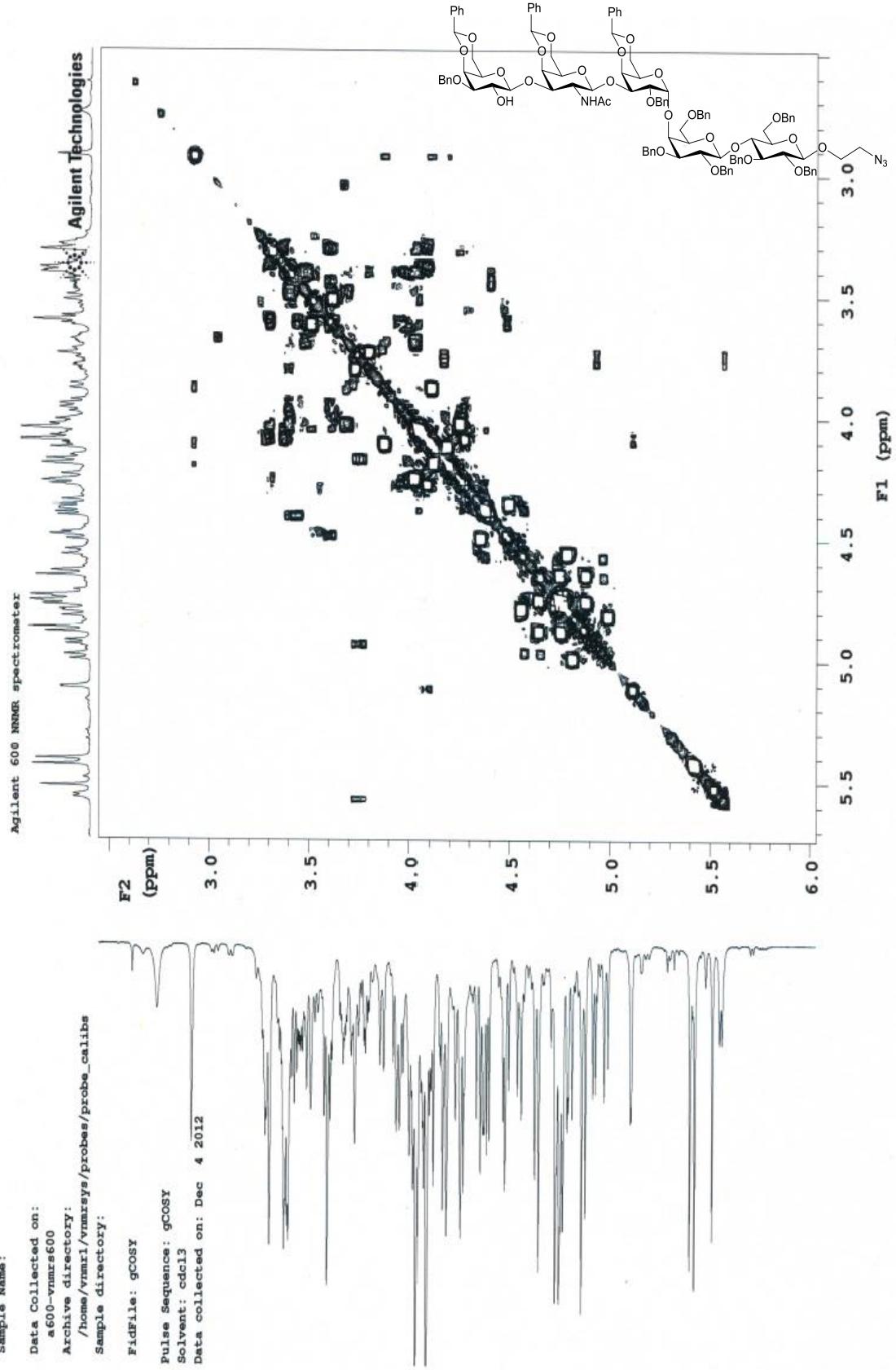
Agilent 600 NMR spectrometer



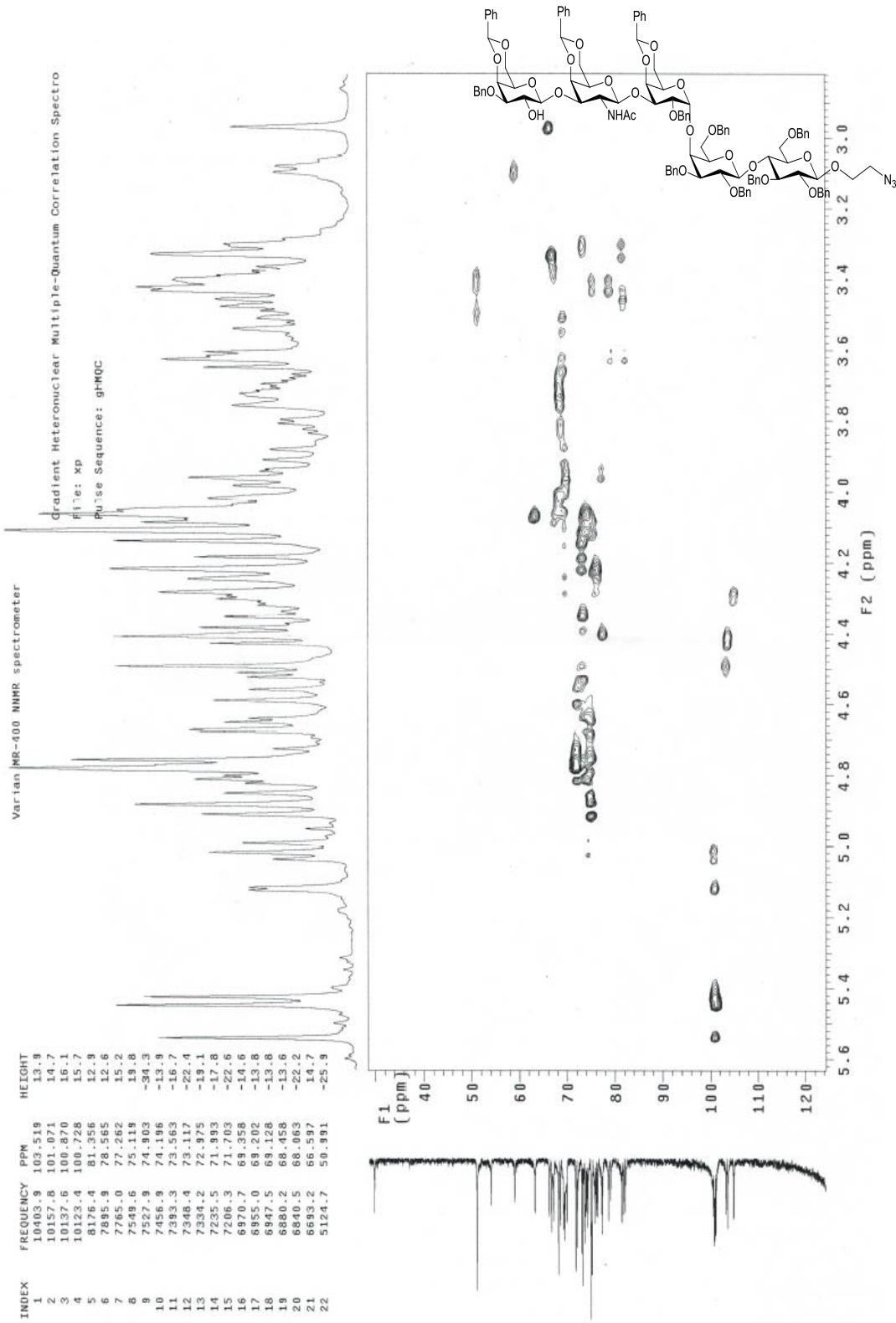
```

Sample Name:                               FidFile:  gcosy
Data Collected on:                         Solvent:  cdc13
                                           Data collected on: Dec 4 2012
   a600/vmr600                           Archive directory:
                                           /home/vmrml/vmrssys/probes/probe_calibs
Archive directory:                         Sample directory:
                                           /home/vmrml/vmrssys/probes/probe_calibs
Sample directory:                         FidFile:  gcosy

```



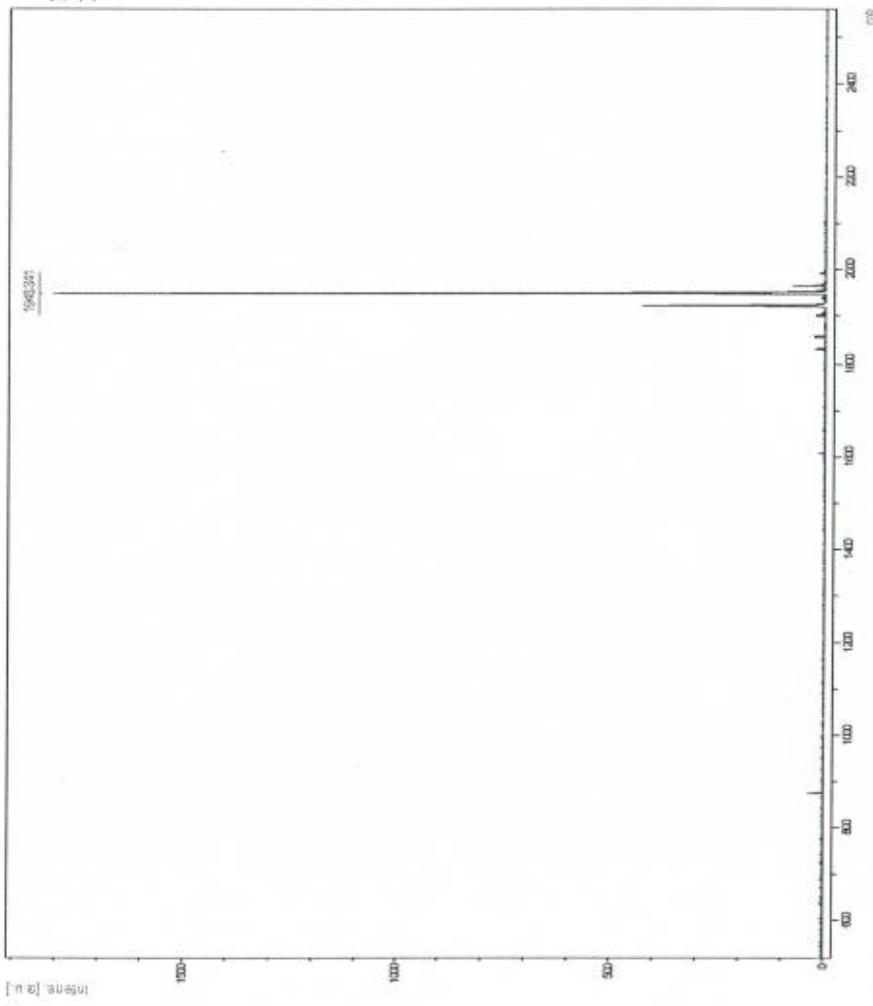
<sup>1</sup>H-<sup>1</sup>H COSY Spectrum of compound **21** (CDCl<sub>3</sub>, 600 MHz)



$^1\text{H}$ - $^{13}\text{C}$  HMQC NMR Spectrum of compound **21** ( $\text{CDCl}_3$ , 400/100 MHz)

D:\Data\Guo\_Lab\S S Mandal\Globo-H\penta-OH-NHAc-pure

Comment 1      pent-OH-NHAc-pure  
Comment 2



MALDI-TOF MS spectrum of compound 21

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

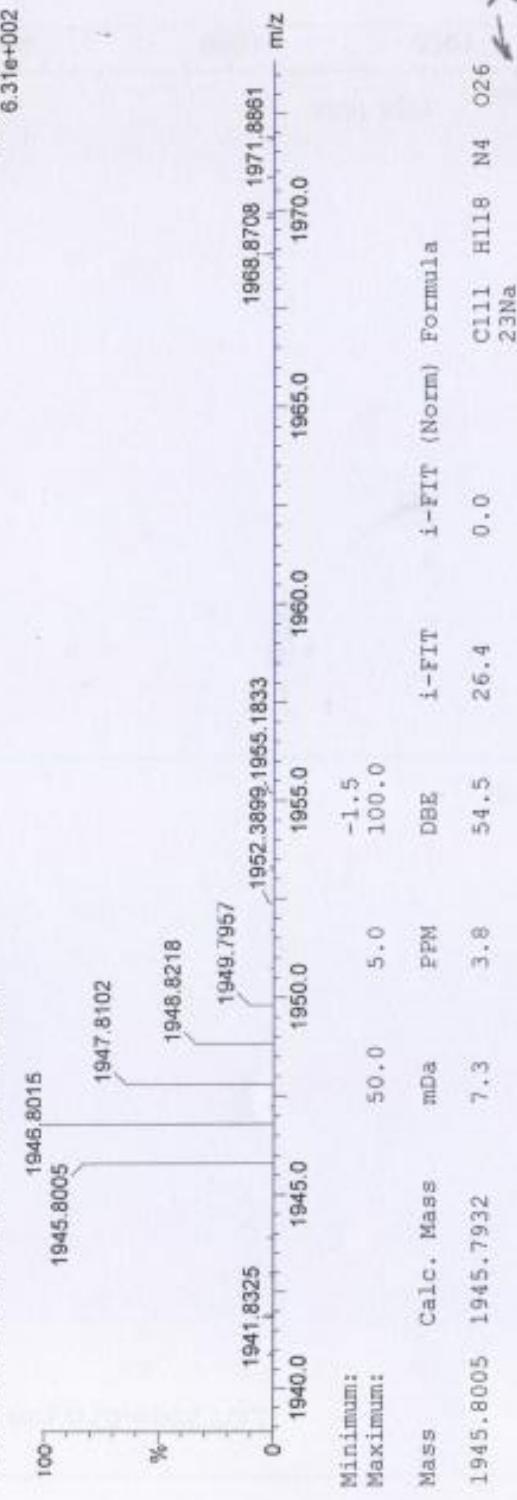
Element prediction: Off

Number of isotope peaks used for i-FIT = 6

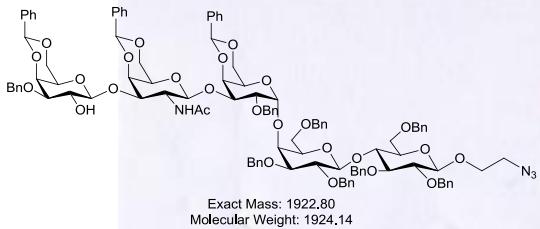
Monoisotopic Mass, Even Electron Ions  
421 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)  
Elements Used:

C: 0-111 H: 0-1000 N: 0-4 O: 0-26 23Na: 0-1  
SSM-OH-NHAc-Penta  
SATARDU S MANDAL

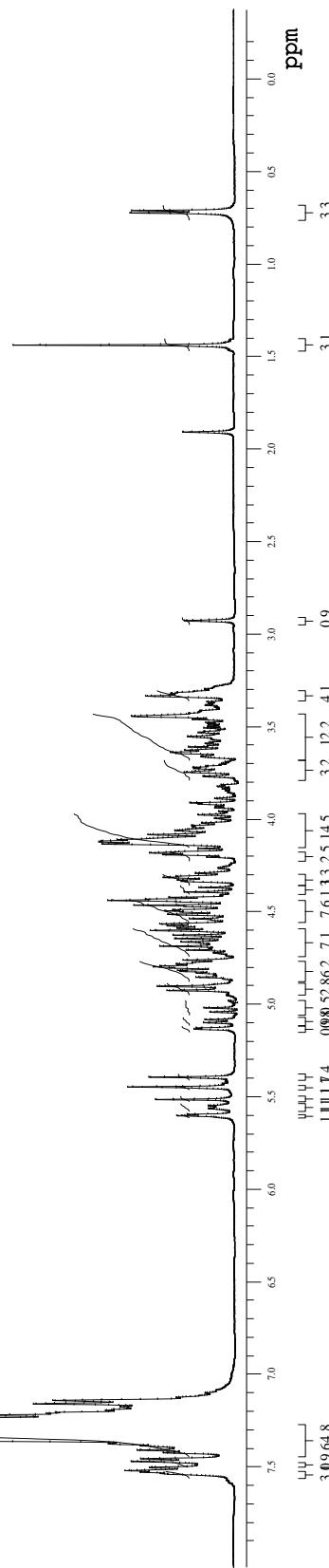
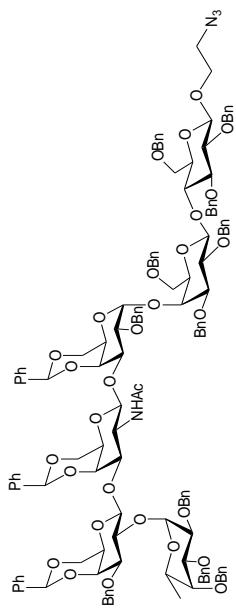
2013\_0218\_3105\_16 (0.317) Cm (16.20-1.7x2.000)



Page 1

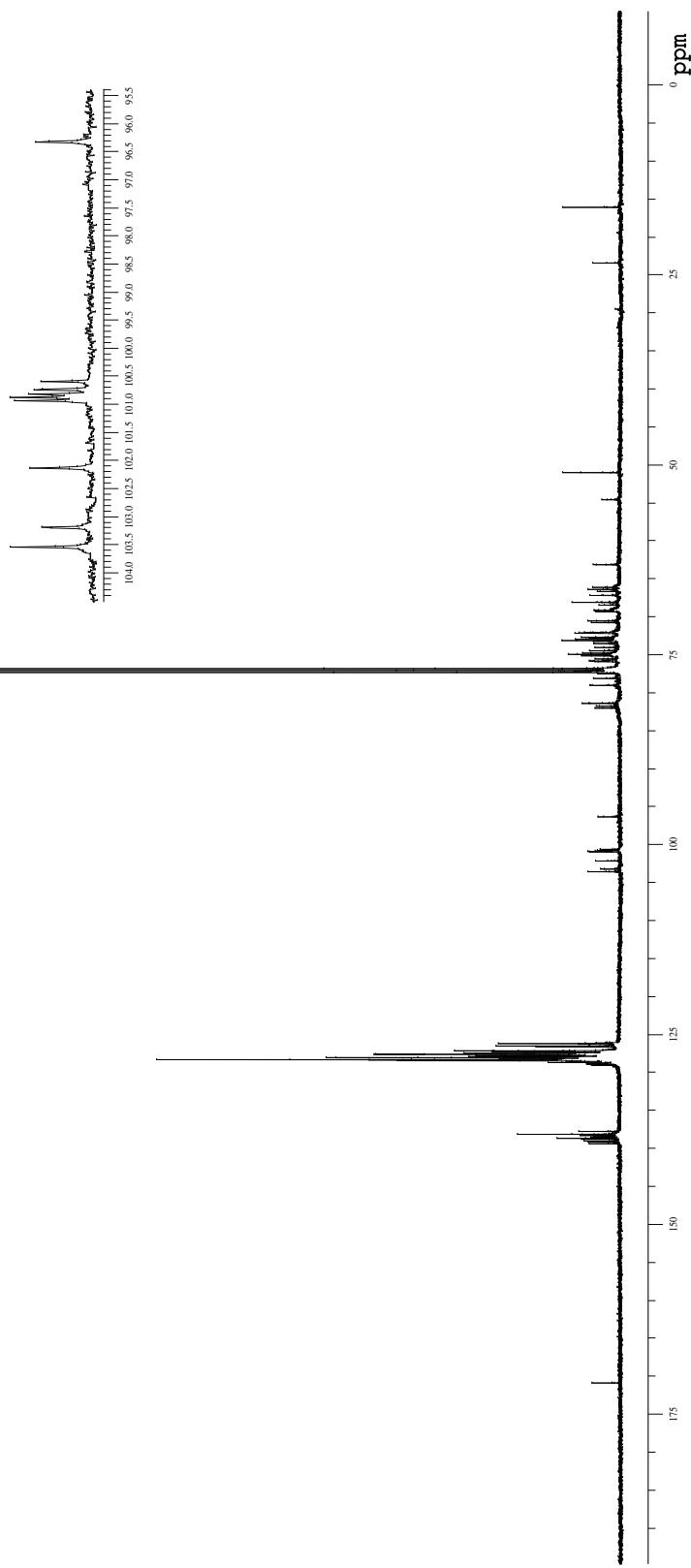
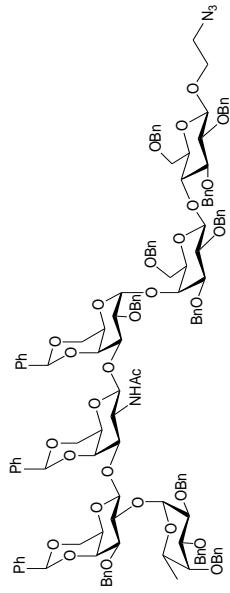


Varian 500 NMR spectrometer



<sup>1</sup>H NMR Spectrum of compound **22** (CDCl<sub>3</sub>, 500 MHz)

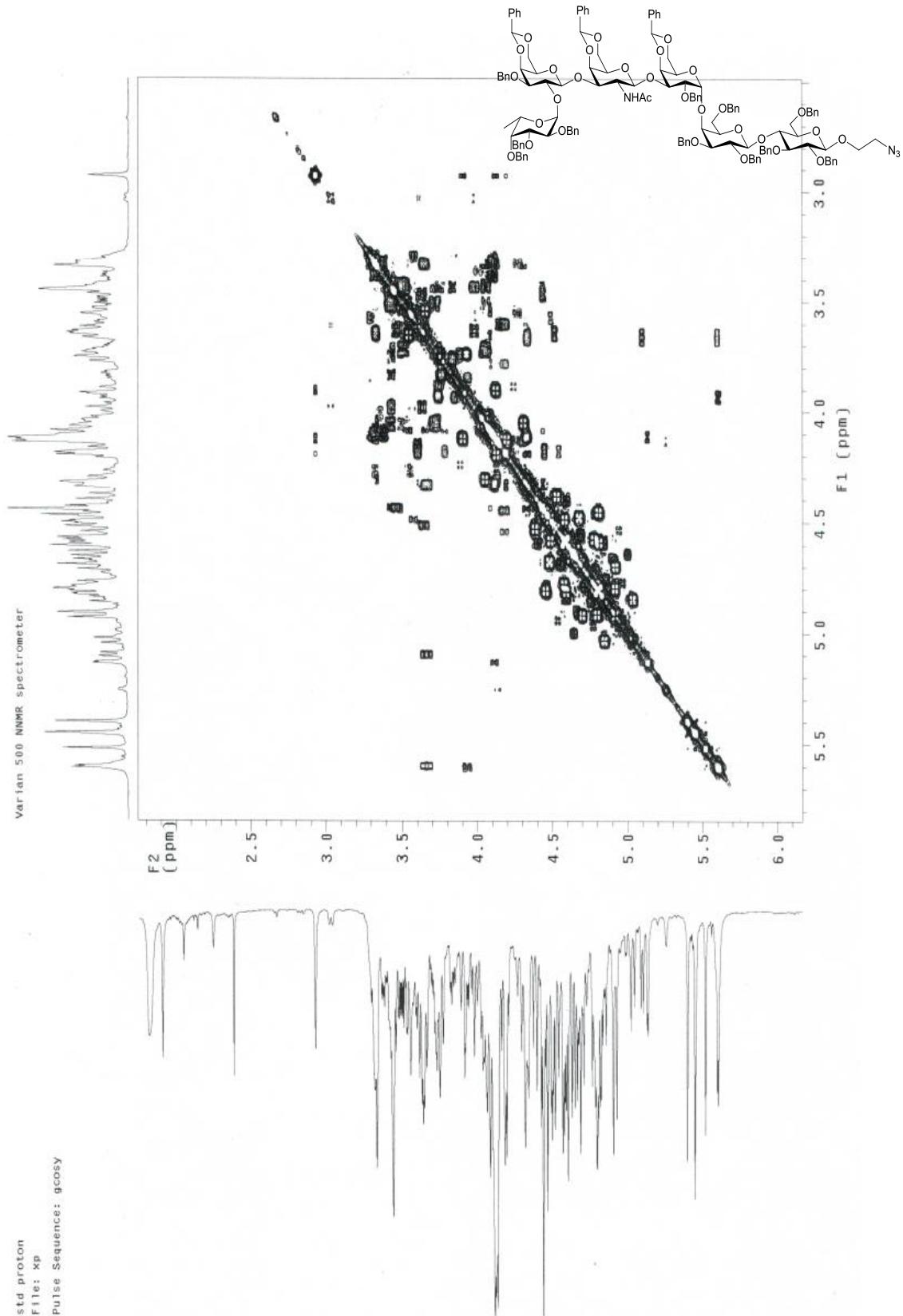
Varian 500 NMR spectrometer



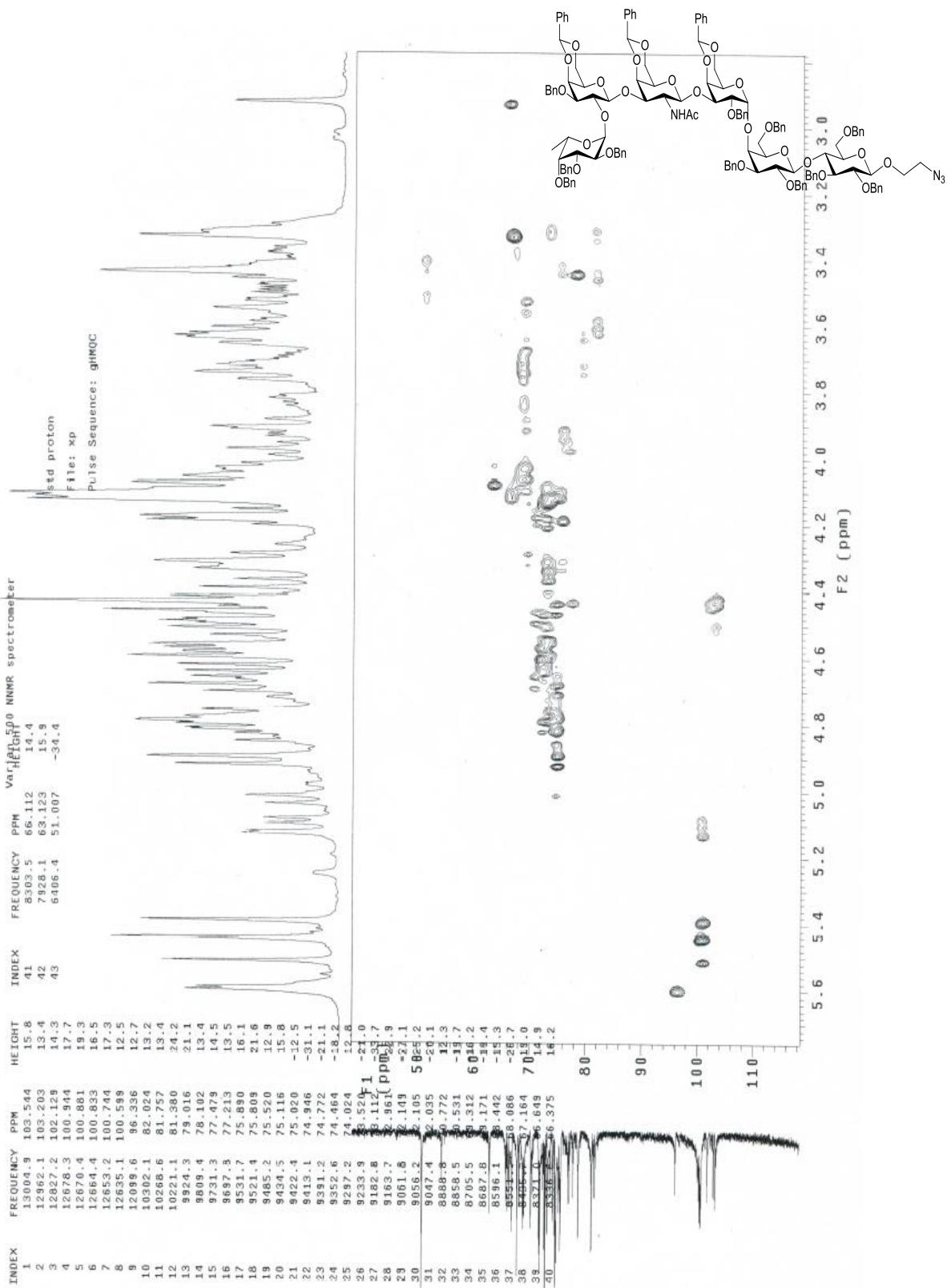
$^{13}\text{C}$  NMR Spectrum of compound **22** ( $\text{CDCl}_3$ , 500 MHz)

std proton  
File: xp  
Pulse Sequence: gcosy

Variian 500 NMR spectrometer



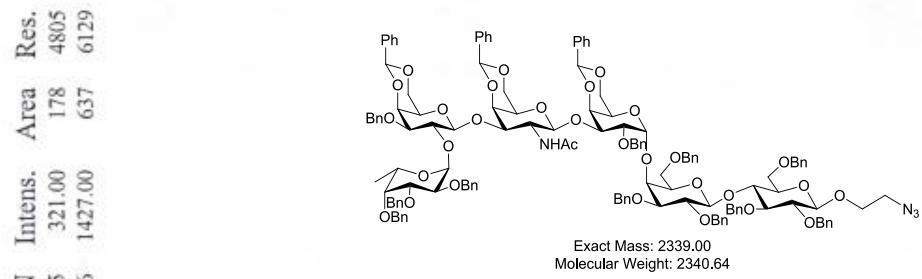
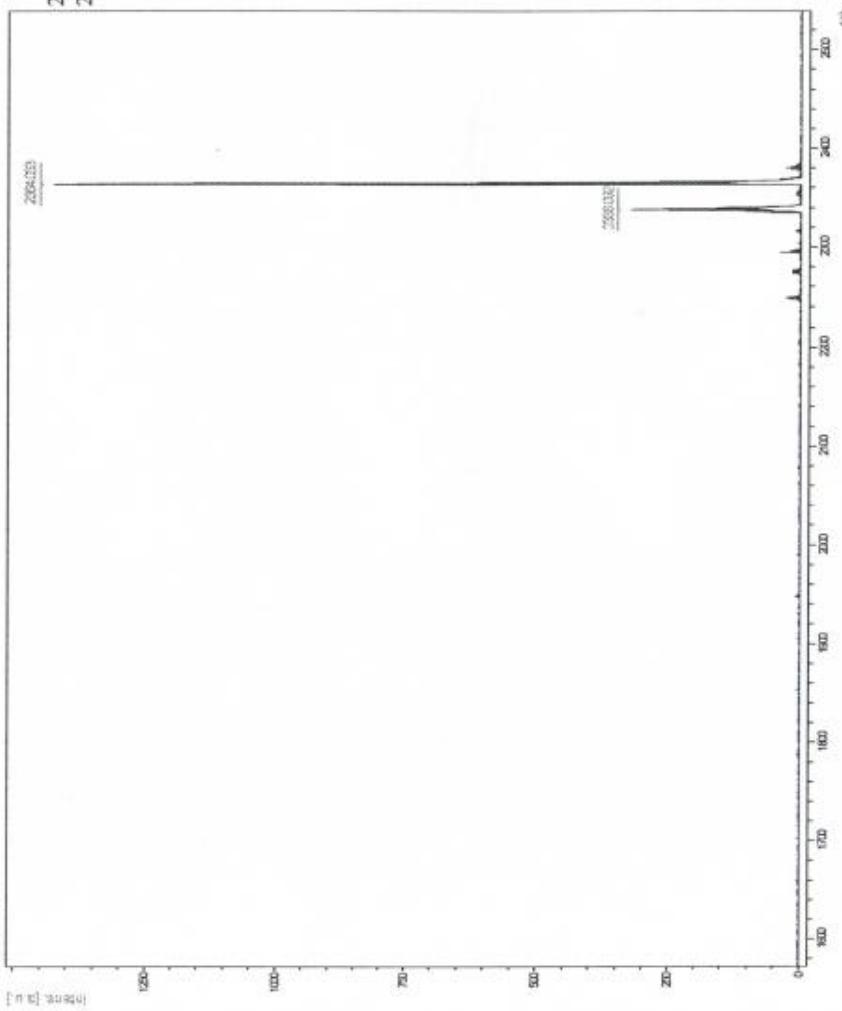
$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound **22** ( $\text{CDCl}_3$ , 500 MHz)



<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **22** ( $\text{CDCl}_3$ , 500/125 MHz)

D:\Data\Guo\_Lab\S Mandal\Globo-H\hexa-pure

Comment 1      hexa-pure  
Comment 2



**Acquisition Parameter**  
D:\Methods\1User\_Methods\GuoRP\_3147\_PepMixDHB\_June 1-2011.par

MALTI-TOF MS spectrum of compound 22

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

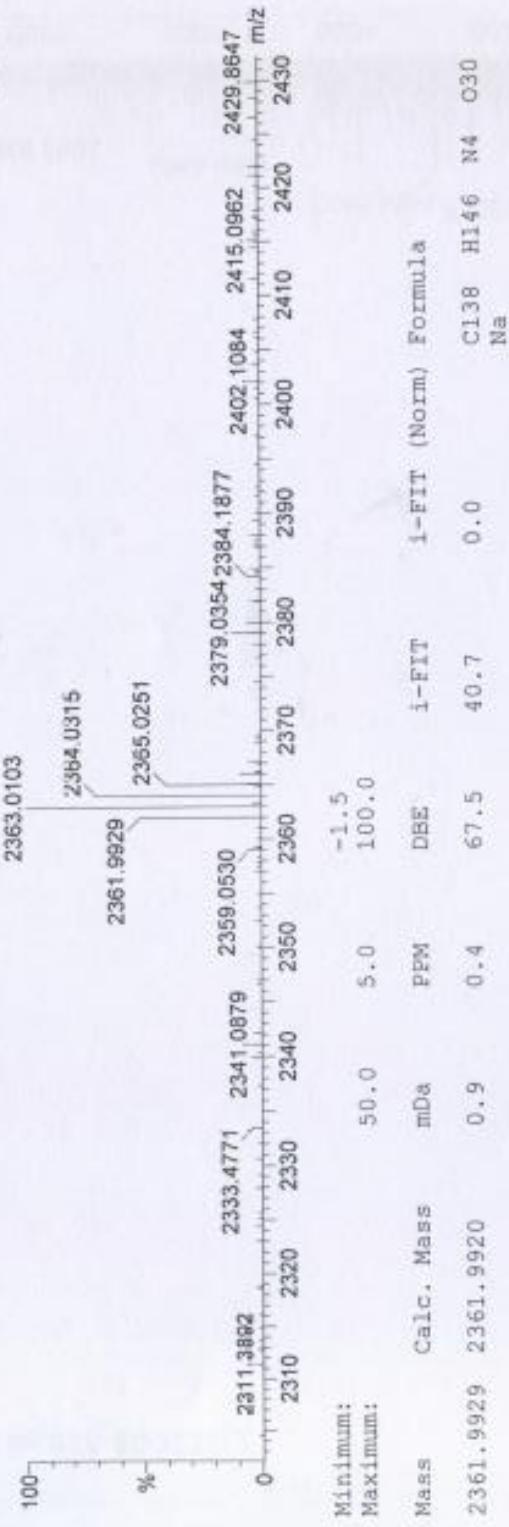
Monoisotopic Mass, Even Electron Ions  
343 formula(e) evaluated with 1 results within limits (all results (up to 1000) for each mass)

Elements Used:

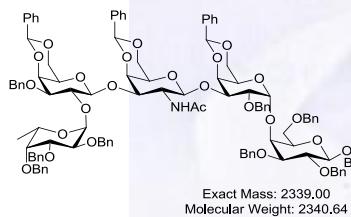
C: 0-130 H: 0-200 N: 0-5 O: 0-30 Ne: 0-1

SATARDU S MANDAL SSM-Hexasaccharide

2013\_0228\_3145\_14 (0.283) Cm ((14+26+34)-1.8x2.000)

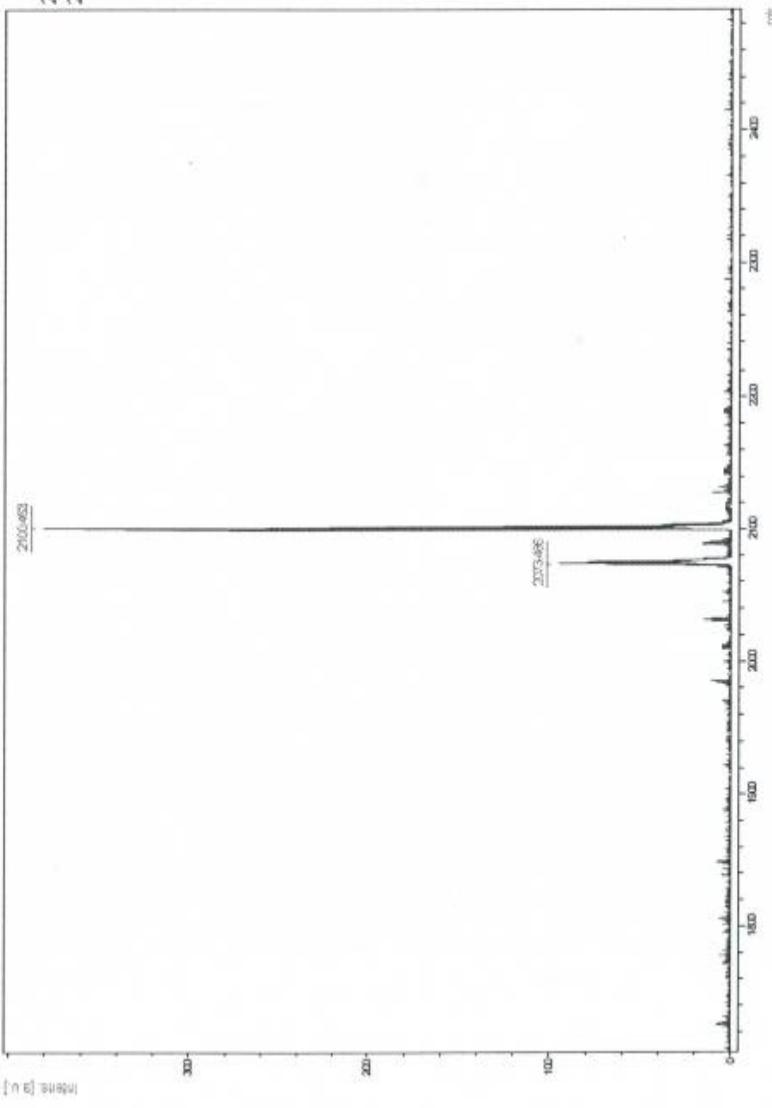


HRMS (ESI MS) spectrum of compound 22.

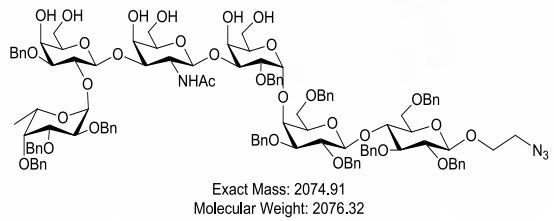


D:\Data\Guo\_lab\SSManda\Globo-H\Hexasaccharide-Debenzilidine

*Comment 1* Hexasaccharide-Debenzilidine  
*Comment 2* DHB



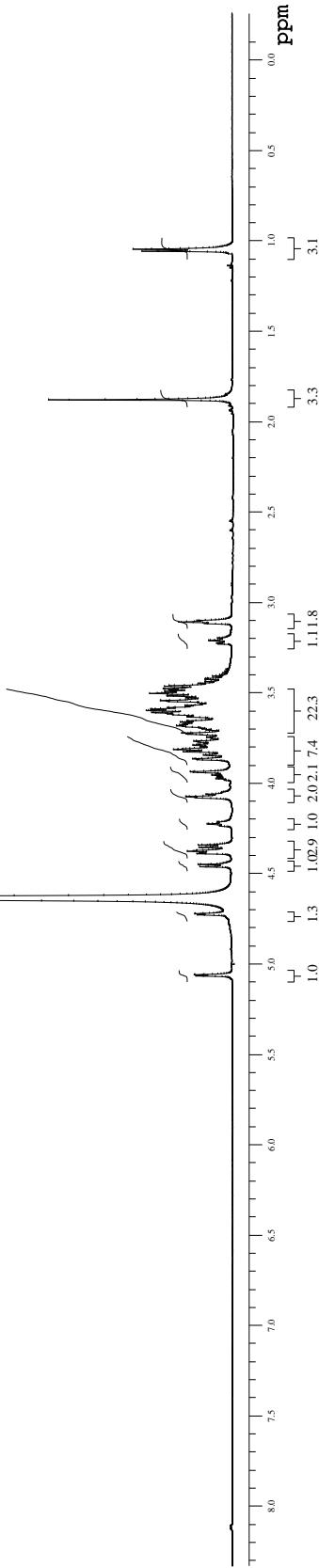
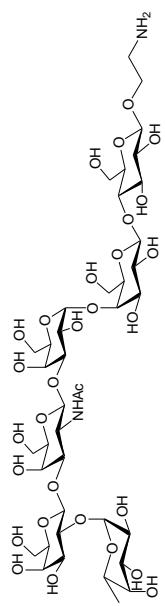
### MALDI-TOF MS spectrum of compound **22a**



Exact Mass: 2074.91  
Molecular Weight: 2076.32

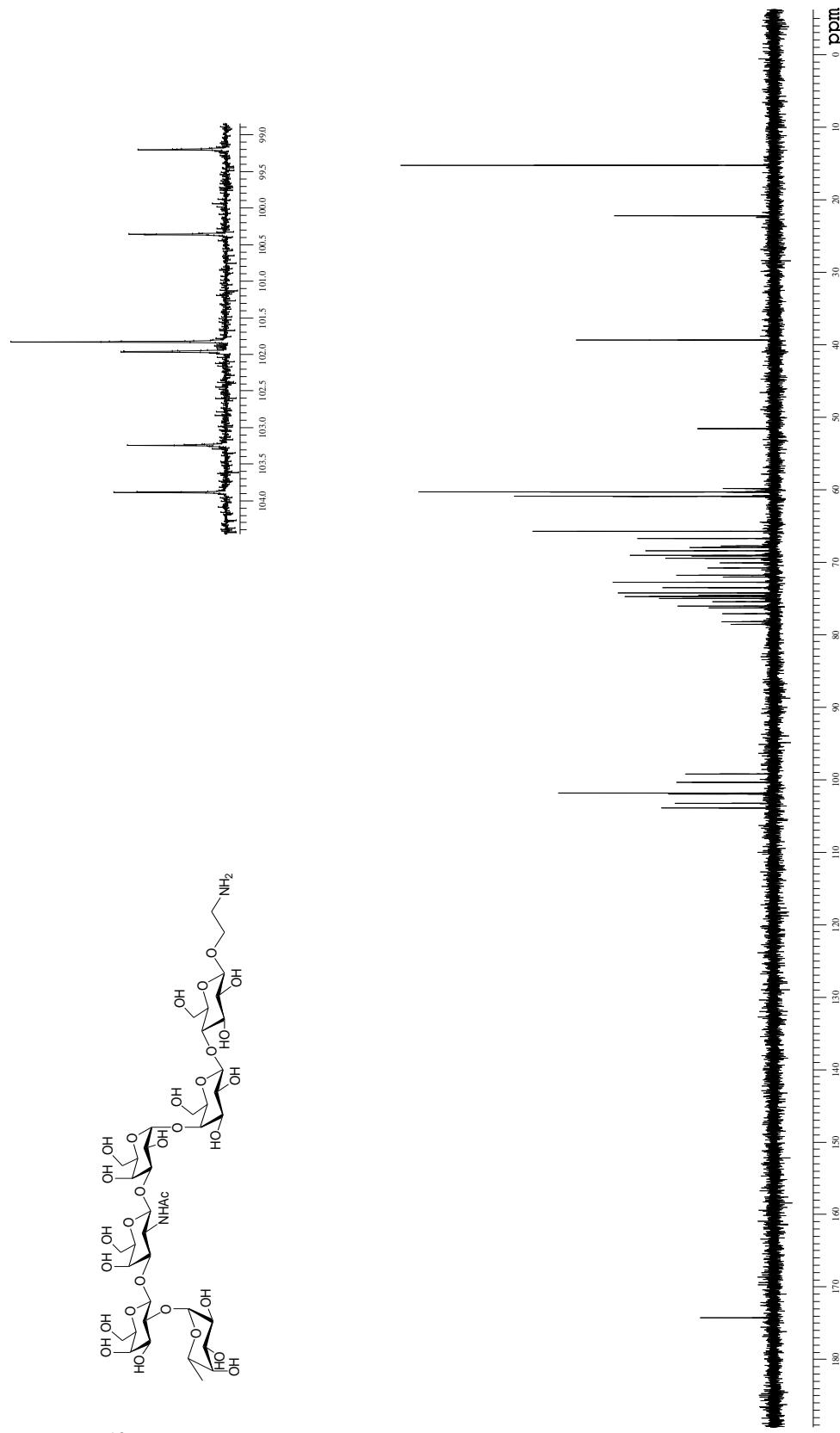
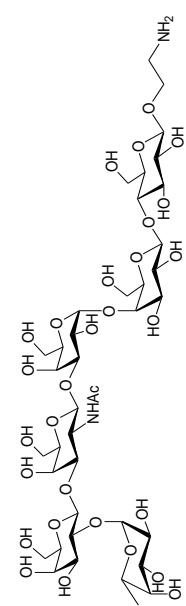
Acquisition Parameter

Agilent 600 NMR spectrometer

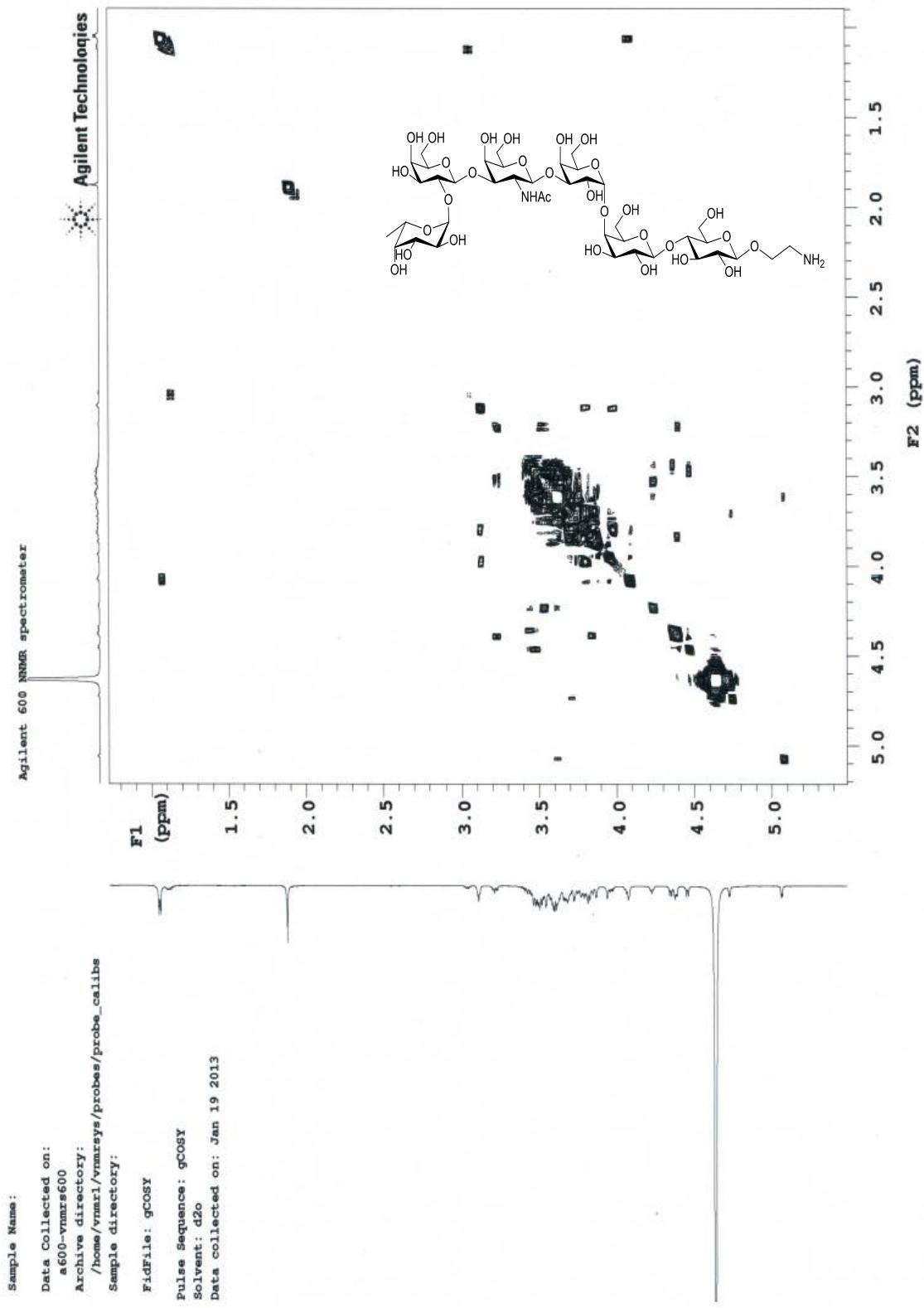


$^1\text{H}$  NMR Spectrum of compound 2 ( $\text{D}_2\text{O}$ , 600 MHz)

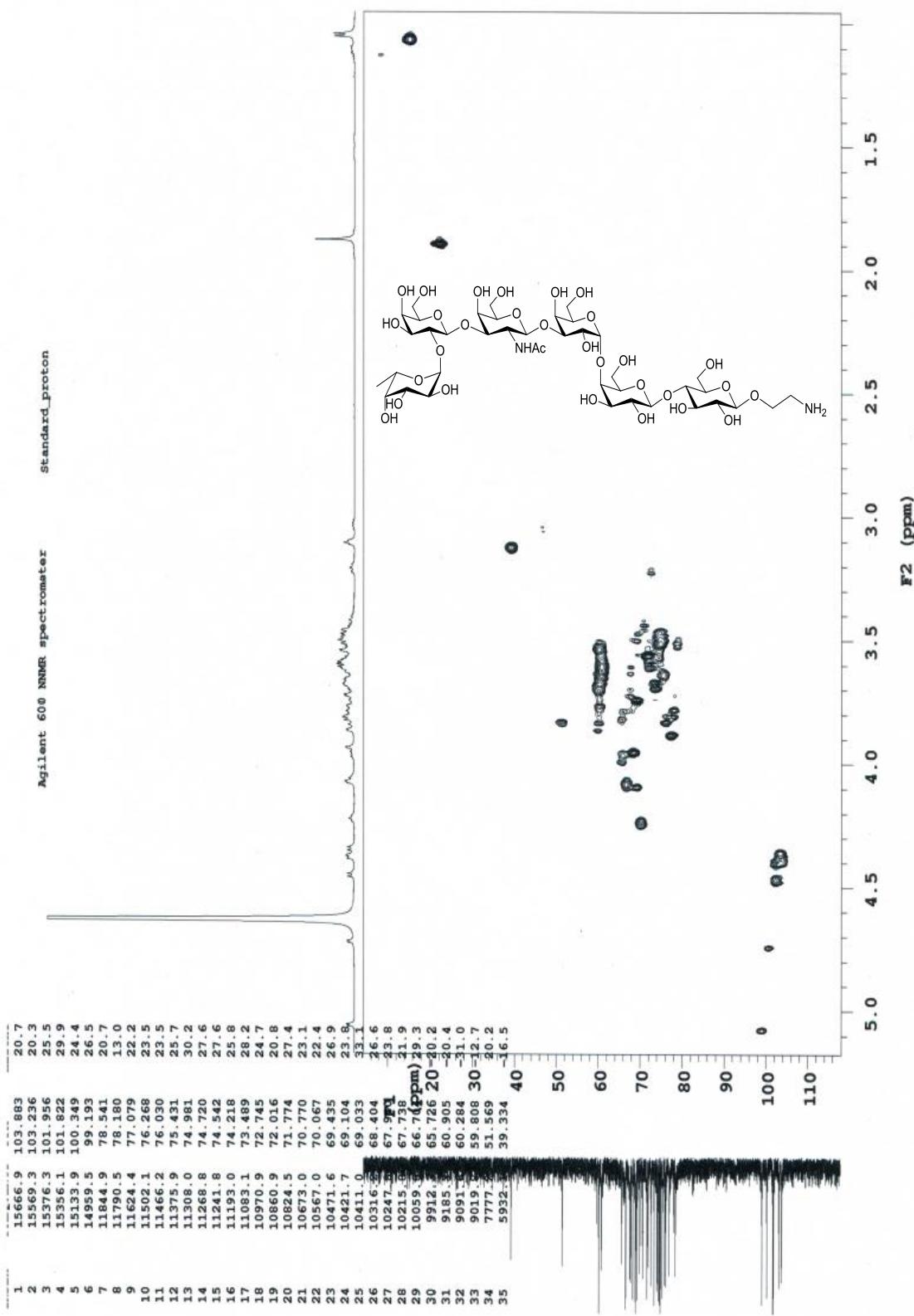
Agilent 600 NMR spectrometer



<sup>13</sup>C NMR Spectrum of compound 2 (D<sub>2</sub>O, 600 MHz)



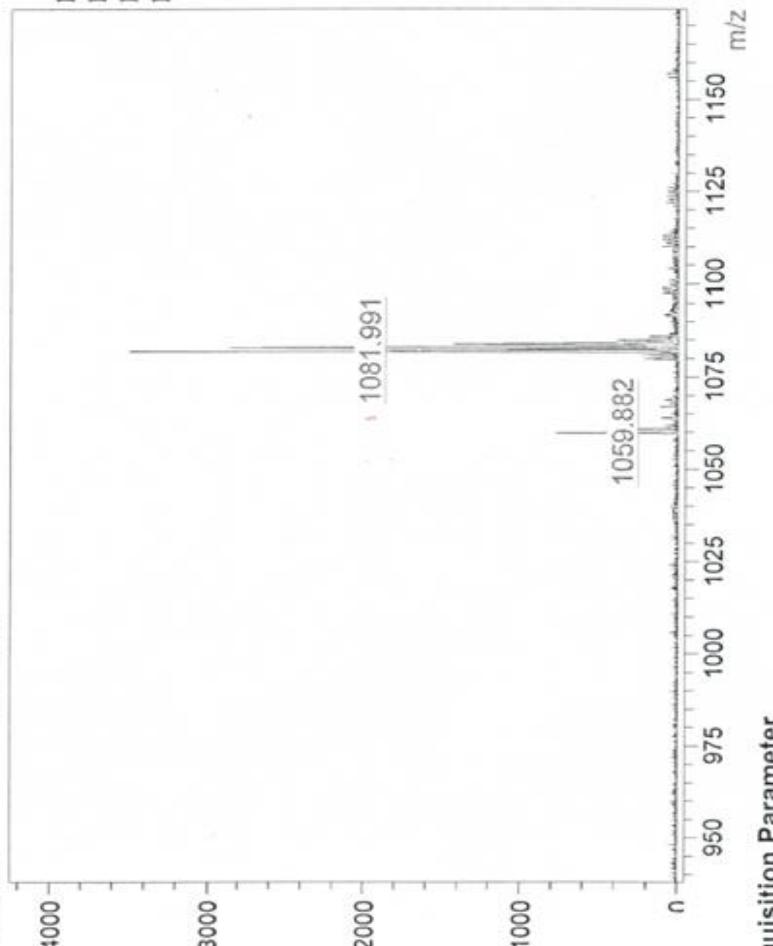
$^1\text{H}$ - $^1\text{H}$  COSY Spectrum of compound 2 ( $\text{D}_2\text{O}$ , 600 MHz)



<sup>1</sup>H-<sup>13</sup>C HMQC NMR Spectrum of compound **2** ( $\text{D}_2\text{O}$ , 600/150 MHz)

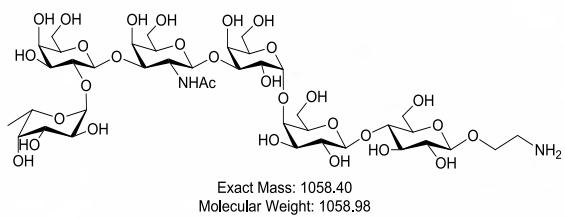
D:\Data\Guo\_Lab\S S Mandal\New Folder\Hexa-GD

Comment 1      DHB-Positive  
Comment 2      Hexa-GD



Acquisition Parameter

D:\Methods\1User\_Methods\Guo\RP\_3147\_PepMixDHB\_June 1-2011.par



MALTI-TOF MS spectrum of compound 2

## Elemental Composition Report

### Single Mass Analysis

Tolerance = 5.0 PPM / DBE: min = -1.5, max = 100.0

Element prediction: Off

Number of isotope peaks used for i-FIT = 6

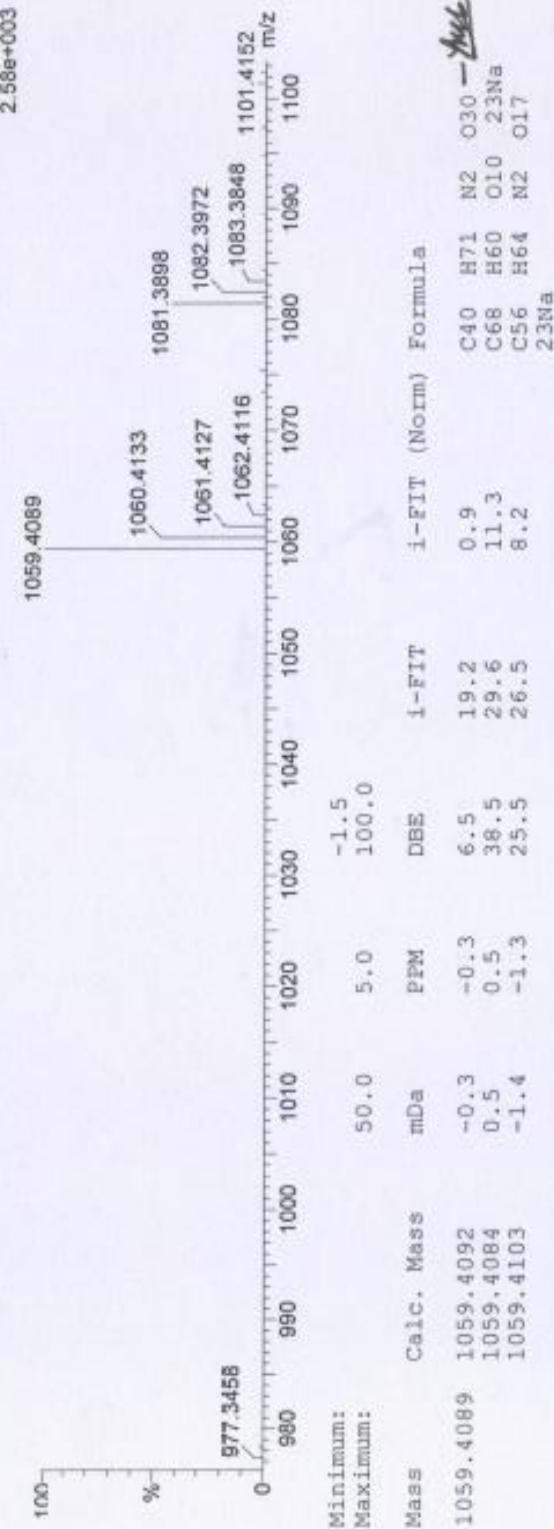
Monoisotopic Mass, Even Electron Ions  
2885 formula(e) evaluated with 14 results within limits (all results (up to 1000) for each mass)

Elements Used:

C: 0-100 H: 0-1000 N: 0-3 O: 0-35 23Na: 0-1

SATARDU S MANDAL SSM-Globo-H

2013\_0215\_3096 14 (0.283) Cm (10:14-1.6x2,000)



HRMS (ESI MS) spectrum of compound 2

