# Supporting information:

# Enantioselective sulfoxidation reaction catalyzed by G-

# quadruplex DNA metalloenzyme

Mingpan Cheng,<sup>ab</sup> Yinghao Li,<sup>ab</sup> Jun Zhou,<sup>a</sup> Guoqing Jia,<sup>a</sup> Sheng-Mei Lu,<sup>a</sup> Yan Yang <sup>a</sup> and Can Li<sup>\*a</sup>

<sup>a</sup> State Key Laboratory of Catalysis, Dalian Institute of Chemical Physics, Chinese Academy of Sciences, Dalian 116023, China. E-mail: canli@dicp.ac.cn.
<sup>b</sup> University of Chinese Academy of Sciences, No. 19A Yuquan Road, Beijing, 100049, China.

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## **Materials**

All DNA oligodeoxynucleotides (purified by PAGE) and 3-(N-morpholino) propanesulfonic acid (MOPS) were purchased from Sangon (Shanghai, China). The concentration of DNA was determined in ddH<sub>2</sub>O by the absorbance value at 260 nm at 20 °C. The extinction coefficient of each oligodeoxynucleotide was obtained from IDT website (http://www.idtdna.com/calc/analyzer). All sulfides, phenyl methyl sulfoxide, phenyl methyl sulfone, 1,10-phenanthroline (L1), 2,2'-bipyridine (L3), 4,4'-bimethyl-2,2'-bipyridine (L4) and 4,4'-bimethoxy-2,2'-bipyridine (L5) were purchased from J&K (Beijing, China). 5,6-dimethyl-1,10-phenantiroline (L2) was purchased from TCI. Cu(NO<sub>3</sub>)<sub>2</sub>•3H<sub>2</sub>O (>99%), KCl (>99%) and NaCl (>99%) were purchased from the Shanghai Chemical Reagent Company of the Chinese Medicine Group. Copper complexes were prepared according to the previous procedure (Yinghao Li, Can Li, *et al., Tetrahedron*, 2013, **69**, 6585-6590). All reagents and solvents were used without further purification. Water was distilled and deionized using a Milli-Q A10 water purification system.

# **Typical oxidation procedure**

Sulfides oxidation procedure: DNA (10  $\mu$ M) was added to a 3-(N-morpholino) propanesulfonic acid (MOPS) buffer (1 mL, 20 mM, pH 7.0) containing KCl (150 mM), heated for 3 min at 98 °C and annealed slowly over the course of 2 h to room temperature. Then, an aqueous solution of copper complex (50  $\mu$ M) was added, stirring for 0.5 h at 15 °C. Then, sulfide dissolved in CH<sub>3</sub>CN (10  $\mu$ L of a 0.5 M solution) was added. The reaction was started with the addition of H<sub>2</sub>O<sub>2</sub> (7.5  $\mu$ L of 3 % aqueous solution). The reaction mixture was stirred for another 5 h at reaction temperature. Then diethyl ether (3mL×3) was added to extract the products and remained substrate. After drying the extracted mixture with anhydrous Na<sub>2</sub>SO<sub>4</sub> and removal of the solvent under reduced pressure, the crude products were directly analyzed by a chiral HPLC.

## **Characterization Experiments**

DNA samples (5  $\mu$ M) dissolved in MOPS buffer (20 mM, pH 7.0) containing KCl (150 mM) or NaCl (150 mM) were heated for 3 min at 98 °C and slowly annealed over the course of 2 h to room temperature. After that, these aqueous samples were incubated at 4 °C overnight to ensure the equilibrium between folding and unfolding. CD measurements (220 to 360 nm) were done on a dual beam DSM 1000 CD spectrophotometer (Olis, Bogart, GA) at 20 °C. The CD spectra presented are the average of five consecutively measured scans. UV-Vis absorption (250 to 360 nm) experiments were carried out on Shimadzu 2450 spectrophotometer (Shimadzu, Japan) at room temperature. UV melting (20~95 °C, 0.5 °C/min) experiments were done on Shimadzu 2450 spectrophotometer equipped with a Peltier temperature control accessory. The concentration of CuL4 was 25  $\mu$ M. All samples were dissolved in 20 mM MOPS buffer (pH 7.0). All spectra were recorded using a sealed quartz cell with a path length of 1.0 cm. High-performance liquid chromatography (HPLC) analysis was performed on an Agilent 1100 Series instrument with the eluents of hexane and isopropanol (*i*-PrOH), using a Daicel Chiralcel ODH or ASH column.

# Calculation the conversion of 1a~m sulfides

Conversion of **1a** is calculated using the following formula:

Conversion of  $1a (\%) = PA_{2a}/(PA_{2a} + PA_{1a}/f) *100$ ,

Where  $PA_{1a}$  and  $PA_{2a}$  are the HPLC peak areas of 1a and 2a, respectively. *f* is the correction factor determined to be 0.45 from a fitting curve (Figure S1).



*Fig. S1* Determination of the correction cofactor between **1a** and **2a**. The HPLC ratios of peak areas  $(PA_{2a}/PA_{1a})$  were determined with the standard molar ratios  $(n_{2a}/n_{1a})$  of 1/10, 1/5, 1/2, 1, 2, 5, 10.

	1	2	R <sub>1</sub>	R <sub>2</sub>	$oldsymbol{f}^{a}$	HPLC Method <sup>b</sup>	<b>Retention</b> Time /		
Entry							min		
							1	2	2
1	1a	2a	phenyl	methyl	0.45	Ox-1	3.9	18.3	25.3
2	1b	2b	2'-chlorophenyl	methyl	0.39	Ox -2	4.3	26.1	33.7
3	1c	2c	3'-chlorophenyl	methyl	0.42	Ox -2	4.0	33.9	53.2
4	1d	2d	3'-tolyl	methyl	0.37	Ox -2	3.8	32.7	41.6
5	1e	2e	4'-tolyl	methyl	0.50	Ox -1	3.8	18.0	29.2
6	1f	2f	4'-methoxyphenyl	methyl	1.14	Ox -1	6.2	32.3	51.7
7	1g	2g	2'-naphthyl	methyl	0.41	Ox -1	4.3	23.3	32.5
8	1h	2h	phenyl	ethyl	0.24	Ox -1	3.8	14.4	16.3
9	1i	2i	phenyl	benzyl	0.80	Ox -3	5.8	22.9	28.7
10	1j	2j	2'-pyridine	methyl	0.53	Ox -1	4.3	12.1	19.8

*Table S1* Correction factor and HPLC conditions of substrates  $(1a \sim m)$  and products

<sup>*a*</sup>f is the correction factor.

<sup>*b*</sup> HPLC methods:

Ox-1 (Daicel chiralpak-ASH, hexane/i-PrOH 70:30, 1.0 mL/min, 250 nm);

Ox-2 (Daicel chiralpak-ASH, hexane/i-PrOH 85:15, 1.0 mL/min, 250 nm);

Ox-3 (Daicel chiralcel-ODH, hexane/i-PrOH 95:5, 1.0 mL/min, 250 nm).





*Fig. S2* CD spectra of HT21 alone and HT21•CuL4 in the presence of (a) K<sup>+</sup>, (b) Na<sup>+</sup> ion and (c) in the absence of metal ion, respectively.

Fig. S3



Fig. S3 CD spectra of rHT21 and cgmHT21 in  $K^+$  (150 mM) solution.





*Fig. S4* Optimization of HT21•CuL4 catalyzing sulfoxidation reaction conditions on conversion ( $\Box$ ) and enantioselectivity (•): (a) Concentration of potassium ion; (b) Molar ratio between CuL4 and HT21; (c) Reaction temperature and (d) pH.

Fig. S5



Fig. S5 CD spectra of HT21 in various concentrations of K<sup>+</sup>.

Fig. S6



*Fig. S6* UV/Vis absorption spectra of (I) CuL4 (25  $\mu$ M) alone; (II) CuL4 (25  $\mu$ M) with HT21 (0.5  $\mu$ M) and (III) HT21 (2.5  $\mu$ M) in MOPS buffer (20 mM, pH 7.0).

Fig. S7



Fig. S7 UV-melting curves of HT21 with and without CuL4.

## **Oxidative degradation of DNA**

In order to figure out whether the degradation of DNA happened or not, CD spectroscopy and electrophoresis experiments were conducted. CD spectra of HT21 G-quadruplex/CuL complexes were obtained after adding 7.5 mM H<sub>2</sub>O<sub>2</sub> (Fig. S8). The profile of CD spectra are not significantly changed: the characteristic positive bonds at 295 and 270 nm exist. And the little decrease of intensity at 295 and 270 nm may indicate the slight oxidative degradation of DNA. Thus, we can conclude that slight degradation of DNA happens and structure of G-quadruplex remains. In addition, the DNA integrity was investigated by using SDS-PAGE assay in which HT21/CuL4 samples were incubated with or without 7.5 mM H<sub>2</sub>O<sub>2</sub> (Fig. S9). The slight degradation of DNA metalloenzyme can be observed by using loading samples with high concentration (20  $\mu$ L of 200  $\mu$ M DNA). Considering that the oxidation of thioanisole catalyzed by H21 G-quadruplex/CuL4 metalloenzyme can achieve 99 % conversion, this slight degradation of DNA may not influence the catalytic performance significantly.



*Fig. S8* CD spectra of HT21 G-quadruplex (5  $\mu$ M)•CuL2 (left) or CuL4 (right) (25  $\mu$ M) complexes without (black) and with (red) 7.5 mM H<sub>2</sub>O<sub>2</sub>. After the H<sub>2</sub>O<sub>2</sub> was added, the spectra was recorded at 0, 10, 20 minutes and 5 hours.



*Fig. S9* Electrophoresis of HT21 G-quadruplex•CuL4 with and without treated by 7.5 mM  $H_2O_2$ . The experiments were conducted twice.









#### HPLC trace 3. Racemic 2b:





HPLC trace 4. Sulfoxidation of 1b catalyzed by HT21 G-quadruplex•CuL4.

### HPLC trace 5. Racemic 2c:





HPLC trace 6. Sulfoxidation of 1c catalyzed by HT21 G-quadruplex•CuL4.

## HPLC trace 7. Racemic 2d:





HPLC trace 8. Sulfoxidation of 1d catalyzed by HT21 G-quadruplex•CuL4.

### HPLC trace 9. Racemic 2e:





*HPLC trace 10.* Sulfoxidation of 1e catalyzed by HT21 G-quadruplex•CuL4.

## HPLC trace 11. Racemic 2f:



HPLC trace 12. Sulfoxidation of 1f catalyzed by HT21 G-quadruplex•CuL4.









HPLC trace 14. Sulfoxidation of 1g catalyzed by HT21 G-quadruplex•CuL4.

### HPLC trace 15. Racemic 2h:





HPLC trace 16. Sulfoxidation of 1h catalyzed by HT21 G-quadruplex•CuL4.

## HPLC trace 17. Racemic 2i:





HPLC trace 18. Sulfoxidation of 1i catalyzed by HT21 G-quadruplex•CuL4.

#### HPLC trace 19. Racemic 2j:





HPLC trace 20. Sulfoxidation of 1j catalyzed by HT21 G-quadruplex•CuL4.