

**Supplemental Information**

**Impacts of Coal Ash on Methylmercury Production and the  
Methylating Microbial Community in Anaerobic Sediment Slurries**

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**Table of Contents**

Total number of pages: 18

<b>1. Methods</b>	
Speciation of Hg in Porewater.....	S3

<b>2. Tables</b>	
Table S1. Stability constants used in Hg speciation calculations.....	S4
Table S2. Experiment #1: Hg Speciation calculation inputs.....	S5
Table S3. Experiment #2: Hg Speciation calculation inputs.....	S5
Table S4. Experiment #3: Hg Speciation calculation inputs.....	S6
<b>3. Figures</b>	
SI Figure S1. Map of sampling sites on the Emory and Clinch Rivers.....	S7
SI Figure S2. Total AVS in Slurry.....	S8
SI Figure S3. Porewater Acid Volatile Sulfide concentrations in sediment-ash microcosms.....	S9
SI Figure S4. Total dissolved Iron concentrations in sediment-ash microcosms.....	S10
SI Figure S5. Porewater Ferrous Iron in sediment-ash microcosms.....	S11
SI Figure S6. Dissolved Organic Carbon concentrations in sediment-ash microcosm porewater.....	S12
SI Figure S7. Porewater total Hg in sediment-ash microcosms.....	S13
SI Figure S8: Hg Speciation in microcosm porewater.....	S14
SI Figure S9: Saturation indices for metacinnabar in microcosm porewater.....	S15
SI Figure S10: Relative abundances of methylating microorganisms.....	S16
SI Figure S11: pH in sediment-ash microcosms.....	S17
<b>4. SI References.....</b>	S18

## Methods

**Mercury Speciation Calculations.** Mercury equilibrium speciation calculations with Visual MINTEQ.<sup>1</sup> The Hg speciation calculations utilized stability constants shown in Table S1, which were selected based on the analysis described in Hsu-Kim et al. (2013).<sup>2</sup> Ionic strength was set to 0 M. Visual MINTEQ data inputs (SI Tables S4-S6) were the microcosm porewater concentrations of: H<sup>+</sup>, Fe(II), AVS, Cl, Hg, and thiols (estimated from DOC measurements). All of the microcosms were amended with 10 mM pyruvate (30 mmol L<sup>-1</sup> of organic carbon), which does not act as a strong ligand for Hg. In Experiment #3, we observed that the DOC in the porewater of the slurries was initially 33 mM right after construction of the microcosms. Thus, we estimated that approximately 10% of the measured DOC value for other microcosms was attributed to dissolved organic matter with thiol functional groups that bind Hg<sup>2+</sup>. This assumption results in approximately 1.5 – 3.8 mM (18 - 46 mg L<sup>-1</sup>) of DOC with the capacity to strongly bind Hg. We then assumed that the DOC had 5 nmol mg<sup>-1</sup> of thiols that strongly bind Hg, based on previous studies with dissolved organic matter isolates.<sup>3</sup>

**Table S1.** Stability constants used in Hg Speciation Calculations.

Reaction	$\log K (I=0, T=25^\circ C)$	Reference
$H_2S \rightleftharpoons HS^- + H^+$	-7.02	<sup>4</sup>
$HS^- \rightleftharpoons S^{2-} + H^+$	-17.4	<sup>4</sup>
$\beta\text{-HgS}_{(s)} + H^+ \rightleftharpoons Hg^{2+} + HS^-$	$\log K_{s0} = -38.7 \pm 2$	<sup>2</sup>
$Hg^{2+} + HS^- \rightleftharpoons HgSH^+$	30.2	<sup>2</sup>
$Hg^{2+} + 2HS^- \rightleftharpoons Hg(SH)_2^0$	37.7	<sup>2</sup>
$Hg^{2+} + 2HS^- \rightleftharpoons HgSH_2^-$	31.5	<sup>2</sup>
$Hg^{2+} + 2HS^- \rightleftharpoons HgS_2^{2-} + 2H^+$	23.2	<sup>4</sup>
$Hg^{2+} + RS_2^{2-} \rightleftharpoons Hg(RS_2)$	28.7	<sup>2</sup>
$RS_2^{2-} + H^+ \rightleftharpoons RS_2H^-$	8.4	<sup>2</sup>
$RS_2H^- + H^+ \rightleftharpoons RS_2H_2$	8.4	<sup>2</sup>
$Hg^{2+} + H_2O \rightleftharpoons HgOH^+ + H^+$	-3.4	<sup>4</sup>
$Hg^{2+} + 2H_2O \rightleftharpoons Hg(OH)_2^0 + 2H^+$	-6.2	<sup>4</sup>
$Hg^{2+} + 3H_2O \rightleftharpoons Hg(OH)_3^- + 3H^+$	-21.1	<sup>4</sup>
$Hg^{2+} + Cl^- \rightleftharpoons HgCl^+$	7.3	<sup>4</sup>
$Hg^{2+} + 2Cl^- \rightleftharpoons Hg(Cl)_2^0$	14	<sup>4</sup>
$Hg^{2+} + 3Cl^- \rightleftharpoons Hg(Cl)_3^-$	15	<sup>4</sup>
$Hg^{2+} + Cl^- + H_2O \rightleftharpoons HgOHCl^0 + H^+$	4.2	<sup>4</sup>
$Fe^{2+} + HS^- \rightleftharpoons FeS_{(s)}, \text{mackinawite} + H^+$	3.6	<sup>4</sup>
$Fe^{2+} + HS^- \rightleftharpoons Fe(HS)^+$	5.62	<sup>5</sup>
$Fe^{2+} + 2HS^- \rightleftharpoons Fe(HS)_{2(aq)}$	8.95	<sup>4</sup>
$Fe^{2+} + 3HS^- \rightleftharpoons Fe(HS)_3^-$	10.99	<sup>4</sup>
$Fe^{2+} + H_2O \rightleftharpoons FeOH^+ + H^+$	-9.40	<sup>4</sup>
$Fe^{2+} + 2H_2O \rightleftharpoons Fe(OH)_{2(aq)} + 2H^+$	-20.49	<sup>4</sup>
$Fe^{2+} + 3H_2O \rightleftharpoons Fe(OH)_3^- + 3H^+$	-30.99	<sup>4</sup>
$Fe^{2+} + Cl^- \rightleftharpoons FeCl^+$	-0.20	<sup>4</sup>

**Table S2.** Experiment 1: Microcosm porewater concentrations used as input parameters in the Visual MINTEQ calculations.

<b>Experiment 1: Sediment-Only</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	6.48	0.118	0.009	0.085	1.62E-04	2.17E-07
24	6.55	0.003	0.008	0.090	1.70E-04	1.48E-07
48	6.50	0.003	0.011	0.092	1.66E-04	1.52E-07
96	6.39	0.003	0.010	0.091	1.63E-04	1.01E-07
168	6.84	0.001	0.003	0.095	1.36E-04	1.84E-07

<b>Experiment 1: Sediment-Only + Ash</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	6.93	1.52E-01	0.005	0.092	1.62E-04	1.77E-07
24	6.91	4.86E-02	0.008	0.096	1.62E-04	1.14E-07
48	6.97	6.08E-02	0.009	0.093	1.69E-04	1.19E-07
96	6.92	1.39E-02	0.007	0.092	1.32E-04	6.93E-08
168	7.13	6.13E-02	0.002	0.095	1.64E-04	1.13E-07

**Table S3.** Experiment 2: Microcosm porewater concentrations used as input parameters in the Visual MINTEQ calculations.

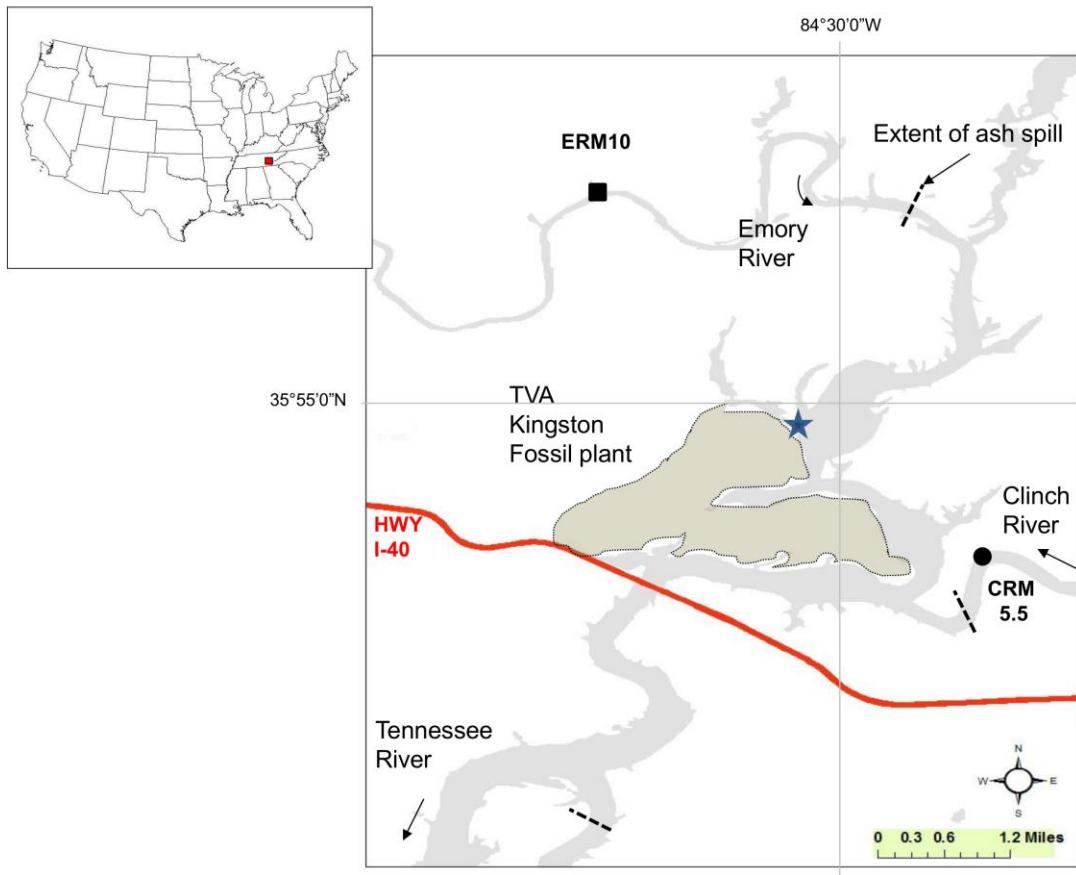
<b>Experiment 2: Sediment Only</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	6.33	0.306	2.37E-03	0.025	1.99E-04	1.29E-07
12	6.17	0.315	4.32E-03	0.022	1.93E-04	6.00E-08
24	6.33	0.306	7.44E-03	0.011	2.25E-04	9.50E-08
48	6.40	0.312	8.10E-03	0.012	2.22E-04	7.76E-08
120	6.42	0.317	4.88E-03	0.011	1.64E-04	7.27E-08

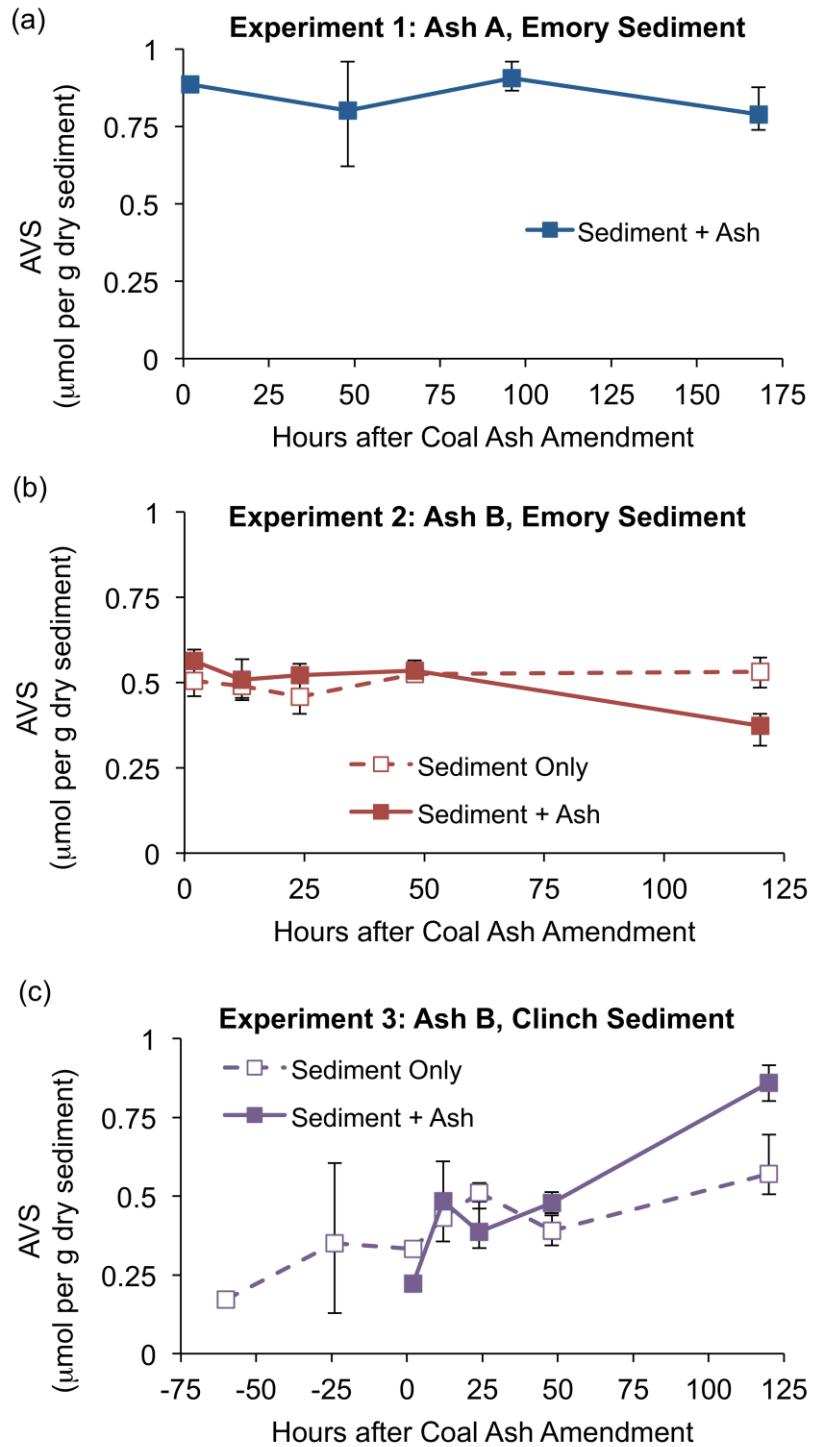
<b>Experiment 2: Sediment Only + Ash</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	6.91	0.305	3.12E-05	0.017	1.67E-04	4.45E-08
12	6.68	0.303	3.12E-05	0.016	1.83E-04	3.69E-08
24	6.91	0.266	2.05E-03	0.014	1.79E-04	6.02E-08
48	6.96	0.260	5.95E-03	0.013	1.81E-04	4.28E-08
120	7.06	0.238	3.72E-03	0.011	1.40E-04	4.21E-08

**Table S4.** Experiment 3: Microcosm porewater concentrations used as input parameters in the Visual MINTEQ calculations.

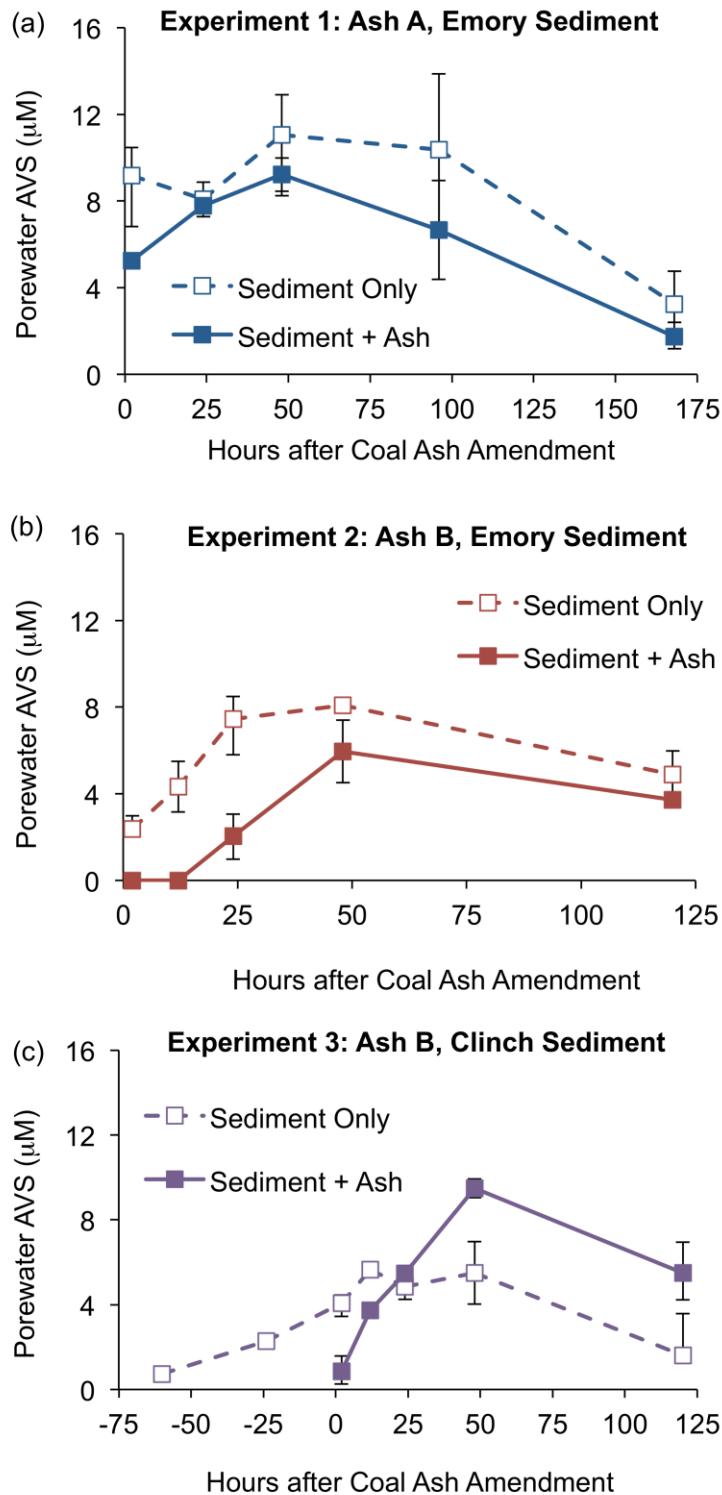
<b>Experiment 3: Sediment Only</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	7.26	0.187	4.08E-03	0.161	1.32E-04	1.29E-07
12	7.11	0.164	5.66E-03	0.167	8.65E-05	2.87E-07
24	7.11	0.176	4.85E-03	0.158	1.28E-04	1.92E-07
48	7.15	0.165	5.50E-03	0.205	1.33E-04	1.73E-07
120	7.15	0.230	1.61E-03	0.238	1.36E-04	3.65E-07
<b>Experiment 3: Sediment Only + Ash</b>						
Timepoint (Hours)	pH	Fe(II) (mM)	HS <sup>-</sup> (mM)	Cl <sup>-</sup> (mM)	Thiols (mM)	Hg(II) <sub>T</sub> (mM)
2	7.62	0.122	8.58E-04	0.278	1.32E-04	3.02E-08
12	7.31	0.018	3.74E-03	0.189	8.13E-05	2.02E-07
24	7.43	0.017	5.48E-03	0.156	1.24E-04	1.01E-07
48	7.48	0.048	9.48E-03	0.184	1.19E-04	1.79E-08
120	7.62	0.038	5.50E-03	0.189	1.29E-04	3.33E-07



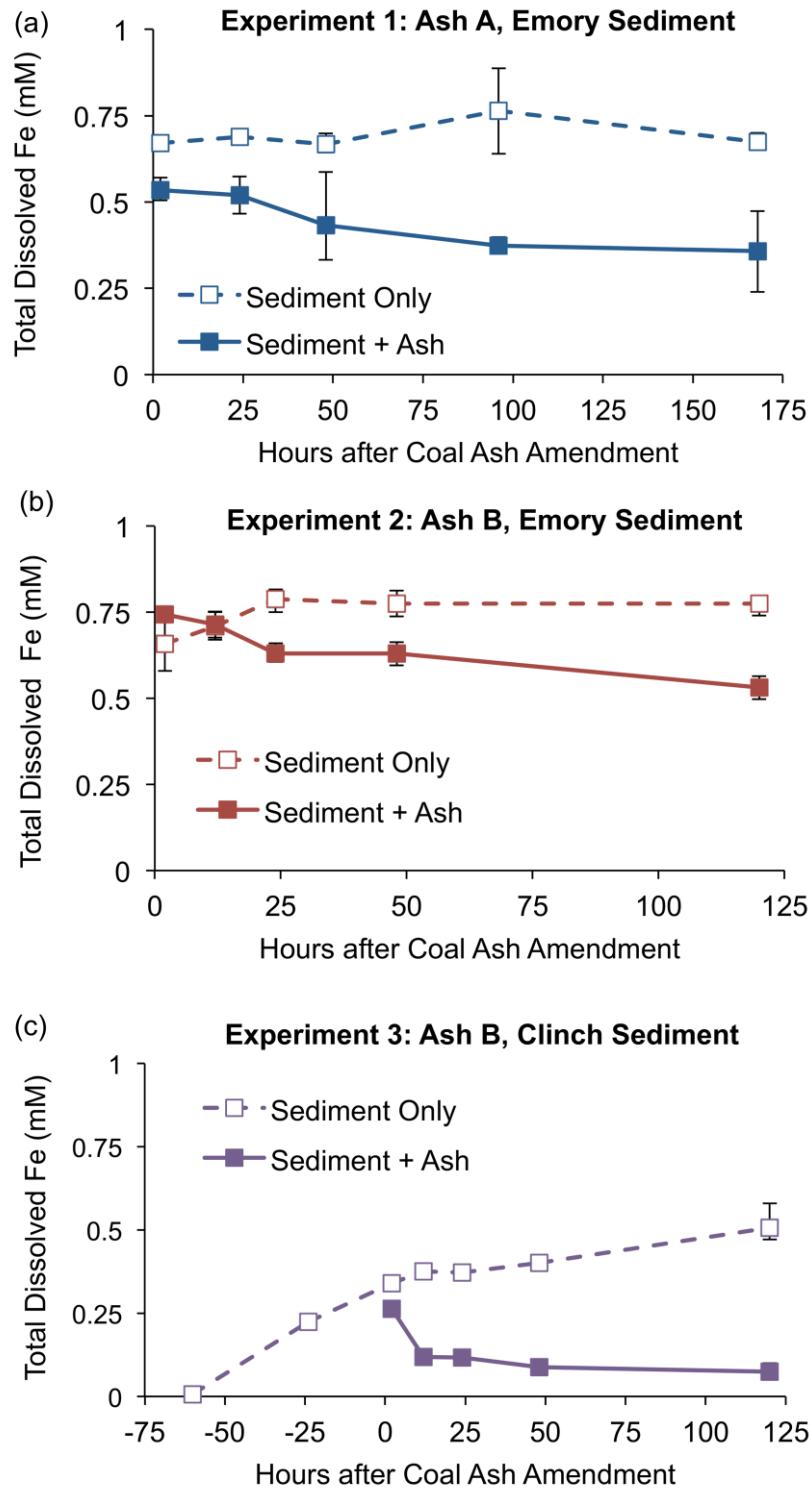
**Figure S1.** Locations of the sampling sites from which sediment and water samples were collected for the microcosm experiments. Emory River Mile Marker 10 (ERM 10) and Clinch River Mile Marker 5.5 (CRM 5.5) are located upstream of the coal ash spill. The star represents the location of the ash release from the impoundment. The dashed lines indicate the extent of the ash spill.



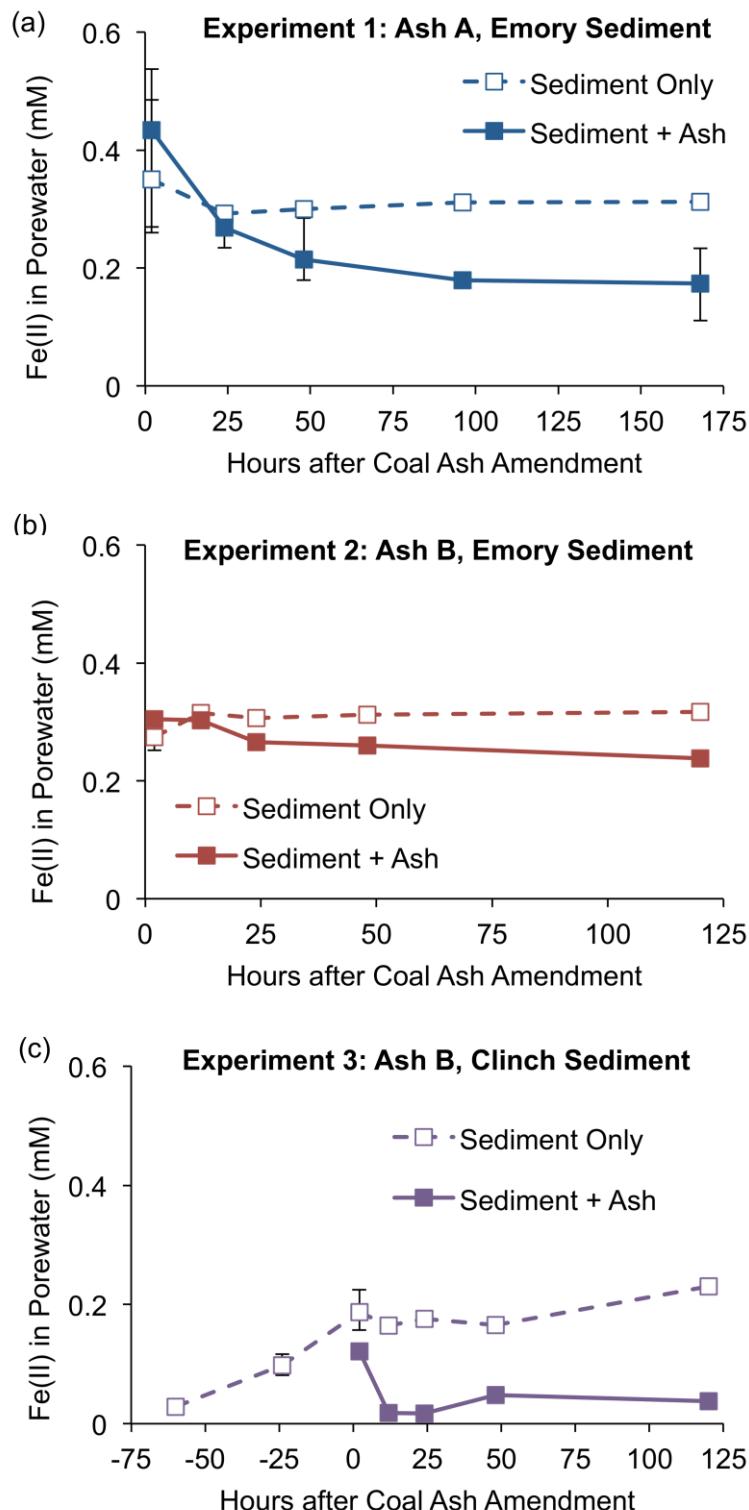
**Figure S2.** Acid volatile sulfide (AVS) contents in aliquots of whole slurries from the sediment-ash microcosms. (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Experiment 3: Clinch Sediment/Ash B. Data points represent replicate microcosms ( $n=2-3$ ). Error bars represent the range of the samples. AVS data is not available for the Sediment Only treatment from Experiment 1.



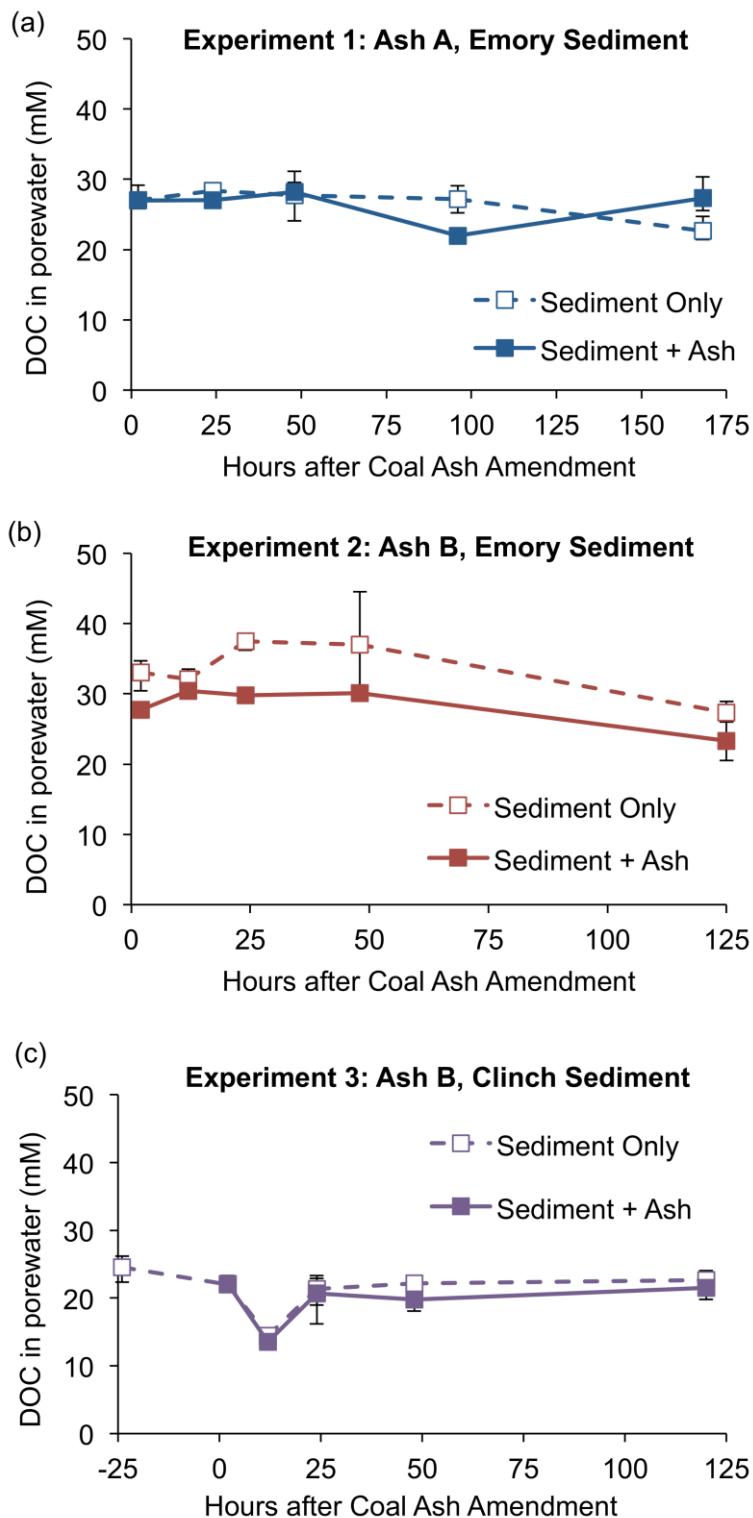
**Figure S3.** Dissolved acid volatile sulfide concentrations in the porewater of sediment-ash microcosms. (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Experiment 3: Clinch Sediment/Ash B. Data points represent the mean of replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.



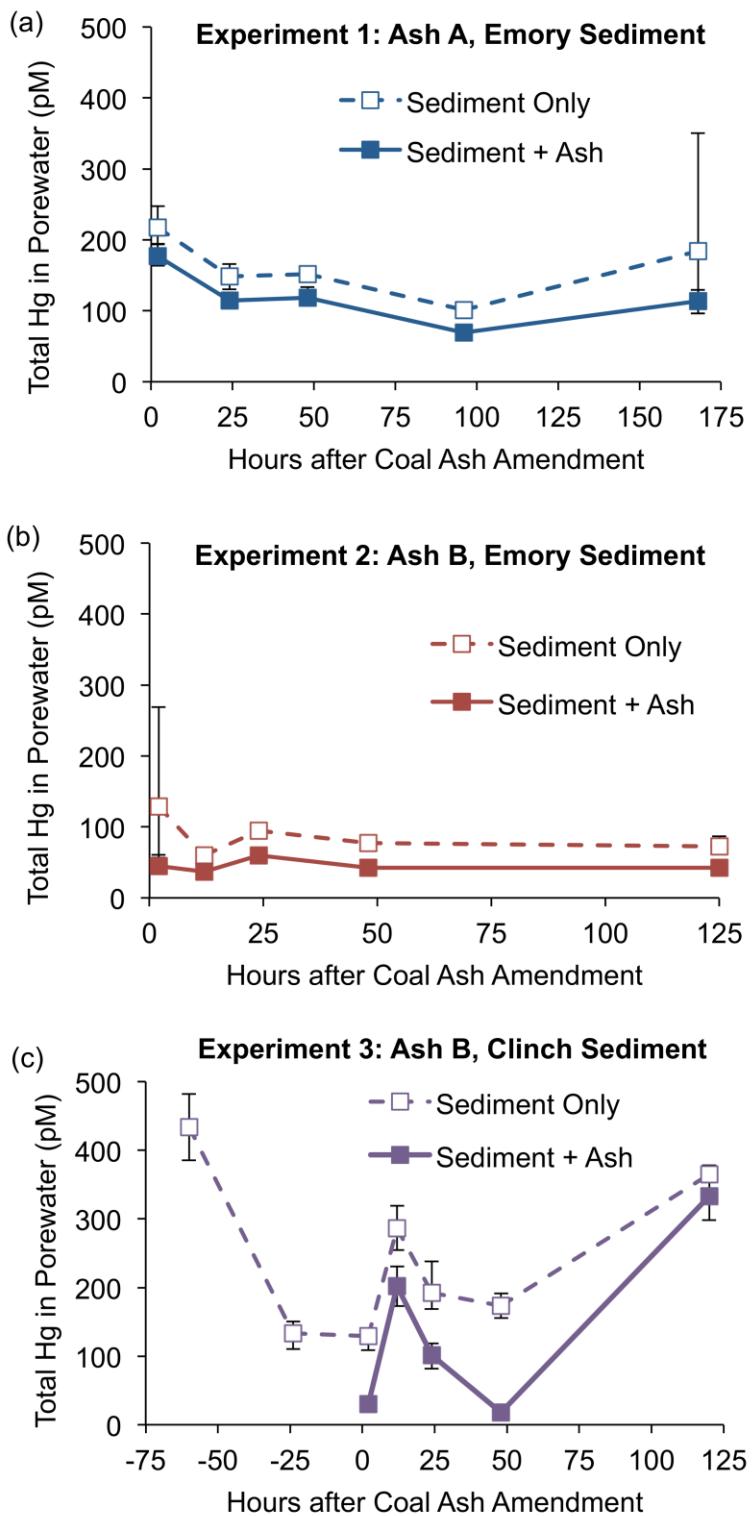
**Figure S4.** Dissolved iron concentrations in the porewater of sediment-ash microcosms. (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Experiment 3: Clinch Sediment/Ash B. Each data point represents replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.



**Figure S5.** Ferrous iron (Fe(II)) concentrations in the porewater of sediment-ash microcosms; (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B (c) Clinch Sediment/Ash B. Data points represent the mean of replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.

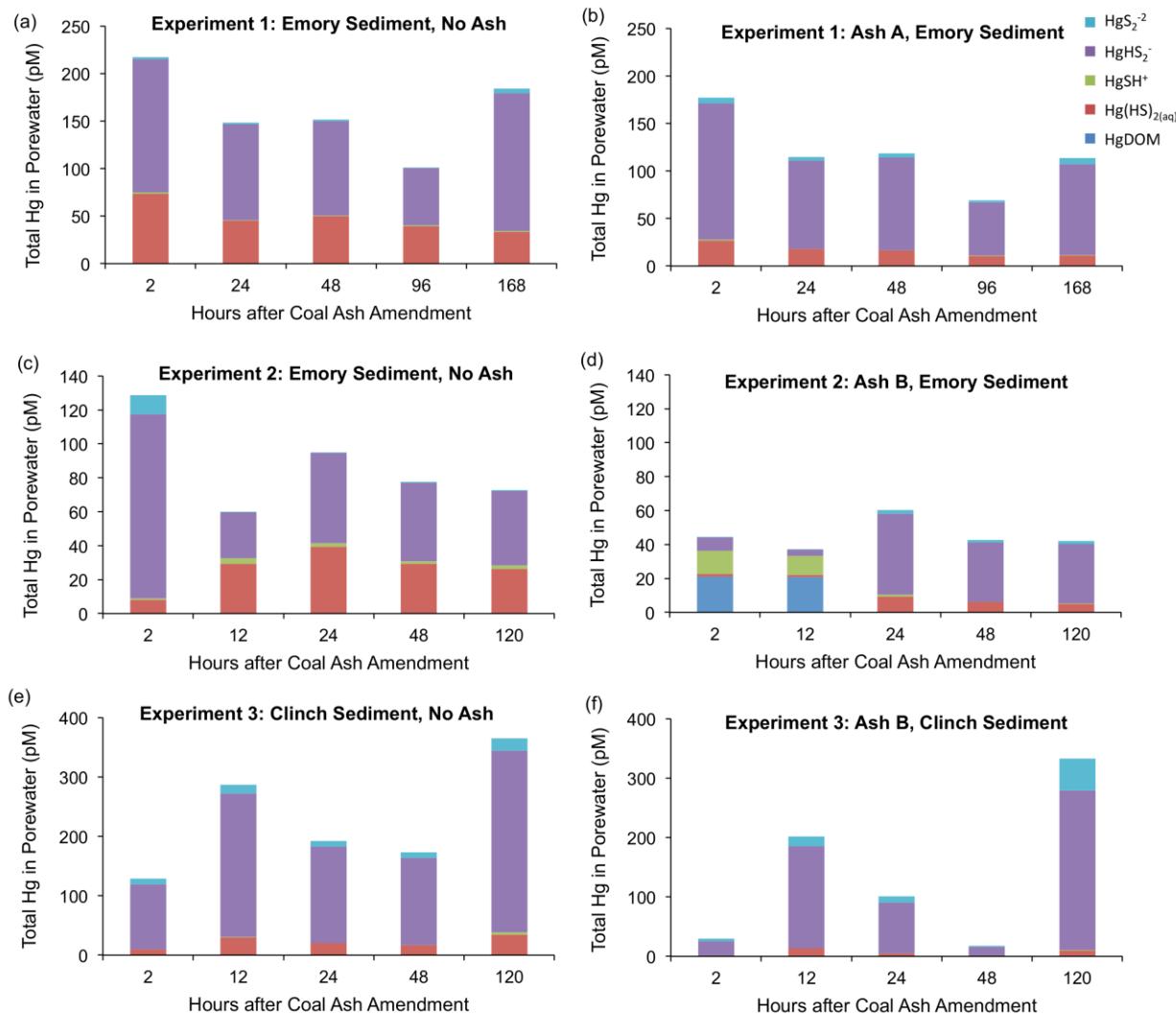


**Figure S6.** Dissolved organic carbon (DOC) concentrations in the porewater of sediment-ash microcosms (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Experiment 3: Clinch Sediment/Ash B. Data points represent replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.

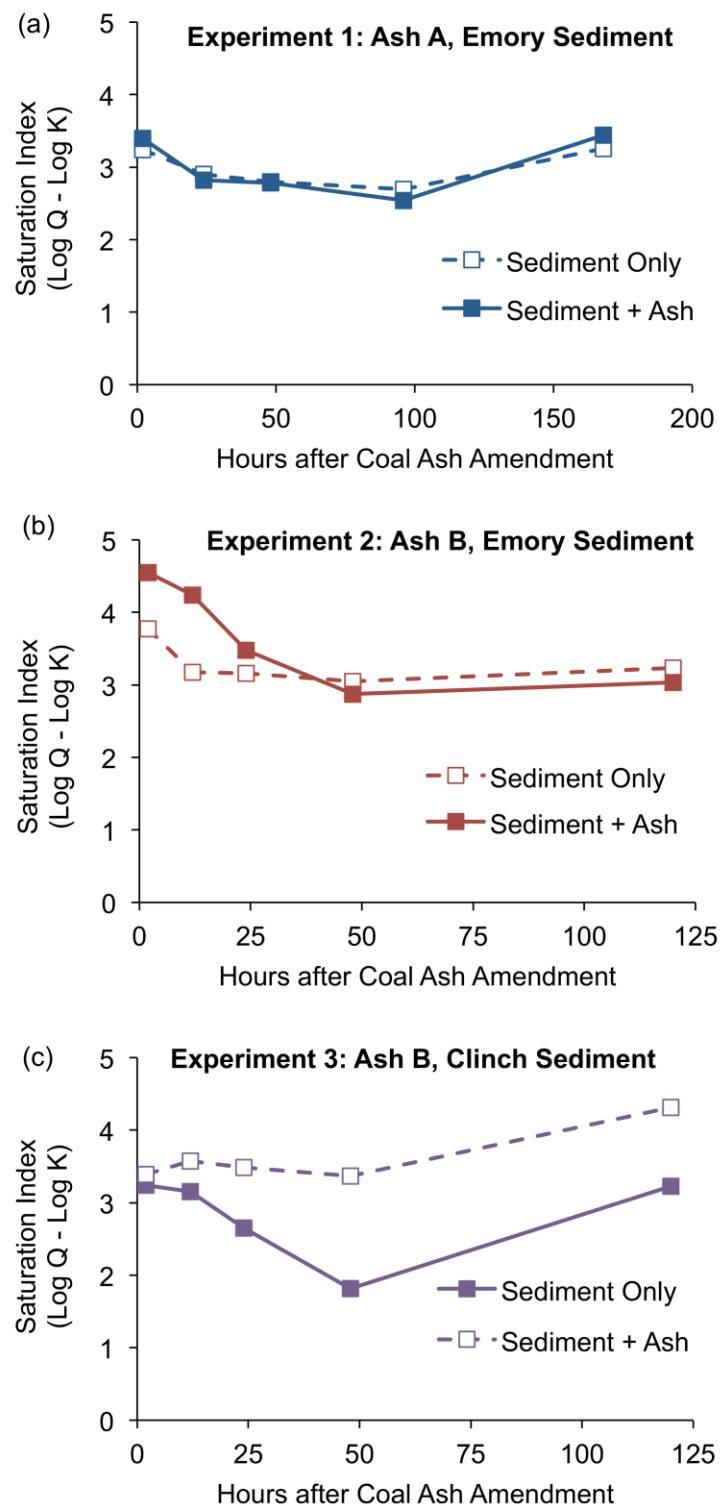


**Figure S7.** Total Hg concentrations in the porewater of sediment-ash microcosms; (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Clinch

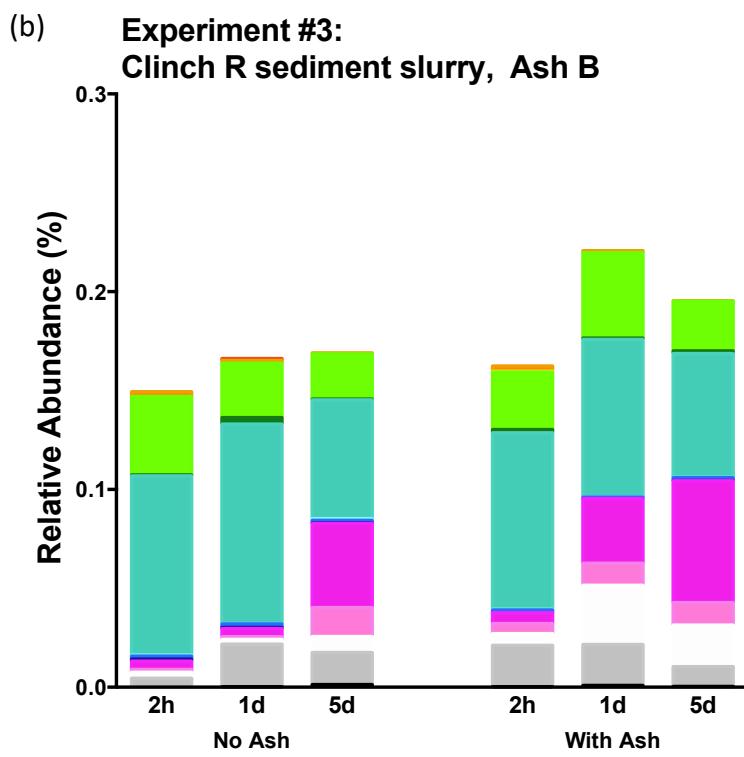
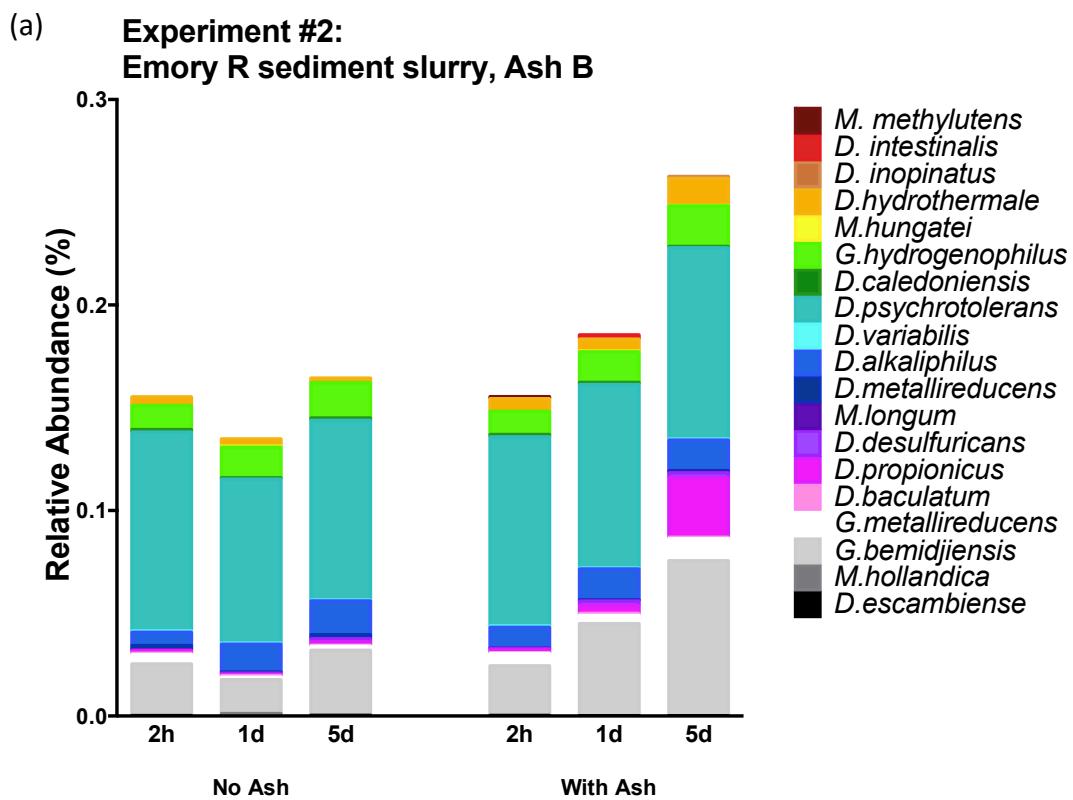
Sediment/Ash B. Data points represent the mean of replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.



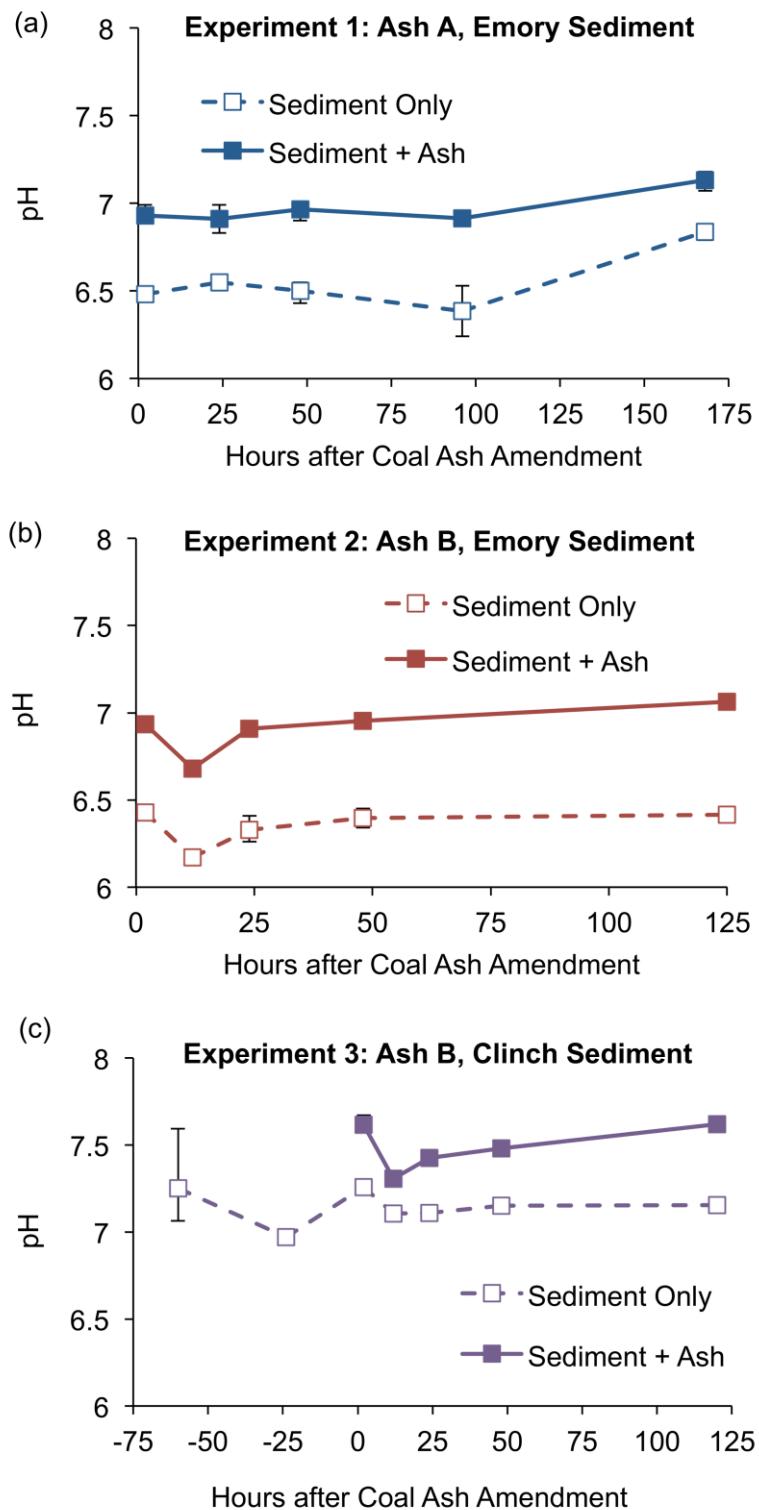
**Figure S8.** Calculated speciation of porewater Hg in sediment-ash microcosms (a) Experiment 1: Emory Sediment-only microcosms; (b) Experiment 1: Emory Sediment/Ash A microcosms (c) Experiment 2: Emory Sediment-only microcosms; (d) Experiment 2: Emory Sediment/Ash B microcosms; (e) Experiment 3: Clinch Sediment-only microcosms; (f) Clinch Sediment/Ash B microcosms.



**Figure S9.** Saturation indices for metacinnabar in sediment-ash microcosm porewater (a) Experiment 1: Emory Sediment/Ash A microcosms; (b) Experiment 2: Emory Sediment/Ash B microcosms; (c) Experiment 3: Clinch Sediment/Ash B microcosms. Positive saturation index values indicate oversaturation with respect to  $\text{HgS}_{(s)}$ .



**Figure S10.** Fraction of the total relative abundance of methylating species in sediment-ash microcosms from (a) Experiment #2 and; (b) Experiment #3.



**Figure S11.** pH values in the sediment-ash microcosms. (a) Experiment 1: Emory Sediment/Ash A; (b) Experiment 2: Emory Sediment/Ash B; (c) Experiment 3: Clinch Sediment/Ash B. Data

points represent replicate microcosms ( $n=2-3$ ). Error bars represent the range of replicate samples.

## References

1. Gustafsson, J. P. *Visual MINTEQ*, 3.1; Stockholm, Sweden, 2014.
2. Hsu-Kim, H.; Kucharzyk, K. H.; Zhang, T.; Deshusses, M. A., Mechanisms Regulating Mercury Bioavailability for Methylating Microorganisms in the Aquatic Environment: A Critical Review. *Environ. Sci. Technol.* **2013**, 47, (6), 2441-2456.
3. Haitzer, M.; Aiken, G. R.; Ryan, J. N., Binding of Mercury(II) to Aquatic Humic Substances: Influence of pH and Source of Humic Substances. *Environ. Sci. Technol.* **2003**, 37, (11), 2436-2441.
4. National Institute of Standards and Technology, NIST Critically Selected Stability Constants of Metal Complexes Database. In 8.0 ed.; Gaithersburg, MD, 2004.
5. Luther, G. W.; Rickard, D. T.; Theberge, S.; Olroyd, A., Determination of metal (bi)sulfide stability constants of Mn<sup>2+</sup>, Fe<sup>2+</sup>, Co<sup>2+</sup>, Ni<sup>2+</sup>, Cu<sup>2+</sup>, and Zn<sup>2+</sup> by voltammetric methods. *Environ. Sci. Technol.* **1996**, 30, (2), 671-679.