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Scalable and continuous nanomaterial integration with transgenic fibers for enhanced photoluminescence

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Reflectance and fluorescence confocal microscopy measurements

In addition to electron microscopy and energy dispersive X-ray, we made extensive use of reflectance confocal microscopy (RCM) to directly visualize AgNPs embedded inside silk fibers. The typical configuration of RCM can be summarized as follows: confocal aperture size = 50 or 80 μ m (i.e., ~ 0.5 airy unit), numerical aperture = 0.4, and scan speed (pixel dwell time) = 10 μ s/pixel. For optically sectioned images, we generated 2D RCM images of AgNP-embedded silk under three different illumination wavelengths of 488, 543, and 633 nm on the same optically sectioned plane. For 3D visualizations, we stacked 64 image slices with a slice thickness of 0.2 μ m along the z-axis (total thickness = 12.6 μ m), covering an area up to ~ 211.7 × 211.7 μ m². For fluorescence (mKate2) confocal images, we used a laser excitation source at 543 nm and a detection bandpass of 600 – 700 nm.

Mechanical property measurements

We explored the integration effect of AgNPs into silk fibers on the mechanical properties. For each silk cocoon, at least 15 single fibers were tested. A universal electromechanical test machine (100P/Q, TestResources) was used with an extension rate of 1 mm min⁻¹ and a gauge length of 10 mm under ambient conditions (25 ± 2 °C and $50 \pm 10\%$ relative humidity). There were no significant variations in the diameter of AgNP-embedded silk fibers compared to the native bare silk fibers before the integration of AgNPs, measured under an optical microscope. As shown in Table S2 and Fig. S9, after AgNP hybridization with silk, the common mechanical properties of individual silk fibers, including the maximum strain, the maximum stress, and the Young's modulus, were not significantly changed. A one-way ANOVA test showed that the three groups of the Young's moduli (i.e., native bare control silk fibers and AgNP-embedded silk fibers at 0.5-mM AgNO₃ and 1.0-mM AgNO₃) were not different statistically with a p-value of 0.3. Thus, the integration of AgNPs into silk did not lead to considerable variations in the mechanical properties of individual silk fibers. We note that the previous studies have reported the maximum strain and stress values of degummed silk fibers in wide ranges of 4 - 26% and 300 - 600 MPa, respectively.¹⁻³ Such large variations for different samples are often caused by different feeding environments, degumming processes, testing parameters, and analysis methods. In our analyses, we only investigated the integration effect of AgNPs into silk fibers on the mechanical properties using degummed silk cocoons under the same conditions.

Mass density of mKate2/Fibroin H-chain fusion recombinant protein

We estimated the mass density of mKate2/Fibroin H-chain fusion recombinant protein in the transgenic silk cocoon as ~ 12.6 %. Using quantitative densitometry of SDS/PAGE, we measured the protein SDS/PAGE spot optical density, which is proportional to the mass of the expressed fusing protein. 25 mg of each cocoon shell was dissolved in 1 mL of 60% (w/v) lithium thiocyanate by vortex-mixing for 10 minutes and was incubated at room temperature for 1 hour followed by centrifugation at 10,000 rpm in a microcentrifuge for five minutes. 5µL samples were subjected to SDS/PAGE [10% (w/v) polyacrylamide slab gel] by dissolving in an equal volume of a sample loading buffer [5% (v/v) glycerol, 1% (w/v) SDS, 0.05% (w/v) bromophenol blue, 0.0625 M Tris/HCl, pH 6.8] with 2% (v/v) 2-mercaptoethanol and were boiled for five minutes. After electrophoresis, the gel was stained with a staining buffer [0.1% (w/v) Coomassie Brilliant Blue R-250, 10% (v/v) acetic acid, 50% (v/v) methanol]. 2 μ L of each sample was diluted five times with 10 mm Tris/HCl (pH 6.8) and then 2 μ L of the diluted sample was dissolved in a sample loading buffer and was subjected to SDS/PAGE. The gel was transferred onto a polyvinylidene difluoride (PVDF) membrane, which was incubated at room temperature for 1 hour with TBST containing 5000-fold diluted tRFP antibody (Evrogen, Russia). The membrane was incubated at room temperature for one hour

in TBST containing 30,000-fold diluted horseradish peroxidase-labeled rabbit IgG secondary antibody. The immunoreactive bands were visualized with ECL Plus Western Blotting Detection Reagents (Amersham Biosciences). The signal intensity of the bands of Western blotting were calculated with Quantity One 1D Analysis Software (Bio-Rad Laboratories), in comparison with the intensity of rKFP-Red as a control.

Mole ratio of AgNO ₃ (@ 5 minutes)	0.3 mM	0.5 mM	0.7 mM	1.0 mM
(%) ^{a)}	15.6	32.5	67.2	96.4
Treatment time (@ 0.5 mM)	1 minute	5 minutes	10 minutes	20 minutes
(%)	15.9	32.5	84.5	95

Table S1. Absorption changes in absorption spectra (400 - 800 nm) for AgNP-embedded silk relative to native bare control silk under different conditions.

^{a)} Percentage changes with respect to bare control silk.

Sample ^{a)}	Average diameter of fibers	Maximum strain	Maximum stress	Young's modulus
	(µm)	(%)	(MPa)	(GPa)
Native bare silk (Control)	$9.8\pm0.5^{\text{b})}$	12.7 ± 3.9	383.6 ± 57.3	9.2 ± 0.8
AgNP-embedded silk (@ 0.5 mM and 5 minutes)	9.6 ± 0.6	12.9 ± 4.0	372.0 ± 48.8	8.7 ± 0.6
AgNP-embedded silk (@ 1.0 mM and 5 minutes)	9.7 ± 0.6	13.1 ± 4.2	360.2 ± 54.8	9.0 ± 0.8

Table S2. Mechanical properties of bare native (control) silk fibers and AgNP-embedded silk fibers with different mole ratios of AgNO₃.

^{a)} For each silk cocoon, at least 15 single fibers were tested. ^{b)} Mean \pm standard deviation. The Young's moduli are calculated from the first linear regime of the strain-stress curve before the first bend in Fig. S9.



Fig. S1 (a) Photograph of the rearing environment for transgenic silkworms to produce mKate2 expressing silk cocoons. (b) Molar absorption and fluorescence spectra of far-red fluorescent mKate2 protein. (c) Transformation vector for germline transgenesis. (i) pBac-3xP3-EGFP. (ii) pGEMT-pFibH-NTR. (iii) pGEMT-CTR. (iv) pFibHNC-null. (v) pFibHNC-mKate2. (vi) p3xP3-EGFP-pFibH-mKate2. The nucleotide sequences of pFibH-NTR and CTD were derived from Genebank Accession No. AF226688. pFibH: fibroin heavy chain promoter domain (1124 bp), NTR1: N-terminal region 1 (142 bp), intron: first intron (871 bp), NTR2: N-terminal region 2 (417 bp), CTR: C-terminal region (179 bp), PolyA: poly(A) signal region (301 bp), EGFP: enhanced green fluorescent protein gene, mKate2: monomeric far-red fluorescent protein, ITR: inverted repeat sequences of *piggyBac* arms, 3xP3: 3xP3 promoter, and SV40: SV40 polyadenylation signal sequence. The restriction enzyme sites for the construction of recombinant vectors are indicated with the arrows.



Fig. S2 Simplified schematic diagram of synthesizing AgNP-embedded silk via facile and green chemistry.



Fig. S3 Customized mesoscopic (between microscopic and macroscopic) imaging setup. This system provides a matrix of intensity as a function of x and y at each wavelength λ with a spectral resolution of 4 nm in a range of 400 – 800 nm. The imaging area of our mesoscopic imaging setup is ~ 15 mm × 15 mm with a pixel size of 50 µm × 50 µm.



Fig. S4 Calculated scattering cross section of an AgNP inside silk with a diameter of 59 nm. The plasmon resonance occurs $\lambda = 488$ nm where the scattering cross section of AgNP is 8.1 times larger than the geometrical cross section. This simple example supports the use of RCM for directly visualizing AgNPs.



Fig. S5 EDX spectrum and SEM image on the surface of AgNP-embedded silk fibers with a low density of AgNPs (i.e., mole ratio of $AgNO_3 = 0.5$ mM and treatment time = 5 minutes), also confirming the presence of AgNPs on the surface, in addition to internalized AgNPs (Fig. 1).



Fig. S6 SEM images of (a&b) native bare silk fibers (control) and (c) AgNP-embedded silk fiber synthesized under a relatively high-density condition (i.e., mole ratio of $AgNO_3 = 1.0$ mM and treatment time = 5 minutes). (d&e) Comparisons of EDX elements of native bare silk (control) and AgNP-embedded silk inside fibers and on the fiber surface. The native bare silk fibers do not show any element of Ag.



Fig. S7 TEM images of (a) AgNP-embedded fibers of silk cocoons, (b) isolated AgNPs synthesized in aqueous solutions (left) and TEM-derived size distribution of AgNPs (right), and (c) AgNP-embedded fibers of silk fabrics. All of the samples are prepared for low-density AgNPs at 0.5-mM AgNO₃ for five minutes. The sizes of sphere-like AgNPs synthesized in aqueous solutions are in a range of 24 - 44 nm, with a mean size of ~ 35 nm. The randomly distributed AgNPs are present inside fibers of both AgNP-embedded silk cocoons and fabrics.



Fig. S8 (a) Photograph of aqueous AgNP solutions under different mole ratios of AgNO₃. (b) Measured extinction spectra of the corresponding AgNP solutions. Broad spectra indicates larger sizes of AgNPs with higher densities without the nanostructural frame of native silk.



Fig. S9 Representative strain–stress curves of bare native (control) silk fibers and AgNPembedded silk fibers with different mole ratios of AgNO₃. The error bars show the standard deviations of elongation at break (horizontal axis) and fracture strength (vertical axis).



Fig. S10 Tandem mass spectra of (a) 1827.88 m/z, (b) 1685.76 m/z, (c) 945.43 m/z, and (d) 843.43 m/z of mKate2 tryptic fragments and derived amino acid sequences.



Fig. S11 (a&b) Reflectance images under two different illumination bands as illustrated on top. Changes in the scattering strength among different specimens are almost negligible, due to the low density of AgNPs. The specimens are placed on top of a white reflectance standard with a reflectivity of 99% (Labsphere SRS-99-010).



Fig. S12 (a) Emission spectra of AgNP-embedded mKate2 silk cocoons synthesized with different AgNO₃ mole ratios. (b) Photoluminescent intensity of the corresponding samples averaged by a bandwidth of $\lambda = 600 - 700$ nm. The dotted line in (b) is the average emission intensity of the control mKate2 silk cocoon. The synthesis condition of 0.5-mM AgNO₃ and 5-minute treatment time provides an optimal hybridization for plasmon-enhanced photoluminescence of mKate2 silk.

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