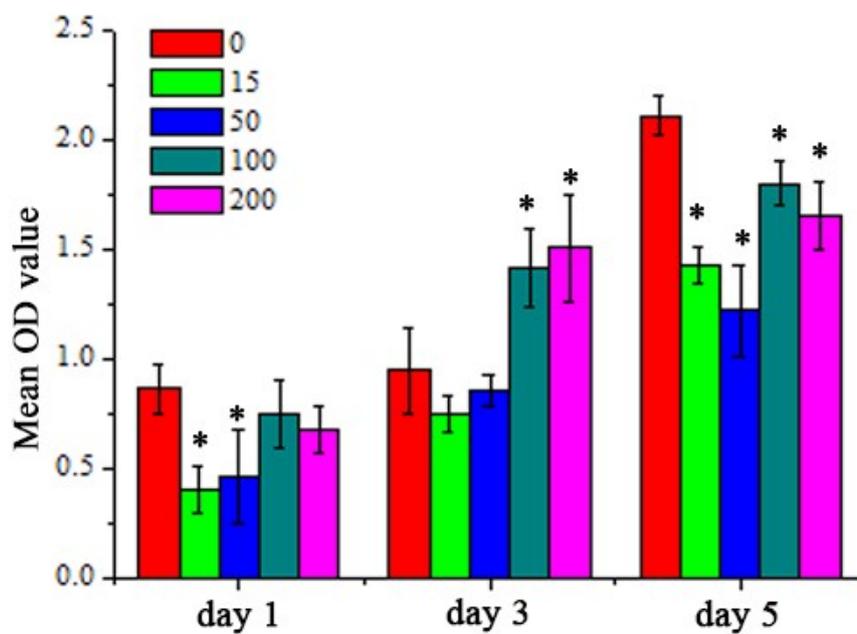
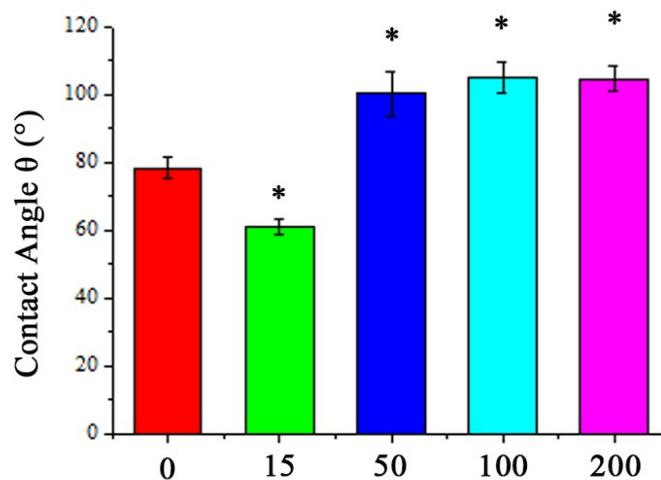


### Supplementary Table 1 RNA Primers applied in this study

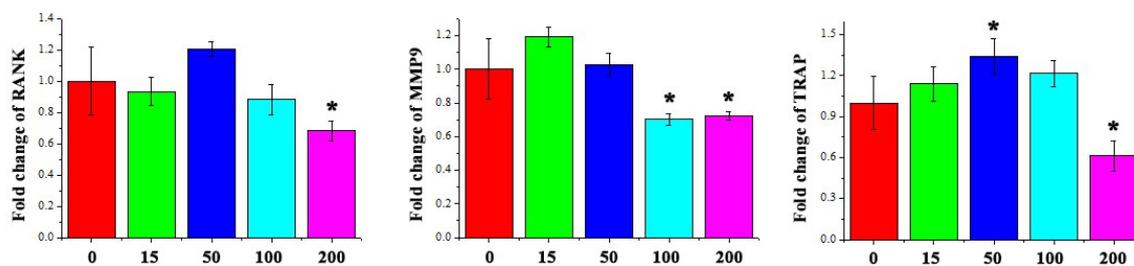
Genes	Primer sequences
TNF $\alpha$	<b>Forward:</b> 5'-CTGAACTTCGGGGTGATCGG-3' <b>Reverse:</b> 5'-GGCTTGCTCACTCGAATTTTGAGA-3'
IL-18	<b>Forward:</b> 5'-TGGCCGACTTCACTGTACAAC-3' <b>Reverse:</b> 5'-TGGGGTTCCTGGCACTTTG-3'
IL1 $\beta$	<b>Forward:</b> 5'-TGGAGAGTGTGGATCCCAAG-3' <b>Reverse:</b> 5'-GGTGCTGATGTACCAGTTGG-3'
OSM	<b>Forward:</b> 5'-ACGGTCCACTACAACACCAG-3' <b>Reverse:</b> 5'-CCATCGTCCCATTCCCTGAAG-3'
IL6	<b>Forward:</b> 5'-ATAGTCCTTCTACCCAATTTCC-3' <b>Reverse:</b> 5'-GATGAATTGGATGGTCTTGGTCC-3'
CD86	<b>Forward:</b> 5'-CTGCTCATCATTGTATGTCAC-3' <b>Reverse:</b> 5'-ACTGCCTTCACTCTGCATTTG-3'
CD11c	<b>Forward:</b> 5'-ACTTCACGGCCTCTCTCC-3' <b>Reverse:</b> 5'-CACCAGGGTCTTCAAGTCTG-3'
iNOS	<b>Forward:</b> 5'-CAGAAGTGCAAAGTCTCAGACAT-3' <b>Reverse:</b> 5'-GTCATCTTGATTGTTGGGCT-3'
CCR7	<b>Forward:</b> 5'-ATGACGTCACCTACAGCCTG-3' <b>Reverse:</b> 5'-CAGCCCAAGTCCTTGAAGAG-3'
CD206	<b>Forward:</b> 5'-AGACGAAATCCCTGCTACTG-3' <b>Reverse:</b> 5'-CACCCATTGGAAGGCATTC-3'
BMP2	<b>Forward:</b> 5'-GCTCCACAAACGAGAAAAGC-3' <b>Reverse:</b> 5'-AGCAAGGGGAAAAGGACACT-3'
BMP6	<b>Forward:</b> 5'-TGGCAGGACTGGATCATTGC-3' <b>Reverse:</b> 5'-ACCAAGGTCTGTACAATGGCG-3'
WNT10b	<b>Forward:</b> 5'-CCAGGTGGTAACGGAAAACC-3' <b>Reverse:</b> 5'-TGCCCTCCAACAGGTCTTG-3'
TGF $\beta$ 1	<b>Forward:</b> 5'-CAGTACAGCAAGTCCTTGC-3' <b>Reverse:</b> 5'-ACGTAGTAGACGATGGGCAG-3'
VEGF $\alpha$	<b>Forward:</b> 5'-GTCCCATGAAGTGATCAAGTTC-3' <b>Reverse:</b> 5'-TCTGCATGGTGTGTTGCTCTCTG-3'
TRAP	<b>Forward:</b> 5'-CACTCCCACCCTGAGATTTGT-3' <b>Reverse:</b> 5'-CATCGTCTGCACGGTTCTG-3'
RANK	<b>Forward:</b> 5'-GCAGCTCAACAAGGATACGG-3' <b>Reverse:</b> 5'-GGTGCAGTTGGTCCAAGTT-3'
MMP9	<b>Forward:</b> 5'-GGGCGTGTCTGGAGATTCG-3' <b>Reverse:</b> 5'-CACCTGGTTCACCTCATGGTC-3'
SMAD1	<b>Forward:</b> 5'-GAGATCAATAGAGGAGATGTTCC-3' <b>Reverse:</b> 5'-TCGGTCTTATTGTTGGAAG-3'
SMAD4	<b>Forward:</b> 5'-TACCACCATAACAGCACTAC-3' <b>Reverse:</b> 5'-GAACACCAATATTCAGGAGC-3'
SMAD5	<b>Forward:</b> 5'-GTACTATGAACTGAACAACGG-3' <b>Reverse:</b> 5'-TATAGATGGACACCTTTCCC-3'
BMPR2	<b>Forward:</b> 5'-CATTTGAGGATATGCAGGTTCC-3' <b>Reverse:</b> 5'-CTGCCAGGCTATTTTCTTTC-3'
BMPR1a	<b>Forward:</b> 5'-GACACGTGCGAATTGGACAATG-3' <b>Reverse:</b> 5'-CGTCTGATTTTATACCAGTAC-3'
BMPR1b	<b>Forward:</b> 5'-GAAGCTTATGACAGAGTGCTG-3' <b>Reverse:</b> 5'-CTGACGTCAGAGTTTAATGTC-3'
18S	<b>Forward:</b> 5'-CGGAACTGAGGCCATGATTAAG-3' <b>Reverse:</b> 5'-GTATCTGATCGTCTTCGAACCTCC-3'



**Supplementary Figure 1.** Proliferation rates of macrophages grown on different alumina surfaces at day 1, 3 and 5 of culture, determined using the MTT assay; \*Significant difference ( $P < 0.05$ ) compared to the 0 group.



**Supplementary Figure 2.** Contact angle  $\theta$  (°) of different nanoporous structures. The nanoporous structures significantly affected hydrophilicity. 15nm nanoporous surface decreased contact angle while the other surfaces increased the contact angle. \*Significant difference ( $P < 0.05$ ) compared to the 0 group.



**Supplementary Figure 3.** Osteoclastic activities of macrophages on grown on different nanoporous surfaces. The expression levels of osteoclastic activity marker genes (RANK, MMP9, and TRAP) were determined using RT-qPCR. \*Significant difference ( $P < 0.05$ ) compared to the 0 group.