

# First Total Synthesis of (+)-Broussonetine W: Glycosidase Inhibition of Natural Product & Analogs

Ying-Ying Song,<sup>a,e</sup> Kyoko Kinami,<sup>b</sup> Atsushi Kato,<sup>\*,b</sup> Yue-Mei Jia,<sup>a</sup> Yi-Xian Li,<sup>a</sup> George W. J. Fleet<sup>c,d</sup> and Chu-Yi Yu<sup>\*,a,d</sup>

<sup>a</sup> Beijing National Laboratory of Molecular Science (BNLMS), CAS Key Laboratory of Molecular Recognition and Function, Institute of Chemistry, Chinese Academy of Sciences, Beijing 100190, China

<sup>b</sup> Department of Hospital Pharmacy, University of Toyama, 2630 Sugitani, Toyama 930-0194, Japan

<sup>c</sup> Chemistry Research Laboratory, Department of Chemistry, University of Oxford, Mansfield Road, Oxford OX1 3TA, U.K.

<sup>d</sup> National Engineering Research Center for Carbohydrate Synthesis, Jiangxi Normal University, Nanchang, 330022, China

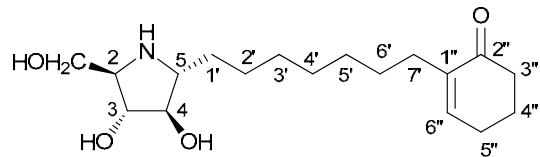
<sup>e</sup> University of the Chinese Academy of Sciences, Beijing 100049, China

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# 1. Comparison of $^{13}\text{C}$ NMR of synthetic broussonetine W (4) with natural product.

**Table 1.** Comparison of the synthetic broussonetine W (4) with natural product



		W (lit.) <sup>a</sup>	W (4) <sup>b</sup>
Pyrrolidine moiety	CH <sub>2</sub> OH	63.45	-0.46
	2	64.97	0.24
	3	80.07	-0.47
	4	84.06	-0.53
	5	62.74	0.11
Octyl moiety	1'	35.43	-0.63
	2'	27.23	0.00
	3'	30.12	0.02
	4', 5' }	29.74, 29.70	0.08, 0.12
	6'	28.98	0.11
	7'	29.96	0.13
	1''	139.79	0.12
Cyclohexenone moiety	2''	198.81	0.00
	3''	38.81	0.13
	4''	23.43	0.14
	5''	26.09	0.13
	6''	145.09	0.06

a)  $\delta$  in pyridine-d<sub>5</sub> at 125 MHz. b)  $\delta$  in pyridine-d<sub>5</sub> at 126 MHz.

## 2. Table for all the new compounds.

Experimental procedures and characterization data for all the new compounds see the general procedures in essay for the details.

**Table 1.** New Compounds in the total synthesis of (**4**, *ent*-**4**, *3-epi*-**4**)

	Synthesis of <b>4</b>	Synthesis of <i>ent</i> - <b>4</b>	Synthesis of <i>3-epi</i> - <b>4</b>
<b>Cyclic Nitrones:</b> (Starting Materials)	 <b>D-arabino-nitrone (10)</b>	 <b>L-arabino-nitrone (ent-10)</b>	 <b>D-lyxo-nitrone (3-epi-10)</b>
<b>Step 1-3:</b> Grignard addition Reduction Protection of amino group	 <b>17</b> 87% yield in 3 steps starting from <b>10</b>	 <b>ent-17</b> 98% yield in 3 steps starting from <i>ent</i> - <b>10</b>	 <b>3-epi-17</b> 66% yield in 3 steps starting from <i>3-epi</i> - <b>10</b>
<b>Step 4:</b> Hydroboration-Oxidation	 <b>18</b> 73% yield derived from olefin <b>17</b>	 <b>ent-18</b> 93% yield derived from olefin <i>ent</i> - <b>17</b>	 <b>3-epi-18</b> 100% yield derived from olefin <i>3-epi</i> - <b>17</b>
<b>Step 5:</b> Appel reaction	 <b>15</b> 97% yield derived from alcohol <b>18</b>	 <b>ent-15</b> 85% yield derived from alcohol <i>ent</i> - <b>18</b>	 <b>3-epi-15</b> 88% yield derived from alcohol <i>3-epi</i> - <b>18</b>
<b>Step 6:</b> $\alpha$ -alkylation of imine ( <i>N</i> -cyclohexyldienecyclohexanamine <b>14</b> )	 <b>19</b> 73% yield derived from bromide <b>15</b>	 <b>ent-19</b> 71% yield derived from bromide <i>ent</i> - <b>15</b>	 <b>3-epi-19</b> 60% yield derived from alcohol <i>3-epi</i> - <b>15</b>
<b>Step 7:</b> 10% Pd/C-catalyzed hydrogenolysis reaction	 <b>20</b> 96% yield derived from ketone <b>19</b>	 <b>ent-20</b> 84% yield derived from ketone <i>ent</i> - <b>19</b>	 <b>3-epi-20</b> 75% yield derived from ketone <i>3-epi</i> - <b>19</b>

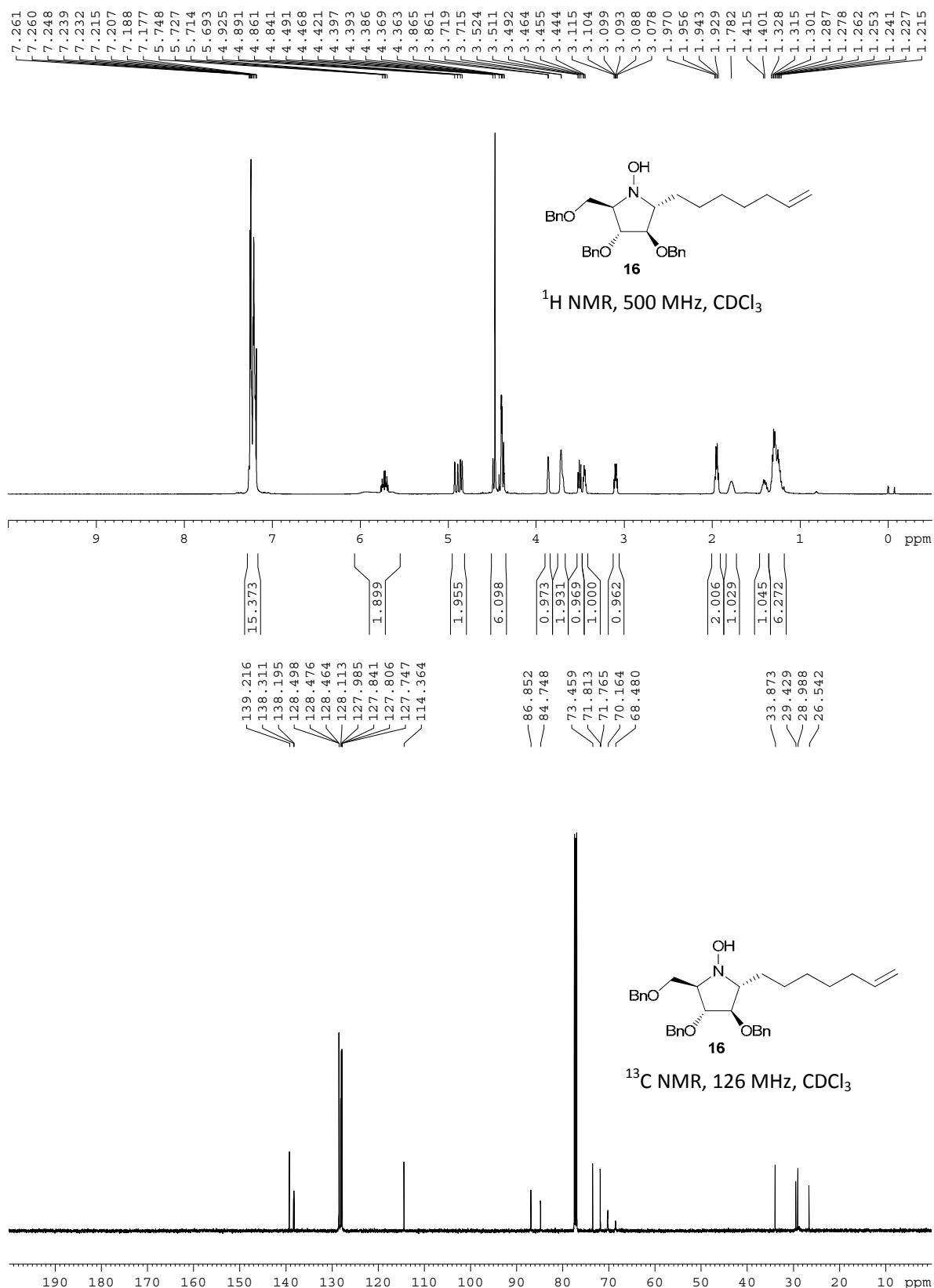
<b>Step 8:</b> Protection of the amino and hydroxyl groups with Ac <sub>2</sub> O			
	<b>95%</b> yield derived from ketone <b>20</b>	<b>94%</b> yield derived from ketone <b>ent-20</b>	<b>76%</b> yield derived from ketone <b>3-epi-20</b>
<b>Step 9-10:</b> Bromination-elimination of ketone			
	<b>83%</b> yield in 2 steps derived from ketone <b>21</b>	<b>85%</b> yield in 2 steps derived from ketone <b>ent-21</b>	<b>78%</b> yield in 2 steps derived from ketone <b>3-epi-21</b>
<b>Step 11:</b> Removal of acetyl groups			
	<b>92%</b> yield derived from compound <b>22</b>	<b>78%</b> yield derived from compound <b>ent-22</b>	<b>89%</b> yield derived from compound <b>3-epi-22</b>
<b>Total Yield In 10 Steps:</b>	<b>31%</b>	<b>29%</b>	<b>14%</b>

**Table 2.** New Compounds in the total synthesis of (*ent*-3-*epi*-4, *ent*-4a, *ent*-4b)

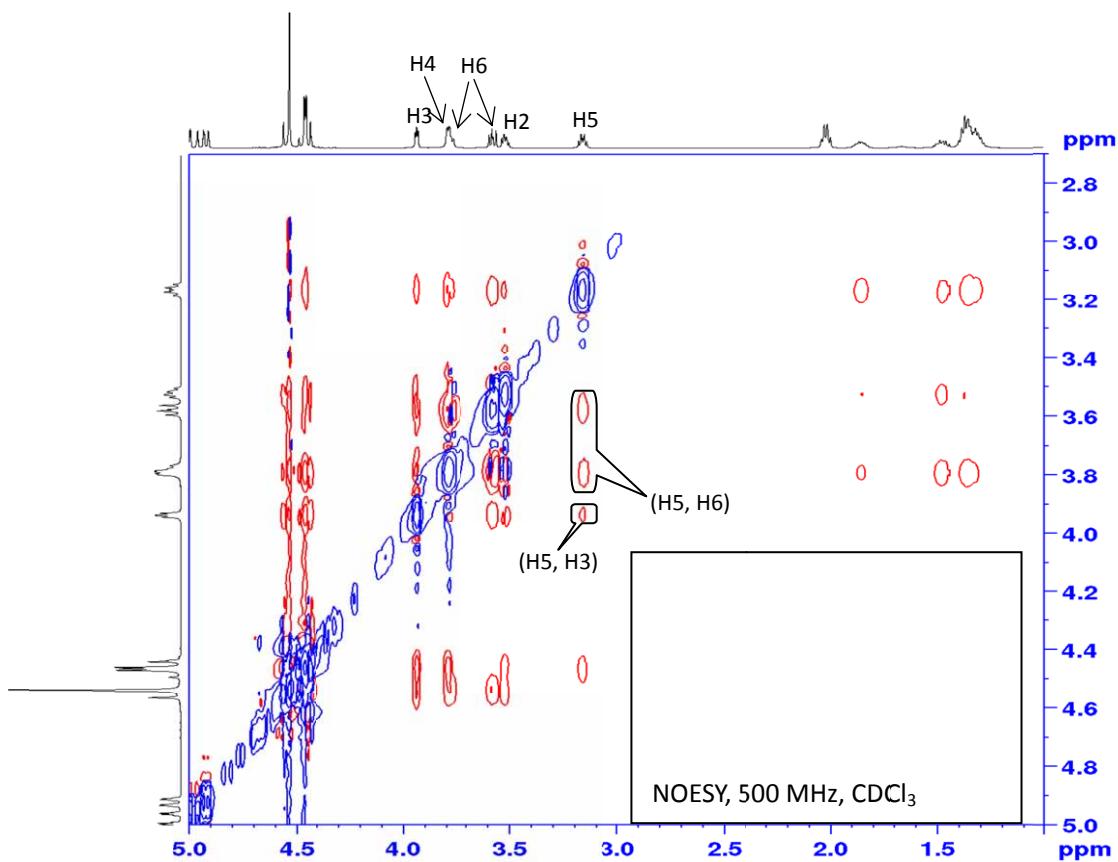
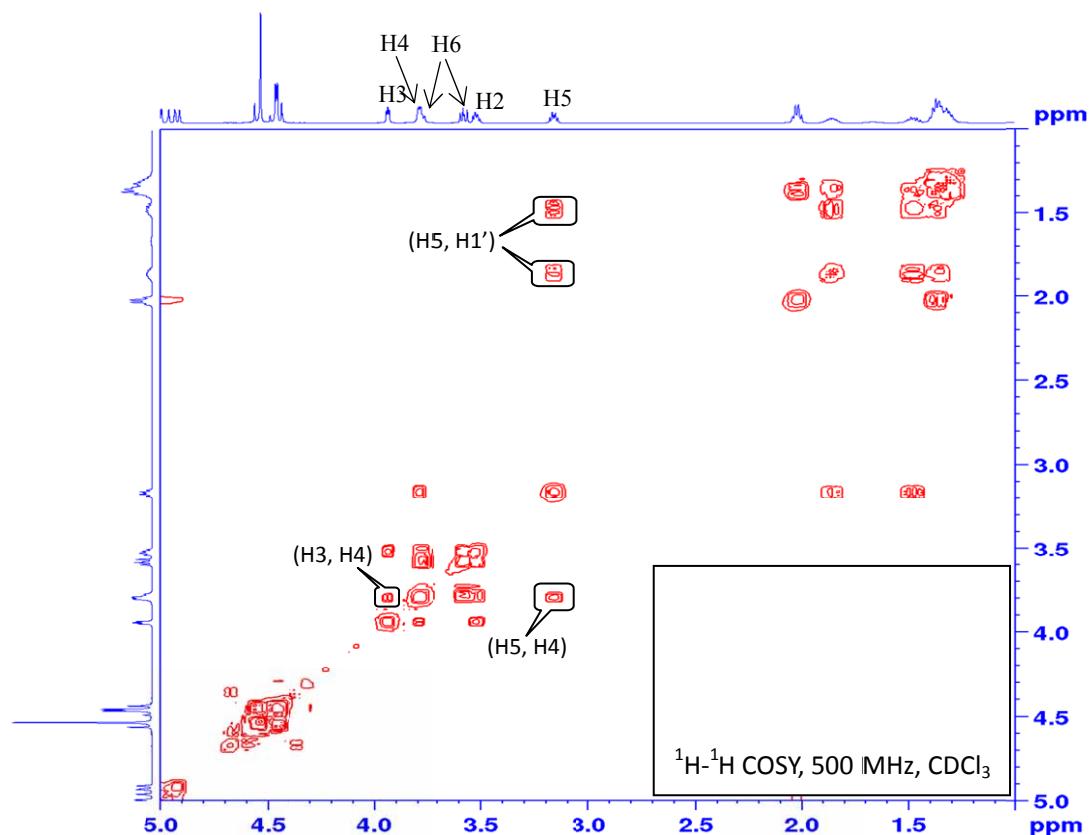
	Synthesis of <i>ent</i> -3- <i>epi</i> -4	Synthesis of <i>ent</i> -4a	Synthesis of <i>ent</i> -4b
<b>Cyclic Nitrones:</b> (Starting Materials)			
<b>Step1-3:</b> Grignard addition Reduction Protection of amino group			
	<b>76%</b> yield in 3 steps starting from <b>ent-3-epi-10</b>	<b>84%</b> yield in 3 steps starting from <b>ent-10</b>	<b>93%</b> yield in 3 steps starting from <b>ent-10</b>
<b>Step 4:</b> Hydroboration-Oxidation			
	<b>73%</b> yield derived from olefin <b>ent-3-epi-17</b>	<b>91%</b> yield derived from olefin <b>ent-17a</b>	<b>91%</b> yield derived from olefin <b>ent-17b</b>

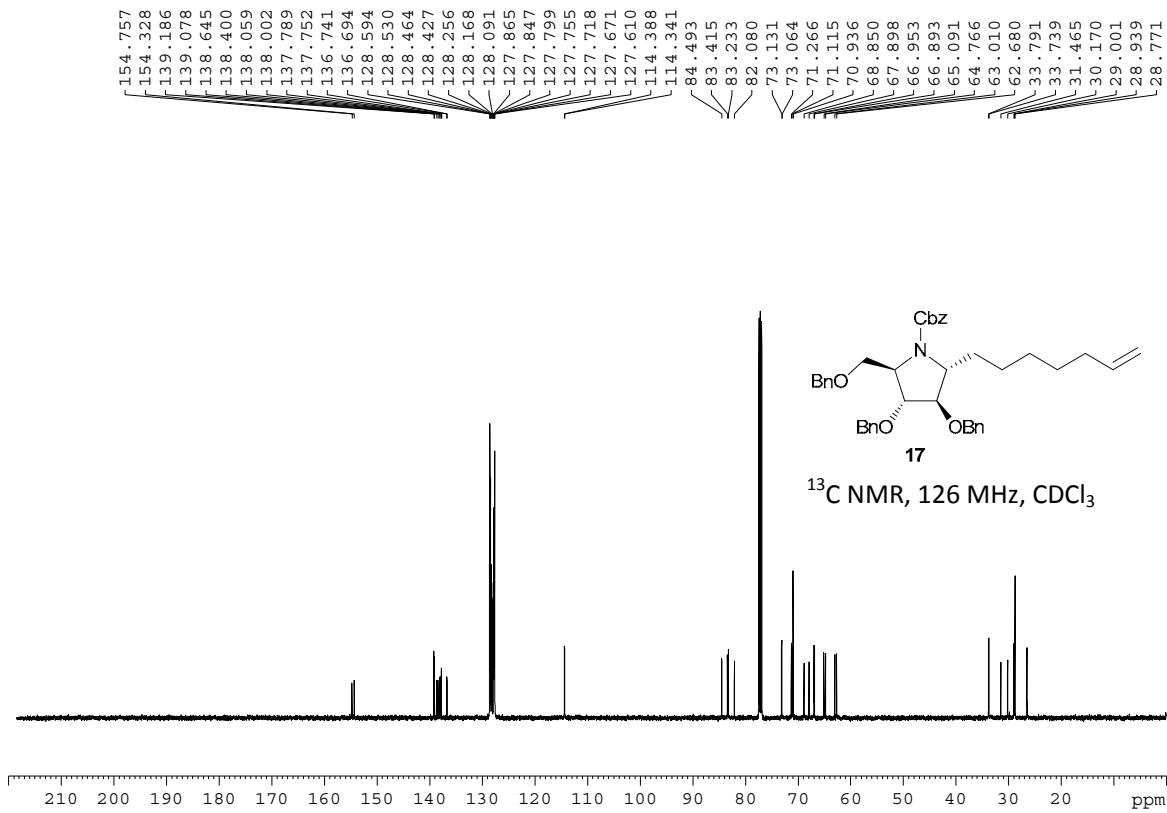
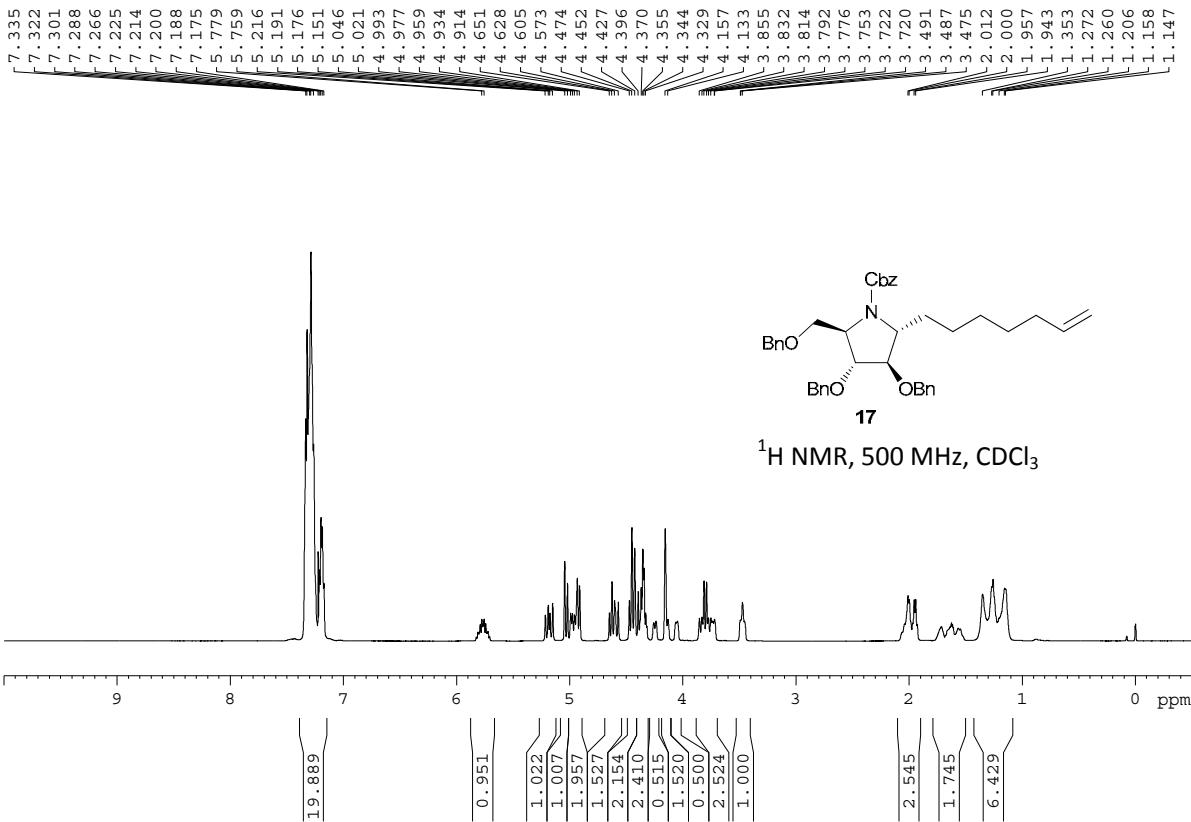
<b>Step 5:</b> Appel reaction			
	<b>96%</b> yield derived from alcohol <i>ent-3-epi-18</i>	<b>92%</b> yield derived from alcohol <i>ent-18a</i>	<b>93%</b> yield derived from alcohol <i>ent-18b</i>
<b>Step 6:</b> $\alpha$ -alkylation of imine ( <i>N</i> -cyclohexylidenecyclohexanamine <b>14</b> )			
	<b>71%</b> yield derived from bromide <i>ent-3-epi-15</i>	<b>68%</b> yield derived from bromide <i>ent-15a</i>	<b>71%</b> yield derived from alcohol <i>ent-15b</i>
<b>Step 7:</b> 10% Pd/C-catalyzed hydrogenolysis reaction			
	<b>79%</b> yield derived from ketone <i>ent-3-epi-19</i>	<b>82%</b> yield derived from ketone <i>ent-19a</i>	<b>83%</b> yield derived from ketone <i>ent-19b</i>
<b>Step 8:</b> Protection of the amino and hydroxyl groups with Ac <sub>2</sub> O			
	<b>94%</b> yield derived from ketone <i>ent-3-epi-20</i>	<b>93%</b> yield derived from ketone <i>ent-20a</i>	<b>90%</b> yield derived from ketone <i>ent-20b</i>
<b>Step 9-10:</b> Bromination-elimination of ketone			
	<b>83%</b> yield in 2 steps derived from ketone <i>ent-3-epi-21</i>	<b>74%</b> yield in 2 steps derived from ketone <i>ent-21a</i>	<b>74%</b> yield in 2 steps derived from ketone <i>ent-21b</i>
<b>Step 11:</b> Removal of acetyl groups			
	<b>77%</b> yield derived from compound <i>ent-3-epi-22</i>	<b>86%</b> yield derived from compound <i>ent-22a</i>	<b>74%</b> yield derived from compound <i>ent-22b</i>
<b>Total Yield In 10 Steps:</b>	<b>18%</b>	<b>23%</b>	<b>23%</b>

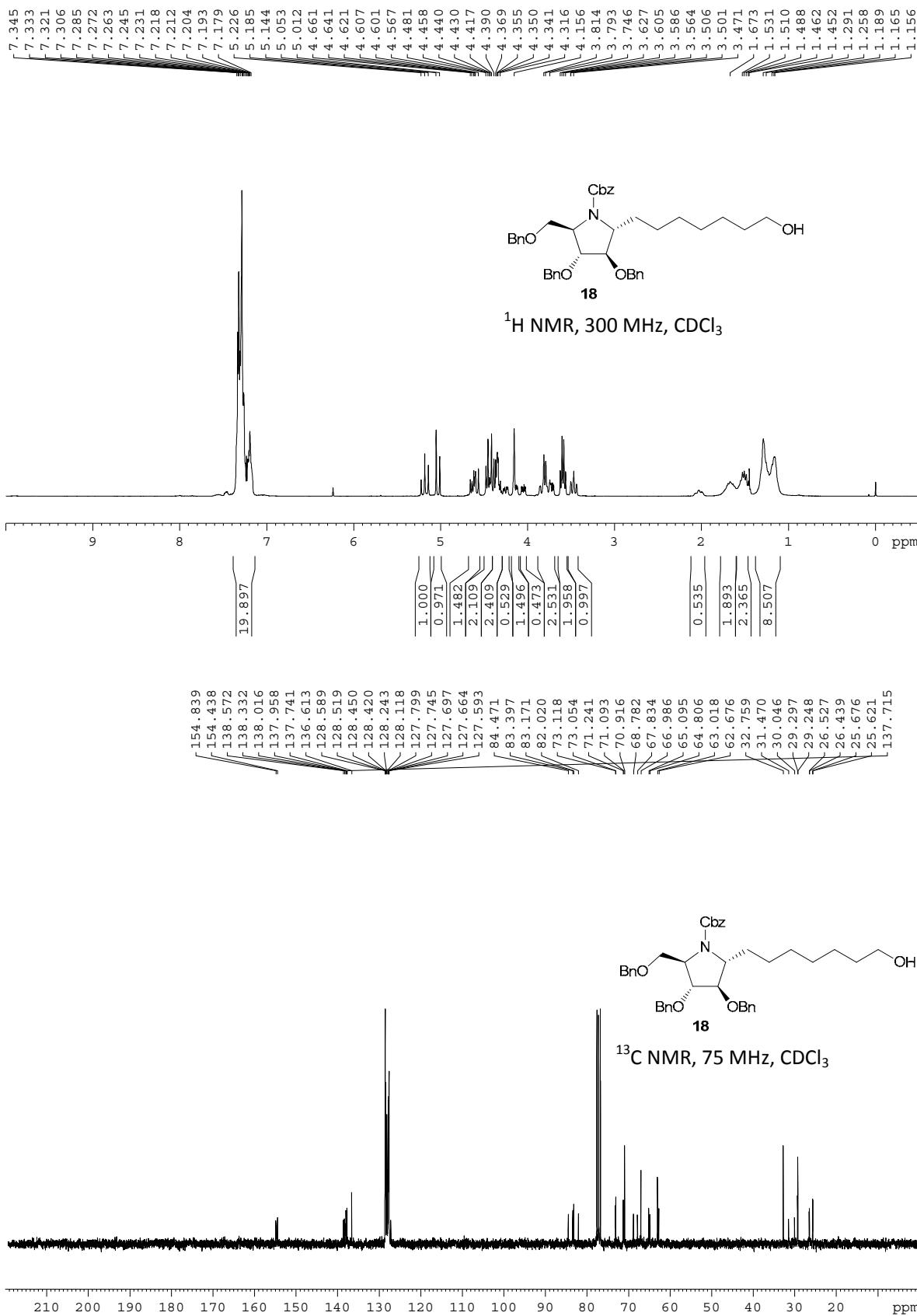
### 3. Copies of $^1\text{H}$ , $^{13}\text{C}$ and 2D NMR spectra of new compounds.

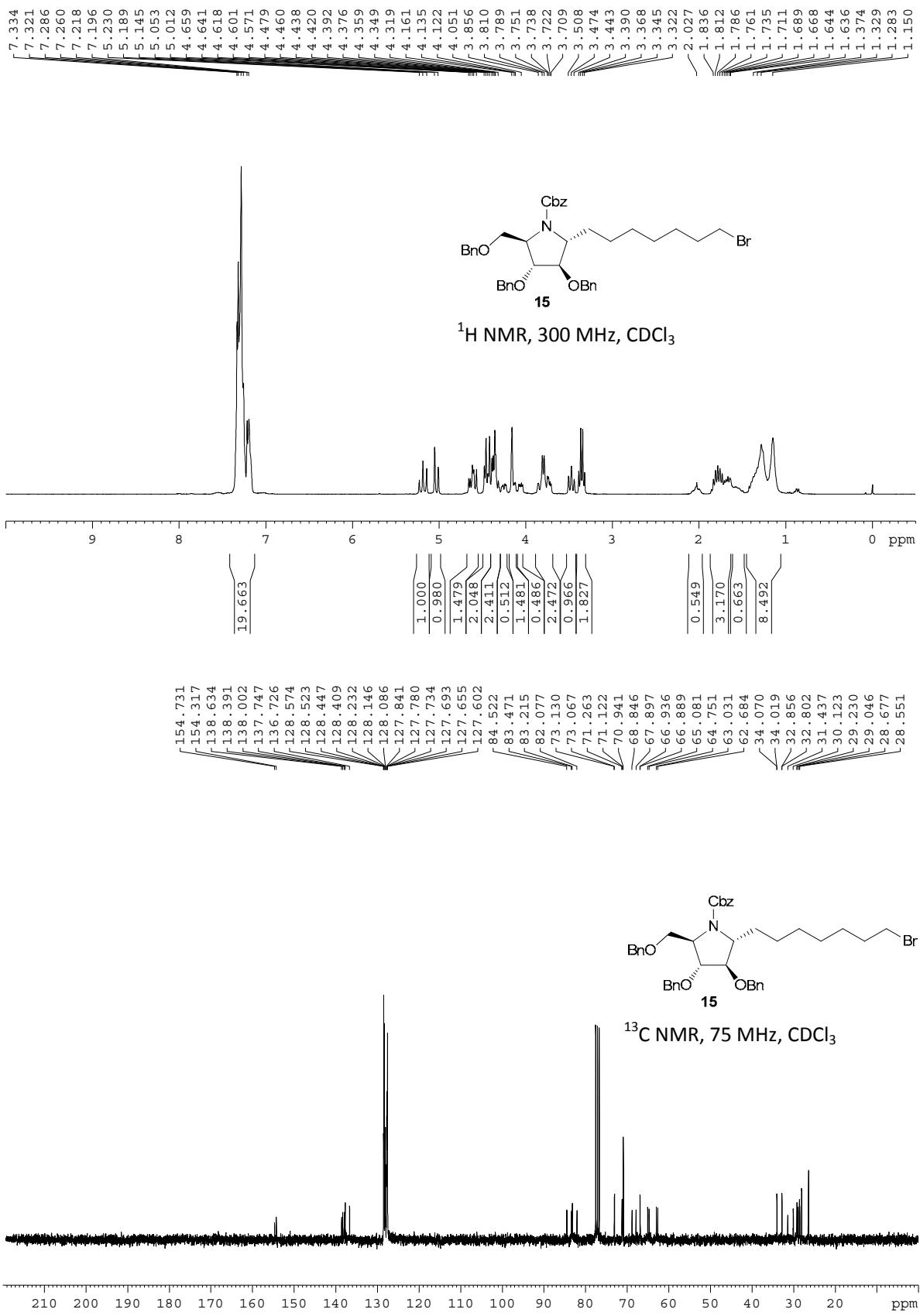


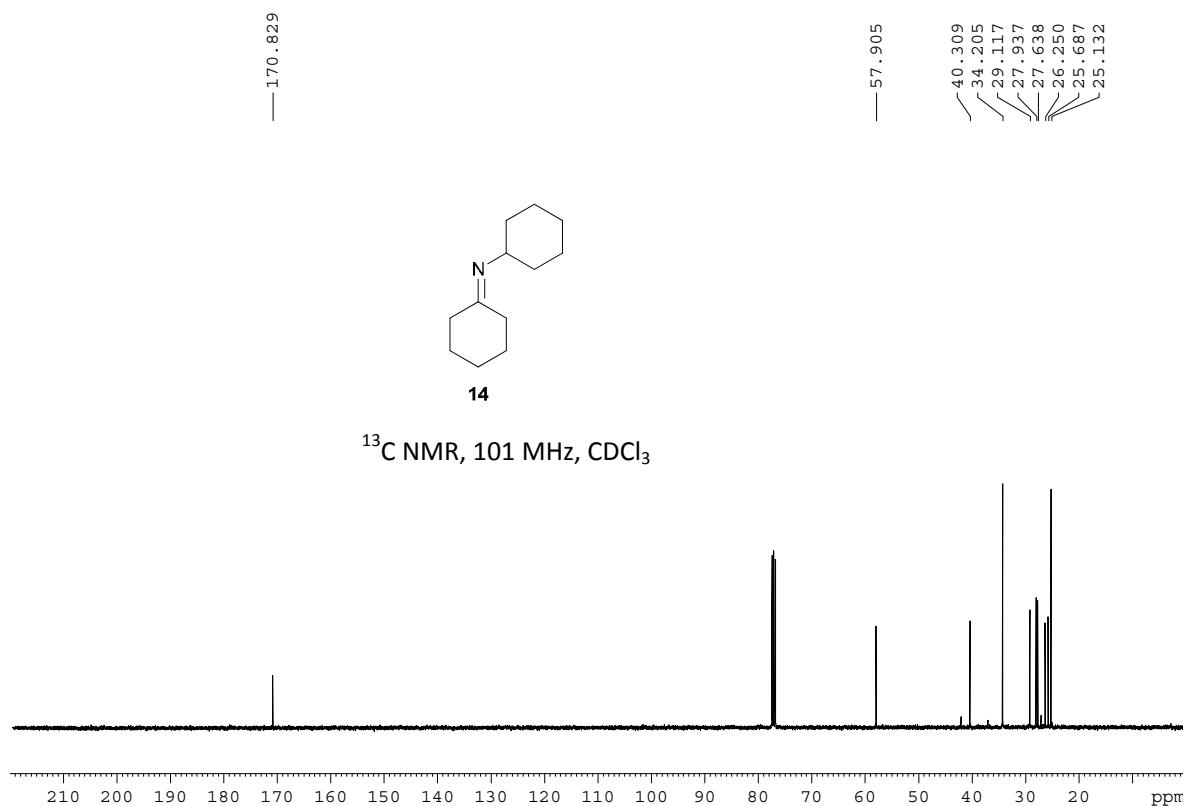
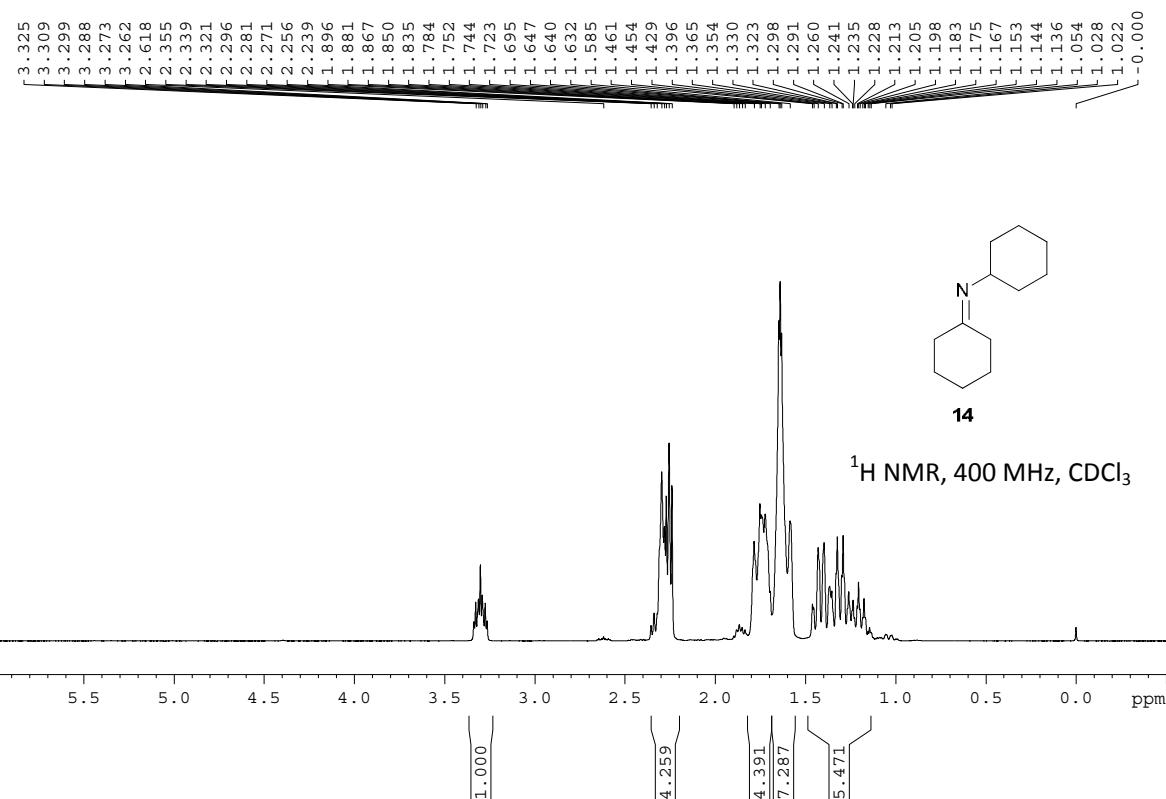
COSY of compound 16

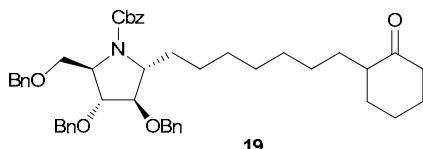
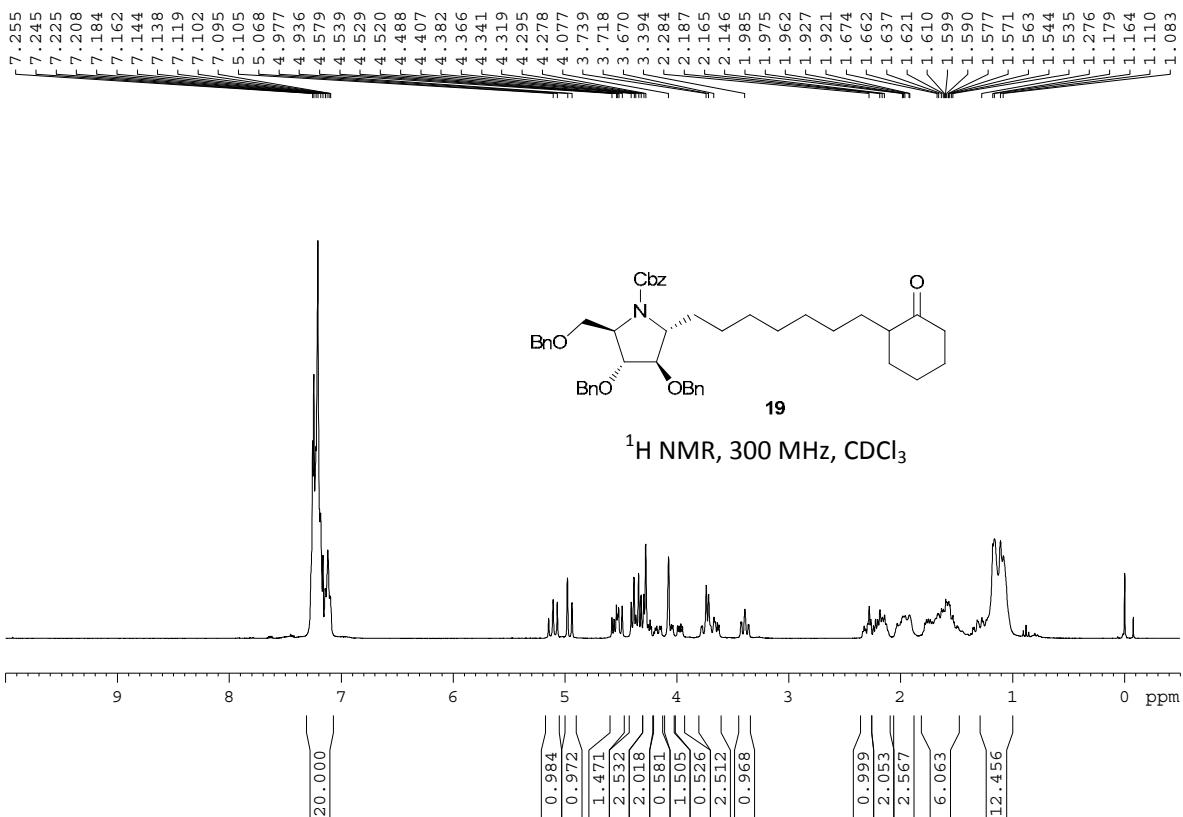




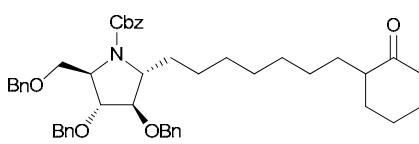
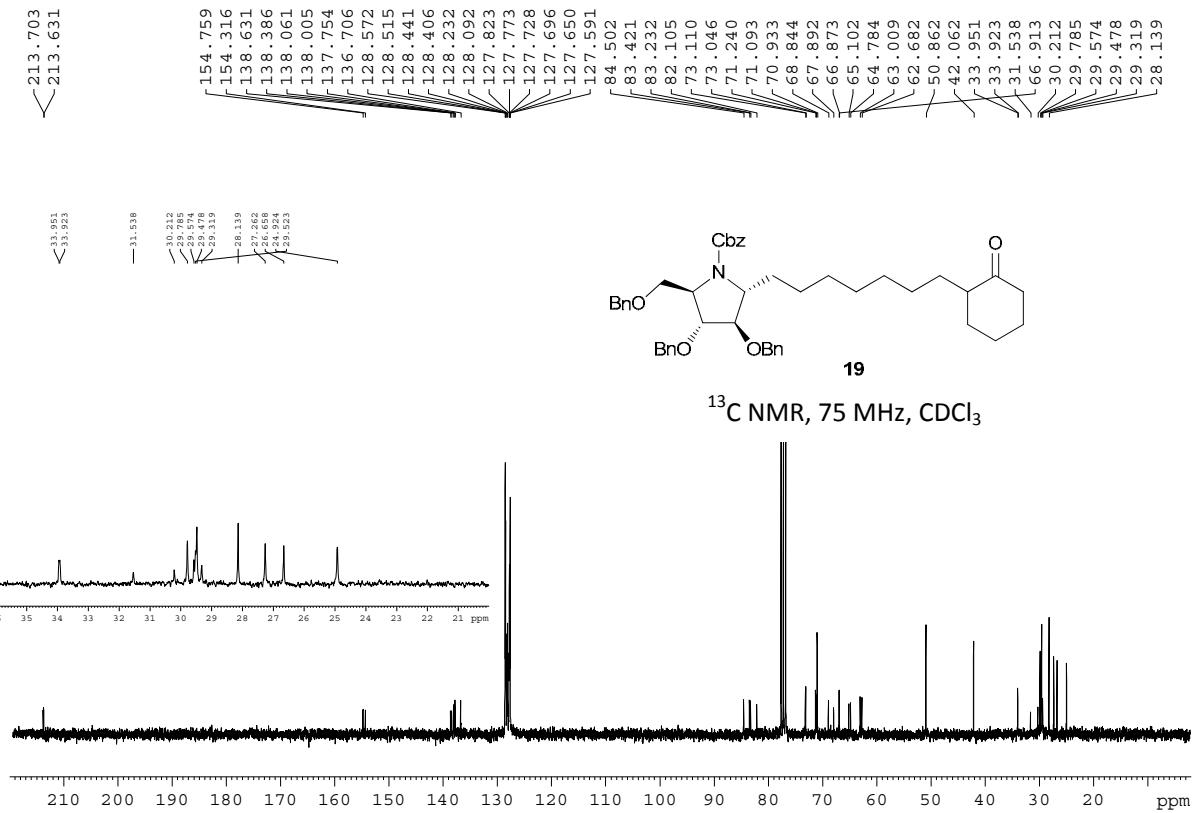




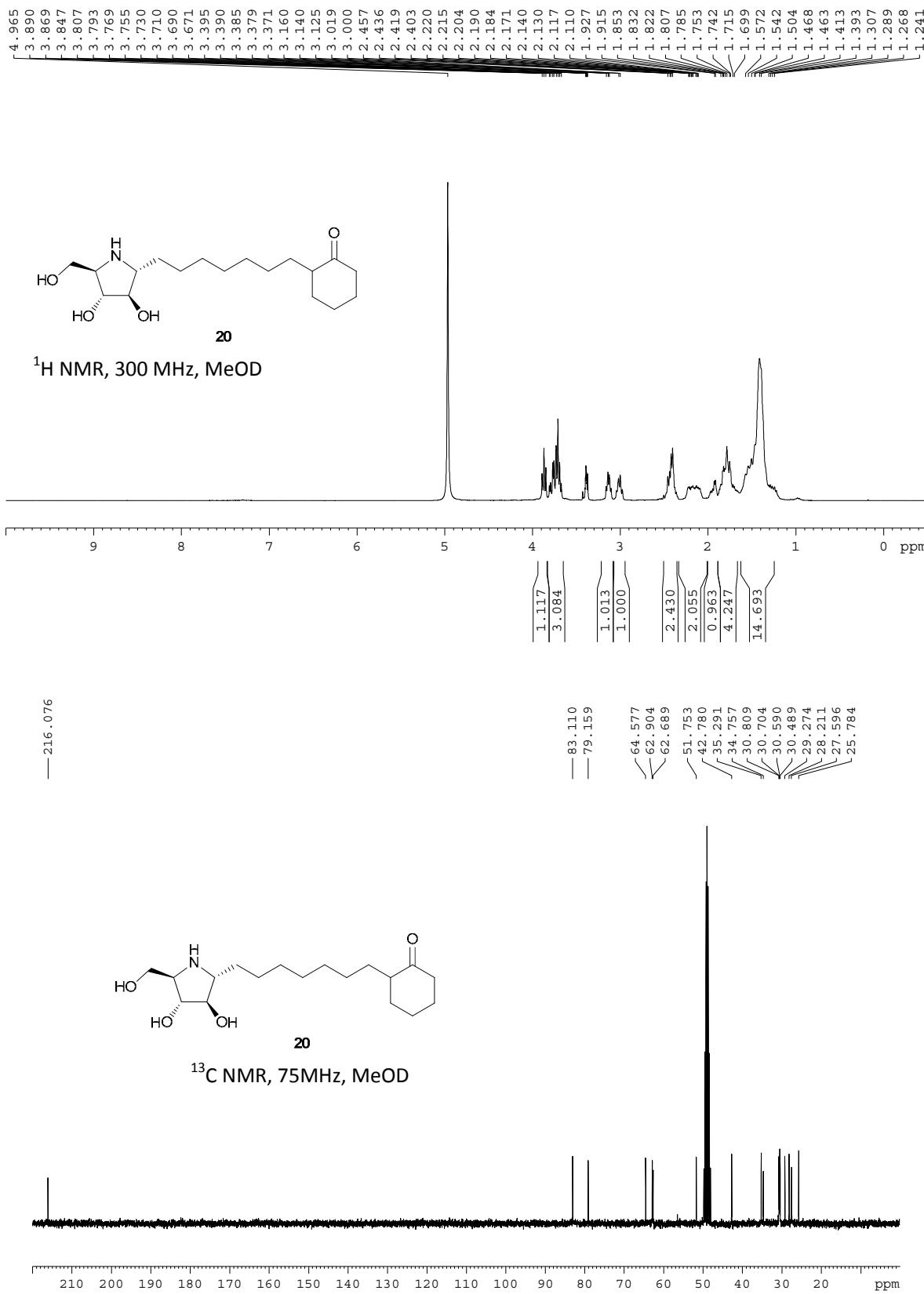


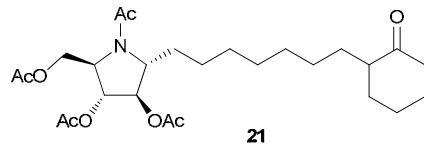
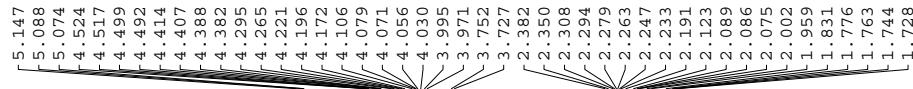


<sup>1</sup>H NMR, 300 MHz, CDCl<sub>3</sub>

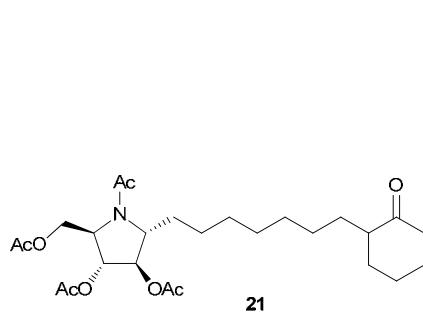
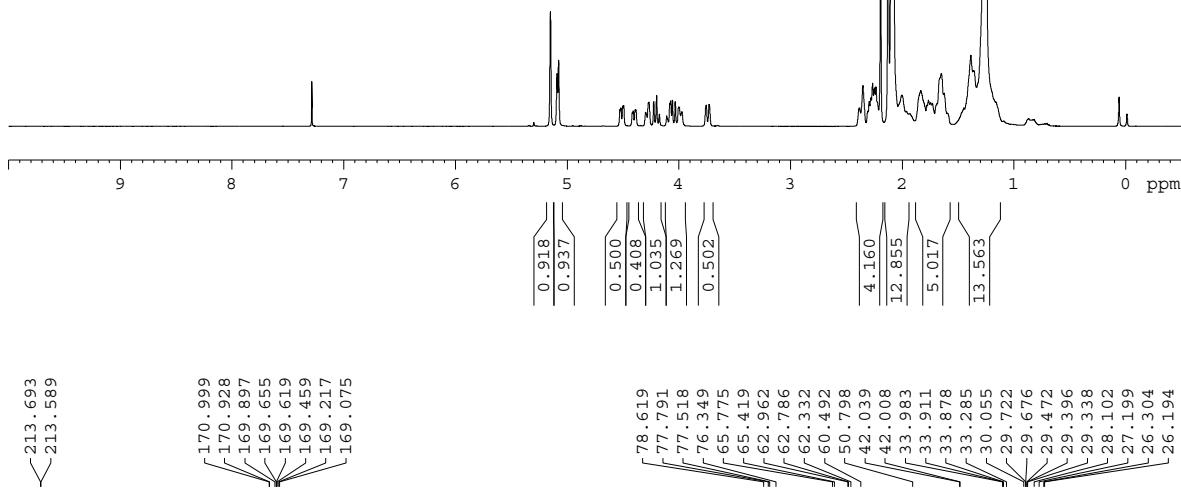


<sup>13</sup>C NMR 75 MHz CDCl<sub>3</sub>

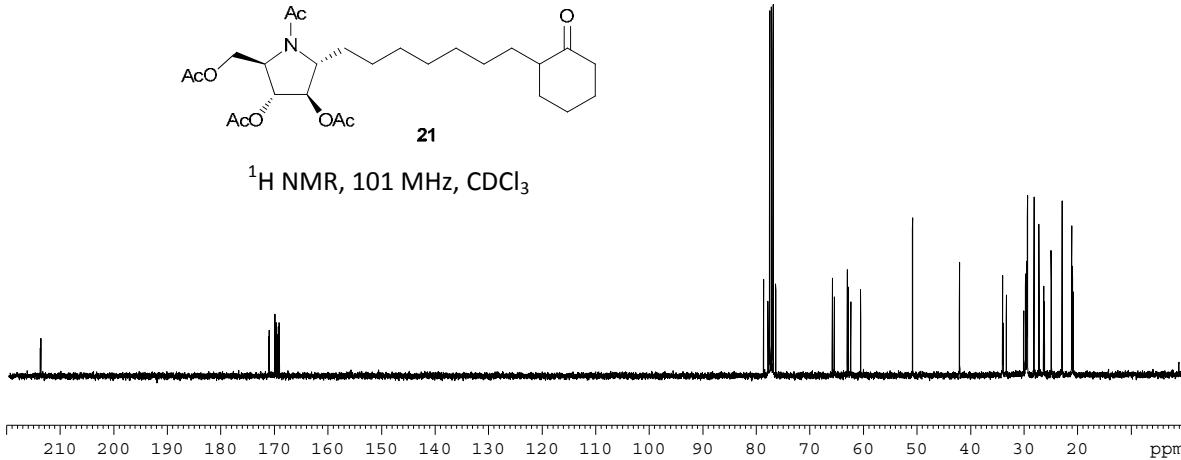


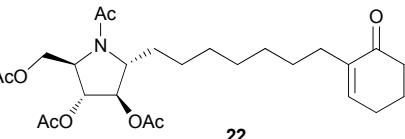
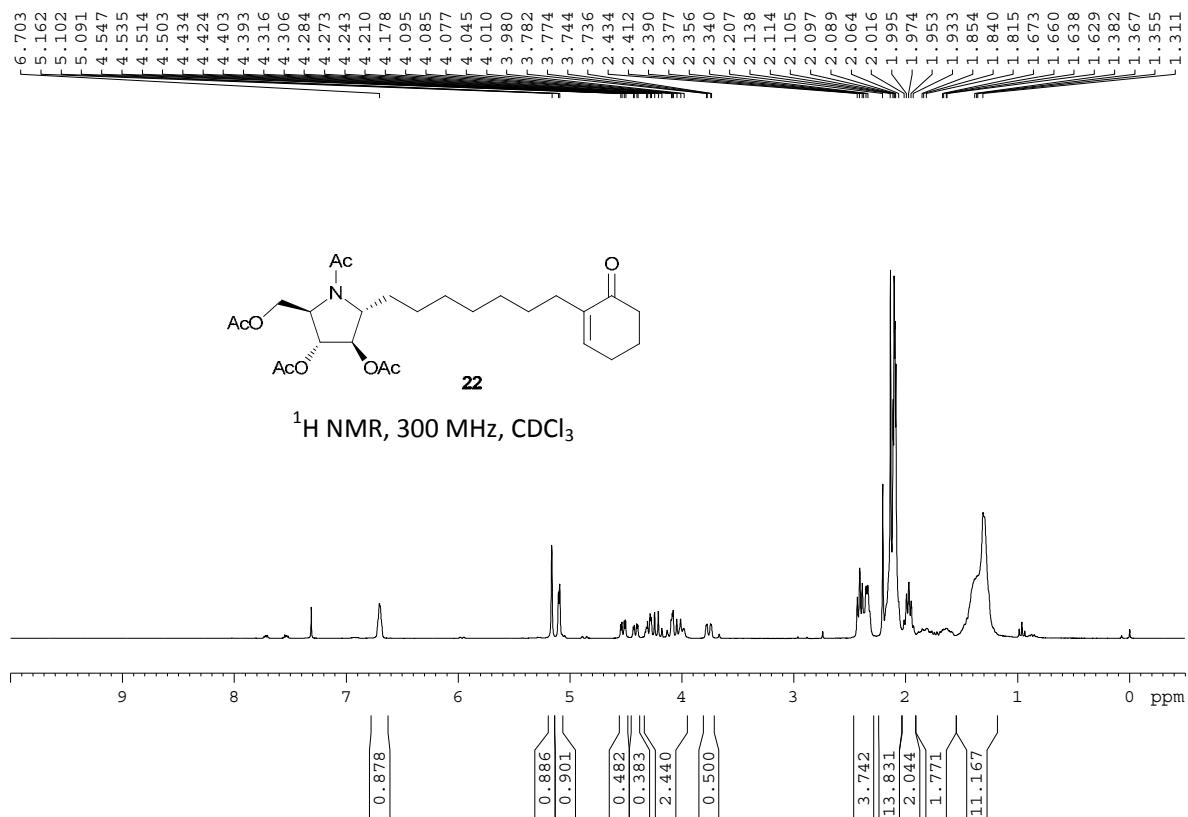


$^1\text{H}$  NMR, 400 MHz,  $\text{CDCl}_3$

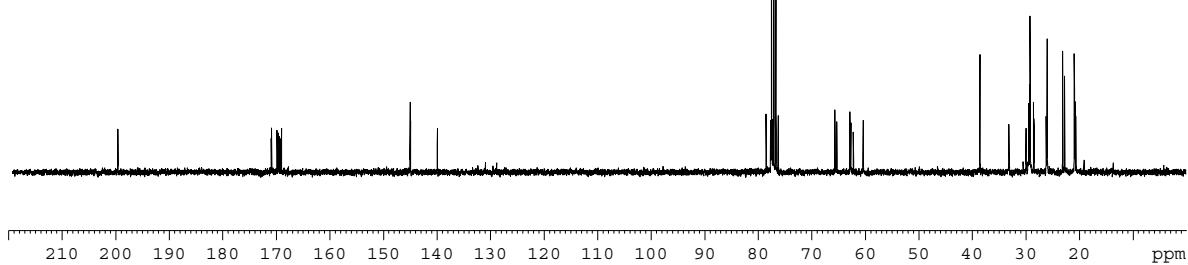


$^1\text{H}$  NMR, 101 MHz,  $\text{CDCl}_3$

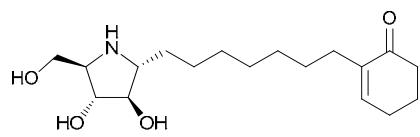




<sup>13</sup>C NMR, 75 MHz, CDCl<sub>3</sub>

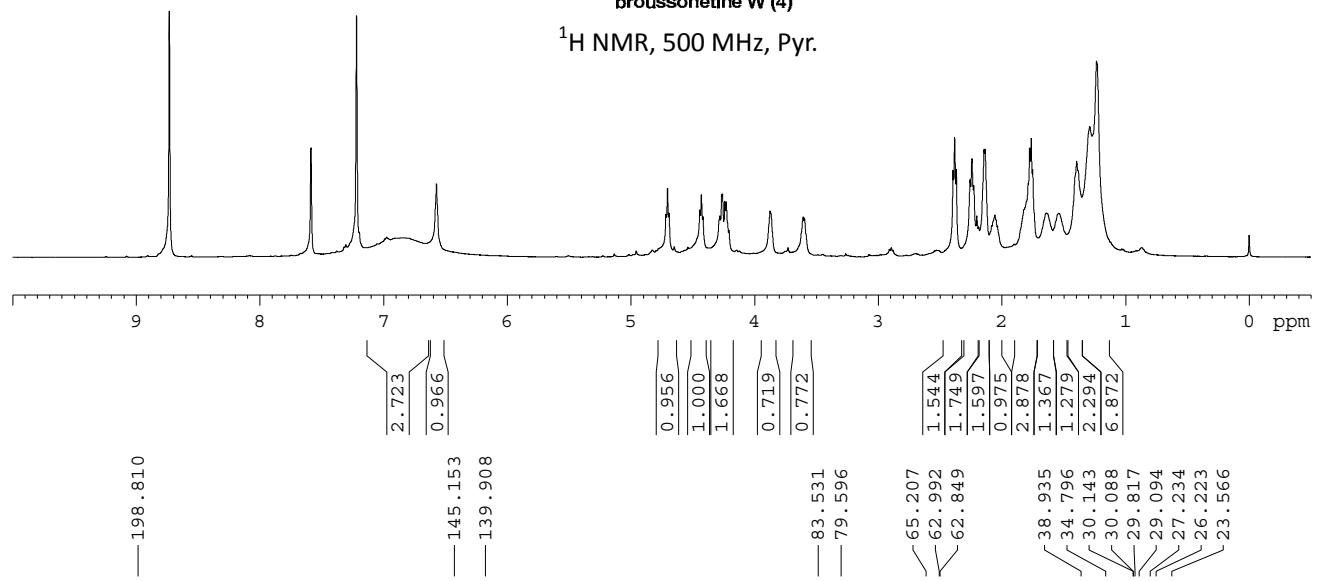


— 6.909  
 — 6.572  
 — 4.716  
 — 4.704  
 — 4.691  
 — 4.443  
 — 4.429  
 — 4.417  
 — 4.280  
 — 4.265  
 — 4.259  
 — 4.239  
 — 4.228  
 — 4.206  
 — 3.874  
 — 3.867  
 — 3.607  
 — 3.597  
 — 3.294  
 — 2.381  
 — 2.368  
 — 2.256  
 — 2.241  
 — 2.227  
 — 2.187  
 — 2.143  
 — 2.135  
 — 2.082  
 — 2.055  
 — 2.047  
 — 2.035  
 — 1.802  
 — 1.787  
 — 1.775  
 — 1.762  
 — 1.750  
 — 1.638  
 — 1.629  
 — 1.539  
 — 1.395



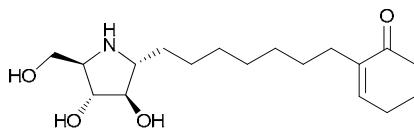
**broussonetine W (4)**

$^1\text{H}$  NMR, 500 MHz, Pyr.



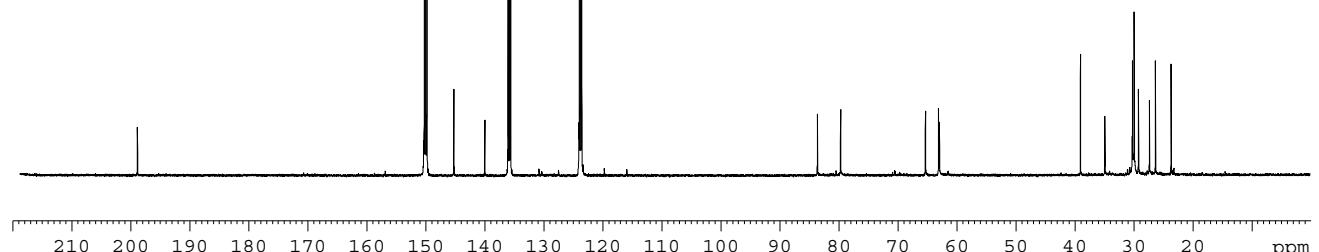
— 198.810

— 145.153  
 — 139.908

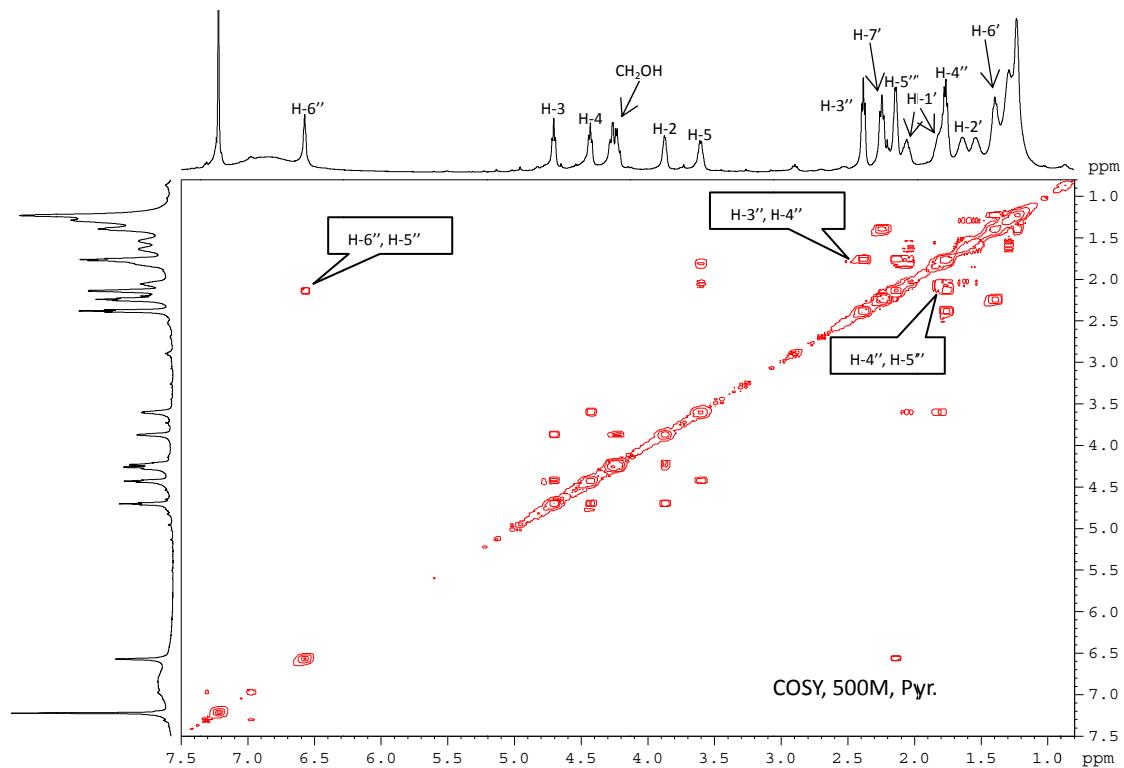


**broussonetine W (4)**

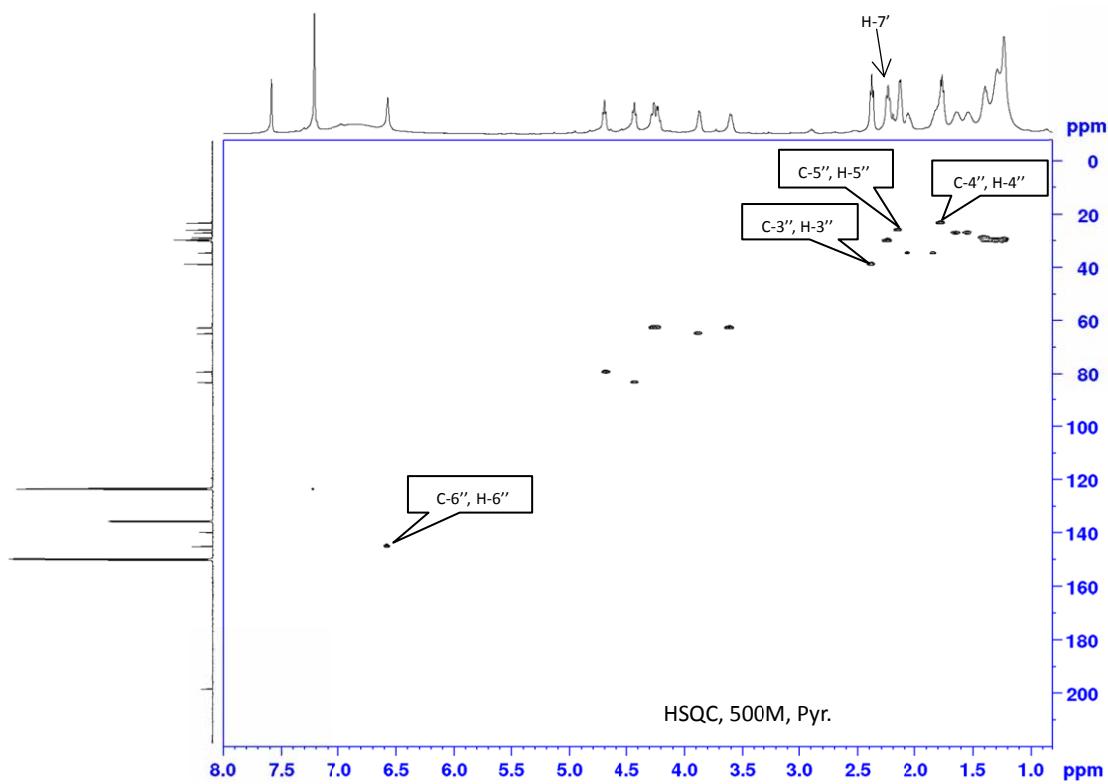
$^1\text{H}$  NMR, 126 MHz, Pyr.



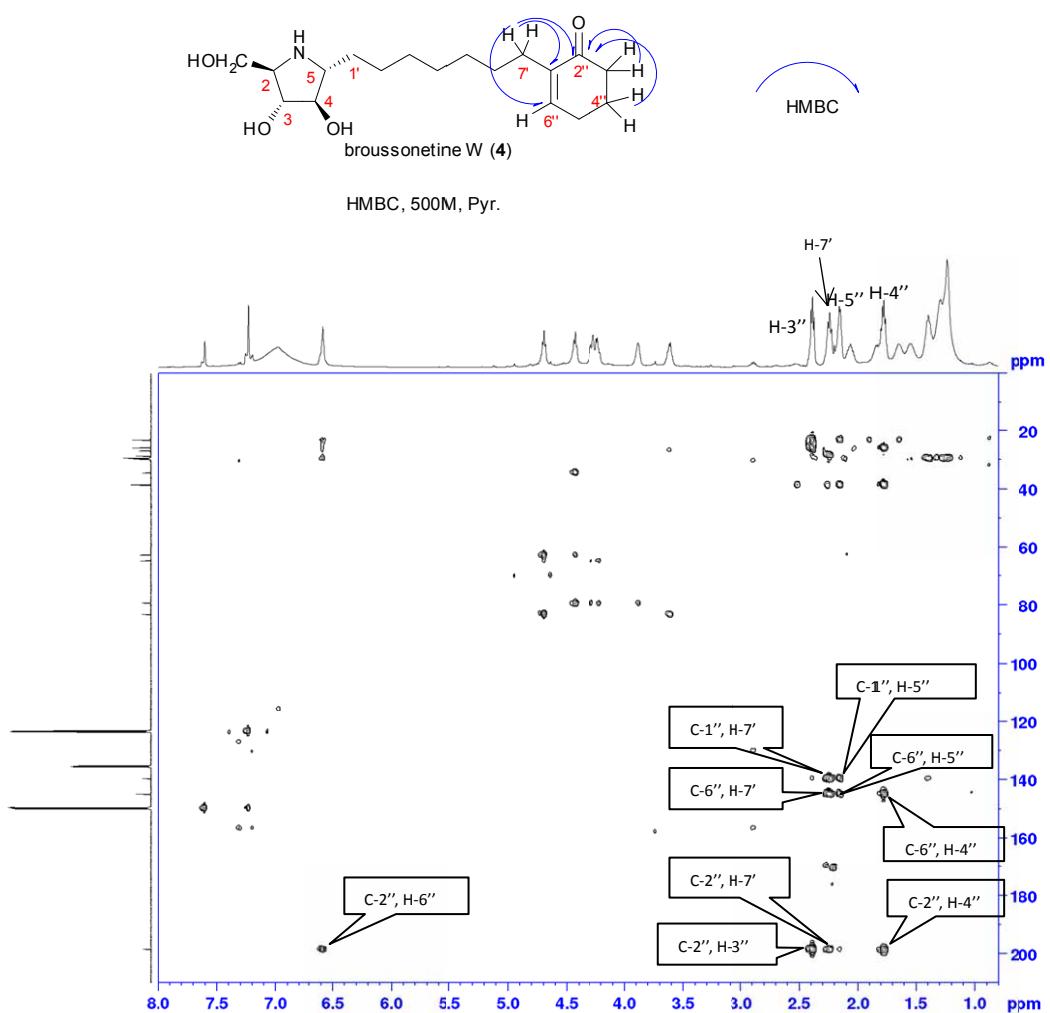
COSY of the synthetic broussonetine W (**4**)

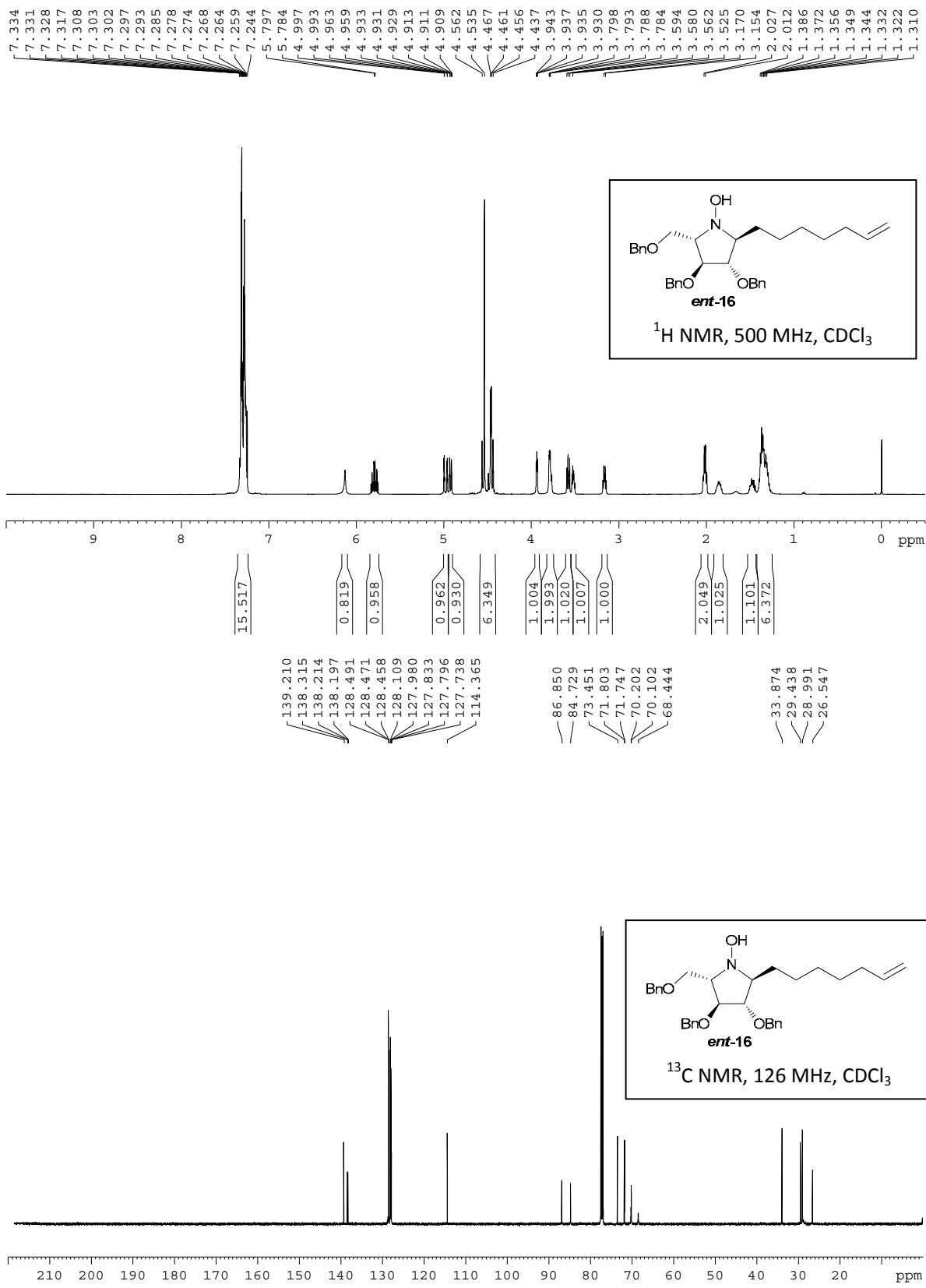


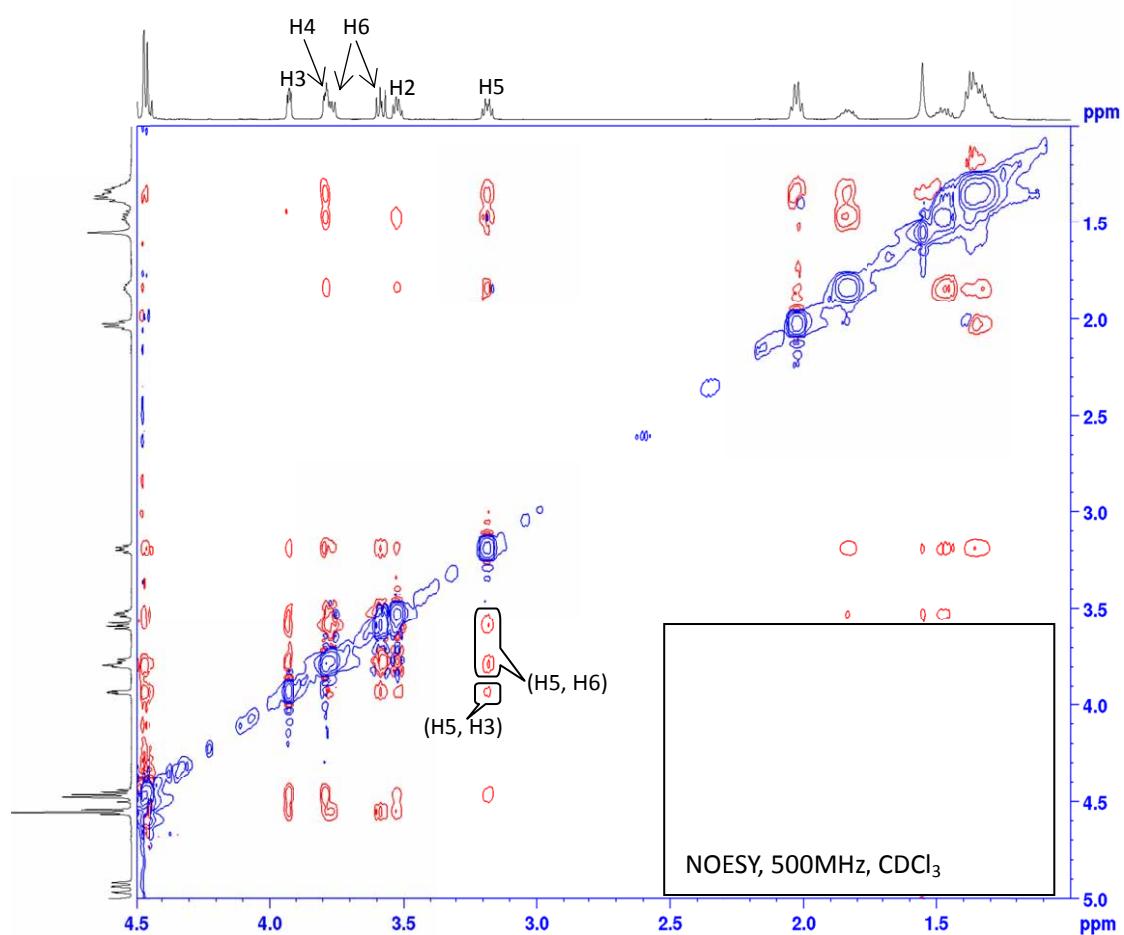
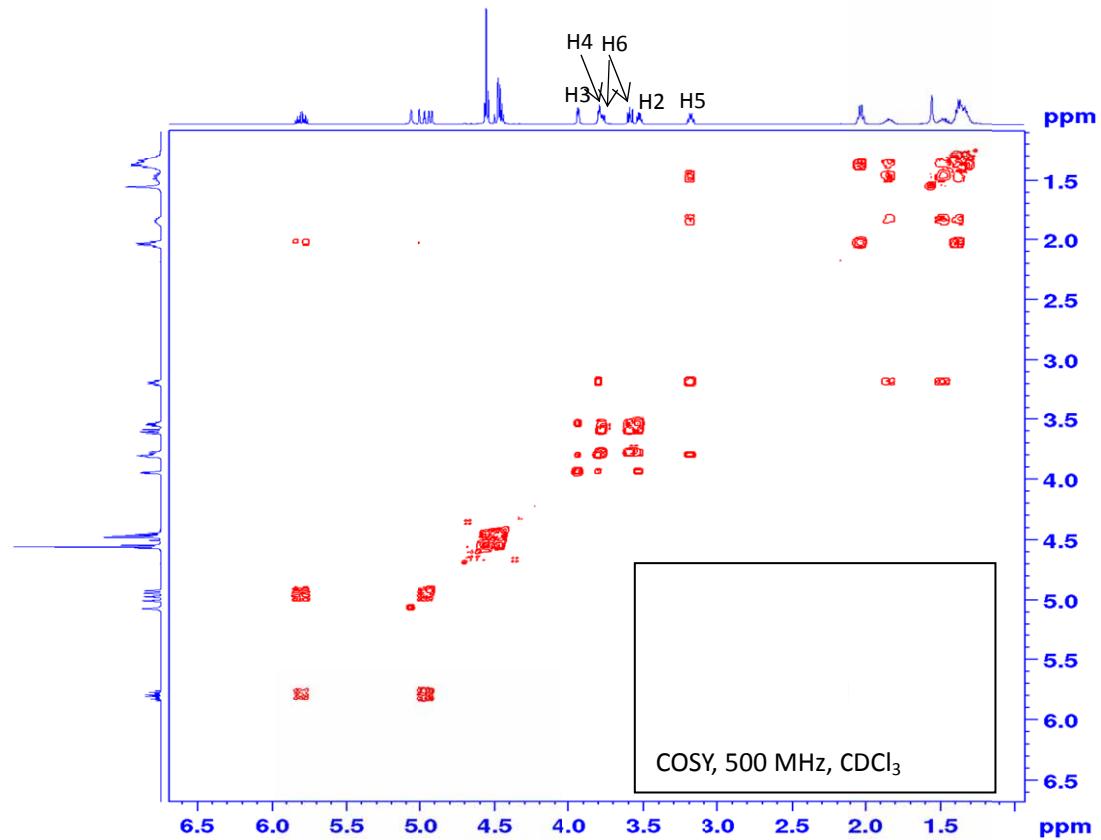
HSQC of the synthetic broussonetine W (**4**)

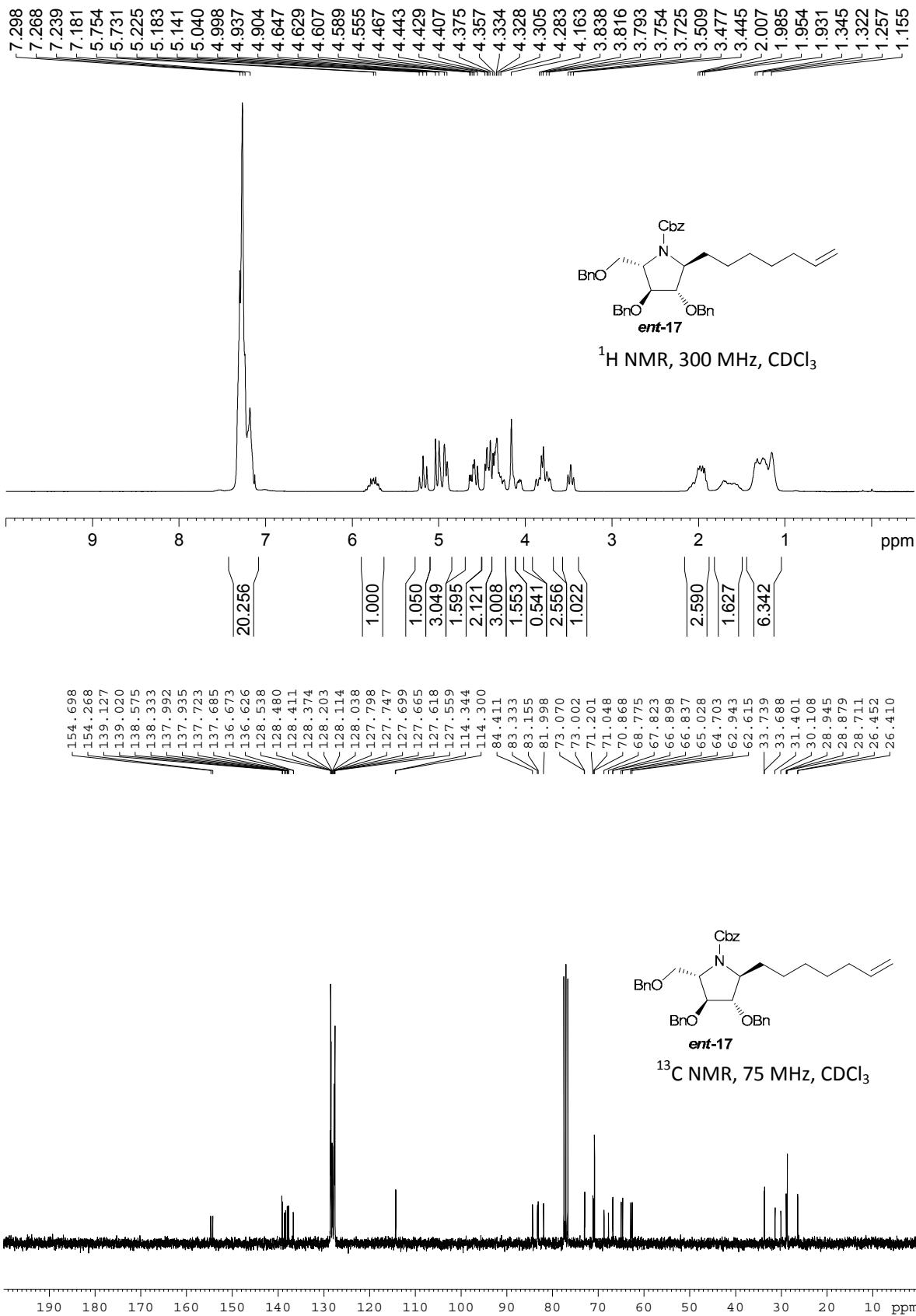


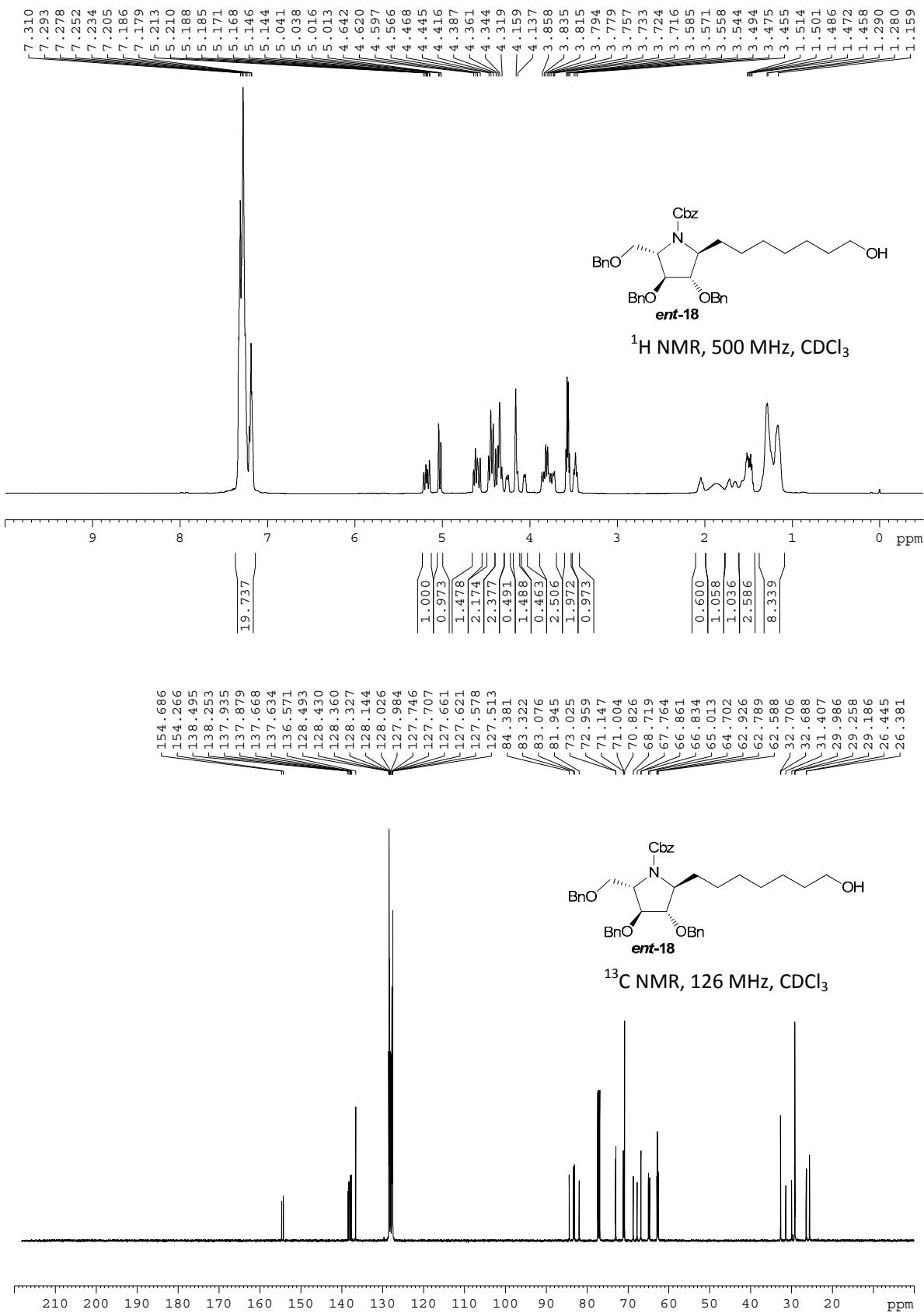
HMBC of the synthetic broussonetine W (**4**)

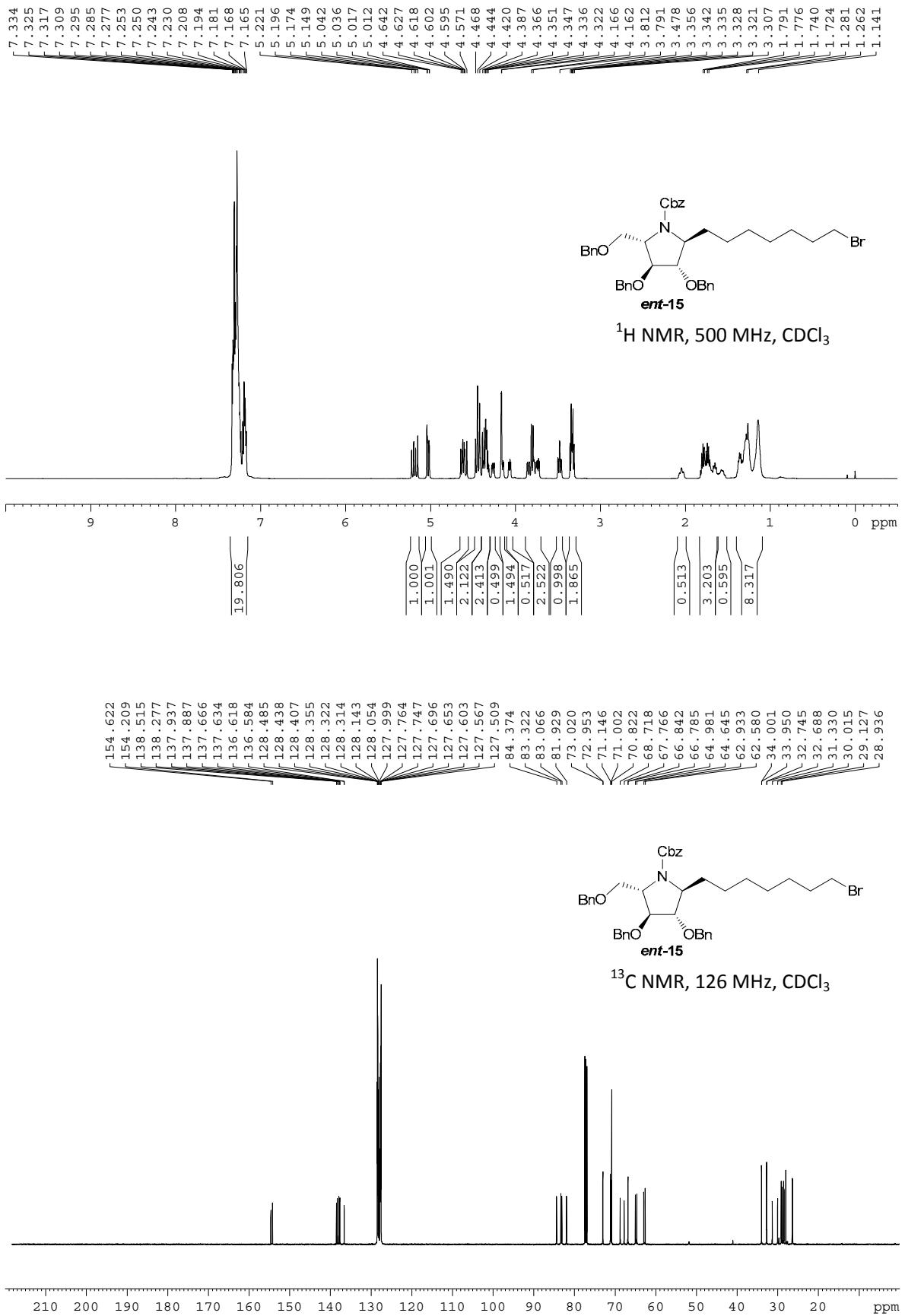


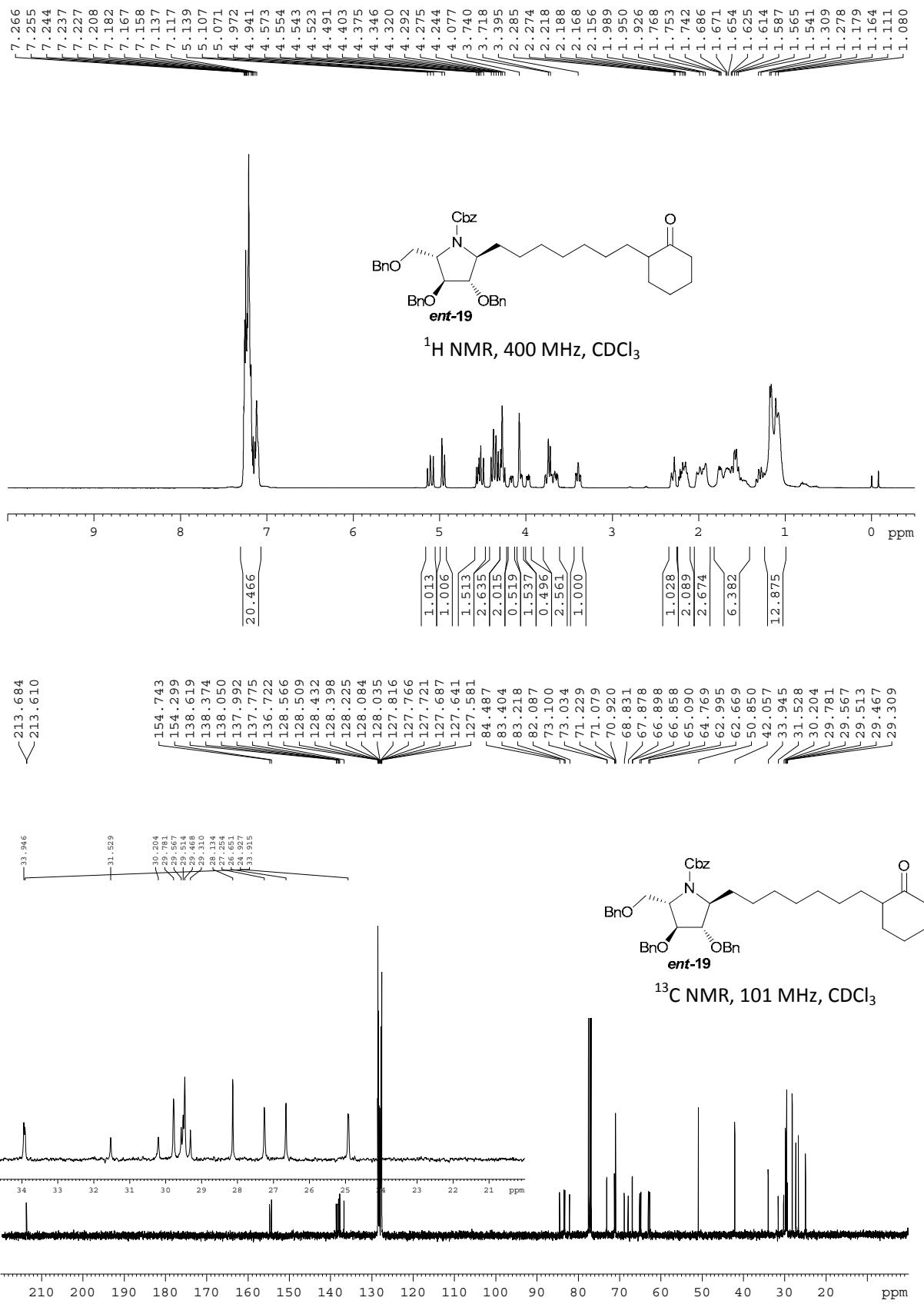


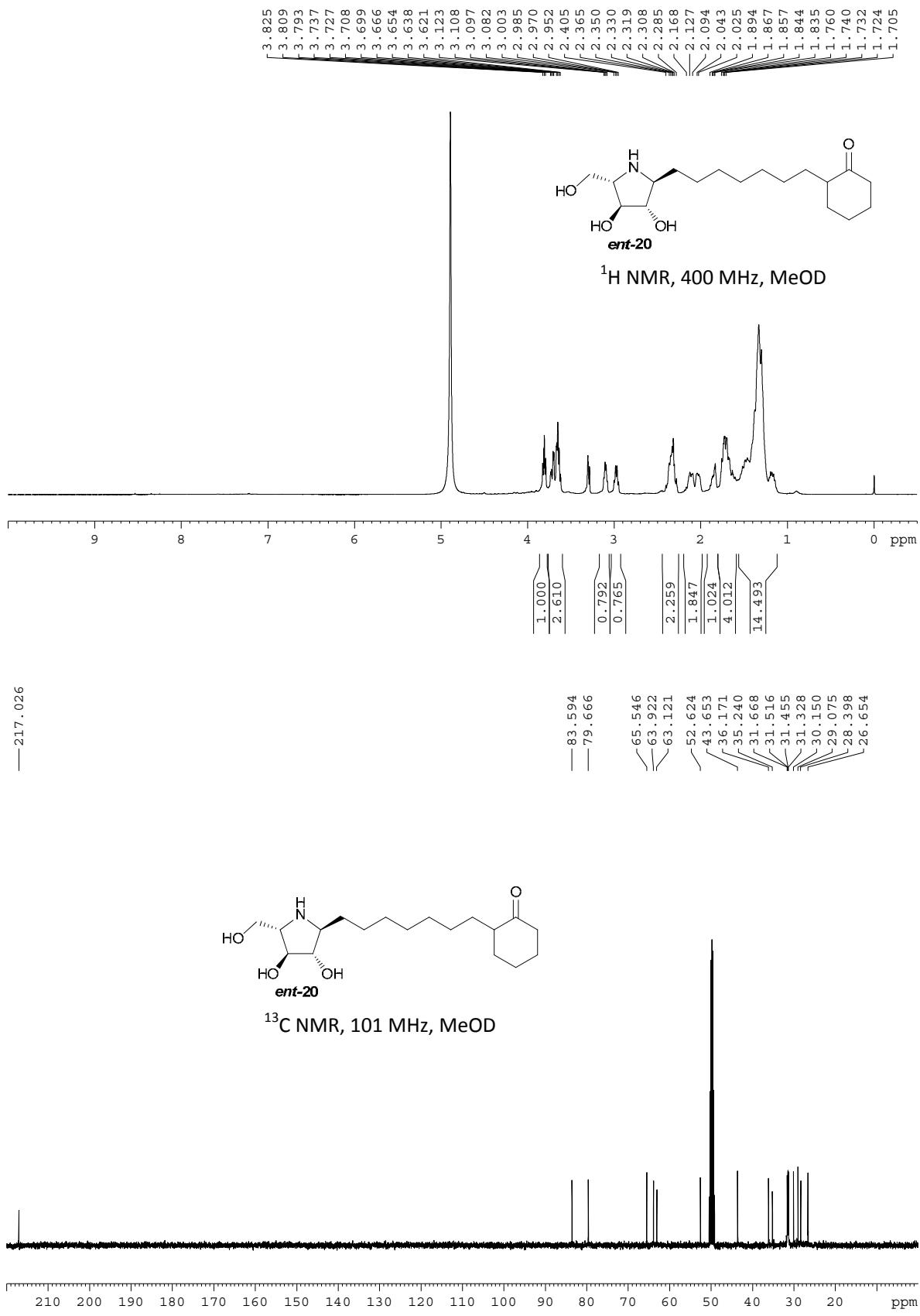


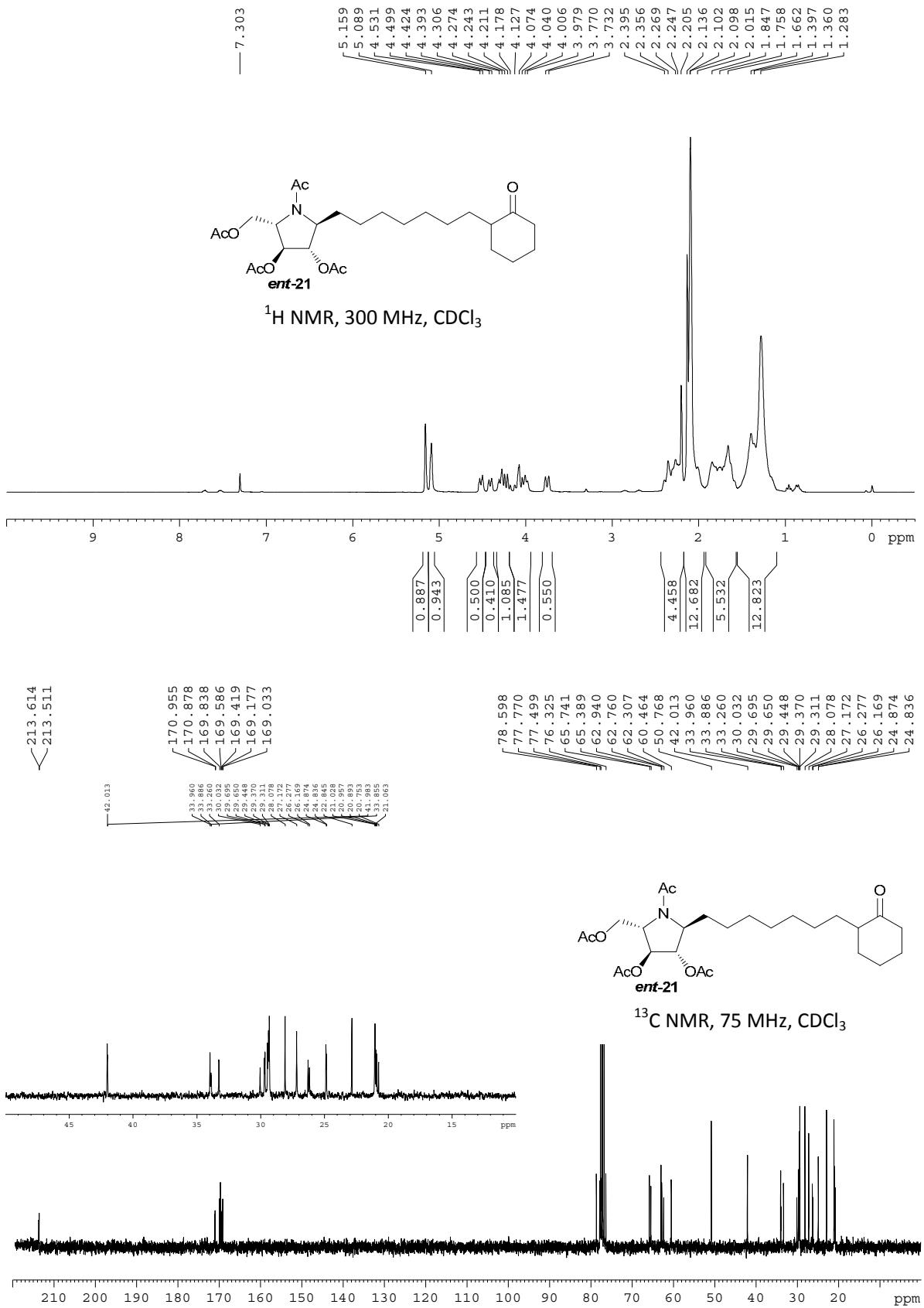


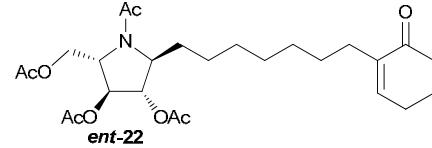
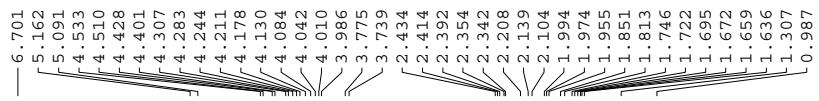




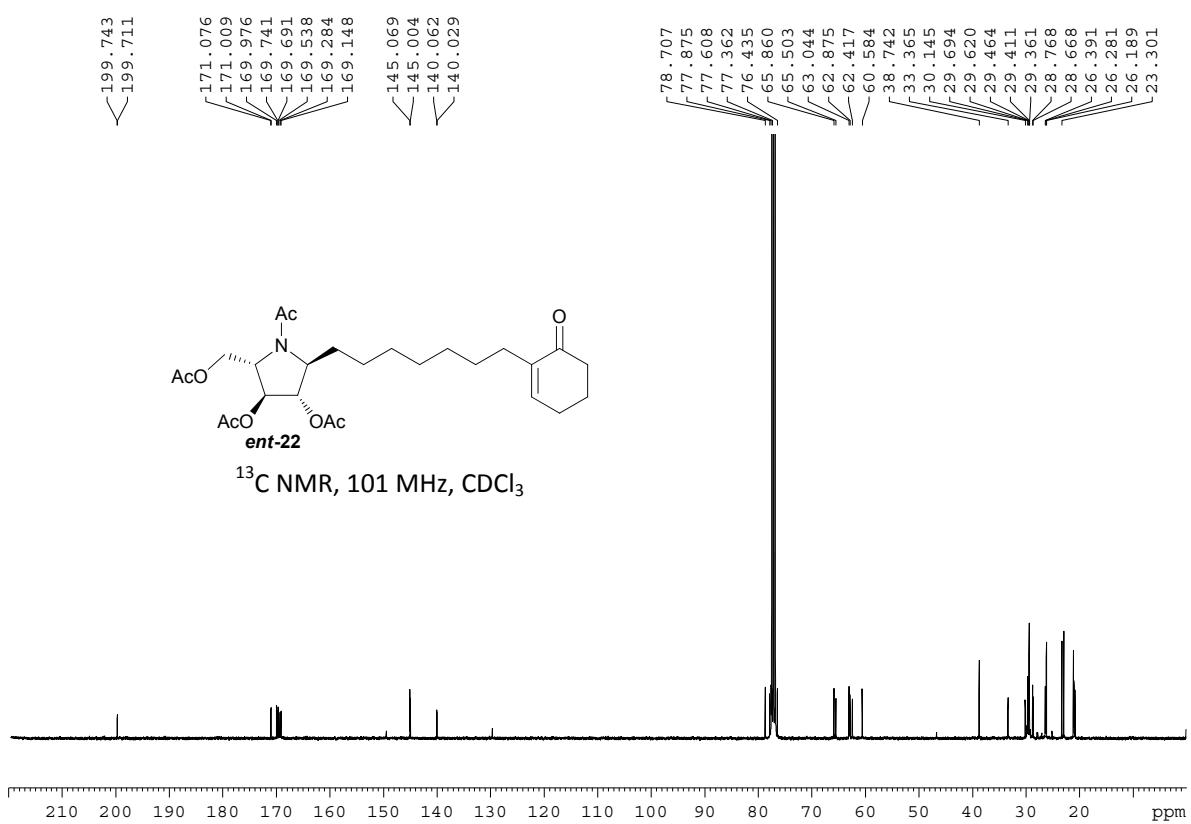


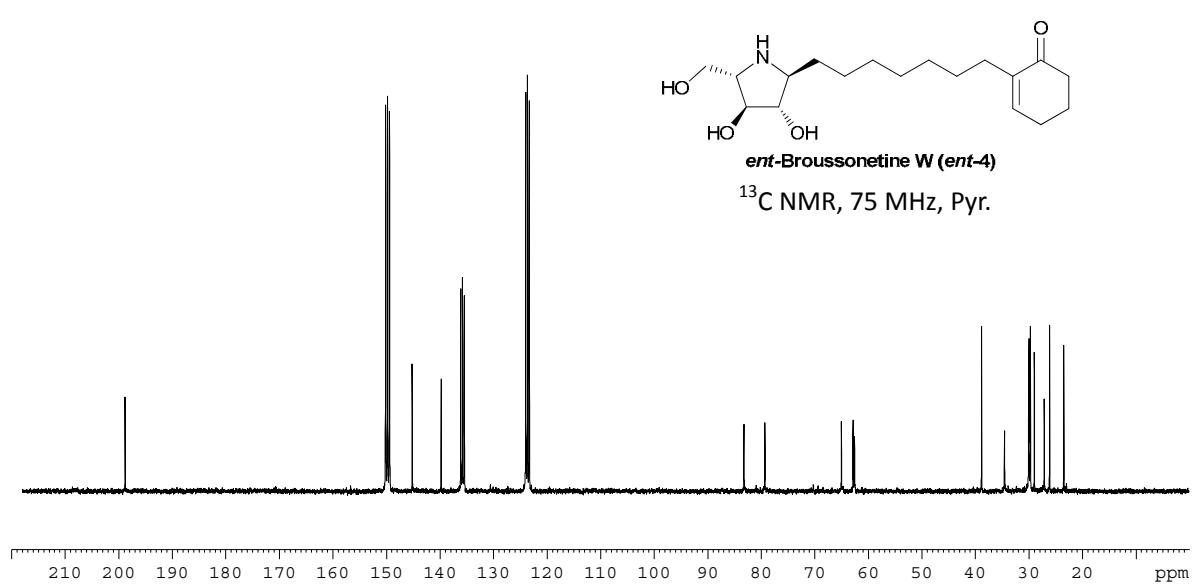
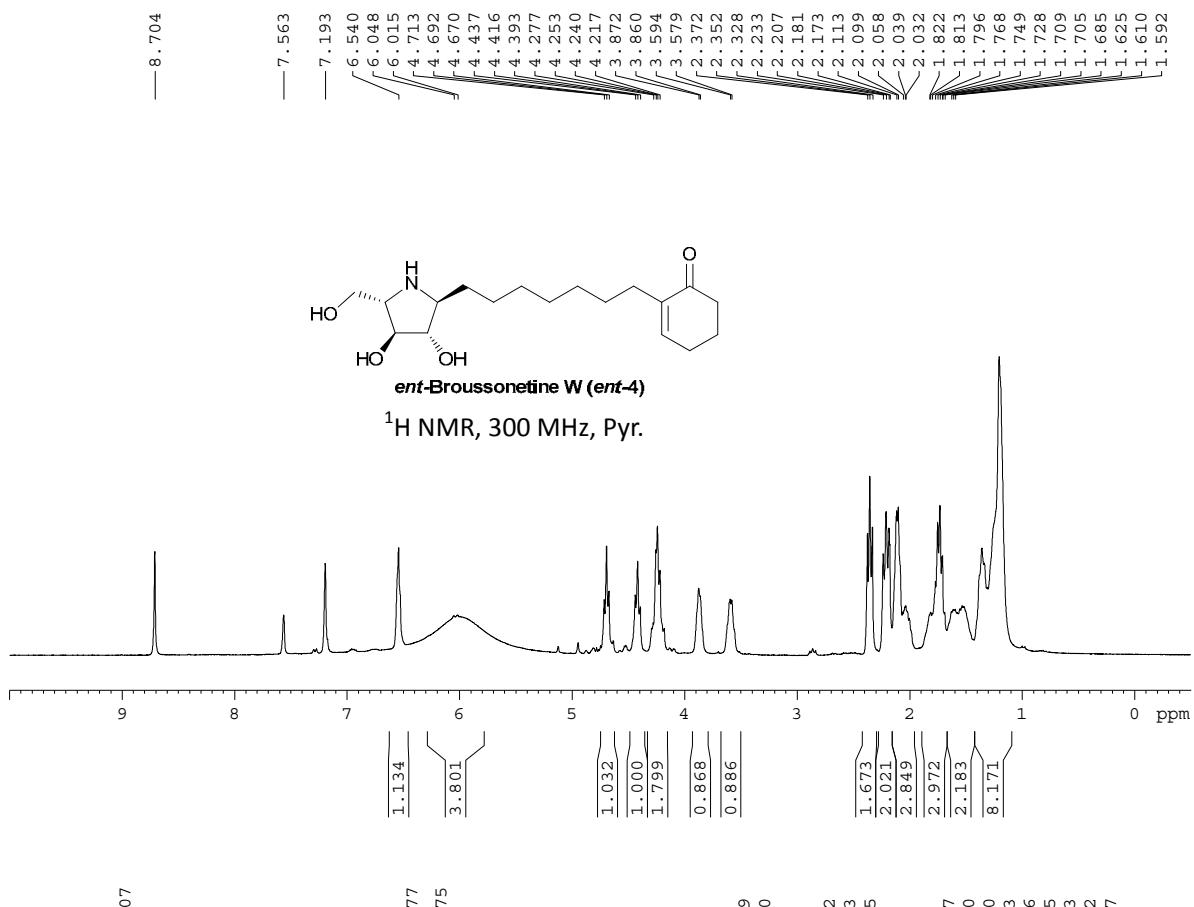


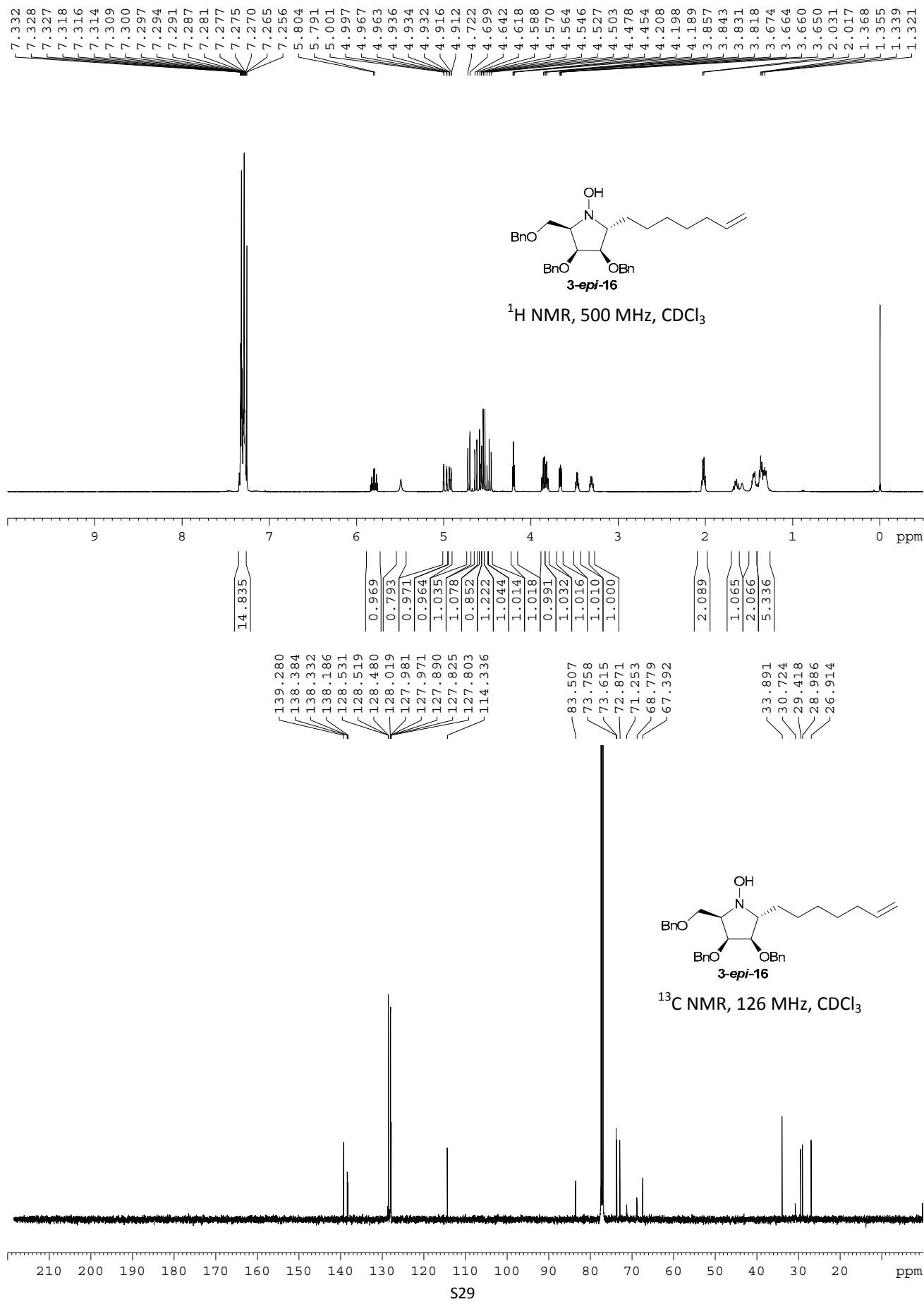


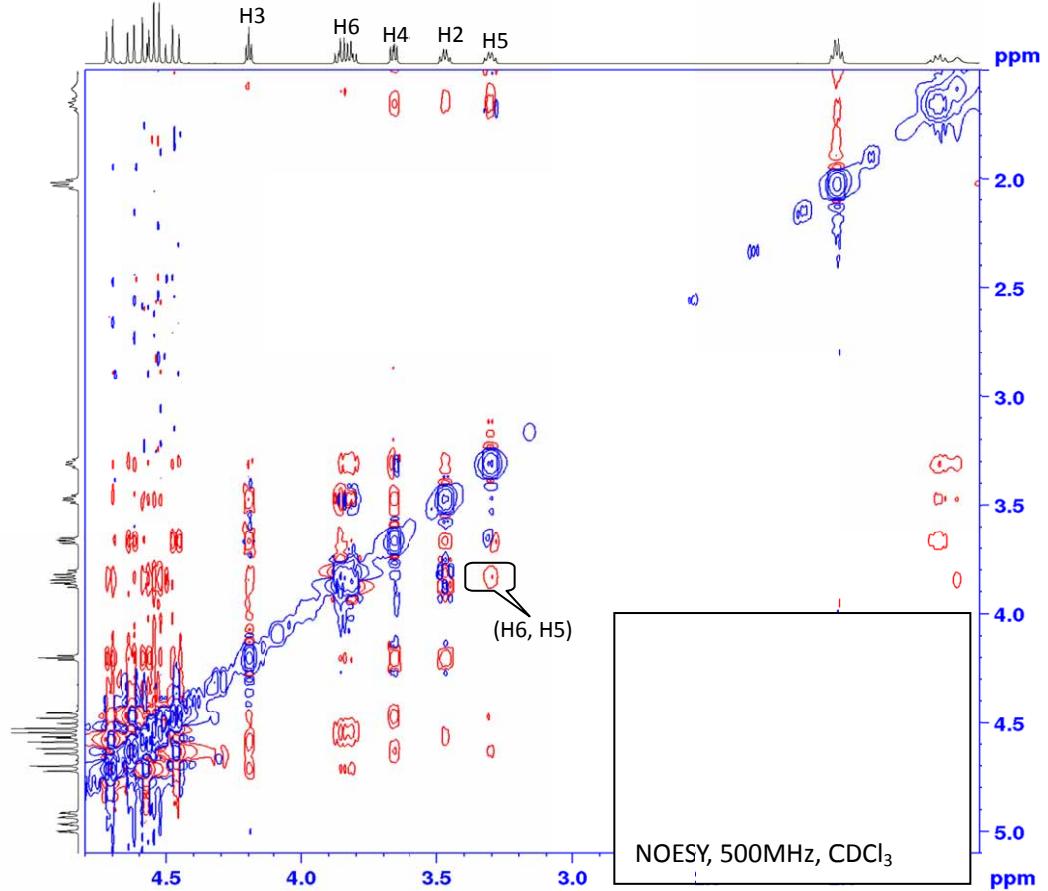
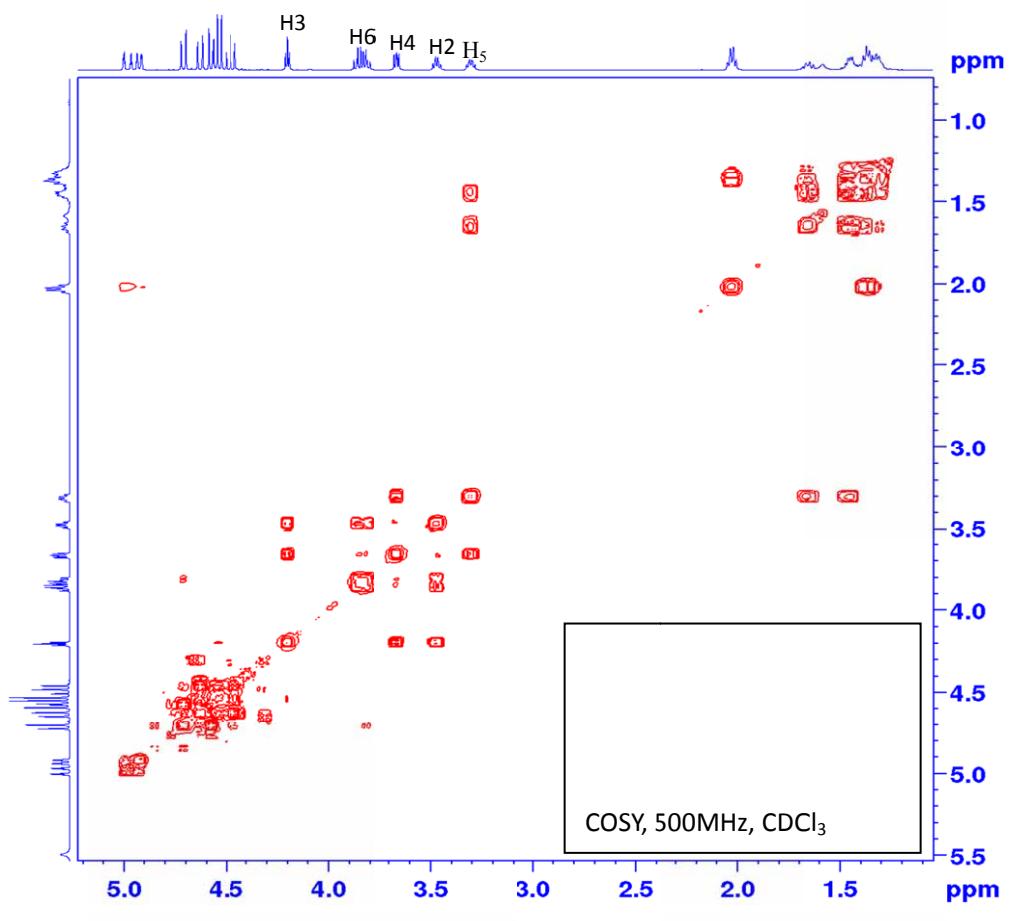


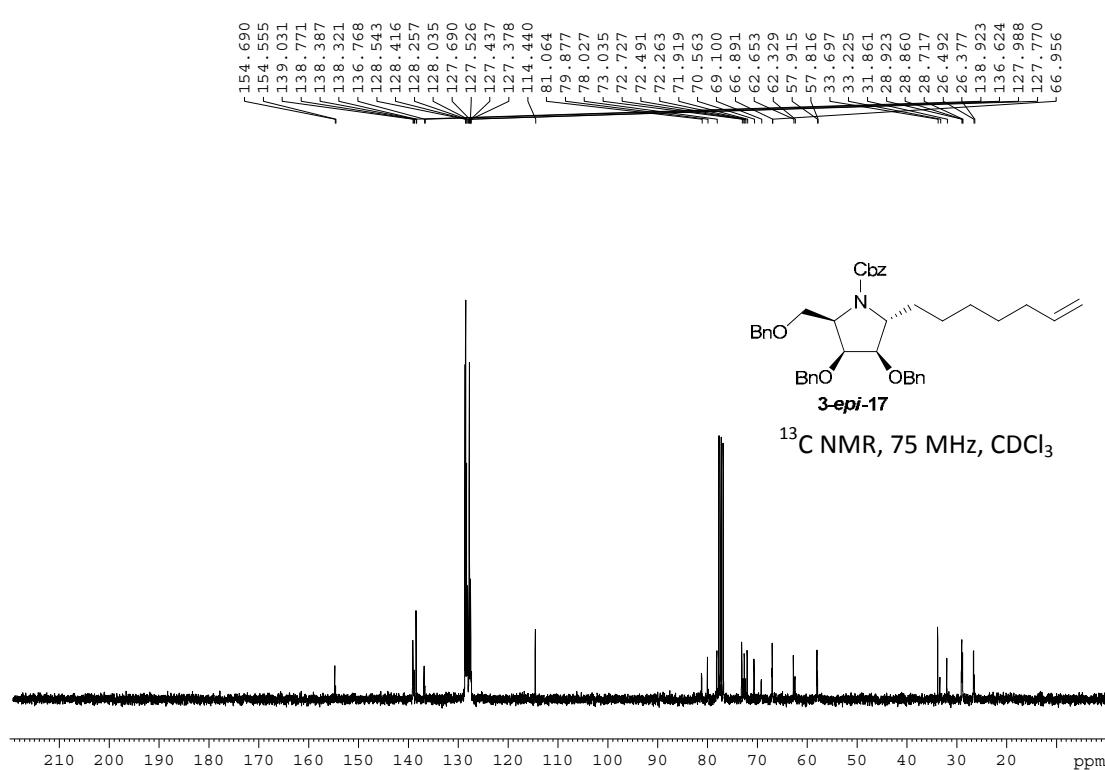
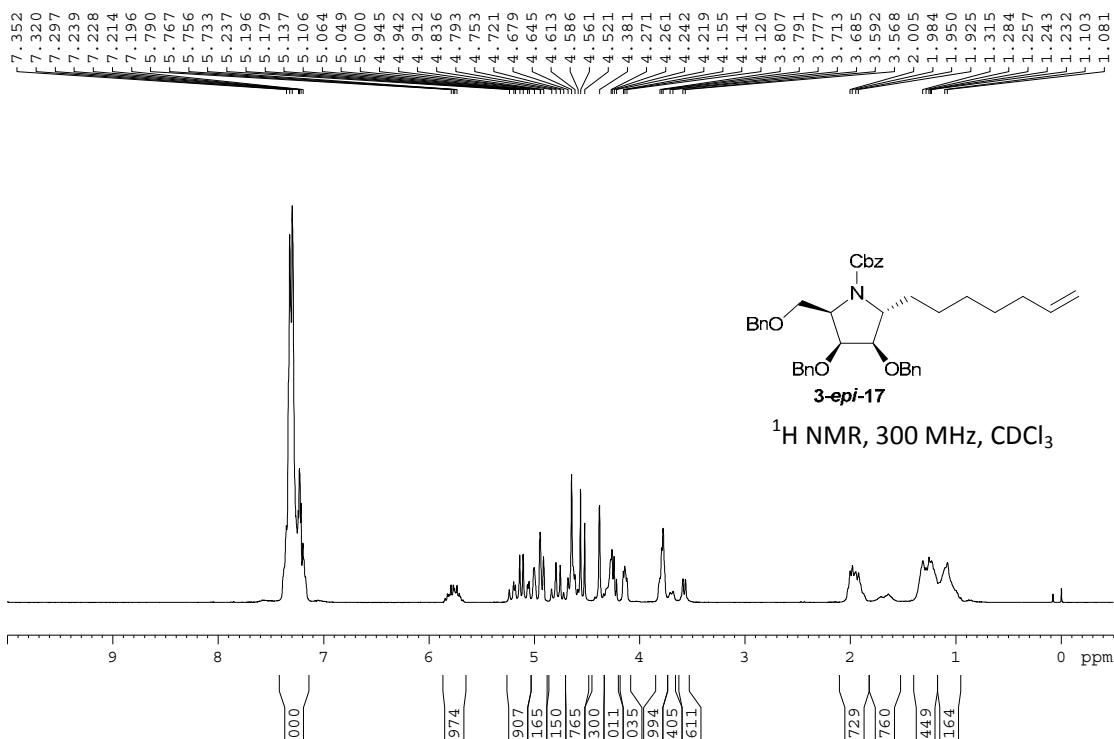
<sup>1</sup>H NMR, 300 MHz, CDCl<sub>3</sub>

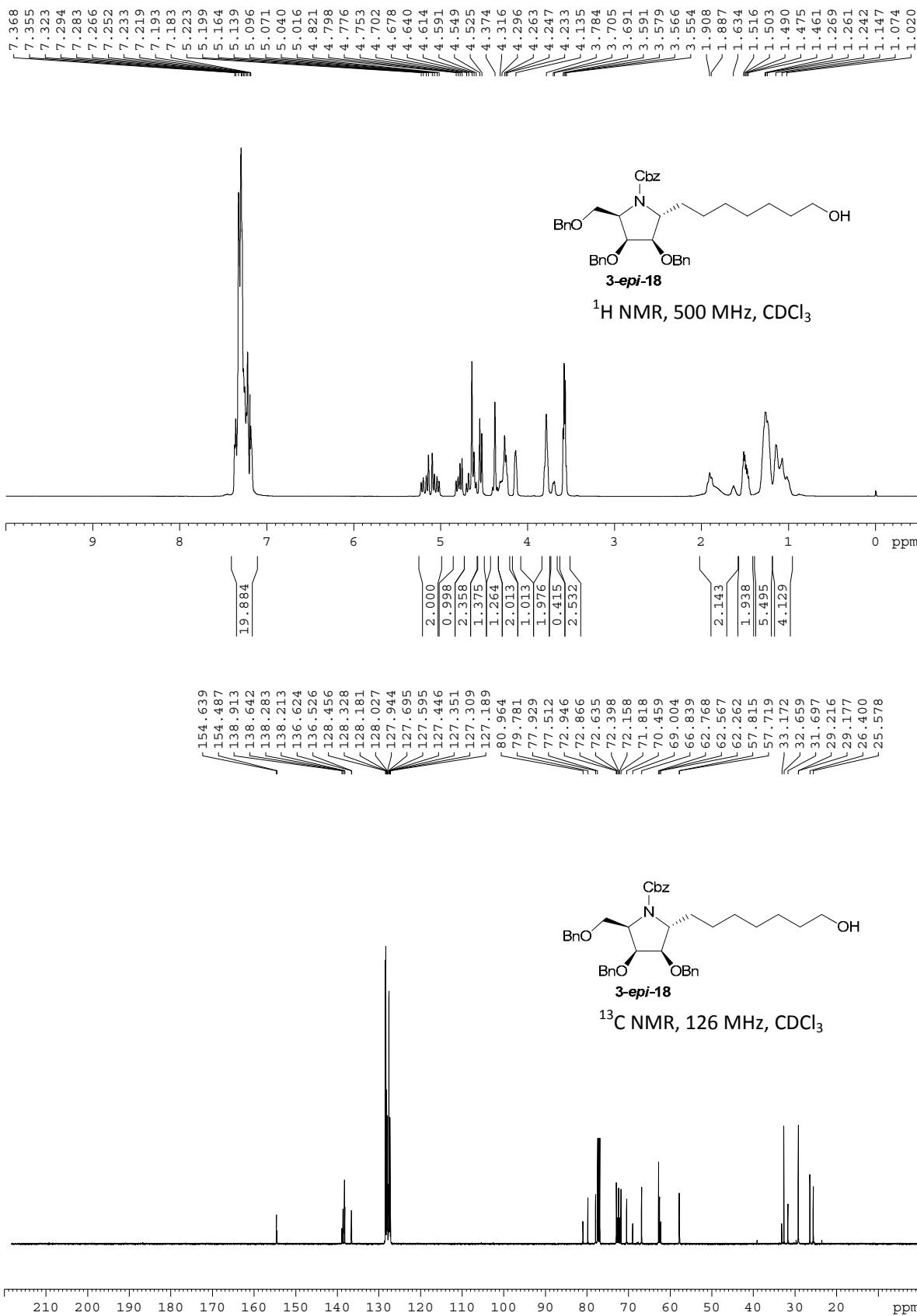


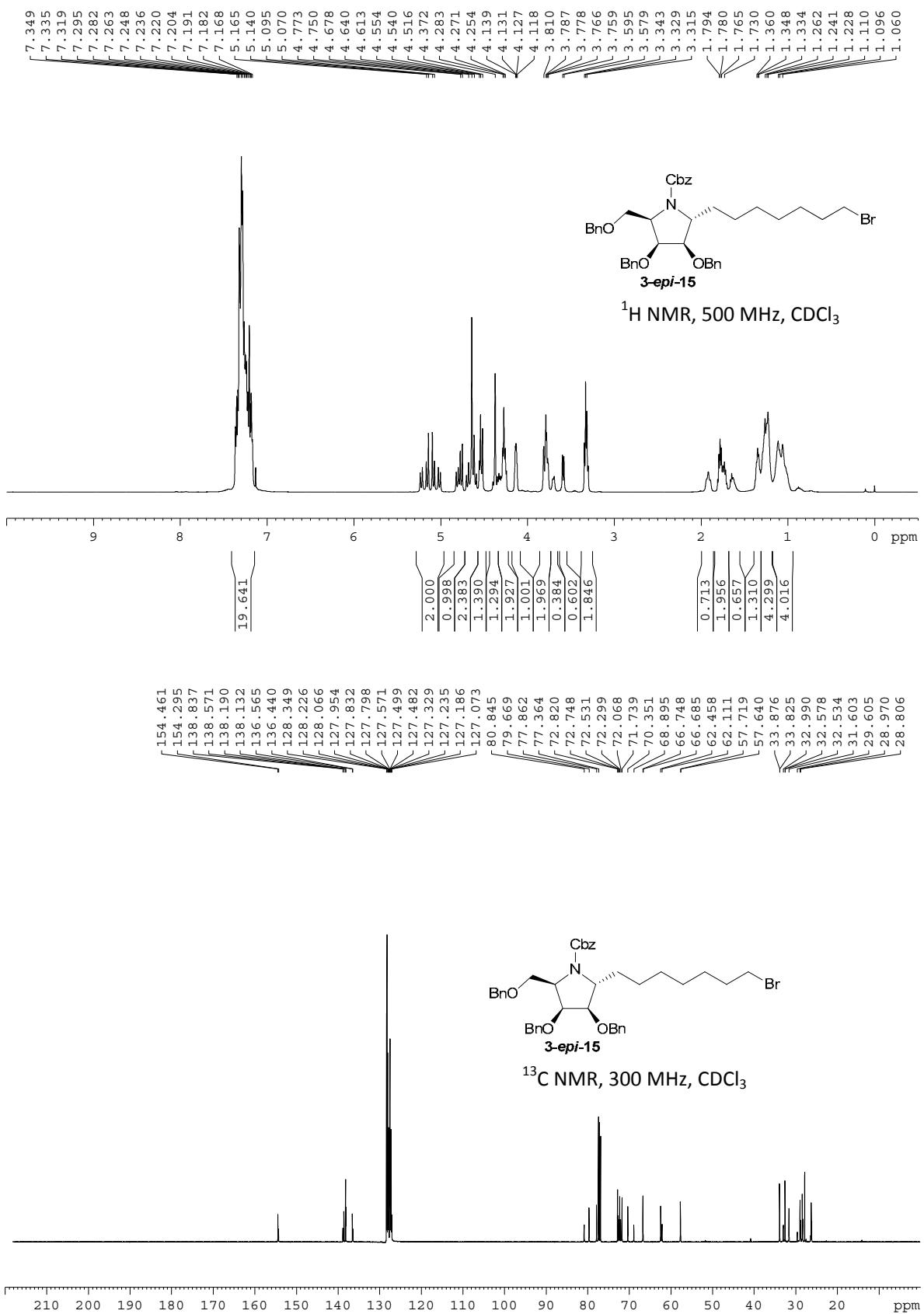


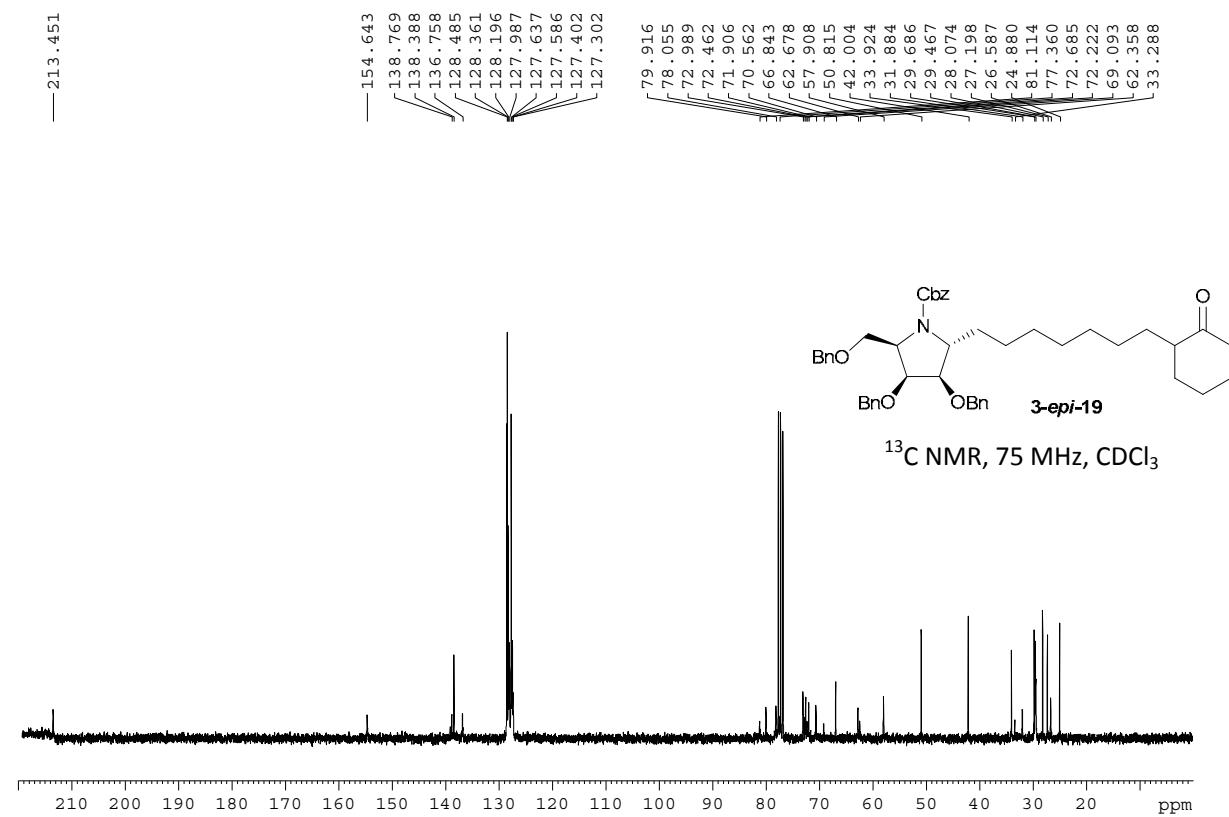
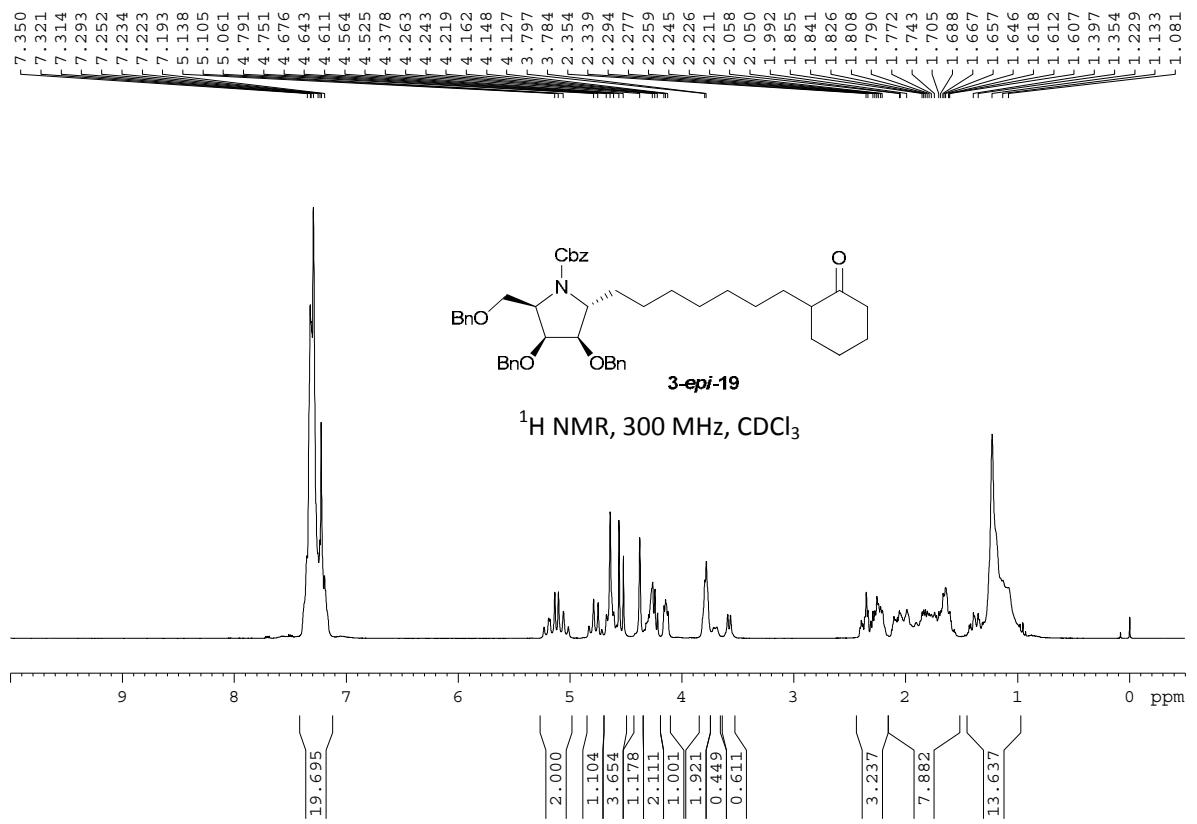


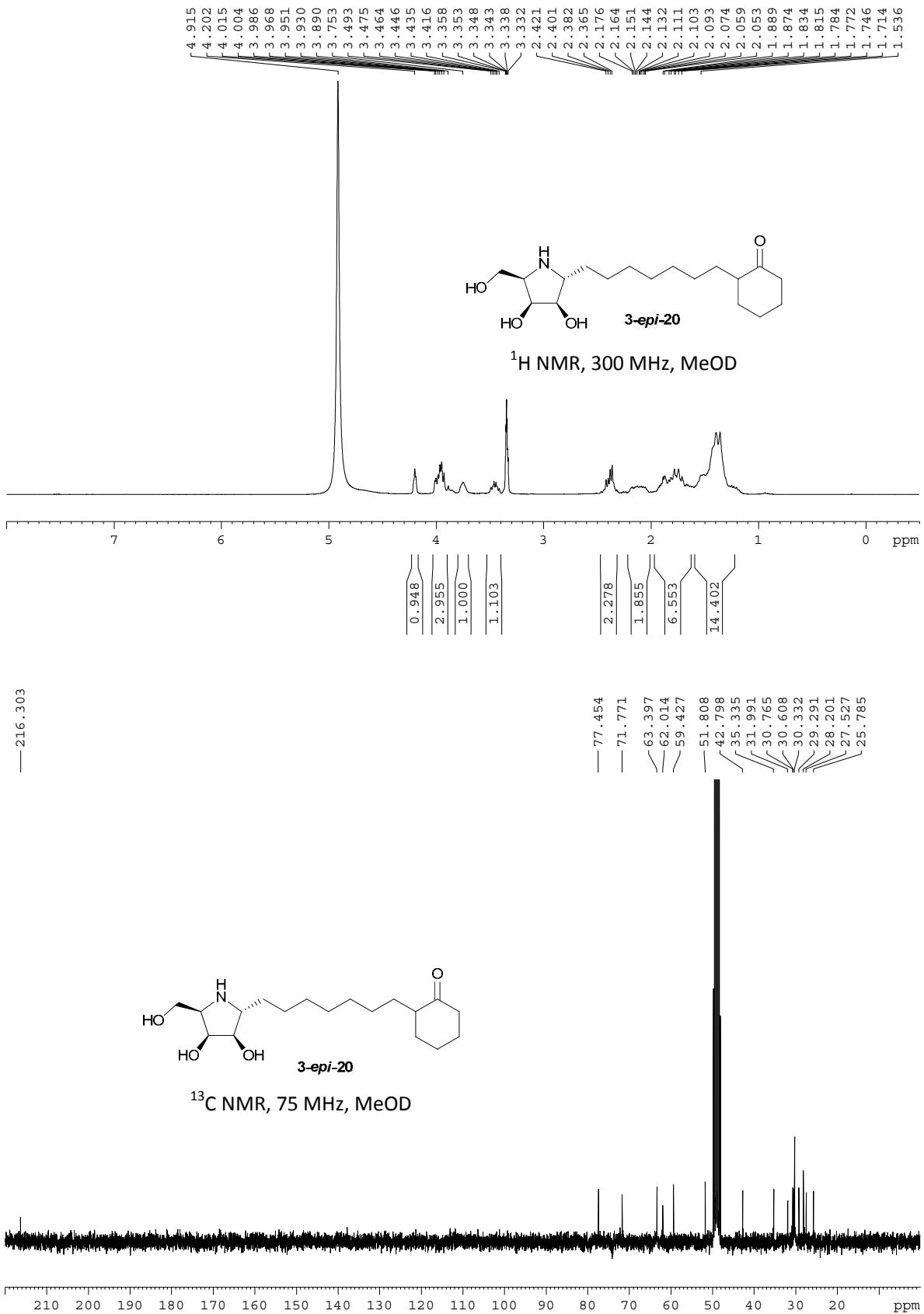


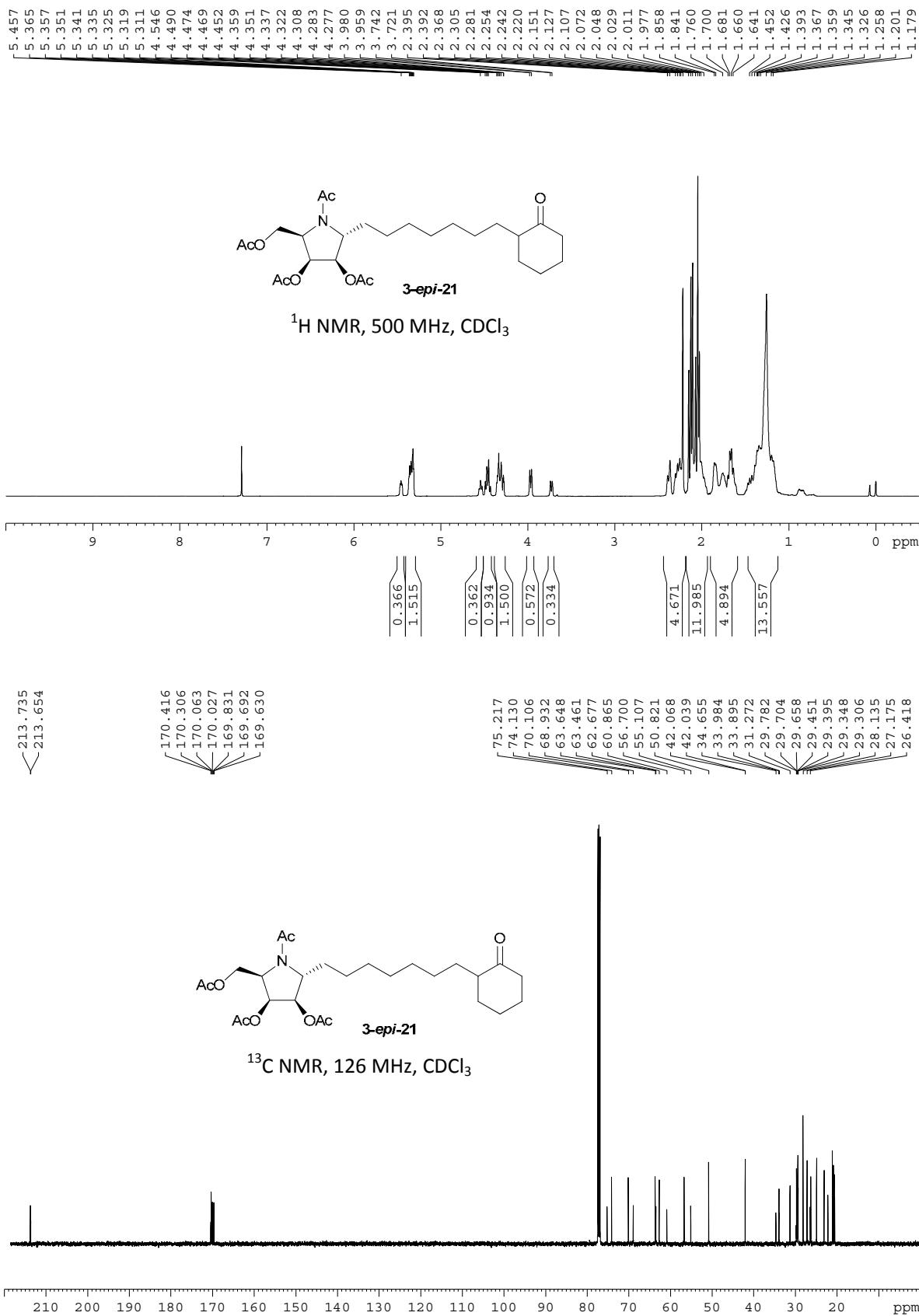


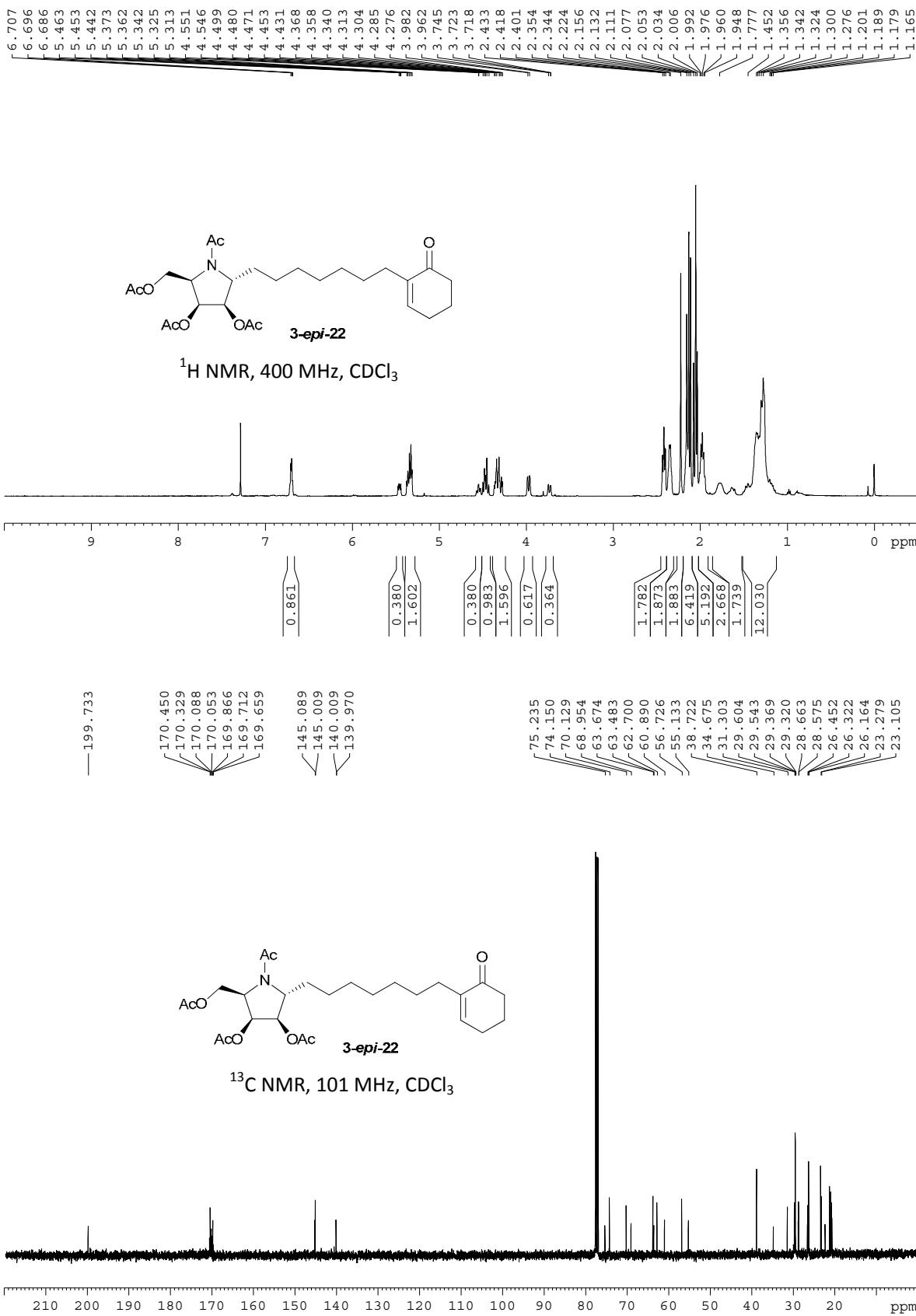


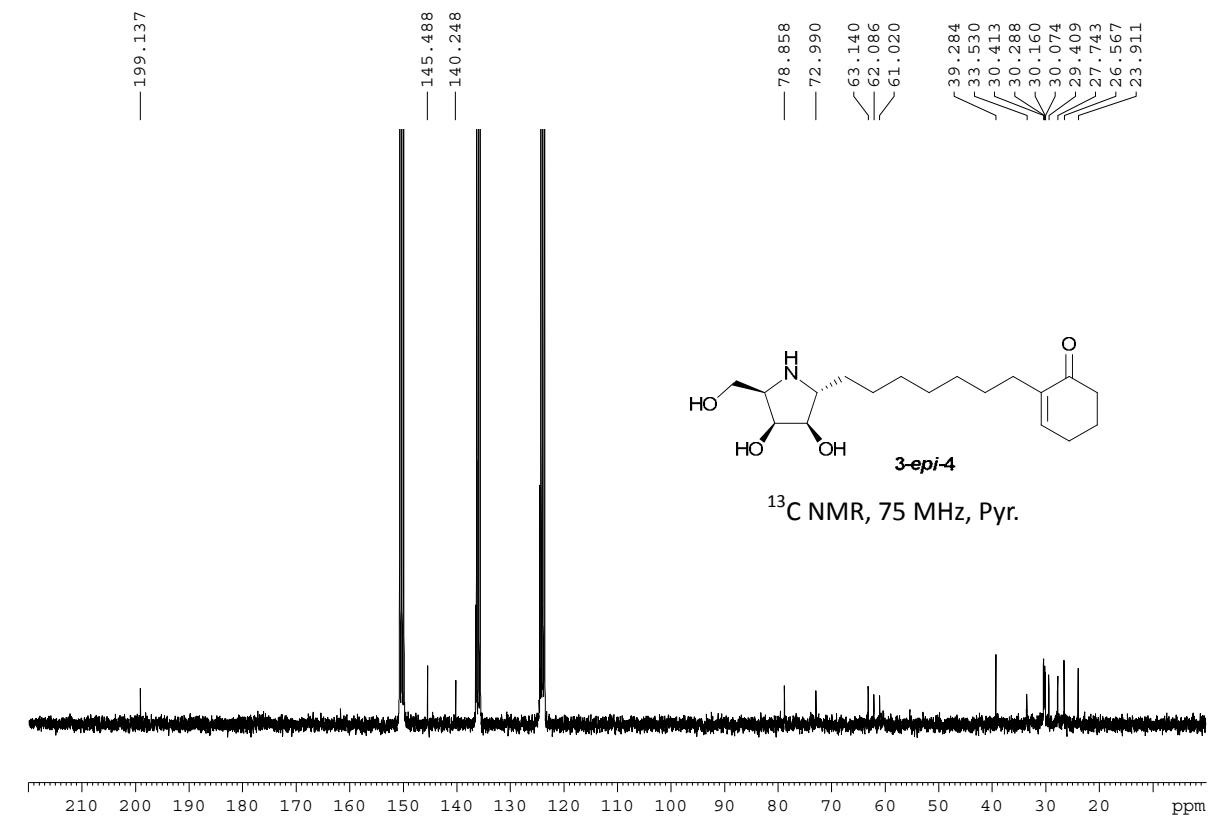
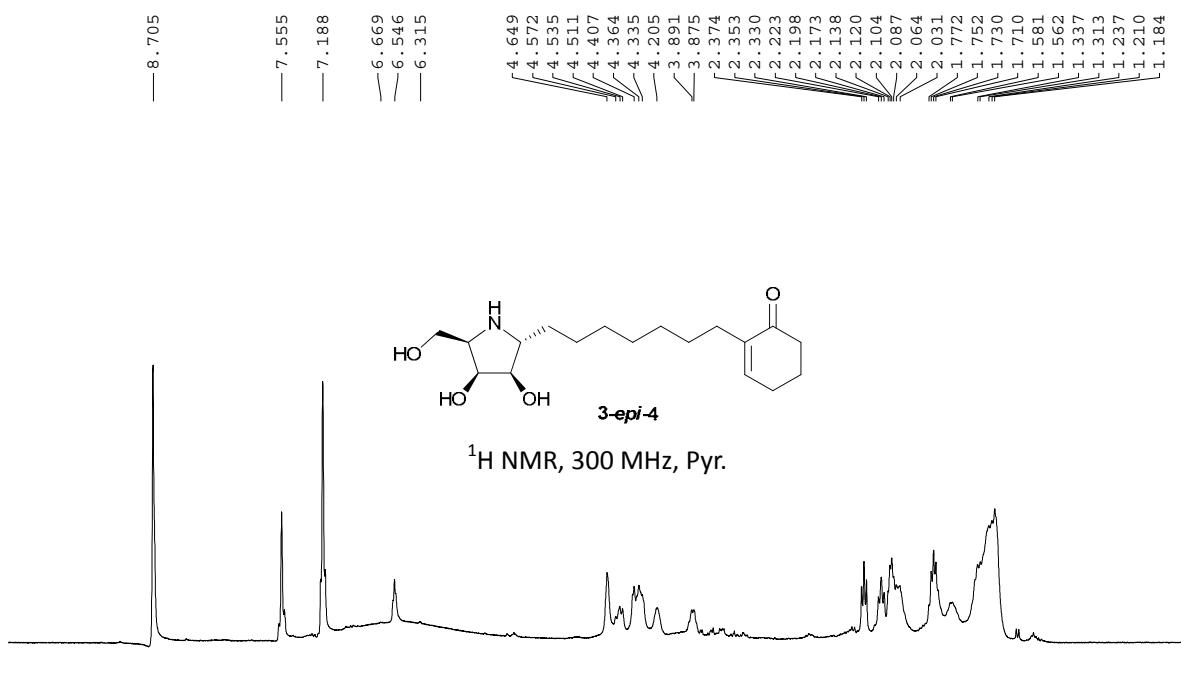


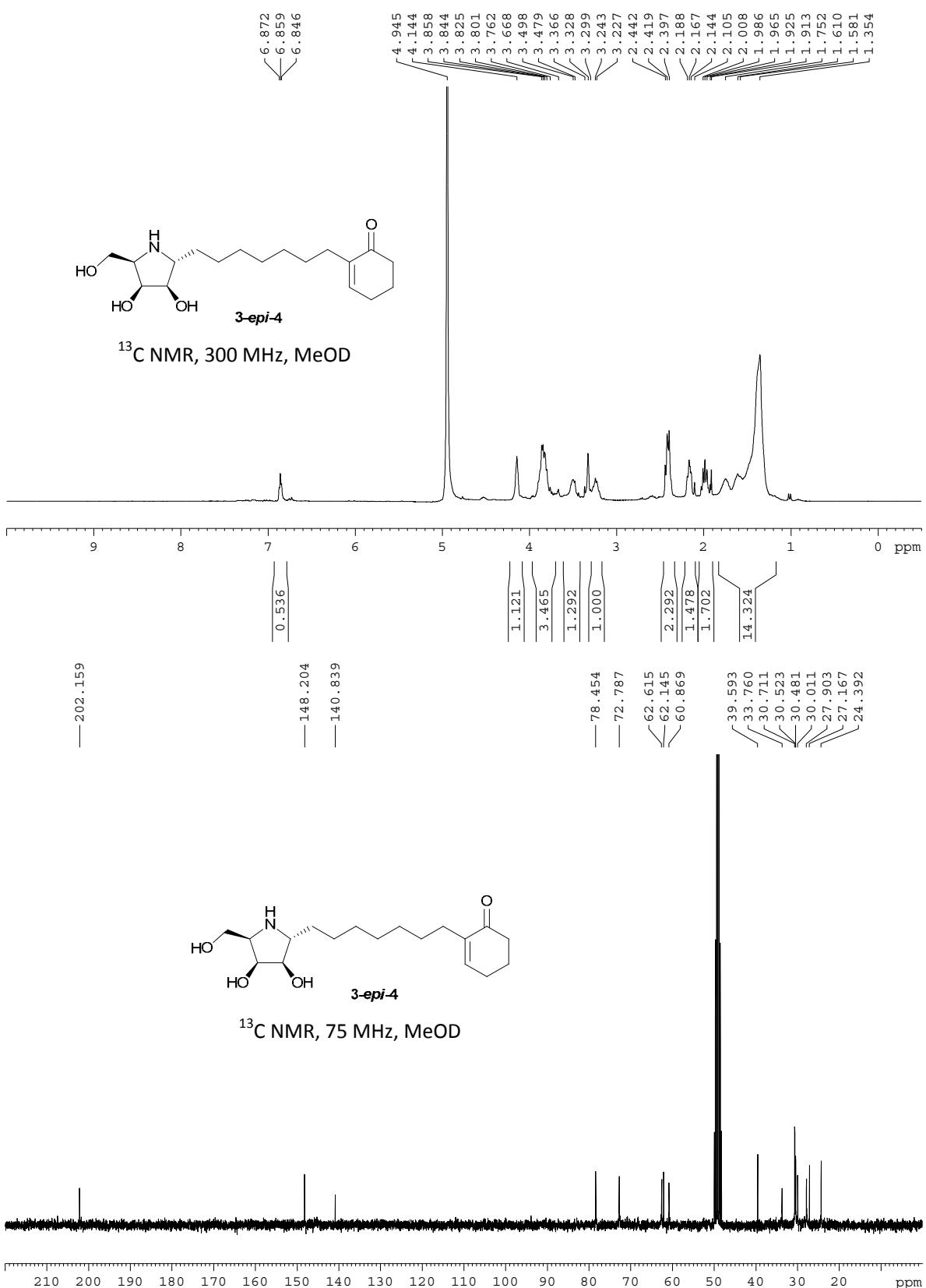


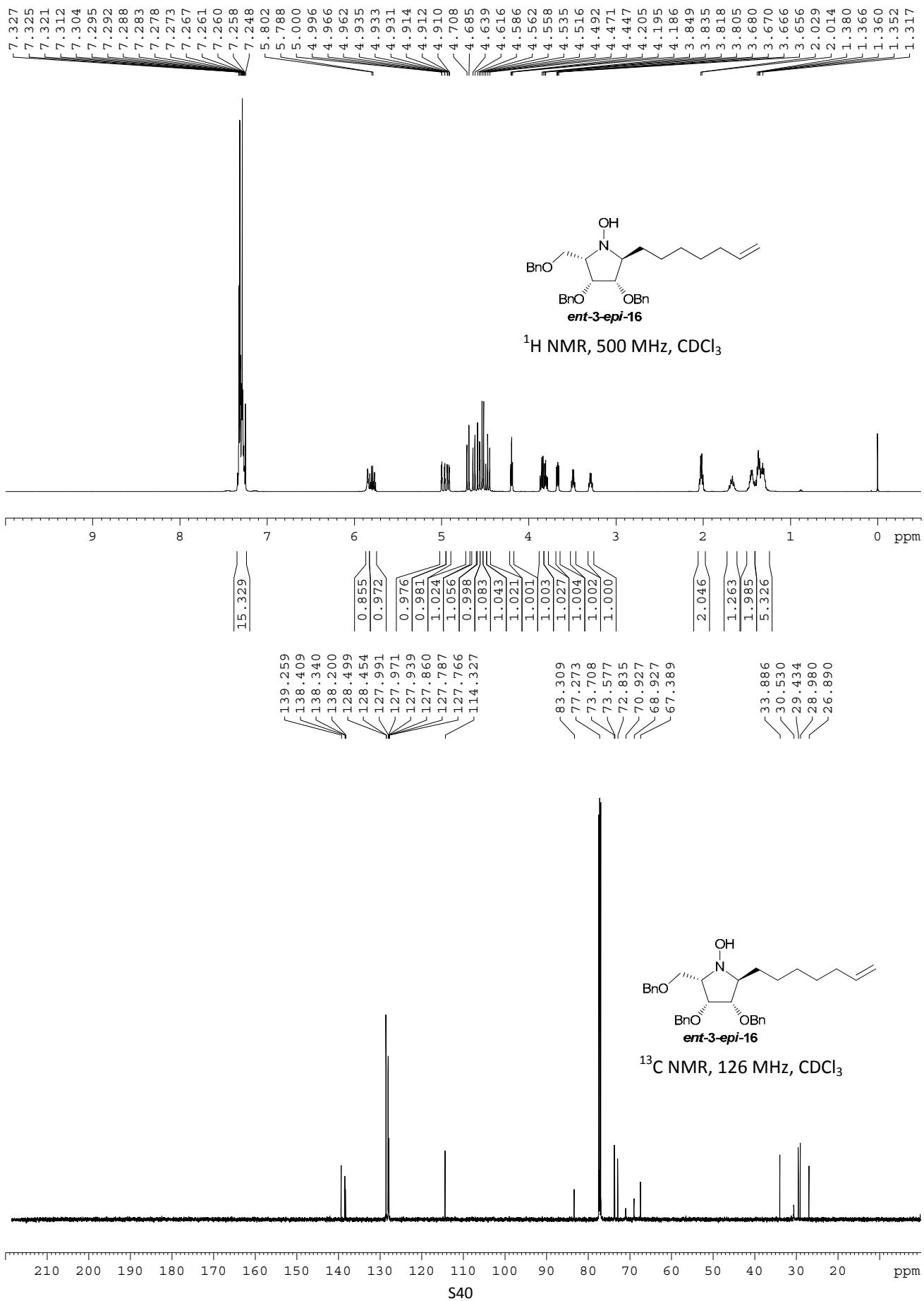


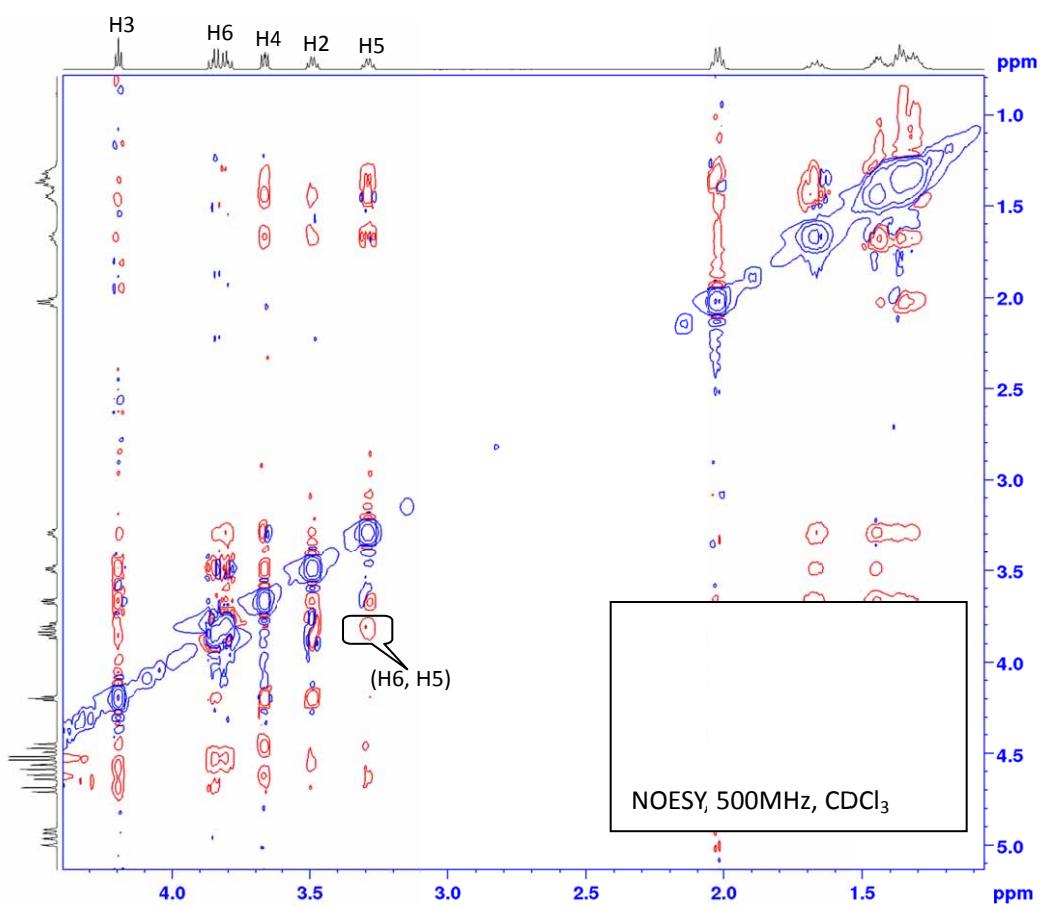
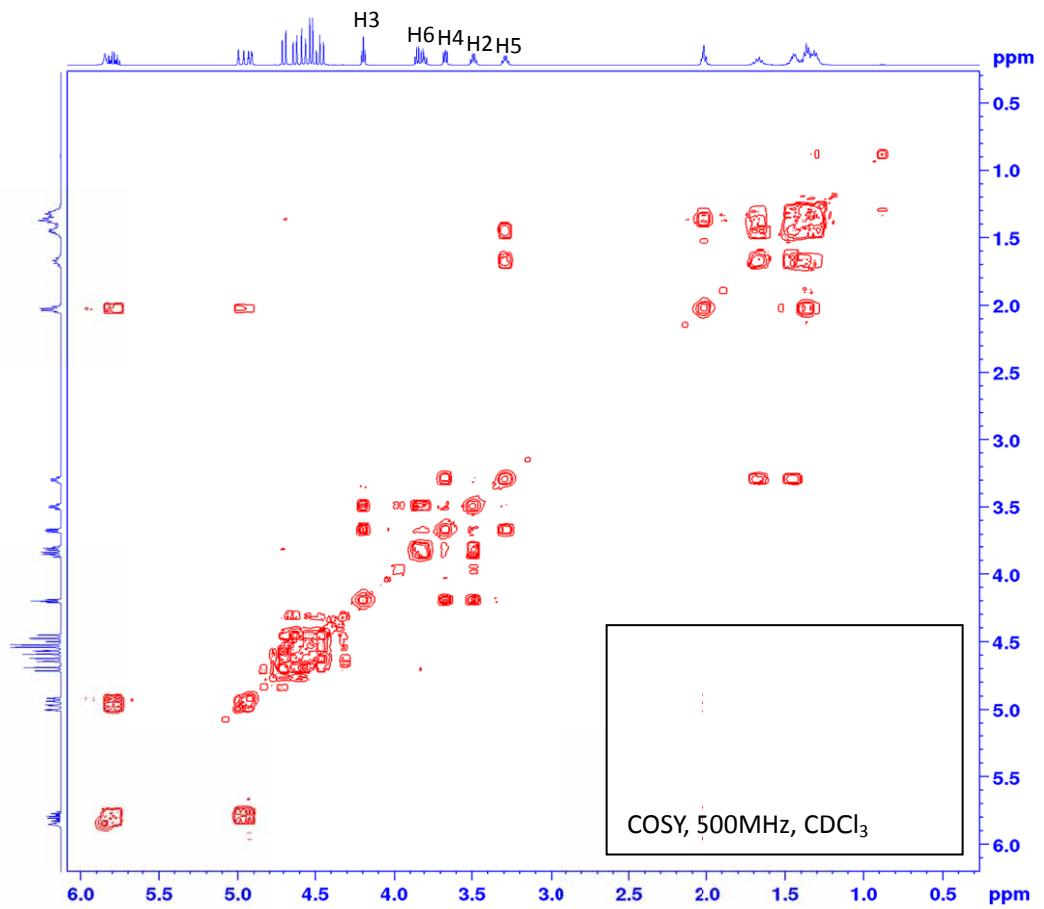


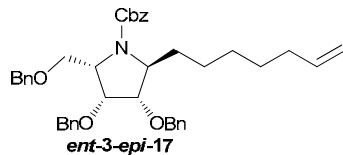
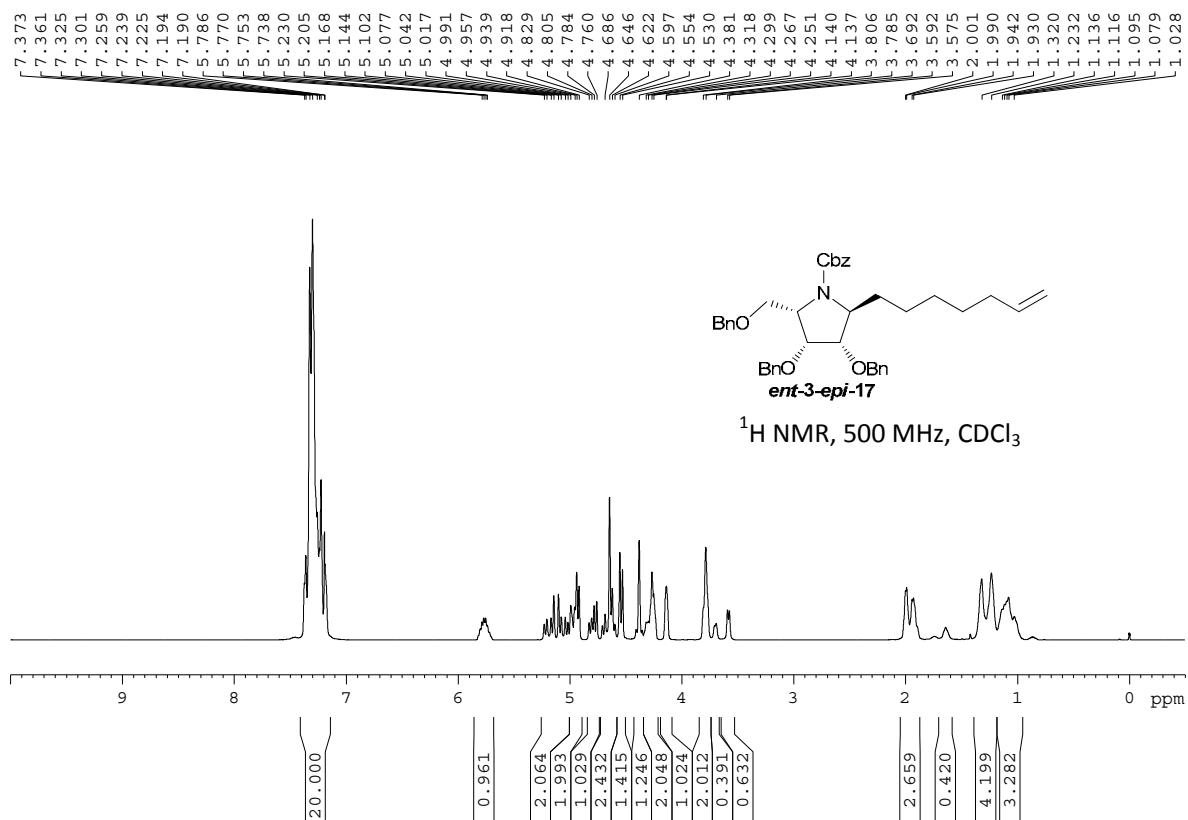




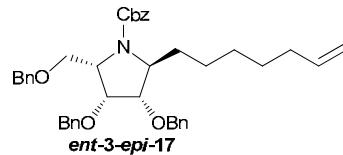
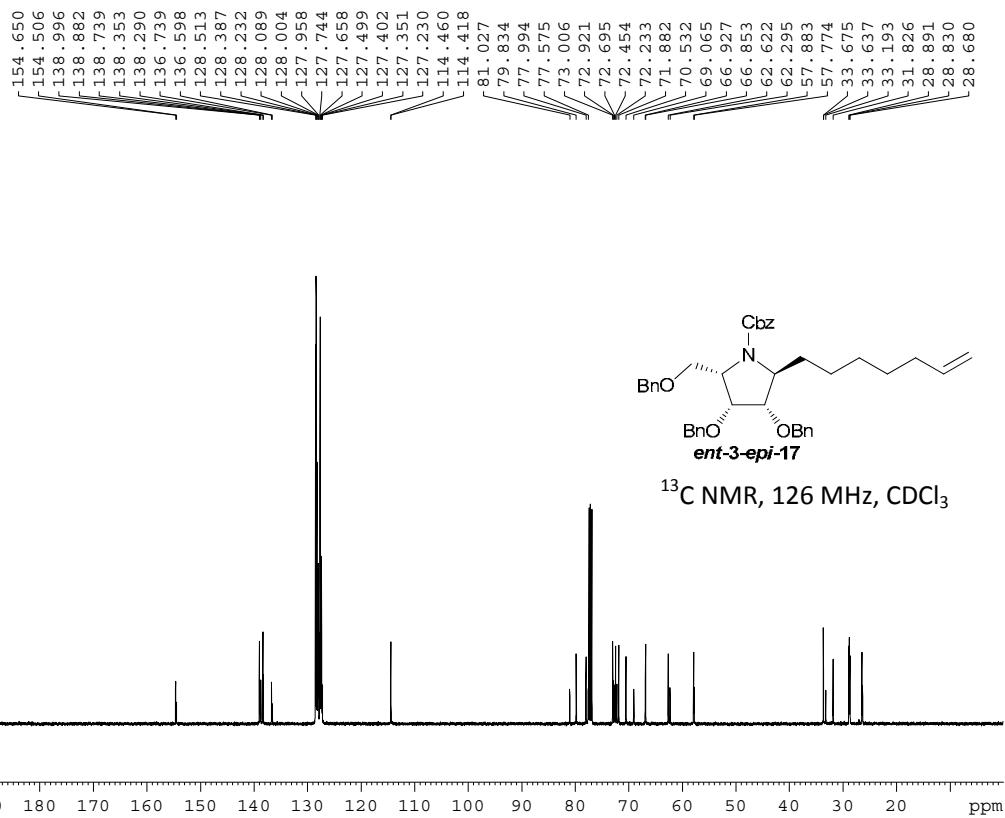




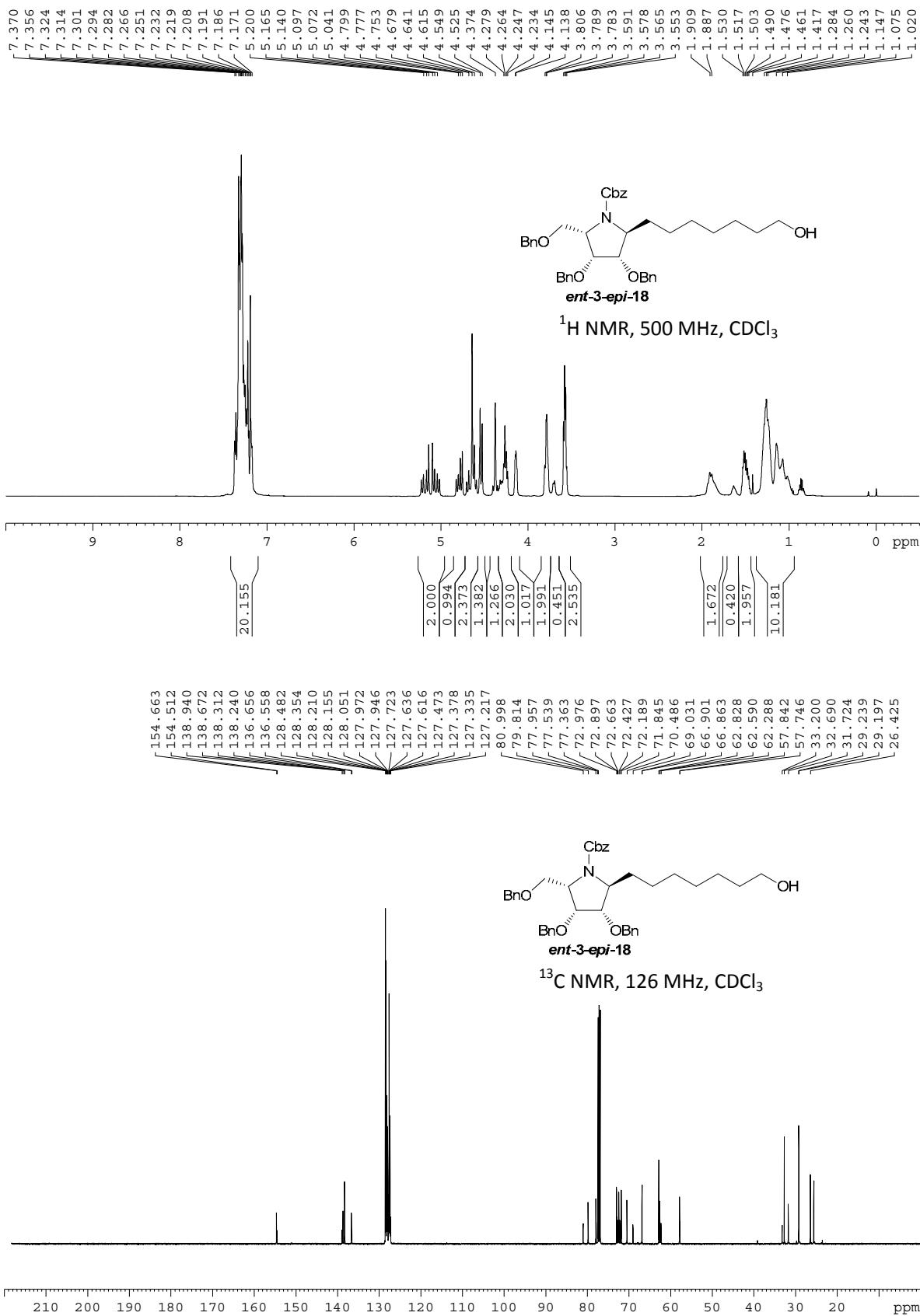


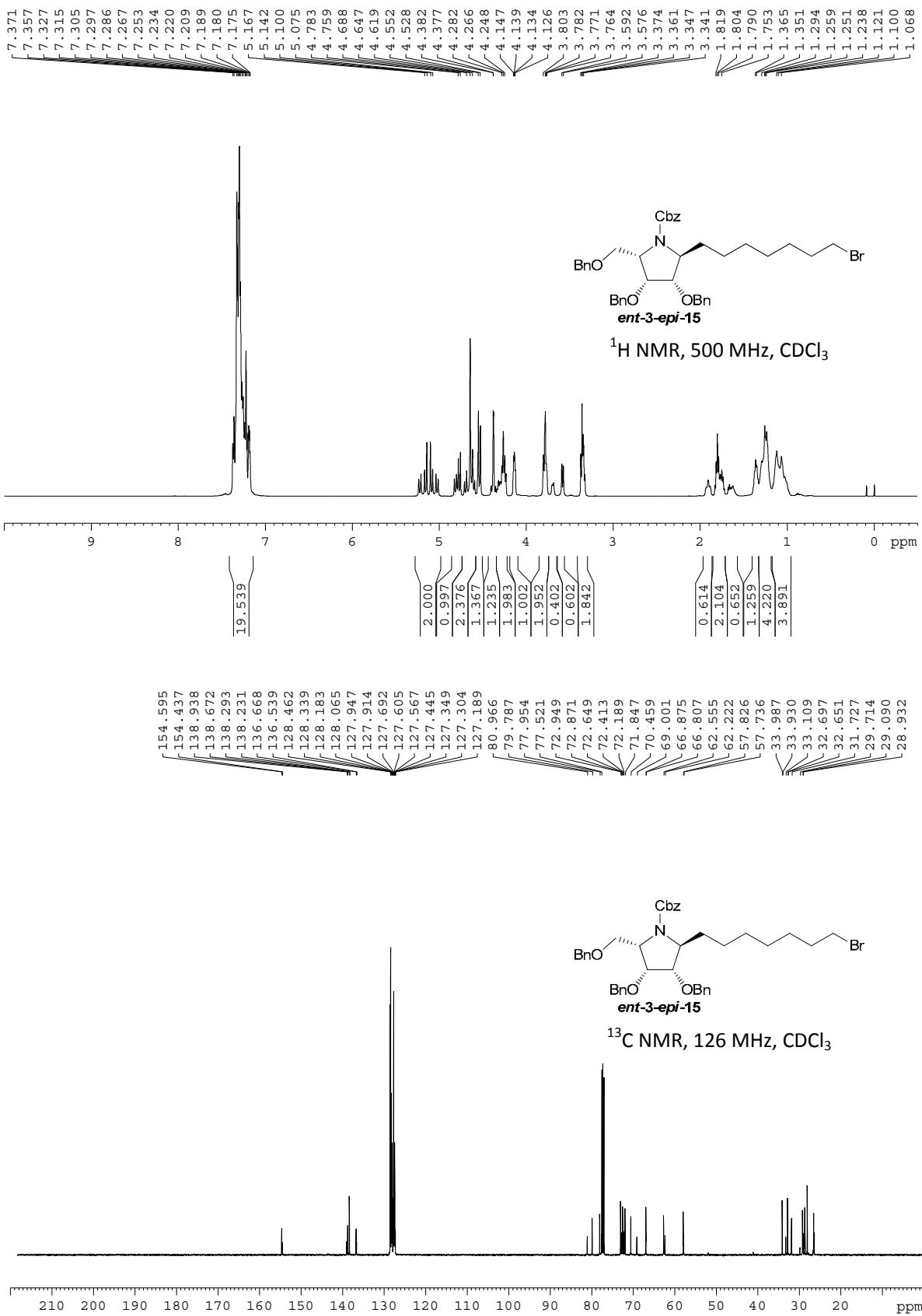


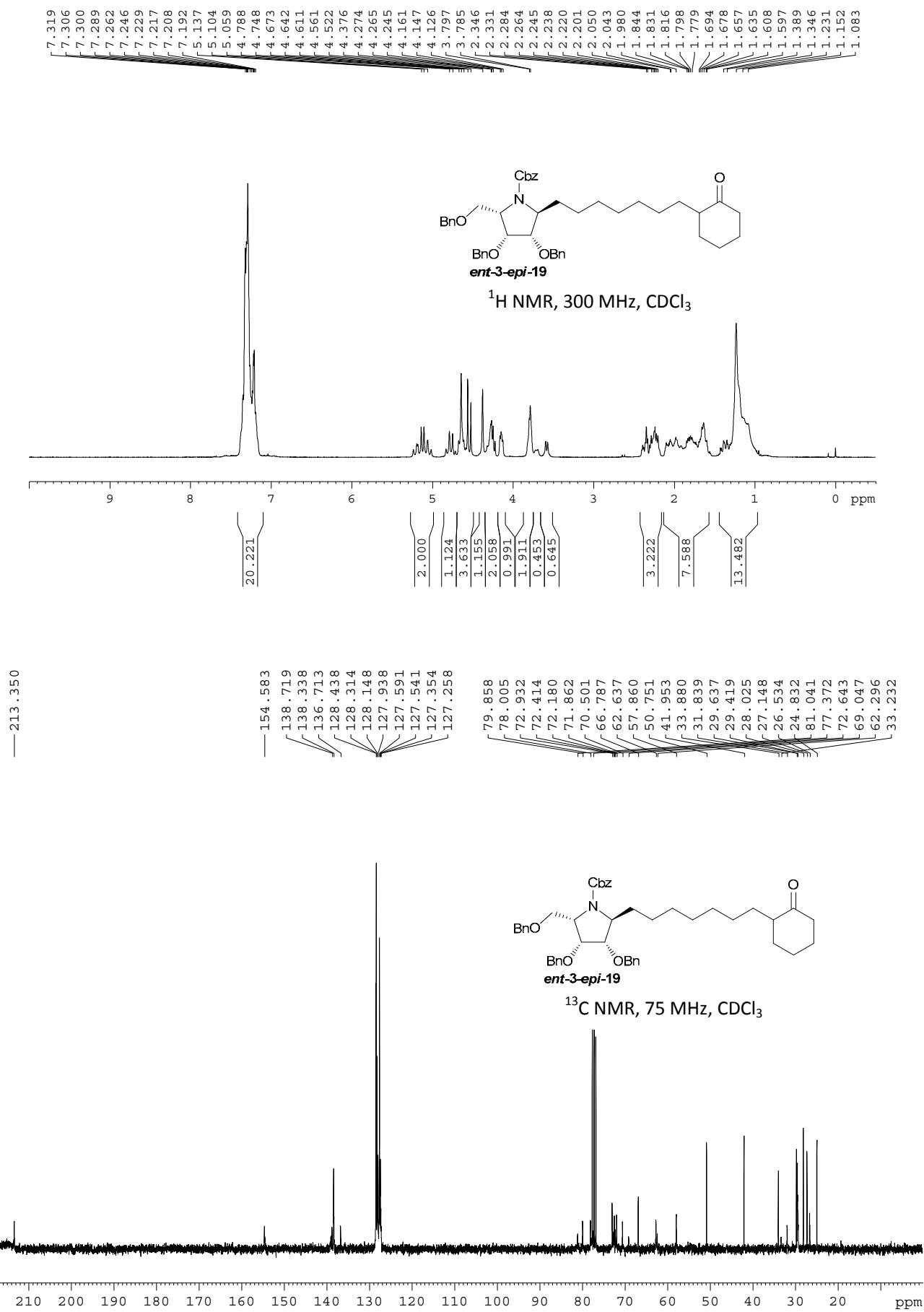
<sup>1</sup>H NMR, 500 MHz, CDCl<sub>3</sub>

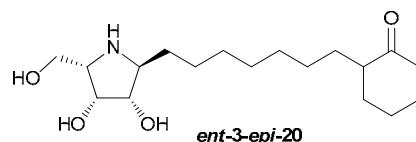
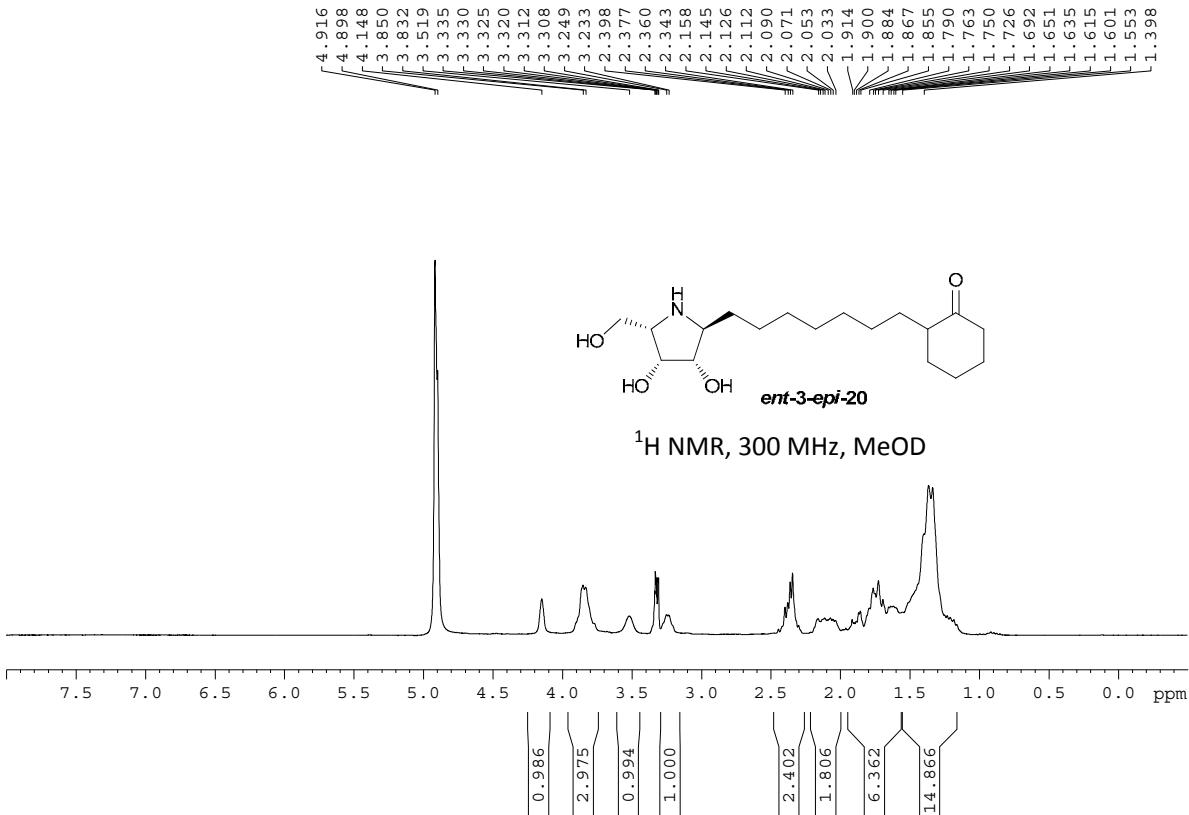


<sup>13</sup>C NMR, 126 MHz, CDCl<sub>3</sub>

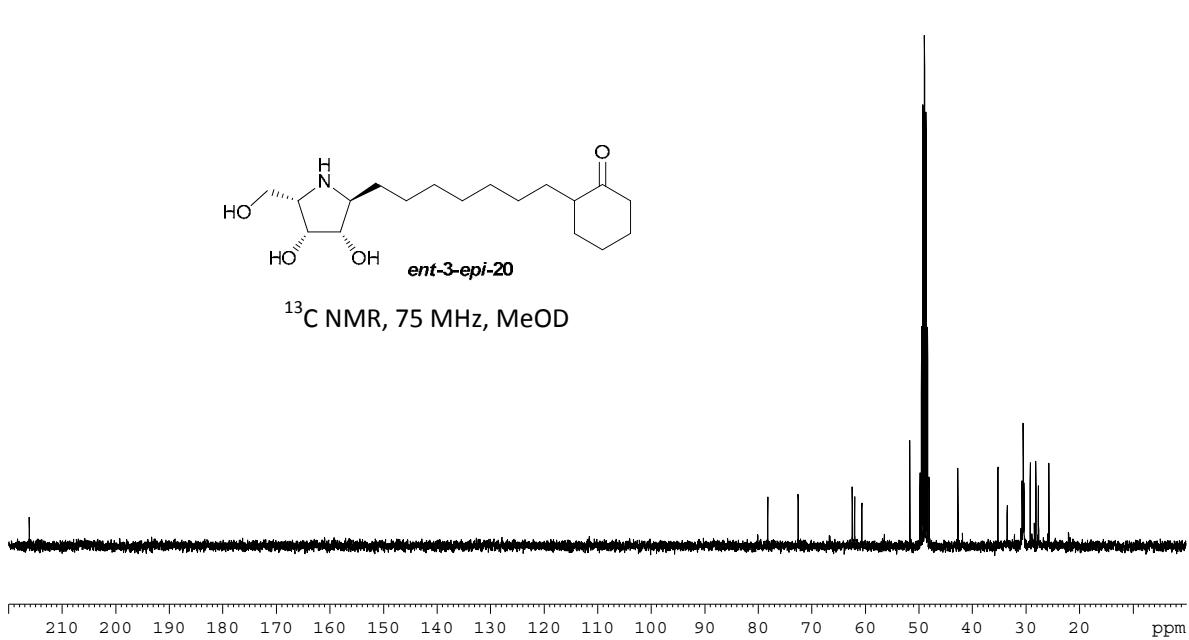


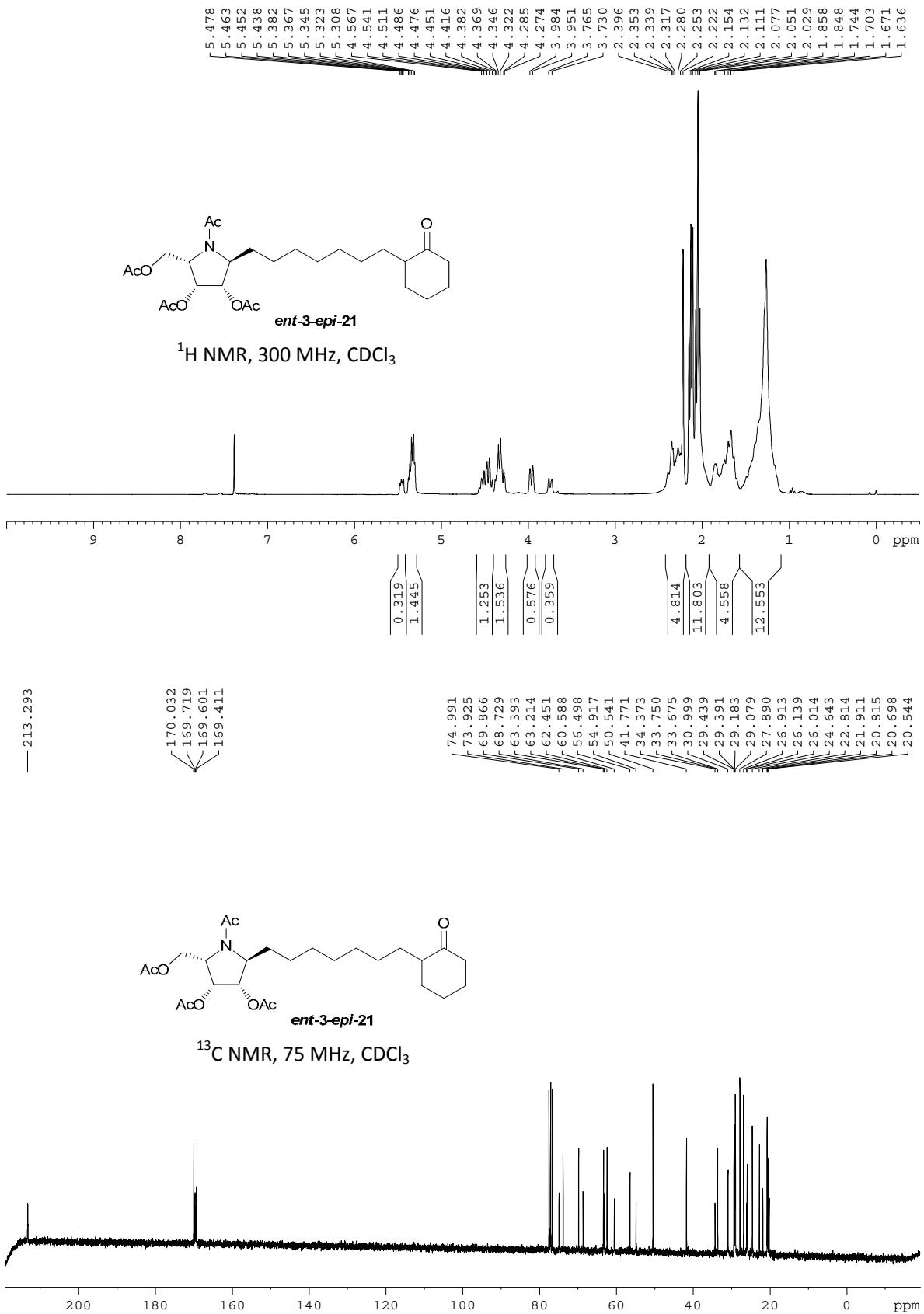


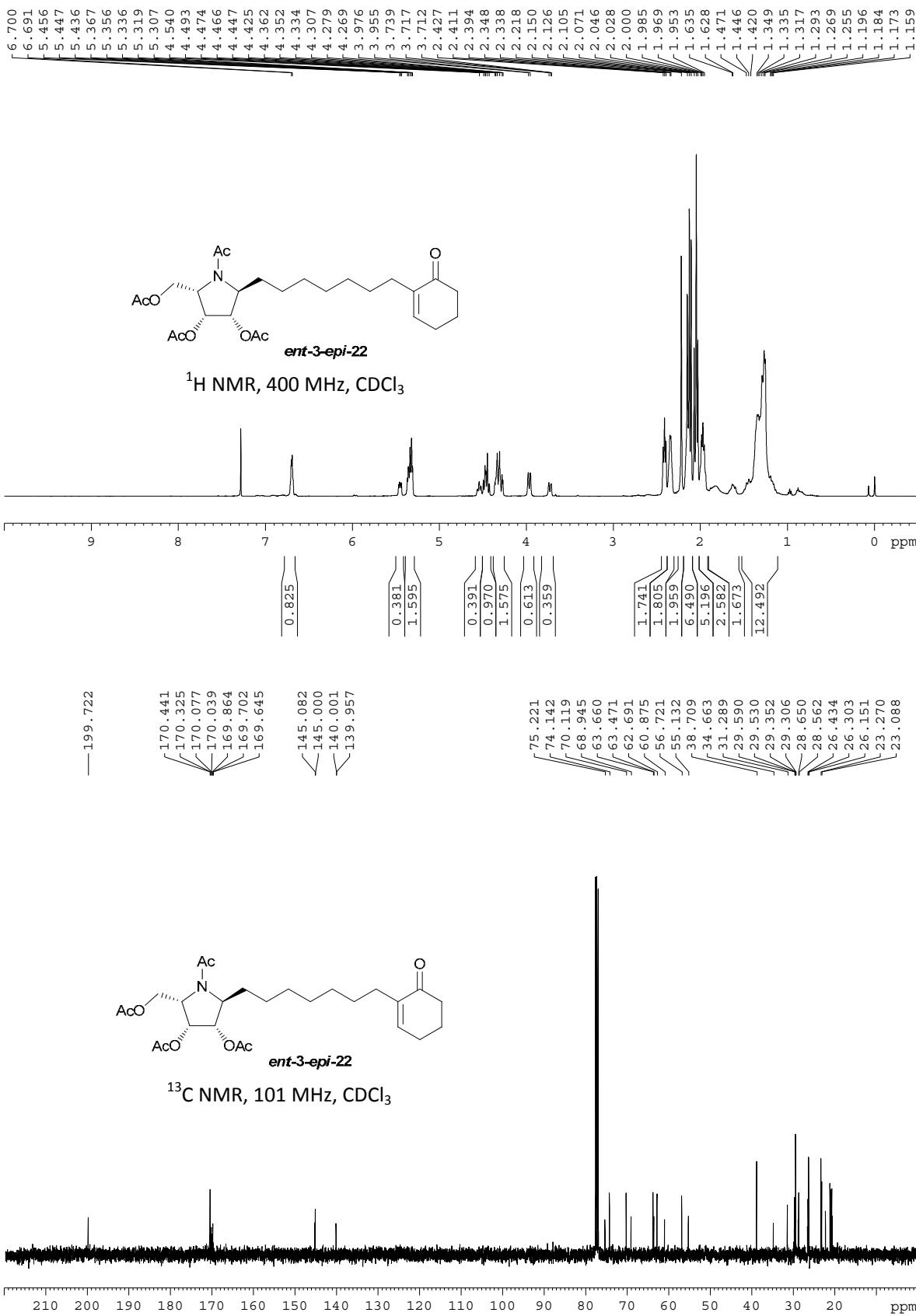


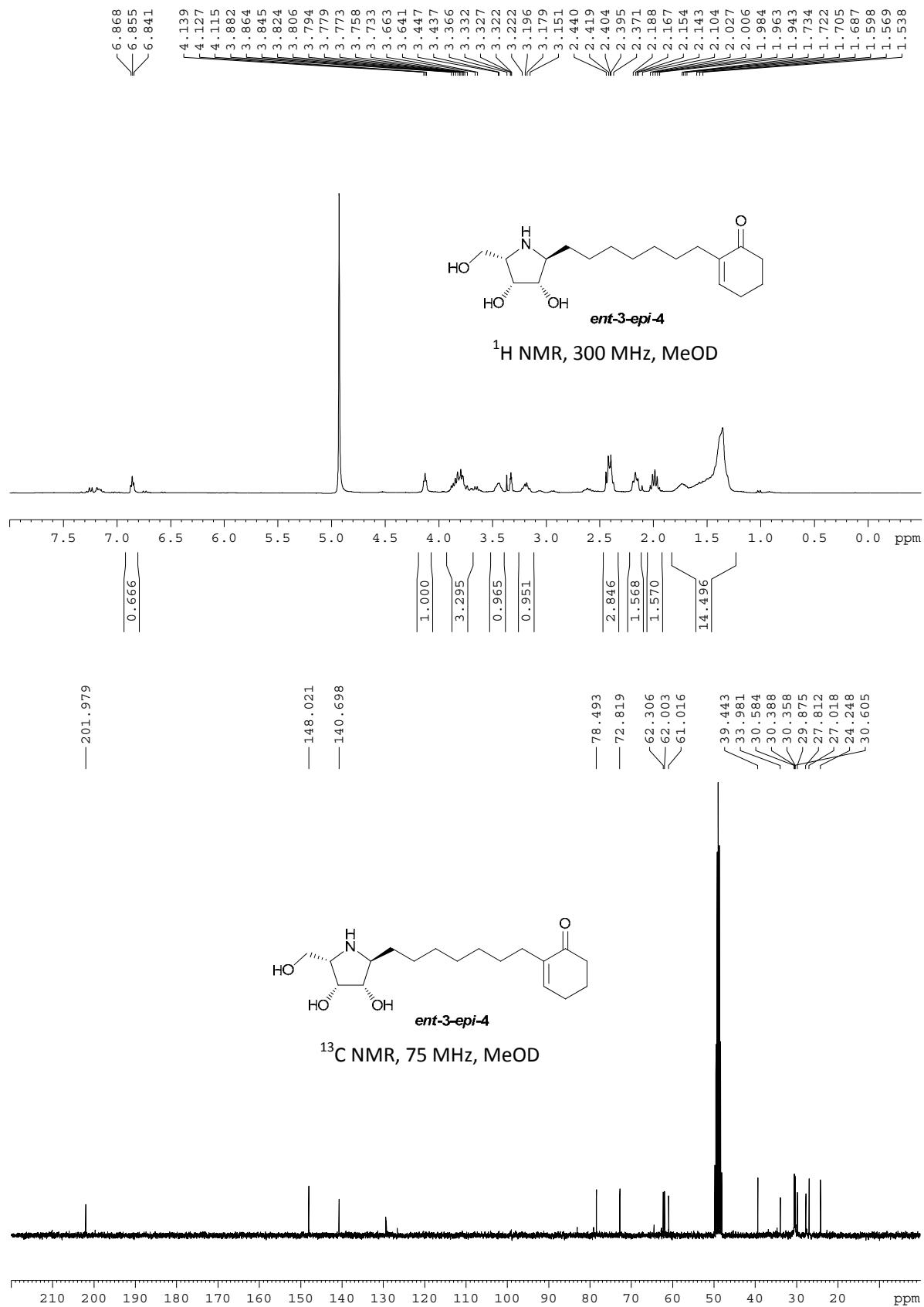


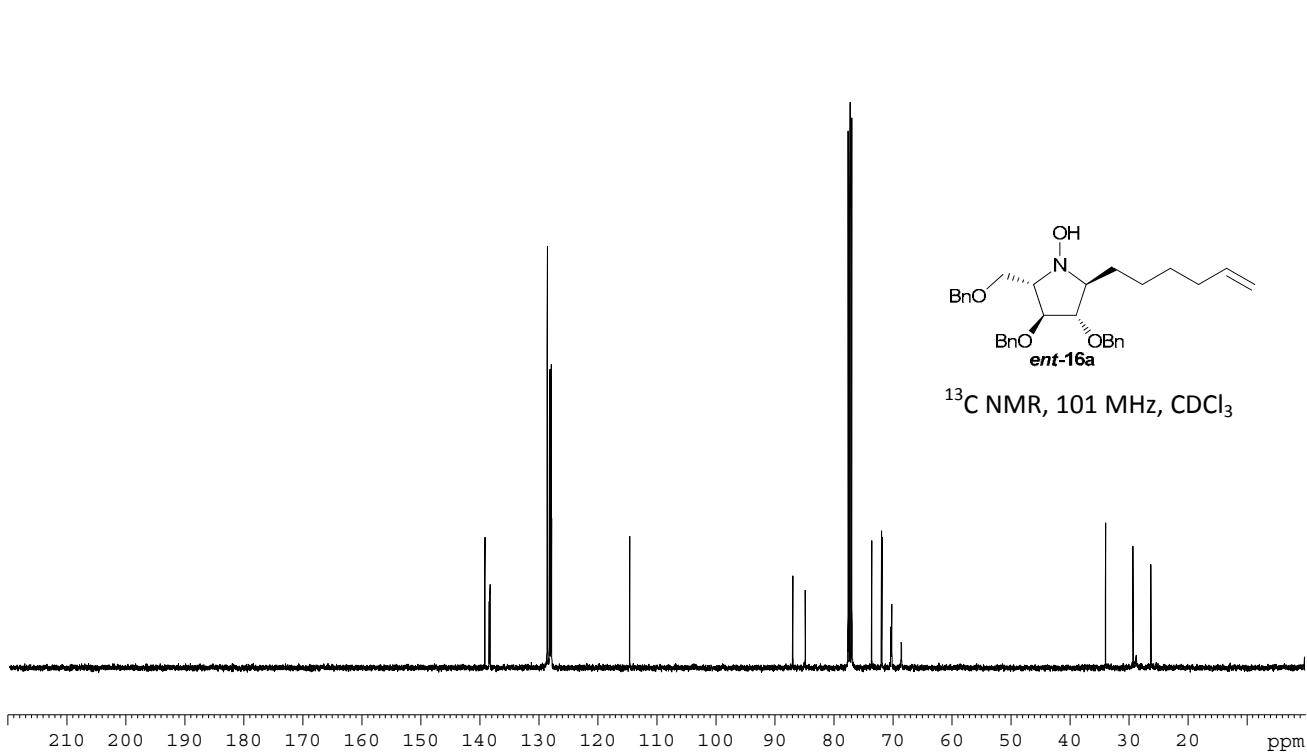
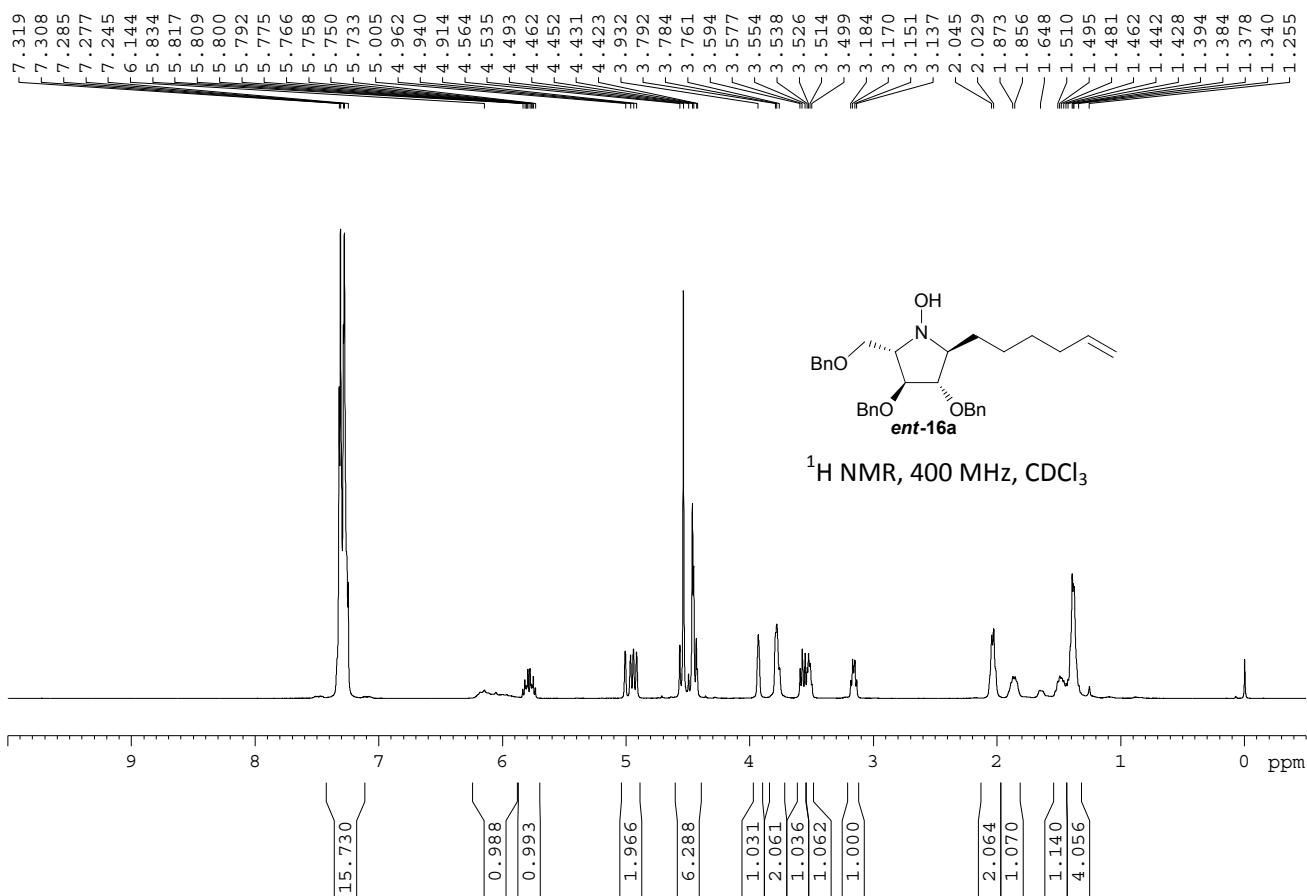
<sup>13</sup>C NMR, 75 MHz, MeOD

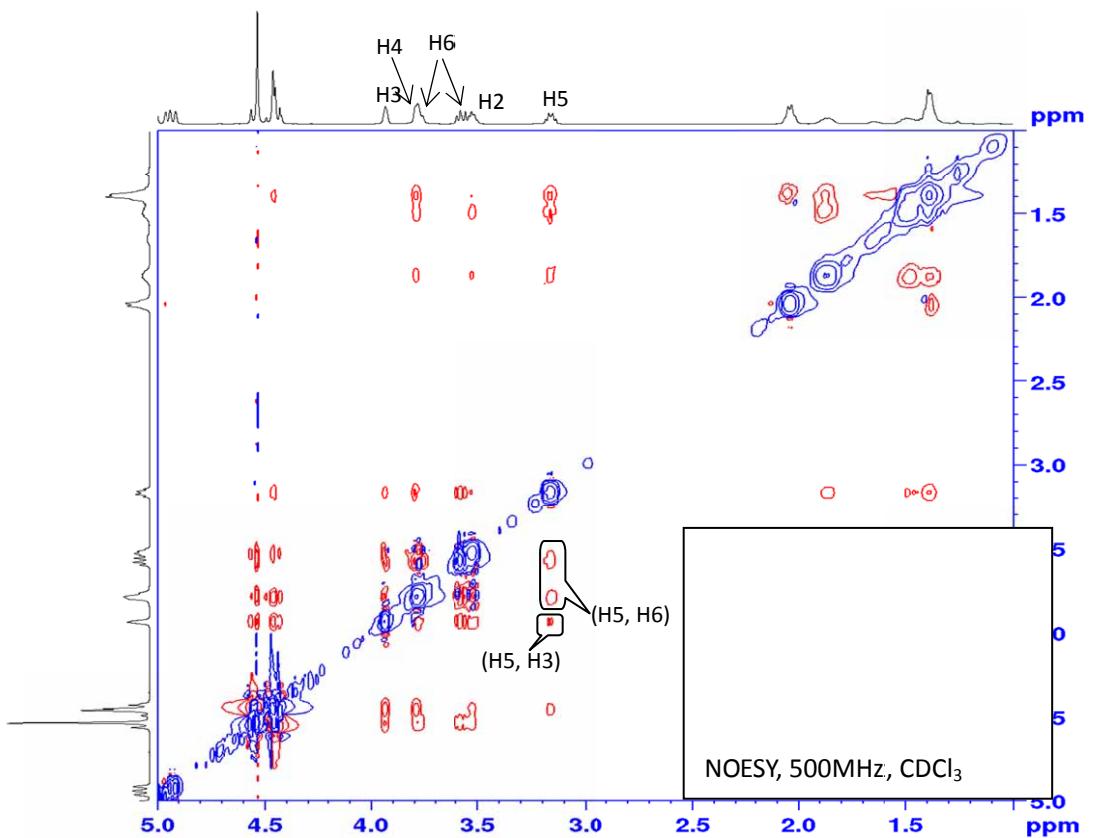
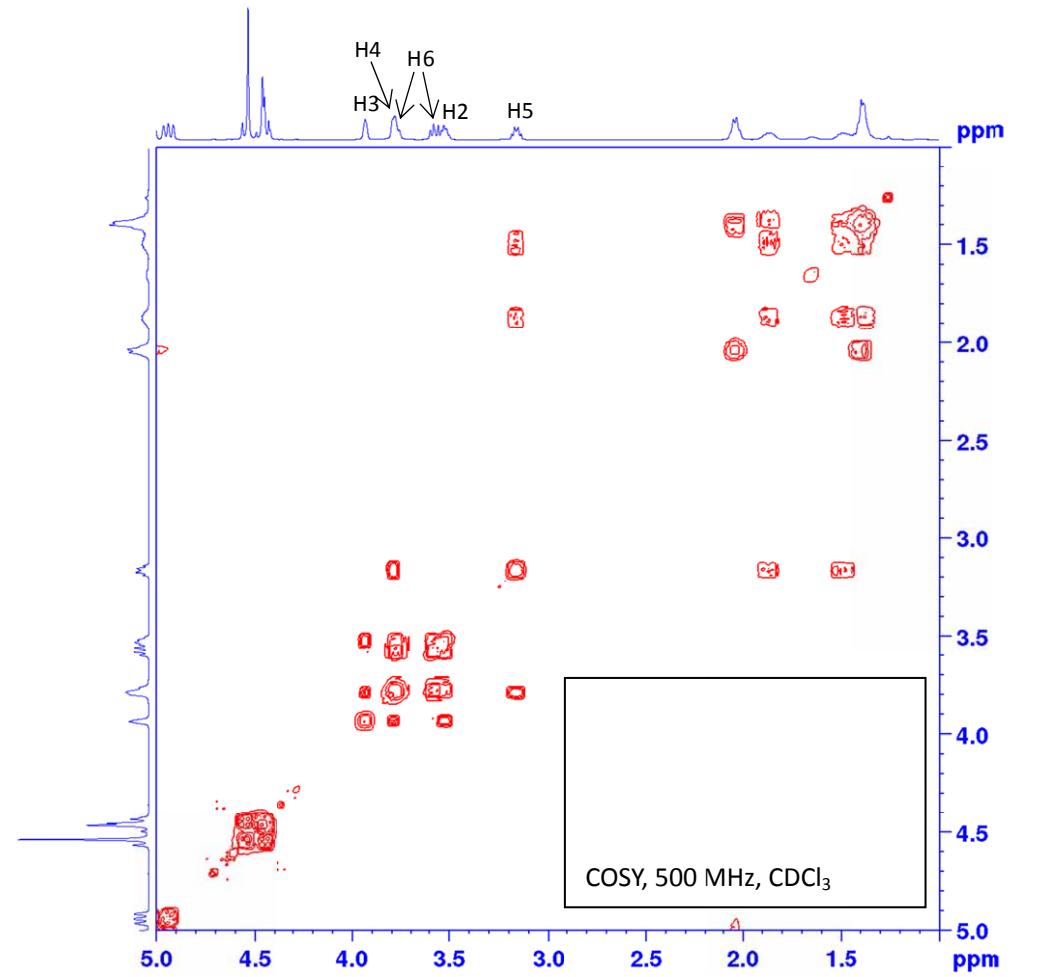


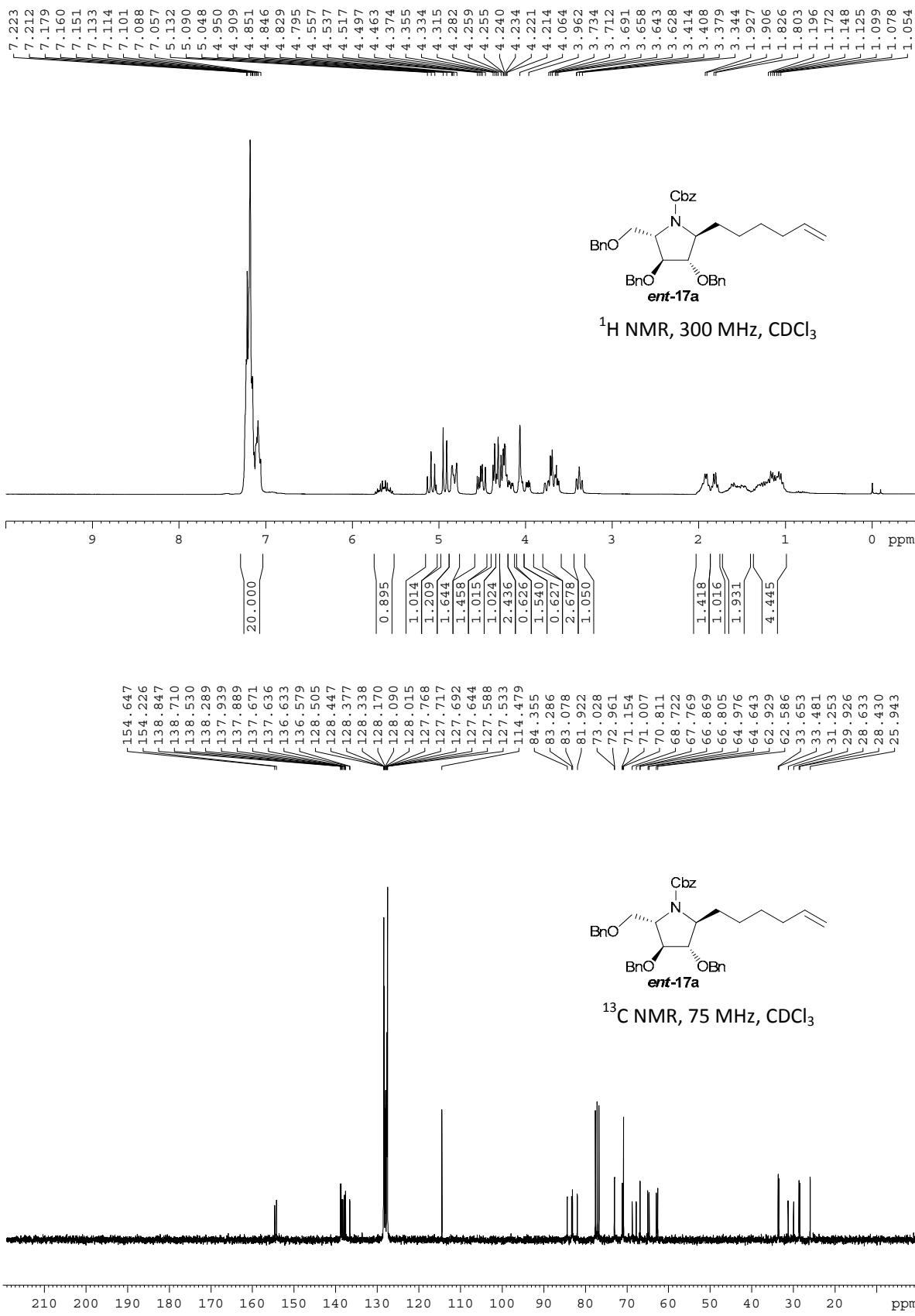


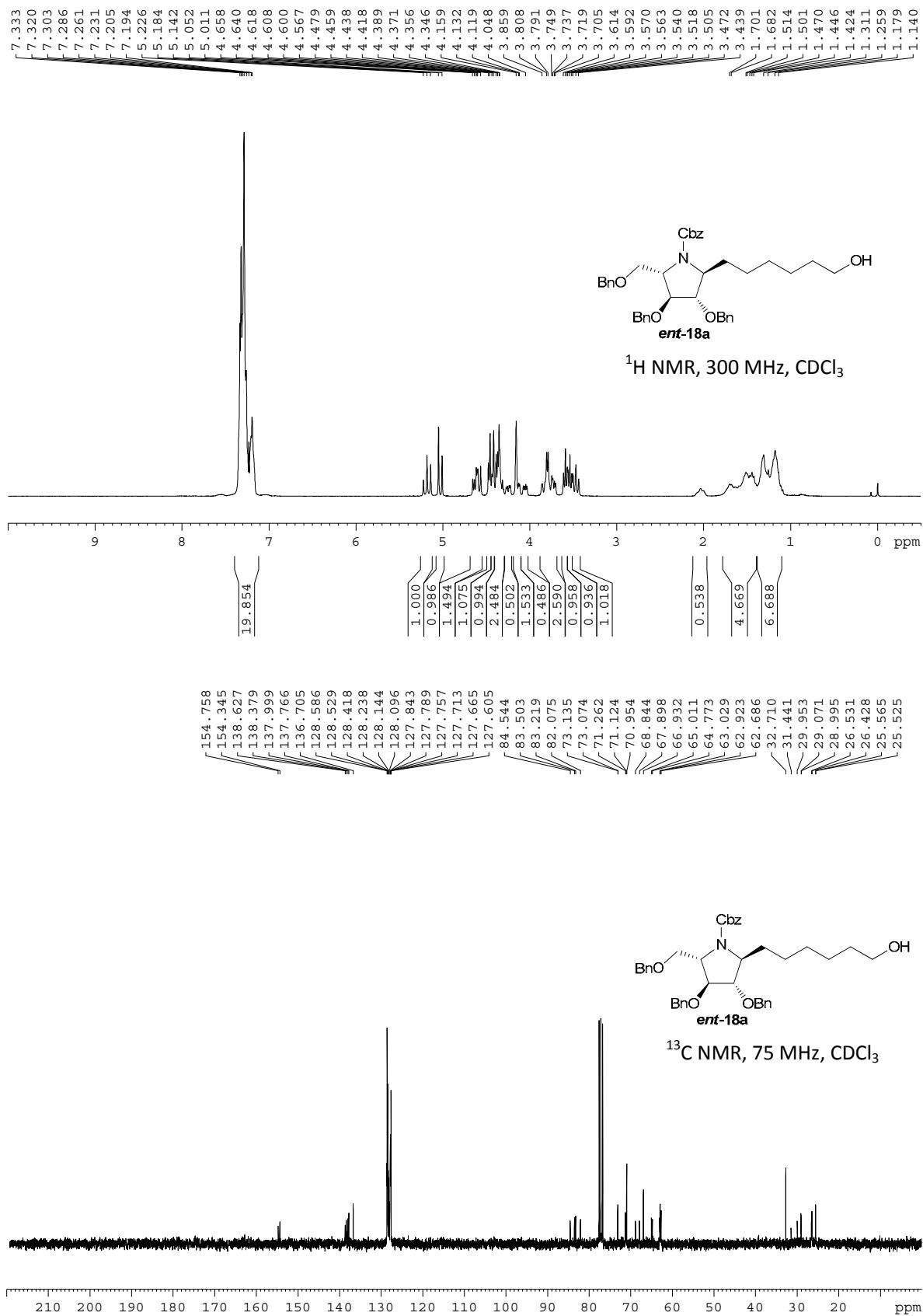


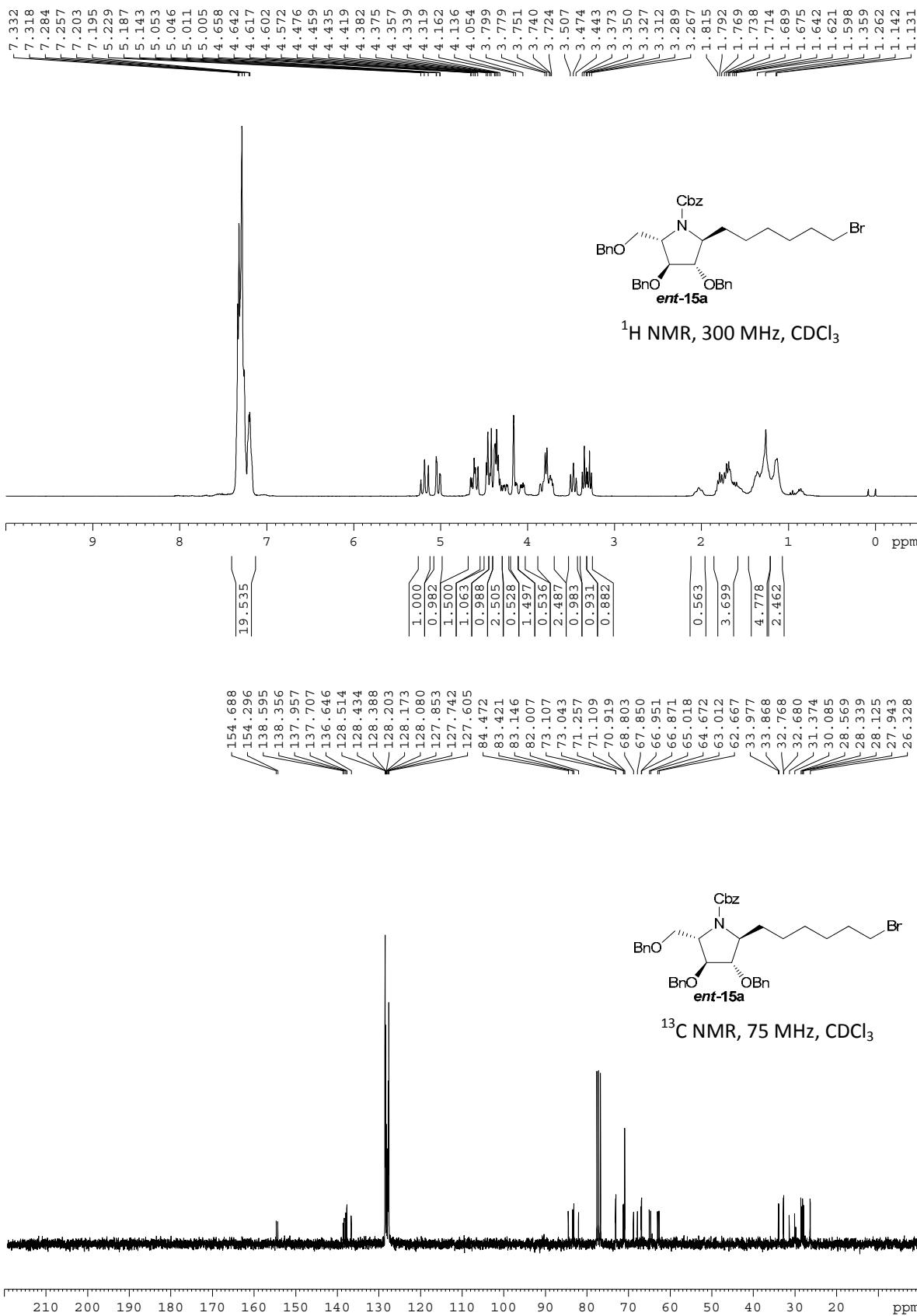


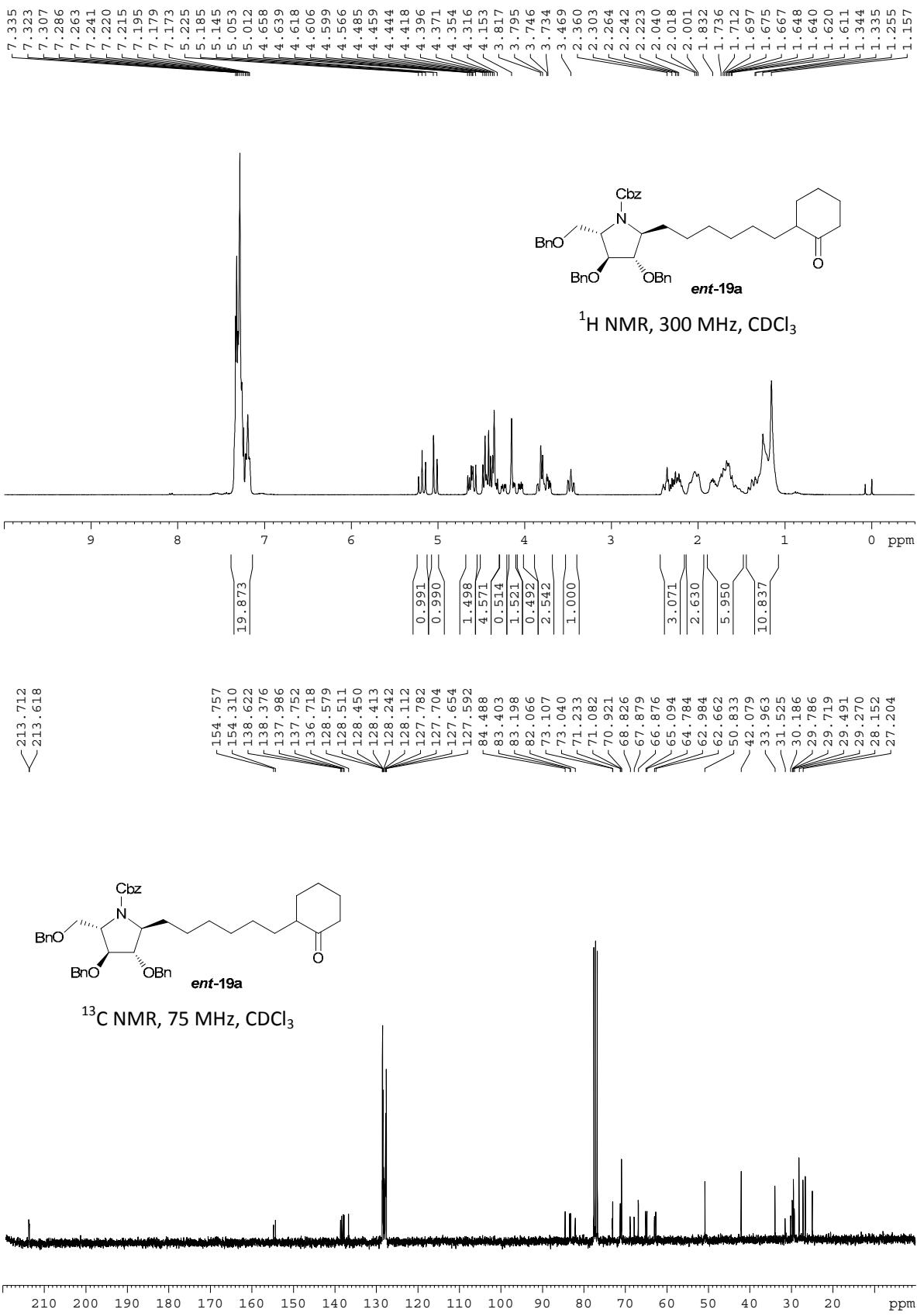




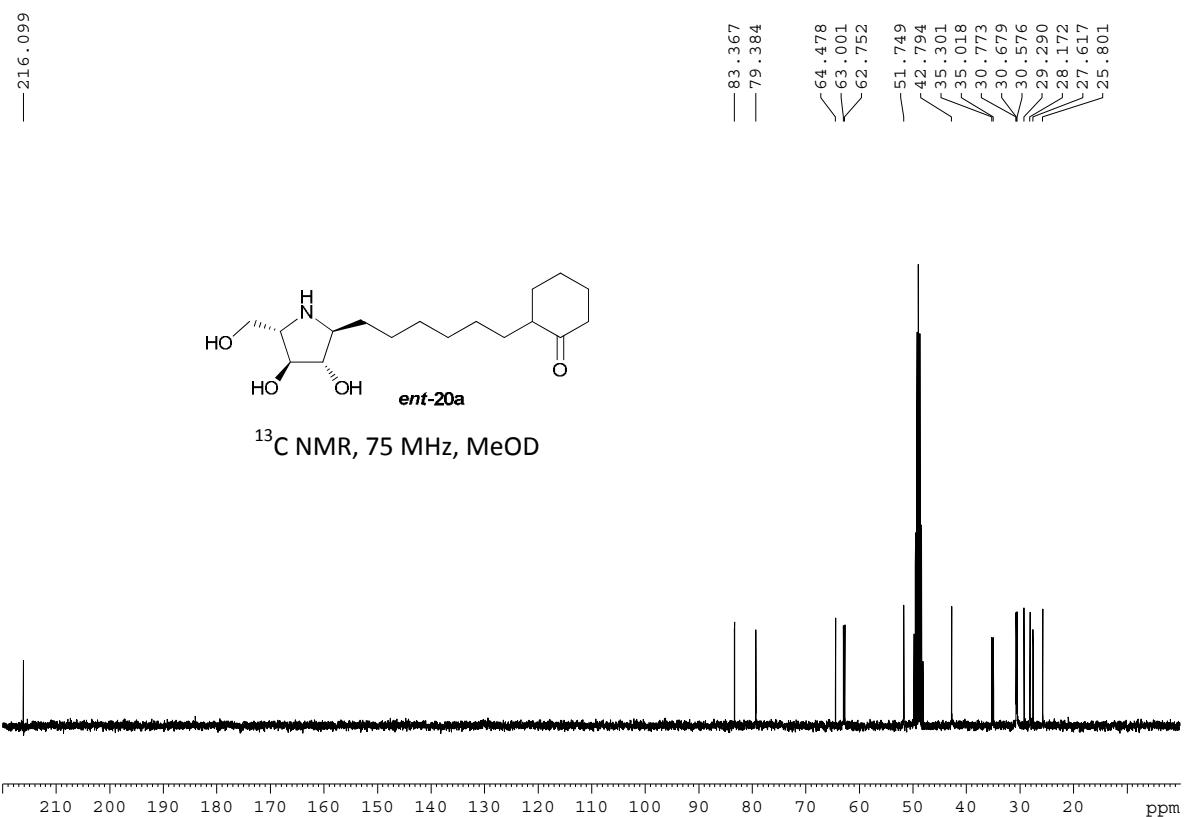
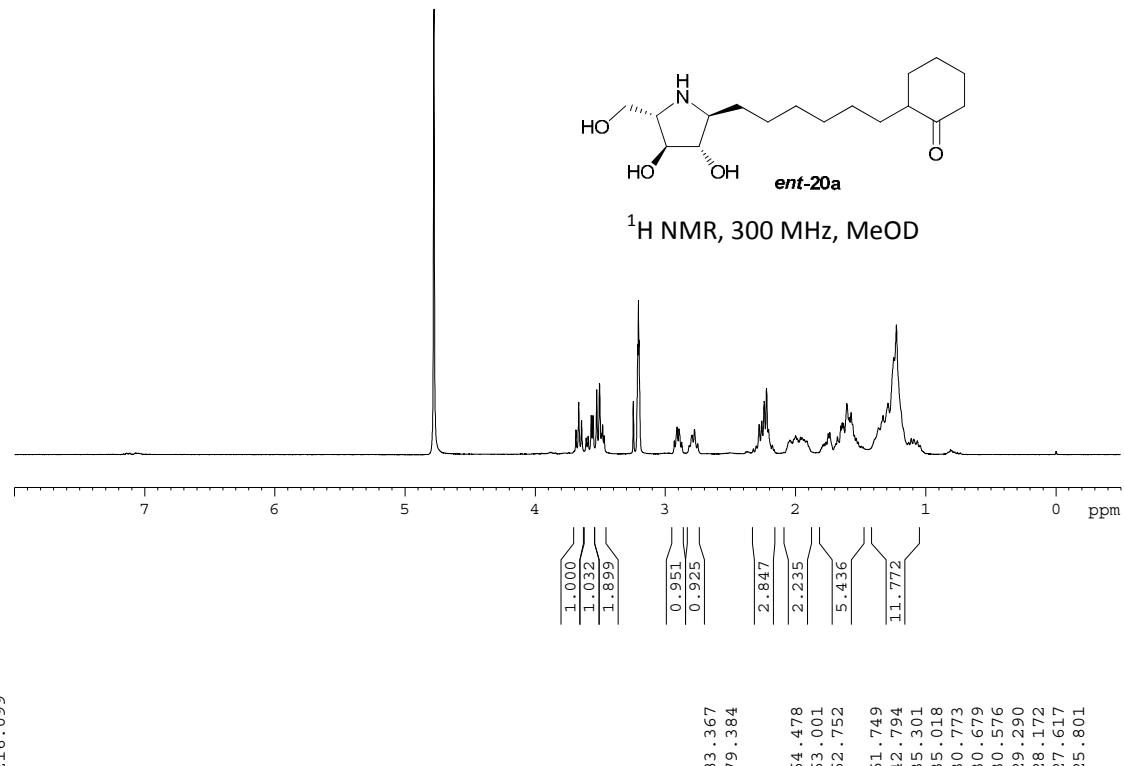


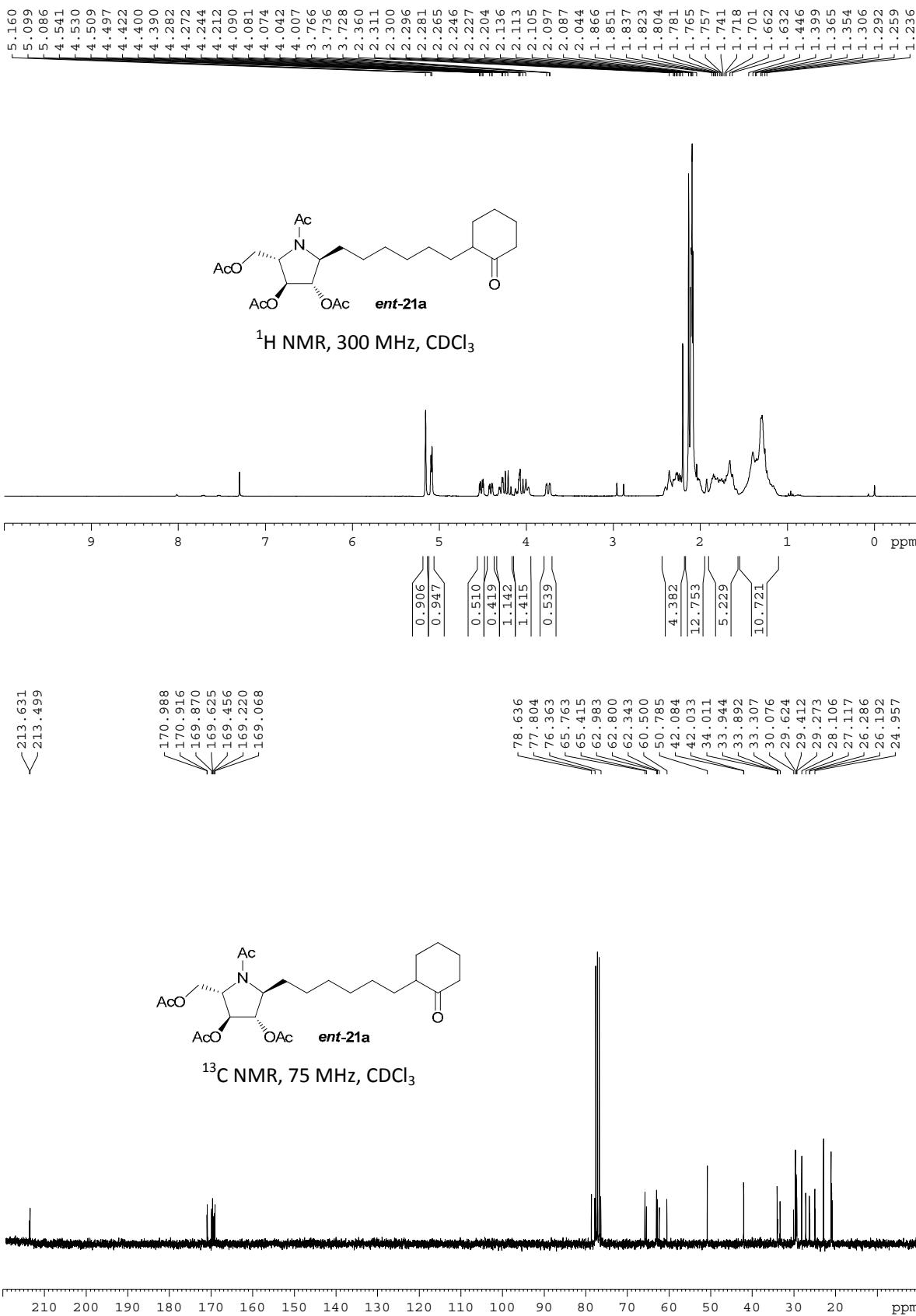


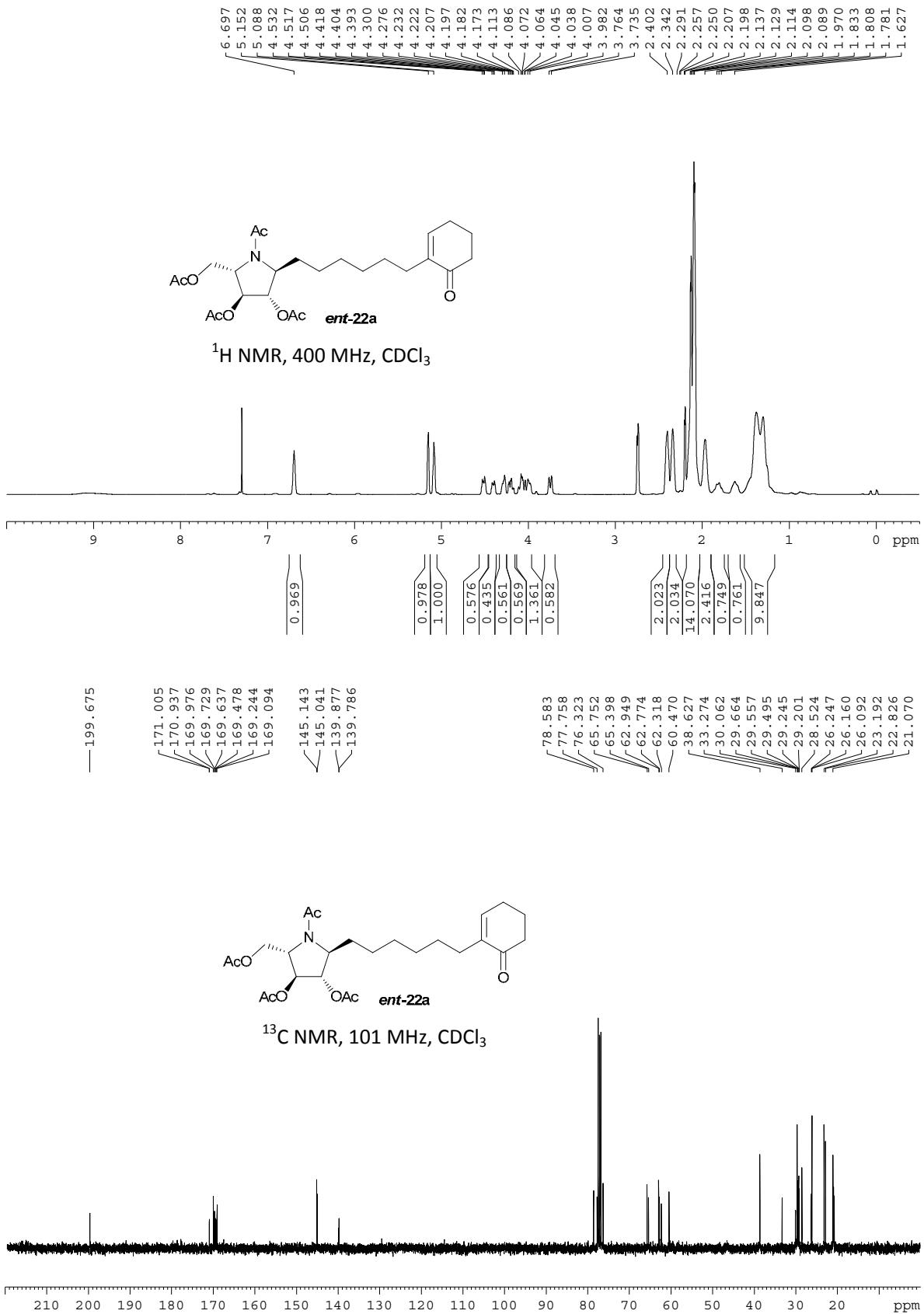


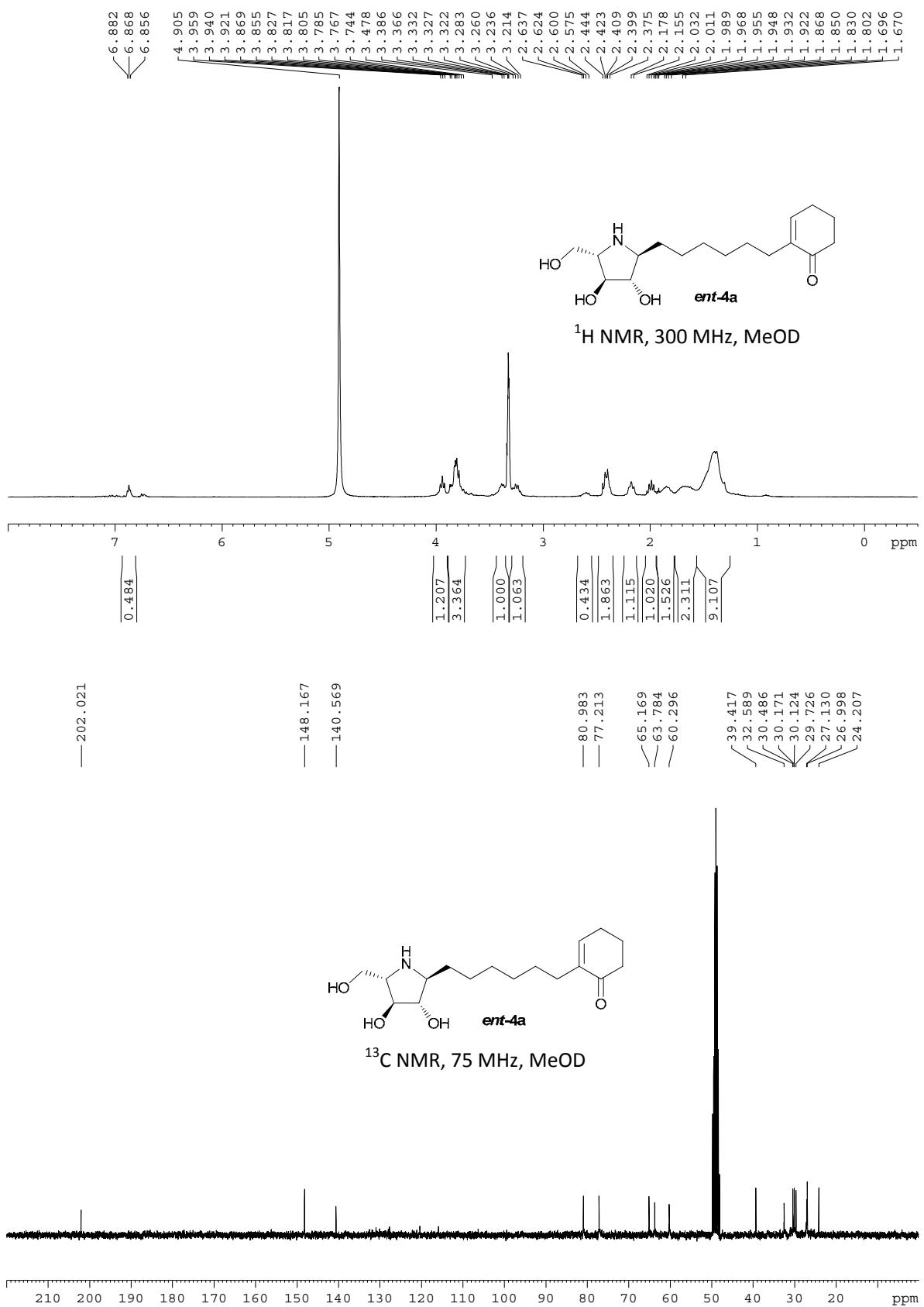


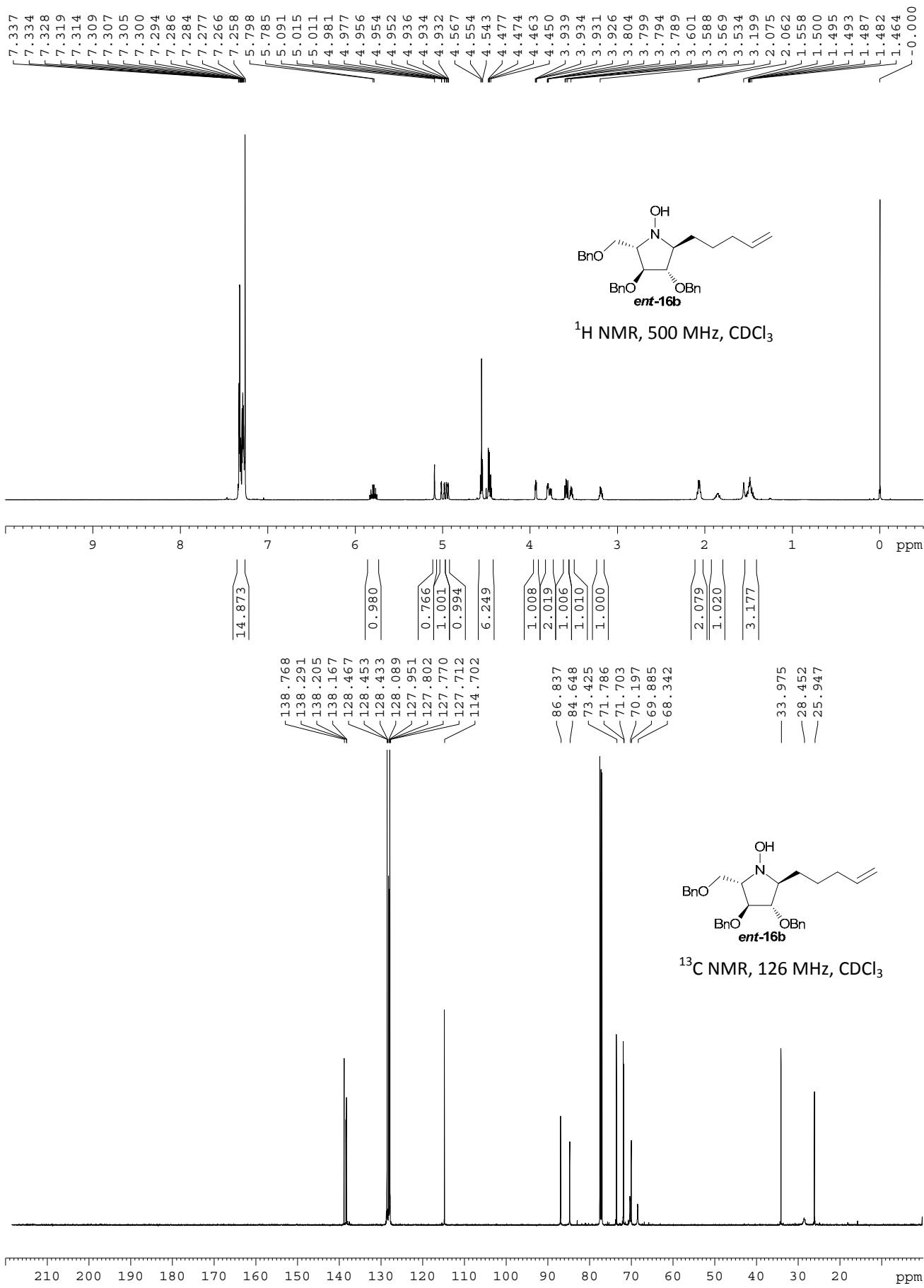
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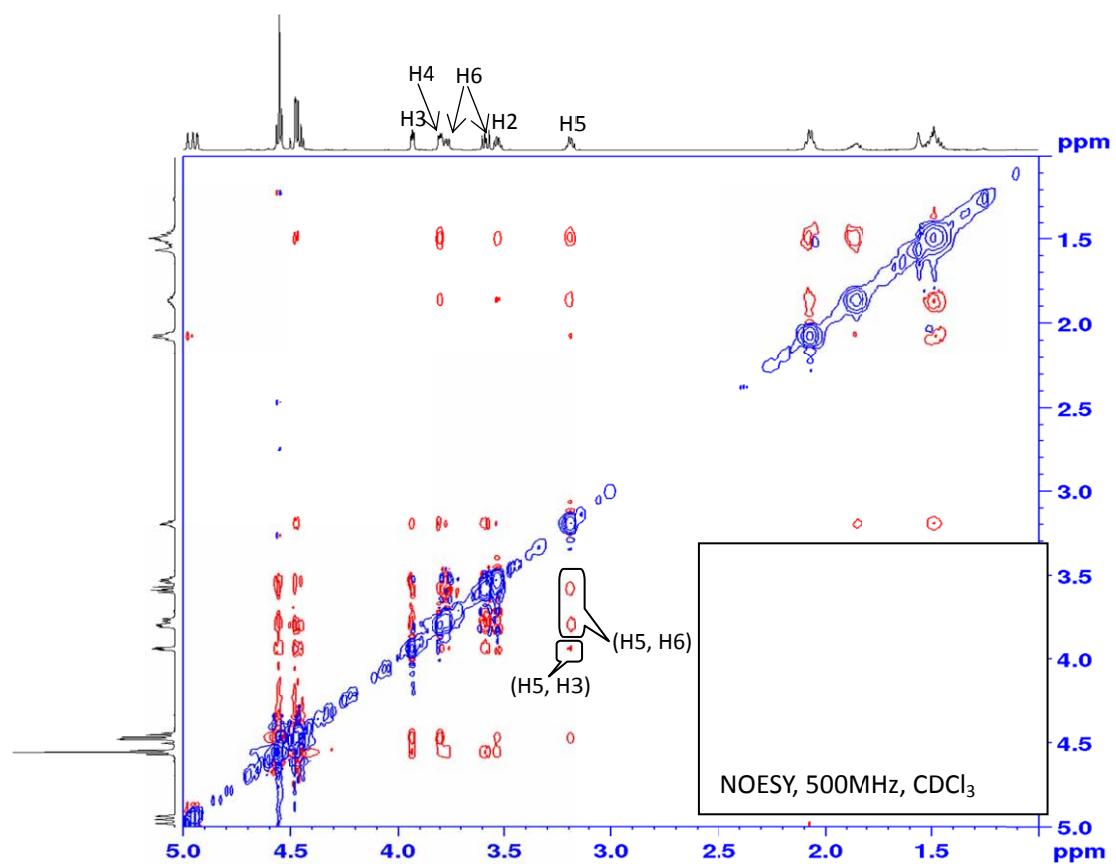
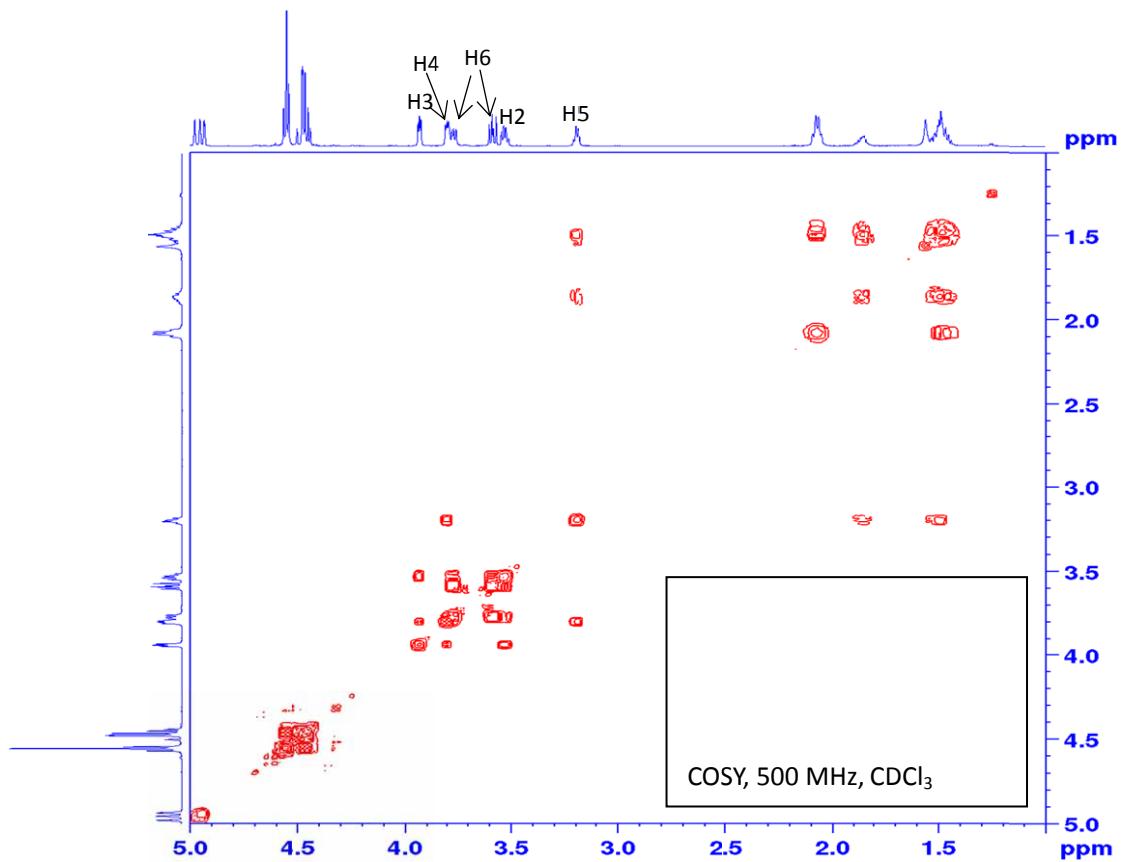


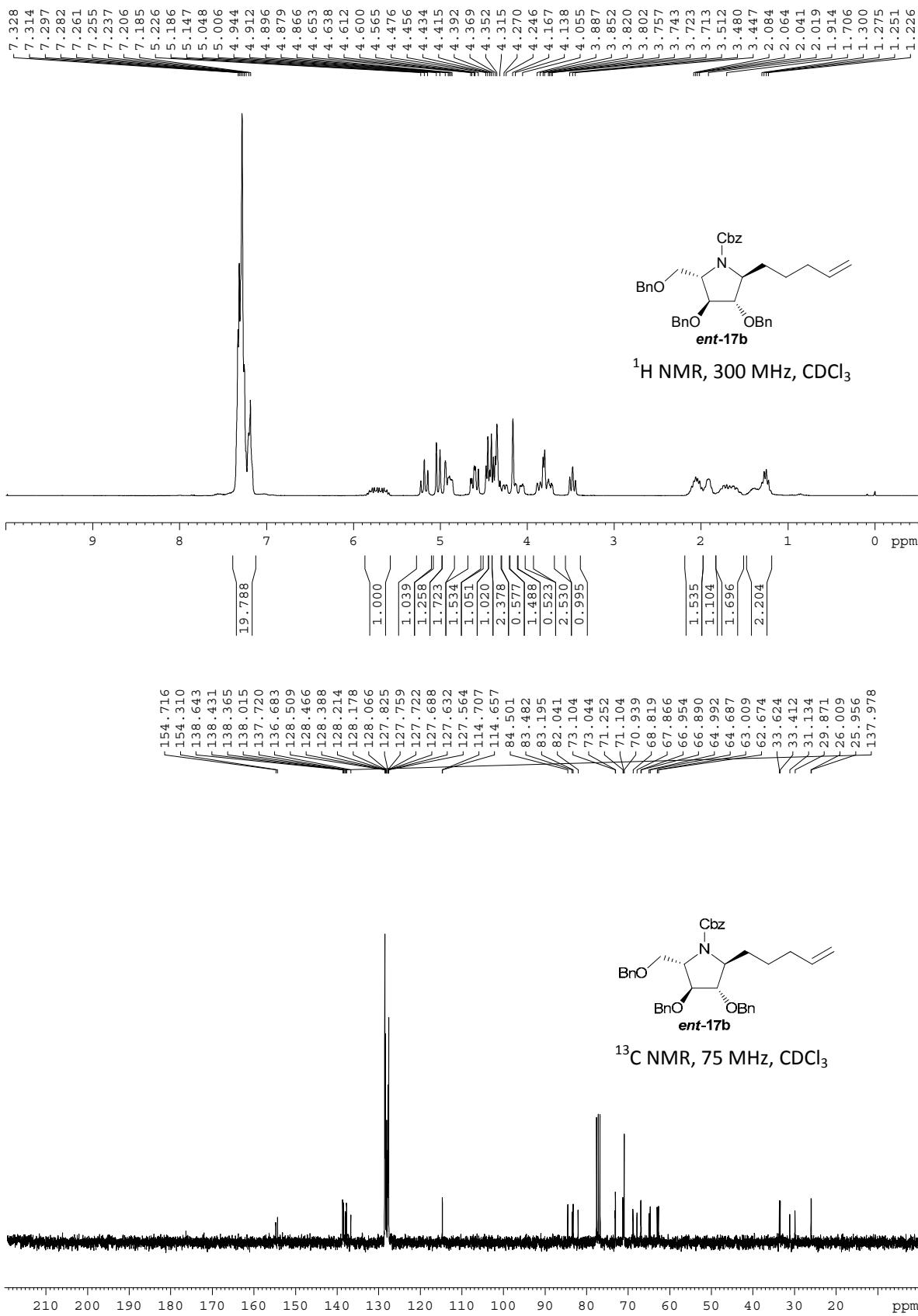


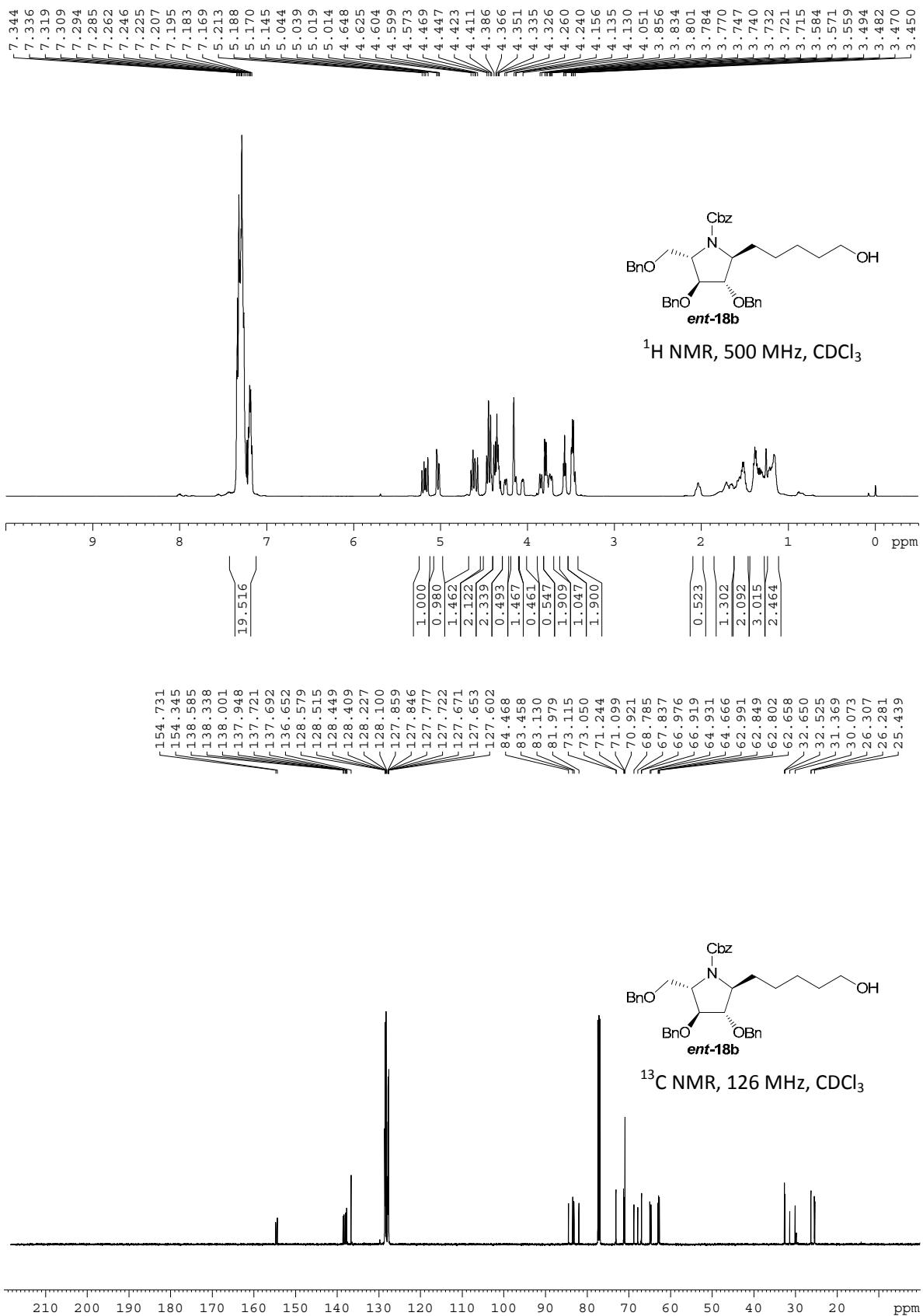


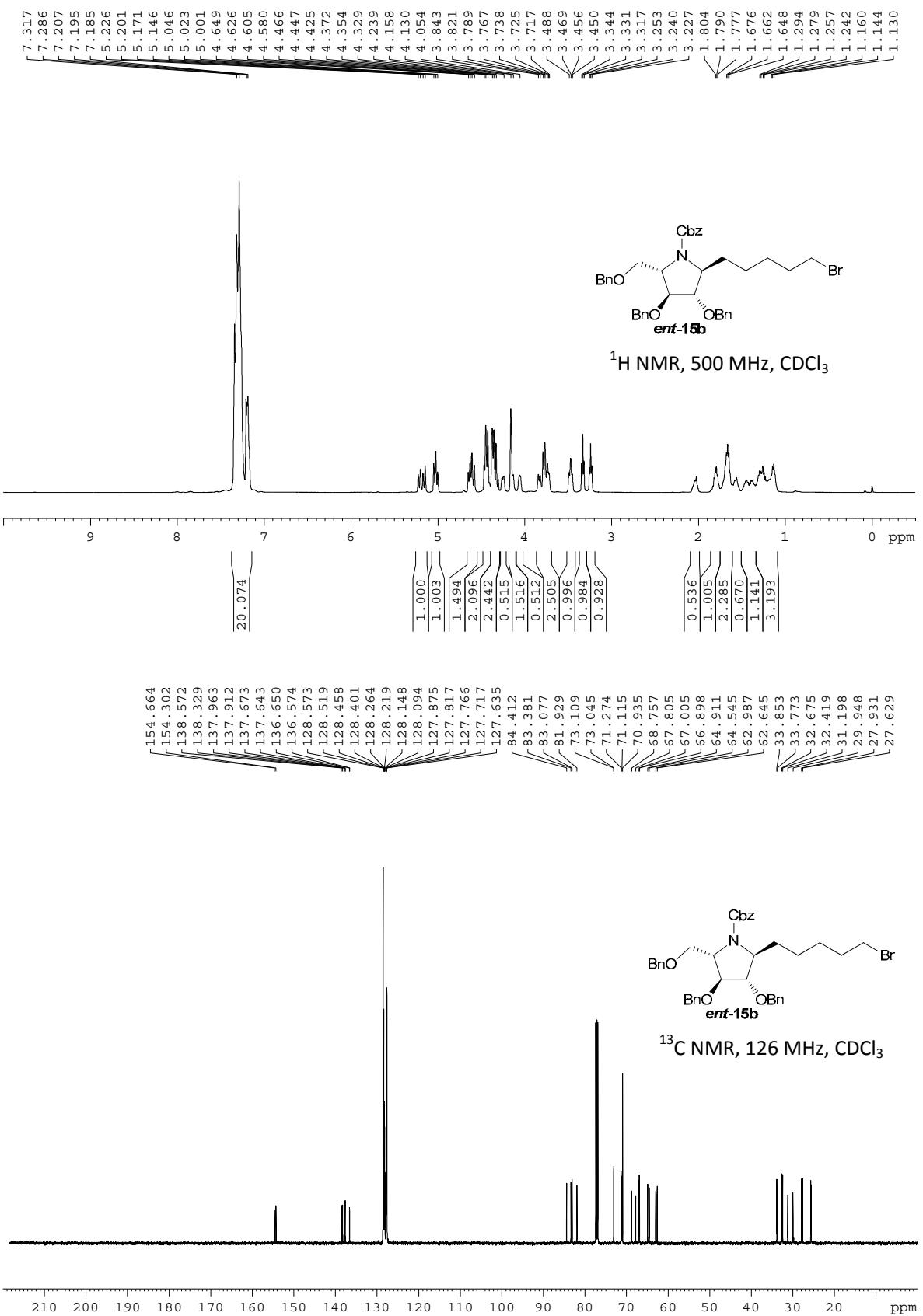


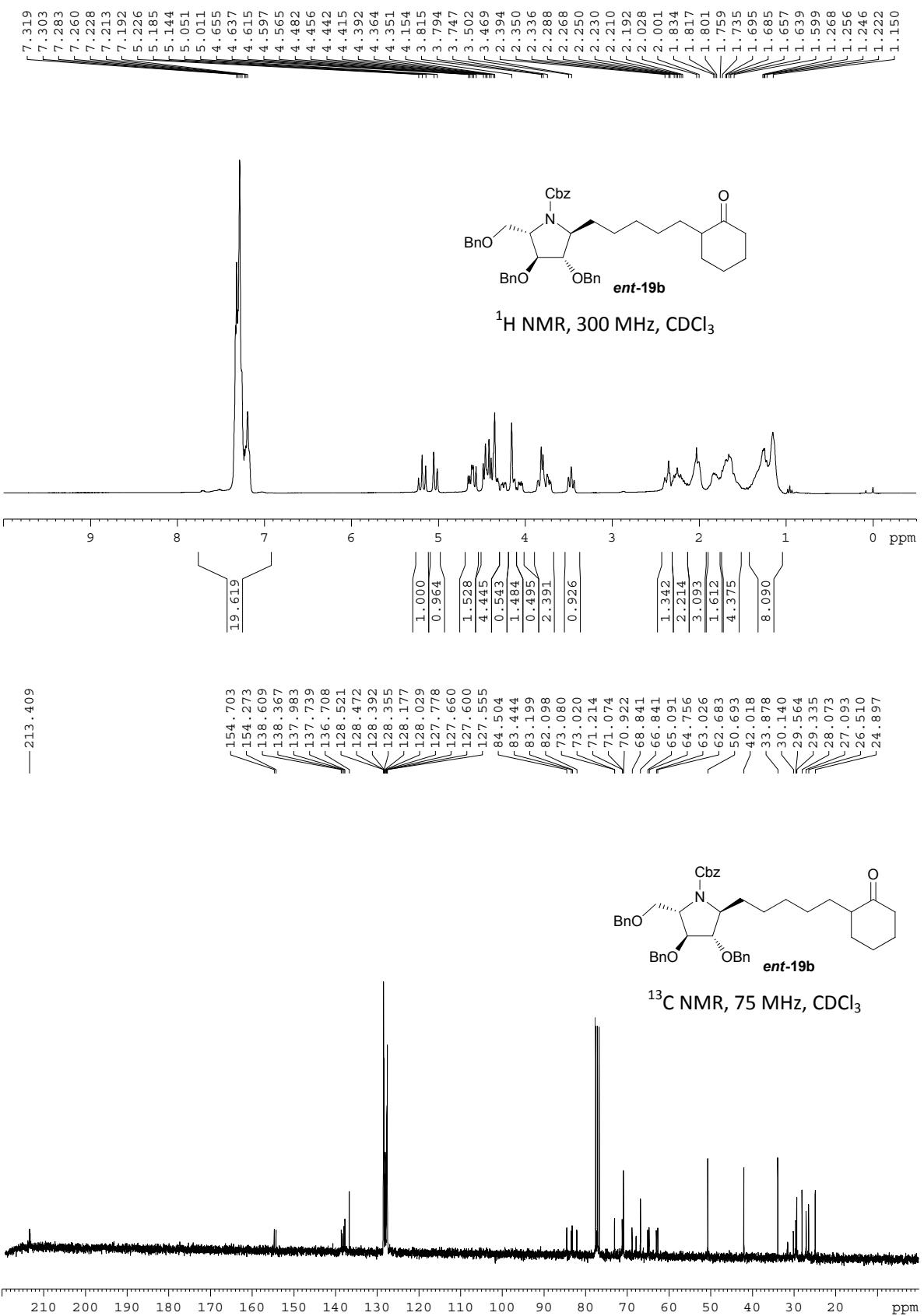


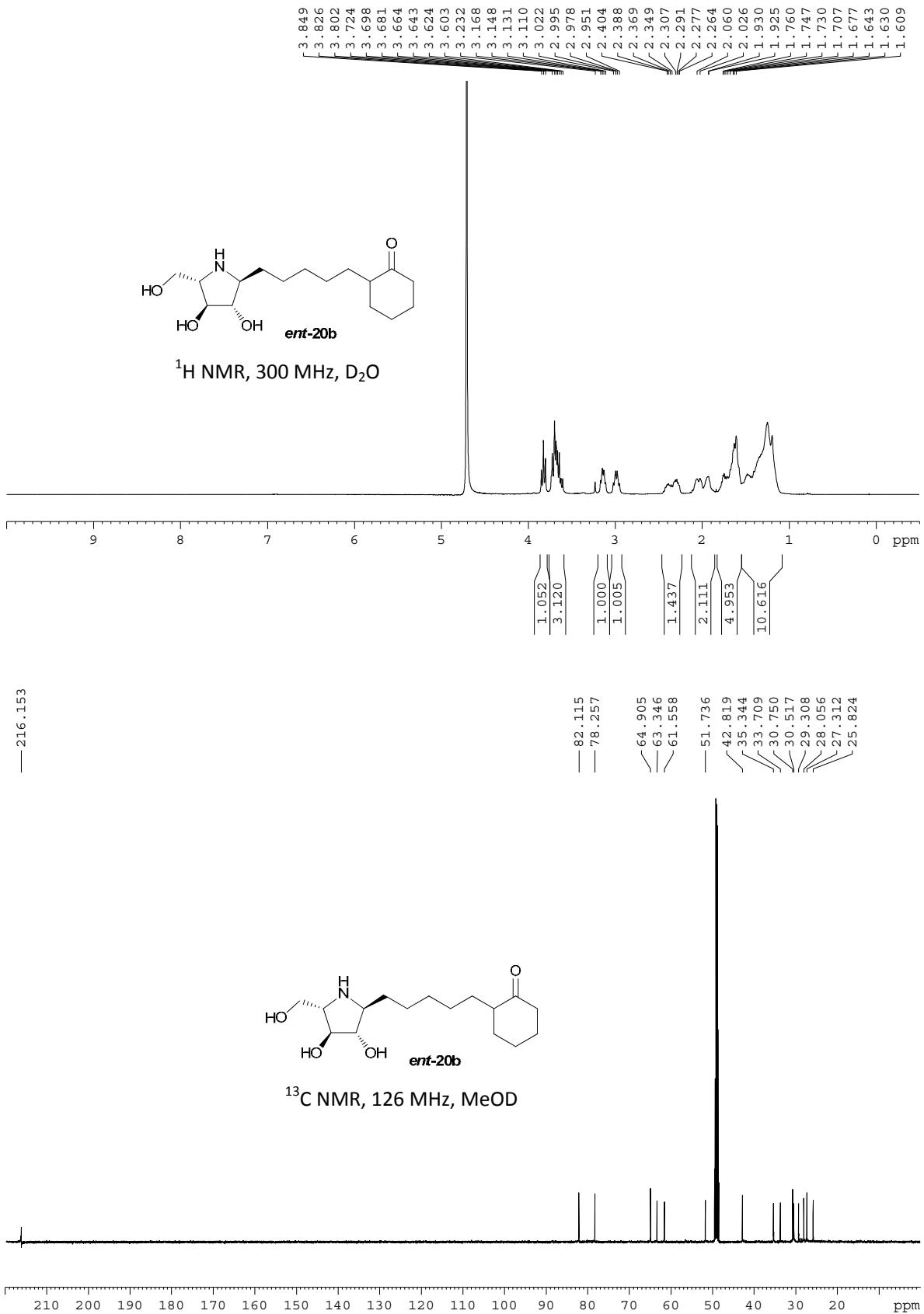


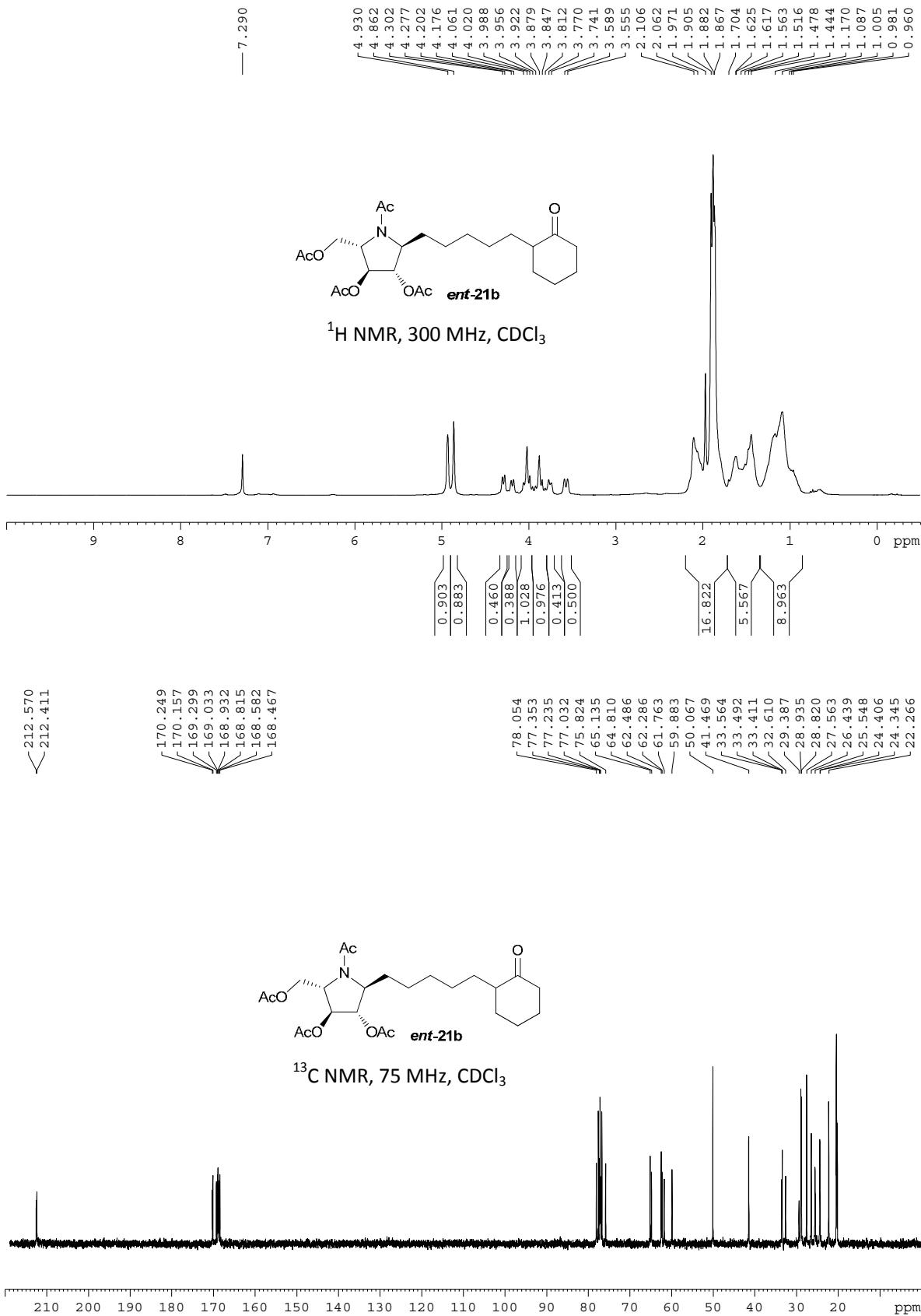


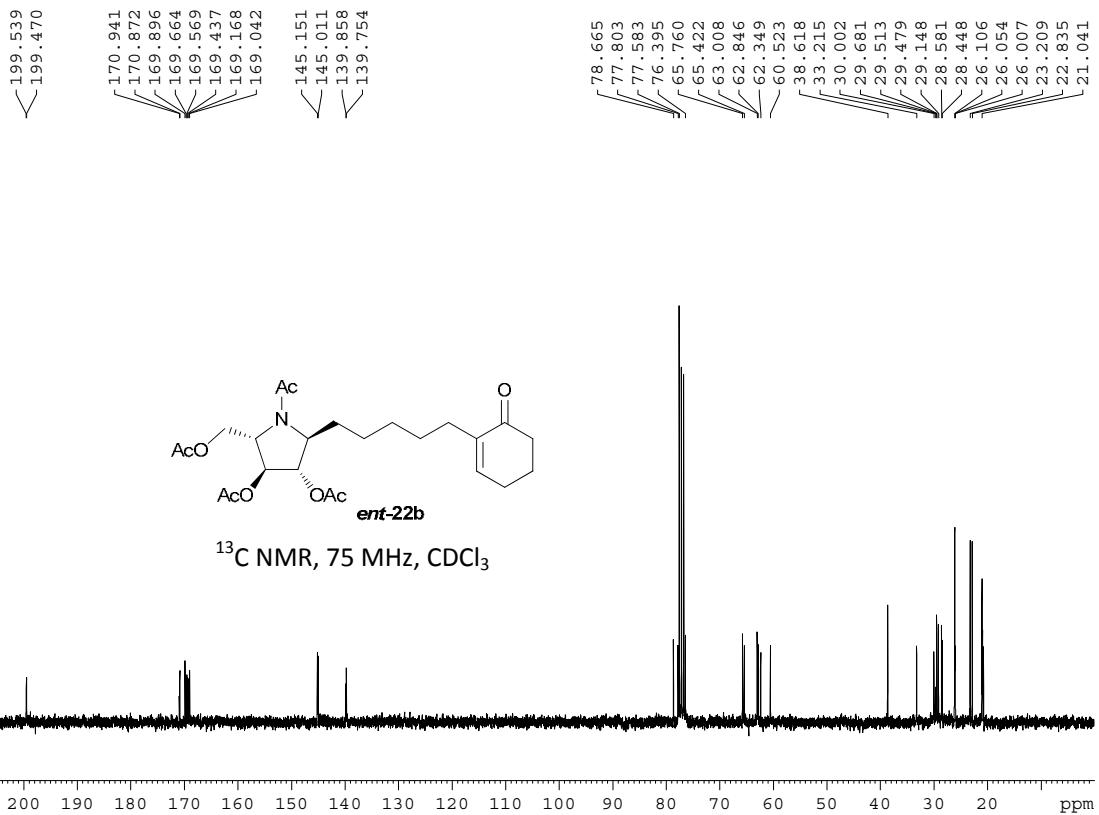
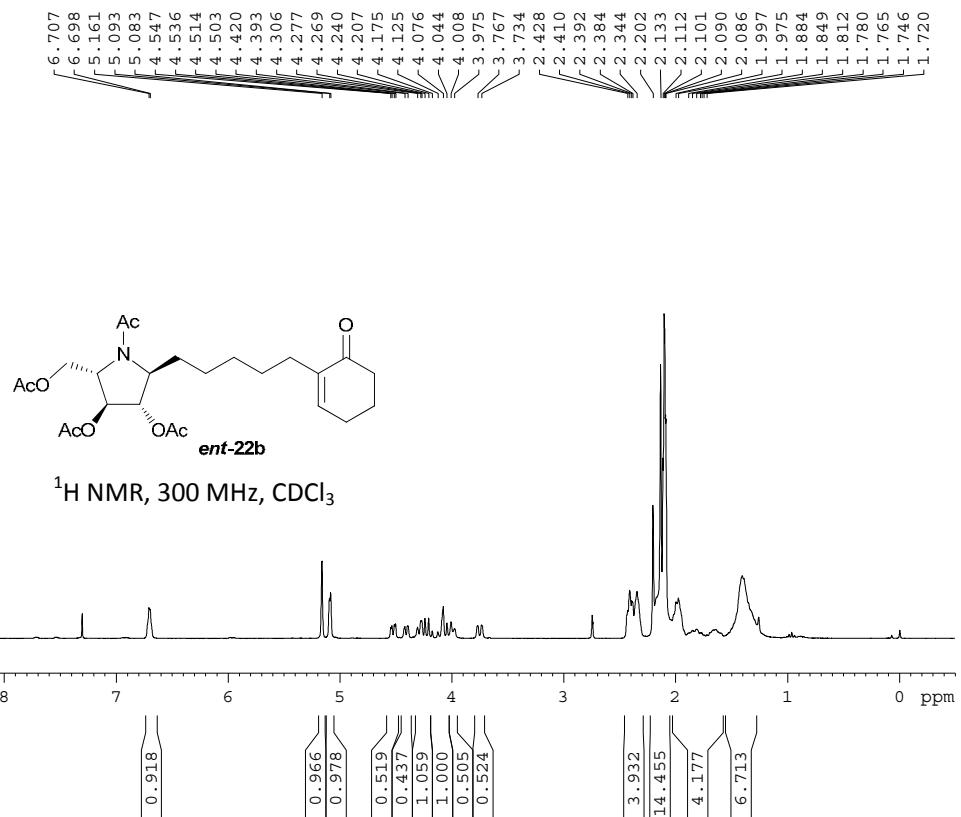


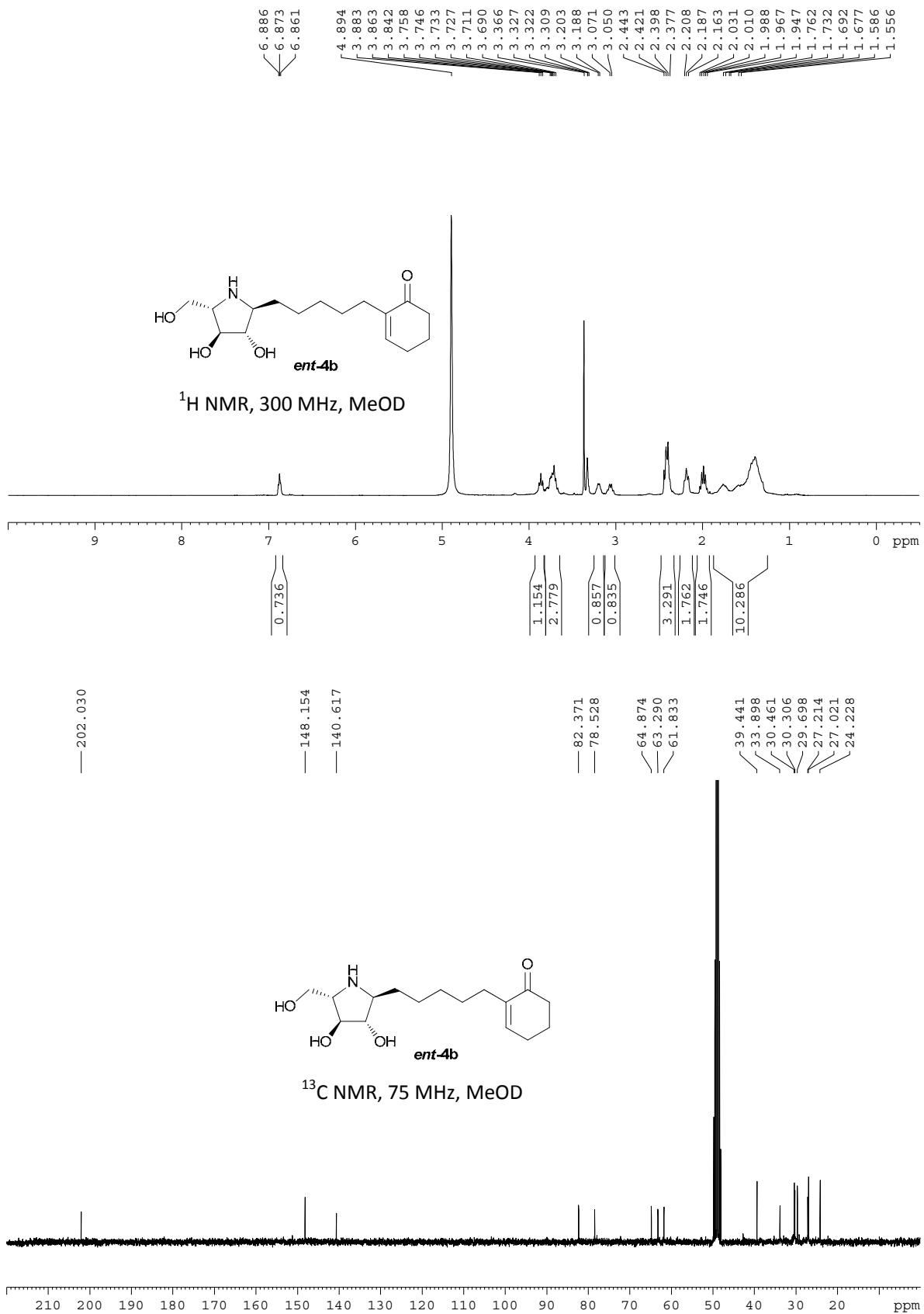












## 4. Results of the bioassay

The enzymes  $\alpha$ -glucosidase (from yeast; from rice),  $\beta$ -glucosidase (from almond; from bovine liver),  $\alpha$ -galactosidase (from coffee beans),  $\beta$ -galactosidase (from bovine liver),  $\alpha$ -mannosidase (from jack bean),  $\beta$ -mannosidase (from snail),  $\alpha$ -L-fucosidase (from bovine kidney),  $\alpha,\alpha$ -trehalase (from porcine kidney), amyloglucosidase (from *A. niger*),  $\alpha$ -L-rhamnosidase (from *Pdecumbens*),  $\beta$ -glucuronidases (from bovine liver; from *E. coli*), *p*-nitrophenyl glycosides, and various disaccharides were purchased from Sigma-Aldrich Co. Brush border membranes were prepared from the rat small intestine according to the method of Kessler et al, (Kessler et al 1978)<sup>1</sup> and were assayed at pH 6.8 for rat intestinal maltase using maltose. ER  $\alpha$ -glucosidase II was prepared from rat liver according to the method of Hino et al.<sup>2</sup> For rat intestinal glucosidases and porcine kitrehalase activities, the reaction mixture (0.2 mL) contained 25 mM substrate and the appropriate amount of enzyme, and the incubations were performed for 10 min at 37 °C. The reaction was stopped by heating at 100 °C for 3 min. After centrifugation (600 g; 10 min), 0.035 mL of the resulting reaction mixture were added to 2.1 mL of the Glucose CII-test Wako (Wako Pure Chemical Ind., Osaka, Japan). The absorbance at 505 nm was measured to determine the amount of the released D-glucose. Other glycosidase activities were determined using an appropriate *p*-nitrophenyl glycoside as substrate at the optimum pH of each enzyme. The reaction mixture (0.2 mL) contained 2 mM of the substrate and the appropriate amount of enzyme. The reaction was stopped by adding 0.4 mL of 400 mM Na<sub>2</sub>CO<sub>3</sub>. The released *p*-nitrophenol was measured spectrometrically at 400 nm. The assay results are summarized in the following Table 3 and Table 4.

**Table 3.** Concentrations of Broussonetine W and Its Analogues (**4**, *ent-4*, *3-epi-4*, *ent-3-epi-4*, *ent-4a*, *ent-4b*) Giving 50% Inhibition of Various Glycosidases.

enzyme	IC <sub>50</sub> (μM)					
	<b>4</b>	<i>ent-4</i>	<i>3-epi-4</i>	<i>ent-3-epi-4</i>	<i>ent-4a</i>	<i>ent-4b</i>
<b>α-glucosidase</b>						
yeast	22	407	NI <sup>a</sup> (23.7%) <sup>b</sup>	310	758	622
rice	119	<b>0.73</b>	63	24	<b>1.1</b>	<b>1.3</b>
rat intestinal maltase	67	<b>0.047</b>	NI (43.9%)	<b>3.5</b>	<b>0.19</b>	<b>0.2</b>
intestinal isomaltase	NI (29.3%)	<b>1.5</b>	NI (0%)	114	<b>3.0</b>	<b>5.1</b>
rat intestinal sucrase	216	<b>0.20</b>	111	<b>3.4</b>	<b>0.25</b>	<b>0.28</b>
ER α-glucosidase II	NI (43.5%)	7.5	NI (13.2%)	200	15	21
<b>β-glucosidase</b>						
almond	<b>9.8</b>	NI (39.5%)	NI (31.8%)	92	NI (27.2%)	NI (13.4%)
bovine liver	<b>0.12</b>	180	44	<b>0.99</b>	59	263
<b>α-galactosidase</b>						
coffee bean	NI (19.6%)	NI (4.2%)	NI (46.9%)	NI (15.8%)	NI (1.1%)	NI (19.1%)
<b>β-galactosidase</b>						
bovine liver	<b>0.03</b>	127	11	<b>0.18</b>	11	105
<b>α-mannosidase</b>						
jack bean	NI (15.1%)	NI (0%)	385	NI (5.4%)	NI (0%)	NI (6.5%)
<b>β-mannosidase</b>						
snail	282	NI (0%)	NI (7.7%)	NI (0%)	NI (0%)	NI (1.2%)
<b>α-L-fucosidase</b>						
bovine kidney	NI (16.8%)	NI (0%)	NI (12.3%)	20.4	NI (0%)	NI (43.3%)
<b>α, α-trehalase</b>						
porcine kidney	NI (0.8%)	NI (8.3%)	NI (2.2%)	NI (3.3%)	NI (16.4%)	NI (10.0%)
<b>amyloglucosidase</b>						
<i>Aspergillusniger</i>	30	NI (0%)	NI (2.8%)	448	NI (16.2%)	NI (0%)
<b>α-L-rhamnosidase</b>						
<i>Penicillium decumbens</i>	NI (35.1%)	174	NI (47.3%)	166	591	1000
<b>β-glucuronidase</b>						
<i>E.coli</i>	<b>3.3</b>	38	14	83.3	25	60
bovine liver	NI (46.1%)	379	27	NI (40.2%)	NI (46.7%)	186

<sup>a</sup> NI: No inhibition (less than 50% inhibition at 1000 μM).

<sup>b</sup> ( ): inhibition % at 1000 μM.

**Table 4.** Concentrations of Broussonetine W's Analogues (**20**, *ent-20*, *3-epi-20*, *ent-3-epi-20*, *ent-20a*, *ent-20b*) Giving 50% Inhibition of Various Glycosidases

enzyme	IC <sub>50</sub> (μM)					
	<b>20</b>	<i>ent-20</i>	<i>3-epi-20</i>	<i>ent-3-epi-20</i>	<i>ent-20a</i>	<i>ent-20b</i>
<b><i>α</i>-glucosidase</b>						
yeast	80	NI <sup>a</sup> (32.7%) <sup>b</sup>	NI (44.0%)	NI (16.7%)	NI (46.8%)	NI (39.1%)
rice	70	<b>1.7</b>	<b>7.9</b>	33	<b>1.4</b>	<b>3.7</b>
rat intestinal maltase	32	<b>0.15</b>	<b>1.6</b>	13	<b>0.11</b>	<b>0.14</b>
intestinal isomaltase	663	<b>3.7</b>	40.3	366	<b>2.1</b>	<b>3.8</b>
rat intestinal sucrase	80	<b>0.36</b>	<b>3.1</b>	18	<b>0.14</b>	<b>0.17</b>
ER <i>α</i> -glucosidase II	NI (46.9%)	14	157	458	12	25
<b><i>β</i>-glucosidase</b>						
almond	43	NI (37.2%)	NI (25.4%)	NI (35.5%)	NI (35.8%)	NI (39.0%)
bovine liver	<b>0.10</b>	14	27	40	237	372
<b><i>α</i>-galactosidase</b>						
coffee bean	NI (13.8%)	NI (14.6%)	NI (29.4%)	NI (13.5%)	NI (0%)	NI (0.7%)
<b><i>β</i>-galactosidase</b>						
bovine liver	<b>0.046</b>	<b>8.5</b>	35	14	42	129
<b><i>α</i>-mannosidase</b>						
jack bean	NI (33.7%)	NI (0%)	245	NI (2.4%)	NI (1.2%)	NI (0%)
<b><i>β</i>-mannosidase</b>						
snail	224	NI (15.2%)	NI (0%)	NI (0%)	NI (0.9%)	NI (0%)
<b><i>α-L</i>-fucosidase</b>						
bovine kidney	NI (25.2%)	NI (0.7%)	NI (18.9%)	28	NI (35.7%)	NI (7.6%)
<b><i>α, α</i>-trehalase</b>						
porcine kidney	NI (0.8%)	NI (3.3%)	NI (0%)	NI (0%)	NI (10.0%)	NI (13.2%)
<b>amyloglucosidase</b>						
<i>Aspergillusniger</i>	41	NI (28.3%)	NI (7.7%)	NI (8.6%)	NI (4.6%)	NI (1.5%)
<b><i>α-L</i>-rhamnosidase</b>						
<i>Penicillium decumbens</i>	NI (31.4%)	152	895	443	330	410
<b><i>β</i>-glucuronidase</b>						
<i>E.coli</i>	<b>9.5</b>	79	21	45	428	435
bovine liver	NI (39.7%)	NI (45.2%)	NI (39.0%)	NI (12.3%)	NI (31.9%)	597

<sup>a</sup> NI: No inhibition (less than 50% inhibition at 1000 μM).

<sup>b</sup> ( ): inhibition % at 1000 μM

(1) Kessler, M.; Acuto, O.; Strelli, C.; Murer, H.; Semenza, G. A. *Biochem. Biophys. Acta.* **1978**, 506, 136.

(2) Hino, Y., Rothman, J. E. *Biochemistry* **1985**, 24, 800-805.