# SUPPORTING INFORMATION

# A Core Switching Strategy to Pyrrolo[2,3-b]quinolines and Diazocino[1,2-a]indolinones

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Scheme S1 Chiral memory experiment with enantio-enriched *M*-atropisomer of 1a<sup>S1</sup> with NaOMe lead to ±2a



		Observed		
Nucleophile	[M+H]⁺	, [M+Na]⁺	[M-H] <sup>-</sup>	m/z
sodium methoxide	353.3	375.3	351.3	351.3ª
<i>n</i> -butylamine	394.4	416.4	392.4	392.4ª
1,4-diaminobutane	409.4	431.4	407.4	407.4ª
acetylethylenediamine	423.4	445.4	421.4	421.4ª
Benzylamine	428.4	450.4	426.4	426.4ª
Aniline <sup>d</sup>	414.4	436.4	412.4	394.0 <sup>*,d</sup>
morpholine	408.4	430.4	406.4	406.4ª
(15,25)-2-benzyloxycyclopentylamine <sup>d</sup>	512.6	534.6	510.6	492.0 <sup>*,d</sup>
N-methylhydrazine	367.4	389.4	365.4	365.4ª
benzyl carbazate	487.5	509.5	485.5	485.5ª
sodium azide	364.3	386.3	362.3	362.3ª
glycine <i>t</i> -butyl ester. HCl <sup>d</sup>	452.5	474.5	450.5	466.0 <sup>*,d</sup>
propan-2-ol <sup>c</sup>	381.4	403.4	379.4	319.0 <sup>c</sup>
propargyl alcohol <sup>c</sup>	377.4	399.4	375.4	319.0 <sup>c</sup>
methyllithium.LiBr complex	337.3	359.3	335.3	335.3ª
allyl magnesium bromide	363.4	385.4	361.4	361.4ª
ethynyl magnesium chloride	347.3	369.3	345.3	345.3ª
methyl magnesium bromide	337.3	359.3	335.3	335.3ª
trimethylsilyl cyanide <sup>c</sup>	348.3	370.3	346.3	319.0 <sup>c</sup>
triphenylmethanethiol	597.7	619.7	595.7	595.7ª
Ethylthioglycolate <sup>d</sup>	427.4	449.4	425.4	439.0 <sup>*,d</sup>
Benzylmercaptan	445.5	467.5	443.5	445.5 <sup>b</sup>

Table S1 Nucleophile screen of the scope of the transformation of 1a to examples of 2

**Details:** Using a Radleys<sup>®</sup> Greenhouse parallel synthesiser, to 18 stirred solutions of **1a** (20 mg) in dry THF (5mL) were added 1.1 equivalents of the relevant nucleophile at room temperature for 16 hours. In a separate study, the four reactions using Grignard reactions were carried out at -78 °C and then allowed to warm to 0 °C after 3 hours. LCMS analysis (ES<sup>+</sup>/ES<sup>-</sup>) of the crude reaction mixtures revealed that 14 / 22 reaction mixtures contained a new product with a *m/z* from either the ES<sup>-</sup> (<sup>a</sup> in table) or ES<sup>+</sup> (<sup>b</sup> in table) spectra coupled with the UV<sub>254nm</sub> trace;

c no reaction was observed and observed m/z consistent with the presence of starting material 1a; <sup>d</sup> structure of product not determined.

Entry	NaOMe (eq.)	Solvent	Temperature (°C)	Time (min)	Yield (%)
1	1	MeOH	25	10	95
2	2	MeOH	25	10	99
3	10	MeOH	25	10	95
4	2	THF	25	960	45

Table S2 Optimisation of the sodium methoxide-induced rearrangement of 1a to 2a

**Details:** Variation of reaction conditions and the resulting isolated yields for the methoxide induced azepinoindole rearrangement of **1a** to give **2a**.

entry	eq. <i>n</i> BuNH₂	time (min)	percentage conversion
1	1	10	40
2	1	60	80
3	1	180	95 (74ª)
4	1	960	b
5	2	180	70 (51ª)
6	4	180	b
7	10	180	b

Table S3 Optimisation of the butylamine-induced rearrangement of  ${\bf 1a}$  to  ${\bf 2i}$ 

**Details:** Percentage conversion as estimated by <sup>1</sup>H NMR spectra of the transformation of **1a** to **2i**. <sup>a</sup>Isolated yield (after flash column chromatography); <sup>b</sup>Substantial degradation was observed.

Proton Assignment	δ ¹H (ppm)	Multiplicity	Integration	J value (Hz)	Carbon Assignment	δ <sup>13</sup> C (ppm)
C2- <u>H</u> 2	4.19	t	2H	7.5	C2	49.8
C3- <u>H</u> 2	3.36	t	2H	7.4	C3	25.3
					C3a	119.6
C4-O <u>H</u>	8.55	br s	1H	-	C4	153.1
					C5	178.3
					C5a	126.5
C6- <u>H</u>	8.42	d	1H	2.6	C6	129.1
					C7	129.6
C8- <u>H</u>	7.41	dd	1H	8.9, 2.6	C8	134.8
C9- <u>H</u>	7.28	d	1H	8.8	C9	134.7
					C9a	148.6
					C10a	157.8
					C1′	139.1
					C2′	129.6
C3'- <u>H</u>	7.90	dd	1H	7.7, 1.5	C3′	130.5
C4'- <u>H</u>	7.37	dd	1H	7.6, 1.2	C4'	126.5
C5'- <u>H</u>	7.58	ddd	1H	7.7, 7.7, 1.7	C5′	132.5
C6'- <u>H</u>	7.32	dd	1H	8.0, 1.0	C6'	125.4
					Ar <u>C</u> O₂Et	167.0
OC <u>H</u> ₂CH₃	3.96	q	2H	7.2	$OCH_2CH_3$	61.1
OCH₂C <u>H</u> ₃	1.06	q	3H	7.2	OCH₂ <u>C</u> H₃	14.1

**Table S4** <sup>1</sup>H and <sup>13</sup>C NMR based assignment of **4** (Derived from 2D <sup>1</sup>H-<sup>1</sup>H COSY, <sup>1</sup>H-<sup>13</sup>C HSQC and <sup>1</sup>H-<sup>13</sup>C HMBC NMR experiments).

Table S5 Optimisation of the sodium alkoxide-induced rearrangement of 1e to 3a

Entry	NaOMe (eq.)	Solvent	Temperature (°C)	Time (h)	Yield (%)
1	2	MeOH	25	16	77
2	2	MeOH	64	16	59
3	10	MeOH	25	16	71

**Details:** Variation of the reaction conditions for the alkoxide induced pyrrolo[2,3-*b*]quinoline rearrangement **3a** from **1e** and the resulting isolated yields.



Table S6 Crystal data and structure refinement details for compound 2a.

	C19.30 118 N2 05.30	
Formula weight	368.36	
Temperature	125(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/n	
Unit cell dimensions	a = 7.7594(12) Å	α= 90°.
	b = 19.459(3) Å	β= 106.142(2)°.
	c = 11.8044(19) Å	γ = 90°.
Volume	1712.1(5) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.429 Mg/m <sup>3</sup>	
Absorption coefficient	0.106 mm <sup>-1</sup>	

F(000)	772
Crystal size	.1 x .1 x .01 mm <sup>3</sup>
Theta range for data collection	2.08 to 25.49°.
Index ranges	-9<=h<=9, -23<=k<=16, -12<=l<=14
Reflections collected	9839
Independent reflections	3068 [R(int) = 0.0244]
Completeness to theta = 25.49°	96.2 %
Absorption correction	MULTISCAN
Max. and min. transmission	1.00000 and 0.891243
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	3068 / 1 / 257
Goodness-of-fit on F <sup>2</sup>	1.024
Final R indices [I>2sigma(I)]	R1 = 0.0416, wR2 = 0.0971
R indices (all data)	R1 = 0.0552, wR2 = 0.1051
Extinction coefficient	0.0039(9)
Largest diff. peak and hole	0.455 and -0.445 e.Å <sup>-3</sup>



 Table S7 Crystal data and structure refinement details for compound 2i.

Formula weight	434.49	
Temperature	93(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/n	
Unit cell dimensions	a = 10.8627(13) Å	⊵= 90°.
	b = 19.295(2) Å	₽= 111.323(6)°.
	c = 11.3504(14) Å	<b>?</b> = 90°.
Volume	2216.1(5) Å <sup>3</sup>	
Z	4	
Density (calculated)	1.302 Mg/m <sup>3</sup>	
Absorption coefficient	0.090 mm <sup>-1</sup>	

F(000)	920
Crystal size	0.200 x 0.010 x 0.010 mm <sup>3</sup>
Theta range for data collection	2.11 to 25.34°.
Index ranges	-13<=h<=12, -16<=k<=23, -12<=l<=13
Reflections collected	15426
Independent reflections	3727 [R(int) = 0.0552]
Completeness to theta = 25.34°	92.0 %
Absorption correction	Multiscan
Max. and min. transmission	1.0000 and 0.8690
Refinement method	Full-matrix least-squares on F <sup>2</sup>
Data / restraints / parameters	3727 / 2 / 300
Goodness-of-fit on F <sup>2</sup>	1.192
Final R indices [I>2sigma(I)]	R1 = 0.0654, wR2 = 0.1112
R indices (all data)	R1 = 0.0893, wR2 = 0.1211
Extinction coefficient	0.0060(8)
Largest diff. peak and hole	0.234 and -0.224 e.Å <sup>-3</sup>

Copies of <sup>1</sup>H and <sup>13</sup>C NMR spectra: **Compound 1b** <sup>1</sup>H NMR (CDCl<sub>3</sub>)



Compound 1b <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2a <sup>1</sup>H NMR (CDCl<sub>3</sub>)



# Compound 2a <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2b <sup>1</sup>H NMR (CDCl<sub>3</sub>)



Compound 2b <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2c <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2c <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2d <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2d <sup>13</sup>C NMR (CDCl<sub>3</sub>)



## Compound 2e <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2e <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2f <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2f <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2g <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2g <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2h <sup>1</sup>H NMR (CDCl<sub>3</sub>)



# Compound 2h <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2i <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2i <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2j <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2j <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2k <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 2k <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 2I <sup>1</sup>H NMR (CDCI<sub>3</sub>)



## Compound 2I <sup>13</sup>C NMR (CDCI<sub>3</sub>)



Compound 2m <sup>1</sup>H NMR (d<sub>6</sub>-DMSO)



# Compound 2m <sup>13</sup>C NMR (d<sub>6</sub>-DMSO)



Compound 2n <sup>1</sup>H NMR (CDCl₃)



Compound 2n <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 20 <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 20 <sup>13</sup>C NMR (CDCl<sub>3</sub>)







## Compound 3a <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 3b <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 3b <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 3c <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 3c <sup>13</sup>C NMR (d<sub>6</sub>-DMSO)



Compound 3d <sup>1</sup>H NMR (CDCl<sub>3</sub>)



## Compound 3d <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 3e <sup>1</sup>H NMR (CDCl<sub>3</sub>)





Compound 3e<sup>13</sup>C NMR (CDCl<sub>3</sub>)

220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 ppm

Compound 3f <sup>1</sup>H NMR (d<sub>6</sub>-DMSO)



## Compound 3f <sup>13</sup>C NMR (d<sub>6</sub>-DMSO)



Compound 3g <sup>1</sup>H NMR (CDCI<sub>3</sub>)



## Compound 3g <sup>13</sup>C NMR (d<sub>6</sub>-DMSO)



## Compound 3h <sup>1</sup>H NMR (CDCl<sub>3</sub>)



Compound 3h <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 4 <sup>1</sup>H NMR (CDCl<sub>3</sub>)



# Compound 4 <sup>13</sup>C NMR (CDCl<sub>3</sub>)



Compound 4 2D NMR (CDCl<sub>3</sub>)





Figure S1: <sup>1</sup>H-<sup>13</sup>C HMBC NMR spectrum of 4 showing selected correlations that were used to determine the structure of ring B.

#### References

**S1** A. M.Jones, G. Liu, M. M. Lorion, S. Patterson, A. M. Z. Slawin, N. J. Westwood, *Chem. Eur. J.*, 2011, **17**, 5714.

**S2** J. L. C. Marais, W. Pickl, B. Staskun, *J. Org. Chem.*, 1990, **55**, 1969.