

Lipid Bilayer Thickness Determines Cholesterol's Location in Model Membranes

Drew Marquardt,^{1,2} Frederick A. Heberle,³⁻⁵ Denise V. Greathouse,⁶ Roger E. Koeppe II,⁶

Robert F. Standaert,^{7,8} Brad J. Van Oosten,¹ Thad A. Harroun,¹ Jacob J. Kinnun,⁹ Justin A.

Williams,⁹ Stephen R. Wassall*⁹ and John Katsaras*^{1,3,5,10,11}

¹Department of Physics, Brock University, St. Catharines, Ontario L2S 3A1, Canada

²Institute of Molecular Biosciences, University of Graz, Graz, Austria

³The Bredesen Center for Interdisciplinary Research and Graduate Education, University of Tennessee, Knoxville, Tennessee 37996, United States

⁴Joint Institute for Biological Sciences, Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, United States

⁵Biology and Soft Matter Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, United States

⁶Department of Chemistry and Biochemistry, University of Arkansas, Fayetteville, Arkansas 72701, United States

⁷Biosciences Division, Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, United States

⁸Department of Biochemistry and Cellular & Molecular Biology, University of Tennessee, Knoxville, TN 37996.

⁹Department of Physics, Indiana University - Purdue University, Indianapolis, Indiana 46202, United States

¹⁰Shull Wollan Center — a Joint Institute for Neutron Sciences, Oak Ridge National Laboratory, Oak Ridge, Tennessee 37831, United States

¹¹Department of Physics and Astronomy, University of Tennessee, Knoxville, Tennessee 37996, United States

Supplementary Information

^2H NMR

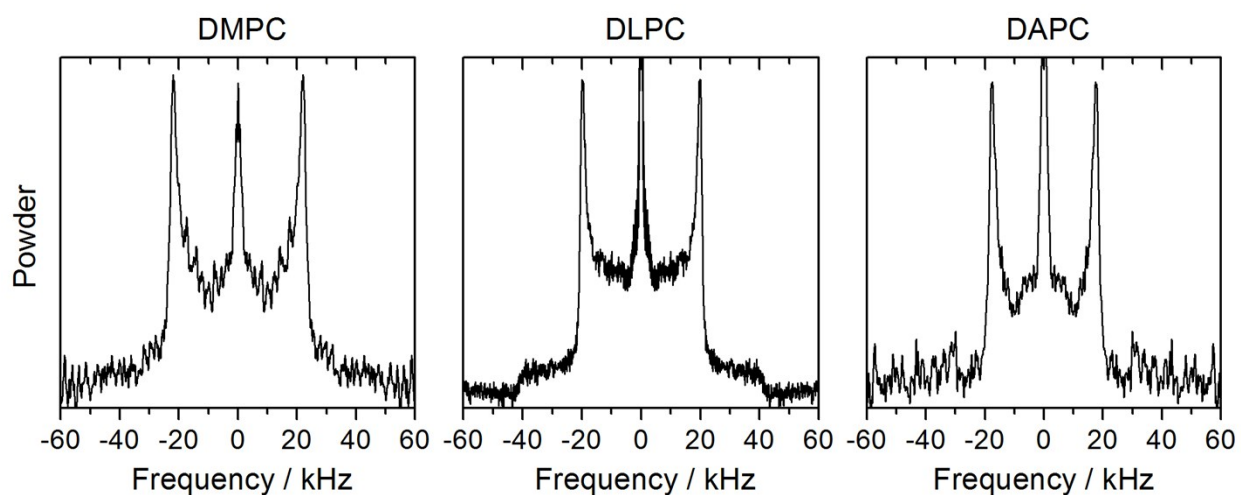


Figure S1 ^2H NMR spectra for 10 mol% [$3\alpha\text{-}^2\text{H}_1$]cholesterol in aqueous multilamellar dispersions of DLPC, DMPC and DAPC (50 wt% in 50 mM Tris, pH 7.5) at 30°C. The spectra are plotted in powder pattern format that were obtained by conventional FFT. The experiments were conducted on resonance, and the out-of-phase channel was zeroed, improving the signal-to-noise by a factor of $\sqrt{2}$ for the spectra that were reflected about their central (resonant) frequency. Residual ^2HHO together with [$3\alpha\text{-}^2\text{H}_1$]cholesterol in isotropic phospholipid phase produced during sample mixing are responsible for the central spike, which was ignored for data analysis.

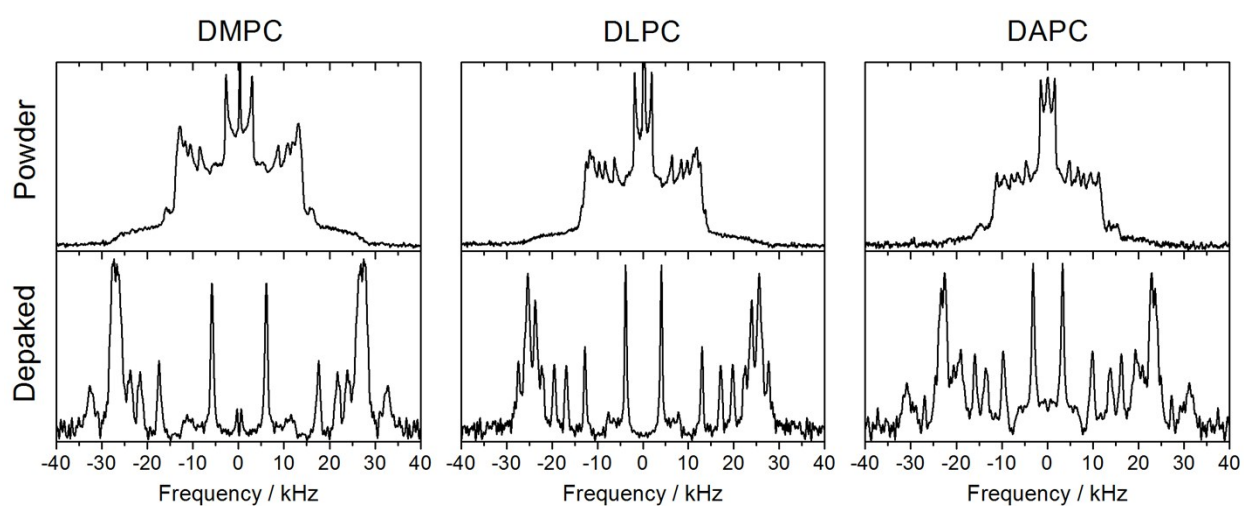


Figure S2 ^2H NMR spectra recorded at 30 °C with LA- d_{23} (5 mol%) in 50 wt% aqueous multilamellar dispersions of DMPC, DLPC and DAPC in 50 mM Tris (pH 7.5). The upper panels are powder patterns and the lower panels are FFT depaked. In processing the intrinsically noisy depaked data, the out-of-phase channel was zeroed before Fourier transformation. The resultant spectra were reflected about the central frequency and possess an improvement in signal/noise of a factor of $\sqrt{2}$.

MD simulations

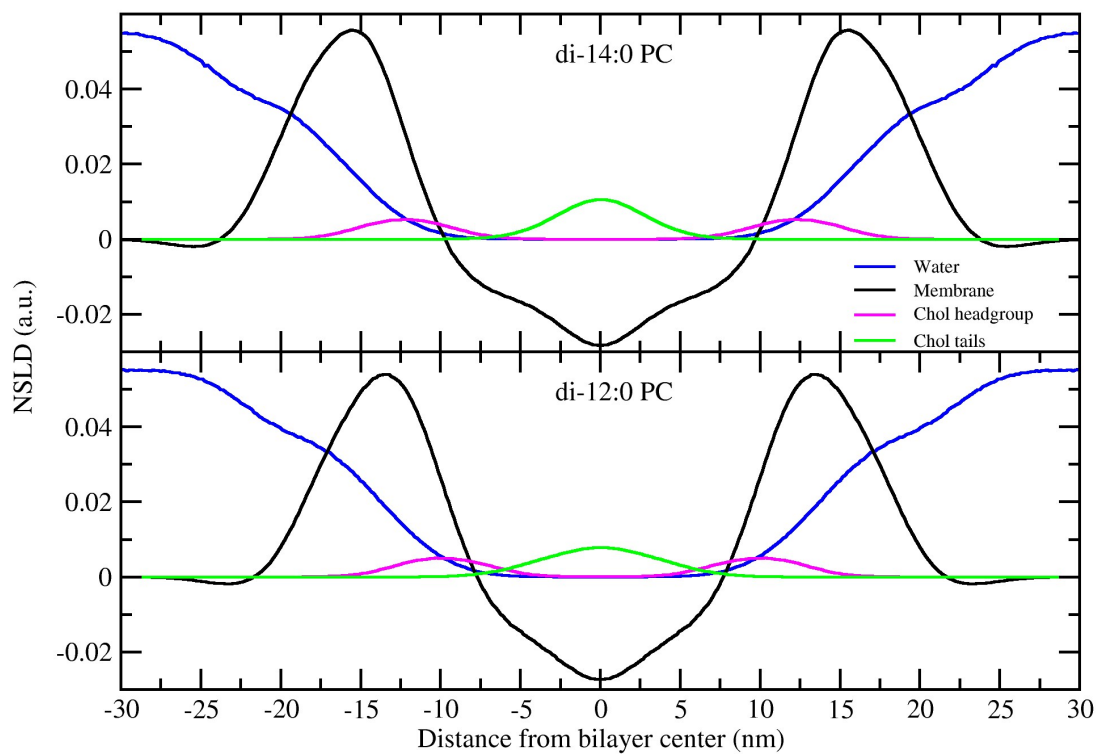


Figure S3 NSLD profiles of bilayers containing cholesterol calculated from MD simulations for DMPC + 10 mol % cholesterol (upper panel), and DLPC + 10 mol % cholesterol (lower panel). Shown are profiles of the lipid (black), water (blue), [2,3,4]-carbons of the cholesterol headgroup (purple), and [25,26,27]-carbons of the cholesterol tail (green).

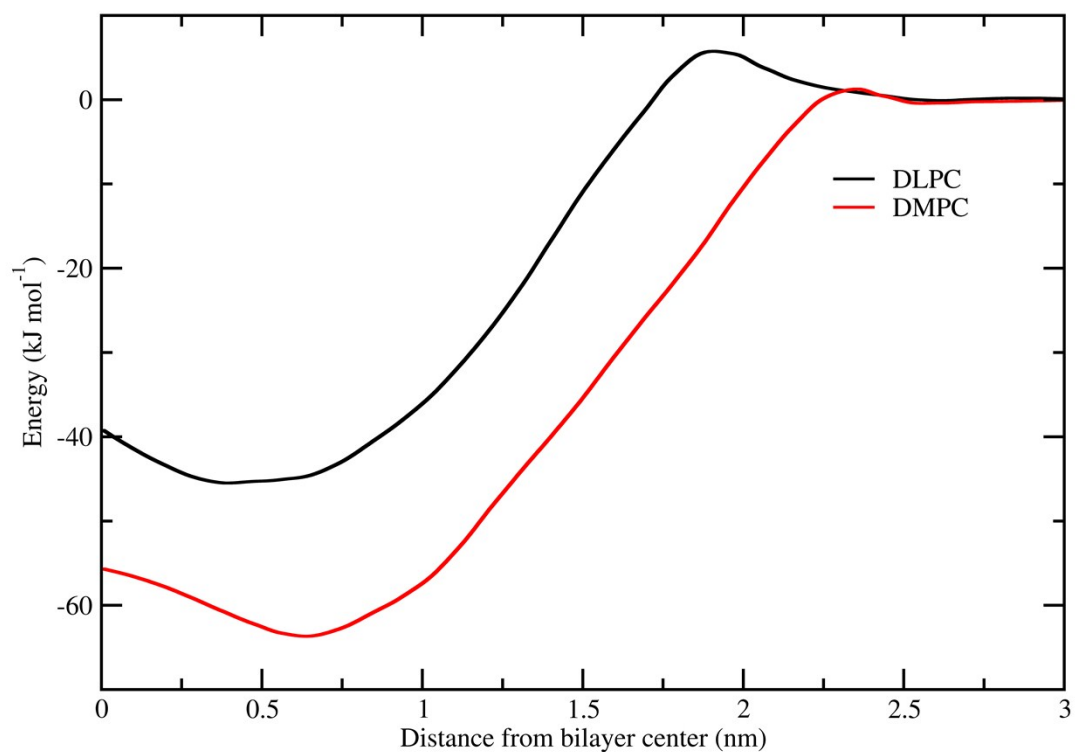


Figure S5 Potentials of mean force calculations reveal a larger energy barrier at the center of the membrane for cholesterol in DMPC (red) compared to DLPC (black) bilayers. The energy barrier for cholesterol flip-flop is 8 kJ/mol in DMPC bilayers, compared to 5.4 kJ/mol in DLPC bilayers.

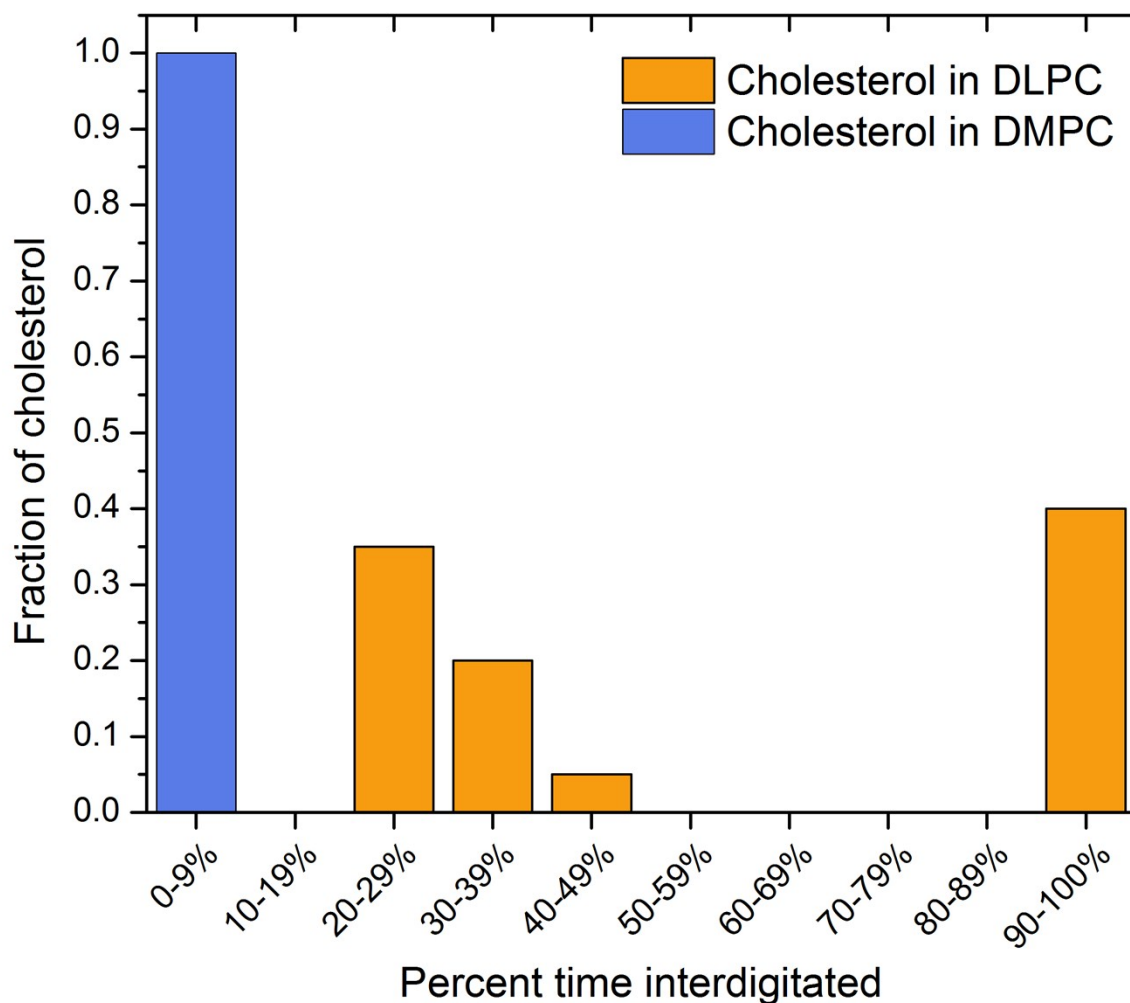


Figure S5 MD results illustrating cholesterol's propensity for interdigitation in DLPC (orange bars) and DMPC (blue bars) bilayers. 20 cholesterol molecules were incorporated into the simulated bilayer. A cholesterol molecule is defined as being interdigitated when at least 25% of cholesterol atoms are present in each bilayer leaflet, simultaneously. The percentages indicated by the plot represent the amount of time cholesterol spends interdigitated, with the size of each bar corresponding to the number of cholesterol molecules.

References

- 1) S. Jo, H. Rui, J.B. Lim, J.B. Klauda and W. Im, Cholesterol Flip-Flop: Insights from Free Energy Simulation Studies. *J. Phys. Chem. B*, 2010, **114**, 13342-13348.
- 2) W.F.D. Bennett, J.L. MacCallum, M.J. Hinner, S.J. Marrink and D.P. Tieleman, Molecular View of Cholesterol Flip-Flop and Chemical Potential in Different Membrane Environments. *J. Am. Chem. Soc.*, 2009, **131**, 12714-12720.