## Enzyme-guided crystal growing surface-enhanced Raman

## scattering active Core shell for bisphenol A detection

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Fig.S1 The dynamic light scattering (DLS) of AuNS at different structures.



Fig.S2 The Vis-NIR spectra of AuNS, DNA2-AuNS, GOx-AuNS and Core/shell.



Fig.S3 The SERS intensity of biosensors with the different concentration of GOx.



Fig.S4 The Vis-NIR spectra after addition of different concentrations of GOx.

Comments: The Vis-NIR spectra (**Fig. S4**) and TEM images (**Fig. S5**) of the core/shell nanostructures with various concentrations of GOx was characterized. As increasing the concentration of GOx in the range from 0 to  $10^{-13}$  g mL<sup>-1</sup>, we could observed the surface plasmon resonance band could display gradually blue shift from ~860 nm to ~680 nm along with gradually thickens of the silver shell. As the superimposing effect of the charge transfer and the enhancement of electromagnetic field, the SERS signals could be gradually increased with increasing the concentration of GOx.



Fig.S5 TEM images of Core/shell structure in the presence of different concentrations of GOx: 0 g mL<sup>-1</sup> (A), 10<sup>-19</sup> g mL<sup>-1</sup> (B), 10<sup>-16</sup> g mL<sup>-1</sup> (C), 10<sup>-13</sup> g mL<sup>-1</sup> (D).



Fig.S6 (A) Reproducibility of the SERS spectra of 4-NTP collected at 10 randomly selected spots on the substrate ( $10^{-15}$  g/mL BPA). (B) Signal intensity of the 1339 cm<sup>-1</sup> line from 4-NTP collected at 10 randomly selected spots on the substrate ( $10^{-15}$  g/mL BPA). SERS detection parameters:  $\lambda_{\text{excitation}} = 633$  nm, accumulation time = 15 s, laser power = 8 mW



Fig.S7 Stability of the SERS spectra of 4-MBA within 7 days (A) (10<sup>-15</sup> g/mL BPA). Signal intensity of the 1339 cm<sup>-1</sup> line from 4-NTP within 7 days (B). SERS detection parameters:  $\lambda_{\text{excitation}} = 633$  nm, accumulation time = 15 s, laser power = 8 mW.



Fig.S8 SERS spectrum of different BPA analogs at the same concentration.

Detection method	Limit of detection	Reference	
GC-MS	62 mg kg <sup>-1</sup>	Food chemistry, 2017, 232, 501-507	
Fluorescence	6.62×10 <sup>-9</sup> g mL <sup>-1</sup>	Biosensors & Bioelectronics, 2017, 92, 147-153	
Fluorescence	9.82×10 <sup>-13</sup> g mL <sup>-1</sup>	Chemical Communications, 2013, 49, 5960-5962	
Plasmonic chiral aptasensor	7.99×10 <sup>-12</sup> g mL <sup>-1</sup>	ACS applied materials & interfaces, 2014, 6, 364-369	
SERS	2.28×10 <sup>-15</sup> g mL <sup>-1</sup>	Biosensors & Bioelectronics, 2015, 64, 560-565	
SERS	$5.00 \times 10^{-17} \text{ g mL}^{-1}$	This work	

Table S1 Comparison of this work with other methods based on aptamer for detecting BPA

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Table S2 Analytical parameters of the proposed method for determining BPA in tap water

Sample	Linear range (g mL <sup>-1</sup> )	Linearity (R <sup>2</sup> )	LOD <sup>a</sup> (g mL <sup>-1</sup> )	RSD <sup>b</sup> (%)	RSD <sup>c</sup> (%)
Tap water	10-16-10-12	0.9901	5×10 <sup>-17</sup>	3.3%	5.7%

<sup>a</sup>LOD for the present method, based on the signal being three times as large as the baseline noise (S/N=3). <sup>b</sup>intraday and n=5. <sup>c</sup>interday and n=5.