

## Halogenation of glycopeptide antibiotics occurs at the amino acid level during non-ribosomal peptide synthesis

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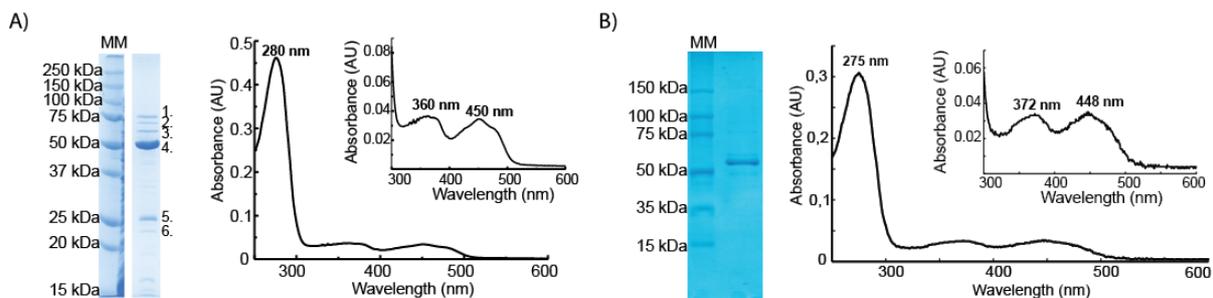
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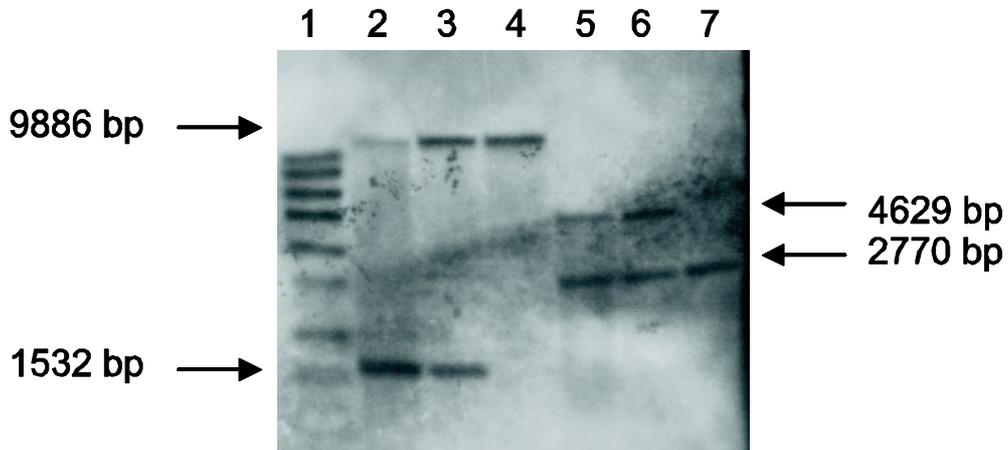
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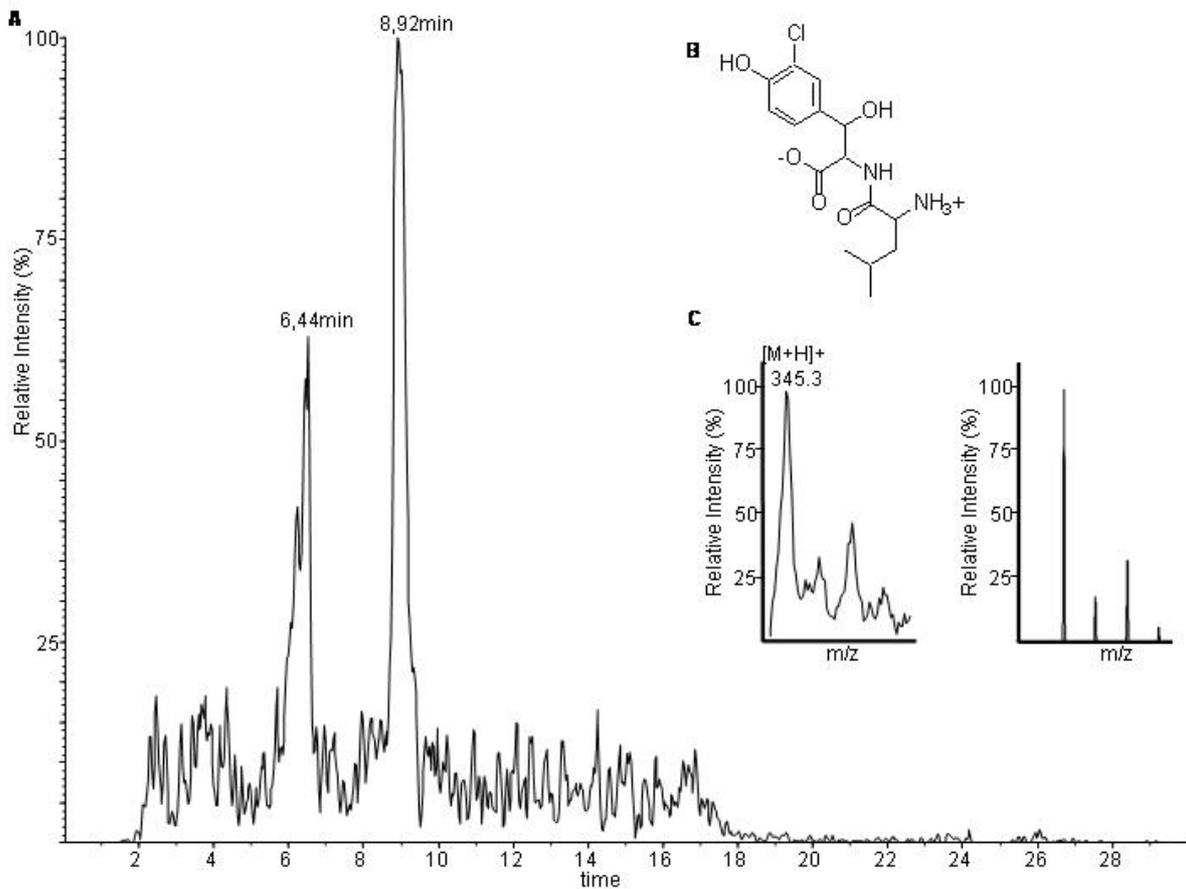
### Supplementary Information



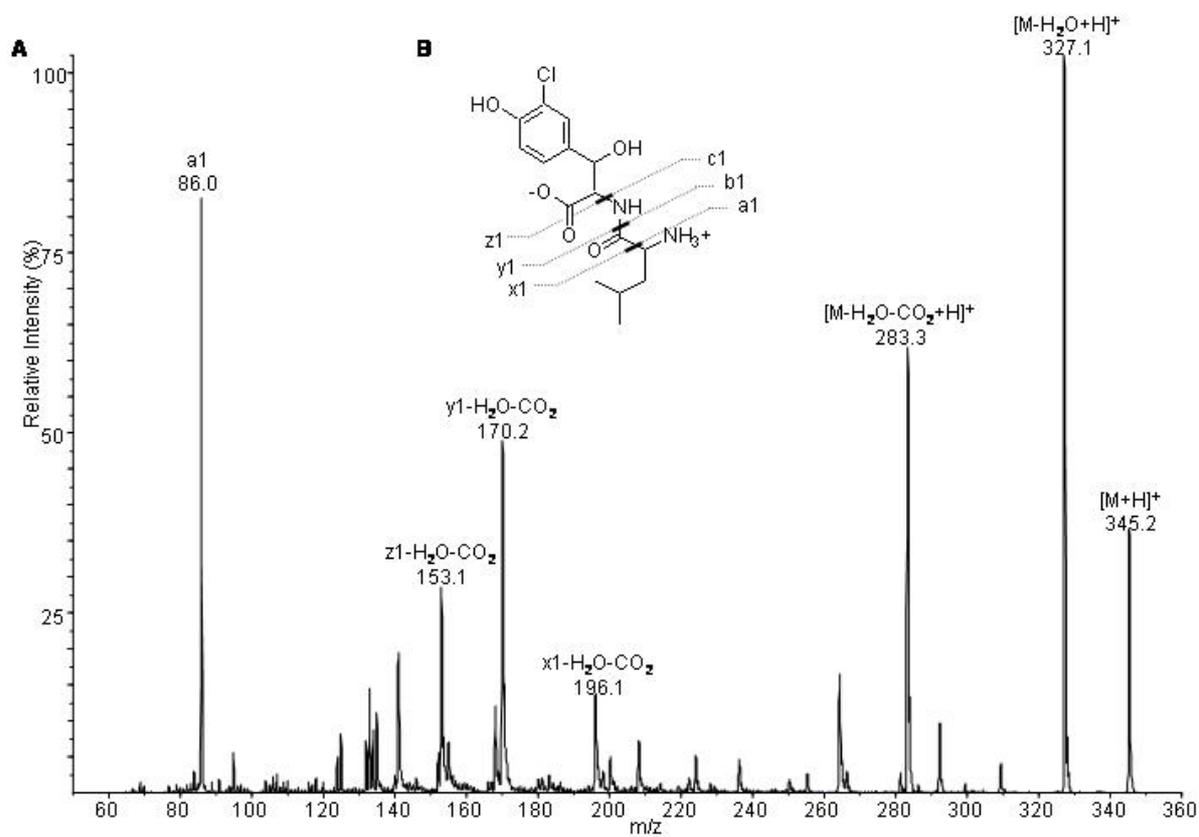
**Figure S1.** Purification and spectral characterisation of halogenases. A) SDS-PAGE and spectra of Tcp21, which was purified in a single affinity chromatography step with a bright yellow colour indicating the presence of FAD. Co-purifying enzymes were analysed but no reduction partner could be identified. Proteins were identified by peptide map fingerprinting as (1) Bi-functional UDP-glucuronic acid decarboxylase; (2) Glucosamine-fructose-6-phosphate aminotransferase; (3) GroEL; (4) Tcp21; (5) Peptidylprolyl isomerase; and (6) Crp/Fnr family transcriptional regulator. B) Representative SDS-PAGE and spectra of BhaA.



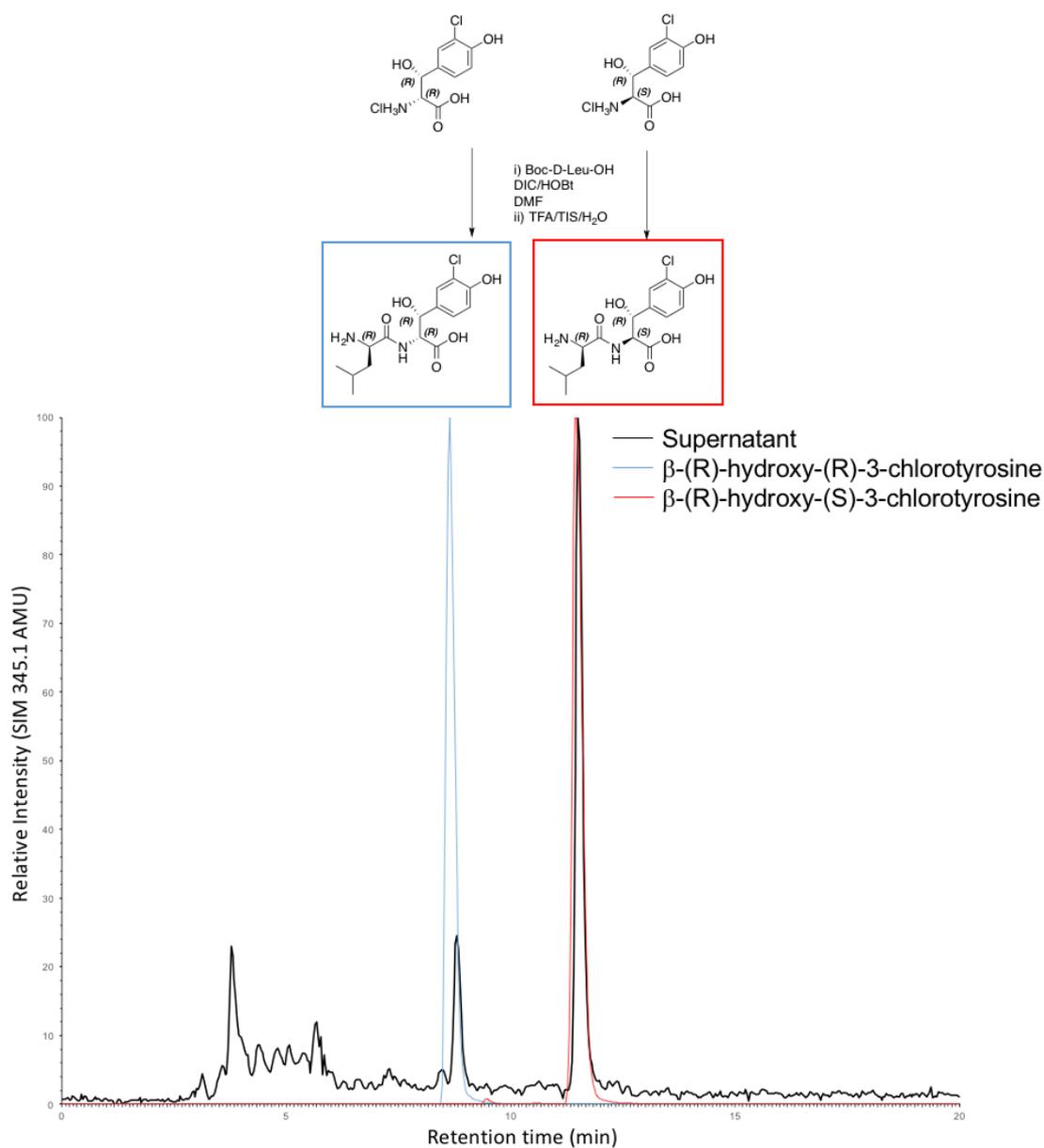
**Figure S2.** Southern blot analyses of wildtype and the truncation mutants CK2.1 and CK2.2. The total DNA was digested either with NcoI (N) or with BamHI. As a probe the TE domain was used, resulting in one band for the wildtype (NcoI: 9886 bp; BamHI: 2770 bp) and in two bands for the mutants (NcoI: 9886bp, 1532 bp; BamHI: 2770 bp, 4629 bp). Lane 1: DIG-labelled DNA Molecular Weight Marker VII; lane 2 and 3: mutants (NcoI), lane 4: WT (NcoI); lane 5 and 6: mutants (BamHI) and lane 7: WT (BamHI).



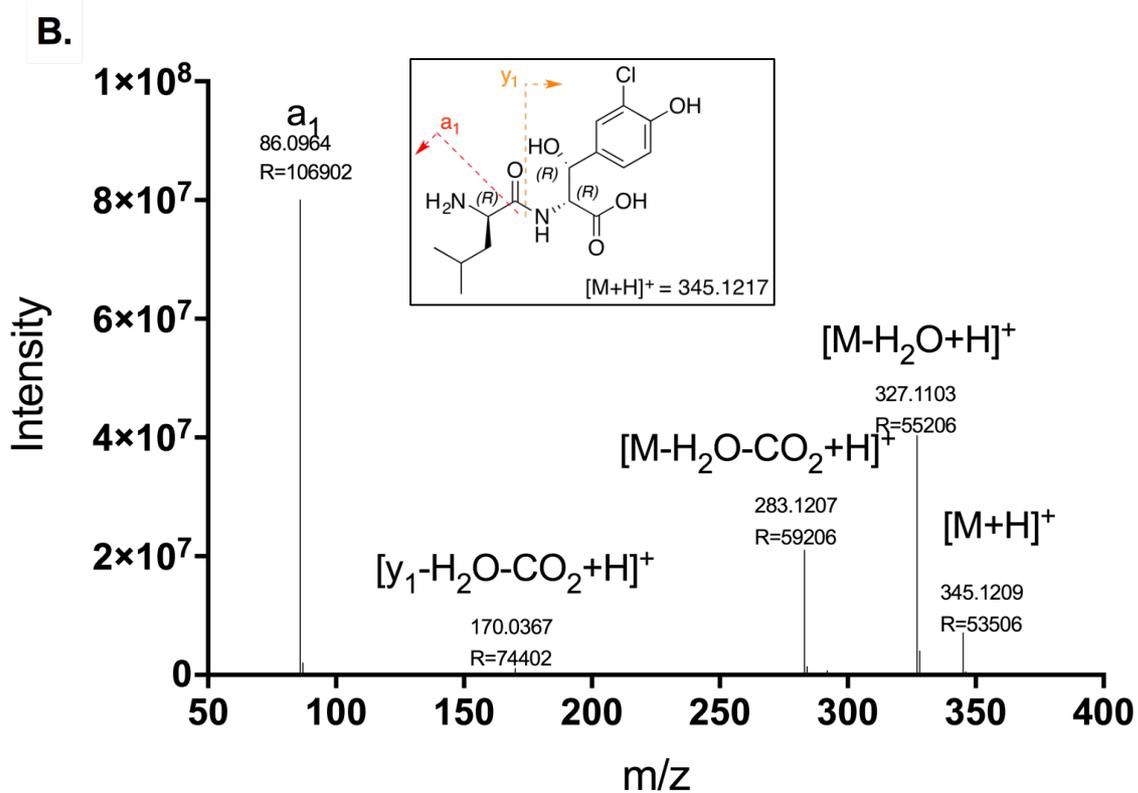
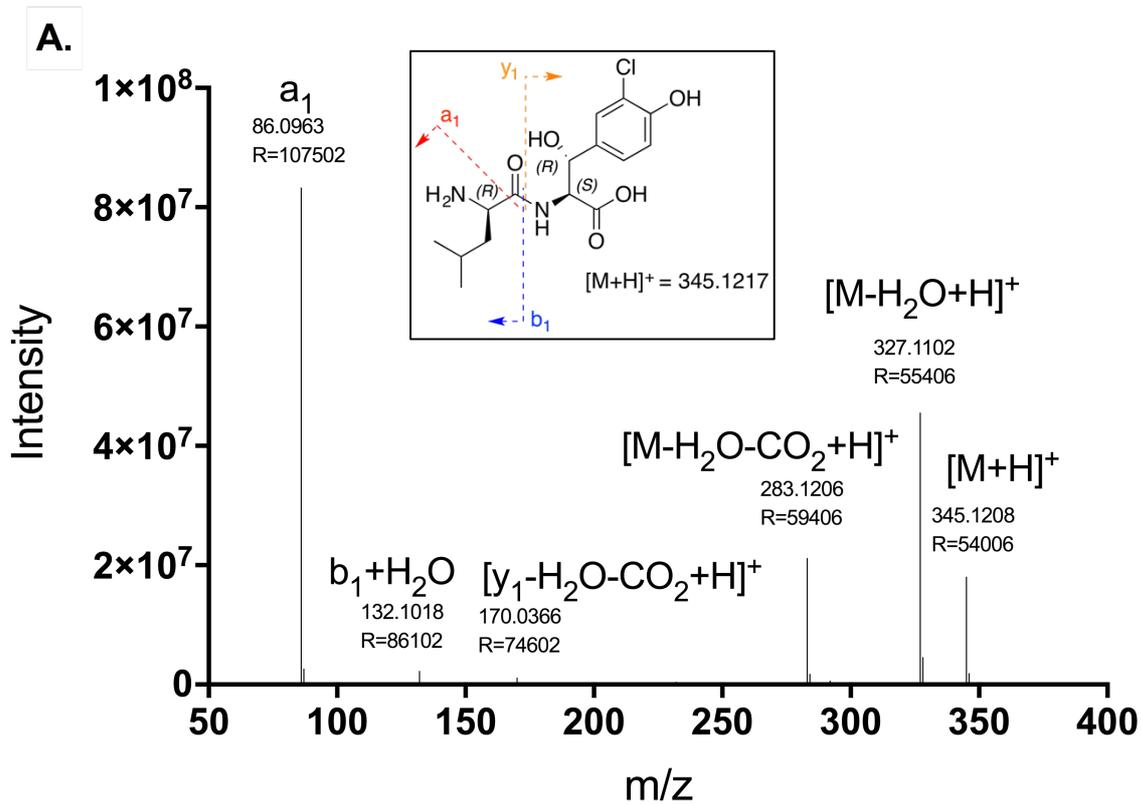
**Figure S3.** (A) Extracted ion chromatogram (EIC, smoothed) shows two retention times of  $m/z$  345.3 of the chlorinated dipeptides. (B) Structure formula of the chlorinated dipeptide. (C) The isotopic pattern found in the spectrum (left) in comparison to the theoretically calculated isotopic pattern (right).



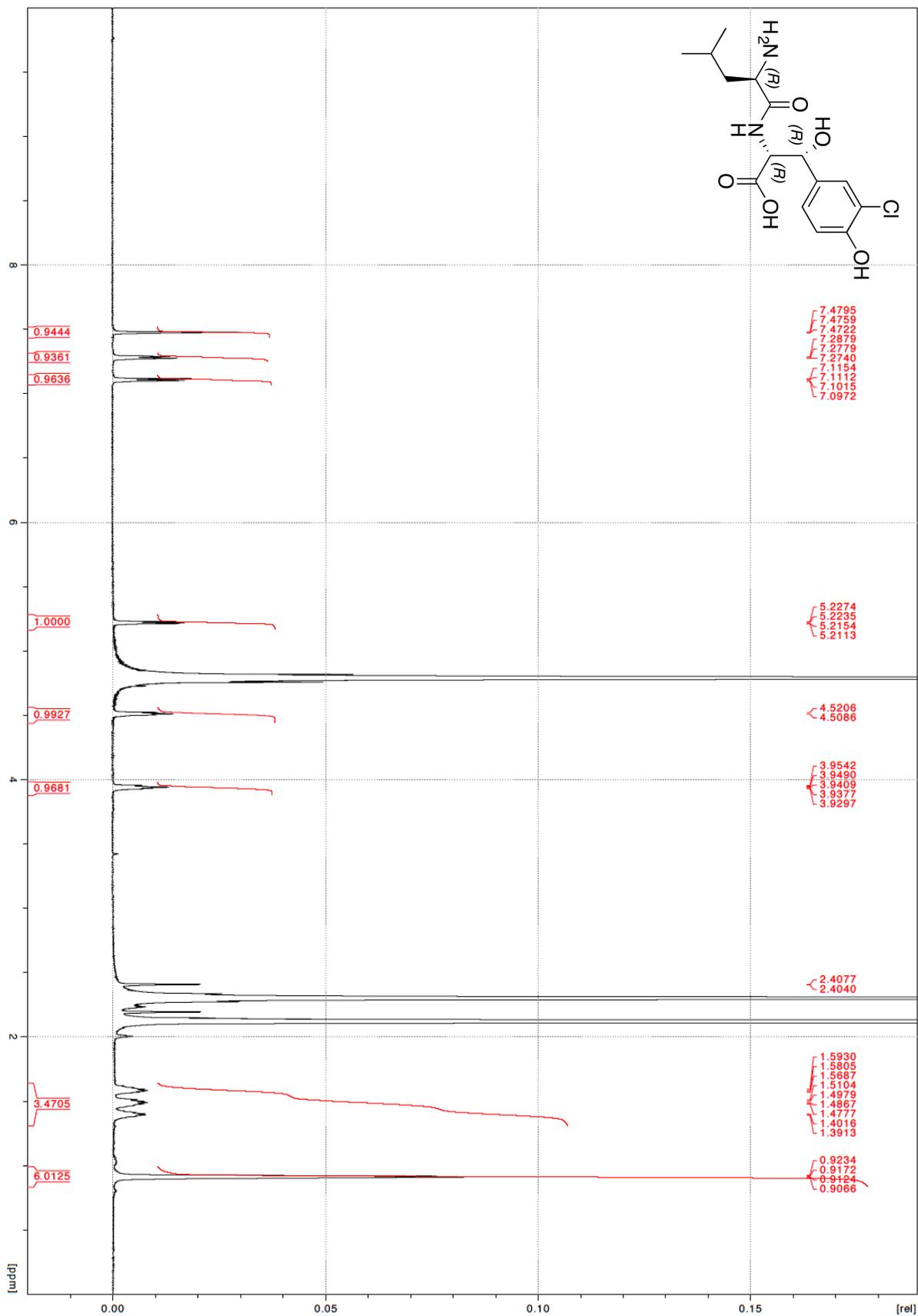
**Figure S4.** (A) Daughter ion spectrum of the chlorinated dipeptide (retention time: 8.92 min) and (B) assignment of the Roepstorff fragmentation along the peptide chain.



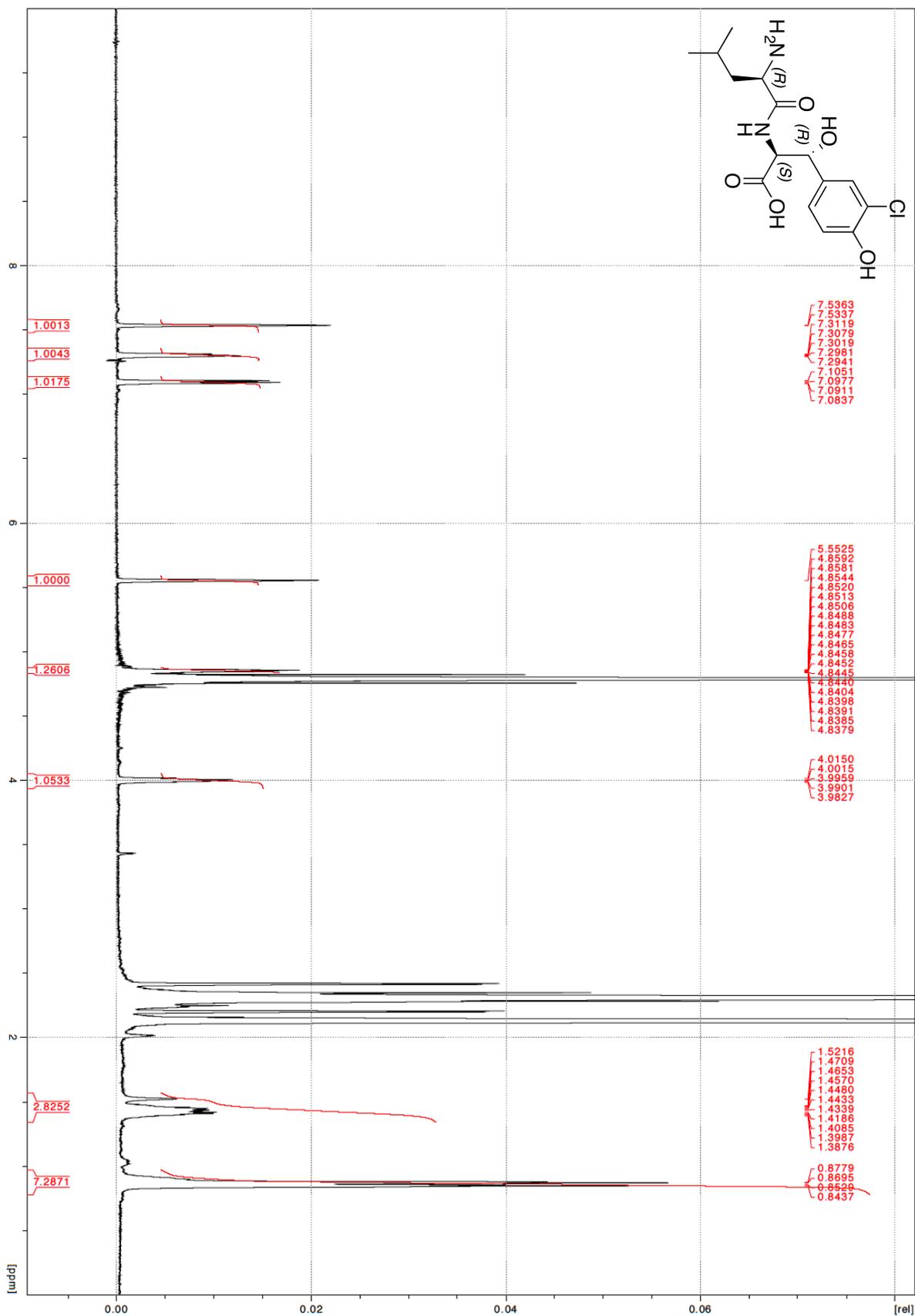
**Figure S5.** LC traces of the culture filtrate of the *A. balhimycina* mutant strain (black) overlaid with the traces of the authentic Leu-OH-Cl-Tyr dipeptide standards containing either (2R, 3R)- $\beta$ -hydroxy-3-chlorotyrosine (blue) or (2S, 3R)- $\beta$ -hydroxy-3-chlorotyrosine (red). Traces shown are in single ion monitoring mode (SIM) at 345.1 AMU, positive mode. Co-injection of synthetic Leu-OH-Cl-Tyr dipeptide standards and supernatant confirmed the retention time of the standards and dipeptide produced by the mutant strain are identical.



**Figure S6.** MS<sup>2</sup> fragmentation of authentic Leu-OH-Cl-Tyr dipeptide standards containing (2S, 3R)-β-hydroxy-3-chlorotyrosine (A) or (2R, 3R)-β-hydroxy-3-chlorotyrosine (B) with fragmentation pattern displayed for each dipeptide (boxed).



**Figure S7.**  $^1\text{H}$  NMR (600MHz;  $\text{D}_2\text{O}$ ) of authentic Leu-OH-Cl-Tyr dipeptide standard containing (2R, 3R)- $\beta$ -hydroxy-3-chlorotyrosine.



**Figure S8.** <sup>1</sup>H NMR (600MHz; D<sub>2</sub>O) of authentic Leu-OH-Cl-Tyr dipeptide standard containing (2S, 3R)-β-hydroxy-3-chlorotyrosine.