

Supporting Information

A dual signal amplification method for exosome detection based on DNA dendrimer self-assembly

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Supporting figures and table

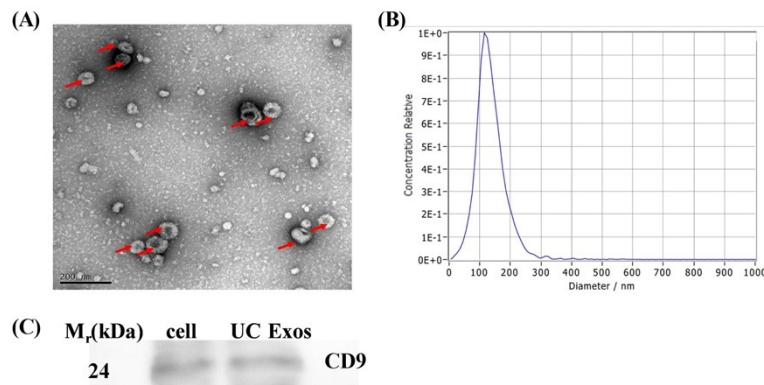


Fig. S1. Characterization of exosomes purified from HepG2 cell culture supernatant. (A) Isolated exosomes were identified by transmission electron microscopy. The red arrows indicate exosomes with a mean diameter of ~100 nm. (B) Size distribution of exosomes by dynamic light scattering measurements. (C) Western blotting images of CD9 proteins from HepG2 cell lysates and exosomes.

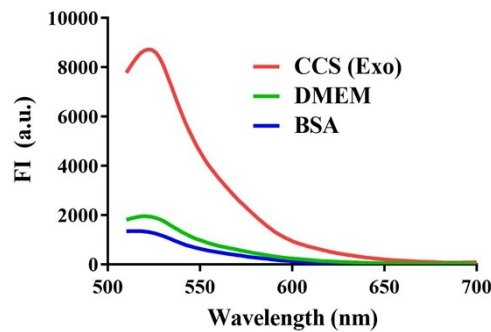


Fig. S2. Fluorescence spectrum of exosome capture process in the presence of CCS, DMEM and 0.1% BSA.

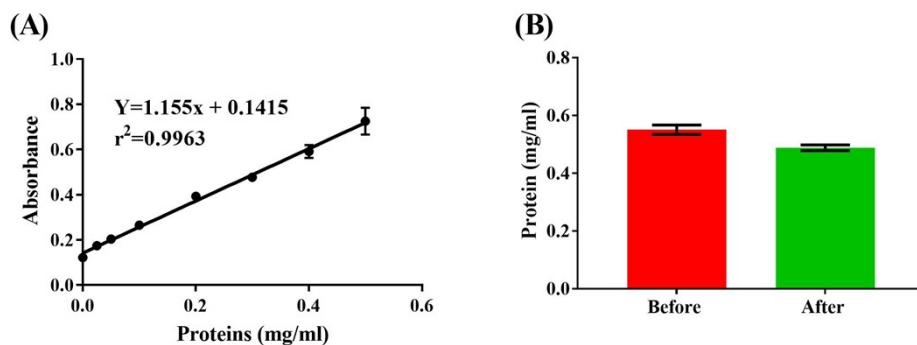


Fig. S3. (A) Standard work curve of the concentration of exosomal proteins and absorbance value obtained by BCA protein quantitation kit. (B) Quantification of the exosomal protein

concentration before the capture (before) and the protein concentration of the captured exosomes (after), using BCA protein quantitation kit.

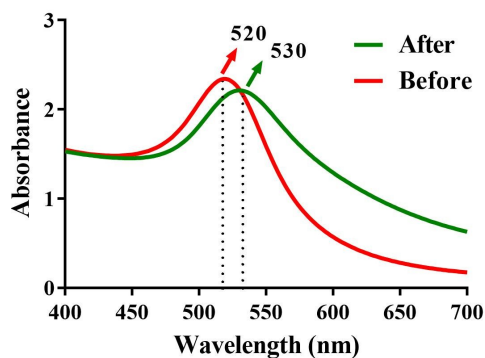


Fig. S4. Absorption spectra of the AuNPs before and after modification of HP₁.

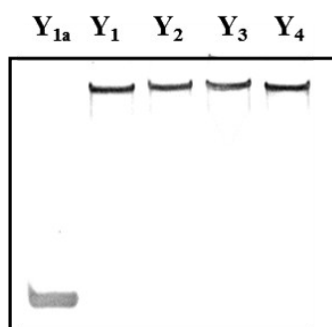


Fig. S5. Native PAGE analysis of Y₁, Y₂, Y₃, and Y₄, while Y_{1a} is as control.

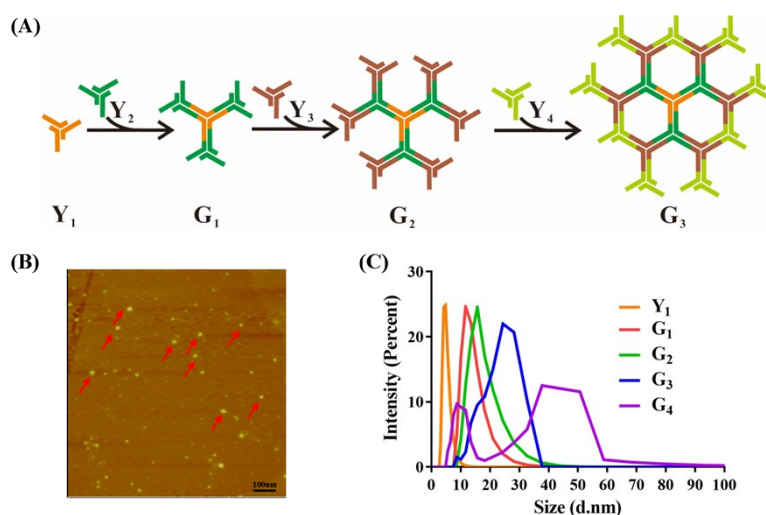


Fig. S6. (A) Schematic illustration of DNA dendrimers generation. (B) AFM imaging for the last generation G₃. (C) DLS experiments for particle size analysis for different dendrimers.

Table S1. Sequences of oligonucleotides used in this work

Name	^a Sequence (5'-3')
A ₁	<i>ATATACACCCACCTCGCTCCCGTGACACTAATGCTATTTTTT</i> -Biotin
S ₁	<i>TGGGGTGTATAT</i> -FAM
A ₂	<i>CACCCACCTCGCTCCCGTGACACTAATGCTATTTTTT</i> -Biotin
S ₂	<i>CACGGGAGCGAGATATA</i> -FAM
S ₃	<u><i>TGGGGTGTATATTT</i></u> <i>CATATATATA</i>
R	ATATACACCCATTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTTT
HP ₁	<i>CACACACAATATTT</i> <u><i>CATATATATATTGATGTGTACCTATATATATGAA</i></u> <i>ATATACACCCATTTTTT</i> -SH
HP ₂	<i>TTGATGTGTACCATATTT</i> <u><i>CATATATATAGGTACACATCAATATATATA-</i></u> <u><i>FAM</i></u>
Y _{1a}	<i>TGAAATAT TGTGTGTG CCTGTCTGCCTAA TGTGCGTCGTAAG</i> -FAM
Y _{1b}	<i>CTGTCATCGGTCA CTTACGACGCACA AGGAGATCATGAG</i>
Y _{1c}	<i>CTGTCATCGGTCA CTCATGATCTCCT TAGGCAGACAGG</i>
Y _{2a}	<i>TGACCGATGACAG CCTGTCTGCCTAA TGTGCGTCGTAAG</i> -FAM
Y _{2b}	<i>GACACACTGAGGT CTTACGACGCACA AGGAGATCATGAG</i>
Y _{2c}	<i>GACACACTGAGGT CTCATGATCTCCT TTAGGCAGACAGG</i>
Y _{3a}	<i>ACCTCAGTGTGTC CCTGTCTGCCTAA TGTGCGTCGTAAG</i> -FAM
Y _{3b}	<i>TGCTGTCTGTCCA CTTACGACGCACA AGGAGATCATGAG</i>
Y _{3c}	<i>TGCTGTCTGTCCA CTCATGATCTCCT TTAGGCAGACAGG</i>
Y _{4a}	<i>TGGACAGACAGCA CCTGTCTGCCTAA TGTGCGTCGTAAG</i> -FAM
Y _{4b}	<i>AGGTCAGAACTGT CTTACGACGCACA AGGAGATCATGAG</i>
Y _{4c}	<i>AGGTCAGAACTGT CTCATGATCTCCT TTAGGCAGACAGG</i>

^a The color of the characters corresponds to the color of probes illustrated in Scheme 1. Italicized characters in A₁, S₁, A₂, and S₂ represent the bases complementary with each other. Underlined characters in S₃ represent the sequence that can open the hairpin structure of HP₁. Underlined characters in HP₂ represent the toehold that is complementary with the hairpin structure of HP₁.