Electronic Supplementary Material (ESI) for Biomaterials Science.


Supporting Information

# Celastrol-loaded PEG-*b*-PPS nanocarriers as an anti-inflammatory treatment for atherosclerosis

Sean D. Allen, Yu-Gang Liu, Taehyeung Kim, Sharan Bobbala, Sijia Yi, Xiaohan Zhang, Jaehyuk Choi, and Evan A. Scott

This Supporting Information Includes:

Figure S1. Confocal images of RAW 264.7 cells not treated with DiI-labeled micelles.

Figure S2. Mouse body weight and food consumption analysis.

Figure S3. Additional flow cytometric cell population comparisons between treatment groups.

Figure S4. Flow cytometry gating strategy contour plots.

**Fig. S1. Confocal images of RAW 264.7 cells not treated with DiI-labeled micelles. RAW 264.7 cells stained with Hoechst 33342 for nuclei and LysoTracker Green for lysosomes. As cells were not treated with DiI-labeled micelles, the red channel is devoid of signal, demonstrating low bleed through of LysoTracker Green signal.**

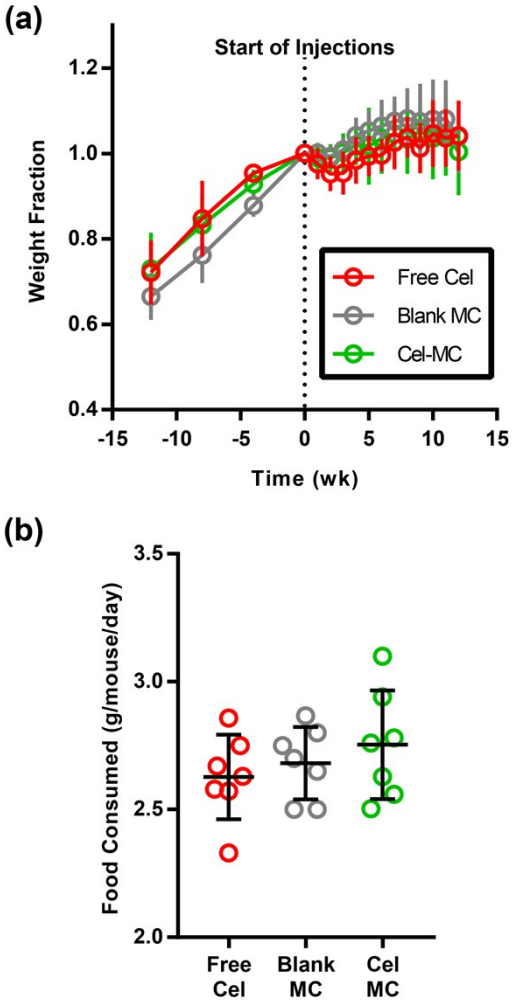


**Fig. S2. Mouse body weight and food consumption analysis.** (a) Average mouse body weights in the weeks before and after the initiation of treatment with free celastrol, Blank MC, and Cel-MC. n=8 for free celastrol and Blank MC groups and n = 9 for Cel-MC group. Error bars are standard deviation, x-axis time 0 is the initiation of treatment. (b) Average food consumed during treatment by mice within the three treatment groups, n=7 for all treatment groups, bars represent the mean and standard deviation.

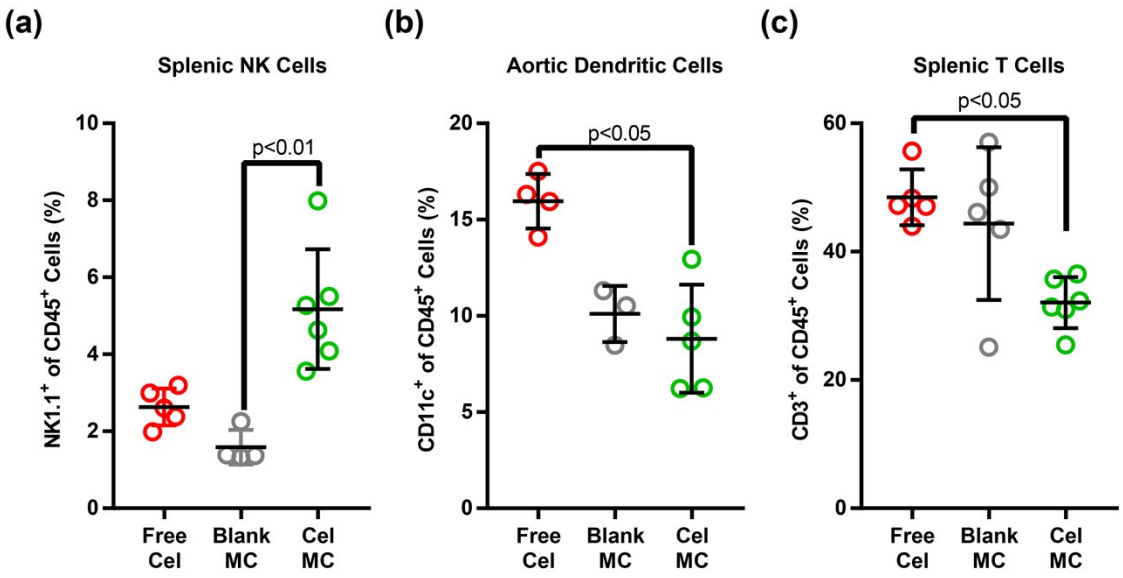


**Fig. S3. Additional flow cytometric cell population comparisons between treatment groups.** Comparison of each cell population as a percent of CD45+ cells in that organ between free celastrol, Blank MC, and Cel-MC treatments for (a) splenic NK cells, (b) aortic dendritic cells, and (c) splenic T cells. P values obtained using Dunn's multiple comparison test, n=6. Bars represent the mean and standard deviation. All data points are shown on graphs.

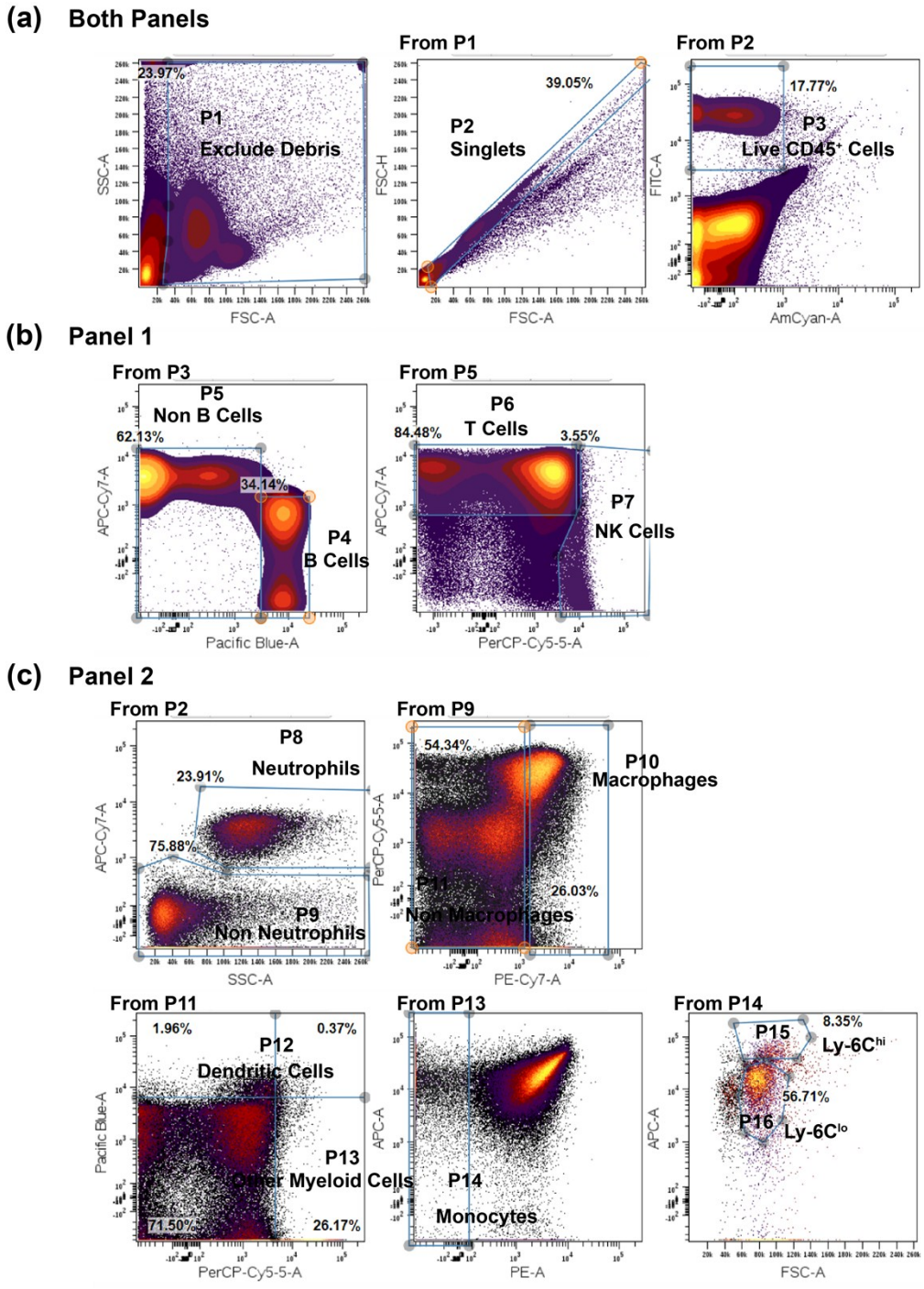


**Fig. S4. Flow cytometry gating strategy contour plots.** Gating strategy for flow cytometry for (a) both panels, (b) panel 1, and (c) panel 2.