Facile Cu(II) mediated conjugation of thioesters and thioacids to peptides and proteins under mild conditions

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Figure S1. The MTT experiment on L929 cell lines for Cu(OAc)₂ and HOBt.



Figure S2. ESI-MS of Cy5.5COSH



Figure S3. ESI-MS of Cy5.5-RGDfK







Figure S5. MALDI-TOF-MS of Cy5.5-AE105



Figure S6. ESI-MS of Cy5.5-E[c(RGDyK)₂]

General procedure for the modification of Ub protein with thioester and the sample preparation for MS analysis.

The thioester (100 to 200 eq) and Ub (86.0 μ g, 0.01 μ mmol, 1 eq) were dissolved in the mixture of PBS/DMF (5:5 to 7:3, total 150 μ L). Then Cu(OAc)₂ (2.00 μ g to 4.00 μ g, 1 eq to 2 eq) and HOBt (13.5 μ g, 10 eq) were added and the reaction mixture was stirred at room temperature for 6 h to 16 h. The crude product was in-gel digested with trypsin overnight. The peptides mixtures were extracted with extraction buffer (5 % FA and 50 % ACN in water) and analyzed through LC-MS/MS.

The crude protein was purified by SDS-PAGE. The gel was stained with Coomassie Blue G-250 and cut into about 1 mm³ cubes, followed by destaining and in-gel digestion with 10 ng/ μ L of trypsin at 37 °C overnight. The peptides were extracted with extraction buffer (5 % FA and 50 % ACN in water) and then ACN and finally dried. Peptide mixtures were loaded onto an in-house packed capillary column (75 μ m I.D. and 15 cm) with 3 μ m C18 reverse-phase fused-silica (Michrom Bioresources, Inc., Auburn, CA) with a flow rate of 0.3 μ L/min, and eluted with a 100 minutes gradient developed as follows: 0-5 % B for 10 minutes, 5-10 % B for 10 minutes, 10-20 % B for 30 minutes, 20-45 % B for 40 minutes, and 45-80 % B for 10 minutes (Buffer A: 0.1 % FA; Buffer B: 0.1 % FA and 100 % ACN).



Figure S7. The result of MALDI-TOF-MS spectrum of Ub modification with 100 eq of Cbz-L-Phe thioester under $1 eq Cu(OAc)_2$ and 10 eq HOBt



Figure S8. The result of MALDI-TOF-MS spectrum of Ub modification with Cbz-L-Phe thioester under 2eq Cu(OAc)₂, 200 eq thioester and 16 h conditions.



Figure S9. LC-MS spectrum of the resulting peptide fragment of trypsinized Ub conjugated with Cbz-L-Phe thioester.



Figure S10. MS and MS_2 spectrums of the peptide fragment of Ub K_{33} modified with Cbz-L-Phe thioester.



Figure S11. MS and MS_2 spectrums of the peptide fragment of Ub K_{48} modified with Cbz-L-Phe thioester.



Figure S12. MS and MS_2 spectrums of the peptide fragment of Ub K_6 modified with biotin thioester.



Figure S13. MS and MS_2 spectrums of the peptide fragment of Ub K_{33} modified with biotin thioester.



Figure S14. MS and MS_2 spectrums of the peptide fragment of Ub K_{48} modified with biotin thioester.

General procedure for the modification of Ub protein with thioacid and the sample preparation for MS analysis.

The thioacid (100 to 200 eq) and Ub (86.0 μ g, 0.01 μ mmol, 1 eq) were dissolved in the mixture of PBS/DMF (5:5, total 150 μ L) or PBS. Then Cu(OAc)₂ (2.00 μ g to 4.00 μ g, 1 eq to 2 eq) and HOBt (2 eq to 4 eq) were added and the reaction mixture was stirred at room temperature for 6 h. The crude product was in-gel digested with trypsin overnight. The peptides mixtures were extracted with extraction buffer (5 % FA and 50 % ACN in water) and analyzed through LC-MS/MS.

The crude protein was purified by SDS-PAGE. The gel was stained with Coomassie Blue G-250 and cut into about 1 mm³ cubes, followed by destaining and in-gel digestion with 10 ng/ μ L of trypsin at 37 °C overnight. The peptides were extracted with extraction buffer (5 % FA and 50 % ACN in water) and then ACN and finally dried. Peptide mixtures were loaded onto an in-house packed capillary column (75 μ m I.D. and 15 cm) with 3 μ m C18 reverse-phase fused-silica (Michrom Bioresources, Inc., Auburn, CA) with a flow rate of 0.3 μ L/min, and eluted with a 100 minutes gradient developed as follows: 0-5 % B for 10 minutes, 5-10 % B for 10 minutes, 10-20 % B for 30 minutes, 20-45 % B for 40 minutes, and 45-80 % B for 10 minutes (Buffer A: 0.1 % FA; Buffer B: 0.1 % FA and 100 % ACN).



Figure S15. MALDI-TOF-MS spectrum result of Ub modification with thioacid under 1eq $Cu(OAc)_2$, 2 eq HOBt, 100 eq thioester and 6 h conditions.



Figure S16. MALDI-TOF-MS spectrum result of Ub modification with thioacid in PBS solution.

Peptide sequence	Mass(z=1)	Mass(z=2)
*MQIFVK ₆	849.453	425.230

Ub-CH3COOSH #15196 RT: 54.42 AV: 1 NL: 2.24E5 T: FTMS + c NSI Full ms [300.00-1600.00]



Figure S17. MS and MS_2 spectrums of the peptide fragements of Ub N-terminal and K_6 modified with thioacid.

Peptide sequence	Mass(z=1)	Mass(z=2)
TLTGK11TITLEVEPSDTIENVK	2330.233	1165.620

Ub-CH3COOSH #12429 RT: 44.77 AV: 1 NL: 1.55E7 T: FTMS + c NSI Full ms [300.00-1600.00]



Figure S18. MS and MS_2 spectrums of the peptide fragments of Ub K_{11} modified with thioacid.

Peptide sequence	Mass(z=1)	Mass(z=2)
TITLEVEPSDTIENVKAK29	2029.069	1015.038

Ub-CH3COOSH #11600 RT: 42.39 AV: 1 NL: 4.50E7 T: FTMS + c NSI Full ms [300.00-1600.00]



Figure S19. MS and MS_2 spectrums of the peptide fragments of Ub K_{29} modified with thioacid.

Peptide sequence	Mass(z=1)	Mass(z=2)
IQDK33EGIPPDQQR	1565.791	783.399



Figure S20. MS and MS_2 spectrums of the peptide fragments of Ub K_{33} modified with thioacid.



Figure S21. MS and MS_2 spectrums of the peptide fragments of Ub K48 modified with thioacid.



Figure S22. MS and MS_2 spectrums of the peptide fragments of Ub K63 modified with thioacid.

General methods

All chemicals were purchased from commercial sources (such as Aldrich, conju-probe, Lumiprobe and peptide international). The ¹H and ¹³C NMR spectra were acquired on a Bruker 400 MHz magnetic resonance spectrometer. Data for ¹H NMR spectra are reported as follows: chemical shifts are reported as δ in units of parts per million (ppm) relative to chloroform-d (δ 7.26, s); multiplicities are reported as follows: s (singlet), d (doublet), t (triplet), q (quartet), dd (doublet of doublets), m (multiplet), or br (broadened); coupling constants are reported as a *J* value in Hertz (Hz); the number of protons (n) for a given resonance is indicated nH, and based on the spectral integration values. MALDI-MS spectrometric analyses were performed on an Applied Biosystems 4700 MALDI TOF mass spectrometer. HPLC was performed on a Dionex HPLC System (Dionex Corporation) and a reversed-phase C18 column was used for analysis (Phenomenax, 5 µm, 4.6 mm × 250 mm) and semi-preparation (Agilent, 5 µm, 10 mm × 250 mm).

Chemical synthesis and characterization



Synthesis of 1a: The mixture of Boc-Phe-*L*-amino acid (265.0 mg, 998.9 µmol), 4-Methylbenzenethiol (136.4 mg, 1.098 mmol), diisopropylcarbodiimide (189.3 mg, 1500 µmol) and 1-hydroxybenzotriazole (202.6 mg, 1500 µmol) were dissolved in ethyl acetate (2000 µL). The reaction mixture was stirred for overnight and purified by flash chromatography affording the desired product as a white solid (315.4 mg, 85 %). ¹H NMR (400 MHz, CDCl₃) δ = 7.37-7.21 (m, 9H), 5.00 (d, *J*=12.0, 1H), 4.79-4.77 (m, 1H), 3.20-3.14 (m, 2H), 2.40 (s, 3H), 1.46 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ = 199.7, 154.9, 139.8, 135.6, 134.5, 130.1, 129.4, 128.6, 127.1, 123.7, 119.0, 80.4, 60.8, 38.4, 28.3, 21.3; HRMS (ESI) Calcd for: C₂₁H₂₆NO₃S: 372.1628. Found: 372.1622 ([M+H]⁺).



Synthesis of 1b: The mixture of biotin (244.0 mg, 998.9 μmol), 4-Methylbenzenethiol (136.4 mg, 1.098 mmol) diisopropylcarbodiimide (189.3 mg, 1500 μmol) and 1-hydroxybenzotriazole (202.6 mg, 1500 μmol) were dissolved in DMF (2000 μL). The reaction mixture was stirred for overnight and purified by flash chromatography affording the desired product as a white solid (273.1 mg, 78 %). ¹H NMR (400 MHz, CDCl₃) δ = 8.27 (br, 1H), 7.62 (d, *J*=8.0, 1H), 7.40 (d, *J*=8.0, 1H), 7.27-7.14 (m, 6H), 7.06 (br, 1H), 5.16 (d, *J*=8.0, 1H), 4.86-4.81 (m, 1H), 3.45-3.29 (m, 2H), 2.39 (s, 3H), 1.48 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ = 200.6, 155.2, 139.7, 136.1, 134.6,

130.0, 127.6, 123.8, 123.2, 119.8, 119.0, 111.2, 109.7, 80.4, 60.5, 28.3, 28.1, 21.3; HRMS (ESI) Calcd for: C₁₇H₂₃N₂O₂S₂: 351.1195. Found: 351.1193 ([M+H]⁺).



Synthesis of 1c: The mixture of Cbz-Phe-*L*-amino acid (299.0 mg, 999.0 µmol), 4-Methylbenzenethiol (136.4 mg, 1.098 mmol) diisopropylcarbodiimide (189.3 mg, 1500 µmol) and 1-hydroxybenzotriazole (202.6 mg, 1500 µmol) were dissolved in ethyl acetate (2000 µL). The reaction mixture was stirred for overnight and purified by flash chromatography affording the desired product as a white solid (344.3 mg, 85%). ¹H NMR (400 MHz, CDCl₃) δ = 7.29-7.08 (m, 14H), 5.17 (d, *J*=8.0, 1H), 5.04 (s, 2H), 4.78-4.73 (m, 1H), 3.12-3.02 (m, 2H), 2.30 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ = 199.1, 155.6, 140.0, 136.0, 135.3, 134.5, 130.1, 129.4, 128.7, 128.5, 128.3, 128.1, 127.3, 123.4, 67.3, 61.3, 38.4, 21.4,; HRMS (ESI) Calcd for: C₂₄H₂₄NO₃S: 406.1471. Found: 406.1471 ([M+H]⁺).



Synthesis of 1d: The mixture of Boc-L-Phe-Gly-OH (322.0 mg, 999.1 µmol), 4-Methylbenzenethiol (136.4 mg, 1.098 mmol) diisopropylcarbodiimide (189.3 mg, 1500 µmol) and 1-hydroxybenzotriazole (202.6 mg, 1500 µmol) were dissolved in ethyl acetate (2000 µL). The reaction mixture was stirred for overnight and purified by flash chromatography affording the desired product as a white solid (325.4 mg, 76 %). ¹H NMR (400 MHz, CDCl₃) δ = 8.21(s, 1H), 7.61 (d, *J*=8, 1H), 7.41 (d, *J*=8, 1H), 7.26-7.08 (m, 7H), 5.14 (d, *J*=12, 1H), 4.85-4.80 (m, 1H), 3.45-3.28 (m, 2H), 2.39 (s, 3H), 1.47 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) δ = 199.8, 139.8, 135.6,

134.5, 130.1, 129.4, 128.7, 127.1, 123.7; HRMS (ESI) Calcd for: C₂₃H₂₈N₂O₄S: 429.1843. Found: 429.1843 ([M+H]⁺).



Synthesis of Cy5.5COSH: The Cy5.5-NHS (2.100 mg, 2.932 µmol) was dissolved in distilled DMF (300.0 µL). NaSH (300.0 µg, 5.355 µmol) was added to this stirring solution under N₂ atmosphere. The reaction mixture was allowed to stir for 4h. Then the reaction mixture was purified by HPLC to get the desired Cy5.5 thioacid as a blue solid . ESI-MS Calcd for:C₄₀H₄₃N₂OS: 599.3,Found: 599.5 ([M+H]⁺).



Synthesis of 3a: Boc-Phe-*p*-toluene thiol ester (371.0 mg, 998.7 µmol) was added to DMF (2000 µL) or DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added catalyst (1 eq) or catalyst and HOBt (1eq or 2eq) and Try-OMe (327.0 mg, 1.498 mmol) in it. The mixture was stirred at 37 °C, then cooled to room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate (15.00 mL) then washed successively with 1M aqueous HCl, 2M aqueous NaOH and brine. The organic layer was dried with Na₂SO₄, filtered, and concentrated to dryness. The crude product was purified by flash chromatography (first use 25% ethyl acetate in petroleum ether, then use 10% MeOH in DCM wash the chromatography to get the product. The title compound was prepared as a white solid.

¹H NMR (400 MHz, CDCl₃) $\delta = 8.20$ (br, 1H),7.36-7.09 (m, 10H), 6.41 (br, 1H), 4.84-4.76 (m, 2H), 4.36 (br, 1H), 3.64 (s, 3H), 3.28-3.27 (m, 2H), 3.04-3.03 (m, 2H), 1.38 (s, 9H); ¹³C NMR (101 MHz, CDCl₃) $\delta = 171.7$, 170.8, 136.5, 136.0, 129.4, 128.6, 126.9, 122.9, 122.2, 119.6, 118.4, 111.3, 109.7, 52.9, 52.3, 38.3, 28.2, 27.6; HRMS (ESI) Calcd for: C₂₆H₃₂N₃O₅: 466.2336. Found: 466.2347 ([M+H]⁺).



Synthesis of 3b: Boc-Phe-*p*-toluene thiol ester (371.0 mg, 998.7 μmol) was added to DMF/PBS (2000 μL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and Gly (116.0 mg, 1.125 mmol) in it. The mixture was stirred at 37 °C for 1h, then cooled to room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate then washed successively with 1M aqueous HCl, 2M aqueous NaOH and brine. The organic layer was dried with Na₂SO₄, filtered, and concentrated to dryness. The crude product was purified by flash chromatography (first use 25% ethyl acetate in petroleum ether, then use 10% MeOH in DCM wash the chromatography to get the product). The title compound was prepared as a white solid (308.9 mg, 88 %). ¹H NMR (400 MHz, CDCl₃) δ = 7.22-6.79 (m, 5H), 6.79 (br, 1H), 5.25-5.22 (m, 1H), 4.40 (br, 1H), 4.13 (dd, J_1 =8.0, J_2 =16.0, 2H), 4.38 (br, 1H), 3.92-3.86 (m, 2H), 3.09-2.91 (m, 2H), 1.30 (s, 9H), 1.20 (t, J=4.0, 3H), ¹³C NMR (101 MHz, CDCl₃) δ = 171.5, 170.4, 169.4, 136.6, 129.2, 128.5, 126.9, 61.5, 54.2, 41.3, 38.2, 23.0, 14.1; HRMS (ESI) Calcd for: C₁₆H₂₃N₂O₅: 351.1914. Found: 351.1914 ([M+H]⁺).



Synthesis of 3c: Biotin-p-toluene thiol ester (350.0 mg, 998.6 µmol) was added to DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and Phe-OEt (289.5 mg, 1.498 mmol) in it. The mixture was stirred at 37 °C for 1h, then cooled to room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate then washed successively with 1M aqueous HCl, 2M aqueous NaOH and brine. The organic layer was dried with Na₂SO₄, filtered, and concentrated to dryness. The crude product was purified by flash chromatography (first use 25% ethyl acetate in petroleum ether, then use 10% MeOH in DCM wash the chromatography to get the product). The title compound was prepared as a white solid (327.0 mg, 80 %). ¹H NMR (400 MHz, MeOD) $\delta =$ 7.32-7.23 (m, 5H), 4.69-4.64 (m, 1H), 4.52-4.49 (m, 1H), 4.30-4.27 (m, 1H), 4.19-4.12 (m, 2H), 3.16-3.15 (m, 2H), 2.97-2.92 (m, 2H), 2.74 (d, J=12.0, 1H), 2.21-2.18 (m, 2H), 1.60-1.57 (m, 4H), 1.24 (t, J=8.0, 4H),0.94-0.90 (m, 2H), ¹³C NMR (101 MHz, MeOD) $\delta = 174.5$, 171.9, 136.8, 128.8, 128.1, 126.4, 61.8, 61.0, 60.2, 55.5, 53.8, 39.6, 37.0, 34.8, 28.0, 27.9, 25.3, 13.0; HRMS (ESI) Calcd for: C₂₁H₃₀N₃O₄S: 420.1952. Found: 420.1945 ([M+H]⁺).



Synthesis of 3d: Cbz-Phe-*p*-toluene thiol ester (405.0 mg, 998.8 µmol) was added to DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and Ser-OEt (199.5 mg, 1.499 mmol) in it. The mixture was stirred at 37 °C for 1h, then cooled to room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate then washed successively with 1M aqueous HCl, 2M aqueous NaOH and brine. The organic layer was dried with Na₂SO₄, filtered, and concentrated to dryness. The crude product was purified by flash chromatography (first use 25% ethyl acetate in petroleum ether, then use 10% MeOH in DCM wash the chromatography to get the product). The title compound was prepared as a white solid (352.0 mg, 85%). ¹H NMR (400 MHz, CDCl₃) δ = 7.27-7.06

(m, 11H), 5.58 (d, *J*=8.0, 1H), 5.00-4.89 (m, 2H), 4.52-4.50 (m, 2H), 4.13-4.10 (m, 2H), 3.82 (s, 2H), 3.08-2.67 (m, 2H), 1.20 (t, *J*=8.0, 3H), ¹³C NMR (101 MHz, CDCl₃) δ = 171.5, 170.1, 136.2, 136.0, 129.3, 128.6, 128.5, 128.2, 128.0, 127.0, 67.1, 62.8, 61.9, 56.2, 54.9, 38.4, 14.1; HRMS (ESI) Calcd for: C₂₂H₂₇N₂O₃: 415.1864. Found: 415.1856 ([M+H]⁺).



Synthesis of 3e: Boc-L-Phe-Gly-*p*-toluene thiol ester (428.0 mg, 998.9 µmol) was added to DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and L-Phe-Gly-OEt (375.0 mg, 1.498 mmol) in it. The mixture was stirred at 37 °C for 2 h, then cooled to room temperature. The reaction mixture was purified by HPLC to get desire product as a white solid (415.0 mg, 75 %). ¹H NMR (400 MHz, MeOD) δ = 7.31-7.21 (m, 10H), 4.65 (s, 3H), 4.29-4.19 (m, 2H), 3.95-3.94 (m, 3H), 3.67-3.62 (m, 2H), 3.27-3.23 (m, 1H), 3.16-3.11 (m, 1H), 2.98-2.95 (m, 1H), 2.87-2.85 (m, 1H), 1.37-1.18 (m, 12H), ¹³C NMR (101 MHz, CDCl₃) δ = 171.5, 170.1, 136.2, 136.0, 129.3, 128.6, 128.5, 128.2, 128.0, 127.0, 67.1, 62.8, 61.9, 56.2, 54.9, 38.4, 14.1; HRMS (ESI) Calcd for: C₂₉H₃₉N₄O₇: 555.2813. Found: 555.2816 ([M+H]⁺).



Synthesis of 3f: Cbz-L-Phe-Gly-*p*-toluene thiol ester (462.0 mg, 998.7 µmol) was added to DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and L-Thr-Gly-L-Ala-OEt (412.0 mg, 1.497 mmol) in it. The mixture was stirred at 37 °C for 4 h, then cooled to room temperature. The reaction mixture was purified by HPLC to get desire product as a white solid (348.0 mg, 60.0 %). ¹H NMR (400 MHz, MeOD) δ = 8.34-8.31 (m, 1H), 8.16-8.08 (m, 2H), 7.84 (d, *J*=8.0, 1H), 7.56 (d, *J*=8.0, 1H), 7.35-7.20 (m, 10H), 4.95-4.93 (m, 2H), 4.28-4.21

(m, 3H), 4.10-4.00 (m, 3H), 3.87(t, *J*=4.0, 2H), 3.76(t, *J*=4.0, 2H), 3.07-3.02 (m, 2H), 2.79-2.73 (m, 1H), 1.74 (br, 1H), 1.28(d, *J*=8.0, 3H), 1.19(d, *J*=8.0, 3H), 1.09(d, *J*=8.0, 3H), 13 C NMR (101 MHz, CDCl₃) δ = 172.8, 170.7, 169.1, 137.4, 129.6, 128.7, 128.5, 127.9, 126.7, 67.0, 60.9, 48.1, 20.0, 17.4, 14.5; HRMS (ESI) Calcd for: C₂₇H₄₂N₅O₉: 580.2977. Found: 580.2971 ([M+H]⁺).



Synthesis of 3g: Thiol acid (76.00 mg, 998.4 µmol) was added to DMF/PBS (2000 µL, 1:1) in round-bottom flask, then added Cu(OAc)₂/HOBt (1 eq) and Phe-OEt (289.5 mg, 1.498 mmol) in it. The mixture was stirred at 37 °C for 0.1 h, then cooled to room temperature. The solvent was evaporated under reduced pressure, and the residue dissolved in ethyl acetate then washed successively with 1M aqueous HCl, 2M aqueous NaOH and brine. The organic layer was dried with Na₂SO₄, filtered, and concentrated to dryness. The crude product was purified by flash chromatography (first use 25% ethyl acetate in petroleum ether, then use 10% MeOH in DCM wash the chromatography to get the product). The title compound was prepared as a white solid (211.0 mg, 90 %). ¹H NMR (400 MHz, CDCl₃) δ 7.46 – 7.19 (m, 3H), 7.16 – 7.05 (m, 2H), 6.11 (d, *J* = 7.4 Hz, 1H), 4.87 (dt, *J* = 7.8, 5.9 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 3.46 – 2.93 (m, 2H), 1.99 (s, 3H), 1.25 (t, *J* = 7.1 Hz, 3H), ¹³C NMR (101 MHz, CDCl₃) δ 171.74, 169.67, 135.95, 129.32, 128.52, 127.08, 61.52, 53.17, 37.92, 23.14, 14.12; ESI-MS Calcd for: C₁₃H₁₈NO₃: 236.13. Found: 235.88 ([M+H]⁺).



Synthesis of 3h: Cy5.5 thiol acid (635.0 μ g, 999.5 μ mol) was added to DMF/PBS (100.0 μ L, 1:1) in EP tube, then added Cu(OAc)₂/HOBt (1 eq) and c(RGDfk) (900.0

 μ g, 1.542 μ mol) in it. The mixture was stirred at 37 °C for 0.5 h, then cooled to room temperature. The reaction mixture was purified by HPLC to get the product as a blue solid (759.4 μ g, 60 %). ESI-MS Calcd for: C₆₇H₈₂N₁₁O₈⁺: 1168.6. Found: 1168.9 ([M+H]⁺).



Synthesis of 3i: Cy5.5 thiol acid (635.0 µg, 0.9995 µmol) was added to DMF/PBS (100.0 µL, 1:1) in EP tube, then added Cu(OAc)₂/HOBt (1 eq) and JMV594 (1600 µg, 1.438 µmol) in it. The mixture was stirred at 37 °C for 0.5 h, then cooled to room temperature. The reaction mixture was purified by HPLC to get the product as a blue solid (1.057 mg, 63 %). ESI-MS Calcd for: C₉₅H₁₂₁N₁₆O₁₀S⁺: 1677.9. Found: 1679.4 ([M+H]⁺).



Synthesis of 3j: Cy5.5 thiol acid (635.0 μ g, 0.9995 μ mol) was added to DMF/PBS (100.0 μ L, 1:1) in EP tube, then added Cu(OAc)₂/HOBt (1 eq) and AE105 (2100 μ g, 1.453 μ mol) in it. The mixture was stirred at 37 °C for 0.5 h, then cooled to room temperature. The reaction mixture was purified by HPLC to get the product as a blue solid (1170 μ g, 58 %). ESI-MS Calcd for: C₁₁₀H₁₄₁N₁₈O₁₉: 2018.0, Found: 2017.5 ([M+H]⁺).



Synthesis of 3k: Cy5.5 thiol acid (635.0 μ g, 0.9995 μ mol) was added to DMF/PBS (100.0 μ L, 1:1) in EP tube, then added Cu(OAc)₂/HOBt (1 eq) and E[c(RGDyK)₂] (2.020 mg, 1.496 μ mol) in it. The mixture was stirred at 37 °C for 0.5 h, then cooled

to room temperature. The reaction mixture was purified by HPLC to get the product as a blue solid (1.111 mg, 58 %). ESI-MS Calcd for: $C_{99}H_{129}N_{21}O_{19}^+$: 1915.9. Found: 1915.1 ([M+H]⁻).

NMR Spectra

¹H NMR and ¹³C NMR for 1a



¹H NMR and ¹³C NMR for 1b



¹H NMR and ¹³C NMR for 1c



¹H NMR and ¹³C NMR for 1d





¹H NMR and ¹³C NMR for 3a



80 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10

¹H NMR and ¹³C NMR for 3b



¹H NMR and ¹³C NMR for 3c























¹H NMR and ¹³C NMR for 3g

