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Supporting Information

Ring-closing metathesis of unprotected peptides in water

Shun Masuda, Shugo Tsuda, and Taku Yoshiya*

Peptide Institute, Inc., Ibaraki, Osaka 567-0085, Japan

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1. General Information

a. Materials

All reagents and solvents were obtained from Peptide Institute, Inc. (Osaka, Japan), FUJIFILM Wako Pure Chemical Corporation (Osaka, Japan), Tokyo Chemical Industry Co., Ltd. (Tokyo, Japan), Nacalai Tesque, Inc. (Kyoto, Japan), Watanabe Chemical Industries, Ltd. (Hiroshima, Japan), Funakoshi Co., Ltd. (Tokyo, Japan), and Sigma-Aldrich Co. LLC. (St. Louis, MO).

b. HPLC, MS and NMR

Preparative HPLC was carried out on a Shimadzu liquid chromatograph Model LC-8A (Kyoto, Japan) with a YMC-Pack ODS-A (30 x 250 mm) and the following solvent systems: 0.1% TFA in H₂O and 0.1% TFA in CH₃CN at a flow rate of 20 mL min⁻¹ with detection at 220 nm. Analytical HPLC was performed on a Shimadzu liquid chromatograph Model LC-10A (Kyoto, Japan) with a YMC-Pack ODS-A (4.6 x 150 mm) and the following solvent systems: 0.1% TFA in H₂O and 0.1% TFA in CH₃CN at a flow rate of 1 mL min⁻¹ (40 °C) with detection at 220 nm. Low resolution mass spectra (LRMS) were observed with an Agilent G1956B LC/MSD detector using an Agilent 1100 series HPLC system. ¹H NMR spectra were recorded on a JEOL-ECX400 spectrometer (Tokyo, Japan), as solutions in deuterated solvents as specified. Chemical shift values (δ) are given in parts per million (ppm) using residual solvent protons as internal standard.

c. SPPS

Automated peptide synthesis by Fmoc SPPS was performed on an ABI 433A peptide synthesizer. The peptide chain was elongated using the coupling protocol of Fmoc-amino acid/DIC/OxymaPure.^[1] The acetyl capping was performed using acetic anhydride/NMP in the presence of DIEA after each coupling step. The following side-chain-protected amino acids were employed: Arg(Pbf), Cys(Trt), Glu(OtBu), His(Trt), Lys(Boc), Ser(*t*Bu), Thr(*t*Bu), Trp(Boc), Tyr(*t*Bu). Fmoc-Pgl (2 equiv) was manually condensed using DIC/OxymaPure (2/2 equiv) in NMP for 14–20 h. After HPLC purification and lyophilization of the target peptide, the identity was confirmed by MS and ¹H NMR (in case of a cyclic peptide), and the purity was determined to be higher than 95% by HPLC unless otherwise noted. RCM fundamentally affords peptide cis-trans mixture, which shows two different peaks in HPLC in all cases of this study. Each isomer, named sequentially as peptide **Xa/Xb** for cyclic peptide **X**, was isolated and analysed separately, while yield of RCM was calculated as the sum of both isomers.

2. Experimental Section

a. Preparation of unprotected peptides for RCM

Ac-Ser-His-Arg-Glu-Gly-Agl-Pro-Tyr-Agl-Val-NH₂(1)

The peptide was assembled on a Rink amide resin (0.125 mmol) using Fmoc SPPS as described in general information (Fmoc-Xaa: 8 equiv). The subsequent deprotection of the resin was carried out by TFA/TIS/H₂O (v/v, 95/2.5/2.5) for 1.5 h to give a crude product, which was purified by preparative HPLC to yield the title peptide **1** (65 mg, 44%). Analytical HPLC: $t_{\rm R} = 12.9$ min (1–60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₃H₇₉N₁₆O₁₅ 1179.6, found 1179.5.



Figure S1. Analytical HPLC chromatogram and ESI-MS spectrum of peptide 1.

Ser-His-Arg-Glu-Gly-Agl-Pro-Tyr-Agl-Val-NH₂(1')

The peptide was assembled on a Rink amide resin (0.125 mmol) using Fmoc SPPS as described in general information (Fmoc-Xaa: 8 equiv). The subsequent deprotection of the resin was carried out by TFA/TIS/H₂O (v/v, 95/2.5/2.5) for 1.5 h to give a crude product, which was purified by preparative HPLC to yield the title peptide **1'** (107 mg, 75%). Analytical HPLC: $t_{\rm R} = 12.1$ min (1–60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₁H₇₇N₁₆O₁₄ 1137.6, found 1137.5.



Figure S2. Analytical HPLC chromatogram and ESI-MS spectrum of peptide 1'.

2,7-Agl-octreotide (3)

The peptide was assembled on an H-Thr(*t*Bu)-ol-Trt(2-Cl)-resin (0.25 mmol) using Fmoc SPPS as described in general information (Fmoc-Xaa: 4 equiv). The subsequent deprotection of the resin was carried out by TFA/TIS/H₂O (v/v, 95/2.5/2.5) for 1.5 h to give a crude product, which was purified by preparative HPLC to yield the title peptide **3** (181 mg, 72%). Analytical HPLC: $t_R = 17.8 \text{ min} (1-60\% \text{ CH}_3\text{CN}/0.1\% \text{ TFA for 25 min})$; LRMS (M+H) calcd for C₅₃H₇₃N₁₀O₁₀ 1009.6, found 1009.5.



Figure S3. Analytical HPLC chromatogram and ESI-MS spectrum of peptide 3.

2,7-Pgl-octreotide (4)

The peptide was assembled on an H-Thr(*t*Bu)-ol-Trt(2-Cl)-resin (0.1 mmol) using Fmoc SPPS as described in general information (Fmoc-Xaa: 10 equiv, Fmoc-Pgl: 2 equiv). The subsequent deprotection of the resin was carried out by TFA/TIS/H₂O (v/v, 95/5/5) for 1.5 h to give a crude product, which was purified by preparative HPLC to yield the title peptide **4** (62 mg, 58%). Analytical HPLC: $t_R = 20.5 \text{ min} (1-60\% \text{ CH}_3\text{CN}/0.1\% \text{ TFA} \text{ for})$





Figure S4. Analytical HPLC chromatogram and ESI-MS spectrum of peptide 4.

2,7-Sac-octreotide (5)

The peptide was assembled on an H-Thr(*t*Bu)-ol-Trt(2-Cl)-resin (0.125 mmol) using Fmoc SPPS as described in general information (Fmoc-Xaa: 8 equiv). The subsequent deprotection of the resin was carried out by TFA/TIS/H₂O (v/v, 95/5/5) for 1 h to give a crude product. Then, the obtained peptide dissolved in H₂O (5 mL) and CH₃CN (3 mL) was reacted with allyl bromide (21.3 μ L, 0.252 mmol) and 1 M NaOH (360 μ L, 0.360 mmol) at room temperature. After 10 min, the reaction mixture was quenched with 50 % AcOH aq. (1 mL) and the solution was subjected to preparative HPLC to yield the title peptide **5** (89 mg, 56%). Analytical HPLC: $t_R = 19.6 \text{ min}$ (1–60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₅H₇₇N₁₀O₁₀S₂ 1101.5, found 1101.5.



Figure S5. Analytical HPLC chromatogram and ESI-MS spectrum of peptide 5.

b. Synthesis of cyclic peptides using RCM

Cyclised Ac-Ser-His-Arg-Glu-Gly-Agl-Pro-Tyr-Agl-Val-NH₂(2)

Peptide 1 (20.0 mg, 17.0 µmol) and MgCl₂·6H₂O (1.43 g, 7.04 mmol) were dissolved in 6.0 M Gn·HCl, 0.2 M phosphate (pH 7) (3.5 mL). To the mixture was added a solution of AquaMet (4.3 mg, 5.35 µmol) in H₂O (0.9 mL). After stirring for 2 h at 60 °C, the reaction mixture was quenched by TFA, and then subjected to preparative HPLC to obtain 2 (10.9 mg, 56%, 2a:2b = 40:60 determined by HPLC). Peptide 2a: ¹H NMR (400 MHz, $D_{2}O(\delta) = 8.60 (1H, s), 7.29 (1H, s), 7.14 (2H, d, J = 8.2 Hz), 6.84 (2H, d, J = 8.2 Hz),$ 5.59-5.49 (1H, m), 5.44-5.34 (1H, m), 4.61 (1H, t, J = 5.0 Hz), 4.39-4.30 (2H, m), 4.25 (1H, dd, J = 8.2 and 6.0 Hz), 4.16-4.08 (2H, m), 3.92 (2H, AB quartet, J = 16.9 Hz), 3.80(2H, d, J = 5.5 Hz), 3.73-3.64 (1H, m), 3.51-3.41 (1H, m), 3.37-3.26 (2H, m), 3.23-3.11 (3H, m), 2.77 (1H, dd, J = 14.2 and 11.4 Hz), 2.68-2.52 (2H, m), 2.47-2.26 (4H, m), 2.13-1.93 (7H, m), 1.93-1.67 (4H, m), 1.66-1.54 (2H, m), 1.27-1.17 (1H, m), 0.96 (6H, d, J = 6.4 Hz); Analytical HPLC: $t_{\rm R} = 11.2 \text{ min} (1-60\% \text{ CH}_3 \text{CN}/0.1\% \text{ TFA for 25 min}); LRMS$ (M+H) calcd for C₅₁H₇₅N₁₆O₁₅ 1151.6, found 1151.6. Peptide **2b**: ¹H NMR (400 MHz, D_2O) $\delta = 8.61$ (1H, d, J = 1.4 Hz), 7.28 (1H, s), 7.10 (2H, d, J = 8.2 Hz), 6.85 (2H, d, J =8.7 Hz), 5.54-5.43 (1H, m), 5.35-5.24 (1H, m), 4.66 (1H, dd, J = 10.5 and 4.1 Hz), 4.41-4.29 (3H, m), 4.22 (1H, dd, J = 8.2 and 6.0 Hz), 4.16 (1H, d, J = 7.3 Hz), 4.02 (1H, dd, J= 8.7 and 6.4 Hz), 3.91 (2H, AB quartet, J = 16.9 Hz), 3.79 (2H, d, J = 5.5 Hz), 3.72-3.63 (1H, m), 3.57-3.46 (1H, m), 3.36-3.25 (2H, m), 3.22-3.13 (3H, m), 3.04-2.86 (2H, m), 2.82 (1H, dd, J = 14.2 and 11.0 Hz), 2.42-2.32 (3H, m), 2.15-1.96 (7H, m), 1.96-1.68 (5H, m), 1.65-1.55 (2H, m), 1.49-1.38 (1H, m), 0.97 (6H, d, J = 6.9 Hz); Analytical HPLC: $t_{\rm R}$ = 11.6 min (1-60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₁H₇₅N₁₆O₁₅ 1151.6, found 1151.6.





Figure S6. Analytical HPLC chromatograms and ESI-MS spectra of peptide 2a and 2b.

Cyclised Ser-His-Arg-Glu-Gly-Agl-Pro-Tyr-Agl-Val-NH₂(2')

Peptide 1' (1.1 mg, 1.0 μ mol) and MgCl₂·6H₂O (81.3 mg, 400 μ mol) were dissolved in 0.1 M HCl aq. (0.8 mL). To the mixture was added a solution of AquaMet (0.25 mg, 0.3 μ mol) in H₂O (0.2 mL). After stirring for 2 h at 60 °C, the reaction mixture was monitored by analytical HPLC (1–60% CH₃CN/0.1% TFA for 25 min). Peptide **2**; LRMS (M+H) calcd for C₄₉H₇₃N₁₆O₁₄ 1109.6, found 1109.5.



Figure S7. Analytical HPLC chromatograms of RCM of model peptide **1'** and ESI-MS spectra of peptide **2'a** and **2'b**: A) reaction mixture (t = 0 h), B) reaction mixture (t = 2 h), C) ESI-MS spectrum of peptide **2'a**, D) ESI-MS spectrum of peptide **2'b**. * indicates internal standard. [RCM fundamentally affords cis-trans isomers, which are sequentially named as peptide **2'a**/**2'b** for cyclic peptide **2'**.]

Cyclised 2,7-Agl-octreotide (6)

Peptide 3 (10.0 mg, 9.91 µmol) and MgCl₂·6H₂O (805 mg, 3.96 mmol) were dissolved in 0.1 M HCl aq. (7.93 mL). To the mixture was added a solution of AquaMet (2.4 mg, 2.97 µmol) in H₂O (1.98 mL). After stirring for 2 h at 60 °C, the reaction mixture was subjected to preparative HPLC to obtain 6 (3.5 mg, 36%, 6a:6b = 63:37 determined by HPLC). Peptide **6a**: ¹H NMR (400 MHz, D_2O) δ = 7.55-7.34 (8H, m), 7.30-7.23 (5H, m), 7.20 (1H, t, J = 7.3 Hz), 7.14 (1H, s), 5.37-5.26 (1H, m), 5.14-5.04 (1H, m), 4.62 (1H, dd, J = 9.2 and 6.4 Hz), 4.42 (1H, dd, J = 11.4 and 2.7 Hz), 4.27-4.18 (2H, m), 4.12 (1H, dd, J = 11.0 Hz and 5.5 Hz), 4.10-4.03 (1H, m), 4.01 (1H, d, J = 4.6 Hz), 4.00-3.92 (2H, m), 3.87-3.80 (1H, m), 3.71 (1H, dd, J = 11.4 and 5.0 Hz), 3.62 (1H, dd, J = 11.4 and 7.3 Hz), 3.22-3.08 (2H, m), 3.07-2.89 (3H, m), 2.80 (1H, dd, J = 13.7 and 5.5 Hz), 2.75-2.55 (3H, m) 2.29-2.01 (3H, m), 1.62-1.50 (1H, m), 1.38-1.25 (5H, m), 1.25-1.07 (4H, m), 0.57-0.43 (1H, m), 0.35-0.22 (1H, m); Analytical HPLC: $t_{\rm R} = 15.2 \text{ min} (1-60\% \text{ CH}_3\text{CN}/0.1\% \text{ m})$ TFA for 25 min); LRMS (M+H) calcd for C₅₁H₆₉N₁₀O₁₀ 981.5, found 981.5. Peptide **6b**: ¹H NMR (400 MHz, D_2O) δ = 7.55 (1H, d, J = 7.8 Hz), 7.50 (1H, d, J = 8.2 Hz), 7.45-7.32 (6H, m), 7.31-7.17 (6H, m), 7.14 (1H, s), 5.48-5.37 (1H, m), 5.11-5.02 (1H, m), 4.65 (1H, dd, J = 8.7 Hz and 6.4 Hz), 4.46 (1H, dd, J = 10.1 Hz and 4.1 Hz), 4.28-4.20 (2H, m), 4.15 (1H, dd, J = 8.7 Hz and 5.5 Hz), 4.08-3.98 (3H, m), 3.98-3.90 (1H, m), 3.87-3.80 (1H, m), 3.70 (1H, dd, J = 11.9 Hz and 5.0 Hz), 3.60 (1H, dd, J = 11.9 Hz and 7.3 Hz), 3.20-2.87 (5H, m), 2.82 (1H, dd, J = 14.2 Hz and 6.0 Hz), 2.75-2.57 (2H, m), 2.53-2.42 (1H, m), 2.37-2.17 (3H, m), 1.59-1.46 (1H, m), 1.38-1.23 (5H, m), 1.23-1.07 (4H, m), 0.59-0.45 (1H, m), 0.41-0.27 (1H, m); Analytical HPLC: $t_{\rm R} = 15.3$ min (1–60%) CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₁H₆₉N₁₀O₁₀ 981.5, found 981.5.





Figure S8. Analytical HPLC chromatograms and ESI-MS spectra of peptide 6a and 6b.

Cyclised 2,7-Pgl-octreotide (7)

Peptide 4 (15.0 mg, 14.1 µmol) and MgCl₂·6H₂O (1.15 g, 5.64 mmol) were dissolved in 0.1 M HCl aq. (22.6 mL). To the mixture was added a solution of AquaMet (3.4 mg, 4.23 µmol) in H₂O (5.6 mL). After stirring for 30 min at 60 °C, the reaction mixture was subjected to preparative HPLC to obtain 7 (7.6 mg, 52%, 7a:7b = 68:32 determined by HPLC). Peptide 7a: ¹H NMR (400 MHz, D₂O) δ = 7.58(1H, d, J = 7.8 Hz), 7.48 (1H, d, J = 8.2 Hz), 7.43-7.28 (6H, m), 7.28-7.19 (5H, m), 7.16 (1H, t, J = 7.3 Hz), 7.11 (1H, s), 5.53-5.41 (1H, m), 5.35-5.23 (1H, m), 4.72-4.66 (1H, m), 4.59 (1H, dd, J = 10.5 Hz and 4.1 Hz), 4.44 (1H, dd, J = 10.1 Hz and 6.4 Hz), 4.35-4.28 (1H, m), 4.25-4.16 (2H, m), 4.13-3.98 (3H, m), 3.88-3.81 (1H, m), 3.70 (1H, dd, J = 11.4 Hz and 5.5 Hz), 3.62 (1H, dd, J = 11.4 Hz and 6.9 Hz), 3.23-3.13 (1H, m), 3.11-2.90 (5H, m), 2.75-2.57 (2H, m), 2.11-1.96(1H, m), 1.94-1.67 (5H, m), 1.61-1.40 (3H, m), 1.40-1.26 (4H, m), 1.26-1.17 (4H, m), 1.14 (3H, d, *J* = 6.4 Hz), 1.02-0.90 (2H, m), 0.64-0.50 (1H, m), 0.49-0.35 (1H, m); Analytical HPLC: $t_{\rm R} = 17.2 \text{ min} (1-60\% \text{ CH}_3\text{CN}/0.1\% \text{ TFA for 25 min})$, purity was 88%; LRMS (M+H) calcd for $C_{55}H_{77}N_{10}O_{10}$ 1037.6, found 1037.6. Peptide **7b**: ¹H NMR $(400 \text{ MHz}, D_2\text{O}) \delta = 7.52 (1\text{H}, \text{d}, J = 8.2 \text{ Hz}), 7.48 (1\text{H}, \text{d}, J = 8.2 \text{ Hz}), 7.43-7.34 (3\text{H}, J = 8.2 \text{Hz}), 7.43-7.34 (3\text{H}, J = 8.2 \text{Hz}),$ m), 7.34-7.20 (6H, m), 7.19-7.10 (3H, m), 7.04 (1H, s), 5.45-5.35 (1H, m), 5.34-5.23 (1H, m), 4.52 (1H, dd, J = 8.2 Hz and 6.9 Hz), 4.44-4.38 (1H, m), 4.34 (1H, dd, J = 10.1 Hz and 4.6 Hz), 4.16-4.03 (5H, m), 4.03-3.95 (1H, m), 3.86-3.78 (1H, m), 3.69 (1H, dd, J =11.4 Hz and 5.5 Hz), 3.58 (1H, dd, J = 11.4 Hz and 7.3 Hz), 3.19 (1H, dd, J = 13.3 Hz and 6.0 Hz), 3.07-2.92 (3H, m), 2.87 (1H, dd, J = 13.3 Hz and 8.7 Hz), 2.84-2.74 (3H, m), 2.07-1.66 (6H, m), 1.61-1.50 (1H, m), 1.50-1.26 (7H, m), 1.26-1.16 (3H, m), 1.11 (3H, d, J = 6.4 Hz), 1.00-0.89 (1H, m), 0.89-0.68 (3H, m); Analytical HPLC: $t_{R} = 17.9$ min (1-60% CH₃CN/0.1% TFA for 25 min), purity was 91%; LRMS (M+H) calcd for C₅₅H₇₇N₁₀O₁₀ 1037.6, found 1037.6.



Figure S9. Analytical HPLC chromatograms and ESI-MS spectra of peptide 7a and 7b.

Cyclised 2,7-Sac-octreotide (8)

Peptide **5** (10.0 mg, 9.08 μmol) and MgCl₂·6H₂O (738 mg, 3.63 mmol) were dissolved in 0.1 M HCl aq. (14.6 mL). To the mixture was added a solution of AquaMet (2.2 mg, 2.72 μmol) in H₂O (3.6 mL). After stirring for 30 min at 60 °C, the reaction mixture was subjected to preparative HPLC to obtain **8** (6.1 mg, 63%, **8a:8b** = 77:23 determined by HPLC). Peptide **8a**: ¹H NMR (400 MHz, D₂O) δ = 7.58 (1H, d, *J* = 7.8 Hz), 7.49 (1H, d, *J* = 7.8 Hz), 7.42-7.22 (9H, m), 7.20-7.13 (3H, m), 7.10 (1H, s), 5.61-5.49 (2H, m), 4.69-4.60 (2H, m), 4.48 (1H, dd, *J* = 9.2 Hz and 6.4 Hz), 4.38 (1H, dd, *J* = 8.2 Hz and 6.0 Hz), 4.26-4.15 (2H, m), 4.09-3.98 (2H, m), 3.97-3.89 (1H, m), 3.89-3.82 (1H, m), 3.71 (1H, dd, *J* = 11.4 Hz and 5.0 Hz), 3.62 (1H, dd, *J* = 11.4 Hz and 6.9 Hz), 3.26-3.12 (2H, m), 3.12-2.91 (9H, m), 2.82-2.70 (3H, m), 2.66 (1H, dd, *J* = 13.7 Hz and 6.0 Hz), 2.45 (1H, dd, *J* = 13.7 Hz and 8.2 Hz), 1.66-1.54 (1H, m), 1.46-1.27 (3H, m), 1.21 (3H, d, *J* = 6.4 Hz), 1.15 (3H, d, *J* = 6.4 Hz), 0.81-0.58 (2H, m); Analytical HPLC: $t_{\rm R}$ = 17.1 min (1–60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₃H₇₃N₁₀O₁₀S₂ 1073.5, found 1073.5. Peptide **8b**: ¹H NMR (400 MHz, D₂O) δ = 7.62 (1H, d, *J* = 7.8 Hz), 7.49 (1H, d, *J* = 8.2 Hz), 7.40-7.28 (6H, m), 7.27-7.20 (5H, m), 7.20-7.15 (1H, m), 7.14 (1H, s), 5.71-

5.62 (1H, m), 5.61-5.52 (1H, m), 5.10 (1H, dd, J = 8.7 Hz and 6.9 Hz), 4.54 (1H, dd, J = 10.5 Hz and 6.0 Hz), 4.33 (1H, d, J = 6.9 Hz), 4.29-4.21 (1H, m), 4.10-4.02 (2H, m), 3.94-3.88 (1H, m), 3.85-3.73 (2H, m), 3.69 (1H, dd, J = 11.4 Hz and 6.9 Hz), 3.28-3.11 (3H, m), 3.11-2.90 (6H, m), 2.88-2.74 (2H, m), 2.73-2.54 (3H, m), 2.47 (1H, dd, J = 14.2 Hz and 9.6 Hz), 2.22 (1H, dd, J = 14.2 Hz and 6.4 Hz), 1.70-1.58 (1H, m), 1.35-1.19 (7H, m), 1.19-1.10 (4H, m), 0.60-0.47 (1H, m), 0.43-0.30 (1H, m); Analytical HPLC: $t_{\rm R} = 17.3$ min (1–60% CH₃CN/0.1% TFA for 25 min); LRMS (M+H) calcd for C₅₃H₇₃N₁₀O₁₀S₂ 1073.5, found 1073.5.



Figure S10. Analytical HPLC chromatograms and ESI-MS spectra of peptide 8a and 8b.





Peptide 2b



Peptide 6a



Peptide 6b



Peptide 7a



Peptide 7b



Peptide 8a



4. Reference

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