Electronic Supporting Information

Hydrogen Peroxide as a Hydride Donor and Reductant under Biologically Relevant Conditions

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**Synthetic Procedures**

**General synthetic considerations.** The compounds Ru1 and ABTS\(^+\) were prepared and measured as previously described.\(^1\)-\(^3\) All other materials were of reagent quality and used as received. All solvents used were HPLC grade. \(^1\)H and \(^13\)C\(\{\)\(^1\)H\}\) NMR spectra were recorded using a Bruker 500 MHz spectrometer. Chemical shifts \(\delta\) (in ppm) for \(^1\)H and \(^13\)C NMR are referenced to SiMe\(_4\) using the residual proto-solvent as an internal standard. For \(^1\)H NMR: CDCl\(_3\), 7.26 ppm; DMSO-\(d_6\), 2.50 ppm. For \(^13\)C NMR: CDCl\(_3\), 77.16 ppm; DMSO-\(d_6\), 39.52 ppm. Coupling constants \(J\) are expressed in hertz (Hz). Fourier transform high-resolution mass spectrometry with electrospray ionization (ESI-MS) were acquired on a Thermo q-Exactive Plus instrument via direct injection (100% CH\(_3\)CN) using a Thermo Ultimate 3000 HPLC at a flow rate of 0.2L/min. A resolving power of 140,000 was used for the data acquisition, and the instrument was calibrated immediately prior to use yielding better than 5 ppm mass accuracy. All reactions were performed under an inert atmosphere under an N\(_2\) atmosphere using standard Schlenk or glovebox techniques with the exclusion of light. All subsequent manipulations were performed under ambient conditions using standard benchtop techniques without the exclusion of light. When required, solvents were dried and deoxygenated using an Innovative Technologies solvent purification system, and then stored over molecular sieves (3 Å) in a drybox.

**Spectroscopic and Kinetic Analysis Procedures**

**General spectroscopic considerations.** UV–visible absorption spectra were acquired on a Varian Cary 50 Bio spectrometer equipped with a water-cooled Quantum Northwest TC-125 peltier temperature controller. All solution measurements were performed at 25.0 ± 0.1 °C in matched gas-tight quartz cuvettes (Precision Scientific) with 1 cm path lengths and 3.0 mL analyte solution volumes. Absorption spectra were acquired from 950 to 200 nm with a scanning speed of 300 nm min\(^{-1}\) and a resolution of 0.5 nm. Each kinetic analysis experiment (5 second intervals) was performed in quadruplicate on four different days. Stock solutions were prepared fresh daily and filtered (0.2 μm PTFE) immediately prior to use.

**General kinetic procedure for analysis of ABTS\(^-\) degradation by Ru1 in PBS.** This procedure was used to generate the data for Figure 1. An aliquot of ABTS\(^2^-\) (30 μL from stock solution in H\(_2\)O) and an aliquot of ABTS\(^+\) (35 μL from stock solution in H\(_2\)O) were added to PBS (pH 7.4) and this working solution was allowed to equilibrate at 25 °C for 10 min. The absorbance spectrum of this working solution was acquired to confirm ABTS\(^+\) concentration and the single wavelength kinetics program was initiated. After 20 s, the cuvette was removed from the spectrometer without stopping the kinetics program, an aliquot of Ru1 (30 μL from stock solution in CH\(_3\)CN) was added, the cuvette was covered and mixed via repeated inversion for 3 s, placed back in the spectrometer, and the kinetics program was allowed to continue. After 5 min, an aliquot of 100 μM H\(_2\)O\(_2\), t-Bu\(_2\)O\(_2\), or t-BuOOH (30 μL from stock solution in H\(_2\)O) was added, the cuvette was covered and mixed via repeated inversion for 3 s, placed back in the spectrometer, and the kinetics program was allowed to continue. After the kinetics program had completed, the absorbance spectrum was acquired to confirm the formation of ABTS\(^2^-\) from the peak at 340 nm. **Standard Conditions** (unless specified otherwise): 100 μM ABTS\(^2^-\), 50 μM ABTS\(^+\), 5 μM Ru1, 100 μM H\(_2\)O\(_2\), t-Bu\(_2\)O\(_2\), or t-BuOOH, PBS (pH 7.4), 25 °C; [ABTS\(^-\)] determined from the absorbance at 734 nm, [ABTS\(^2^-\)] determined from the absorbance at 340 nm. **Notes:** (i) A 100 μM aliquot of ABTS\(^2^-\) is added at the beginning of each experiment to ensure initial ABTS\(^2^-\)/Ru1 ratios are consistent across different experimental conditions. (ii) ABTS\(^+\) reduction is incomplete after 30 min under these conditions due to the large (i.e., >20:1) ABTS\(^2^-\)/Ru1 ratios and the fact that ABTS\(^2^-\) inhibits Ru1-catalyzed ABTS\(^+\) reduction.\(^2\)
Quantitative ABTS\(^{-}\) reduction to ABTS\(^{2-}\) experiments. This procedure was used to generate the data for Figure S1. The concentrations of ABTS\(^{-}\), Ru1, and H\(_2\)O\(_2\) employed in these experiments were obtained by using the general kinetic procedure and standard conditions, holding aliquot volumes constant but varying stock solution concentrations. A 100 μM aliquot of ABTS\(^{2-}\) was not added at the beginning of this experiment because the combined absorbance of this initial aliquot and the ABTS\(^{2-}\) formed during the reaction would saturate the detector and prevent quantification of [ABTS\(^{2-}\)]. Note: ABTS\(^{-}\) reduction is complete after 30 min at 25 °C without an initial 100 μM aliquot of ABTS\(^{2-}\) due to smaller ABTS\(^{2-}\)/Ru1 ratios.

Volumetric quantification of O\(_2\) gas evolution. This procedure was used to generate data for Figure S3. The concentrations of ABTS\(^{-}\), Ru1, and H\(_2\)O\(_2\) employed in these experiments were obtained by using the general kinetic procedure and standard conditions, holding stock solution concentrations constant and varying aliquot volumes to achieve a final volume of 3.00 L. A 100 μM aliquot of ABTS\(^{2-}\) was not added at the beginning of this experiment because the combined absorbance of this initial aliquot and the ABTS\(^{2-}\) formed during the reaction would saturate the detector and prevent quantification of [ABTS\(^{2-}\)]. Note: ABTS\(^{-}\) reduction is incomplete after 30 min at 19 °C without an initial 100 μM aliquot of ABTS\(^{2-}\) due to the lower temperature (i.e., 19 °C temperature in the flask in the hood vs. 25 °C in the Peltier-controlled cuvette holder).

Rate law experiments. This procedure was used to generate the data for Figure 4. The concentrations of ABTS\(^{2-}\), ABTS\(^{-}\), Ru1, and H\(_2\)O\(_2\) employed in these experiments were obtained by using the general kinetic procedure and standard conditions, holding aliquot volumes constant but varying stock solution concentrations. The H\(^{+}\) concentrations were obtained using PBS adjusted to different pH values before the addition of any aliquots. The temperatures were obtained by allowing the working solution to equilibrate at different temperatures before the addition of Ru1. Note: A 100 μM aliquot of ABTS\(^{2-}\) is added at the beginning of each experiment to ensure initial ABTS\(^{2-}\)/Ru1 ratios are consistent across different experimental conditions.\(^2\)

Kinetic isotope effect experiments. The general kinetic procedure and standard conditions were employed as described for the rate law experiments, with minor modifications. Kinetic isotope effect experiments were performed in either protio PBS (pH 7.4) or deutero PBS (pD 7.4). Stock solutions of H\(_2\)O\(_2\) and D\(_2\)O\(_2\) (5.0 M) were prepared in H\(_2\)O and D\(_2\)O, respectively.
Derivation of General Rate Law for Proposed Mechanism

Proposed mechanism. Adapted from Scheme 2 in the manuscript. The intermediates \([\text{L}_n\text{Ru–A}_\text{red}]^{1–}, \ [\text{L}_n\text{Ru–A}_\text{ox}], \ [\text{L}_n\text{Ru–OH}]^{1+}, \ [\text{L}_n\text{Ru–(H}_2\text{O}_2)]^{1+}, \ [\text{L}_n\text{Ru–OOH}] \) and \([\text{L}_n\text{Ru–H}]\) have been abbreviated in the following equations as \([\text{RuA}], \ [\text{RuB}], \ [\text{RuC}], \ [\text{RuD}], \ [\text{RuE}], \) and \([\text{RuF}]\), respectively, for clarity.

(Equation S1) Based on previous mechanistic studies, it is known that the underlying reaction is the oxidation of \([\text{L}_n\text{Ru–H}]\) by \(\text{ABTS}^–\):

\[
\text{rate} = -\frac{d[A_{\text{ox}}]}{dt} = k_6[A_{\text{ox}}][\text{RuF}]
\]

(Equation S2) The sum of the concentrations of the Ru-containing species leading up to the rate-determining step is equal to the total concentration of \(\text{Ru1}\) added at the beginning of the experiment:

\[
[\text{Ru1}]_0 = [\text{RuA}] + [\text{RuB}] + [\text{RuC}] + [\text{RuD}] + [\text{RuE}] + [\text{RuF}]
\]

(Equation S3) Assuming that steps 1-4 achieve equilibrium rapidly (with respect to turnover):

\[
K_1 = \frac{[\text{RuC}][A_{\text{red}}]}{[\text{RuA}]}, \quad K_2 = \frac{[\text{RuC}][A_{\text{ox}}]}{[\text{RuB}]}, \quad K_3 = \frac{[\text{RuD}]}{[\text{RuC}][H_2O_2]}, \quad K_4 = \frac{[\text{RuE}][H^+]}{[\text{RuD}]}
\]

(Equation S4) Assuming the system achieves steady-state rapidly (with respect to turnover):

\[
\frac{d[\text{RuF}]}{dt} = k_5[\text{RuE}] - k_6[A_{\text{ox}}][\text{RuF}]
\]

\[
[\text{RuE}] = \frac{k_6[A_{\text{ox}}]}{k_5}[\text{RuF}]
\]
(Equation S5) Solving Equation S2 for $[\text{RuF}]$ using the relationships established in Equations S3 and S4:

$$[\text{Ru1}]_0 = \left( \frac{[\text{A}_{\text{red}}]}{K_1} + \frac{[\text{A}_{\text{ox}}]}{K_2} \right) [\text{RuC}] + [\text{RuD}] + [\text{RuE}] + [\text{RuF}]$$

$$[\text{Ru1}]_0 = \left( \frac{[\text{A}_{\text{red}}]}{K_1 K_3 [\text{H}_2 \text{O}_2]} + \frac{[\text{A}_{\text{ox}}]}{K_2 K_3 [\text{H}_2 \text{O}_2]} + \frac{1}{K_3 [\text{H}_2 \text{O}_2]} + 1 \right) [\text{RuD}] + [\text{RuE}] + [\text{RuF}]$$

$$[\text{Ru1}]_0 = \left( \frac{[\text{A}_{\text{red}}][\text{H}^+]}{K_1 K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{[\text{A}_{\text{ox}}][\text{H}^+]}{K_2 K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{[\text{H}^+]}{K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{[\text{H}^+]}{K_4} + 1 \right) [\text{RuE}] + [\text{RuF}]$$

$$[\text{Ru1}]_0 = \left( \frac{k_6 [\text{H}^+][\text{A}_{\text{red}}][\text{A}_{\text{ox}}]}{k_5 K_1 K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{k_6 [\text{H}^+][\text{A}_{\text{ox}}]^2}{k_5 K_1 K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{k_6 [\text{H}^+][\text{A}_{\text{ox}}]}{k_5 K_3 K_4 [\text{H}_2 \text{O}_2]} + \frac{k_6 [\text{H}^+][\text{A}_{\text{ox}}]}{k_5 K_4} + \frac{k_6 [\text{A}_{\text{ox}}]}{k_5} + 1 \right) [\text{RuF}]$$

$$[\text{Ru1}]_0 = \left( \frac{k_6 K_2 [\text{H}^+][\text{A}_{\text{red}}][\text{A}_{\text{ox}}] + k_6 K_1 [\text{H}^+][\text{A}_{\text{ox}}]^2 + k_6 K_1 K_2 [\text{H}^+][\text{A}_{\text{ox}}] + k_6 K_1 K_2 K_3 [\text{H}_2 \text{O}_2][\text{H}^+][\text{A}_{\text{ox}}] + k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2][\text{A}_{\text{ox}}] + k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]}{k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]} \right) [\text{RuF}]$$

$$[\text{RuF}] = \left( \frac{k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]}{k_6 K_2 [\text{H}^+][\text{A}_{\text{red}}][\text{A}_{\text{ox}}] + k_6 K_1 [\text{H}^+][\text{A}_{\text{ox}}]^2 + k_6 K_1 K_2 [\text{H}^+][\text{A}_{\text{ox}}] + k_6 K_1 K_2 K_3 [\text{H}_2 \text{O}_2][\text{H}^+][\text{A}_{\text{ox}}] + k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2][\text{A}_{\text{ox}}] + k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]}{[\text{Ru1}]_0} \right) [\text{Ru1}]_0$$
(Equation S6) Plugging the result from Equation S5 into Equation S1 gives the general rate law for the proposed mechanism:

\[
\text{rate} = -\frac{d[A_{ox}]}{dt} = k_6[A_{ox}][RuF]
\]

\[
= k_6[A_{ox}]\left\{\left(\frac{k_5k_1K_2K_3K_4[H_2O_2]}{k_5k_2[H^+][A_{red}][A_{ox}] + k_6K_1[A_{ox}]^2 + k_6K_1K_2[H^+][A_{ox}] + k_6K_1K_2K_3[H_2O_2][H^+][A_{ox}] + k_6K_1K_2K_3K_4[H_2O_2][A_{ox}] + k_5K_1K_2K_3K_4[H_2O_2]}\right)[Ru1]_0\right\}
\]

\[
= \frac{k_5k_6K_1K_2K_3K_4[H_2O_2][A_{ox}][Ru1]_0}{k_6K_2[H^+][A_{red}][A_{ox}] + k_6K_1[A_{ox}]^2 + k_6K_1K_2[H^+][A_{ox}] + k_6K_1K_2K_3[H_2O_2][H^+][A_{ox}] + k_6K_1K_2K_3K_4[H_2O_2][A_{ox}] + k_5K_1K_2K_3K_4[H_2O_2]}
\]

(Equation S7) At very short reaction times, the concentration of species will be very close to their initial concentrations (e.g., \([A_{ox}] \approx [A_{ox}]_0\)), therefore the initial rate \((v_0)\) can be expressed with the following equation:

\[
v_0 = \frac{k_5k_6K_1K_2K_3K_4[H_2O_2]_0[A_{ox}]_0[Ru1]_0}{k_6K_2[H^+]_0[A_{red}]_0[A_{ox}]_0 + k_6[A_{ox}]_0^2 + k_6K_1K_2[H^+]_0[A_{ox}]_0 + k_6K_1K_2K_3[H_2O_2]_0[H^+]_0[A_{ox}]_0 + k_6K_1K_2K_3K_4[H_2O_2]_0[A_{ox}]_0 + k_5K_1K_2K_3K_4[H_2O_2]_0}
\]

(Equation S8) The equation in (7) can be expressed in terms of the different variables \([A_{ox}]_0, [A_{red}]_0, [H^+]_0, [H_2O_2]_0,\) and \([Ru1]_0\) to simplify graphical analysis and data fitting:

(a) If \([A_{ox}]_0\) is varied and the other variables are held constant, a plot of \(v_0\) vs. \([A_{ox}]_0\) should fit the following equation:

\[
v_0 = \frac{(k_5k_6K_1K_2K_3K_4[H_2O_2]_0[Ru1]_0) \cdot [A_{ox}]_0}{(k_6) \cdot [A_{ox}]_0^2 + (k_6K_2[H^+]_0[A_{red}]_0 + k_6K_1K_2[H^+]_0 + k_6K_1K_2K_3[H_2O_2]_0[H^+]_0 + k_6K_1K_2K_3K_4[H_2O_2]_0) \cdot [A_{ox}]_0 + k_5K_1K_2K_3K_4[H_2O_2]_0}
\]

\[
v_0 = \frac{C_1 \cdot [A_{ox}]_0}{C_2 \cdot [A_{ox}]_0^2 + C_3 \cdot [A_{ox}]_0 + C_4}
\]
(b) If \([\text{A}_{\text{red}}]_0\) is varied and the other variables are held constant, a plot of \(v_0\) vs. \([\text{A}_{\text{red}}]_0\) should fit the following equation:

\[
v_0 = \frac{(k_5 k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0 [\text{Ru1}]_0 [\text{A}_{\text{ox}}]_0)}{(k_6 K_2 [\text{H}^+]_0 [\text{A}_{\text{ox}}]_0) \cdot [\text{A}_{\text{red}}]_0 + (k_6 [\text{A}_{\text{ox}}]_0^2 + k_6 K_1 K_2 [\text{H}^+]_0 [\text{A}_{\text{ox}}]_0 + k_6 K_1 K_2 K_3 [\text{H}_2 \text{O}_2]_0 [\text{H}^+]_0 [\text{A}_{\text{ox}}]_0 + k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0 [\text{A}_{\text{ox}}]_0 + k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0)}
\]

\[
v_0 = \frac{C_5}{C_6 \cdot [\text{A}_{\text{red}}]_0 + C_7}
\]

(c) If \([\text{H}^+]_0\) is varied and the other variables are held constant, a plot of \(v_0\) vs. \([\text{H}^+]_0\) should fit the following equation:

\[
v_0 = \frac{(k_5 k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0 [\text{Ru1}]_0)}{(k_6 K_2 [\text{A}_{\text{red}}]_0 [\text{A}_{\text{ox}}]_0 + k_6 K_1 K_2 [\text{A}_{\text{ox}}]_0 + k_6 K_1 K_2 K_3 [\text{H}_2 \text{O}_2]_0 [\text{A}_{\text{ox}}]_0 + \text{[H}^+]_0 + (k_6 [\text{A}_{\text{ox}}]_0^2 + k_6 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0 [\text{A}_{\text{ox}}]_0 + k_5 K_1 K_2 K_3 K_4 [\text{H}_2 \text{O}_2]_0)}
\]

\[
v_0 = \frac{C_8}{C_9 \cdot [\text{H}^+]_0 + C_{10}}
\]

(d) If \([\text{H}_2 \text{O}_2]_0\) is varied and the other variables are held constant, a plot of \(v_0\) vs. \([\text{H}_2 \text{O}_2]_0\) should fit the following equation:

\[
v_0 = \frac{(k_5 k_6 K_1 K_2 K_3 K_4 [\text{A}_{\text{ox}}]_0 [\text{Ru1}]_0) \cdot [\text{H}_2 \text{O}_2]_0}{(k_6 K_1 K_2 K_3 [\text{H}^+]_0 [\text{A}_{\text{ox}}]_0 + k_6 K_1 K_2 K_3 K_4 \cdot [\text{H}_2 \text{O}_2]_0 + (k_6 K_2 [\text{H}^+]_0 [\text{A}_{\text{red}}]_0 [\text{A}_{\text{ox}}]_0 + k_6 [\text{A}_{\text{ox}}]_0^2 + k_6 K_1 K_2 [\text{H}^+]_0 [\text{A}_{\text{ox}}]_0)}
\]

\[
v_0 = \frac{C_{11} \cdot [\text{H}_2 \text{O}_2]_0}{C_{12} \cdot [\text{H}_2 \text{O}_2]_0 + C_{13}}
\]
Figure S1. Overlaid UV–visible spectra for ABTS$^{−−}$ + Ru1 before (red line) and 30 min after the addition of H$_2$O$_2$ (blue line). During the course of this reaction, the ABTS$^{−−}$ concentration decreased by 50 μM (downward grey arrow) and the ABTS$^{2−}$ concentration increased by 50 μM (upward grey arrow). Conditions: [Ru1]$_0$ = 5 μM, [ABTS$^{−−}$]$_0$ = 50 μM, [H$_2$O$_2$]$_0$ = 100 μM, PBS (pH 7.4), 25 °C; [ABTS$^{−−}$] determined from absorbance at 734 nm (ε = 1.5 × 10$^4$ M$^{−1}$ cm$^{−1}$) and [ABTS$^{2−}$] determined from absorbance at 340 nm (ε = 3.7 × 10$^4$ M$^{−1}$ cm$^{−1}$).
Figure S2. Representative digital images acquired during volumetric measurements of O$_2$ gas evolution from Ru1-catalyzed ABTS$^*$ reduction with H$_2$O$_2$ under standard conditions in 3.00 L reaction volumes. Entire apparatus (A) before and (B) 30 min after the addition of H$_2$O$_2$. (C) Cuvettes containing PBS blank (left) and 3.0 mL aliquot from 3.0 L reaction solution before the addition of H$_2$O$_2$ (right). (D) Cuvettes containing PBS blank (left) and 3.0 mL aliquot from 3.0 L reaction solution 30 min after the addition of H$_2$O$_2$ (right). (E) Zoom-in of inverted graduated cylinder 30 min after the addition of H$_2$O$_2$ into which 1.8 mL ± 0.1 of O$_2$ gas (72 ± 2 μmol) had been collected. Conditions: [Ru1]$_0$ = 5 μM, [ABTS$^*$]$_0$ = 50 μM, [H$_2$O$_2$]$_0$ = 100 μM, PBS (pH 7.4), 19 °C, $V_{\text{rxn}} = 3.00$ L.
Figure S3. Overlaid UV–visible spectra, of the 3.0 mL aliquots from the volumetric measurements of O₂ gas evolution shown in Figures S2C and S2D, for ABTS⁻⁺ + Ru1 before (red line) and 30 min after the addition of H₂O₂ (blue line). During the course of this reaction, [ABTS⁻⁺] decreased by 44 ± 1 μM (downward grey arrow) and [ABTS²⁻] increased by 42 ± 1 μM (upward grey arrow). Whereas ABTS⁻⁺ reduction was quantitative at 25 °C (see Figure S1), at 19 °C it was incomplete (this figure). Accounting for the original volume of 3.00 L shown in Figures S2A and S2B, 132 ± 3 μmol of ABTS⁻⁺ were reduced to 126 ± 3 μmol of ABTS²⁻, which was accompanied by the evolution of 72 ± 2 μmol of O₂ gas. Thus, for every 1.0 equiv. of ABTS⁻⁺ reduced, 0.95 ± 0.03 equiv. of ABTS²⁻ were formed and 0.54 ± 0.02 equiv. of O₂ gas were released. Conditions: [Ru1]₀ = 5 μM, [ABTS⁻⁺]₀ = 50 μM, [H₂O₂]₀ = 100 μM, PBS (pH 7.4), 19 °C, V_cuvette = 3.00 mL; [ABTS⁻⁺] determined from absorbance at 734 nm (ε = 1.5 × 10⁴ M⁻¹ cm⁻¹) and [ABTS²⁻] determined from absorbance at 340 nm (ε = 3.7 × 10⁴ M⁻¹ cm⁻¹).
Figure S4. Fourier-transform high-resolution mass spectra with electrospray ionization (ESI-MS) for Ru1-catalyzed ABTS•– reduction with H₂O₂. Samples were administered by direct injection and detected in positive mode. **Bottom:** calculated pattern for the protonated Ru–H intermediate, which would have the formula C₂₆H₂₉N₂O₂Ru. **Top:** observed pattern in the expected region for the protonated Ru–H intermediate, which does not match calculated pattern, but instead corresponds to a cationic Ru-containing species bearing only the cymene and NHC ligands, which has the formula C₂₆H₂₇N₂O₂Ru. **Conditions:** [Ru1]₀ = 5 μM, [ABTS•–]₀ = 50 μM, [ABTS²⁻]₀ = 100 μM, [H₂O₂]₀ = 100 μM, NH₄OAc buffer (10 mM, pH 7.4), 25 °C.
References