

Flavonoid-Alkylphospholipid Conjugates Elicit Dual Inhibition of Cancer Cell Growth and Lipid Accumulation

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Table of Contents

Figure S1.....	S3
Table S1.....	S4
Table S2.....	S5
General.....	S6
Synthesis of I and II	S7
UV spectral recording of I-d and II-c	S18
Cell culture.....	S18
Reagents and antibodies.....	S18
Cell growth inhibition assay.....	S19
Lactate dehydrogenase (LDH) assay.....	S20
Apoptosis and necrosis assay on flow cytometry.....	S20
Western blotting analysis.....	S21
Oil red O staining.....	S21
Measurements of cellular triglyceride (TG) content.....	S22

Figure S1: UV spectral recording of **I-d** (13 μM) and **II-c** (12 μM) in organic solvent MeOH/ CH_2Cl_2 (A) and in a buffer solution with increasing concentration of sodium dodecyl sulfate (SDS) (B).

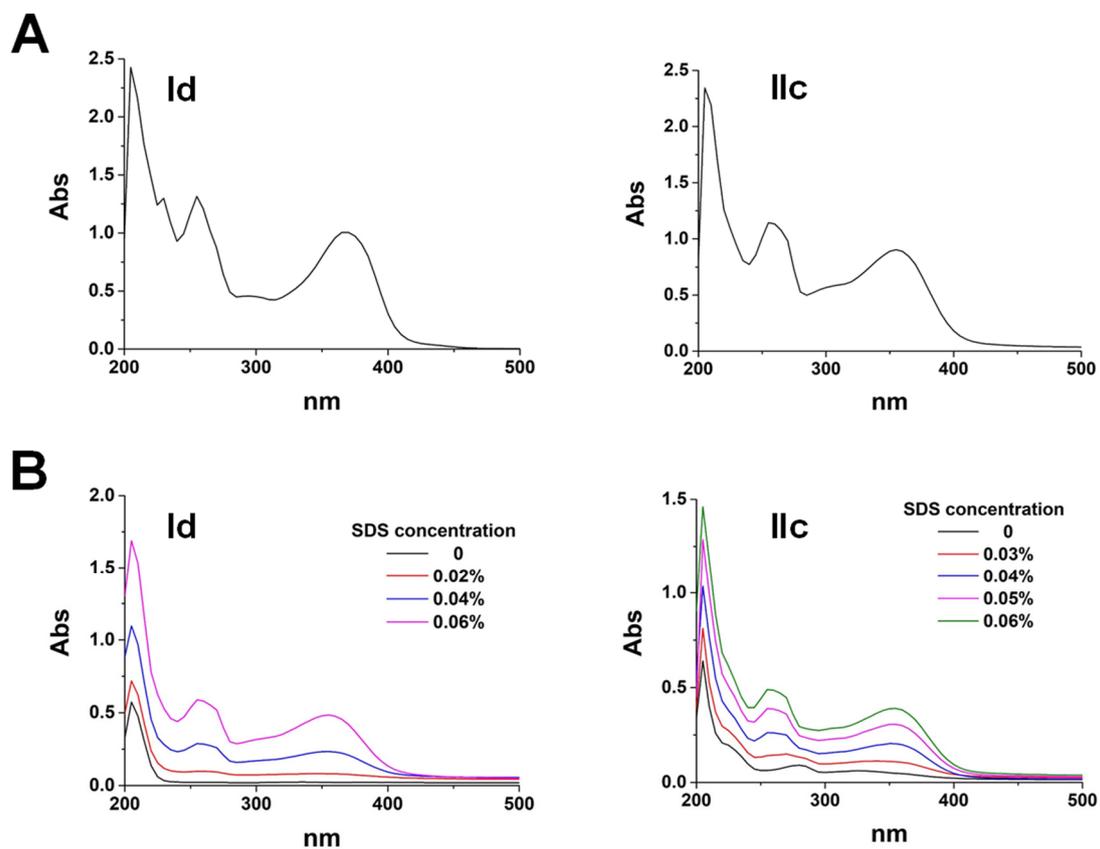
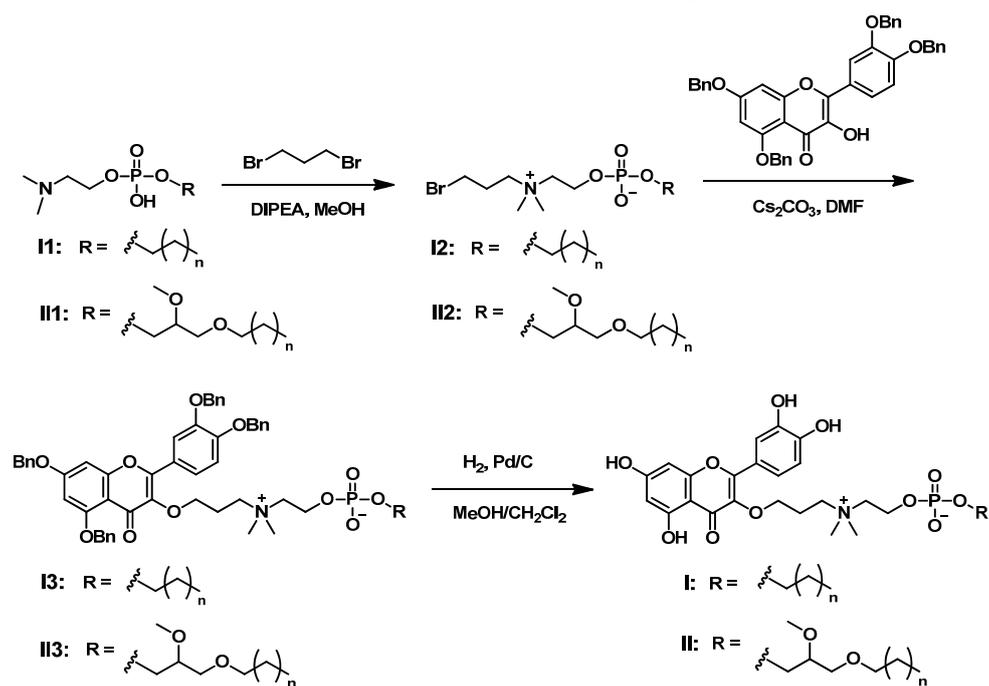


Table S1. Synthesis of quercetin-alkylphospholipid conjugates **I** and **II**.



Entry	n	Comp.	Yield (%)	Comp.	Yield (%)	Comp.	Yield (%)
1	10	I2a	79	I3a	61	Ia	98
2	12	I2b	75	I3b	61	Ib	95
3	14	I2c	79	I3c	71	Ic	95
4	16	I2d	81	I3d	64	Id	97
5	10	II2a	77	II3a	61	IIa	79
6	12	II2b	69	II3b	59	IIb	83
7	14	II2c	67	II3c	66	IIc	77
8	16	II2d	75	II3d	70	IId	81

Table S2. Antiproliferation activity of quercetin-alkylphospholipid conjugates **I** and **II** on different cancer cells

No	Comp.	IC ₅₀ (μM) ^a			
		HepG2	Hep3B	Panc-1	SKOV3
1	I-a	>50	>50	>50	>50
2	I-b	25±1	41±1	36±4	26±3
3	I-c	20±1	33±1	18±1	25±1
4	I-d	9.8±1.1	16±1	8.9±0.2	19±1
5	II-a	>50	>50	49±2	47±1
6	II-b	21±1	31±2	20±1	15±1
7	II-c	13±1	20±2	10±2	18±1
8	II-d	20±1	27±2	20±1	18±2
9	Quercetin	72±1	>100	61±1	>100
10	Miltefosine	>50	35±2	20±1	>50
11	Edelfosine	>50	24±1	12±2	>50

^a IC₅₀ represents the concentration of a compound that is required for 50% inhibition of cancer cell growth.

General

All the reagents were purchased from Adamas-beta or Energy Chemical. Solvents were purchased from a local supplier and used without further purification beside THF. THF was distilled from sodium and benzophenone. All compounds were purified by performing flash chromatography on silica gel (200–300 mesh). ¹H-NMR and ¹³C-NMR spectra were recorded on Agilent DD2 400-MR or DD2 600-MR. The chemical shifts were recorded in parts per million (ppm) with tetramethylsilane as the internal reference. The ESI-MS was recorded on a Waters Acquity SQ Detector mass spectrometer. The high resolution of ESI-MS was recorded on a Bruker Solarix 7.0T mass spectrometer. All MS analysis samples were prepared as solutions in methanol. UV spectral was recorded on Agilent Technologies Cary 60 UV-Vis. Analytical HPLC runs were performed on Agilent Technologies 1260 Infinity using columns packed with Inertsil[®] hypersil C8 4.6×250mm (method 1) and Inertsil[®] ODS-3 5mm 4.6×250mm (method 2) produced by GL sciences Inc, with 1260 VWD VL, wavelength 370 nm. All the samples were dissolved in DMSO (contain 1% TFA). The mobile phase was consisted of a gradient elution of MeOH (contain 0.1% TFA) and H₂O. Elution system was the mixture of MeOH/H₂O from 10/90 to 100/0 in 20 min and then kept 100/0 in 25 min. The flow rate was 1.0 mL/min. Temperature was 30 °C. Compounds of **II** and **III** were synthesized following the reference of previous report.¹

Synthesis of compound I and II

General procedure of synthesis of I2

To a solution of **I1** (0.5 mmol) in MeOH (10 mL) was added 1,3-dibromopropane (1.0 g, 10.0 eq) and diisopropylethylamine (260 mg, 4.0 eq), the mixture was stirred at 50 °C for 48 h. Then the solvent was removed and the residue was purified by column gel chromatograph to afford product.

I2a: 79%, white solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.27 (m, 2H), 3.79-3.82 (m, 4H), 3.70 (m, 2H), 3.56 (t, 2H, *J* = 6.4 Hz), 3.36 (s, 6H), 2.42 (m, 2H), 1.58 (m, 2H), 1.26 (m, 18H), 0.88 (t, 3H, *J* = 6.4 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 65.74, 65.68, 64.21, 58.84, 52.00, 31.90, 31.12, 31.05, 29.69, 29.65, 29.53, 29.34, 29.26, 25.94, 22.66, 14.09. MS (ESI, *m/z*): 458.4, 460.4 [M + H]⁺.

I2b: 75%, white solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.28 (m, 2H), 3.77-3.84 (m, 4H), 3.67-3.71 (m, 2H), 3.56 (t, 2H, *J* = 6.0 Hz), 3.36 (s, 6H), 2.42 (m, 2H), 1.57-1.58 (m, 2H), 1.26 (m, 22H), 0.88 (t, 3H, *J* = 6.4 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 65.84, 64.26, 58.92, 52.00, 31.90, 31.09, 31.01, 29.72, 29.66, 29.56, 29.35, 29.20, 25.95, 22.66, 14.09. MS (ESI, *m/z*): 486.5, 488.5 [M + H]⁺.

I2c: 79%, white solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.27 (m, 2H); 3.77-3.83 (m, 4H); 3.68-3.72 (m, 2H); 3.56 (t, 2H, *J* = 6.4 Hz); 3.36 (s, 6H); 2.42 (m, 2H); 1.57-1.60 (m, 2H); 1.26 (m, 26H); 0.88 (t, 3H, *J* = 6.4 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 65.68, 65.62, 64.14, 58.82, 51.93, 31.86, 31.11, 31.04, 29.69, 29.61, 29.52, 29.30, 29.24, 25.92, 22.62, 14.05. MS (ESI, *m/z*): 514.5, 516.5 [M + H]⁺.

I2d: 81%, white solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.29 (m, 2H); 3.78-3.83 (m, 4H);

3.68-3.72 (m, 2H); 3.55 (t, 2H, $J = 6.0$ Hz); 3.36 (s, 6H); 2.41 (m, 2H); 1.57-1.60 (m, 2H); 1.25 (m, 30H); 0.88 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 65.67, 65.61, 64.18, 58.82, 51.96, 31.88, 31.12, 31.05, 29.70, 29.67, 29.62, 29.52, 29.31, 29.25, 25.93, 22.64, 14.06. MS (ESI, m/z): 542.5, 544.5 $[\text{M} + \text{H}]^+$.

*General procedure of synthesis of **I3***

I2 (0.1 mmol), quercetin (99.3 mg, 1.5 eq) and Cs_2CO_3 (65 mg, 2.0 eq) were suspended in DMF (5 mL) and the mixture was stirred at room temperature overnight. Then the solvent was removed and the residue was purified by column gel chromatograph to afford product.

I3a: 61%, yellow solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.62 (d, 1H, $J = 8.4$ Hz), 7.28-7.50 (m, 21H), 7.06 (d, 1H, $J = 8.4$ Hz), 6.41 (s, 1H), 6.35 (s, 1H), 5.18 (s, 2H), 5.17 (s, 2H), 5.14 (s, 2H), 4.98 (s, 2H), 4.27 (m, 2H), 3.88 (m, 2H), 3.65-3.79 (m, 6H), 3.23 (s, 6H), 2.09 (m, 2H), 1.48-1.52 (m, 2H), 1.12-1.26 (m, 18H), 0.84 (t, 3H, $J = 6.8$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.55, 162.81, 159.58, 158.50, 153.08, 151.22, 148.26, 139.71, 137.08, 136.57, 136.17, 135.51, 128.72, 128.62, 128.54, 128.43, 127.93, 127.54, 127.36, 127.30, 126.86, 123.10, 123.02, 115.05, 113.86, 109.58, 97.88, 93.68, 71.80, 70.69, 70.40, 68.77, 65.85, 65.81, 63.70, 58.87, 51.80, 31.90, 30.93, 30.86, 29.70, 29.53, 29.36, 25.84, 23.78, 22.67, 14.12. MS (ESI, m/z): 1040.8 $[\text{M} + \text{H}]^+$.

I3b: 61%, yellow solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.62 (d, 1H, $J = 8.4$ Hz), 7.28-7.53 (m, 21H), 7.06 (dd, 1H, $J_1 = 8.4$ Hz, $J_2 = 2.4$ Hz), 6.44 (s, 1H), 6.40 (s, 1H), 5.21 (s, 4H), 5.17 (s, 2H), 5.01 (s, 2H), 4.27 (m, 2H), 3.89 (m, 2H), 3.66-3.80 (m, 6H), 3.26

(s, 6H), 2.09 (m, 2H), 1.53 (m, 2H), 1.16-1.25 (m, 22H), 0.86 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.55, 162.83, 159.58, 158.54, 153.09, 151.16, 148.24, 139.69, 137.05, 136.53, 136.16, 135.48, 128.74, 128.58, 128.46, 127.99, 127.95, 127.56, 127.36, 127.29, 126.82, 123.02, 122.96, 115.00, 113.79, 109.59, 97.89, 93.68, 71.73, 70.69, 70.41, 68.73, 65.69, 64.04, 63.74, 58.83, 51.78, 31.92, 31.01, 30.94, 29.73, 29.54, 29.38, 25.88, 23.76, 22.69, 14.15. MS (ESI, m/z): 1068.8 $[\text{M} + \text{H}]^+$.

13c: 71%, yellow solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.61 (d, 1H, $J = 8.4$ Hz), 7.26-7.54 (m, 21H); 7.07 (d, 1H, $J = 8.4$ Hz); 6.45 (s, 1H); 6.40 (s, 1H); 5.20 (s, 4H); 5.16 (s, 2H); 5.02 (s, 2H); 4.27 (m, 2H); 3.90 (m, 2H); 3.74-3.82 (m, 6H); 3.25 (s, 6H); 2.10 (m, 2H); 1.52-1.55 (m, 2H); 1.16-1.23 (m, 26H); 0.86 (t, 2H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.50, 162.86, 159.64, 158.55, 153.09, 151.25, 148.35, 139.75, 137.08, 136.57, 136.19, 135.54, 128.72, 128.55, 128.42, 127.97, 127.93, 127.51, 127.37, 127.29, 126.82, 123.13, 122.98, 115.22, 113.96, 109.67, 97.97, 93.81, 71.83, 70.79, 70.43, 68.80, 65.62, 65.56, 64.13, 63.78, 58.81, 51.78, 31.89, 31.06, 30.98, 29.70, 29.64, 29.52, 29.33, 25.89, 23.81, 22.65, 14.09. MS (ESI, m/z): 1096.9 $[\text{M} + \text{H}]^+$.

13d: 64%, yellow solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.61 (d, 1H, $J = 8.4$ Hz); 7.26-7.50 (m, 21H); 7.06 (d, 1H, $J = 8.8$ Hz); 6.42 (s, 1H); 6.36 (s, 1H); 5.18 (s, 2H); 5.17 (s, 2H); 5.14 (s, 2H); 4.99 (s, 2H); 4.28 (m, 2H); 3.88 (m, 2H); 3.71-3.79 (m, 6H); 3.23 (s, 6H); 2.09 (m, 2H); 1.49-1.52 (m, 2H); 1.12-1.28 (m, 30H); 0.86 (t, 2H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.58, 162.87, 159.65, 158.56, 153.14, 151.30, 148.35, 139.77, 137.13, 136.62, 136.21, 135.57, 128.74, 128.65, 128.57, 128.44, 127.97, 127.55, 127.40, 127.34, 126.91, 123.10, 115.20, 113.97, 109.65, 97.97, 93.79,

71.87, 70.77, 70.44, 68.83, 65.87, 64.19, 63.78, 58.94, 51.85, 31.93, 30.96, 30.91, 29.78, 29.74, 29.68, 29.57, 29.37, 25.88, 23.83, 22.69, 14.12. MS (ESI, m/z): 1125.0 $[M + H]^+$.

General procedure of synthesis of I

I3 (0.1 mmol) and Pd/C (20 mg) was suspended in MeOH/CH₂Cl₂ (5 mL, 1:1) and the mixture was stirred at room temperature under hydrogen atmosphere for 4 h. Then the solid was filtered off and the filtrate was collected. After evaporation, the residue was washed by ethyl acetate to afford the product.

Ia: 98%, yellow solid. HPLC: t_R = 21.762 min (Method 1, purity 97.90 %), t_R = 22.654 min (Method 2, purity 97.51 %). ¹H-NMR (400 MHz, DMSO-d₆): δ 12.67 (s, 1H), 11.57 (br, 1H), 11.23 (br, 1H), 9.58 (br, 1H), 7.74 (s, 1H), 7.19 (d, 1H, J = 8.4 Hz), 6.88 (d, 1H, J = 8.4 Hz), 6.41 (s, 1H), 6.20 (s, 1H), 4.12 (m, 2H), 3.96 (m, 2H), 3.68-3.70 (m, 2H), 3.59 (m, 4H), 3.12 (s, 6H), 2.16 (m, 2H), 1.46-1.48 (m, 2H), 1.15-1.30 (m, 18H), 0.84 (t, 3H, J = 6.8 Hz). ¹³C-NMR (100 MHz, DMSO-d₆): δ 178.01, 164.85, 161.60, 157.11, 156.79, 149.42, 146.34, 136.42, 120.85, 119.70, 116.38, 115.96, 104.39, 99.01, 94.08, 70.20, 69.26, 64.89, 64.83, 62.75, 61.30, 58.63, 52.08, 31.72, 30.87, 30.80, 29.49, 29.47, 29.45, 29.28, 29.14, 25.86, 23.55, 22.52, 14.38. MS (ESI, m/z): 680.6 $[M + H]^+$. HRMS: calcd for C₃₄H₅₁NO₁₁P⁺, $[M + H]^+$, 680.3194; found, 680.3169.

Ib: 95%, yellow solid. HPLC: t_R = 22.617 min (Method 1, purity 96.48 %), t_R = 23.478 min (Method 2, purity 96.49 %). ¹H-NMR (400 MHz, DMSO-d₆): δ 12.68 (br, 1H), 11.71 (br, 1H), 11.00 (br, 1H), 9.50 (br, 1H), 7.76 (s, 1H), 7.19 (d, 1H, J = 8.0 Hz), 6.88 (d, 1H, J = 8.4 Hz), 6.41 (s, 1H), 6.19 (s, 1H), 4.11 (m, 2H), 3.97 (m, 2H), 3.58-3.70

(m, 6H), 3.11 (s, 6H), 2.17 (m, 2H), 1.45-1.47 (m, 2H), 1.16-1.22 (m, 22H), 0.85 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, DMSO- d_6): δ 177.95, 164.92, 161.54, 157.00, 156.73, 149.37, 146.23, 136.35, 120.82, 119.73, 116.30, 115.95, 104.29, 98.99, 94.05, 69.22, 64.94, 64.88, 62.64, 61.31, 58.70, 52.02, 31.74, 30.84, 30.78, 29.52, 29.48, 29.29, 29.18, 25.86, 23.54, 22.55, 14.40. MS (ESI, m/z): 708.6 $[\text{M} + \text{H}]^+$. HRMS: calcd for $\text{C}_{36}\text{H}_{55}\text{NO}_{11}\text{P}^+$, $[\text{M} + \text{H}]^+$, 708.3507; found, 708.3485.

Ic: 95%, yellow solid. HPLC: $t_{\text{R}} = 23.187$ min (Method 1, purity 97.07 %), $t_{\text{R}} = 24.157$ min (Method 2, purity 97.94 %). ^1H -NMR (400 MHz, DMSO- d_6): δ 12.63 (s, 1H), 11.39 (br, 1H), 9.67 (br, 1H), 7.71, (s, 1H), 7.19 (d, 1H, $J = 8.0$ Hz), 6.89 (d, 1H, $J = 8.4$ Hz), 6.40 (s, 1H), 6.19 (s, 1H), 4.16 (m, 2H), 3.96 (m, 2H), 3.72-3.74 (m, 2H), 3.61 (m, 4H), 3.13 (s, 6H), 2.17 (m, 2H), 1.47-1.51 (m, 2H), 1.15-1.21 (m, 26H), 0.84 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, DMSO- d_6): δ 178.00, 164.93, 161.59, 156.99, 156.77, 149.40, 146.22, 136.45, 120.87, 119.88, 116.31, 116.01, 104.36, 99.03, 94.06, 69.27, 65.07, 65.02, 62.89, 61.58, 58.78, 51.95, 31.72, 30.83, 30.76, 29.49, 29.28, 29.14, 25.83, 23.56, 22.52, 14.36. MS (ESI, m/z): 736.6 $[\text{M} + \text{H}]^+$. HRMS: calcd for $\text{C}_{38}\text{H}_{59}\text{NO}_{11}\text{P}^+$, $[\text{M} + \text{H}]^+$, 736.3820; found, 736.3794.

Id: 97%, yellow solid. HPLC: $t_{\text{R}} = 25.641$ min (Method 1, purity 96.51 %), $t_{\text{R}} = 26.438$ min (Method 2, purity 97.22 %). ^1H -NMR (400 MHz, CD_3OD): δ 7.56 (s, 1H), 7.40 (d, 1H, $J = 8.0$ Hz), 6.94 (d, 1H, $J = 8.4$ Hz), 6.35 (s, 1H), 6.17 (d, 1H, $J = 1.6$ Hz), 4.27 (m, 2H), 4.00-4.02 (m, 2H), 3.84-3.89 (m, 2H), 3.63-3.67 (m, 4H), 3.18 (s, 6H), 2.19 (m, 2H), 1.56-1.62 (m, 2H), 1.22-1.35 (m, 30H), 0.89 (t, 3H, $J = 6.8$ Hz). ^{13}C -NMR (100 MHz, DMSO- d_6): δ 178.12, 164.76, 161.63, 156.88, 156.80, 149.37, 146.06,

136.63, 120.90, 120.41, 116.14, 104.50, 99.04, 94.09, 69.33, 65.87, 62.98, 61.97, 59.28, 51.69, 31.72, 30.58, 30.54, 29.47, 29.43, 29.19, 29.13, 25.67, 23.57, 22.52, 14.38. MS (ESI, m/z): 764.7 $[M + H]^+$. HRMS: calcd for $C_{40}H_{63}NO_{11}P^+$, $[M + H]^+$, 764.4133; found, 764.4104..

*General procedure of synthesis of **II2***

To a solution of **III1** (0.5 mmol) in MeOH (10 mL) was added 1,3-dibromopropane (1.0 g, 10.0 eq) and diisopropylethylamine (260 mg, 4.0 eq). The mixture was stirred at 50 °C for 48 h. Then the solvent was removed and the residue was purified by column gel chromatograph to afford the product.

II2a: 77%, white waxy solid. 1H -NMR (400 MHz, $CDCl_3$): δ 4.33 (s, 2H), 3.84-3.97 (m, 4H), 3.69-3.72 (m, 2H), 3.49-3.58 (m, 6H), 3.43-3.49 (m, 2H), 3.41 (t, 2H, $J = 6.8$ Hz), 3.36 (s, 6H), 2.42 (m, 2H), 1.53-1.55 (m, 2H), 1.26 (s, 18H), 0.88 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, $CDCl_3$): δ 79.60, 79.52, 71.79, 70.10, 65.05, 64.28, 58.98, 57.79, 51.98, 31.89, 29.67, 29.64, 29.53, 29.34, 26.06, 25.90, 22.66, 14.10. MS (ESI, m/z): 546.5, 548.5 $[M+H]^+$.

II2b: 69%, white waxy solid. 1H -NMR (400 MHz, $CDCl_3$): δ 4.33 (m, 2H), 3.85-3.96 (m, 4H), 3.69-3.72 (m, 2H), 3.49-3.57 (m, 6H), 3.43-3.47 (m, 2H), 3.41 (t, 2H, $J = 6.8$ Hz), 3.36 (s, 6H), 2.42 (m, 2H), 1.54 (t, 2H, $J = 5.6$ Hz), 1.26 (s, 22H), 0.88 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, $CDCl_3$): δ 79.68, 79.61, 71.78, 70.21, 65.01, 64.32, 58.89, 57.78, 51.95, 31.88, 29.62, 29.52, 29.32, 26.06, 25.90, 22.64, 14.08. MS (ESI, m/z): 574.6, 576.4 $[M+H]^+$, 596.6, 598.6 $[M+Na]^+$.

II2c: 67%, white waxy solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.31 (s, 2H), 3.78-3.95 (m, 5H), 3.62-3.66 (m, 2H), 3.51-3.59 (m, 5H), 3.44-3.48 (m, 4H), 3.41 (t, 2H, *J* = 6.8 Hz), 3.31 (s, 6H), 2.42 (m, 2H), 1.55 (t, 2H, *J* = 6.4 Hz), 1.26 (s, 26H), 0.88 (t, 3H, *J* = 6.8 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 79.58, 79.50, 71.80, 70.09, 65.09, 64.25, 58.98, 57.80, 52.02, 31.89, 29.69, 29.55, 29.48, 29.33, 26.07, 25.91, 22.65, 14.08. MS (ESI, *m/z*): 602.6, 604.6 [M+H]⁺.

II2d: 75%, white waxy solid. ¹H-NMR (400 MHz, CDCl₃): δ 4.33 (s, 2H), 3.84-3.97 (m, 4H), 3.66-3.70 (m, 2H), 3.46-3.58 (m, 6H), 3.42-3.46 (m, 4H), 3.41 (t, 2H, *J* = 6.4 Hz), 3.34 (s, 6H), 2.87 (s, 1H), 2.42 (m, 2H), 1.53-1.56 (m, 2H), 1.25 (s, 30H), 0.88 (t, 3H, *J* = 6.0 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 79.41, 79.33, 71.76, 70.04, 65.15, 59.63, 57.75, 52.05, 43.43, 31.89, 29.71, 29.66, 29.57, 29.33, 26.07, 22.64, 14.06. MS (ESI, *m/z*): 630.7, 632.7 [M+H]⁺.

General procedure of synthesis of II3

II2 (0.1 mmol), quercetin (99.3 mg, 1.5 eq) and Cs₂CO₃ (65 mg, 2.0 eq) were suspended in DMF (5 mL) and the mixture was stirred at room temperature overnight. After that, the solvent was removed and the residue was purified by column gel chromatograph to afford product.

II3a: 61%, yellow waxy solid. ¹H-NMR (400 MHz, CDCl₃): δ 7.60 (d, 1H, *J* = 8.4 Hz), 7.33-7.55 (m, 21H), 7.08 (d, 1H, *J* = 8.4 Hz), 6.52 (s, 1H), 6.46 (s, 1H), 5.27 (s, 2H), 5.24 (s, 2H), 5.20 (s, 2H), 5.09 (s, 2H), 4.31 (m, 2H), 3.78-3.96 (m, 7H), 3.66 (m, 2H), 3.45-3.56 (m, 5H), 3.37 (t, 2H, *J* = 6.8 Hz), 3.24 (s, 6H), 2.22 (t, 1H, *J* = 8.0 Hz), 2.06

(m, 2H), 1.51 (t, 2H, $J = 6.0$ Hz), 1.23 (s, 18H), 0.87 (t, 3H, $J = 6.0$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.54, 162.88, 159.66, 158.57, 153.15, 151.27, 148.36, 139.74, 137.08, 136.59, 136.17, 135.54, 128.72, 128.56, 128.42, 127.94, 127.51, 127.37, 127.30, 126.85, 123.13, 122.98, 115.23, 113.97, 109.66, 97.97, 93.82, 79.73, 79.65, 71.82, 71.65, 70.79, 70.67, 70.44, 68.82, 64.73, 64.68, 64.20, 63.91, 58.84, 57.80, 51.77, 31.88, 29.61, 29.50, 29.31, 26.03, 23.76, 22.64, 14.08. MS (ESI, m/z): 1128.9 $[\text{M}+\text{H}]^+$, 1150.8 $[\text{M}+\text{Na}]^+$.

II3b: 59%, yellow waxy solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.60 (d, 1H, $J = 8.4$ Hz), 7.32-7.55 (m, 15H), 7.08 (d, 1H, $J = 8.8$ Hz), 6.51 (s, 1H), 6.46 (s, 1H), 5.26 (s, 2H), 5.23 (s, 2H), 5.19 (s, 2H), 5.08 (s, 2H), 4.32 (s, 2H), 3.79-3.95 (m, 8H), 3.66 (s, 2H), 3.45-3.56 (m, 4H), 3.37 (t, 2H, $J = 7.6$ Hz), 3.24 (s, 6H), 2.04 (m, 2H), 1.52-1.55 (m, 2H), 1.21-1.27 (m, 22H), 0.87 (t, 3H, $J = 6.4$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.66, 162.92, 159.64, 158.58, 153.27, 151.28, 148.34, 139.71, 137.05, 136.56, 136.14, 135.52, 128.72, 128.61, 128.56, 127.95, 127.53, 127.36, 127.29, 126.88, 123.08, 122.99, 115.19, 113.92, 109.60, 97.98, 93.79, 79.61, 79.54, 71.81, 71.68, 70.77, 70.74, 70.46, 68.78, 64.80, 64.15, 63.92, 58.80, 57.79, 51.74, 50.34, 31.88, 29.67, 29.62, 29.50, 29.32, 26.01, 23.70, 22.65, 14.08. MS (ESI, m/z): 1157.1 $[\text{M}+\text{H}]^+$.

II3c: 66%, yellow waxy solid. ^1H -NMR (400 MHz, CDCl_3): δ 7.61 (d, 1H, $J = 8.0$ Hz), 7.26-7.53 (m, 21H), 7.05 (d, 1H, $J = 8.8$ Hz), 6.42 (s, 1H), 6.38 (s, 1H), 5.20 (s, 2H), 5.17 (s, 2H), 5.15 (s, 2H), 5.00 (s, 2H), 4.38 (m, 2H), 3.78-3.96 (m, 5H), 3.40-3.51 (m, 5H), 3.40-3.51 (m, 4H), 3.33 (t, 2H, $J = 6.8$ Hz), 3.27 (s, 6H), 2.11 (m, 2H), 1.48-1.49 (m, 2H), 1.22 (m, 26H), 0.87 (t, 3H, $J = 5.6$ Hz). ^{13}C -NMR (100 MHz, CDCl_3): δ 173.60,

162.92, 159.67, 158.60, 153.24, 151.27, 148.36, 139.71, 137.07, 136.57, 136.14, 135.53, 128.73, 128.58, 128.44, 127.99, 127.95, 127.53, 127.36, 127.29, 126.86, 123.11, 122.92, 115.24, 113.97, 109.66, 98.00, 93.83, 79.73, 79.66, 71.81, 71.67, 70.81, 70.67, 70.47, 68.78, 64.71, 64.66, 64.23, 64.01, 58.80, 57.82, 51.82, 33.88, 32.70, 31.89, 29.68, 29.63, 29.52, 29.33, 28.53, 28.00, 26.04, 23.75, 22.65, 14.09. MS (ESI, m/z): 1184.9 [M+H]⁺.

II3d: 70%, yellow waxy solid. ¹H-NMR (400 MHz, CDCl₃): δ 7.62 (d, 1H, J = 8.4 Hz), 7.27-7.54 (m, 21H), 7.07 (d, 1H, J = 8.4 Hz), 6.49 (s, 1H), 6.43 (s, 1H), 5.23 (s, 2H), 2.22 (s, 2H), 5.18 (s, 2H), 5.06 (s, 2H), 4.32 (s, 2H), 3.91-3.94 (m, 4H), 3.87 (m, 2H), 3.78 (m, 2H), 3.36-3.53 (m, 4H), 3.33 (t, 2H, J = 6.8 Hz), 3.22 (s, 6H), 2.05 (m, 2H), 1.49 (m, 2H), 1.21-1.25 (m, 30H), 0.87 (t, 3H, J = 6.4 Hz). ¹³C-NMR (100 MHz, CDCl₃): δ 173.53, 162.88, 159.66, 158.57, 153.15, 151.26, 148.36, 139.74, 137.07, 136.58, 136.17, 135.54, 128.72, 128.56, 128.42, 127.97, 127.94, 127.51, 127.37, 127.29, 126.84, 123.13, 122.96, 115.23, 113.97, 109.66, 97.97, 93.82, 79.75, 79.67, 71.81, 71.66, 70.79, 70.70, 70.44, 68.82, 64.65, 64.06, 63.92, 58.83, 57.80, 51.74, 50.09, 33.88, 31.88, 29.62, 29.52, 29.32, 26.04, 23.75, 22.65, 14.09. MS (ESI, m/z): 1213.0 [M+H]⁺.

General procedure of synthesis of II

II3 (0.1 mmol) and Pd/C (20 mg) was suspended in MeOH/CH₂Cl₂ (5 mL, 1:1). The mixture was stirred at room temperature under hydrogen atmosphere for 4 h. After the solid was filtered off, the filtrate was collected and evaporated. Then the residue was washed by ethyl acetate to afford product.

IIa: 79%, yellow solid. HPLC: $t_R = 22.734$ min (Method 1, purity 96.87 %), $t_R = 23.486$ min (Method 2, purity 96.74 %). $^1\text{H-NMR}$ (600 MHz, DMSO-d_6): δ 12.66 (s, 1H), 11.37 (s, 1H), 11.20 (s, 1H), 9.58 (s, 1H), 7.72 (s, 1H), 7.20 (d, 1H, $J = 8.4$ Hz), 6.88 (d, 1H, $J = 8.4$ Hz), 6.40 (s, 1H), 6.19 (s, 1H), 4.14 (m, 2H), 3.96-3.97 (m, 2H), 3.69-3.77 (m, 2H), 3.58-3.60 (m, 3H), 3.40-3.42 (m, 3H), 3.27-3.31 (m, 6H), 3.12 (s, 6H), 2.17 (m, 2H), 1.40 (m, 2H), 1.19-1.23 (m, 18H), 0.84 (t, 3H, $J = 6.6$ Hz). $^{13}\text{C-NMR}$ (150 MHz, DMSO-d_6): δ 178.04, 164.78, 161.61, 157.10, 156.80, 149.43, 146.32, 136.45, 120.85, 119.77, 116.31, 115.99, 104.44, 99.00, 94.08, 79.50, 79.45, 71.04, 70.31, 69.30, 64.36, 62.82, 61.36, 58.71, 57.45, 52.04, 31.72, 29.60, 29.44, 29.29, 29.14, 26.01, 23.55, 22.52, 14.38. MS (ESI, m/z): 768.7 $[\text{M}+\text{H}]^+$, 790.6 $[\text{M}+\text{Na}]^+$. HRMS: calcd for $\text{C}_{38}\text{H}_{58}\text{NNaO}_{13}\text{P}^+$, $[\text{M}+\text{Na}]^+$, 790.3527; found, 790.3538.

IIb: 83%, yellow solid. HPLC: $t_R = 23.207$ min (Method 1, purity 96.17 %), $t_R = 24.179$ min (Method 2, purity 96.04 %). $^1\text{H-NMR}$ (400 MHz, DMSO-d_6): δ 12.69 (s, 1H), 11.59 (s, 1H), 10.93 (s, 1H), 9.51 (s, 1H), 7.74 (s, 1H), 7.21 (d, 1H, $J = 8.0$ Hz), 6.88 (d, 1H, $J = 8.4$ Hz), 6.41 (s, 1H), 6.19 (s, 1H), 4.12 (s, 2H), 3.98 (s, 2H), 3.67-3.74 (m, 2H), 3.58 (s, 4H), 3.39-3.42 (m, 8H), 3.11 (s, 6H), 2.18 (m, 2H), 2.00 (m, 2H), 1.40 (m, 2H), 1.21 (m, 22H), 0.85 (t, 3H, $J = 6.0$ Hz). $^{13}\text{C-NMR}$ (150 MHz, DMSO-d_6): δ 178.04, 164.80, 161.62, 157.00, 156.80, 149.39, 146.19, 136.51, 120.88, 120.01, 116.24, 116.06, 104.44, 99.02, 94.08, 79.46, 79.41, 71.06, 70.25, 69.33, 64.52, 62.96, 61.66, 58.78, 57.47, 51.92, 51.18, 31.72, 29.60, 29.48, 29.46, 29.31, 29.14, 26.01, 23.58, 22.52, 14.37. MS (ESI, m/z): 796.8 $[\text{M}+\text{H}]^+$, 818.9 $[\text{M}+\text{Na}]^+$. HRMS: calcd for $\text{C}_{40}\text{H}_{63}\text{NO}_{13}\text{P}^+$, $[\text{M}+\text{H}]^+$, 796.4054; found, 796.4032.

IIc: 77%, yellow solid. HPLC: $t_R = 23.675$ min (Method 1, purity 97.88 %), $t_R = 24.639$ min (Method 2, purity 97.89 %). $^1\text{H-NMR}$ (400 MHz, DMSO- d_6): δ 12.69 (s, 1H), 11.56 (br, 1H), 10.92 (s, 1H), 9.52 (br, 1H), 7.74 (s, 1H), 7.21 (d, 1H, $J = 8.0$ Hz), 6.88 (d, 1H, $J = 8.0$ Hz), 6.41 (s, 1H), 6.18 (s, 1H), 4.12 (m, 2H), 3.97 (m, 2H), 3.66-3.75 (m, 3H), 3.59 (m, 4H), 3.40-3.41 (m, 3H), 3.30 (m, 4H), 3.11 (s, 6H), 2.18 (m, 2H), 1.40 (m, 2H), 1.21 (d, 26H), 0.85 (t, 3H, $J = 6.8$ Hz). $^{13}\text{C-NMR}$ (150 MHz, DMSO- d_6): δ 178.04, 164.80, 161.62, 157.02, 156.80, 149.40, 146.21, 136.50, 120.86, 119.97, 116.25, 116.04, 104.44, 99.01, 94.07, 79.46, 79.42, 71.05, 70.26, 69.32, 64.47, 62.92, 61.60, 58.76, 57.46, 55.33, 51.94, 31.72, 29.60, 29.31, 29.13, 26.01, 23.56, 22.52, 14.36. MS (ESI, m/z): 824.7 $[\text{M}+\text{H}]^+$, 846.6 $[\text{M}+\text{Na}]^+$. HRMS: calcd for $\text{C}_{42}\text{H}_{66}\text{NNaO}_{13}\text{P}^+$, $[\text{M}+\text{Na}]^+$, 846.4175; found, 846.4164.

IIId: 81%, yellow solid. HPLC: $t_R = 24.388$ min (Method 1, purity 95.10 %), $t_R = 25.355$ min (Method 2, purity 95.85 %). $^1\text{H-NMR}$ (400 MHz, DMSO- d_6): δ 12.71 (s, 1H), 11.70 (br, 1H), 10.94 (br, 1H), 9.55 (br, 1H), 7.76 (s, 1H), 7.21 (d, 1H, $J = 8.0$ Hz), 6.88 (d, 1H, $J = 8.0$ Hz), 6.41 (s, 1H), 6.18 (s, 1H), 4.11 (m, 2H), 3.97 (m, 2H), 3.71 (m, 2H), 3.51-3.58 (m, 6H), 3.29-3.41 (m, 4H), 3.11 (s, 6H), 2.18 (m, 2H), 1.39 (s, 2H), 1.21 (d, 30H), 0.85 (t, 3H, $J = 6.8$ Hz). $^{13}\text{C-NMR}$ (150 MHz, DMSO- d_6): δ 178.04, 164.82, 161.62, 156.98, 156.80, 149.39, 146.17, 136.51, 120.87, 120.04, 116.23, 116.06, 104.44, 99.02, 94.07, 79.46, 79.41, 71.06, 70.26, 69.33, 64.52, 62.98, 61.71, 58.78, 57.46, 51.89, 51.17, 31.73, 29.61, 29.33, 29.14, 26.02, 23.57, 22.52, 14.35. MS (ESI, m/z): 852.8 $[\text{M}+\text{H}]^+$. HRMS: calcd for $\text{C}_{44}\text{H}_{71}\text{NO}_{13}\text{P}^+$, $[\text{M}+\text{H}]^+$, 852.4637; found, 852.4657. calcd for $\text{C}_{44}\text{H}_{70}\text{NNaO}_{13}\text{P}^+$, $[\text{M}+\text{Na}]^+$, 874.4455; found, 874.4477.

UV spectral recording in organic solvent and phosphate buffer

The test samples were prepared in methanol/dichloromethane (1:2). The concentration of **Id** and **Iic** in the solvent was 13 μM and 12 μM , respectively. The UV spectra was recorded on Agilent Technologies Cary 60 UV-Vis.

To 0.1 mg of the compound was added 2.0 mL of 100 mM phosphate buffer (pH = 7.4) and mixed by vortex. After that, the mixture was allowed to stand for 1 minute. The supernatant was used for UV recording. Then the detergent sodium dodecyl sulfate (SDS) was added and the UV spectra of the samples with different concentrations of SDS were recorded.

Cell culture

Human liver cancer HepG2 cell and ovarian cancer SKOV3 cells were purchased from Cell Resource Centre, IBMS, CAMS/PUMC. Human liver cancer Hep3B cell were purchased from China Center for Type Culture Collection, Wuhan University. Human pancreatic cancer Panc-1 cells were the gifts from Prof. Huaizhi Wang (Institute of Hepatopancreatobiliary Surgery, Southwest Hospital, Chongqing). HepG2 and Hep3B cells were grown in Dulbecco's modified eagle's medium (DMEM) (HyClone) supplemented with 10% fetal bovine serum (FBS). Panc-1 cells were grown in Roswell Park Memorial Institute 1640 (RPMI 1640) (GIBCO) supplemented with 10% FBS. SKOV3 cells were grown in McCoy's 5A (GIBCO) supplemented with 10% FBS. All cells were incubated at a humidified atmosphere containing 5% CO_2 at 37 $^\circ\text{C}$.

Reagents and Antibodies

MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl tetrazolium bromide) was purchased from Adamas. The annexin V-FITC/ Propidium Iodide (PI) apoptosis detection kit was purchased from Sangon Biotech (Shanghai, China). Oil Red O and haematoxylin were purchased from Sangon Biotech (Shanghai, China). The BCA Protein Assay Kit was purchased from EnoGene (Nanjing, China). Kits for determining triglyceride (TG) was purchased from Applygen Technologies Inc (Beijing, China). The primary antibodies including Bcl-2 antibody, HSF1 rabbit mAb, HSP70 rat mAb, HSP27 rabbit mAb, eIF4E rabbit mAb, PARP antibody, LXR- β rabbit mAb and PPAR γ rabbit mAb were all obtained from Cell Signaling Technology (Danvers, MA, USA). The AKT antibody was purchased from EnoGene (Nanjing, China). The β -actin rabbit antibody and goat anti-rat (HRP) secondary antibody were purchased from Bioss (Beijing, China). The goat anti-rabbit (HRP) secondary antibody was purchased from Sino Biological Inc. (Beijing, China). The cell lysis buffer, primary antibody dilution buffer and ECL kit were purchased from Beyotime (Shanghai, China).

Cell growth inhibition assay

HepG2, Hep3B and SKOV3 cells were seeded into a 96-well plate at 10000 cells per well, while Panc-1 cells at 5000 cells per well, and allowed to adhere overnight. Then the culture medium was removed and replaced with fresh media alone as control or containing various concentrations of the compounds. After indicated treatment in 72h, the number of viable cells remained was determined by MTT colorimetric assay at 490 nm. All experiments were done in triplicate and repeated three independent times.

The IC₅₀ value were calculated with the software program SPSS 22.0.

Lactate dehydrogenase (LDH) assay.

The LDH assay was measured by using commercial LDH kit (Cytotoxicity Assay Kit, Beyotime). The cancer cells were seeded 10,000 cells per well in 96-well plates, respectively. 24 hours later, the culture medium was removed and replaced with fresh media (1% FBS) alone or containing the test compounds. After 1h treatment, the plates were centrifuged for 5 minutes in a multiwell plate centrifuge (400 g), and 120 μ L supernatant of each well was transferred to a new 96-well plate. The LDH reaction mixture was freshly prepared according to the manufacturer's protocol and 60 μ L of this mixture was added to each well of the plates containing blank, control, or cells in culture. The plate was incubated at 25 °C for 30 min. Control was performed with release agent and blank medium, and set as 100% and 0% LDH release, respectively. The relative LDH release is defined by the ratio of LDH released over total LDH in the cells. All samples were performed in triplicate.

Apoptosis and necrosis assay on flow cytometry

The cancer cells were seeded in 6-well plates (2.5×10^5 cells/well) and allowed to adhere and proliferate overnight. Culture medium was then removed and fresh media containing the compound was added. After 48 hours treatment, the cells were harvested and washed with cold PBS. The samples were pelleted again through centrifugation and resuspended in binding buffer. Annexin-V/FITC and propidium iodide (PI) was then added to the cells and the samples were incubated for 15 minutes at room temperature.

After staining, flow cytometry was performed on Fluorescence Activated Cell Sorting (Beckman Coulter). Each sample was performed in triplicate.

Western blotting analysis

The cancer cells after being treated by the compounds for 48h were lysed in lysis buffer. A total of 10-50 µg protein were quantified and loaded into the 10% SDS-PAGE gel. After electrophoresis, the proteins in the gel were transferred to a PVDF membrane. Then the membrane was blocked in 5% (w/v) skim milk in TBS-T for 1 h and incubated with the primary antibodies at 4 °C overnight. The membrane was washed with TBS-T and incubated with HRP-conjugated secondary antibody for 1 h. Specific proteins were detected using an ECL kit.

Oil red O staining

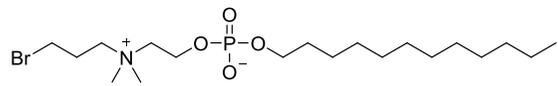
HepG2 cells were grown at an initial density of 2×10^5 cells/well on glass coverslips in 12-well plates and treated with different concentrations of compounds for 48 h. Cells were then washed three times with PBS and fixed with 4% paraformaldehyde for 30 min. After fixation, cells were washed with PBS and stained with freshly diluted Oil Red O working solution (0.5% Oil Red O in isopropanol: H₂O = 3:2) for 1 h at room temperature. Then cells were washed again with PBS several times to remove excess stain. Cell nuclei were stained with haematoxylin for 30 seconds and washed with ddH₂O. The stained samples were observed using a microscope (Leica, DMI8, Germany).

Measurements of cellular triglyceride (TG) content in HepG2 Cells

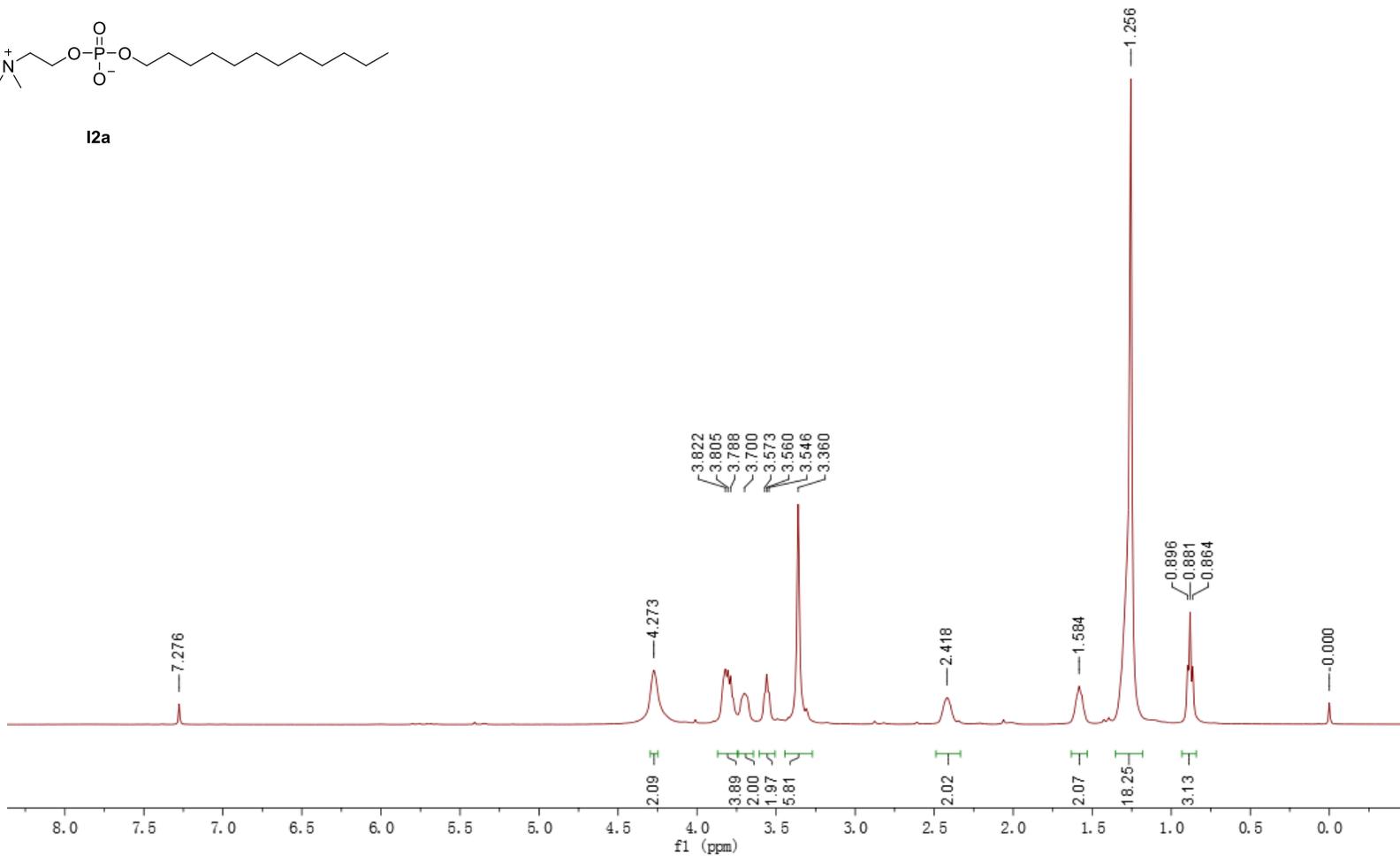
HepG2 cells were grown at an initial density of 2×10^5 cells/well in a 12-well plate and treated with different concentrations of compounds for 48 h. Cells were washed three times with PBS and were lysed in RIPA buffer, then the supernatant was collected. The intracellular TG levels were measured using enzymatic colorimetric assay kits and the protein concentrations were determined using the bicinchoninic acid (BCA) method. Intracellular TG contents were normalized to cellular protein content. All results were presented as the mean \pm SD. Data analysis was performed using the IBM SPSS 22.0 software package for Windows (SPSS, Chicago, IL, USA). Statistical differences were analyzed by one-way ANOVA, followed by Fisher's least significant difference (LSD's) multiple comparison test. Statistical significance was shown as * $p < 0.05$, ** $p < 0.01$, and *** $p < 0.001$.

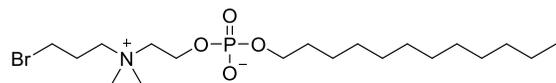
Reference

1. North, E. J.; Osborne, D. A.; Bridson, P. K.; Baker, D. L.; Parrill, A. L. *Bioorg. Med. Chem.* 2009, **17**, 3433–3442.

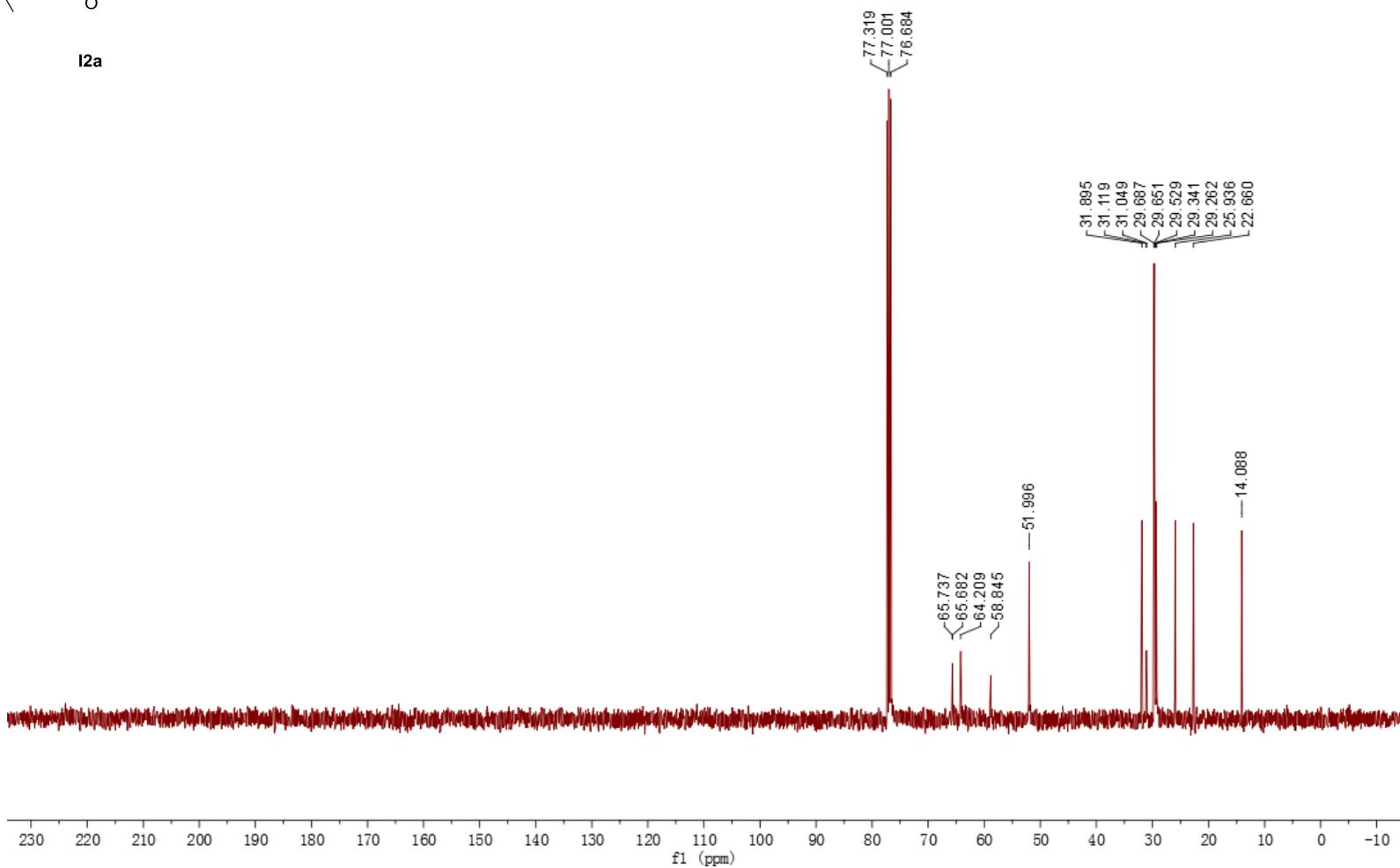


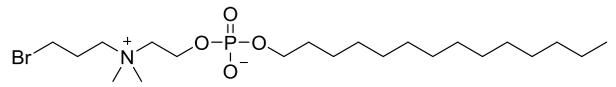
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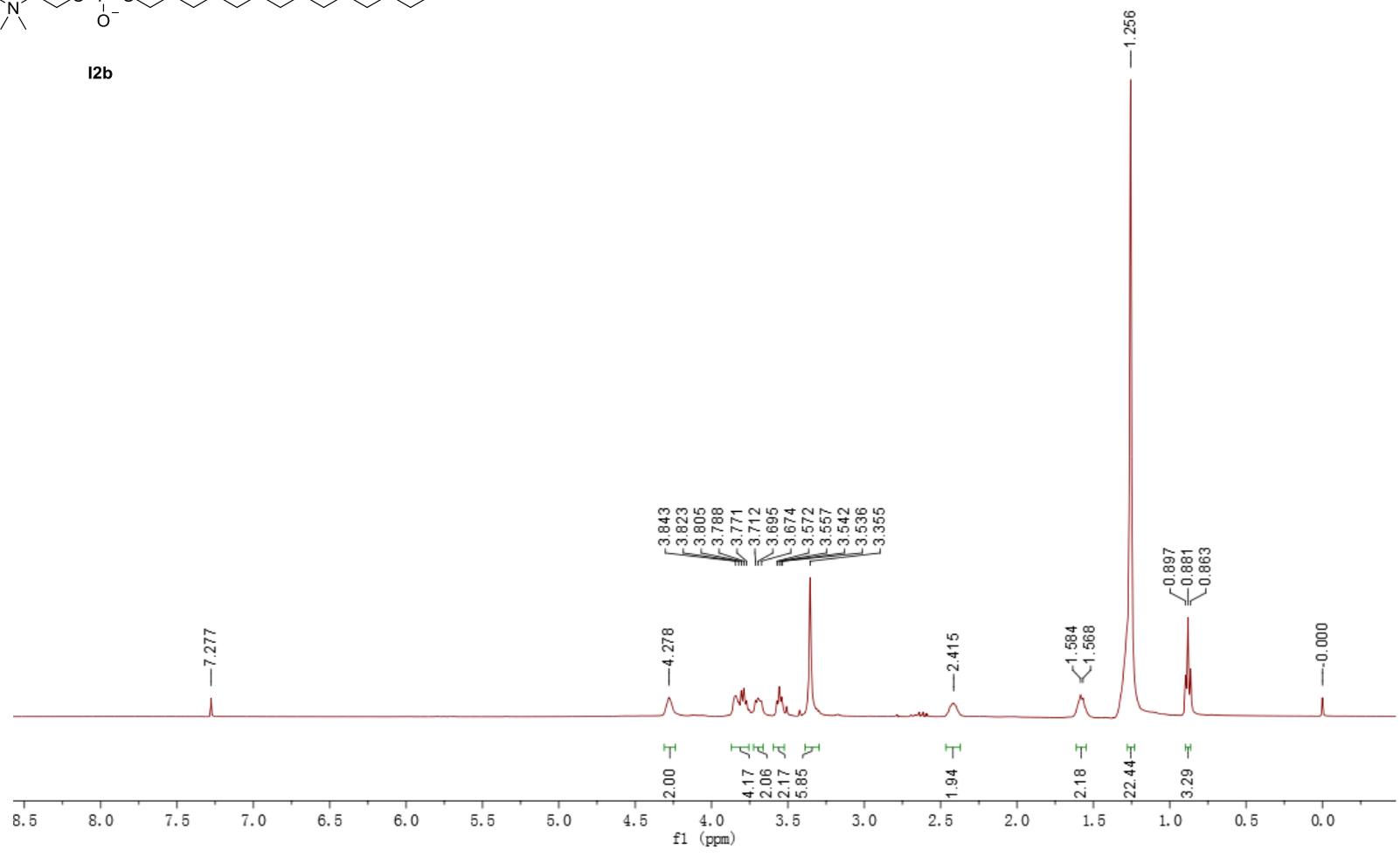


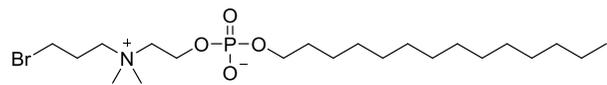
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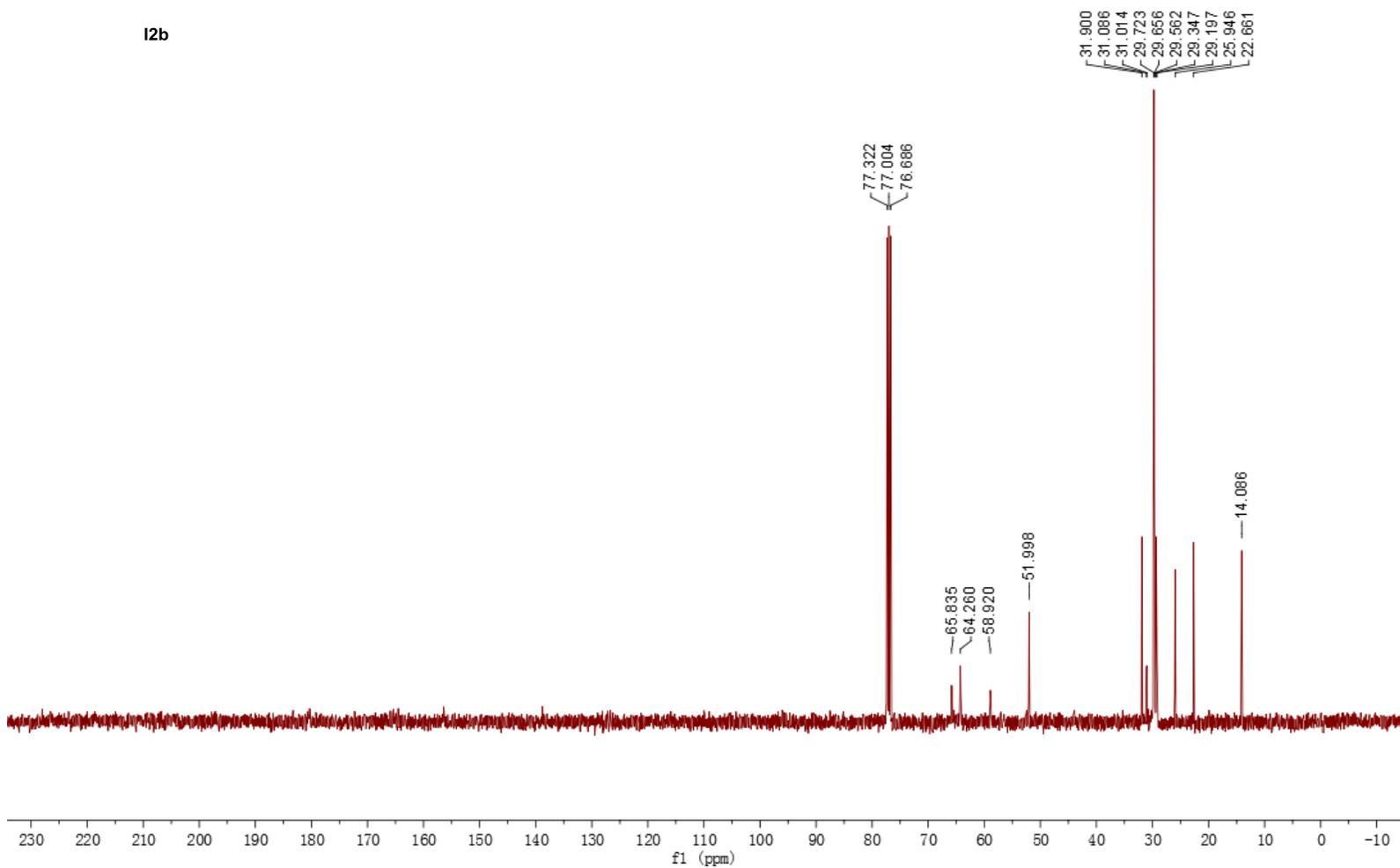


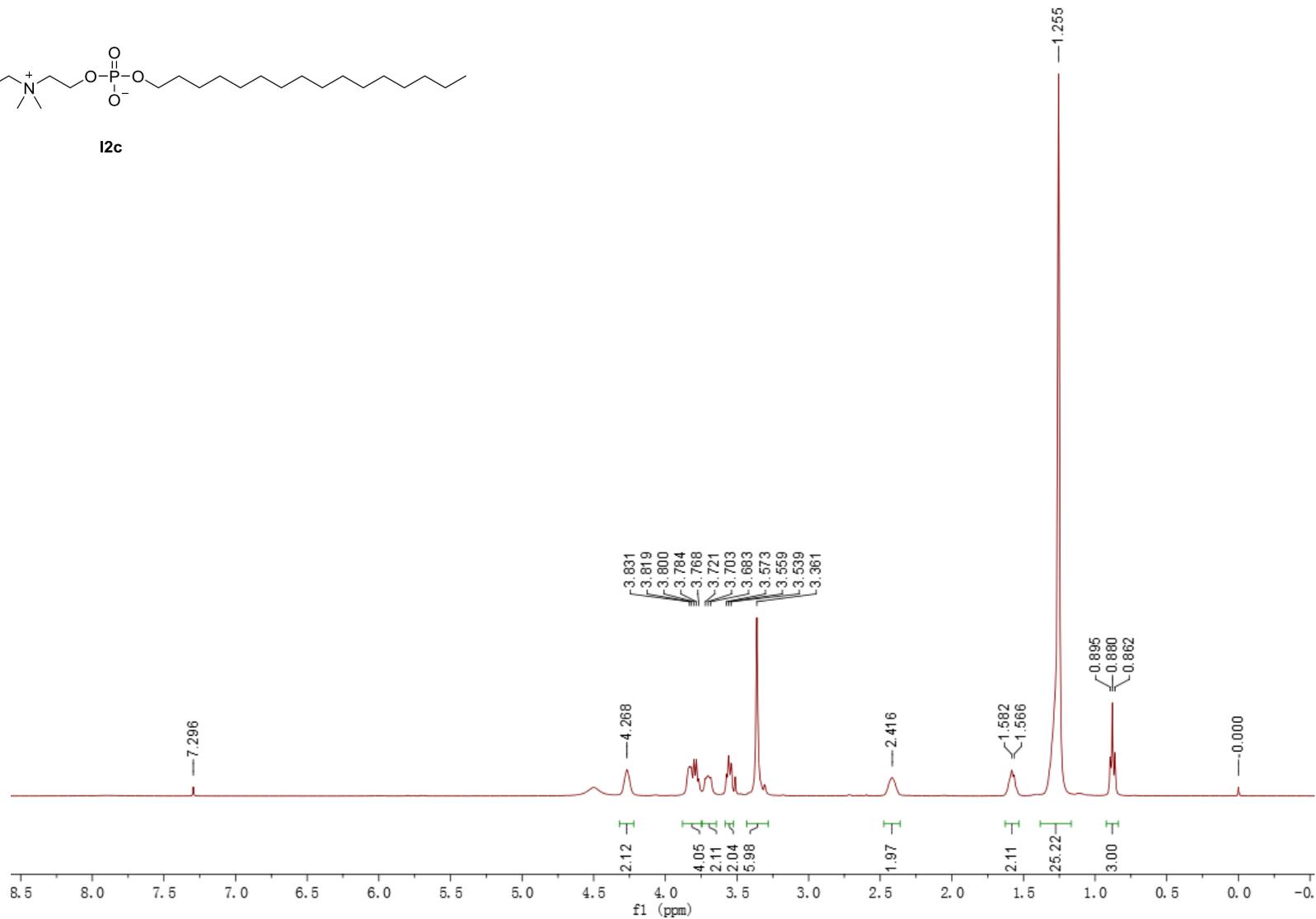
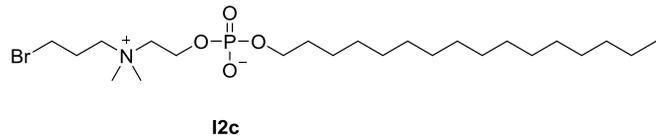
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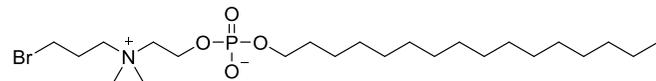




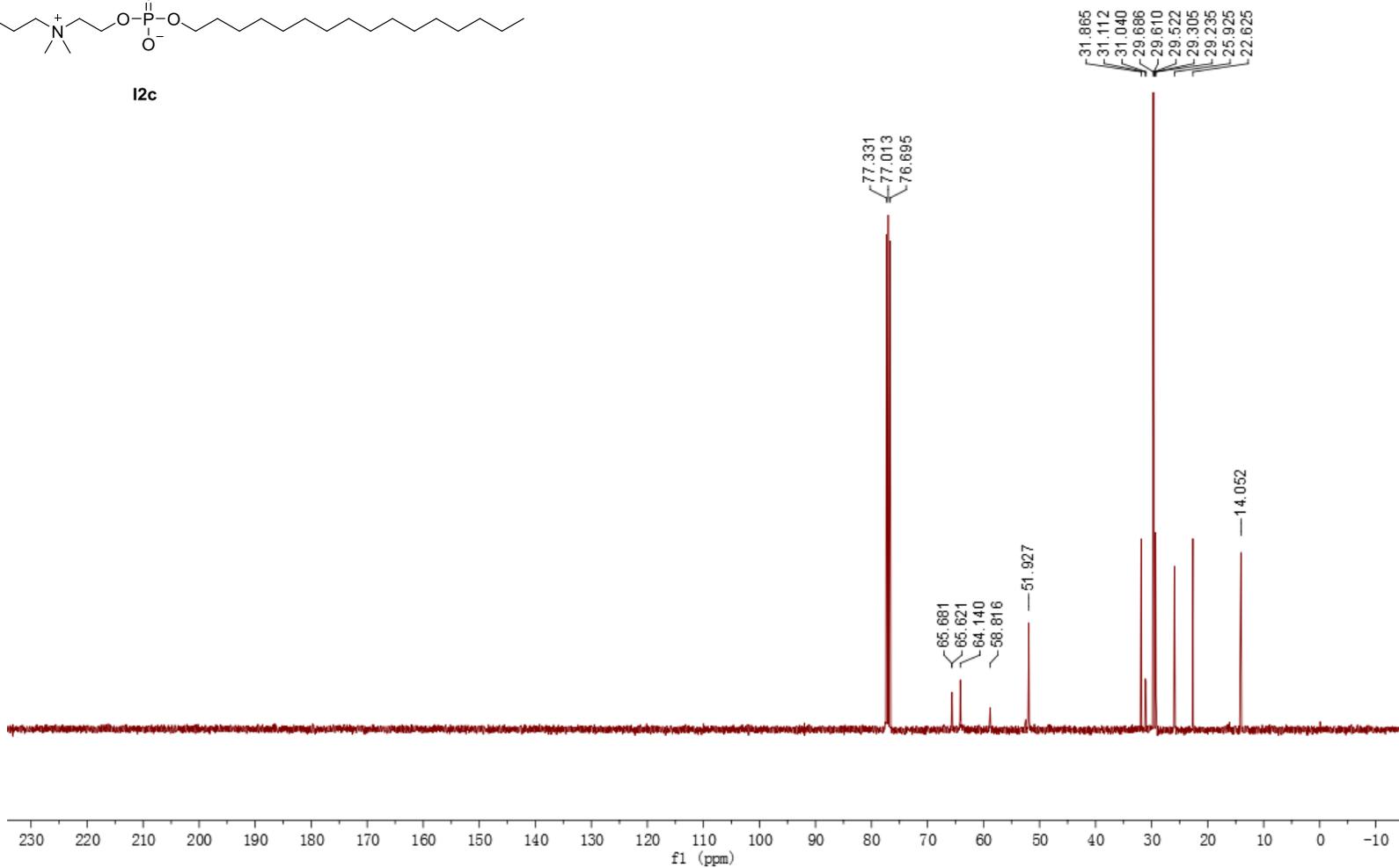
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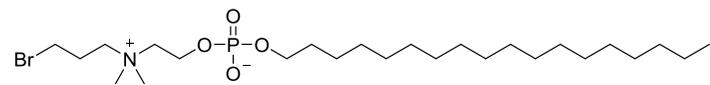




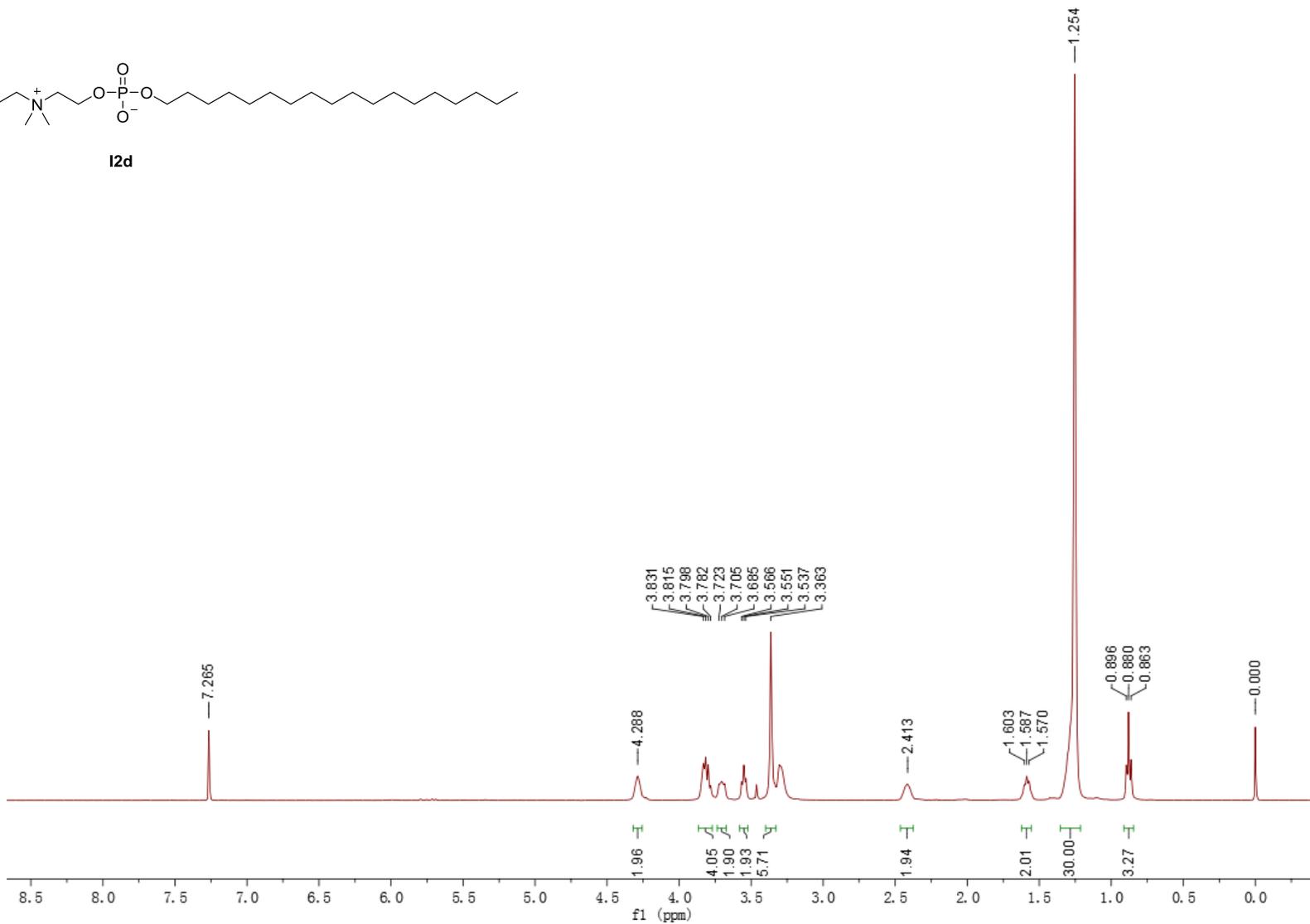


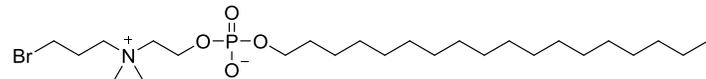
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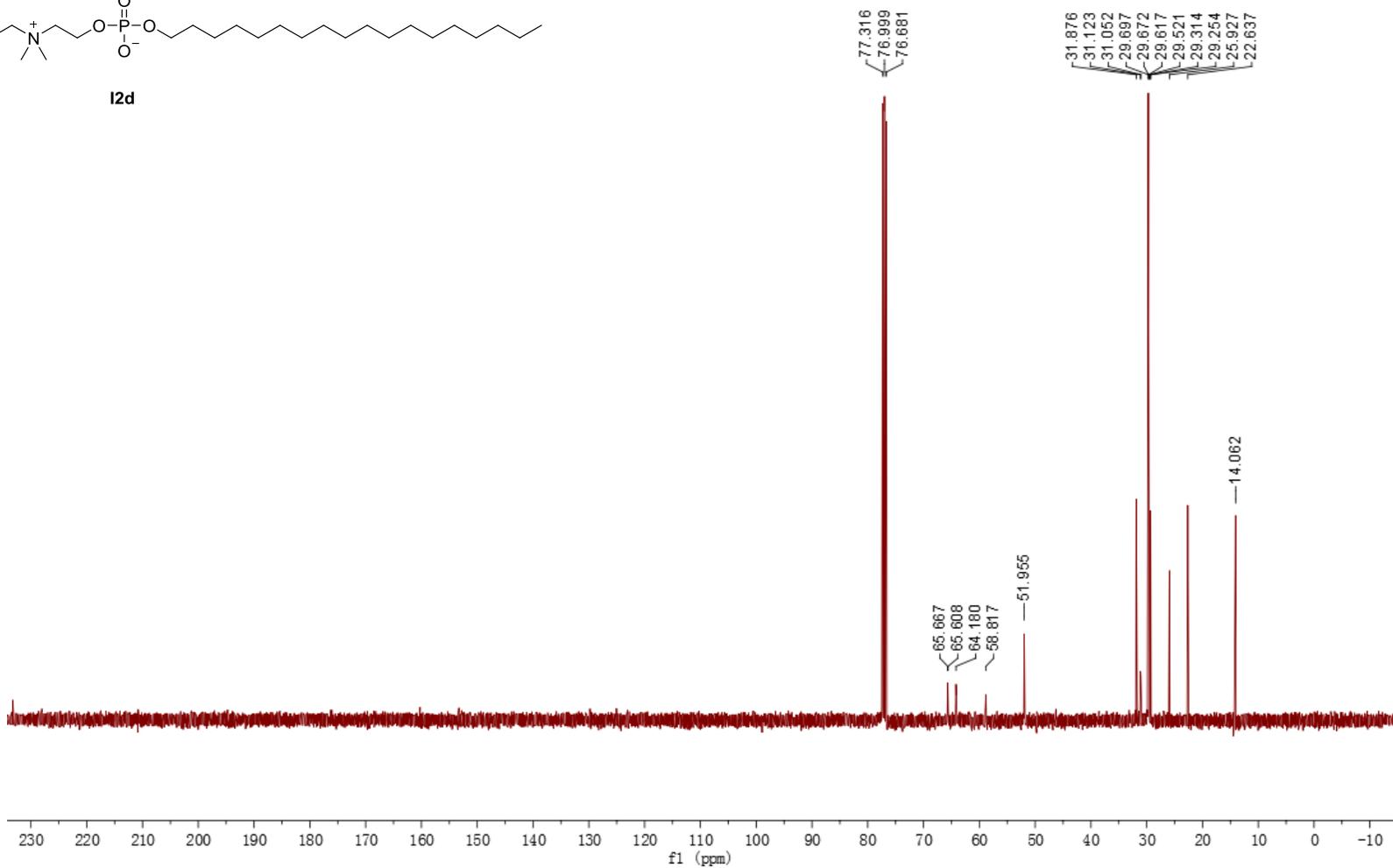


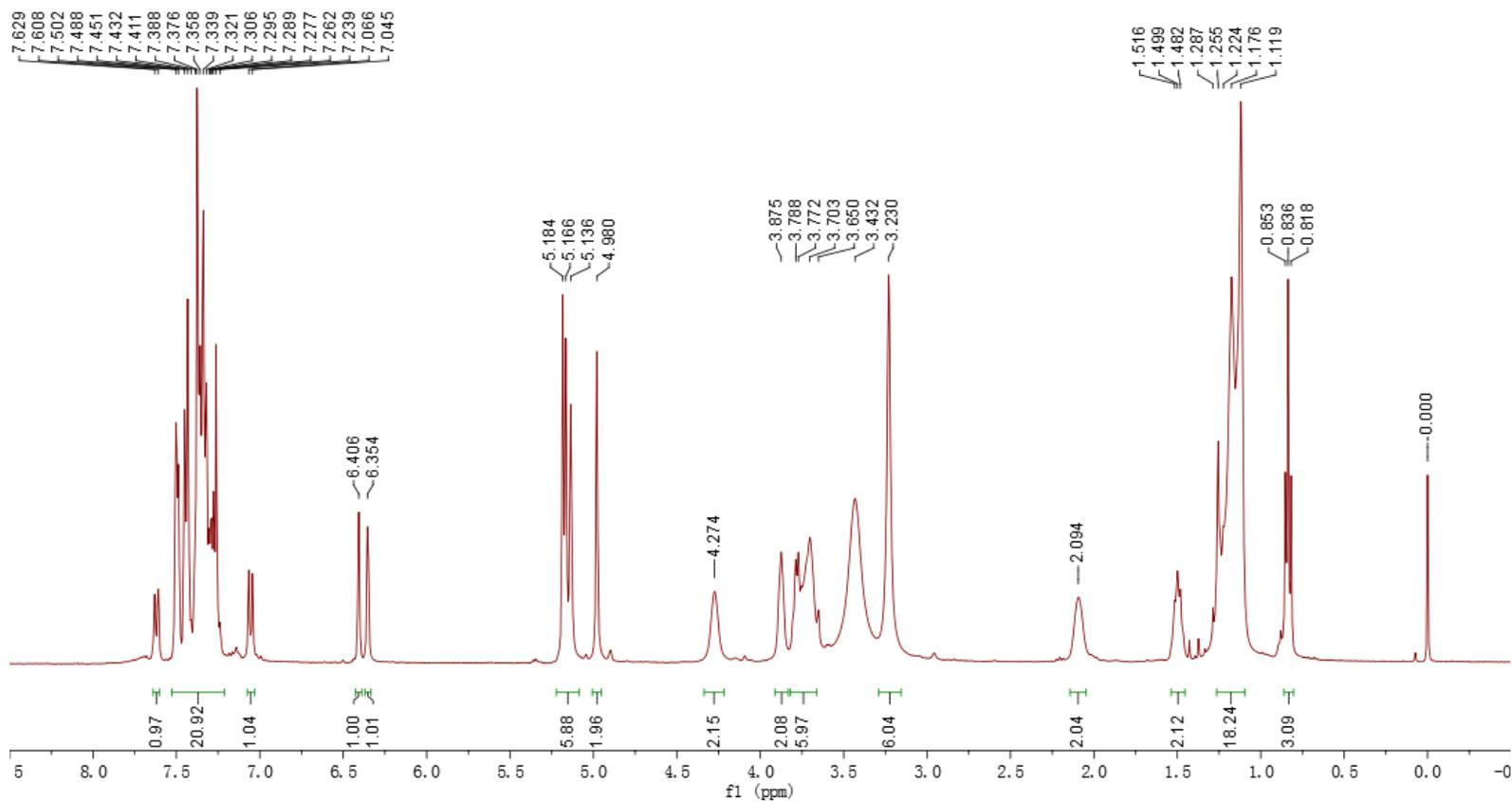
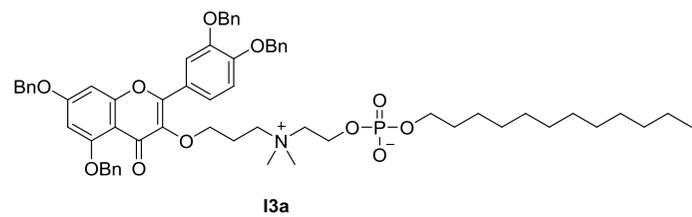
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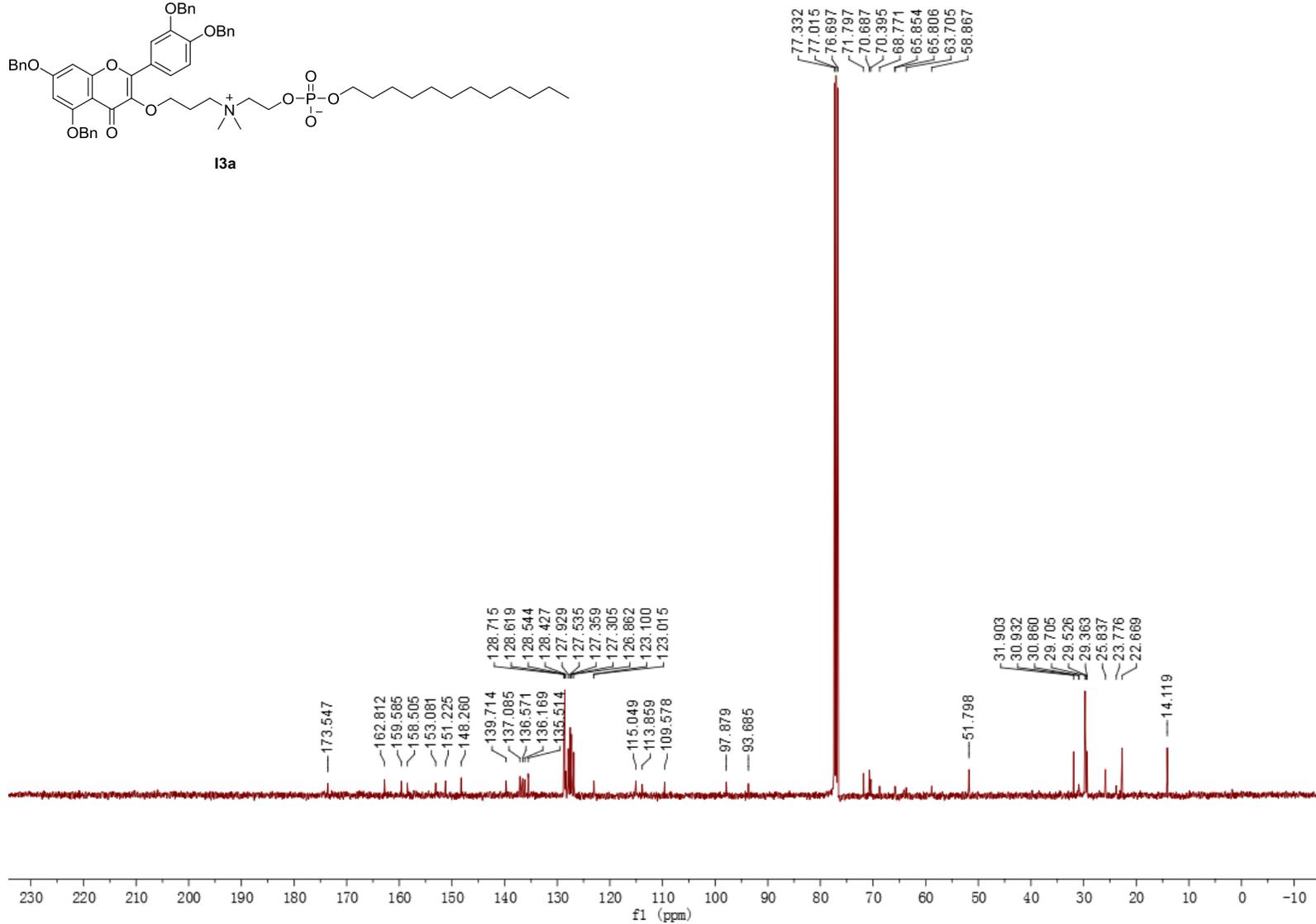
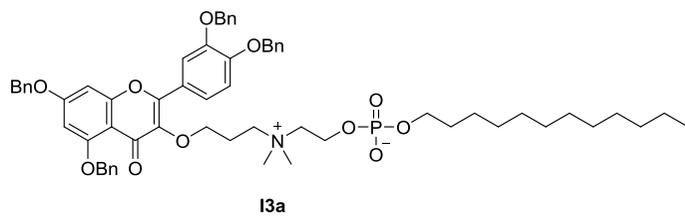


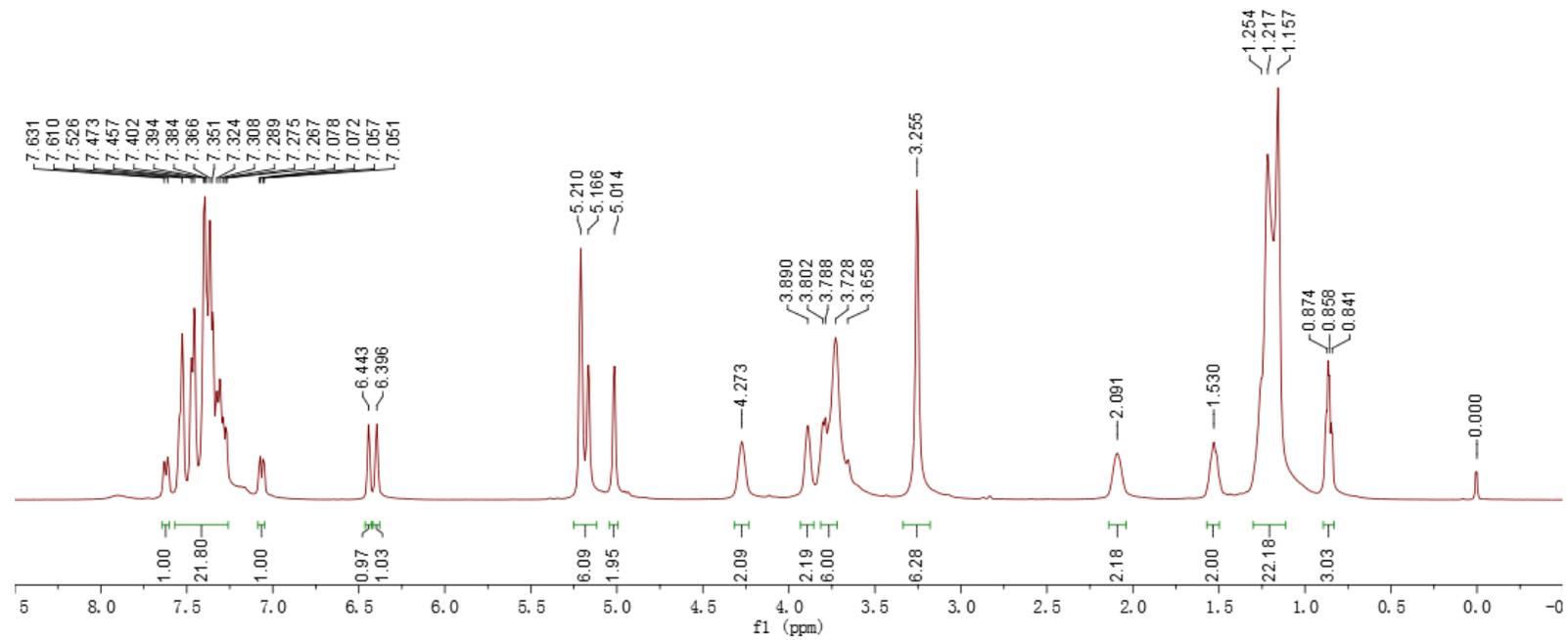
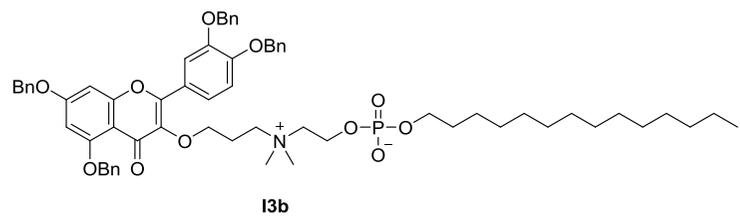


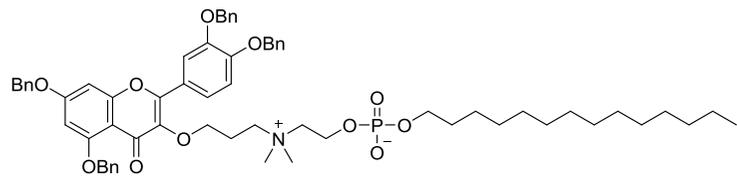
12d



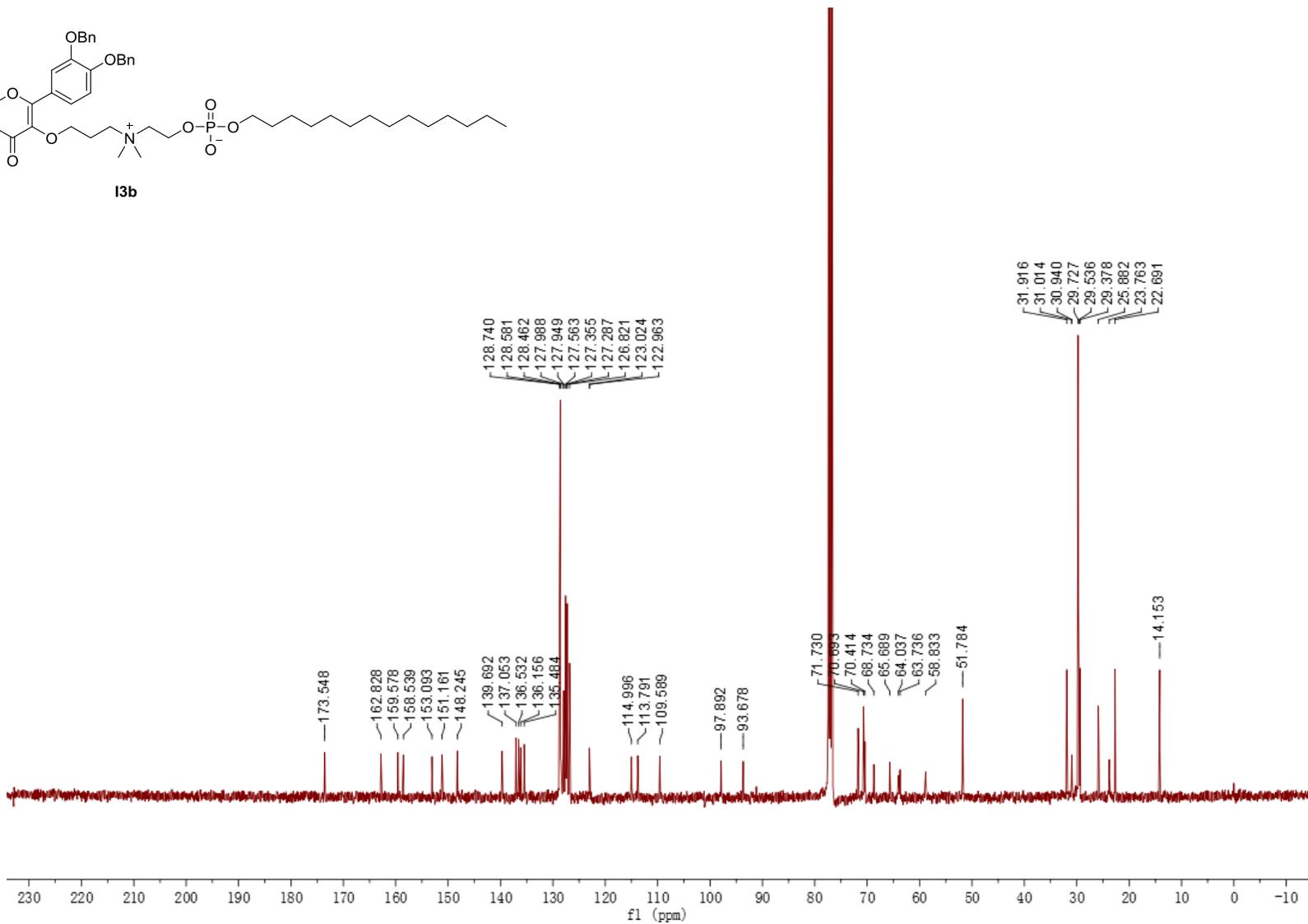


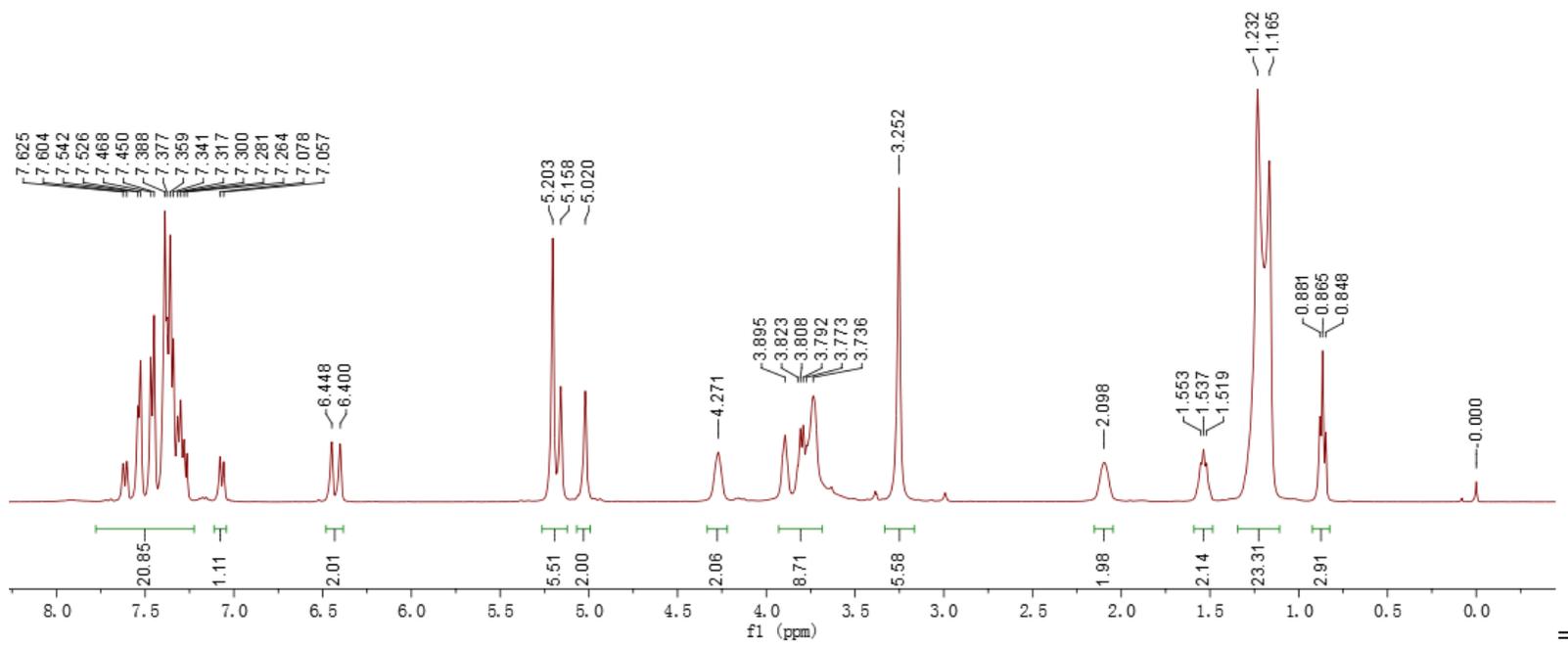
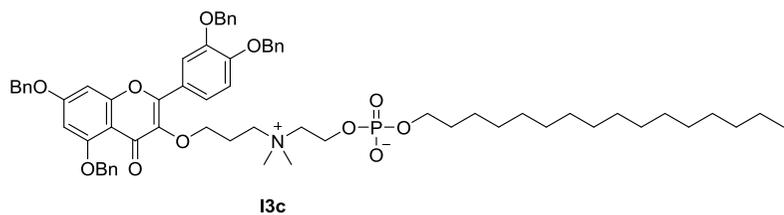


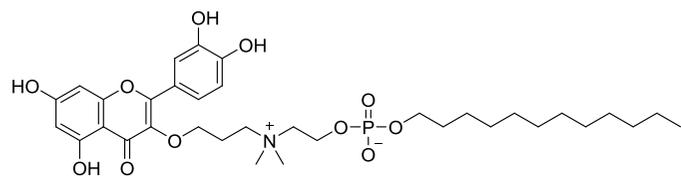




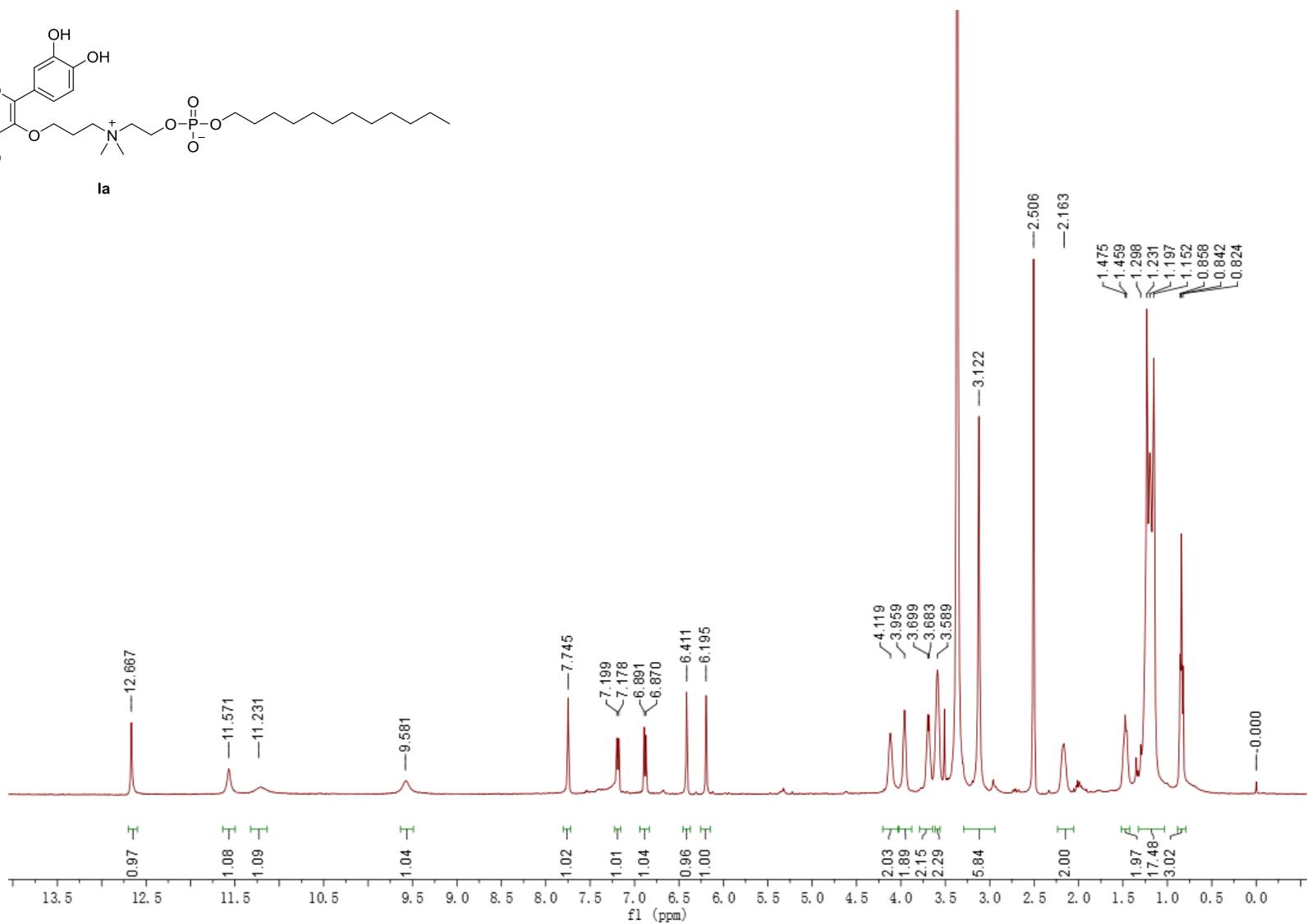
13b

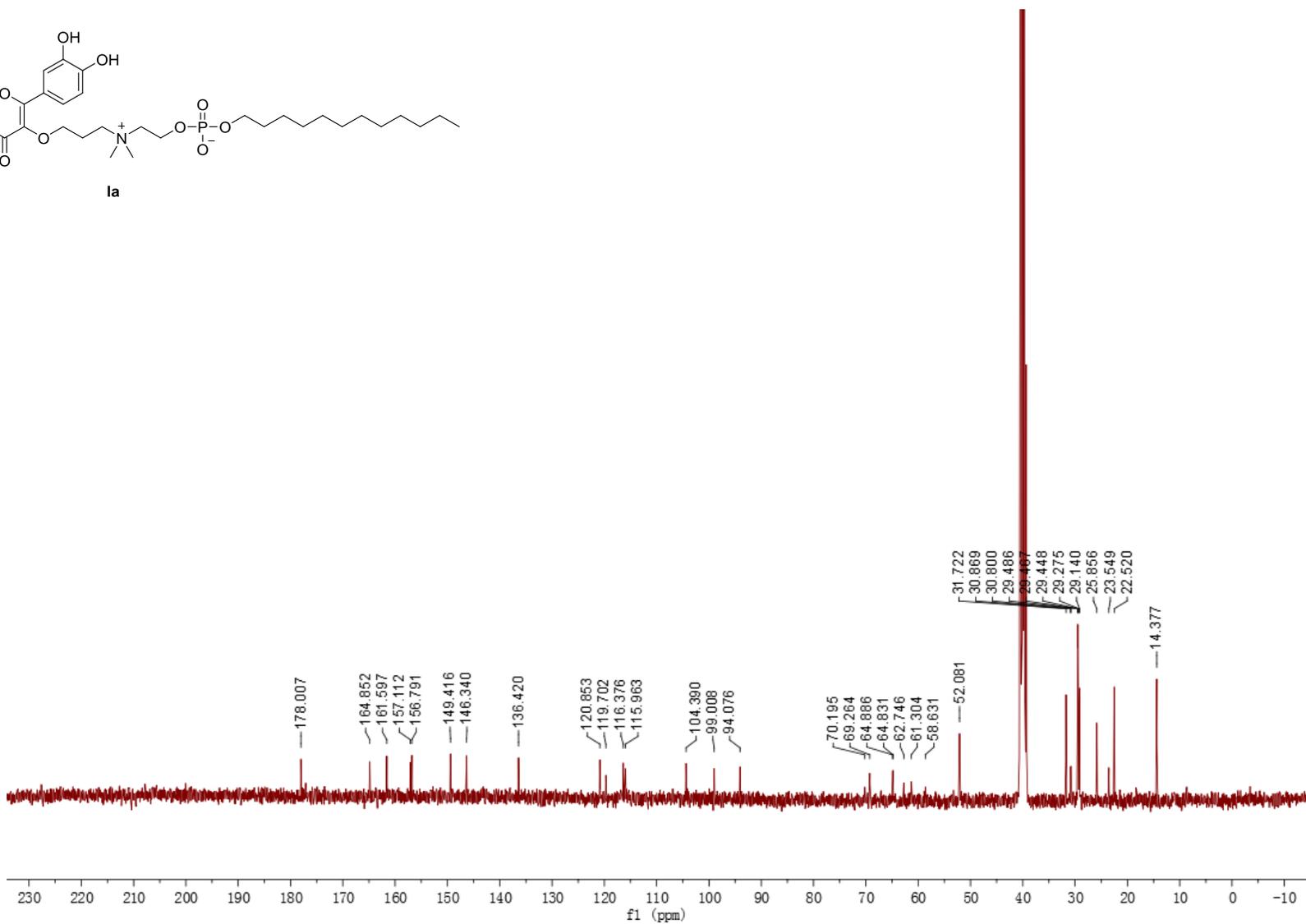
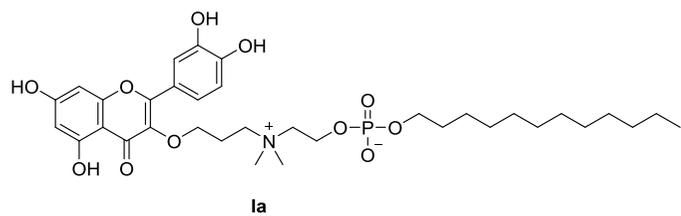


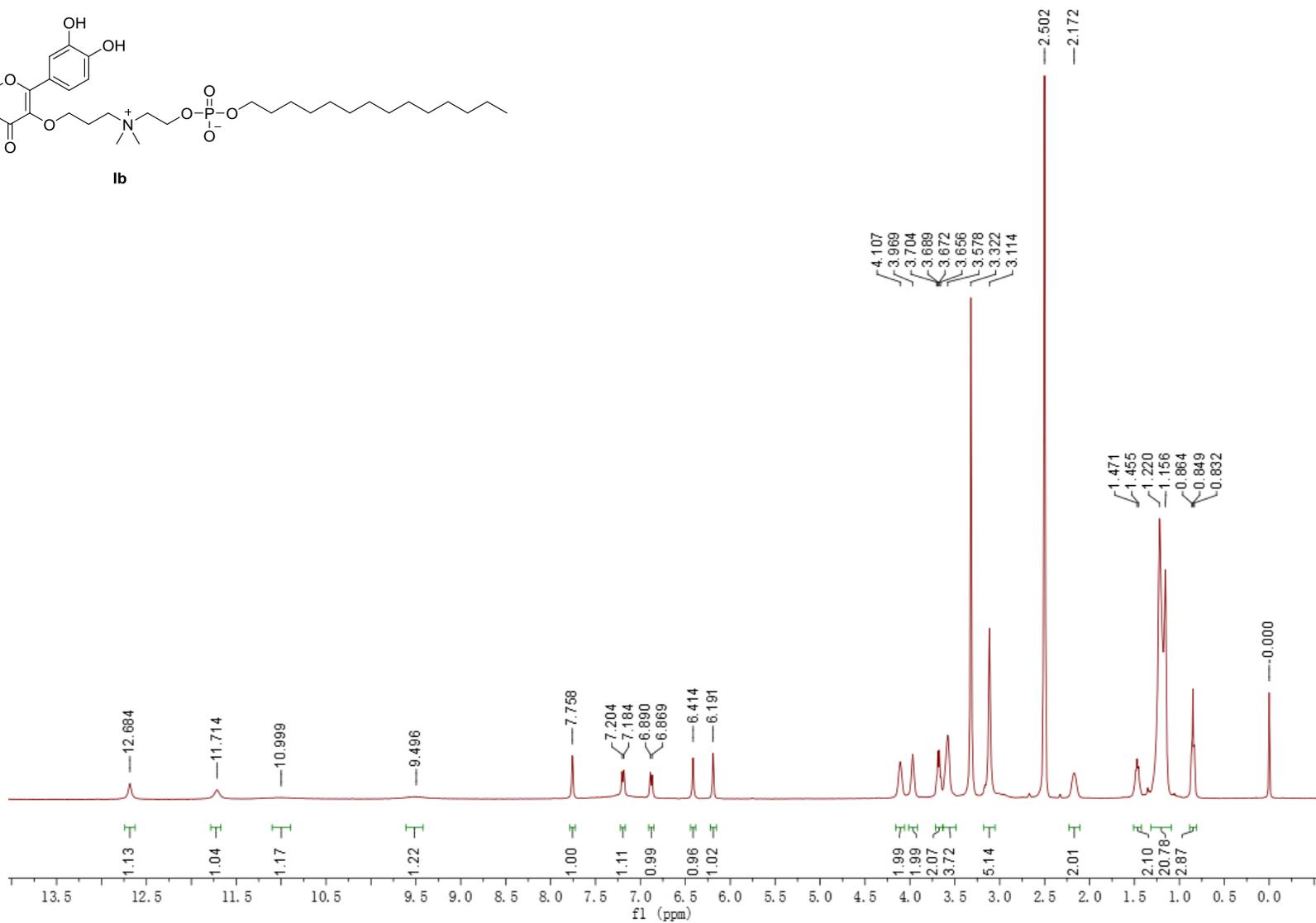
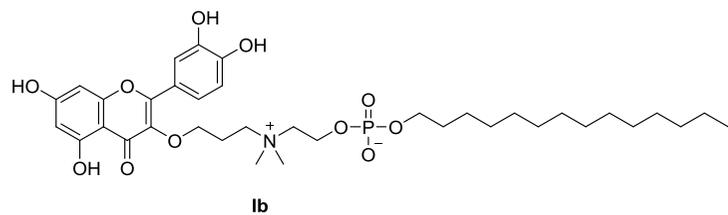


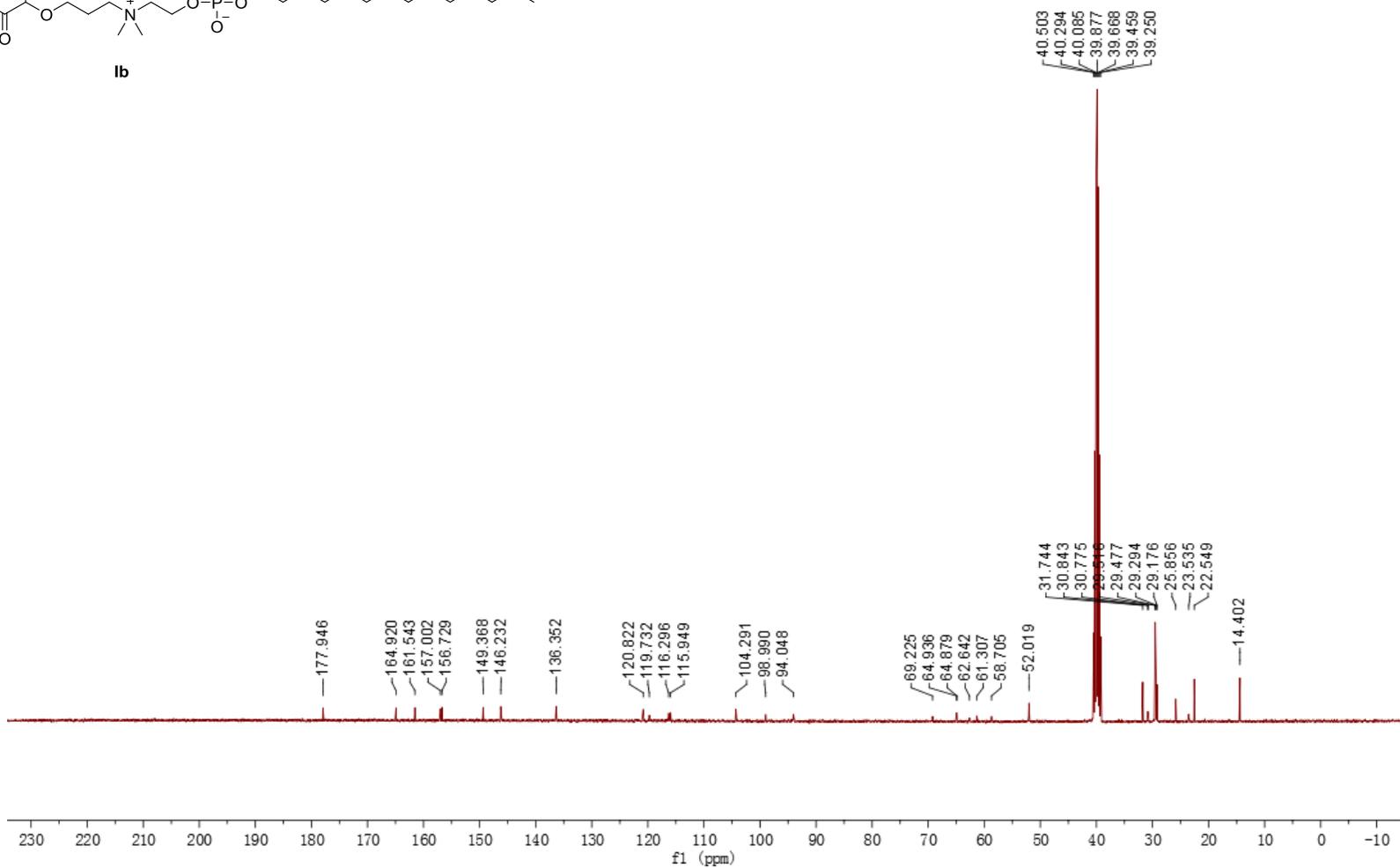
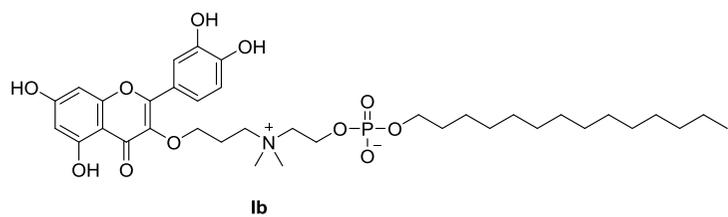


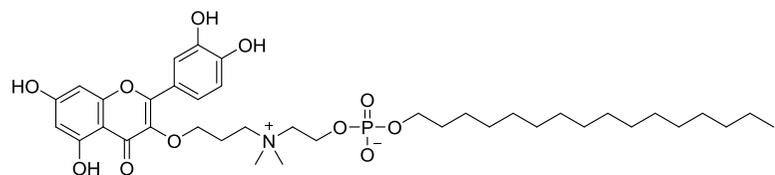
1a



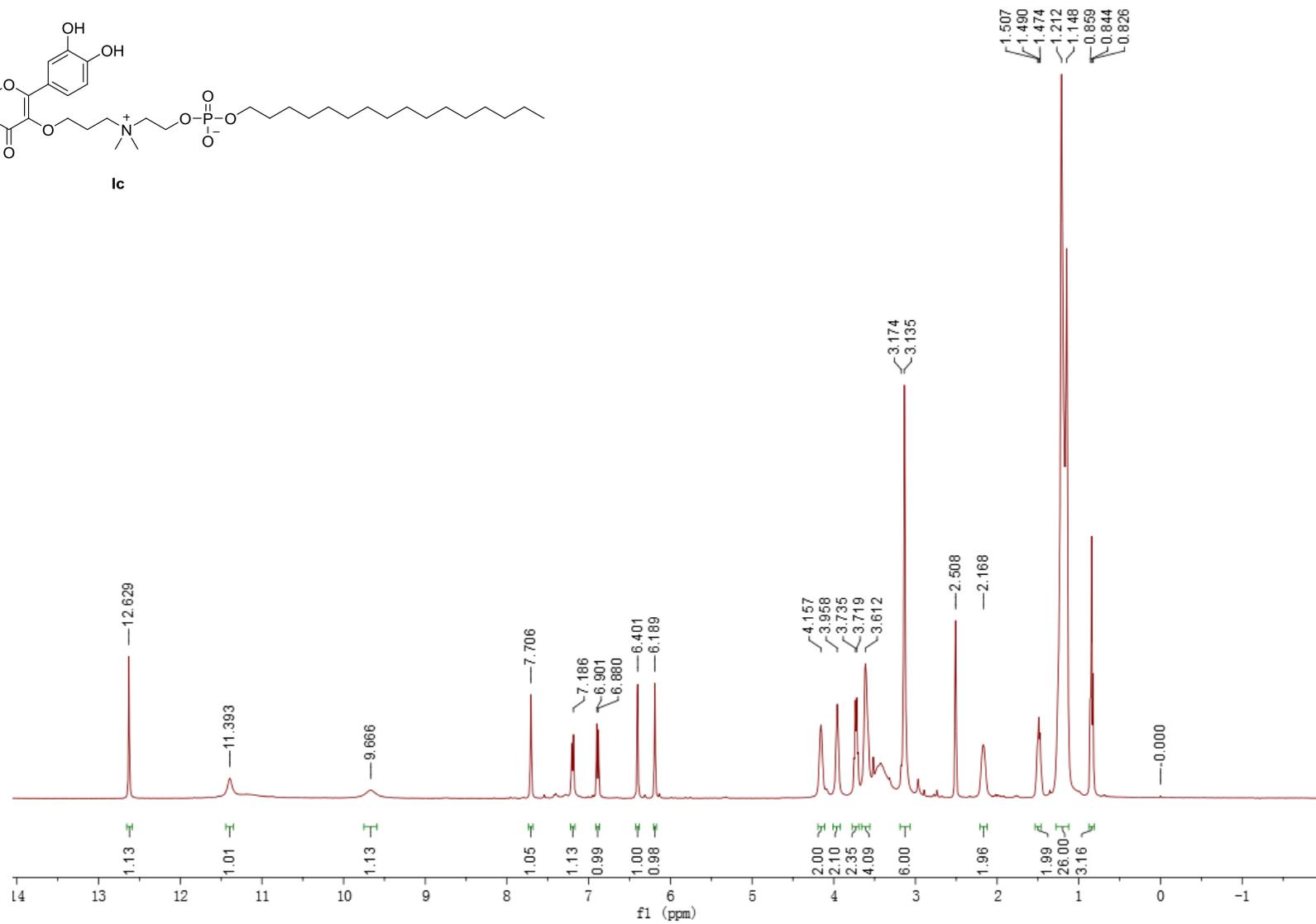


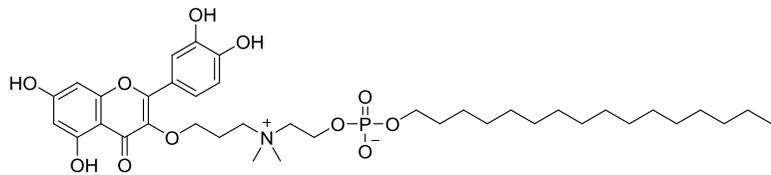




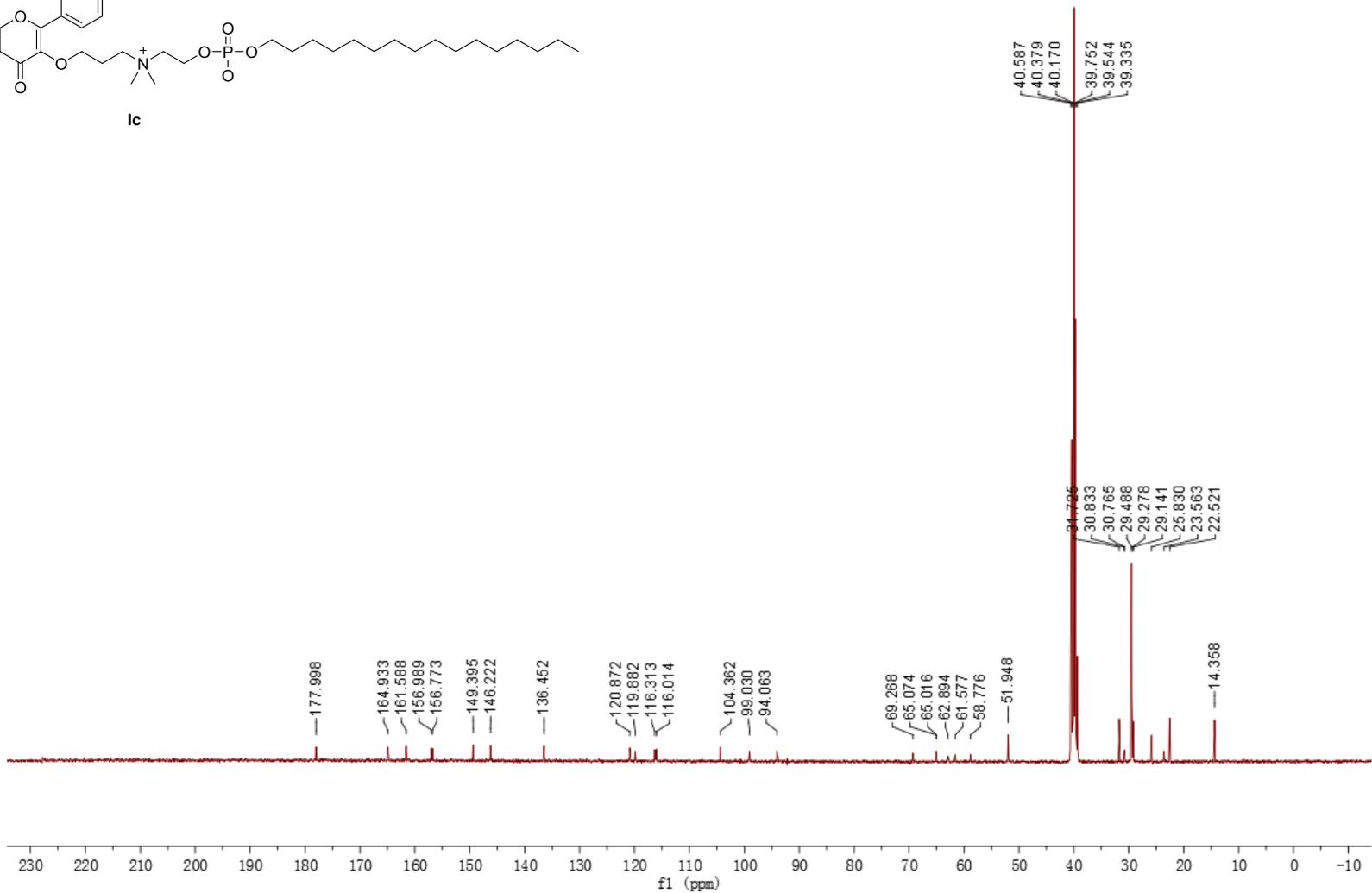


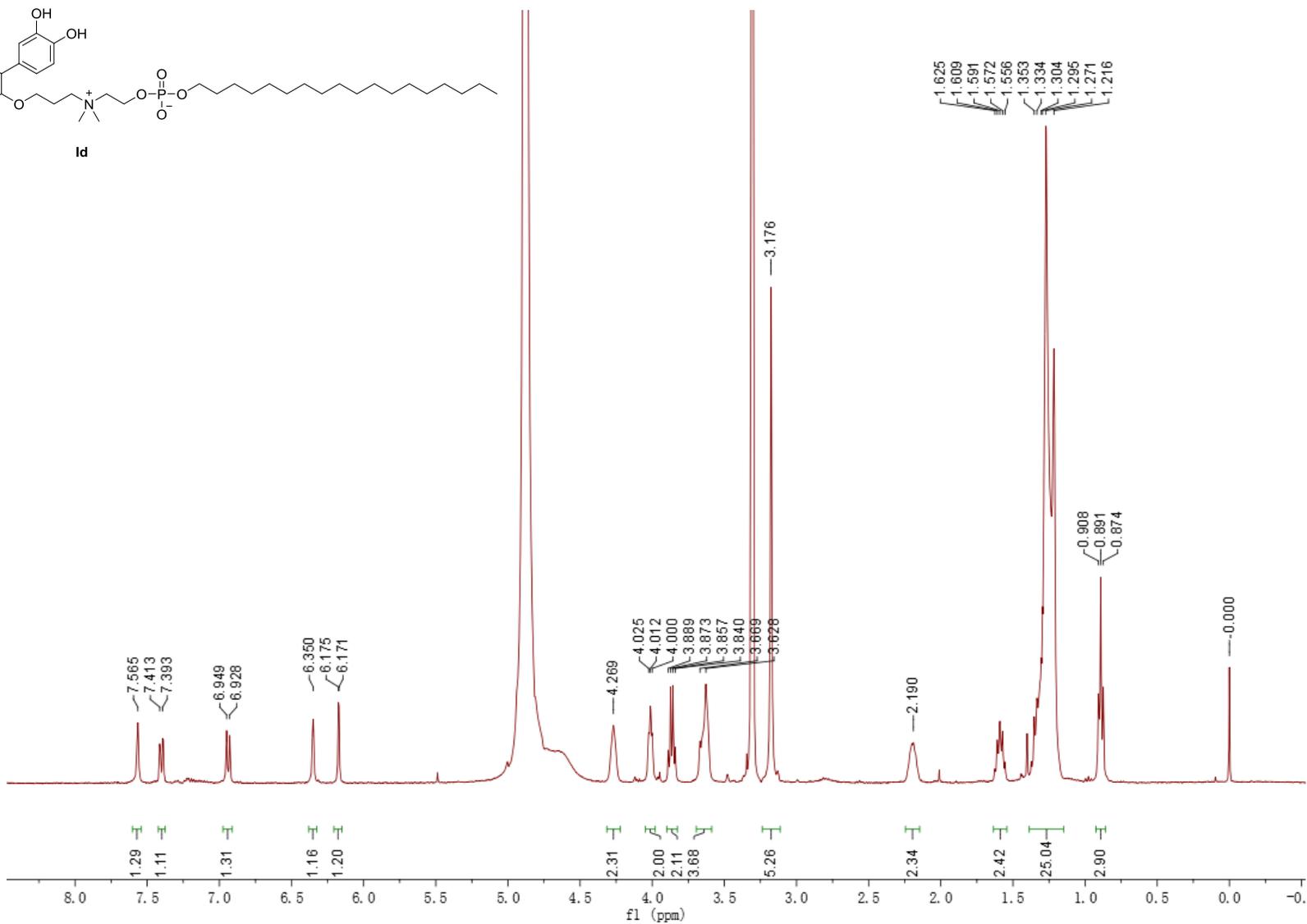
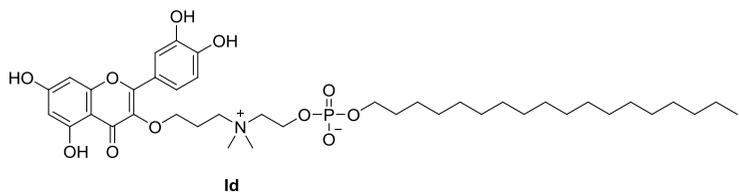
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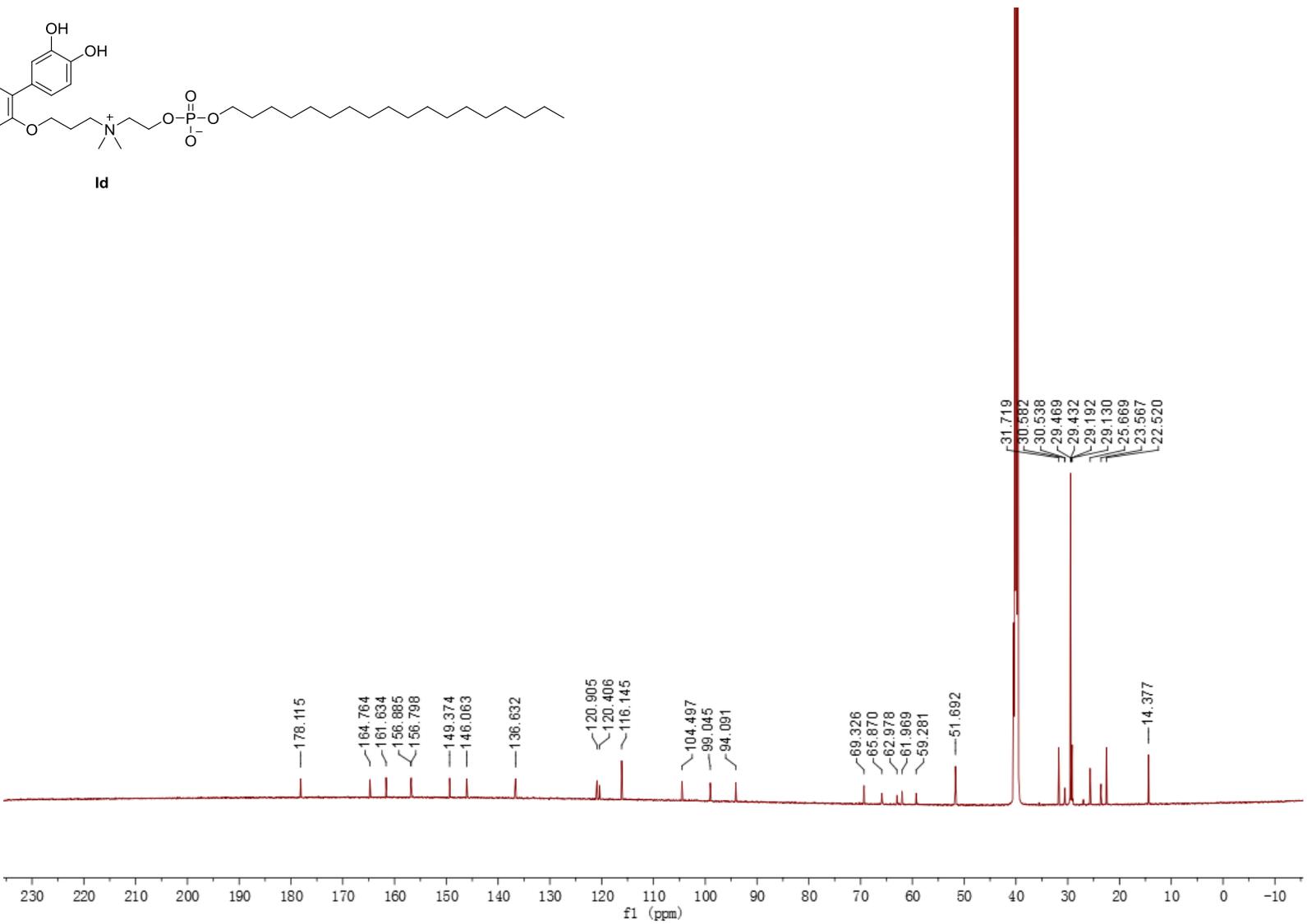
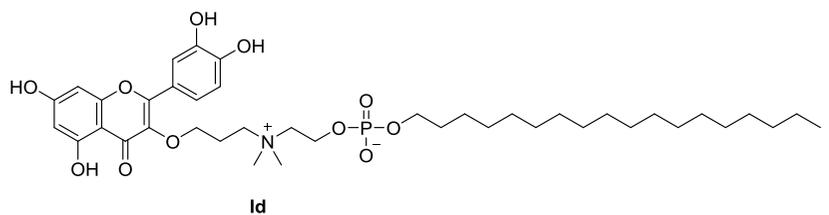


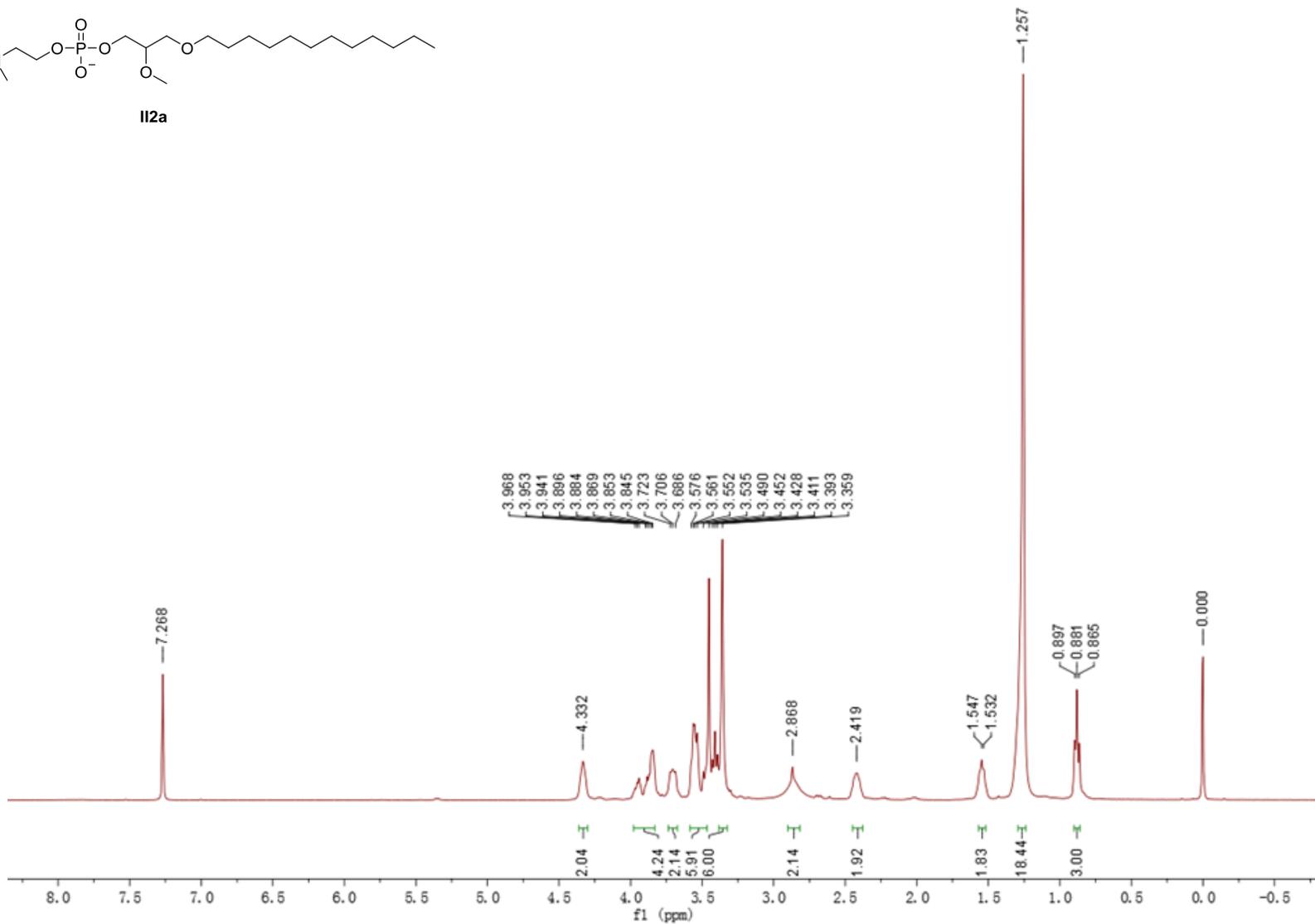
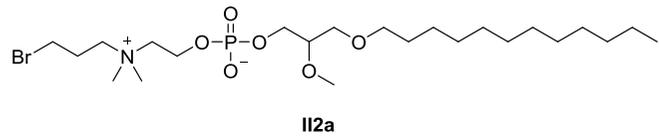


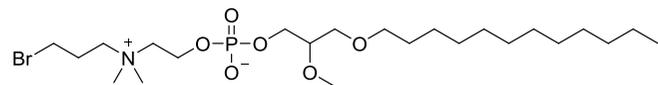
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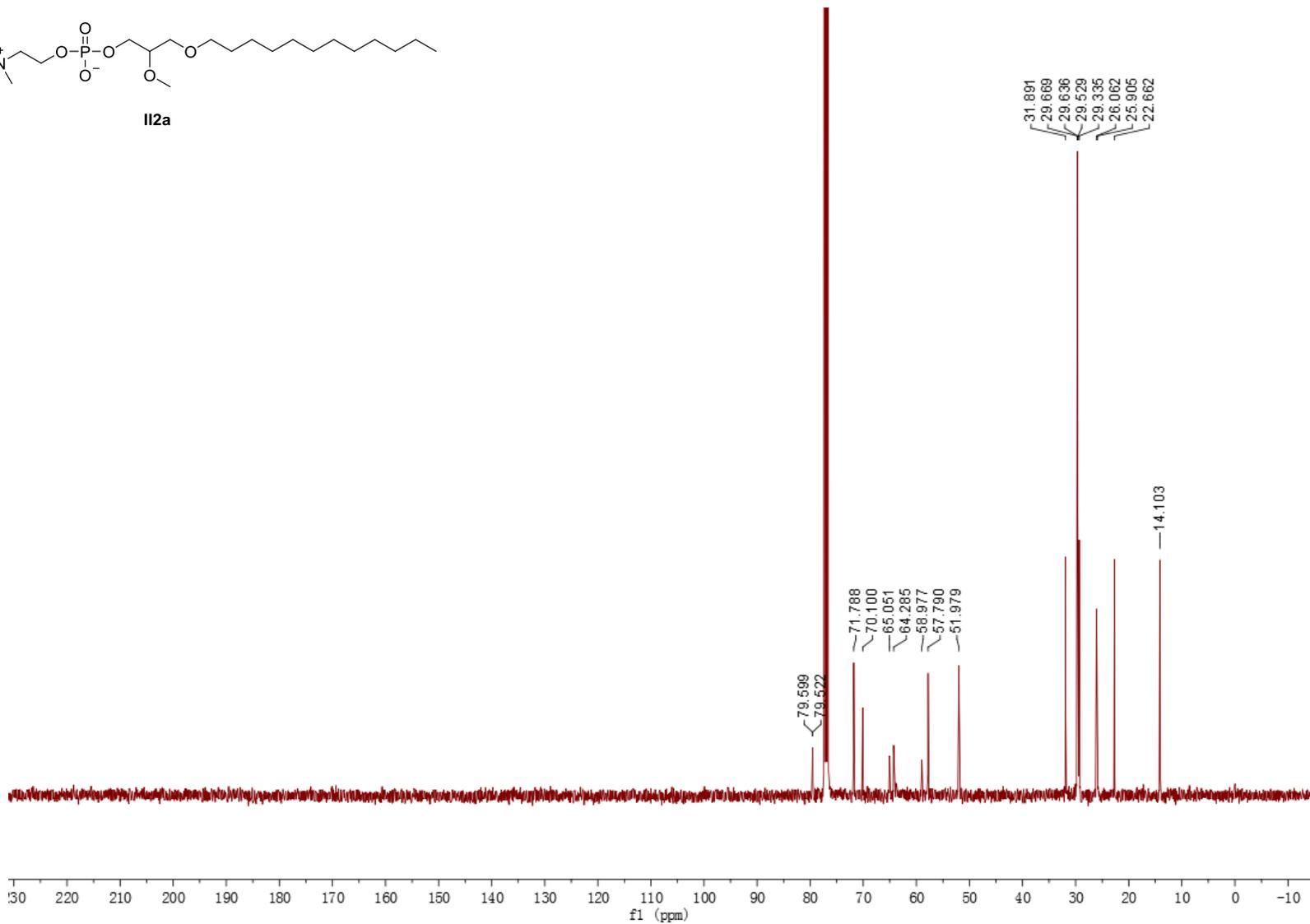


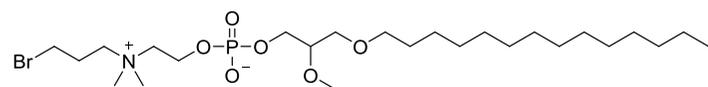




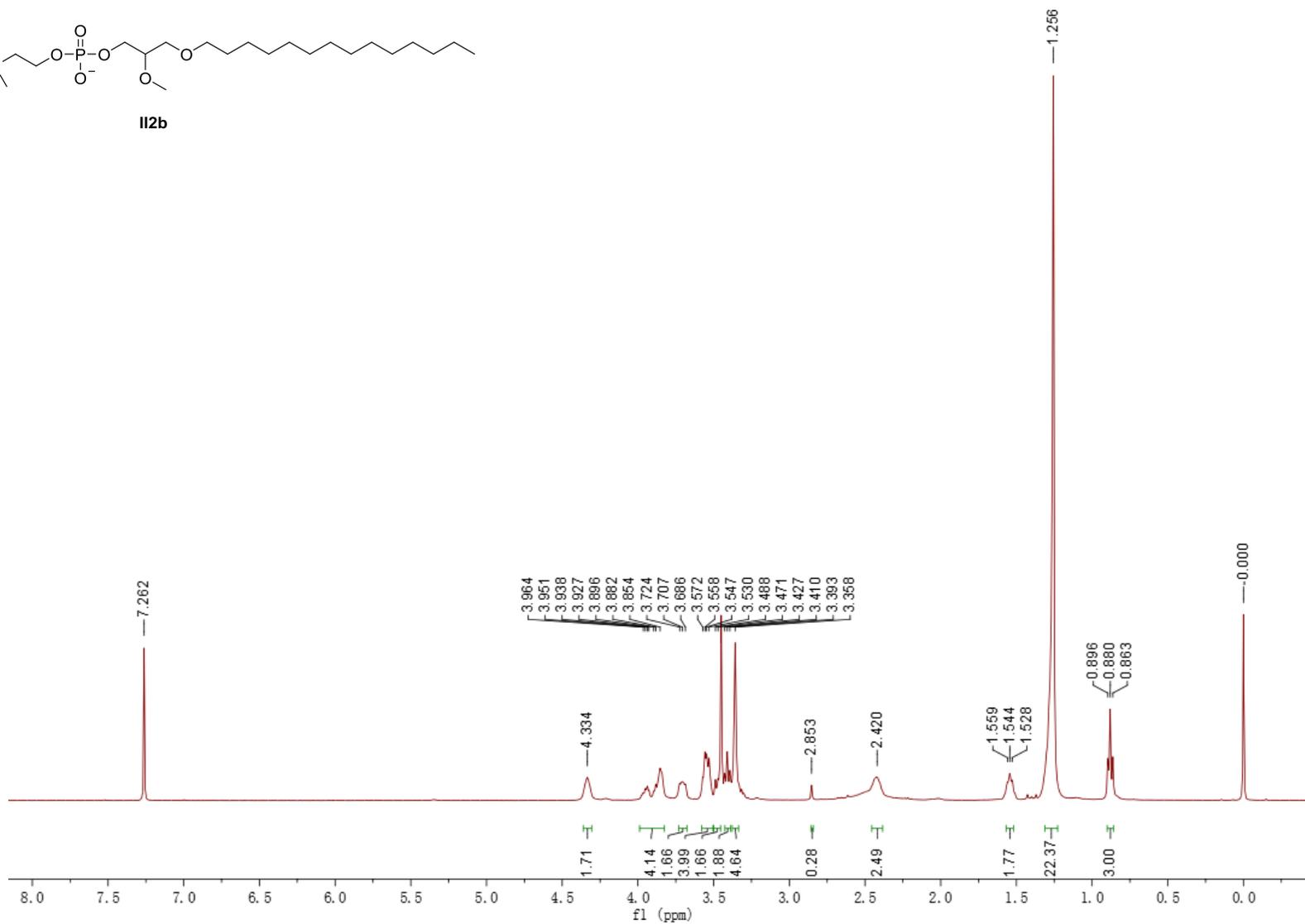


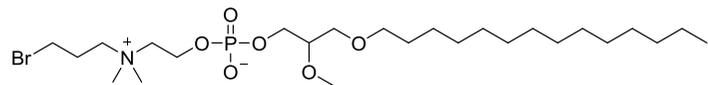
II2a



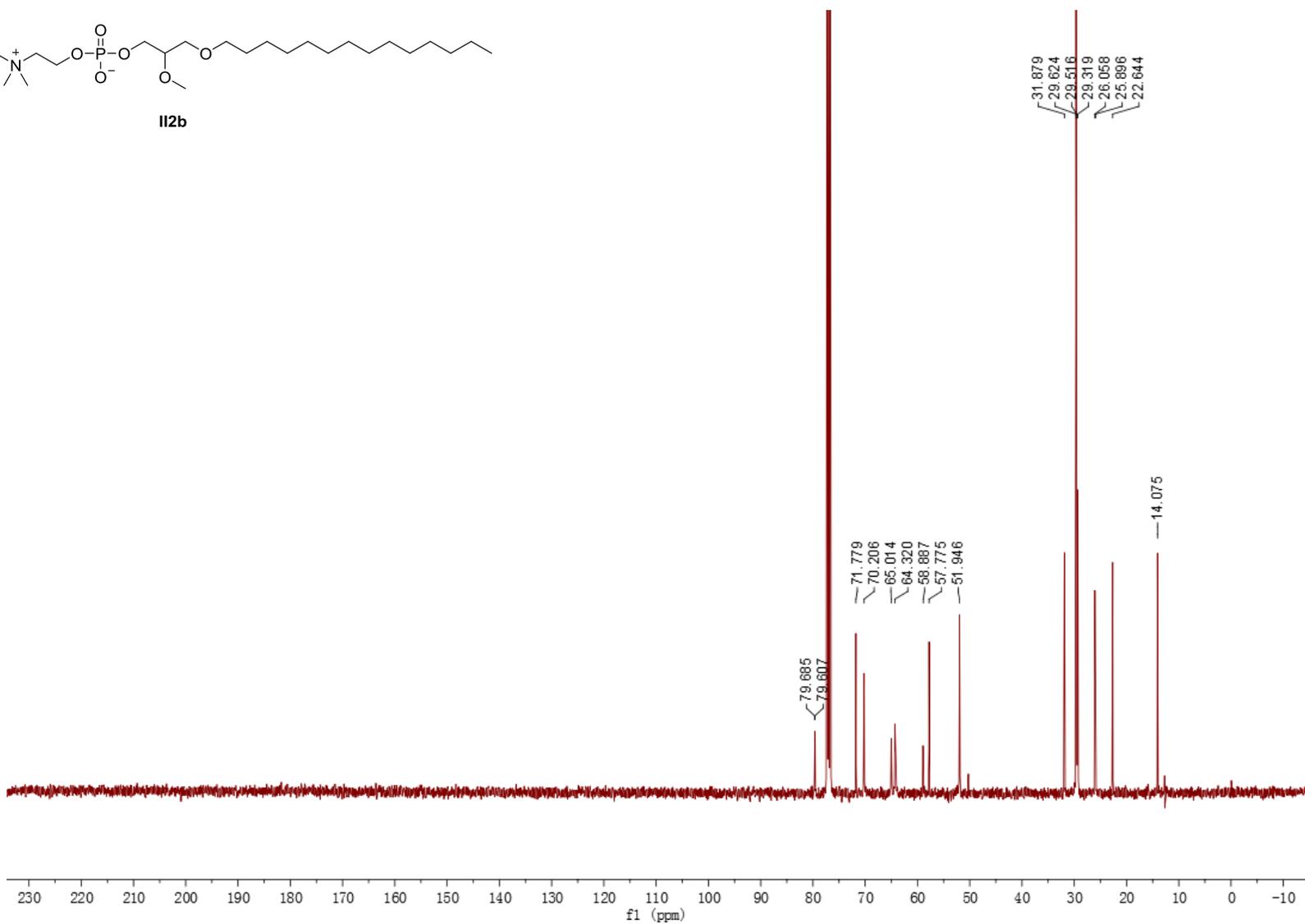


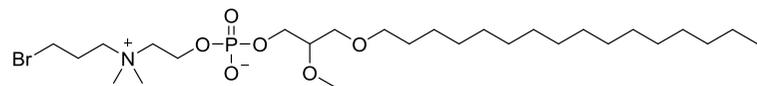
II2b



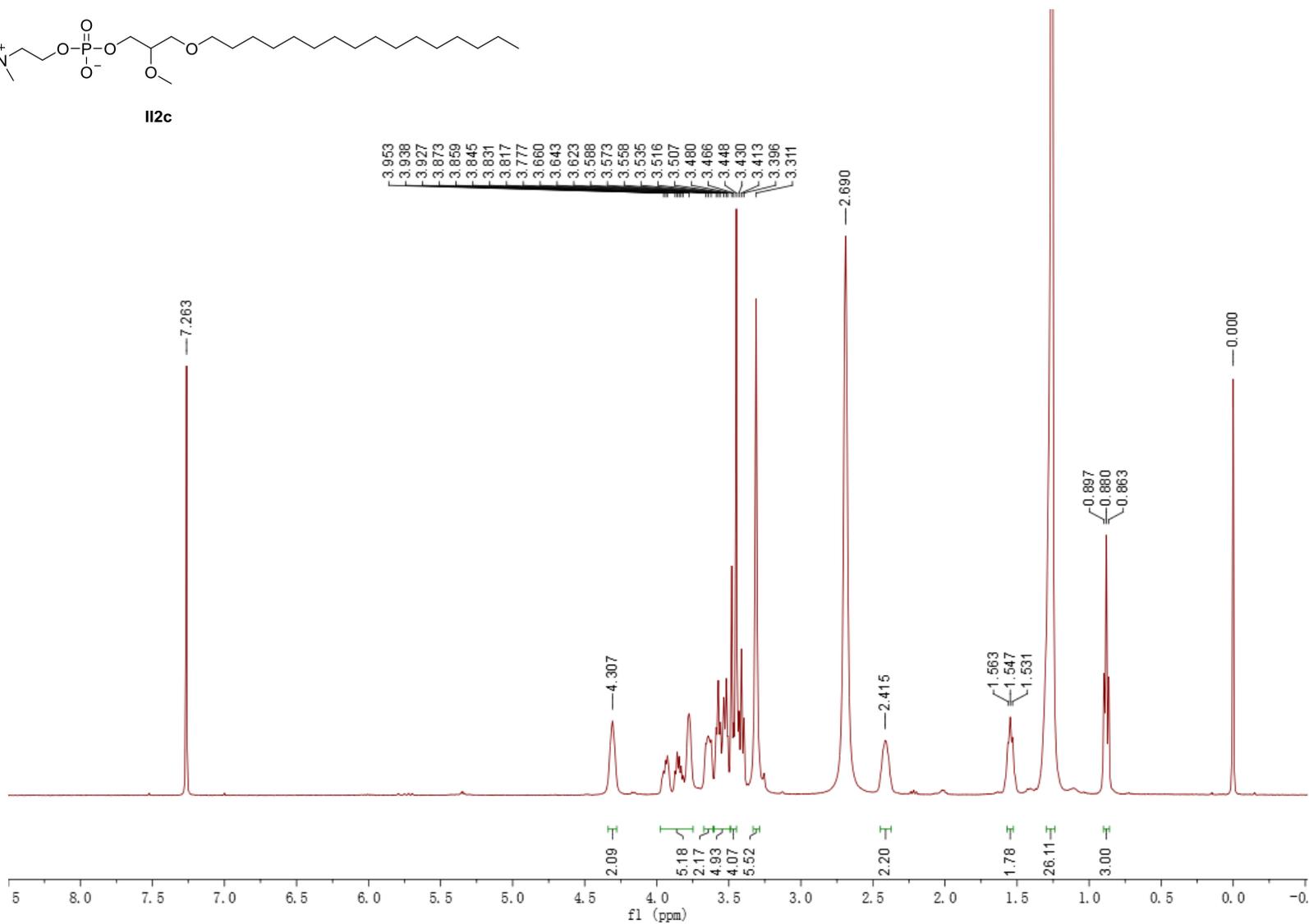


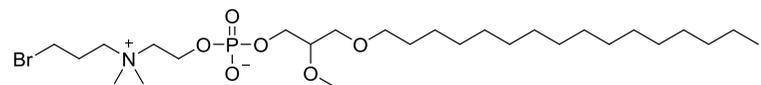
II2b



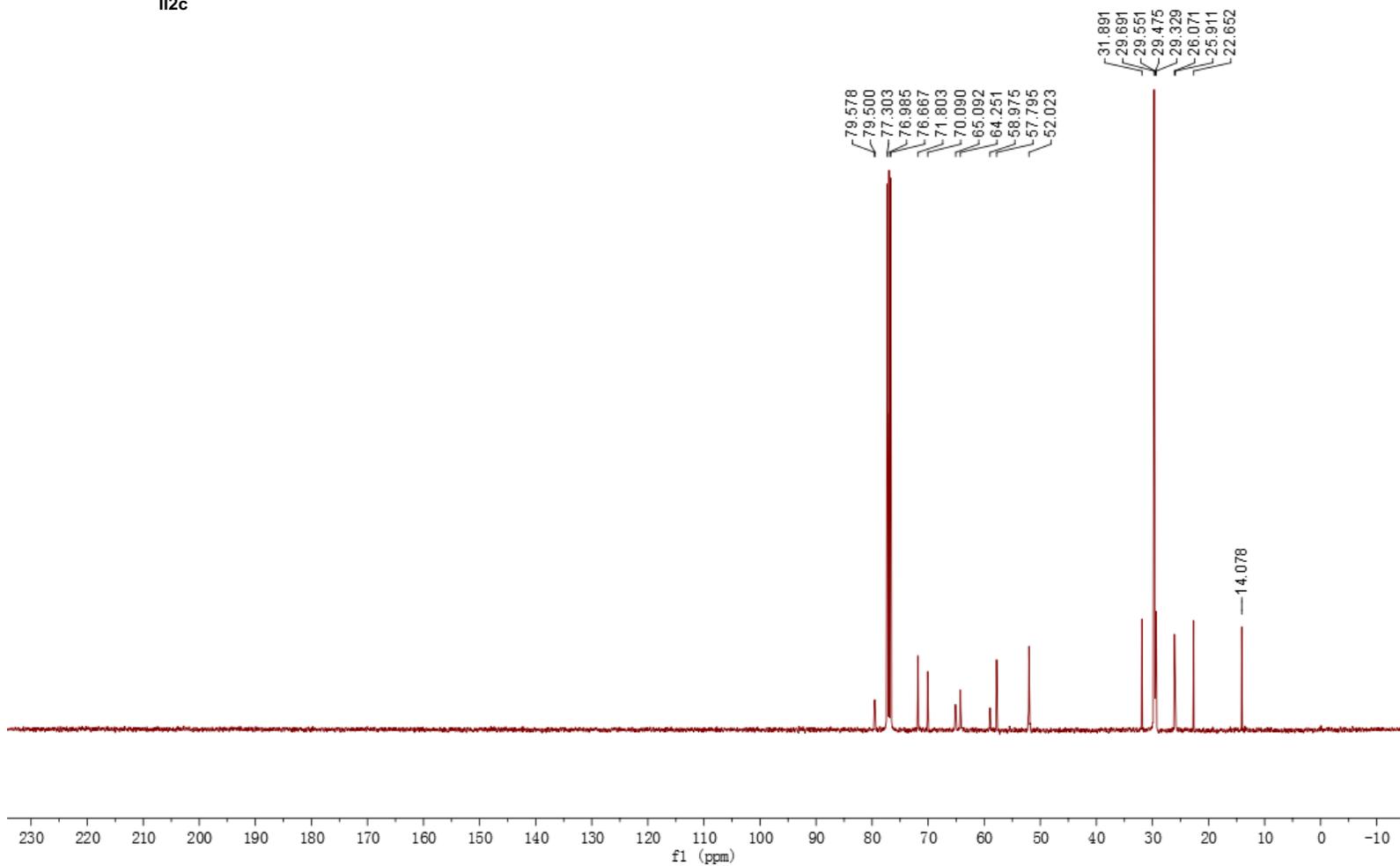


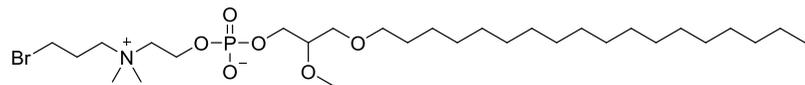
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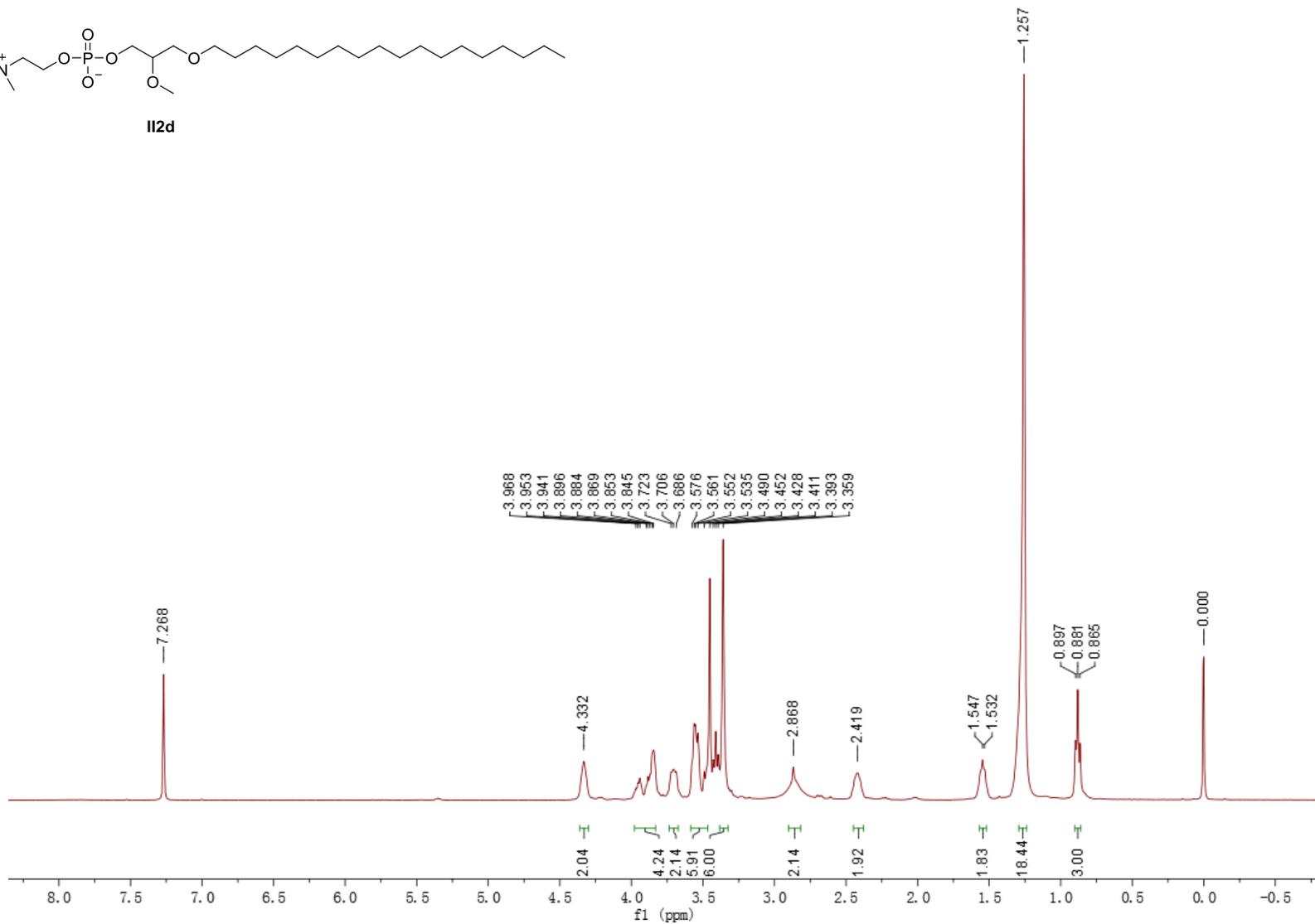


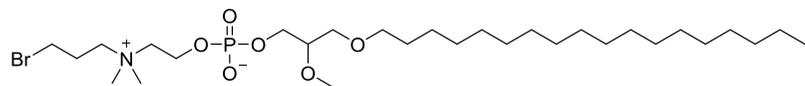
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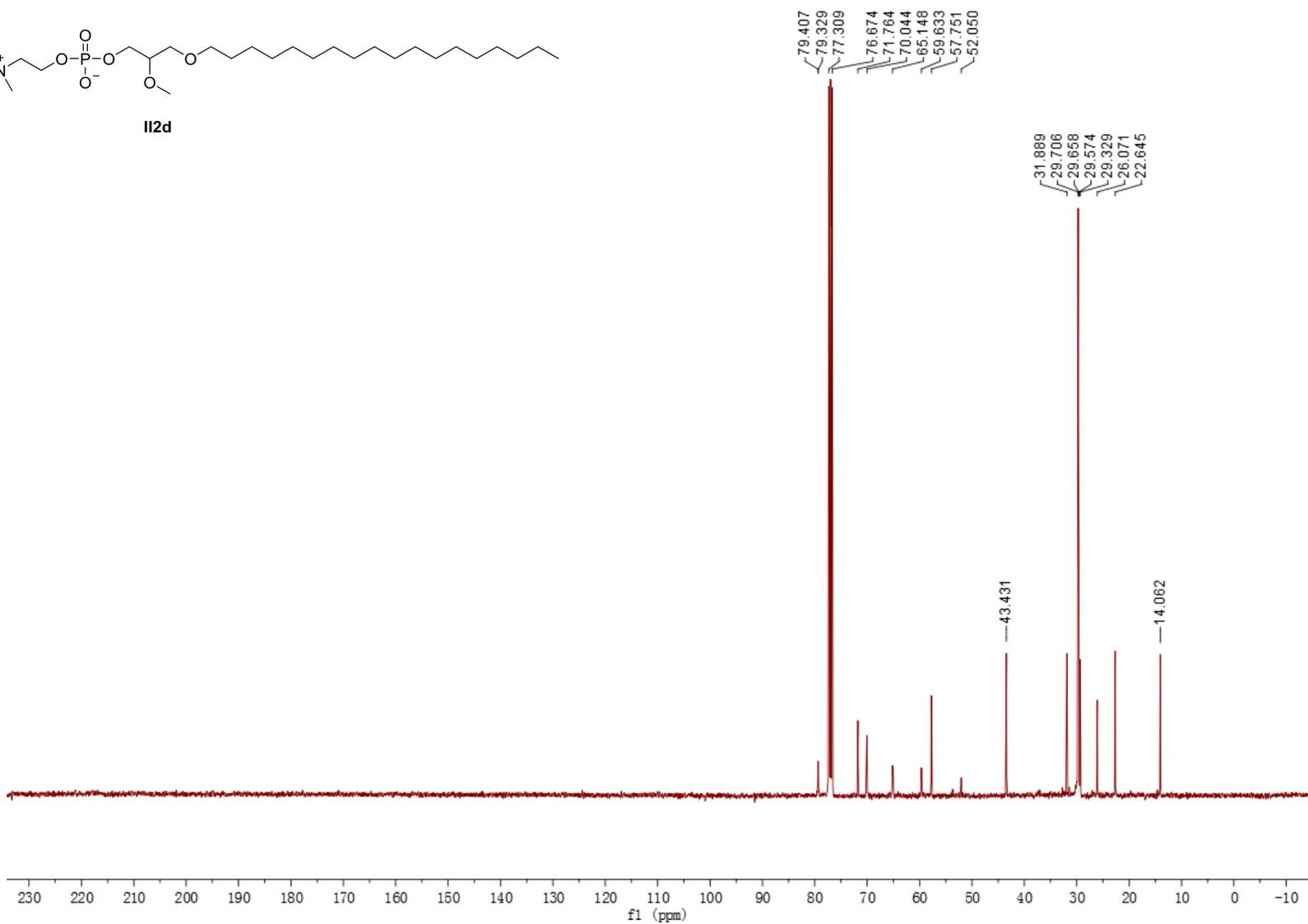


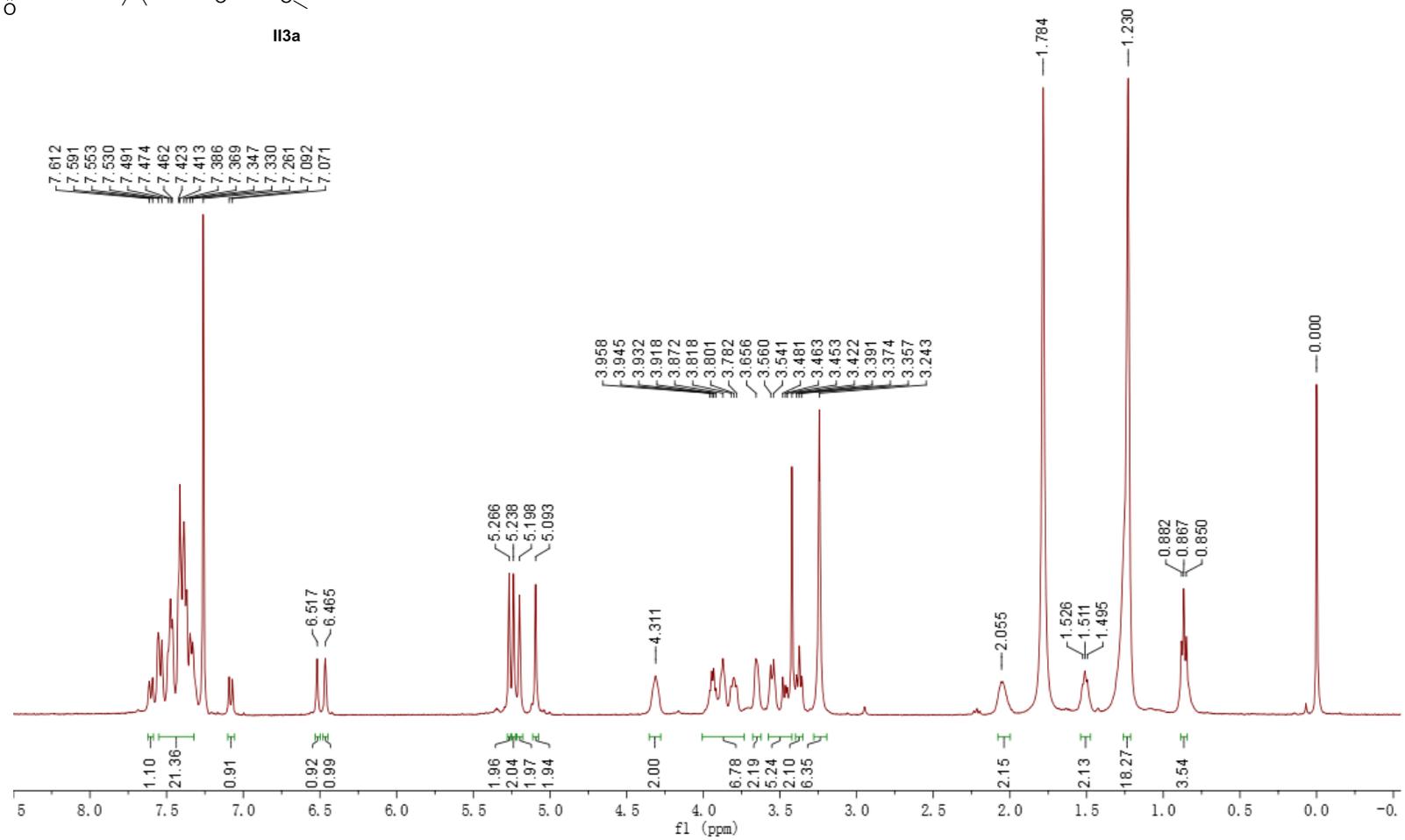
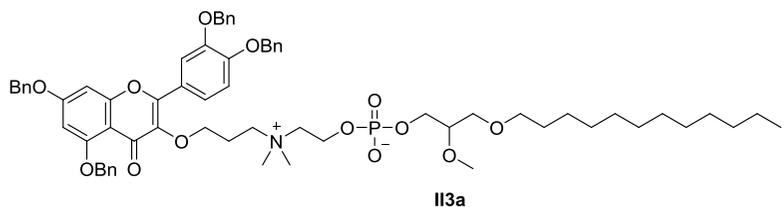
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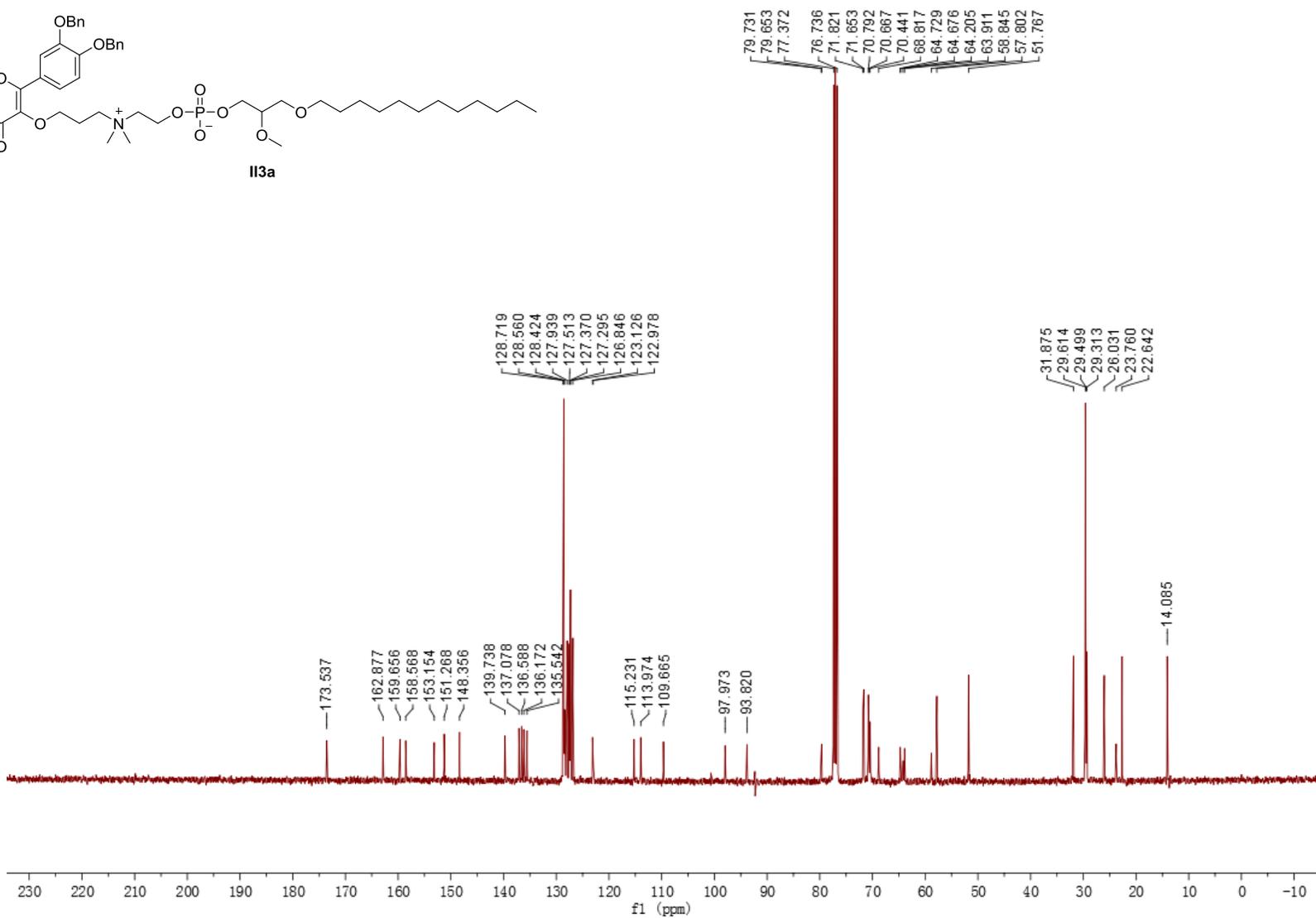
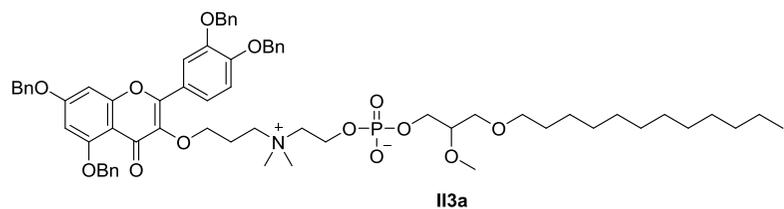


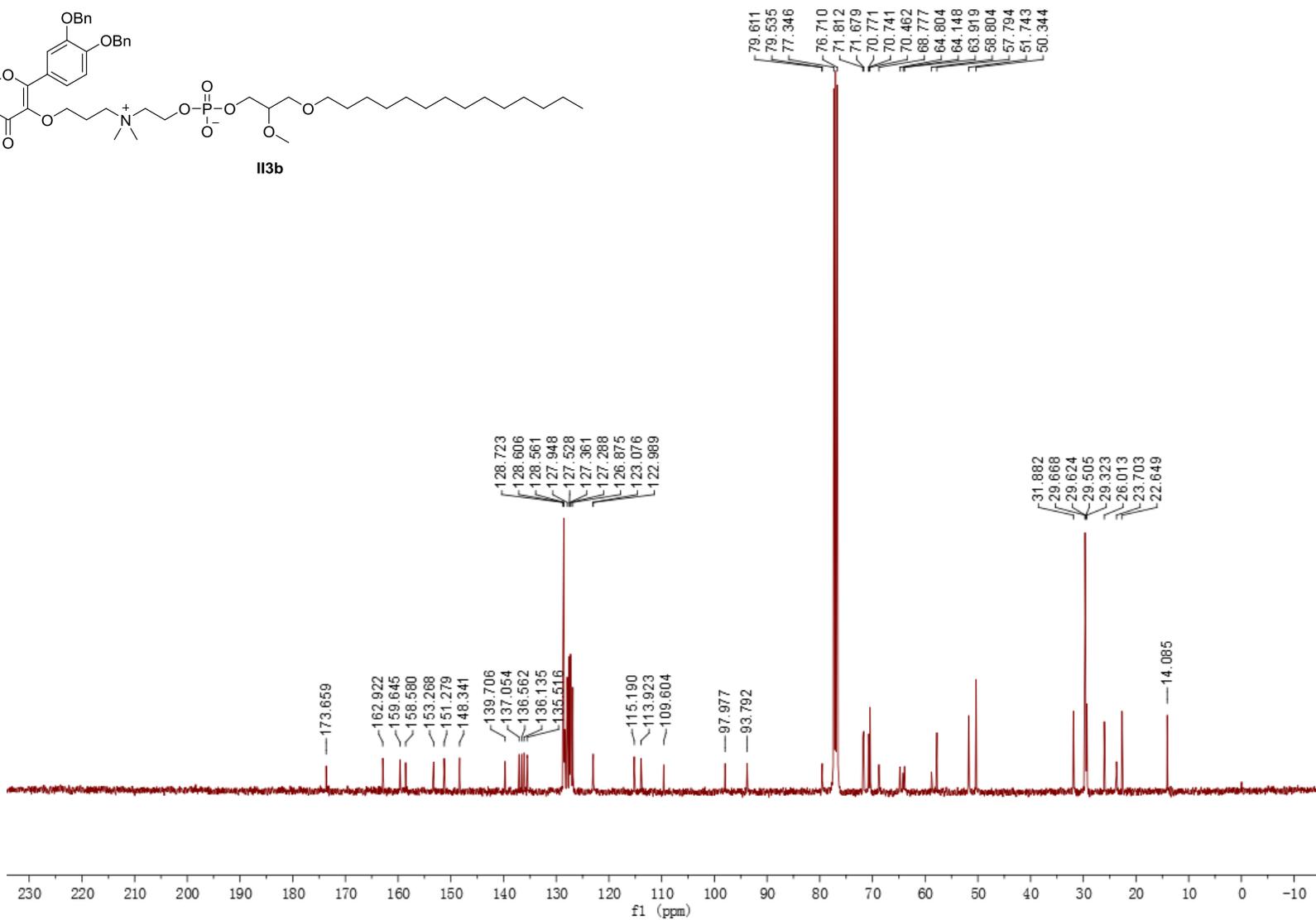
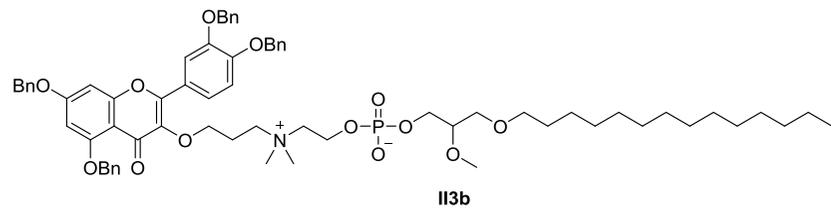


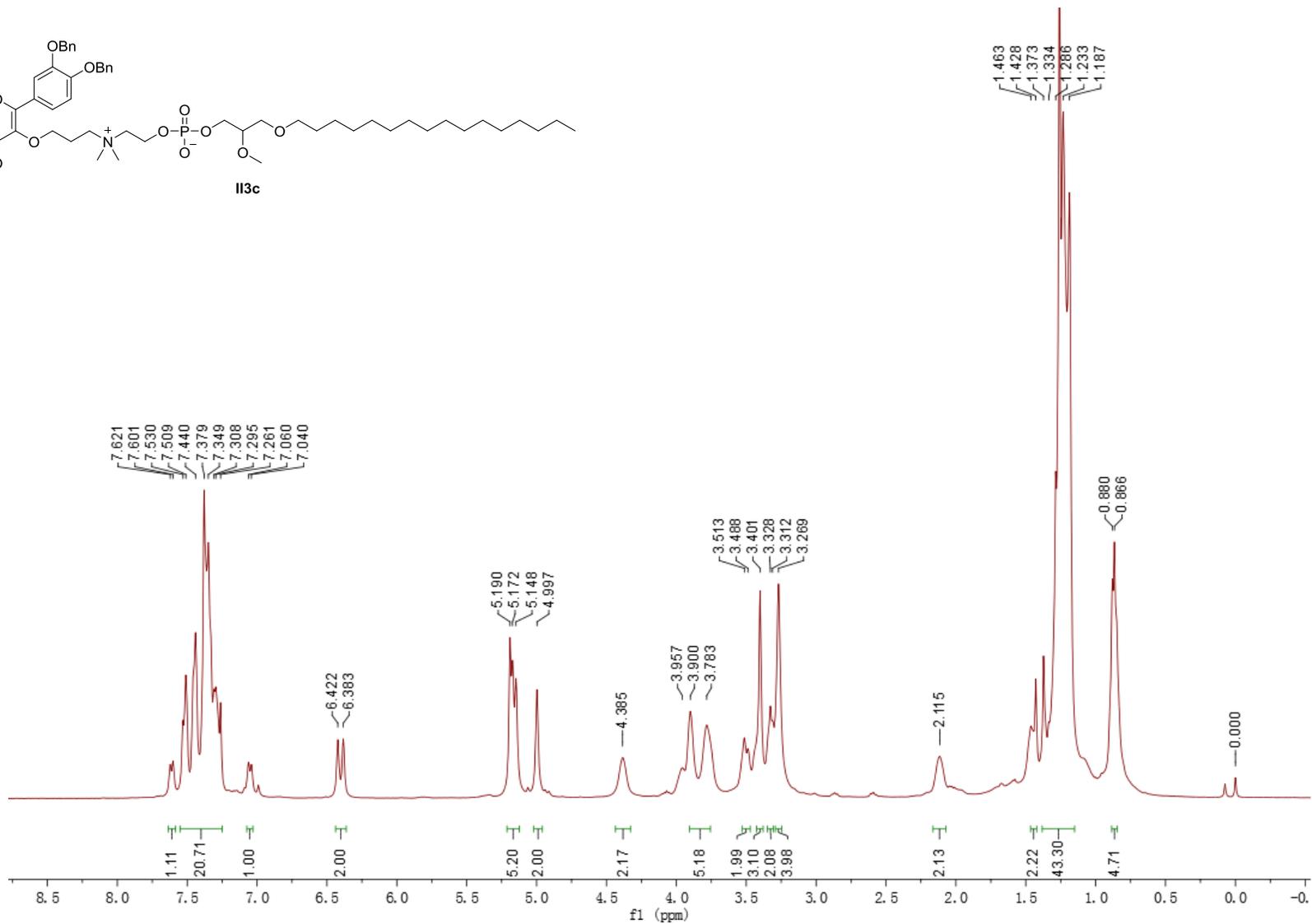
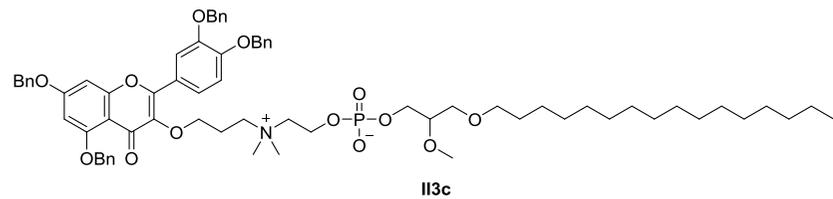
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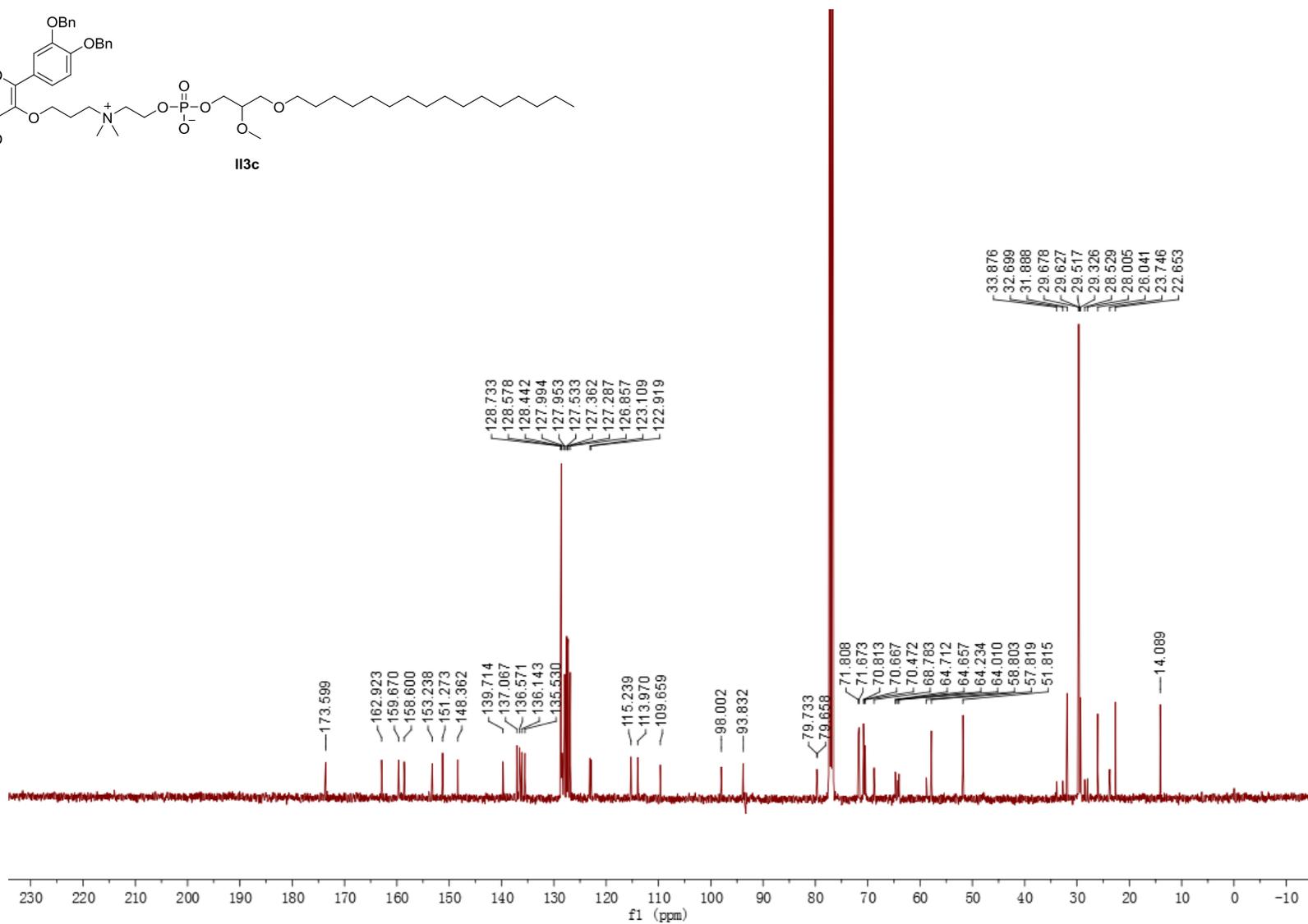
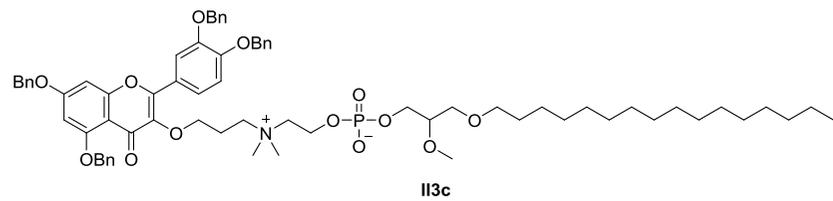


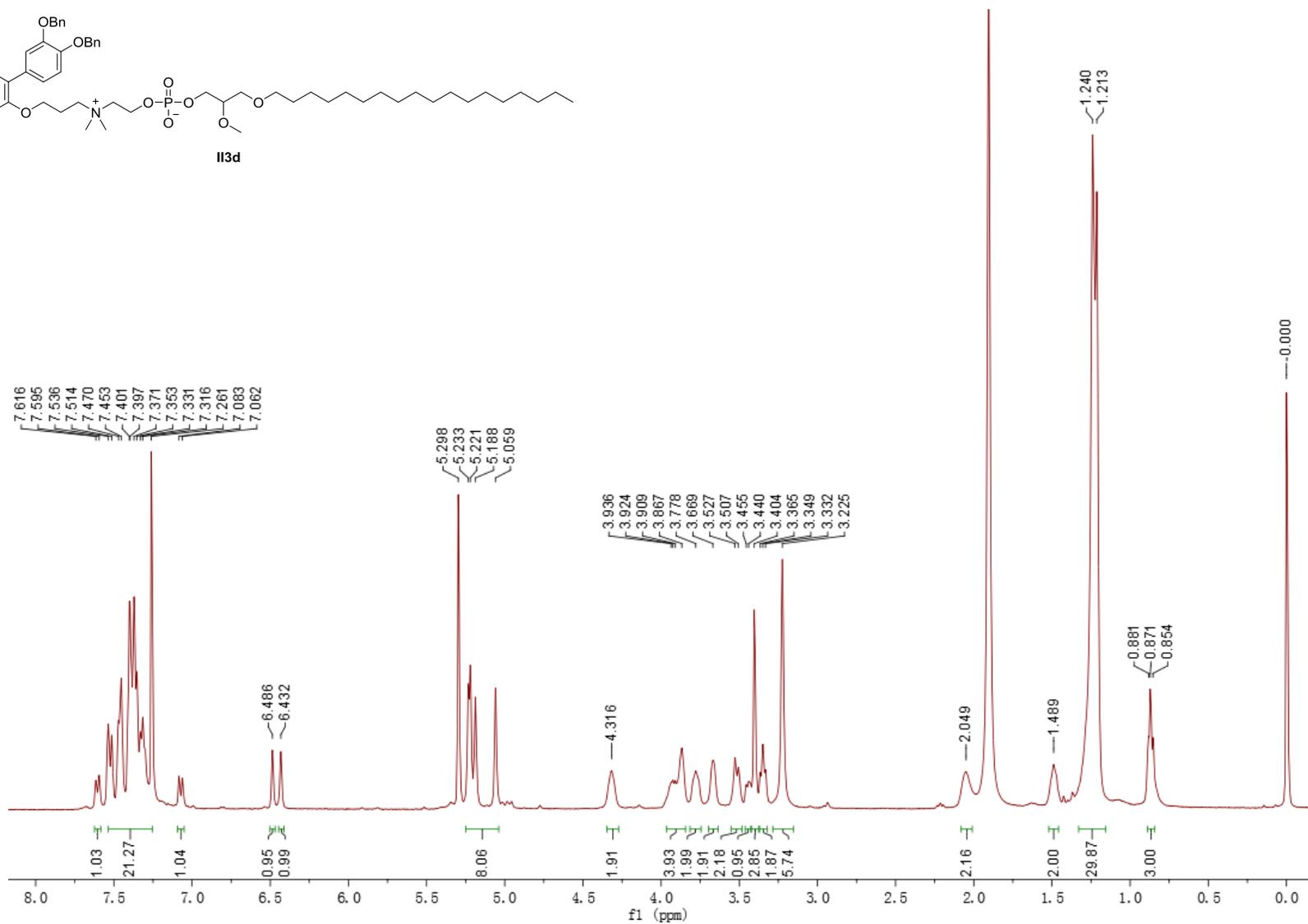
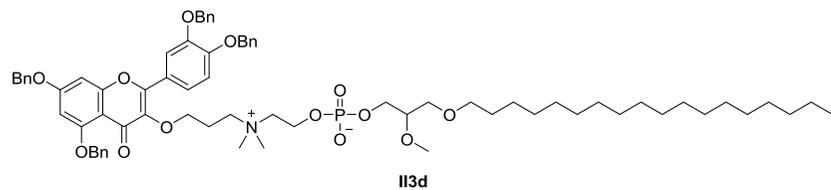


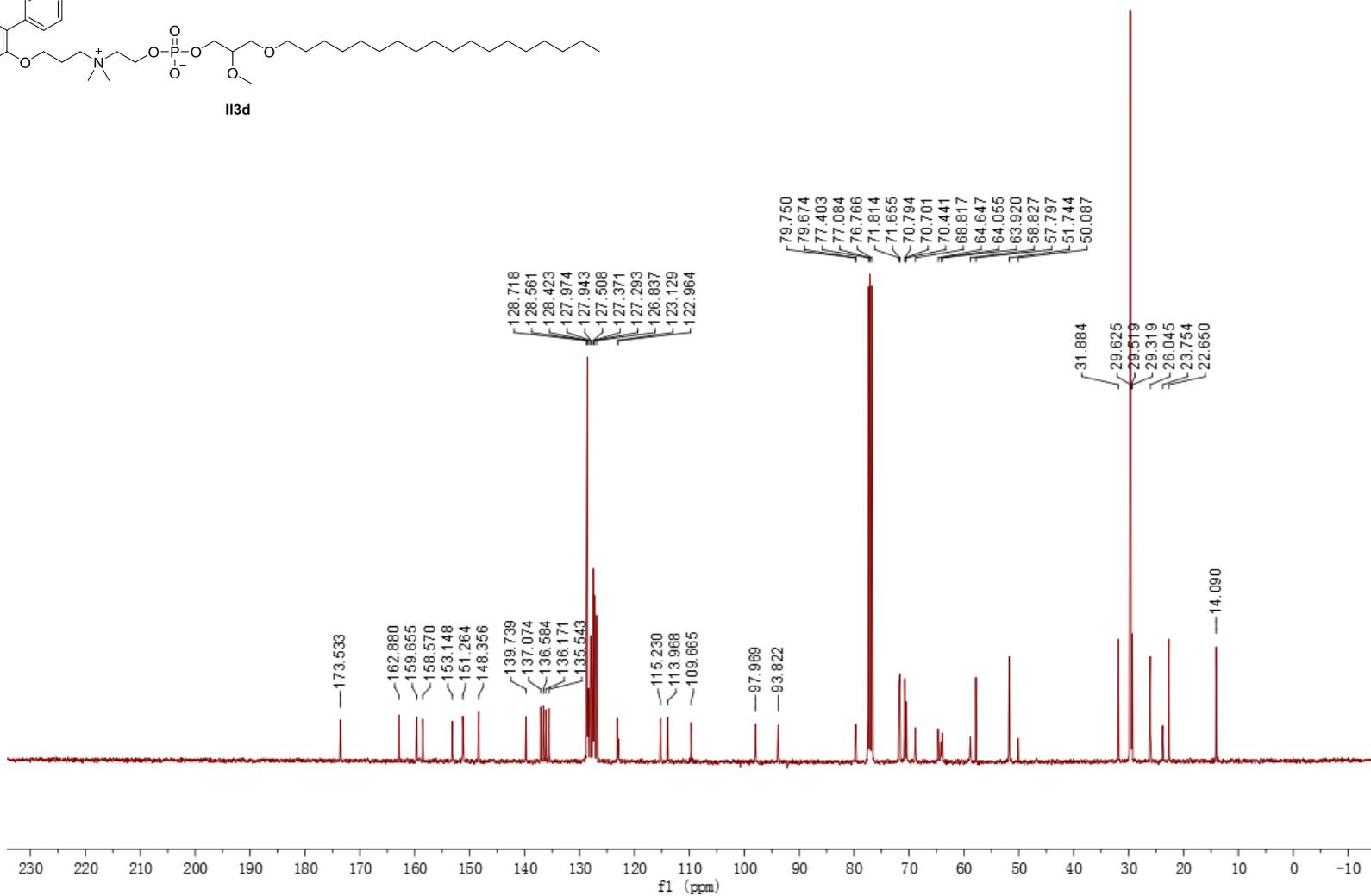
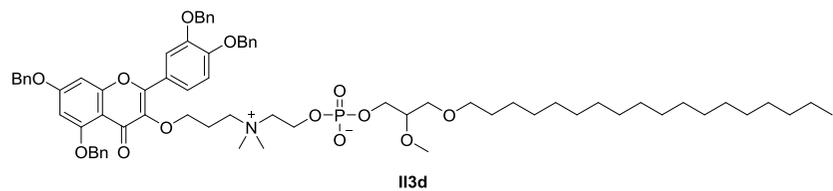


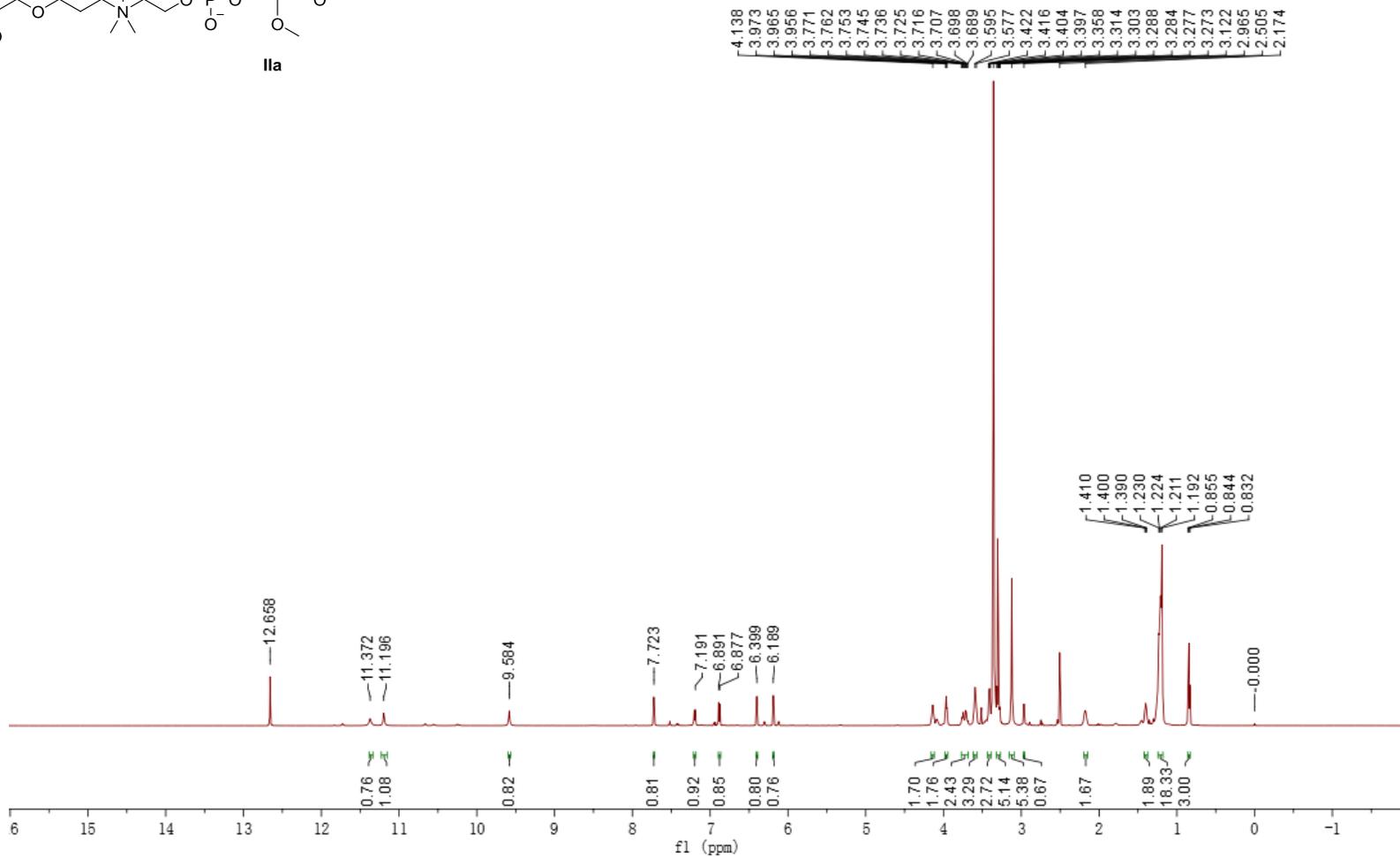
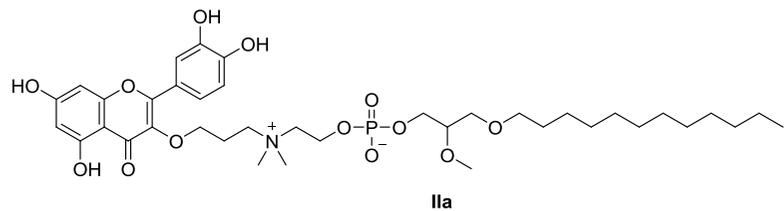


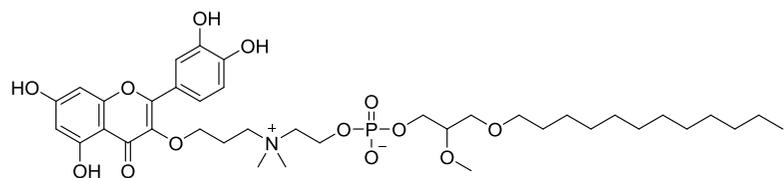




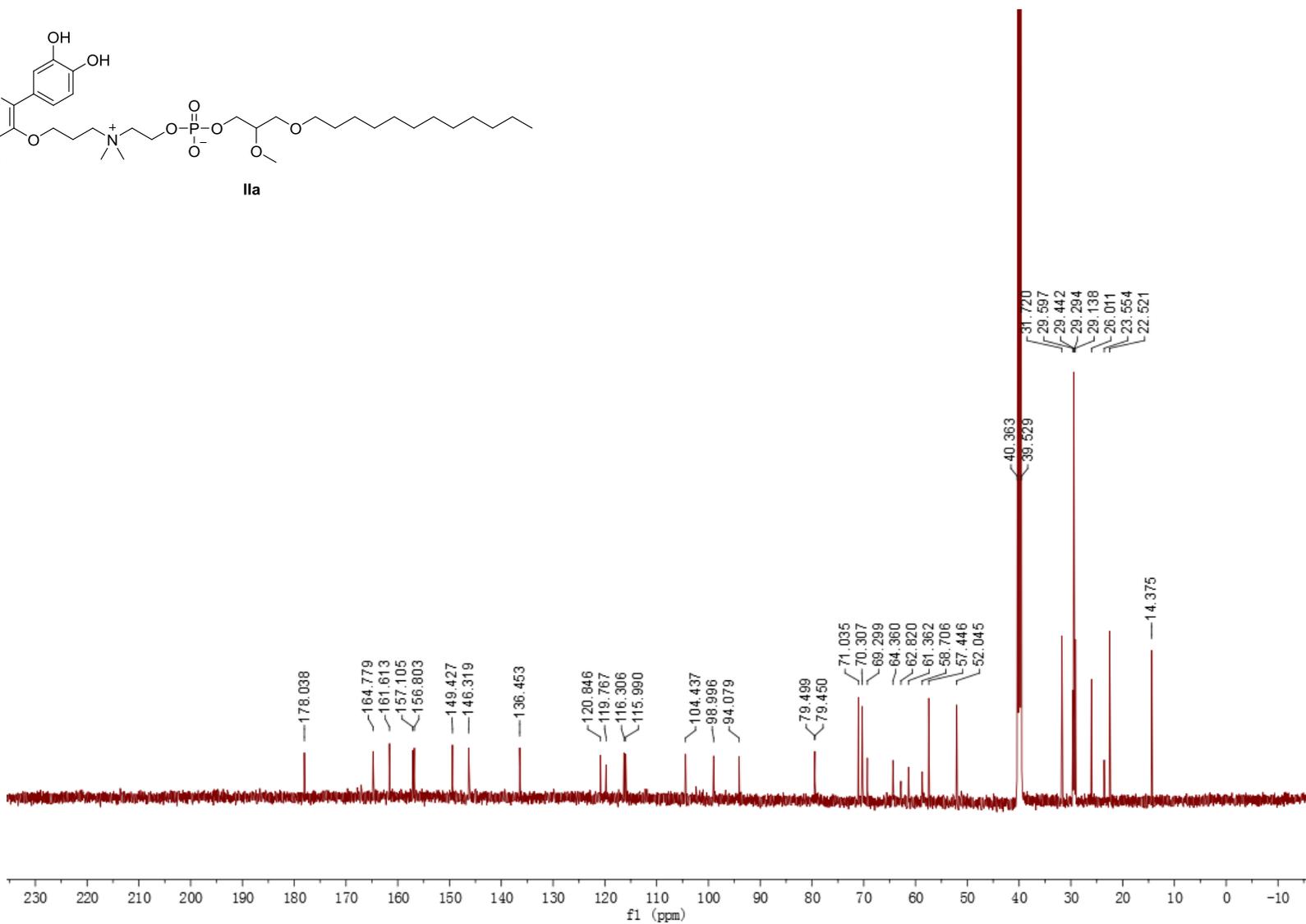


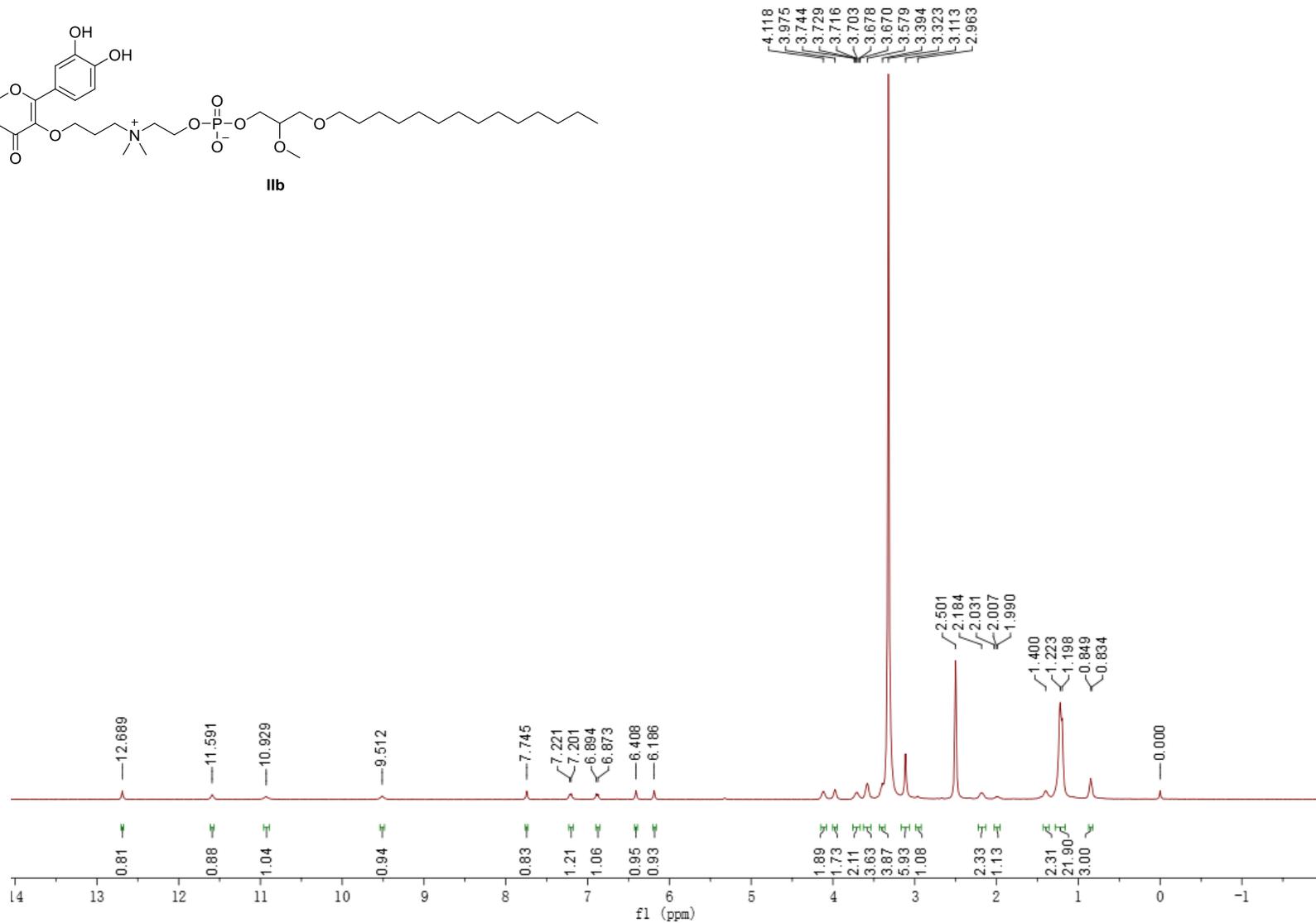
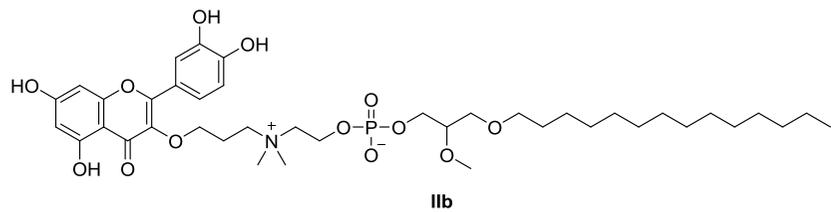


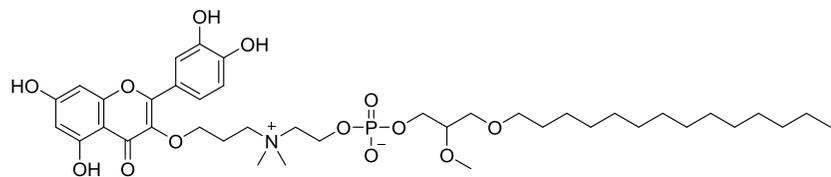




11a







IIb

