Electronic Supplementary Information

Mass production of thin-walled hollow optical fibers enables disposable optofluidic laser immunosensors

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I. Relationship between HOF thickness and fiber drawing speed

Figure S1 Theoretically calculated HOF thickness $\Delta d_2$ versus fiber drawing speed $V_d$. The results were simulated with different outer diameter of HOF, according to the simplified mass conversation law. In the calculations, $OD_2 = 25$ mm, $\Delta d_1 = 1.5$ mm, $V_f = 0.5$ mm/min.
II. Numerical simulation of light distribution

Figure S2 Light intensity along the radial direction of HOF \((x > 0, y = 0)\). In the inset, the evanescent field distribution indicates a penetration depth of 235 nm.
III. The Q-factor of the HOF microcavity

Figure S3 (a) Schematic diagram of the setup for the HOF Q-factor test. (b) The transmission spectrum for the fiber taper-HOF microring resonator system fitted with Lorentz curve. It shows a Q-factor of $1.67 \times 10^6$. 
IV. Experimental setup for disposable OFL immunosensor

Figure S4 (a) The schematic diagram and (b) photo of the experimental setup for disposable OFL immunosensor. The inset of (b) shows details of the V-shaped grooves for replacing the reproducible HOF during the disposable use.
V. The theoretical model and numerical simulation of the sensing mechanism

A simplified theoretical model is established to analyze the sensing mechanism. The mechanism of scattering loss in the microring resonator is the same as that in the Fabry-Perot (FP) cavity,\(^1\) however, the degree of participation for the gain molecules and the resonance mechanism in those two resonators are different.

In the fiber microring resonator, the laser beam would be scattered by the immune complexes through the Beer-Lambert law,\(^2\)

\[
I_r = I_0 e^{-\alpha_L}
\]

(1)

where \(I_r\) and \(I_0\) are the intensity of the residual light after the scattering and incident light, respectively. \(\alpha_L\) is the scattering coefficient (/m). \(L\) is the cavity length. Here we ignore the absorption and reflection by the immune complex and only investigate its scattering effect on the optofluidic laser emission.

According to the resonance mechanism of whispering gallery mode (WGM) microresonators, only the light satisfied the total internal reflection (TIR) and phase matching condition can oscillate in the WGM microresonator. The HOF microring resonator achieves TIR at the HOF-air interface when the incident angle is larger than the critical angle \(\theta_c\) calculated to be \(43.3^\circ\) by \(\theta_c = \arcsin(n_1 / n_2)\), \(n_1 = 1.0\) and \(n_2 = 1.46\) the refractive index (RI) of air and HOF, respectively. That is, the maximum incident angle for light to oscillate in microresonator is about \(46.7^\circ\) (Fig. S5a). We further assume that only the scattering light from \(0^\circ\) to \(46.7^\circ\) can couple into
the microring. According to the Rayleigh scattering theory, the scattering light at an angle of \( \theta \), with respect to the direction of incident beam, is expressed as:

\[
I_\theta = I_0 \frac{8 \pi^4 \varepsilon^2}{\lambda_e^4 r^2} (D + \cos^2 \theta)
\]

\[
I_R = \sum_{\theta=0}^{\theta_i} I_\theta
\]

Here, \( I_\theta \) is the scattering light at the angle of \( \theta \), \( \theta_i=46.7^\circ \), \( \varepsilon \) is the molecular polarizability, \( \lambda_e \) is the optofluidic laser wavelength, and \( r \) is the average distance between the immune complexes. The concentration of immune complexes, \( C_{\text{immuno}} \), can be simplified and denoted by the concentration of protein, \( C_{\text{protein}} \), by \( C_{\text{immuno}} = k t C_{\text{protein}} \). Then the average immune complex distance, \( r \), can be estimated to be

\[
r = \left( \frac{1}{k t N_A C_{\text{protein}}} \right)^{\frac{1}{3}}
\]

Here, \( N_A \) is the Avogadro constant and \( t \) is the immune reaction time. \( k \) is a constant coefficient.

The laser intensity changes with the scattering loss according to\(^4,5\)

\[
I_{\text{laser}} = \frac{\delta I_{\text{sat}}}{2} \left( \frac{\eta g L}{(\alpha_0 + \eta \alpha_e) L + \delta_e} - 1 \right)
\]

Here, we introduce a factor, \( \eta \), as the degree of participation for the gain molecules to participate in lasing. Compared to the conventional FP optofluidic laser, the single-pass gain in the fiber microring resonator is reduced by \( \eta \). \( \delta_e \) and \( g \) are the external coupling factor of the WGM and unsaturated gain coefficient, respectively. \( \alpha_0 = 2 \pi n_2 / \lambda_e Q \) denotes the loss of the WGM cavity with \( \lambda_e \), the emission
wavelength, $Q$ the Q-factor of the HOF microring cavity. $I_{sat} = \frac{hw}{\sigma \tau_{eff}}$ is the saturation intensity with $h$ (6.626×10⁻³⁴ J·s) the Planck constant, $w$ the frequency of pump light, $\sigma$ the fluorophore absorption cross-section, $\tau_{eff}$ the effective recovery time of the transition. The numerical simulation results of temporal evolution and sensing curve for optofluidic laser immunosensor are shown in Figs. S5b and S5c, which can qualitatively describe the trend of optofluidic laser immunosensing and is in agreement with the results in Fig. 4.

Figure S5 (a) The simplified model for disposable OFL immunosensing, in which the dye molecules and immune complex are uniformly distributed in solution within the HOF. $OD_2$ and $\Delta d_2$ are the outer diameter and thickness of HOF, respectively; $r$ is the average distance between immune complexes. (b) Temporal variation of the
simulated OFL intensity at various protein concentrations. (c) Simulated laser intensity $I_{laser}$ versus protein concentration at a fixed reaction time. In the simulation, $I_0 = 1$, $\varepsilon = 13.24 \text{ nm}^3$, $\lambda_L = 600 \text{ nm}$, $L = 480.42 \text{ \mu m}$, $k = 10^{-6}$, $\delta_e = 0.01$, $\sigma = 6 \times 10^{-16} \text{ cm}^2$, $\tau_{eff} = 2.6 \text{ ns}$, $g = 5 \times 10^5 \text{ m}^{-1}$, $n_2 = 1.46$, $\eta = 3.22\%$, and $Q = 1.67 \times 10^6$ are used. This numerical simulation qualitatively reflects the trend for the laser intensity changes with an increasing concentration of protein.

**Reference**


