## Electronic Supplementary Information

In Situ Polymerization and Covalent Functionalisation of Trithiocyanuric acid by MoS<sub>2</sub>

Nanosheets Resulting in Novel Nanozyme with Enhanced Peroxidase Activity, S. Muralikrishna,

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Element	B.E (eV)	FWHM	Area	Atomic%
Mo 3d :	226.667	1.762	6937.2	8.7
Mo 3d :	229.278	0.916	21306.3	26.7
Mo 3d :	230.354	1.541	10536.1	13.2
Mo 3d :	232.445	1.223	20596.0	25.8
Mo 3d :	233.661	1.348	14117.6	17.7
Mo 3d :	236.174	1.806	6324.0	7.9

Table S1: Binding energy and atomic percentage of Mo 3d in  $MoS_2 NS$ 

Table S2: Binding energy and atomic percentage of Mo 3d in  $PTCA-MoS_2 NS$ 

Element	B.E (eV)	FWHM	Area	Atomic%
Mo 3d :	227.059	2.263	20547.2	13.3
Mo 3d :	229.562	1.259	48793.5	31.6
Mo 3d :	230.903	1.259	19207.1	12.4
Mo 3d :	232.704	1.259	27124.2	17.6
Mo 3d :	233.844	1.803	31412.6	20.3
Mo 3d :	236.403	1.616	7434.4	4.8

Table S3: Binding energy and atomic percentage of S 2p in  $MoS_2 NS$ 

Element	B.E	FWHM	Area	Atomic %
	(eV)			
S 2p :	162.145	1.167	14114.2	51.7
S 2p :	163.431	1.116	10683.3	39.1
S 2p :	164.790	1.116	2509.3	9.2

**Table S4**: Binding energy and atomic percentage of S 2p in PTCA-MoS<sub>2</sub> NS

Element	B.E	FWHM	Area	Atomic %
	(eV)			
S 2p :	162.432	1.475	24467.3	40.6
S 2p :	163.967	1.475	26182.5	43.5
S 2p :	165.379	1.475	9576.0	15.9



Figure S1: FT-IR spectrum of PTCA.



Figure S2: SEM image of (a) MoO<sub>3</sub> and (b) PTCA.



Figure S3: ζ-potential of PTCA-MoS<sub>2</sub>-II and MoS<sub>2</sub> recorded in pH 3.5 Na-Ac buffer.



Figure S4: Optical photographic image for peroxidase activity (A)  $MoS_2 NS+TMB+H_2O_2(B) PTCA-MoS_2+TMB+H_2O_2(C) PTCA-MoS_2+TMB$  (D) TMB+H\_2O\_2 (Experimental condition: 1 mL of pH 3.5 Na-Ac buffer containing 40 µg catalyst, 1 mM TMB and 1 mM H\_2O\_2).



**Figure S5**: Absorbance spectrum for mimic peroxidase activity of PTCA (Experimental condition: 250  $\mu$ L of each buffer solution contains 40  $\mu$ g/mL PTCA, 1 mM TMB and 10 mM H<sub>2</sub>O<sub>2</sub>, incubation time 5 min. Inset: Optical photographic image for peroxidase activity with PTCA catalyst).



**Figure S6**: (a) Optical photographic image for  $H_2O_2$  sensing PTCA-MoS<sub>2</sub> as nanozyme catalyst (Experimental condition: 250 µl of pH 3.5 Na-Ac buffer containing 40 µg/mL catalyst, 1 mM TMB with different concentration of  $H_2O_2$ , incubation time 5 min at room temperature)



**Figure S6**: (b) Optical photographic image for  $H_2O_2$  sensing  $MoS_2$  NS as nanozyme catalyst (Experimental condition: 250 µl of pH 3.5 Na-Ac buffer containing 40 µg/mL catalyst, 1 mM TMB with different concentration of  $H_2O_2$ , incubation time 5 min at room temperature)



**Figure S7** (a) Optical photographic image for glucose sensing using PTCA-MoS<sub>2</sub> as nanozyme catalyst (Experimental condition: 250  $\mu$ l of pH 3.5 Na-Ac buffer containing 40  $\mu$ g/mL catalyst, 1 mM TMB with different concentration of glucose in the presence of glucose oxidase, incubation time 15 min at room temperature).



**Figure S7** (b) Optical photographic image for glucose sensing using  $MoS_2 NS$  as nanozyme catalyst (Experimental condition: 250 µl of pH 3.5 Na-Ac buffer containing 40 µg/mL catalyst, 1 mM TMB with different concentration of glucose in the presence of glucose oxidase, incubation time 15 min at room temperature).



Figure S8 SEM image of PTCA-MoS<sub>2</sub>-II after six months.