## Engineering Protein Polymers of Ultrahigh Molecular Weight via Supramolecular Polymerization: Towards Mimicking the Giant Muscle Protein Titin

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## **Electronic Supplementary Information**

**Table S1.** Protein amino acid sequences. Residues colored in red correspond to cysteine mutations in GL5CC as well as W34F mutation in I27. Sequences highlighted in grey are generated from the ligation of BamHI and BgIII sticky ends.

| Name                               | Protein sequence   |
|------------------------------------|--|
| I27-G <sub>N</sub>                 | LIEVEKPLYGVEVFVGETAHFEIELSEPDVHGQFKLKGQPLAASPDCEIIE<br>DGKKHILILHNCQLGMTGEVSFQAANTKSAANLKVKEL RS MDTYKLIL<br>127   |
|                                    | NGKTLKGETTTEAVDAATAEKVFKQYANDNGVGCGLG  |
|                                    | G <sub>N</sub> (GL5CC 1-42)  |
| G <sub>C</sub> -127                | CGDGEWTYDDATKTFTVTE RS <i>LIEVEKPLYGVEVFVGETAHFEIELSE</i><br>G <sub>C</sub> (GL5CC 43-61)<br><i>PDVHGQFKLKGQPLAASPDCEIIEDGKKHILILHNCQLGMTGEVSFQAA</i><br><i>NTKSAANLKVKEL</i><br><i>127</i>        |
| G <sub>C</sub> -I27-G <sub>N</sub> | CGDGEWTYDDATKTFTVTE RS <i>LIEVEKPLYGVEVFVGETAHFEIELSE</i><br><i>PDVHGQFKLKGQPLAASPDCEIIEDGKKHILILHNCQLGMTGEVSFQAA</i><br><i>NTKSAANLKVKEL</i> RS MDTYKLILNGKTLKGETTTEAVDAATAEKVF<br>KQYANDNGVGCGLG |



**Figure S1.** Placing GN at the N-terminus and GC at the C-terminus of two interacting protein partners can lead to steric hindrance when GN and GC undergo protein fragment reconstitution.



**Figure S2.** Stopped-Flow fluorimetry of protein fragment association and reconstitution complex denaturation. A) Association kinetics of mixing equal I27-G<sub>N</sub> and G<sub>C</sub>-I27 solution and adjusting the final concentration to  $2 \times 10^{-5}$  M was shown in red. The rate constant was  $1.29 \times 10^{3} \pm 311$  M<sup>-1</sup> s<sup>-1</sup> fitting to the second-order rate law. B) The blue curve was unfolding kinetics curve of I27-(G<sub>N</sub>-G<sub>C</sub>)-I27 complex at  $2 \times 10^{-5}$  M treated with 4 M GdmCl and the single-exponential fitting gave a rate constant of  $26.4 \pm 0.125$  s<sup>-1</sup>. In both experiments, G<sub>C</sub>-I27 curves colored gray had no obvious change.



**Figure S3.** Native-PAGE of  $G_C$ -I27- $G_N$  polymers reduced by DTT at 4 °C. Lane 1 was standard protein marker whose bands altered position in Native-PAGE; lane 2 showed the native state  $G_C$ -I27- $G_N$  can still polymerize without disulfide bonds.



**Figure S4.** Plot of diffusion coefficient versus polyprotein molecular weight. In DLS experiments, we used Mark-Houwink-Sakurada (MHS) equation to measure the molecular weight:  $D = K \cdot Mw'$  where K and ! are constants and depend on the macromolecule, solvent and temperature, D is the diffusion coefficient. Because the G<sub>C</sub>-127-G<sub>N</sub>-based protein polymers are not globular proteins, we used purified polyproteins (GR)<sub>4</sub>, (NuG2)<sub>8</sub>, (GR)<sub>8</sub>, (GR)<sub>12</sub> and (GR)<sub>16</sub>, which showed a single band on SDS-PAGE (corresponding to a polydispersity index of 1) and have known molecular weights, to calculate K and  $\alpha$  for such protein polymers. DLS experiments were carried out at 25 °C and all proteins were dissolved in water. Fitting the MHS equation to the experimental data yielded a K = 9.71x10<sup>-5</sup>, ! = -0.51. Using this method, we determined the diffusion-averaged molecular weight of the G<sub>C</sub>-127-G<sub>N</sub>-based protein polymers Mw=458±16 kDa. All measurements were done in triplicate.

![](_page_3_Figure_0.jpeg)

**Figure S5.** Competitive effect between  $G_C$ -I27- $G_N$  and  $G_C$ -I27. The amount of higher order of  $G_C$ -I27- $G_N$  polymers decreased with the increasing of  $G_C$ -I27 concentrations due to the competition of the monofunctional monomer. Lane 1: protein molecular weight marker; lane 2: oxidized  $G_C$ -I27- $G_N$  protein polymers; lane 3-5: oxidized  $G_C$ -I27- $G_N$  and  $G_C$ -I27 mixture samples, and the ratio of  $G_C$ -I27- $G_N$ : $G_C$ -I27 is 5:1, 2:1, 1:1, respectively.

![](_page_4_Figure_0.jpeg)

Figure S6. The effect of monomer concentration on the molecular weight of the engineered protein polymers. A) Non-reducing 12% SDS-PAGE analysis of the engineered protein polymers. Monomer concentration is indicated in each lane. B-C) SE-FPLC elution time plot (B) and molecular weight distribution (C) of polymerized  $G_C$ -I27- $G_N$ -based protein polymers.

| Table   | <b>S2.</b> | The    | effect              | of | monomer     | concent   | ration | on | the | molecular | weight | and | molecular | weight |
|---------|------------|--------|---------------------|----|-------------|-----------|--------|----|-----|-----------|--------|-----|-----------|--------|
| distrib | utio       | n of ( | G <sub>C</sub> -I27 | -G | N-based pro | otein pol | ymers  |    |     |           |        |     |           |        |

| Protein conc. (mM) | Mn (kDa) | Mw (kDa) | PDI  |
|--------------------|----------|----------|------|
| 0.5                | 441      | 513      | 1.16 |
| 0.2                | 447      | 505      | 1.13 |
| 0.1                | 464      | 496      | 1.07 |

![](_page_5_Picture_0.jpeg)

**Figure S7**. A photograph of a native PAGE of  $G_C$ -I27- $G_N$ -based protein polymers and depolymerized product after being incubated under reducing condition at 37 °C for 30 minutes.

![](_page_6_Figure_0.jpeg)

**Figure S8.** Protein hydrogels made of  $G_{C}$ -I27- $G_{N}$  (2%) can undergo gel-sol transition under a reducing condition at elevated temperatures. Photographs show the protein hydrogels under different conditions. The yellowish solid are the hydrogel samples.