

Electronic supplementary information (ESI)

**One-Phase Synthesis of Small Gold Nanoparticles Coated by a Horizontal
Porphyrin Monolayer**

Junya Ohyama,^a Yutaka Hitomi,^{*ab} Yasuhiro Higuchi,^a Masashi Shinagawa,^a Hidefumi
Mukai,^a Masahito Kodera,^b Kentaro Teramura,^c Tetsuya Shishido,^a Tsunehiro Tanaka^a

a Department of Molecular Engineering, Graduate School of Engineering, Kyoto
University, Kyoto 615-8510, Japan. Fax: +81 75 383 2561; Tel: +81 75 383 2562

b Department of Molecular Chemistry and Biochemistry, Doshisha University,
Kyotanabe, Kyoto 610-0321, Japan. Tel: +81 774 65 7437; E-mail:
yhitomi@mail.doshisha.ac.jp

c Kyoto University Pioneering Research Unit for Next Generation, Kyoto University,
Kyoto 615-8510, Japan.

General methods. Electronic absorption spectra were measured on a HITACHI U-3500 UV-VIS-NIR spectrometer by using a quartz cell with a 1-cm path length. ^1H , ^{13}C , 2D (HMQC, HMBC) NMR spectra were measured in CDCl_3 on a JEOL JNM-ECX400 spectrometer. Transmission electron microscopy (TEM) images were taken with a JEOL JEM-100SX operating at an accelerating voltage of 100 kV. TEM samples were prepared by depositing of drops of a methanol solution onto a carbon-coated copper grid (Okenshoji Co. LTD.) and dried to the solvent at room temperature. X-ray photoelectron spectra (XPS) were acquired using an ULVAC PHI 5500MT. XPS samples were mounted on an indium foil and the spectra were measured using $\text{Mg } K_{\alpha}$ radiation (15 kV, 400 W) in a chamber with the base pressure of ca. 1×10^{-8} Torr. The take-off angle was set at 45° . All binding energies were corrected for charge shifting by referencing to the C(1s) line from the adventitious carbon at 284.6 eV.

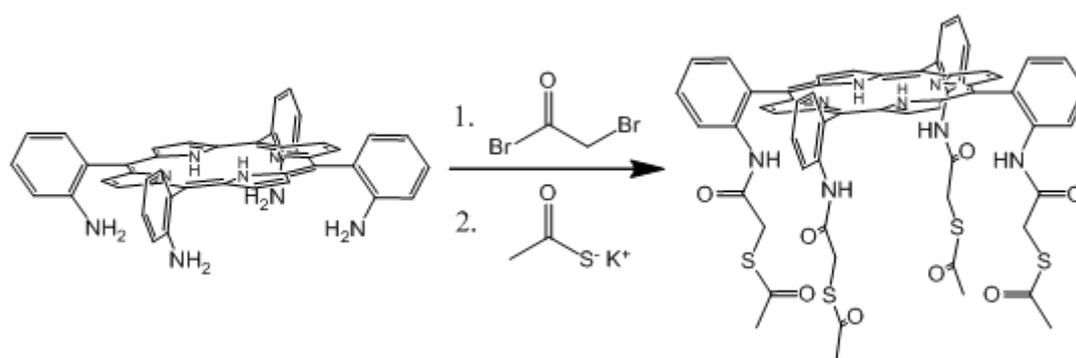
Materials. All solvents were purchased from Wako Pure Chemical Industry.

2-Nitrobenzaldehyde, pyrrole, and potassium thioacetate were purchased from Tokyo Chemical Industry, and the other chemicals were from Wako Pure Chemical Industry.

5,10,15,20-Tetrakis(*o*-nitrophenyl)-porphyrin and 5,10,15,20-tetrakis(*o*-aminophenyl)-

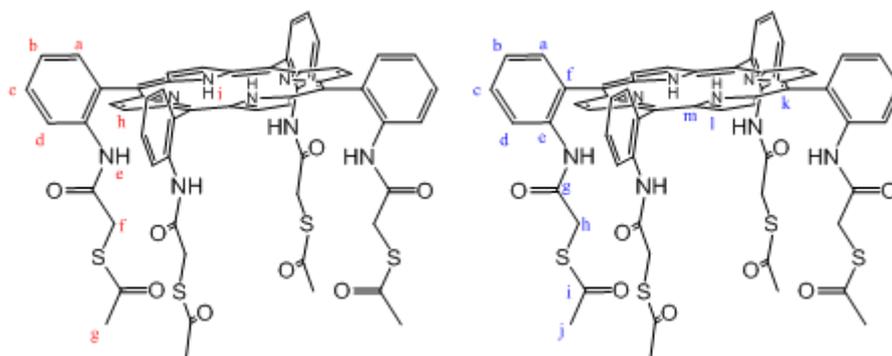
porphyrin were synthesized according to the literature,¹ and the latter was atropisomerized to the $\alpha,\alpha,\alpha,\alpha$ -atropisomer according to the Lindsey's procedure.²

$\alpha,\alpha,\alpha,\alpha$ -5,10,15,20-tetrakis(2-(S-acetyl-thioacetamido)phenyl)porphyrin (1).



To a solution of $\alpha,\alpha,\alpha,\alpha$ -5,10,15,20-tetrakis(2-aminophenyl) porphyrin (0.498 g, 0.738 mmol) in dry THF (80 mL) was diethylaniline (1.6 mL, 10.3 mmol) added under N_2 . Bromoacetyl bromide (641 μ L, 7.34 mmol) in CH_2Cl_2 (11 mL) was added to the mixture dropwise over 5 min at $0^\circ C$ and stirred for 1 h at $0^\circ C$. Then, potassium thioacetate (1.7 g, 14.9 mmol) in dry EtOH (80 mL) was added to the mixture and stirred for 2 h. The solvent was removed under reduced pressure. The residue was purified by column chromatography on silica gel eluted with $CHCl_3$ and subsequent recrystallization from $CHCl_3$ -MeOH. 0.240 g (0.211 mmol, 29%). TLC R_f = 0.62 ($CHCl_3$:MeOH 10:1); HRMS-FAB (m/z): $[M+H]^+$ calcd for $C_{60}H_{50}O_8N_8S_4$, 1138.26; found, 1138.2642; 1H NMR ($CDCl_3$, 400 MHz): δ 8.78 (s, 8H, H_b), 8.57 (d, 4H, H_a), 7.99 (d, 4H, H_d), 7.85

(dd, 4H, H_c), 7.64 (s, 4H, H_e), 7.54 (dd, 4H, H_b), 3.56 (s, 8H, H_f), 0.75 (s, 12H, H_g),
-2.69 (s, 2H, H_i); $^{13}\text{C-NMR}$ (CDCl_3 , 100 MHz): δ 194.1 (C_i), 165.9 (C_g), 138.2 (C_e),
134.6 (C_d), 131.8 (C_f), 131.4 (C_l), 129.9 (C_c), 123.6 (C_b), 121.8 (C_a), 114.8 (C_k), 33.2
(C_h), 28.7 (C_j) (C_m could not be observed, presumably due to N-H tautomerism³;
UV-Vis λ_{max} in DMF: 422, 516, 550, 591, 647 nm.



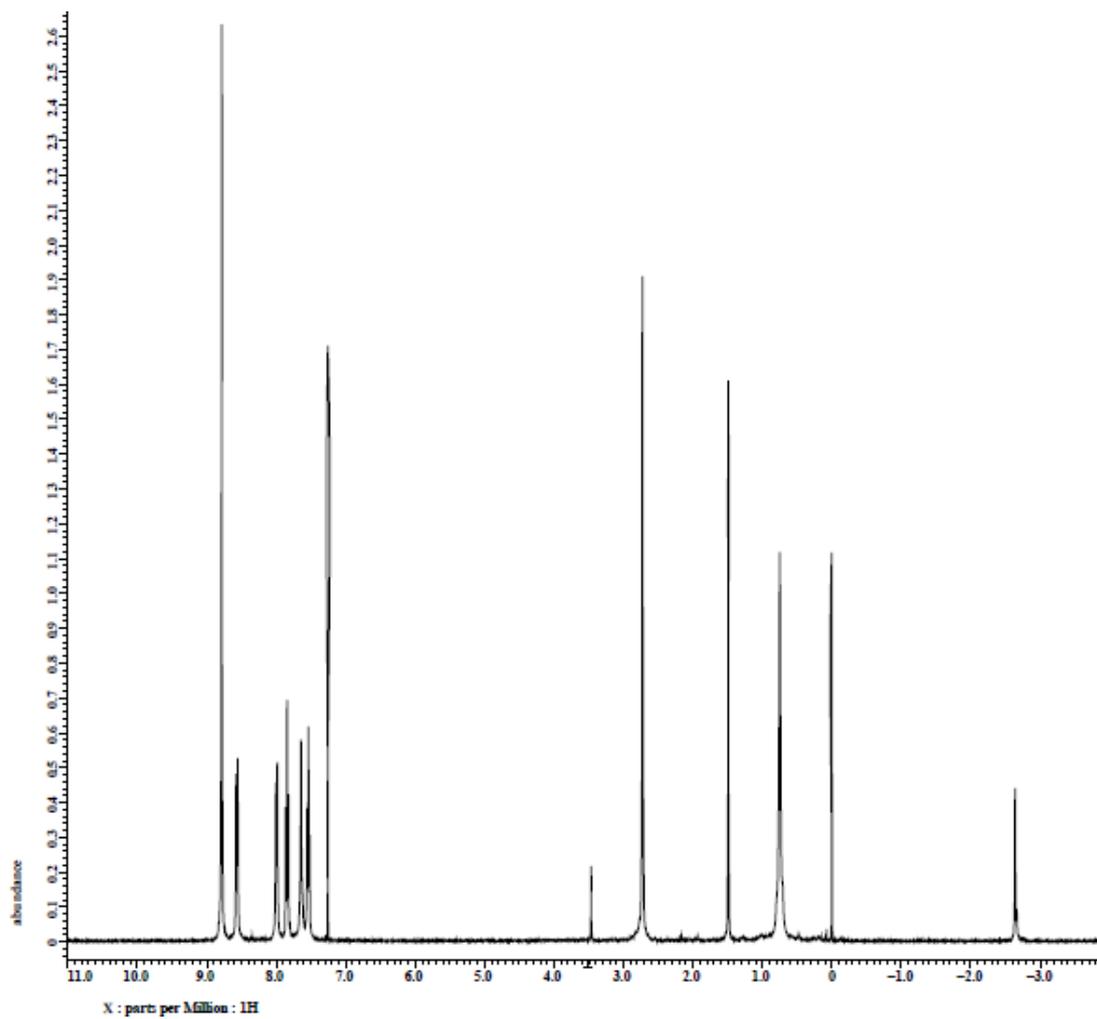


Fig. S1 ¹H-NMR chart of **1**.

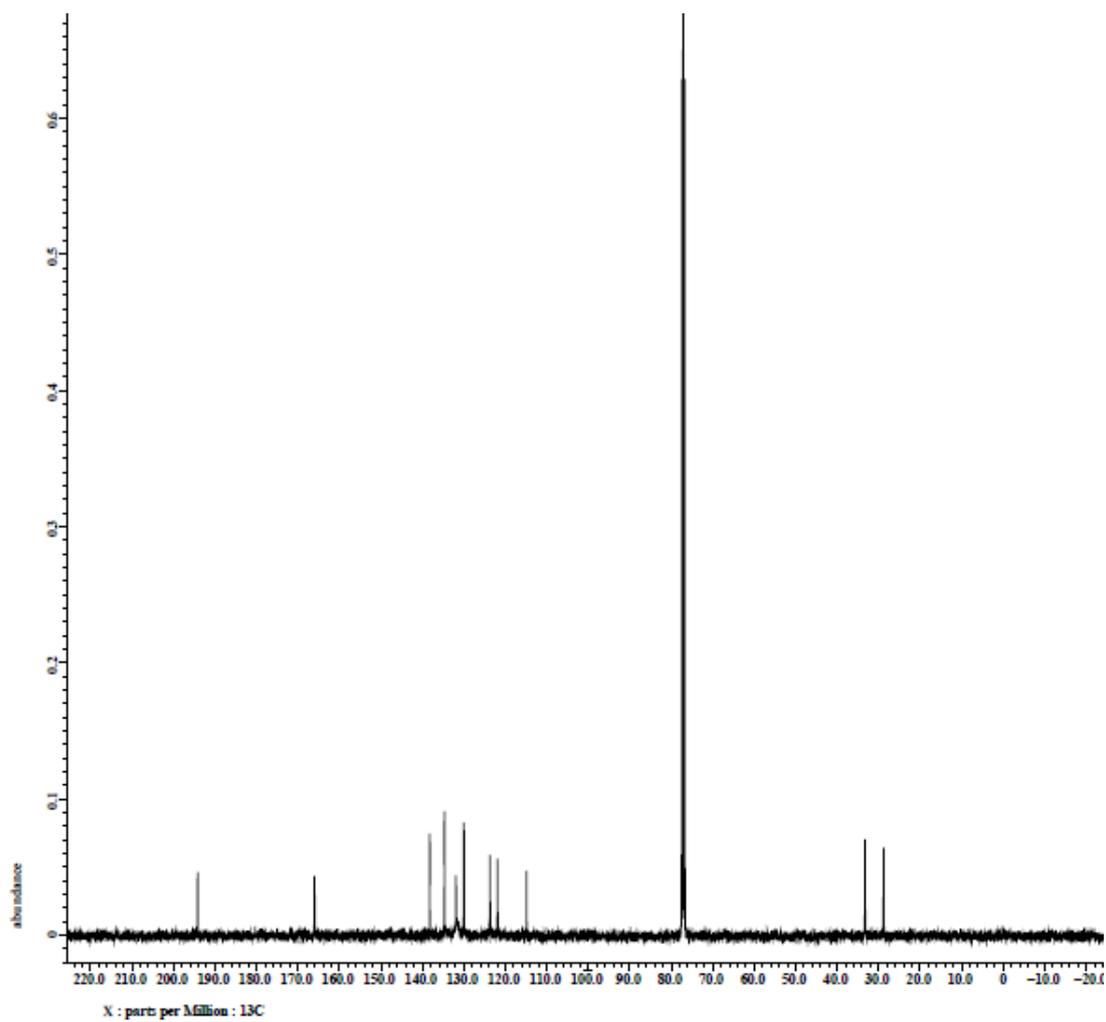


Fig. S2 ^{13}C -NMR chart of **1**.

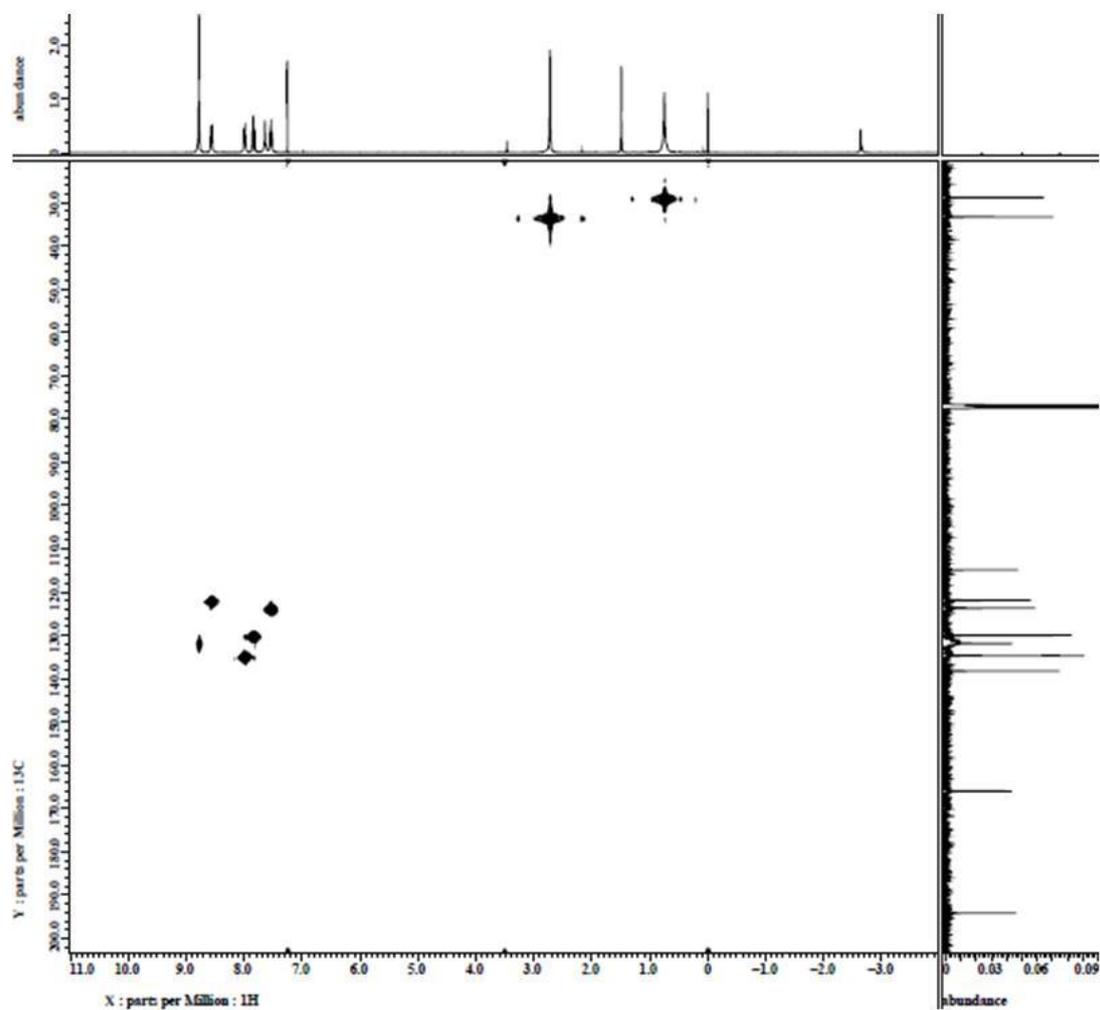


Fig. S3 HMQC chart of **1**.

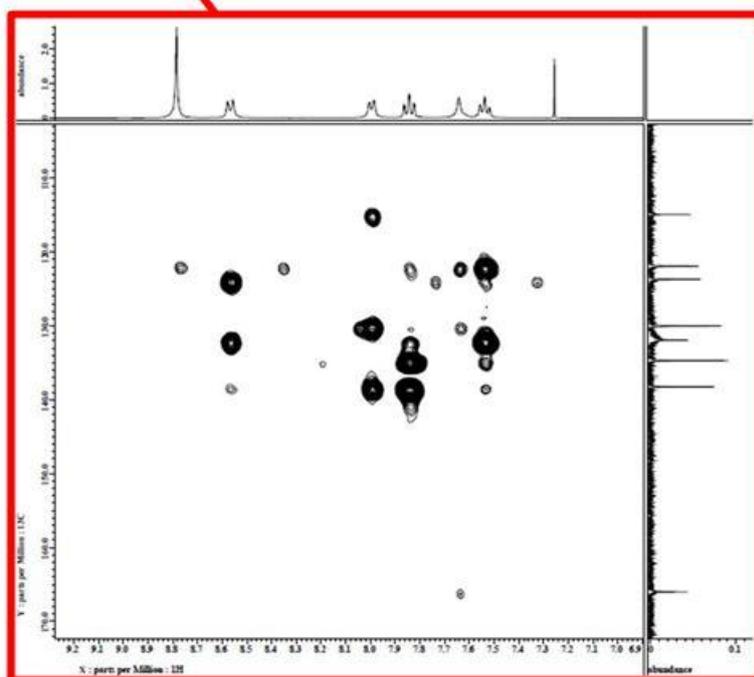
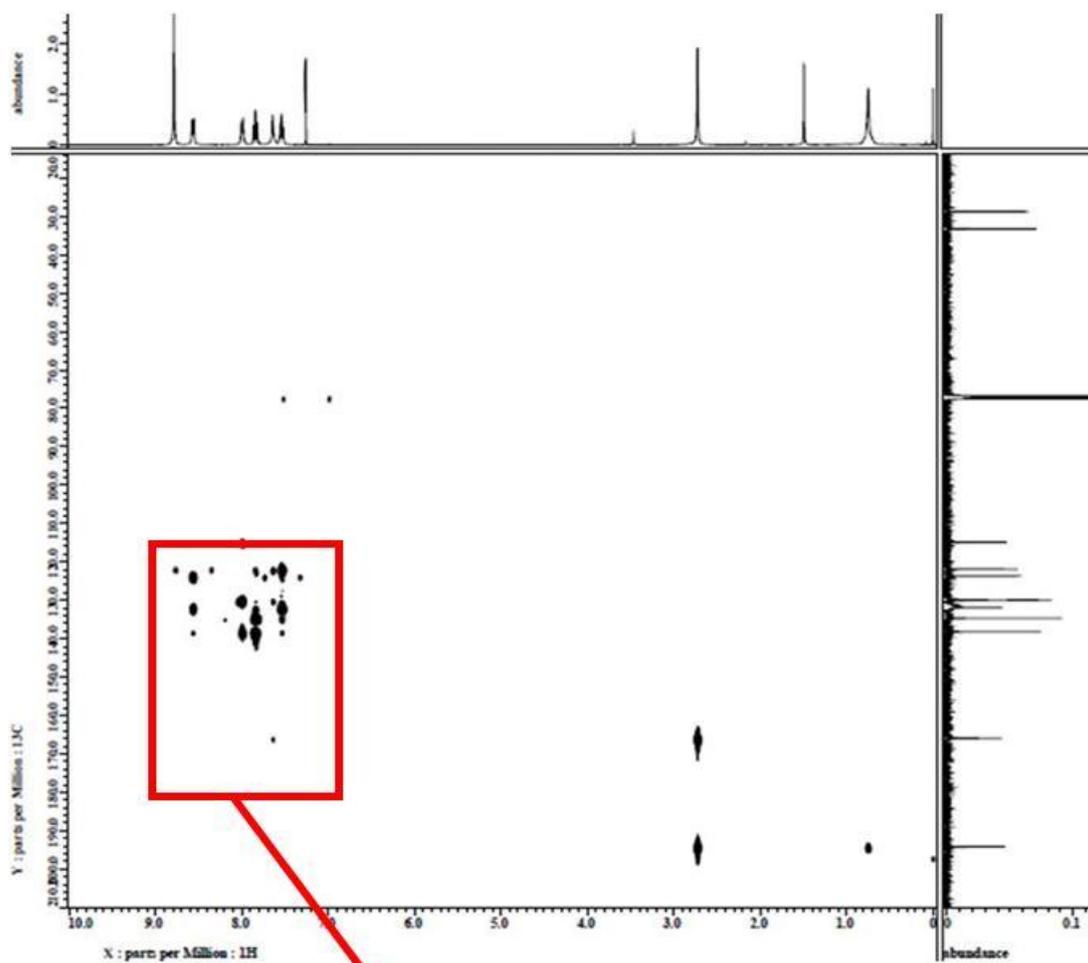
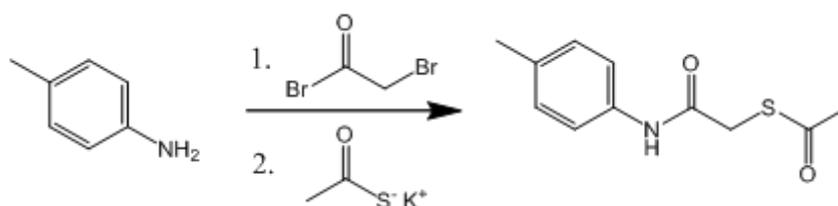


Fig. S4 HMBC chart of **1**.

4-(S-Acetyl-thioacetamido)-toluene (2).



Compound **2** was synthesized by the similar procedure to **1**. *p*-Toluidine (535.8 mg, 5.00 mmol) was reacted with bromoacetyl bromide (1.1 mL, 12.6 mmol) in the presence of diethylaniline (2.5 mL, 15.6 mmol) under N₂ and 0 °C, followed by reaction with potassium thioacetate (2.83 g, 24.8 mmol) under N₂. The product was purified by column chromatography on silica gel with chloroform and 100:1 chloroform:methanol, and followed by recrystallization from hexane-AcOEt. 0.786 g (3.52 mmol, 70%). TLC *R_f* = 0.50 (CHCl₃:MeOH 100:1); ¹H NMR (CDCl₃, 400 MHz): δ 8.02 (s, 4H, **H_d**), 7.37 (d, 2H, **H_c**), 7.12 (d, 2H, **H_b**), 3.65 (s, 2H, **H_e**), 2.45 (s, 3H, **H_a**), 2.31 (s, 3H, **H_f**). ¹³C-NMR (CDCl₃, 100 MHz): δ 197.0 (**C_h**), 166.2 (**C_f**), 135.0 (**C_b**), 134.2 (**C_e**), 129.5 (**C_c**), 120.0 (**C_d**), 34.2 (**C_g**), 30.3 (**C_i**), 20.9 (**C_a**).

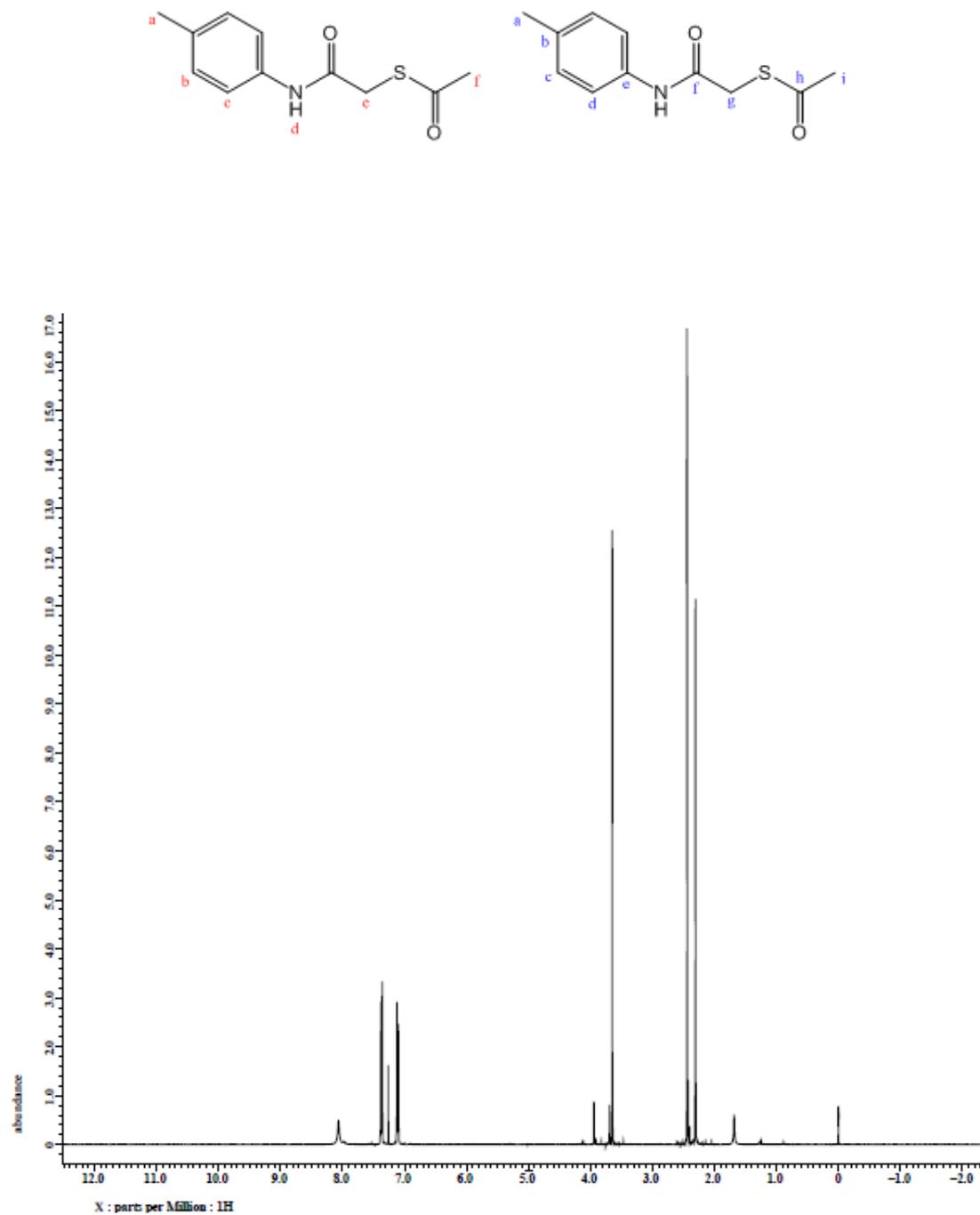


Fig. S5 $^1\text{H-NMR}$ chart of 2.

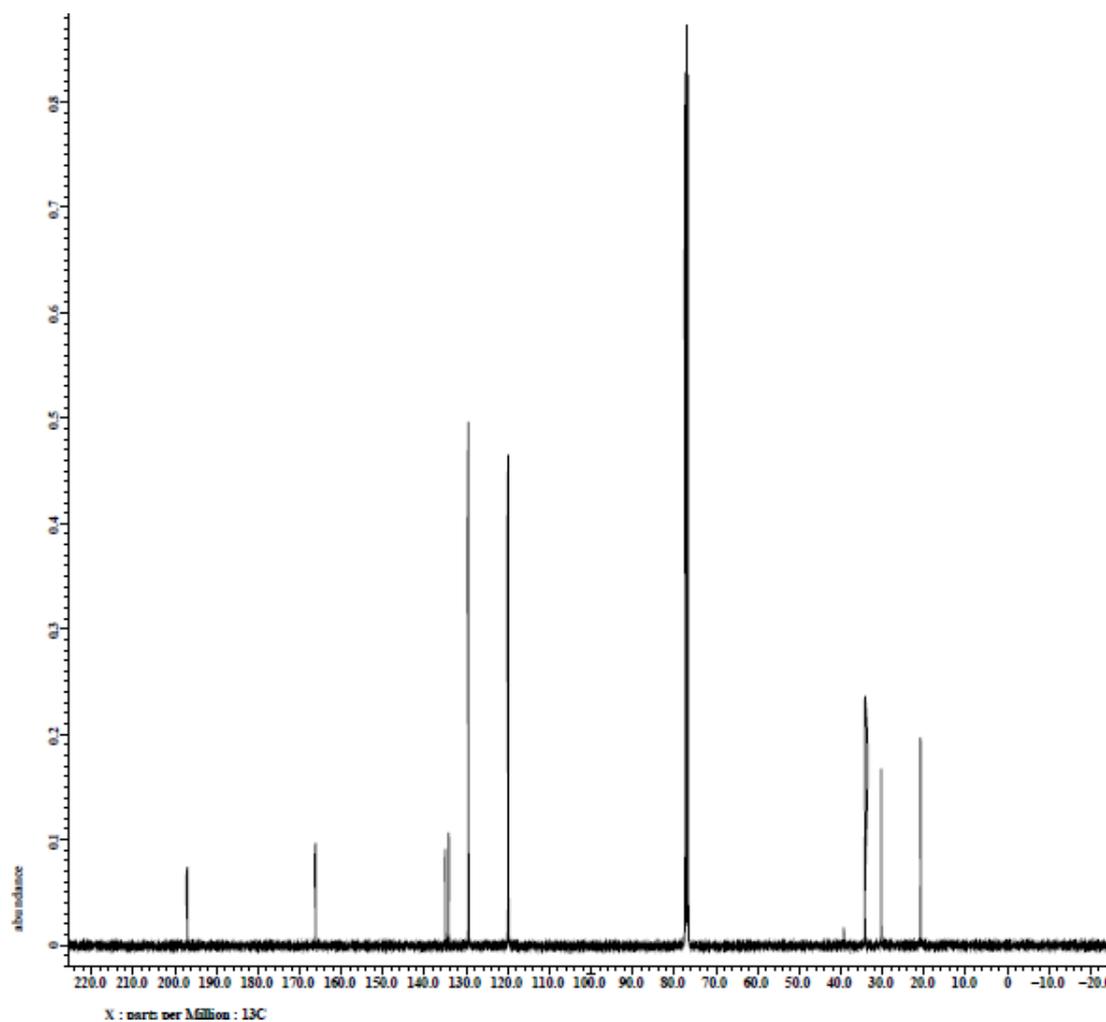


Fig. S6 ^{13}C -NMR chart of **2**.

Gold nanoparticles covered with 1 (GN@1).

A solution of **1** (11.4 mg, 10.0 μmol) in DMF (180 mL) was added to a 300 mL round-bottom flask which was cleaned with piranha solution. 40.0 μM HAuCl_4 solution in DMF (2 mL) was added and the mixture was reduced with NaBH_4 (18.25 mg, 772

μmol) dissolved in DMF. After 2 hour stirring, the reaction mixture was evaporated to ca. 3 mL and precipitated by 30 mL of methanol to remove excess amount of **1** and NaBH_4 . The precipitate was dissolved in 2 mL of DMF and reprecipitated by 30 mL of CHCl_3 . The reprecipitation process was repeated five times. Purified **GN@1** was characterized by TEM, UV-vis and XPS measurements.

Size controlled synthesis of gold nanoparticles. Gold nanoparticles were synthesized in the presence of **1**, **2**, and dodecanethiol under various molar ratios ($\text{S/Au} = 0, 0.1, 0.2, 1, 4$ and 16). All of glass vials for the syntheses of gold nanoparticles were cleaned with piranha solution. A typical method is as follows; 8 mL of 1.01 mM **1** solution of DMF ($8.08 \mu\text{mol}$), $93.5 \mu\text{L}$ of 21.4 mM hydrogen tetrachloroaurate trihydrate solution of DMF ($2.00 \mu\text{mol}$) and 1.8 mL of DMF were added to a reaction vessel. While the mixture was vigorously stirring, $100 \mu\text{L}$ of 0.248 M sodium borohydride (24.8 mmol) was swiftly added and then stirred for 1 hour. The solutions were evaporated and the residue was dispersed in MeOH. All samples were observed by TEM without purification.

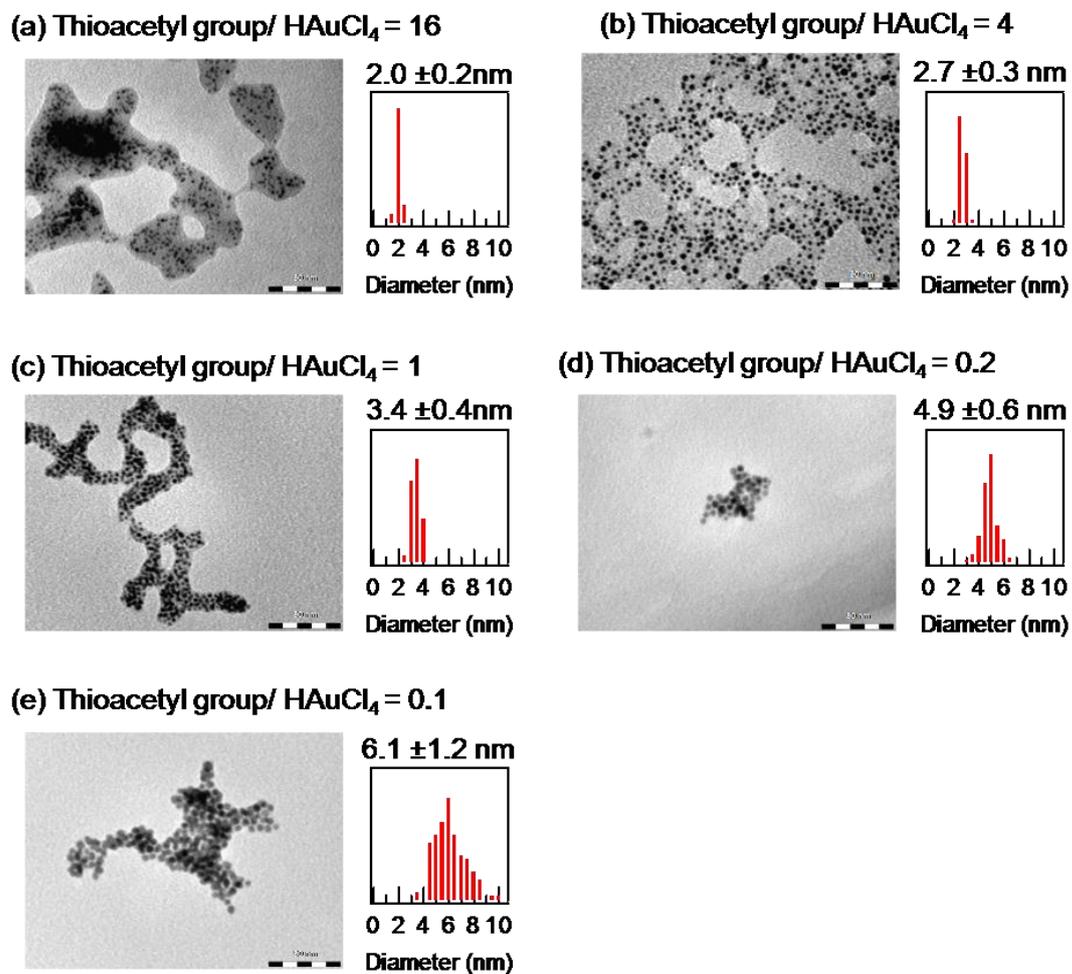
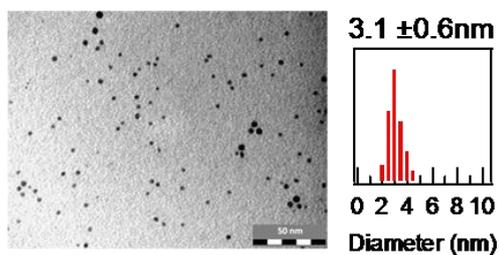
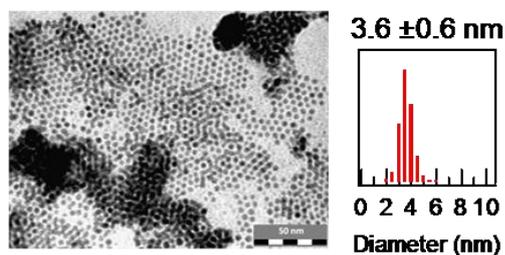


Fig. S7 TEM images and size distributions of as-synthesized GN@1 synthesized at various values of ξ .

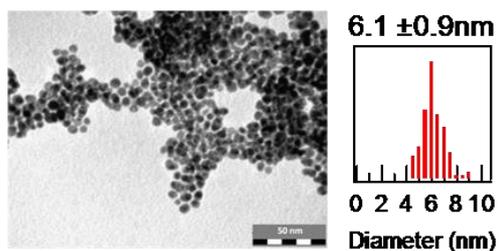
(a) Thioacetyl group/ $\text{HAuCl}_4 = 16$



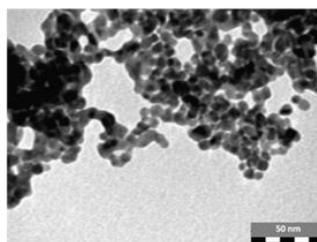
(b) Thioacetyl group/ $\text{HAuCl}_4 = 4$



(c) Thioacetyl group/ $\text{HAuCl}_4 = 1$



(d) Thioacetyl group/ $\text{HAuCl}_4 = 0.2$



(e) Thioacetyl group/ $\text{HAuCl}_4 = 0.1$

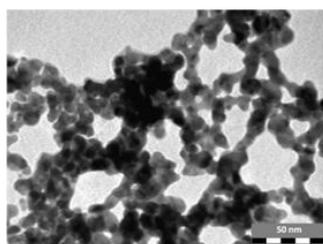


Fig. S8 TEM images and size distributions of as-synthesized GN@2 synthesized at various values of ξ . The TEM images of (d) and (e) didn't show particle boundaries because of aggregation of particles.

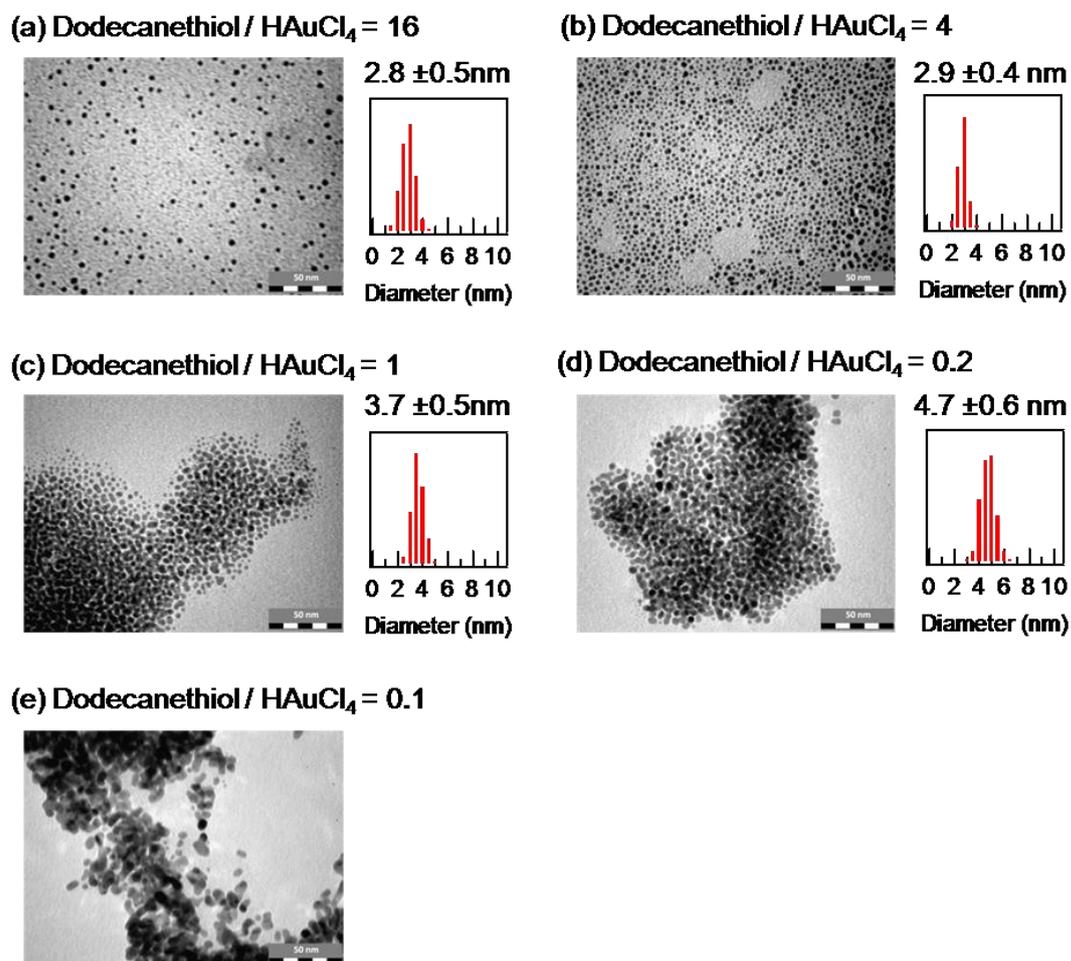


Fig. S9 TEM images and size distributions of as-synthesized gold nanoparticles covered with dodecanethiol synthesized at various values of ξ . The TEM image of (e) didn't show particle boundaries because of aggregation of particles.

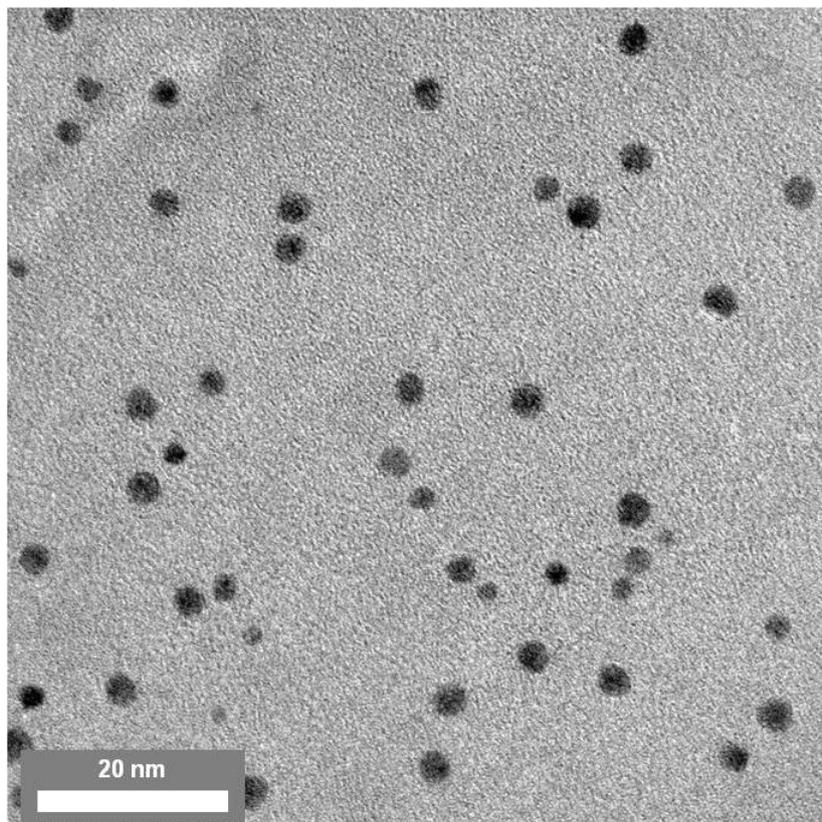


Fig. S10 TEM image of GN@1. The scale bar corresponds to 20 nm.

References

1. J. P. Collman, R. R. Gagne, C. Reed, T. R. Halbert, G. Lang and W. T. Robinson, *J. Am. Chem. Soc.*, 1975, **97**, 1427.
2. J. Lindsey, *J. Org. Chem.*, 1980, **45**, 5215.
3. R. J. Abraham, G. E. Hawkes, M. F. Hudson and K. M. Smith, *J. Chem. Soc. -Perkin Trans. 2*, 1975, 204.