SUPPORTING INFORMATION

for

Chiral Iron Complex Catalyzed Enantioselective Oxidation of Racemic Benzoins

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Table of contents

•	General considerations	6
•	Synthesis of ligand L5	6
•	General experimental procedure for oxidative kinetic resolution	7
•	Spectral data for all benzions and benzils	8
•	Determination of % ee (HPLC spectra for all benzoins)	11
•	Calculation of conversion C (%) using ¹ H NMR	16
•	¹ H and ¹³ C spectra for all compounds	21
•	References	32

General considerations

Iron(II)acetate and 2,2,6,6-tetramethyl-piperidin-1-oxyl (TEMPO) were purchased from Sigma-Aldrich chemical company and used as received. Thin layer chromatography (TLC) was performed using Merck silica gel 60 F_{254} precoated plates (0.25 mm) and visualized by UV fluorescence quenching. Silica gel for column chromatography (particle size 100-200 mesh) purchased from SRL, India. Optical rotations were measured with Autopol IV - Rudolph Research Analytical Polarimeter. ¹H and ¹³C NMR spectra were recorded on a Bruker 400 MHz instrument. ¹H NMR spectra were reported relative to Me₄Si (δ 0.0 ppm) or residual CHCl₃ (δ 7.26 ppm). ¹³C NMR were reported relative to CDCl₃ (δ 77.16 ppm). FTIR spectra were recorded on a Nicolet 6700 spectrometer and are reported in frequency of absorption (cm⁻¹). High resolution mass spectra (HRMS) were recorded on Q-Tof Micro mass spectrometer. The enantiomeric excess (%ee) of benzoins were determined by JASCO uv-2070 plus HPLC using Daicel ChiralPAK AS-H (0.46cm ϕ x 25cm) column. ¹H and ¹³C NMR and HRMS Spectral data have been included for all compounds. HPLC spectra for the % *ee* determination of all optically active benzoins are given in this supporting information.

Synthesis of Ligand L5





Experimental Procedure

Synthesis of L5¹: In a 100 mL two neck round bottom flask equipped with reflux condenser, a mixture of L2 (568 mg, 2 mmol) and 3,5-di-*tert*-butylsalicylaldehyde (983 mg, 4.2 mmol) in methanol (40 mL) (10 ml of CHCl₃ added for better solubility of the reagents) was taken and refluxed for 12 hours. The reaction mixture was cooled to room temperature, solvent was removed under reduced pressure and the residue was purified by column chromatography on basic aluminum oxide to give L5 (1050 mg, 73%) as a yellow compound. Mp 109-113 °C (lit.^{1b} 112-114 °C); R_f 0.4 (in hexanes : ethyl acetate, 95:5 V/V); $[\alpha]_{25}^{D} = -432.28$ (c = 1 in CH₂Cl₂); ¹H NMR (400 MHz, CDCl₃): δ 12.72 (s, 2H), 8.65 (s, 2H), 8.03 (d, J = 8.8 Hz, 2H), 7.94 (d, J =

8.4 Hz, 2H), 7.57 (d, J = 9.2 Hz, 2H), 7.41-7.46 (m, 2H), 7.33 (d, J = 8 Hz, 2H), 7.27-7.31 (m, 2H), 7.24 (d, J = 2.4 Hz, 2H), 7.04 (d, J = 2.4 Hz, 2H), 1.22 (s, 36H); ¹³C NMR (100 MHz, CDCl₃): δ 162.9, 158.3, 144.2, 139.9, 136.7, 133.5, 132.7, 129.8, 129.4, 128.3, 127.6, 126.9, 126.7, 126.6, 125.6, 118.3, 117.4, 35.0, 34.2, 31.5, 29.3; IR (Neat) 3649, 2955, 2871, 1612, 1583, 1467, 818 cm⁻¹; HRMS (*m*/*z*): [M+1]⁺calcd for C₅₀H₅₇N₂O₂, 717.4420; found, 717.4423.

General experimental procedure for oxidative kinetic resolution

A mixture of L5 (35.8 mg, 0.05 mmol) and iron(II)acetate (8.7 mg, 0.05 mmol) in 8 mL of hexanes was taken in a reaction tube and stirred at room temperature for 10 minutes, TEMPO (3.9 mg, 0.025 mmol) was added to the reaction mixture. After stirring for 5 minunets, benzoin (106 mg, 0.5 mmol) was added and then the reaction mixture was stirred under an O_2 atmosphere (using O_2 balloon) at 60 °C for 25 hours. The reaction mixture was concentrated and the resulting residue was purified by silica gel column chromatography (eluents: hexanes-ethyl acetate) to give the benzil (77 mg, yield 73%) and unreacted benzoin (22 mg, yield 21%).



R_f 0.23 (hexanes : ethyl acetate, 90:10 v/v); $[\alpha]_{25}^{D} = -76.0$ (*c* = 1 in acetone); ¹H NMR (400 MHz, CDCl₃): δ 7.88-7.93 (m, 2H), 7.48–7.53 (m, 1H), 7.23-7.41 (m, 7H), 5.95 (d, *J* = 6 Hz, 1H), 4.55 (d, *J* = 6 Hz, 1H) ; ¹³C NMR (100 MHz, CDCl₃): δ 199.1, 139.2, 134.0,

133.7, 129.3, 129.2, 128.8, 128.7, 127.9, 76.4; IR (Neat) 3418, 1679, 1261, 1068 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₄H₁₂O₂Na₁, 235.0735; found, 235.0727. The enantiomeric excess (%*ee*) was determined to be 98% by HPLC using Daicel ChiralPAK AS-H column (15% *i*-PrOH / hexanes, 1 mL/min, 220 nm): t_R (major, 13.250 min), t_R (minor, 8.500 min).



R_f 0.43; (hexanes : ethyl acetate, 90:10 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.96-8.01 (m, 4H), 7.63-7.69 (m, 2H), 7.49-7.55 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ194.7, 135.0, 133.3, 130.1,

129.2; IR (Neat) 3064,1656 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₄H₁₀O₂Na₁, 233.0578; found, 233.0585.

Spectral data for all other optically active benzoins



R_f 0.13; (hexanes : ethyl acetate, 90:10 v/v): $[\alpha]^{D}_{25} = -141.2$ (*c* = 1 in methanol); ¹H NMR (400 MHz, CDCl₃): δ 7.82 (d, *J* = 8.0 Hz, 2H), 7.16–7.23 (m, 4H), 7.12 (d, *J* = 8 Hz, 2H), 5.89 (d, *J* = 6.0 Hz, 1H), 4.54 (d, *J* = 6.4 Hz, 1H), 2.35 (s, 3H), 2.28 (s, 3H); ¹³C NMR

(100 MHz, CDCl₃): δ 198.7, 145.0, 138.5, 136.5, 131.1, 129.9, 129.5, 129.4, 127.8, 75.9, 21.8, 21.3; IR (Neat) 3473, 2917, 1674 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₆H₁₆O₂Na₁, 263.1048; found, 263.1045. The enantiomeric excess (%*ee*) was determined to be 90% by HPLC using Daicel ChiralPAK AS-H column (15% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (major, 12.158 min), t_R (minor, 8.275 min).



R_f 0.22; (hexanes: ethyl acetate, 90:10 v/v): $[\alpha]_{25}^{D} = -76.6$ (*c* = 1 in methanol); ¹H NMR (400 MHz, CDCl₃): δ 7.86 (d, *J* = 8.4 Hz, 2H), 7.19–7.27 (m, 4H), 7.15 (d, *J* = 8 Hz, 2H), 5.90 (d, *J* = 6 Hz, 1H), 4.55 (d, *J* = 6 Hz, 1H), 2.55-2.69 (m, 4H), 1.15-1.24 (m, 6H);

¹³C NMR (100 MHz, CDCl₃): δ 198.7, 151.1, 144.8, 136.7, 131.3, 129.6, 128.8, 128.3, 127.8, 75.9, 29.1, 28.7, 15.5, 15.0; IR (Neat) 3462, 2965, 2873, 1672, 1252, 1041, 973 cm⁻¹; HRMS (*m/z*): [MNa]⁺ calcd for C₁₈H₂₀O₂Na₁, 291.1361; found, 291.1364. The enantiomeric excess (%*ee*) was determined to be 94% by HPLC using Daicel ChiralPAK AS-H column (15% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (major, 8.333 min), t_R (minor, 6.375 min).



R_f 0.23; (hexanes : ethyl acetate, 90:10 v/v): $[\alpha]_{25}^{D} = -127.8$ (*c* = 1.0 in methanol); ¹H NMR (400 MHz, CDCl₃): δ7.76 (s, 1H), 7.69 (d, *J* = 7.6 Hz, 1H), 7.33 (d, *J* = 7.6 Hz, 1H), 7.17-7.27 (m, 2H), 7.11-7.15 (m, 2H), 7.07 (d, *J* = 7.6 Hz, 1H), 5.90 (d, *J* = 6 Hz, 1H),

4.52 (d, J = 6.4 Hz, 1H), 2.35 (s, 3H), 2.30 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 199.2, 139.1, 138.9, 138.6, 134.7, 133.6, 129.6, 129.3, 129.0, 128.5, 128.3, 126.5, 125.0, 76.2, 21.4, 21.3; IR (Neat) 3452, 2920, 1676, 1276 cm⁻¹; HRMS (m/z): [MNa]⁺ calcd for C₁₆H₁₆O₂Na₁, 263.1048; found, 263.1043. The enantiomeric excess (%*ee*) was determined to be 95% by HPLC using Daicel ChiralPAK AS-H column (15% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (major, 10.233 min), t_R (minor, 7.075 min).



R_f 0.14; (hexanes : ethyl acetate, 90:10 v/v): $[\alpha]_{25}^{D} = -75.0$ (c = 1 in methanol); ¹H NMR (400 MHz, CDCl₃): δ7.83-7.87 (m, 2H), 7.27-7.39 (m, 7H), 5.90 (d, J = 6.4 Hz, 1H), 4.48 (d, J = 6.0 Hz, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 198.0, 140.6, 138.8, 131.9,

130.6, 129.4, 129.2, 128.9, 127.9, 76.5; IR (Neat) 3461, 3063, 1678 cm⁻¹; HRMS (m/z): [M]⁺ calcd for C₁₄H₁₁ClO₂Na₁, 269.0345; found, 269.0347. The enantiomeric excess (%*ee*) was determined to be 94% by HPLC using Daicel ChiralPAK AS-H column (15% *i*-PrOH/ hexanes, 1 mL/min, 220 nm): t_R (major, 10.833 min), t_R (minor, 7.767 min).

Spectral data for all other benzils



R_f 0.33; (hexanes : ethyl acetate, 90:10 v/v): ¹H NMR (400 MHz, CDCl₃): δ7.86 (d, J = 8.4 Hz, 4H), 7.30 (d, J = 8.4 Hz, 4H), 2.43 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 194.7, 146.2, 130.9, 130.2, 129.9, 22.1; IR (Neat) 2918, 1659 cm⁻¹; HRMS (*m*/*z*): [MNa]⁺ calcd

for C₁₆H₁₄O₂Na₁, 261.0891; found, 261.0892.



R_f 0.45; (hexanes : ethyl acetate, 90:10 v/v): ¹H NMR (400 MHz, CDCl₃): δ7.89 (d, J = 8.4 Hz, 4H), 7.32 (d, J = 8 Hz, 4H), 2.72 (q, J = 7.6 Hz, 4H), 1.26 (t, J = 7.6 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃): δ194.7, 152.3, 131.0, 130.3, 128.7, 29.3, 15.2; IR (Neat)

2968, 2933, 2875, 1668 cm⁻¹; HRMS (m/z): [M-H]⁺ calcd for C₁₈H₁₉O₂, 267.1385; found, 267.1388.



R_f 0.52; (hexanes : ethyl acetate, 90:10 v/v): ¹H NMR (400 MHz, CDCl₃): δ 7.76 (d, J = 9.2 Hz, 4H), 7.46 (d, J = 7.2 Hz, 2H), 7.39 (t, J = 7.6 Hz, 2H), 2.40 (s, 6H); ¹³C NMR (100 MHz, CDCl₃): δ 195.0, 139.0, 135.7, 133.1, 130.2, 128.9, 127.2, 21.3; IR (Neat)

2922, 1667, 729 cm⁻¹; HRMS (m/z): [M-H]⁺ calcd for C₁₆H₁₅O₂, 239.1072; found, 239.1070.



 R_f 0.43; (hexanes : ethyl acetate, 90:10 v/v): ¹H NMR (400 MHz, CDCl₃): δ7.91-7.99 (m, 4H), 7.65-7.71 (m, 1H), 7.48-7.56 (m, 4H); ¹³C NMR (100 MHz, CDCl₃): δ194.0, 193.2, 141.7, 135.2, 133.0, 131.5, 131.3, 130.1, 129.6, 129.2; IR (Neat) 1666, 1210, 712 cm⁻¹;

HRMS (m/z): $[M]^+$ calcd for C₁₄H₉ClO₂Na₁, 265.0189; found, 265.0187.

Calculation of selectivity (s) = Calculation of selectivity (k_{fast}/k_{slow}) was accomplished by using the method of kagan.^[9]

$$s = \frac{k_{\text{fast}}}{k_{\text{slow}}} = \frac{\ln \left[(1 - C)(1 - ee) \right]}{\ln \left[(1 - C)(1 + ee) \right]}$$

$$s = \text{selectivity factor}$$

$$C = \text{conversion}$$

$$ee = \text{enantiomeric excess of recovered alcohol}$$



m-methylbenzoin



4- Chlorobenzoin



p-ethylbenzoin



p-methylbenzoin





Calculation of conversion C (%) using ¹H NMR





Benzoin Con∨ersion = 76 %











¹H and ¹³C spectra for all compounds





























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