

Brønsted Acid Differentiated Metal Catalysis by Kinetic Discrimination

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Supporting information

General: Unless otherwise noted, all commercially available compounds were used as provided without further purification. Solvents used in reactions were p.A. grade and dried only if indicated. Dichloromethane was dried with CaH_2 before use. Solvents for chromatography were technical grade and distilled prior to use. Analytical thin-layer chromatography (TLC) was performed on Merck silica gel aluminum plates with F-254 indicator, visualized by irradiation with UV light. Column chromatography was performed using silica gel Merck 60 (particle size 0.063 – 0.2 mm). Solvent mixtures are understood as volume/volume.

^1H -NMR and ^{13}C -NMR were recorded on a Bruker AM 250 respectively a Bruker AV 300 or a Varian Inova 400 spectrometer in CDCl_3 . Data are reported in the following order: chemical shift (δ) in ppm; multiplicities are indicated br (broadened singlet), s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet); coupling constants (J) are in Hertz (Hz). Mass spectra (MS-EI, 70 eV) were conducted on GC-MS Shimadzu QP2010 (column: Equity[®]-5, length $\times$ I.D. 30 m $\times$ 0.25 mm, $d = 0.25\ \mu\text{m}$, lot # 28089-U, Supelco). ESI spectra were conducted on a VG-Plattform II (Fisons Instruments). IR spectra were recorded on a Jasco FT/IR-420 and a Perkin Elmer Spectrum 100 spectrometer and are reported in terms of frequency of absorption (cm^{-1}). Melting points were measured on a Büchi 530 apparatus. Optical rotations were measured on a Perkin Elmer 241 polarimeter.

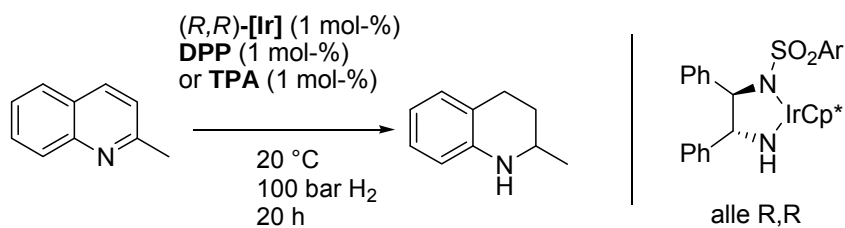
The enantiomeric excesses were determined by capillary GC analysis using a chiral stationary phase column: Shimadzu GC2010 with FID, column: Varian CP-Chirasil-DEX CB (25m $\times$ 0.25mm $\times$ 0.25 μm), carrier gas: H_2 or He. The enantiomeric excess of **4j-4n** were determined by HPLC analysis using a chiral stationary phase column (column: Daicel Co. CHIRALCEL OD-H or CHIRALPAK IA; eluent: hexane/2-propanol). The chiral GC and HPLC methods were calibrated with the corresponding racemic mixtures. Chemical yields refer to pure isolated substances. The yields and enantiomeric excesses are given in table.

Influence of solvent:

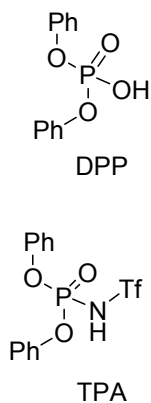
entry	solvent	time	conv ^[b]	e.r. ^[b]
1	o-xylene	7	55	92 : 8
2	toluene	7	45	92 : 8
3	benzene	7	30	92 : 8
4	trifluorotoluene	7	5	72 : 28
5	1,2-dichloroethane	7	15	57 : 43
6	chloroform	7	Spuren	66 : 34

^[a] conditions: 0.15 mmol **5**, 0.8 mL solvent^[b] determined via capillary GC

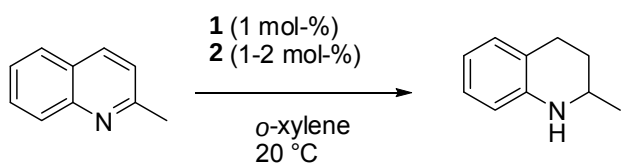
Influence of achiral additives using different enantiopure iridium catalysts



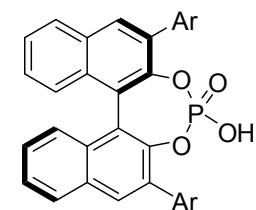
entry	Aryl	DPP	TPA
		%ee	%ee
1	p-Tolyl	ent 42	28
2	Mesityl	ent 60	ent 56
3	2-Naphthyl	ent 38	44
4	Pentamethylphenyl	56	54
5	3,5-Xylyl	ent 22	62
6	3,5-CF ₃ -Phenyl	54	62
7	4-Biphenyl	ent 28	46
8	Me	ent 6	62



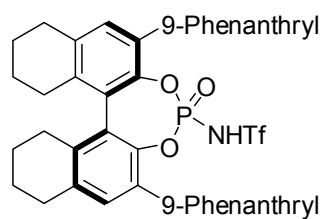
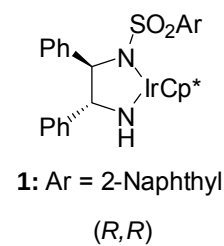
Influence of hydrogen pressure



entry	HA	mol%	pressure	temp	time	conv	%ee
2	2a	2	30	20	18	>95	84
3	2a	2	60	20	22	>95	86
4	2a	2	72	20	20	>95	86
5	2a	1	100	20	19	>95	90
6	2a	2	100	20	19	>95	89.5
7	9b	1	100	20	20	>95	88
8	9b	1	150	20	20	>95	86

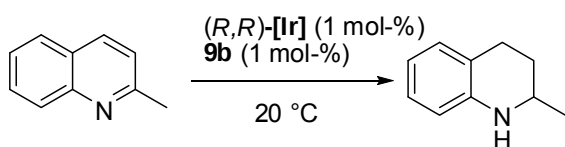


2a: Ar = 9-Phenanthryl

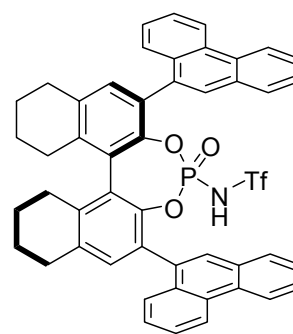
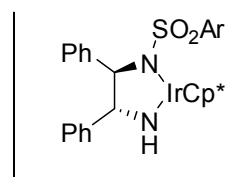


9b

Influence of the sulfonyl moiety



entry	mol%	time	Aryl	pressure	%ee
1*	1	10	p-Tolyl	50	75
2	1	10	Triisopropylphenyl	50->30	28
3	1	10	Mesityl	50->30	ent 13
4	1	14	2-Naphthyl	100	88
5	1	14	1-Naphthyl	100	31
6	1	14	9-Phenanthryl	100	37
7	1	14	3,5-Xylyl	100	54
8	1	14	3,5-CF ₃ -Phenyl	100	52
9	1	24	4-Biphenyl	100	86
10	1	24	Me	100	82



pressure in bar, first number indicates initial pressure, second number is pressure observed at the end of rct

Methods for the determination of enantiomeric excess via capillary GC

Method 1; 100 °C, hold temperature for 1 min, then heating 2 °C/min until a temperature of 135 °C is reached, then heating 20 °C/min until a temperature of 200 °C is reached. This temperature is held for 2 min. Carrier gas: H₂

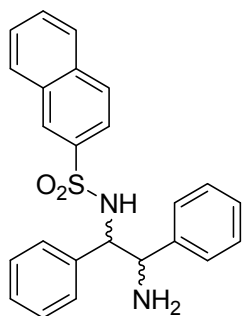
Method 2: 130 °C, then heating 4 °C/min until a temperature of 210 °C is reached. This temperature is held for 1 min. Carrier gas: He

Method 3: 100 °C, hold temperature for 1 min, then heating 2 °C/min until a temperature of 140 °C is reached, then heating 40 °C/min until a temperature of 200 °C is reached. This temperature is held for 1 min. Carrier gas: H₂

Method 4: 100 °C, hold temperature for 1 min, then heating 2 °C/min until a temperature of 160 °C is reached, then heating 40 °C/min until a temperature of 200 °C is reached. This temperature is held for 1 min. Carrier gas: H₂

Method 5: 100 °C, hold temperature for 1 min, then heating 2 °C/min until a temperature of 180 °C is reached, then heating 40 °C/min until a temperature of 200 °C is reached. This temperature is held for 1 min. Carrier gas: H₂

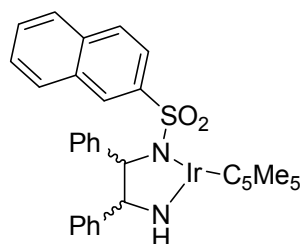
Synthesis of (*rac*)-1,2-Diphenyl-*N*-(2-naphthylsulfonyl)ethylenediamine



The title compound was prepared analogous to the literature method.¹ To an ice-cooled solution of *rac*-1,2-Diphenylethylenediamine (263 mg, 1.2 mmol) in dry Dichloromethane (5 mL) were subsequently added Diisopropylethylamine (0.3 mL, 1.8 mmol, 1.5 eq) and solid 2-Naphthylsulfonylchloride (280 mg, 1.2 mmol, 1 eq). After stirring overnight the crude reaction mixture was purified via column chromatography using hexanes/ethylacetate eluent to yield 342 mg (69 %) of a white fluffy solid.

mp: 183 °C; ¹H-NMR (250 MHz, CDCl₃): δ = 7.97 (s, 1H), 7.79 (d, *J* = 7.9 Hz, 1H), 7.72 (d, *J* = 7.8 Hz, 1H), 7.63 (d, 8.6 Hz, 1H), 7.45-7.6 (m, 2H), 7.42 (dd, *J* = 8.6 Hz, *J* = 1.8 Hz, 1H), 6.9 – 7.2 (m, 10H), 4.47 (d, *J* = 5.1 Hz, 1H), 4.15 (d, *J* = 5.1 Hz, 1H), 1.6 (b, 3H); ¹³C-NMR (62.9 MHz, CDCl₃): δ = 141.079, 139.052, 137.089, 134.501, 131.961, 129.231, 128.855, 128.380, 128.345, 128.229, 128.198, 127.656, 127.528, 127.468, 127.037, 126.980, 126.459, 122.248, 63.335, 60.426; IR (KBr): ν = 3378, 3313, 3169, 3056, 3027, 2907, 1590, 1493, 1455, 1394, 1347, 1300, 1201, 1149, 1129, 1075, 958, 937, 905, 871, 819, 796, 767, 750, 698, 641, 564; MS-ESI: *m/z*(%): 403 [M+H]

Synthesis of *rac*-Cp*Ir[**8**] (**1b**)



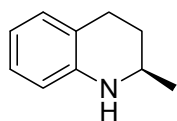
1b was prepared analogous to the literature method.² To a solution of [Cp*IrCl₂]₂ (103 mg, 0.13 mmol) and **8** (104 mg, 0.26 mmol, 2 eq) in 2 mL Dichloromethane Triethylamine (190 μL, 1.3 mmol, 10 eq) was added. The initially orange solution changed to red. After stirring overnight water was added and the red solution immediately turned dark burgundy red. The aqueous layer was extracted three times with Dichloromethane. The combined organic layers were dried over Na₂SO₄. After removal of solvent and thorough drying in high vacuum the residual

dark-red foamy solid was triturated with 4 mL hexanes overnight yielding a dark-burgundy red microcrystalline solid, which was collected after filtration and washing of the filtrate with hexanes (148 mg, 79 %).

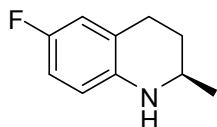
mp: 207 °C; $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 7.4-7.9 (m, 10H), 6.5-7.3 (m, 7H), 5.43 (b, 1H), 4.43 (s, 1H), 4.16 (d, J = 3.4 Hz, 1H), 1.92 (s, 15H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 146.673, 146.445, 141.023, 133.814, 131.993, 128.861, 128.674, 127.728, 127.695, 127.664, 127.415, 127.034, 126.892, 126.634, 126.510, 126.343, 126.127, 125.973, 123.392, 85.241, 80.084, 74.077, 10.186; IR (KBr): ν = 3331, 2911, 1598, 1491, 1448, 1296, 1126, 1074, 1027, 974, 814, 777, 747, 698, 660, 618; MS-ESI: m/z : 729 [M+H]

General procedure for iridium catalyzed hydrogenation

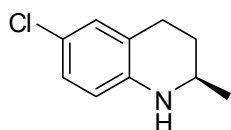
To a vial **1** (0.02mmol) and the appropriate additive and 1.0 mL of solvent were added. After stirring in air for 5 min quinoline (0.2 mmol) was added and the reaction mixture was put in a stainless steel autoclave. The system was connected to a supply of argon/hydrogen and the atmosphere in the autoclave was purged twice with Argon (5 bar). The autoclave was then filled with hydrogen (100 bar). After 16-48 h the pressure was released and the autoclave was purged with twice with Argon (5 bar). The crude reaction mixture was then purified via column chromatography using hexanes/ethylacetate.



2-Methyl-1,2,3,4-tetrahydro-quinoline³: $[\alpha]_{\text{D}}^{\text{RT}}$ = +80.8 (c = 1.0, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 6.9-7.1 (m, 2H), 6.61 (t, J = 7.3 Hz, 1), 6.47 (d, J = 8.2 Hz, 1H), 3.6 (b, 1H), 3.3-3.5 (m, 1H), 2.6-2.95 (m, 2H), 1.85-2.0 (m, 1H), 1.5-1.7 (m, 1H), 1.2 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (62.9 MHz, CDCl_3): δ = 144.623, 129.3, 126.7, 121.2, 117.1, 114.1, 47.2, 30.1, 26.6, 22.5; IR (NaCl): ν = 3393, 3049, 3014, 2961, 2923, 2843, 1607, 1489, 1451, 1375, 1341, 1308, 1276, 1257, 1153, 1124, 1039, 963, 746; MS-EI: m/z (%): 147 (37), 132 (100), 117 (15), 77 (9); Capillary GC: Method 1, minor enantiomer: t_{R} = 9.28 min, major enantiomer: t_{R} = 9.41 min

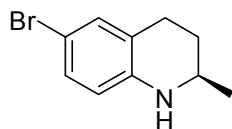


2-Methyl-6-Fluoro-1,2,3,4-tetrahydro-quinoline³: $[\alpha]_{\text{D}}^{\text{RT}}$ = +66.8 (c = 1.0, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 6.6-6.75 (m, 2H), 6.4-6.5 (m, 1H), 4.0 (b, 1H), 3.3-3.4 (m, 1H), 2.6-2.9 (m, 2H), 1.9-2.0 (m, 1H), 1.5-1.7 (m, 1H), 1.21 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 140.5, 122.7 (d, J = 6.9 Hz), 115.4 (d, J = 21.7 Hz), 115.0 (d, J = 7.7 Hz), 113.2 (d, J = 22.4 Hz), 47.4, 29.8, 26.6, 22.3; IR (NaCl): 3397, 2926, 2848, 1599, 1500, 1376, 1340, 1304, 1254, 1232, 1141, 805; MS-EI: m/z (%): 165 (40), 150 (100), 135 (28), 130 (14), 109 (9), 96 (9), 77 (10); Capillary GC: Method 3, minor enantiomer: t_{R} = 8.37 min, major enantiomer: t_{R} = 9.49 min



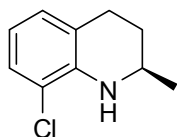
2-Methyl-6-Chloro-1,2,3,4-tetrahydro-quinoline: $[\alpha]_{\text{D}}^{\text{RT}}$ = +108.7 (c = 1.0, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 6.9-7.0 (m, 2H), 6.40 (d, J = 8.2 Hz, 1H), 3.96 (b, 1H), 3.3-3.45 (m, 1H), 2.6-2.9 (m, 2H), 1.9-2.0 (m, 1H), 1.5-1.6 (m, 1H), 1.21 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 143.0, 128.8, 126.5, 122.7, 121.5, 115.1, 47.2, 29.6, 26.4, 22.3; IR (KBr): ν = 3405, 2925, 2847, 1604, 1579, 1491, 1376, 1298, 1132, 876, 806; MS-EI:

$m/z(\%)$: 183 (17), 181 (56), 168 (30), 166 (100), 131 (83), 130 (81), 103 (15), 77 (30);
Capillary GC: Method 2, minor enantiomer: t_R = 13.19 min, major enantiomer: t_R = 13.36 min



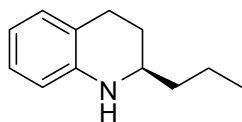
2-Methyl-6-Bromo-1,2,3,4-tetrahydro-quinoline: mp 48-50 °C; $[\alpha]_D^{RT}$ = +64.4 (c = 1.0, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 7.1-7.2 (m, 2H), 6.36 (d, J = 8.3 Hz, 1H), 4.0 (b, 1H), 3.3-3.45 (m, 1H), 2.6-2.9 (m, 2H), 1.85-2.0 (m, 1H), 1.45-1.65 (m, 1H), 1.21 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 143.480, 131.6, 129.3, 123.2, 115.5, 108.5, 47.1,

29.5, 26.3, 22.3; IR (KBr): ν = 3399, 2964, 2925, 2843, 1597, 1575, 1488, 1446, 1297, 1133, 877, 805; MS-EI: $m/z(\%)$: 227 (19), 225 (20), 212 (30), 210 (33), 144 (10), 131 (65), 130 (100), 103 (13), 77 (23); Capillary GC: Method 2, minor enantiomer: t_R = 15.47 min, major enantiomer: t_R = 15.63 min



2-Methyl-8-Chloro-1,2,3,4-tetrahydro-quinoline: $[\alpha]_D^{RT}$ = +63.8 (c = 1.0, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 7.06 (d, J = 7.9 Hz, 1H), 6.87 (d, J = 7.5 Hz, 1H), 6.5 (t, J = 7.7 Hz, 1H), 4.4 (b, 1H), 3.4-3.55 (m, 1H), 2.7-2.9 (m, 2H), 1.9-2.0 (m, 1H), 1.5-1.7 (m, 1H), 1.27 (d, J = 6.3 Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 140.7, 127.4, 126.7, 122.4, 117.8, 116.3, 47.1, 29.6, 26.7,

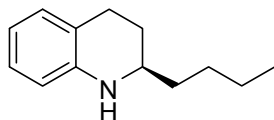
22.5; IR (NaCl): ν = 3413, 3065, 3019, 2924, 1602, 1568, 1493, 1376, 1343, 1330, 1295, 1188, 1139, 1098, 1090, 966, 933, 754, 722; MS-EI: $m/z(\%)$: 183 (13), 181 (44), 168 (26), 166 (88), 131 (67), 130 (100), 104 (27), 77 (34); Capillary GC: Method 2, major enantiomer: t_R = 9.33 min, minor enantiomer: t_R = 9.45 min



2-Propyl-1,2,3,4-tetrahydro-quinoline³: $[\alpha]_D^{RT}$ = +81.6 (c = 1.0, CHCl_3);

$^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 6.9-7.0 (m, 2H), 6.61 (t, J = 7.3 Hz, 1H), 6.51 (d, J = 8.2 Hz, 1H), 4.2 (b, 1H), 3.2-3.3 (m, 1H), 2.6-2.9 (m, 2H), 1.9-2.0 (m, 1H), 1.3-1.7 (m, 5H), 0.96 (t, J = 6.8 Hz, 3H); $^{13}\text{C-NMR}$ (75.4

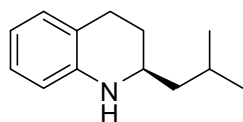
MHz, CDCl_3): δ = 144.4, 129.2, 126.7, 121.6, 117.1, 114.2, 51.4, 38.7, 28.0, 26.3, 18.9, 14.1; IR (NaCl): 3404, 3050, 2955, 2926, 2870, 1607, 1585, 1485, 1353, 1309, 1275, 1253, 1125, 745; MS-EI: $m/z(\%)$: 175 (12), 132 (100), 117 (15), 77 (7); Capillary GC: Method 4, minor enantiomer: t_R = 11.04 min, major enantiomer: t_R = 11.17 min



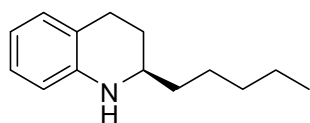
2-Butyl-1,2,3,4-tetrahydro-quinoline³: $[\alpha]_D^{RT}$ = +73.0 (c = 1.0,

CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): δ = 6.9-7.0 (m, 2H), 6.6-6.75 (m, 1H), 6.52 (d, J = 7.9 Hz, 1H), 4.3 (b, 1H), 3.2-3.3 (m, 1H), 2.6-2.9 (m, 2H), 1.9-2.0 (m, 1H), 1.3-1.7 (m, 7H), 0.93 (t, J = 7.0 Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): δ = 129.2, 126.7, 121.6, 117.1, 114.2, 51.6, 36.2, 28.0, 27.9, 26.3,

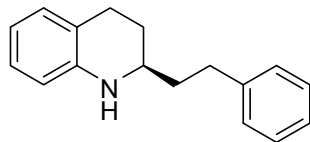
22.8, 14.076; IR (NaCl): ν = 3403, 2926, 2855, 1607, 1585, 1484, 1351, 1309, 1275, 745; MS-EI: $m/z(\%)$: 189 (9), 132 (100), 117 (13), 77 (6); Capillary GC: Method 4, minor enantiomer: t_R = 13.14 min, major enantiomer: t_R = 13.25 min



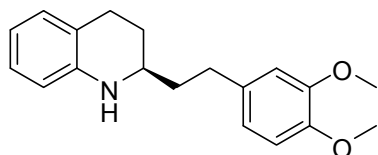
2-iso-Butyl-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{RT} = +79.2$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): $\delta = 6.9\text{--}7.0$ (m, 2H), 6.55–6.65 (m, 1H), 6.47 (d, $J = 8.2$ Hz, 1H), 3.7 (b, 1H), 3.3–3.4 (m, 1H), 2.6–2.9 (m, 2H), 1.9–2.0 (m, 1H), 1.7–1.9 (m, 1H), 1.5–1.7 (m, 1H), 1.3–1.5 (m, 2H), 0.95 (d, $J = 6.6$ Hz, 6H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): $\delta = 144.6$, 129.2, 126.6, 121.4, 116.9, 114.1, 49.2, 45.9, 28.5, 26.4, 24.4, 23.2, 22.4; IR (NaCl): $\nu = 3407$, 3051, 3015, 1606, 1585, 1484, 1384, 1366, 1353, 1308, 1273, 1153, 1001, 927, 744, 716; MS-EI: $m/z(\%)$: 189 (11), 132 (100), 117 (11), 77 (5); Capillary GC: Method 5, minor enantiomer: $t_R = 11.97$ min, major enantiomer: $t_R = 12.12$ min



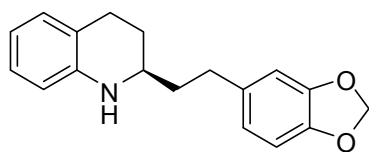
2-Pentyl-1,2,3,4-tetrahydroquinoline³: $[\alpha]_D^{RT} = +75.2$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): $\delta = 6.9\text{--}7.0$ (m, 2H), 6.60 (dt, $J = 7.4$ Hz, $J = 1.0$ Hz, 1H), 6.48 (d, $J = 7.4$ Hz, 1H), 3.88 (b, 1H), 3.2–3.3 (m, 1H), 2.7–2.9 (m, 2H), 1.9–2.05 (m, 1H), 1.55–1.7 (m, 1H), 1.3–1.55 (m, 8H), 0.91 (t, $J = 6.8$ Hz, 3H); $^{13}\text{C-NMR}$ (75.4 MHz, CDCl_3): $\delta = 144.6$, 129.2, 126.7, 121.4, 116.9, 114.1, 51.6, 36.6, 31.9, 28.1, 26.4, 25.4, 22.6, 14.0; IR (NaCl): $\nu = 3407$, 2925, 2853, 1607, 1585, 1484, 1352, 1309, 1253, 1125, 745; MS-EI: $m/z(\%)$: 203 (8), 132 (100), 117 (11), 77 (4); Capillary GC: Method 4, minor enantiomer: $t_R = 15.40$ min, major enantiomer: $t_R = 15.49$ min



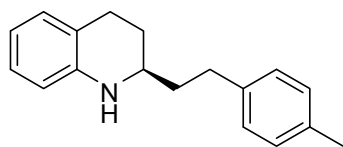
2-Phenethyl-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{RT} = +73.4$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (400 MHz, CDCl_3): $\delta = 7.27\text{--}7.34$ (m 3H), 7.17–7.22 (m, 2H), 6.93–6.96 (m, 2H), 6.60 (td, $J = 7.3$ Hz, $J = 1.1$ Hz, 1H), 6.43–6.48 (m, 1H), 3.75 (b, 1H), 3.25–3.35 (m, 1H), 2.7–2.85 (m, 4H), 1.95–2.05 (m, 1H), 1.8–1.88 (m, 2H), 1.62–1.74 (m, 1H); $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): $\delta = 144.4$, 141.7, 129.2, 128.4, 128.3, 126.6, 125.9, 121.2, 117.0, 114.1, 51.2, 38.3, 32.2, 28.1, 26.3; IR (KBr): $\nu = 3406$, 3055, 3023, 2921, 2849, 1605, 1490, 1451, 1354, 1310, 1116, 748, 699; MS-EI: $m/z(\%)$: 237 (7), 132 (100), 117 (30), 105 (20), 103 (16), 92 (13), 91 (84), 77 (24); HPLC: Daicel Chiralcel OD-H, *n*-hexanes/*i*Propanol 98/2, flow rate 0.6 mL/min, major enantiomer: $t_R = 27.9$ min, minor enantiomer $t_R = 30.5$ min



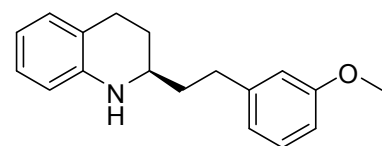
2-[2-(3,4-Dimethoxyphenyl)ethyl]-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{RT} = +57.9$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (400 MHz, CDCl_3): $\delta = 6.94\text{--}7.00$ (m, 2H), 6.81 (d, $J = 7.9$ Hz, 1H), 6.72–6.78 (m, 2H), 6.58–6.64 (m, 1H), 6.44–6.47 (m, 1H), 3.88 (s, 3H), 3.87 (s, 3H), 3.28–3.35 (m, 1H), 2.66–2.87 (m, 4H), 1.96–2.04 (m, 1H), 1.79–1.86 (m, 2H), 1.63–1.74 (m, 1H); $^{13}\text{C-NMR}$ 100.6 MHz, CDCl_3): $\delta = 148.8$, 147.2, 144.4, 134.3, 129.1, 126.6, 121.2, 120.0, 116.9, 114.0, 111.5, 111.2, 56.0, 55.9, 51.2, 38.5, 31.9, 28.1, 26.3; IR (KBr): $\nu = 3395$, 3012, 2931, 2845, 1605, 1513, 1310, 1261, 1151, 1029, 752; MS-EI: $m/z(\%)$: 297 (7), 151 (6), 132 (100), 117 (17), 105 (8), 78 (8); HPLC: Daicel Chiralcel OD-H, *n*-hexanes/*i*Propanol 80/20, flow rate 0.6 mL/min, major enantiomer: $t_R = 24.7$ min, minor enantiomer $t_R = 27.2$ min



2-[2-(1,3-Benzodioxol-5-yl)ethyl]-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{RT} = +63.8$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (250 MHz, CDCl_3): $\delta = 6.92\text{--}7.00$ (m, 2H), $6.70\text{--}6.76$ (m, 2H), $6.64\text{--}6.68$ (m, 1H), 6.61 (dd, $J = 7.3$ Hz, $J = 1.1$ Hz, 1H), $6.44\text{--}6.48$ (m, 1H), 5.93 (s, 2H), 3.76 (b, 1H), $3.25\text{--}3.33$ (m, 1H), $2.70\text{--}2.82$ (m, 2H), $2.63\text{--}2.70$ (m, 2H), $1.95\text{--}2.03$ (m, 1H), $1.75\text{--}1.83$ (m, 2H), $1.62\text{--}1.72$ (m, 1H); $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): $\delta = 147.5, 145.6, 144.4, 135.5, 129.1, 126.6, 121.2, 120.9, 117.0, 114.0, 108.7, 108.1, 100.7, 51.0, 38.5, 31.9, 28.0, 26.3$; IR (KBr): $\nu = 3408, 3011, 2920, 2850, 1606, 1488, 1442, 1354, 1039, 933, 752$; MS-EI: $m/z(\%)$: 281 (6), 145 (6), 132 (100), 117 (24), 91 (15), 77 (12); HPLC: Daicel Chiralcel OD-H, *n*-hexanes/*i*Propanol 90/10, flow rate 1.0 mL/min, major enantiomer: $t_R = 13.9$ min, minor enantiomer $t_R = 18.4$ min



2-(2-[4-Methylphenyl]ethyl)-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{RT} = +71.3$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (400 MHz, CDCl_3): $\delta = 7.14$ (s, 4H), $6.97\text{--}7.03$ (m, 2H), 6.64 (dd, $J = 7.4$ Hz, $J = 1.1$ Hz, 1H), $6.46\text{--}6.50$ (m, 1H), 3.8 (b, 1H), $3.29\text{--}3.36$ (m, 1H), $2.70\text{--}2.90$ (m, 4H), 2.36 (s, 3H), $1.99\text{--}2.06$ (m, 1H), $1.81\text{--}1.88$ (m, 2H), $1.65\text{--}1.76$ (m, 1H); $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): $\delta = 144.5, 138.6, 135.3, 129.2, 129.1, 129.0, 128.9, 128.7, 128.2, 126.6, 121.2, 116.9, 114.1, 51.1, 38.4, 31.8, 28.1, 26.3, 21.1$; IR (KBr): $\nu = 3407, 3012, 2920, 2850, 1606, 1585, 1485, 1449, 1354, 1309, 1216, 1117, 807, 750$; MS-EI: $m/z(\%)$: 251 (5), 144 (6), 132 (100), 117 (47), 105 (56), 91 (36), 77 (25), 65 (10); HPLC: CHIRALPAK IA *n*-hexanes/*i*Propanol 98/2, flow rate 1.0 mL/min, major enantiomer: $t_R = 7.5$ min, minor enantiomer $t_R = 6.95$ min

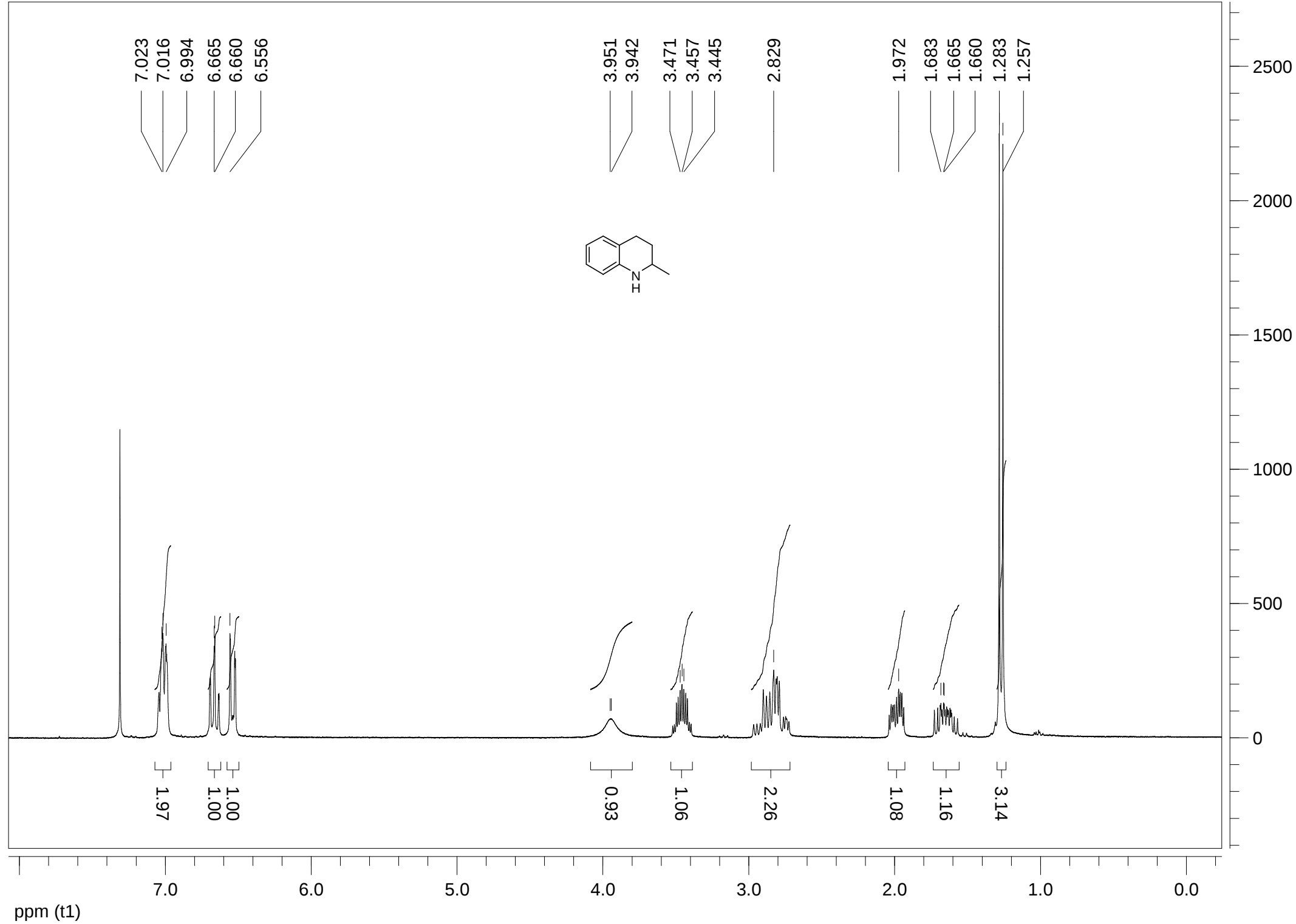
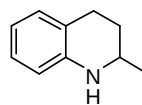


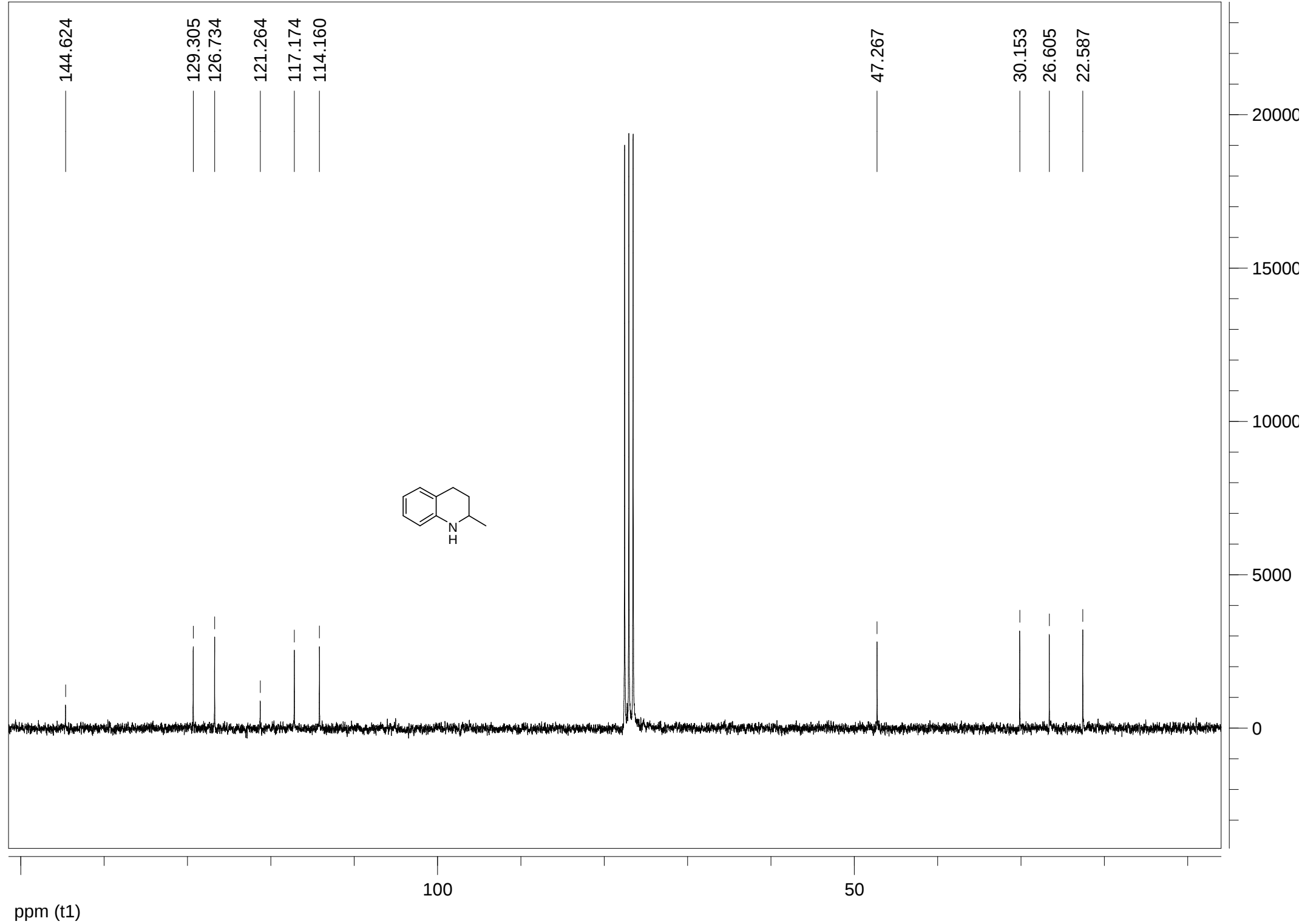
2-(2-[3-Methoxyphenyl]ethyl)-1,2,3,4-tetrahydroquinoline: $[\alpha]_D^{20} = +65.5$ ($c = 1.0$, CHCl_3); $^1\text{H-NMR}$ (400 MHz, CDCl_3): $\delta = 7.14\text{--}7.24$ (m, 1H), $6.92\text{--}6.98$ (m, 2H), $6.72\text{--}6.82$ (m, 3H), 6.61 (dd, $J = 7.3$ Hz, $J = 1.1$ Hz, 1), $6.43\text{--}6.47$ (m, 1H), 3.80 (s, 3H), 3.75 (b, 1H), $3.26\text{--}3.34$ (m, 1H), $2.69\text{--}2.86$ (m, 4H), $1.95\text{--}2.05$ (m, 1H), $1.80\text{--}1.87$ (m, 2H), $1.63\text{--}1.73$ (m, 1H); $^{13}\text{C-NMR}$ (100.6 MHz, CDCl_3): $\delta = 159.6, 144.4, 143.4, 129.4, 129.2, 126.6, 121.2, 120.7, 117.0, 114.1, 114.1, 111.1, 55.2, 51.1, 38.2, 32.3, 28.0, 26.3$; IR (KBr): $\nu = 3402, 3010, 2920, 2848, 1604, 1585, 1488, 1435, 1354, 1310, 1258, 1153, 1044, 874, 749, 695$; MS-EI: $m/z(\%)$: 267 (5), 145 (8), 132 (100), 117 (25), 91 (20), 77 (11); HPLC: Daicel Chiralcel OD-H, *n*-hexanes/*i*Propanol 90/10, flow rate 1.0 mL/min, major enantiomer: $t_R = 16.2$ min, minor enantiomer $t_R = 18.3$ min

¹ D. Xue, Y.-C. Chen, Xi. Cui, Q.-W. Wang, J. Zhu, J.-G. Deng, *J. Org. Chem.*, 2005, 70, 3584

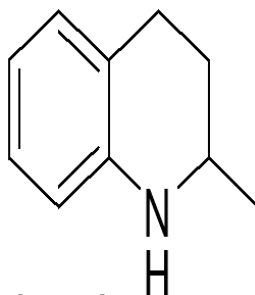
² Z. M. Heiden, T. B. Rauchfuss, *J. Am. Chem. Soc.* **2007**, 129, 14303

³ W.-B. Wang, S.-M. Lu, P.-Y. Yang, X.-W. Han, Y.-G. Zhou, *J. Am. Chem. Soc.* **2003**, 125, 10536

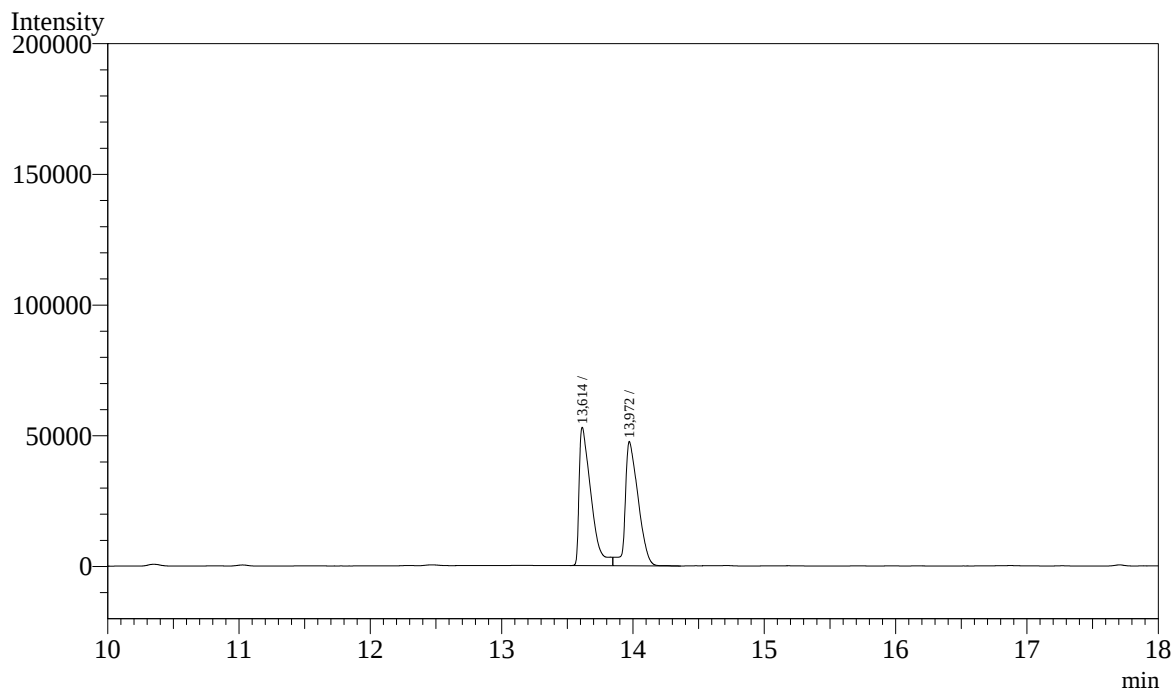




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 Sample ID :
 Sample Type : Unknown
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 ISTD Amount :

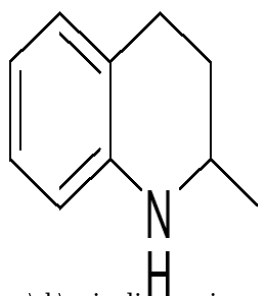


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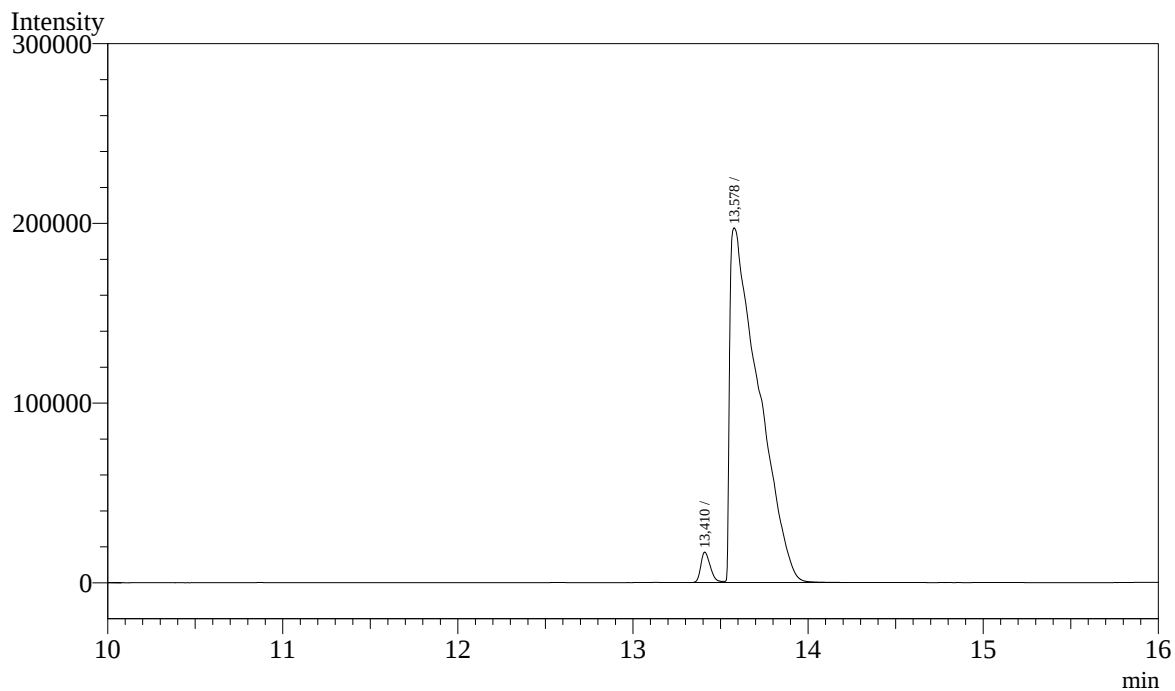


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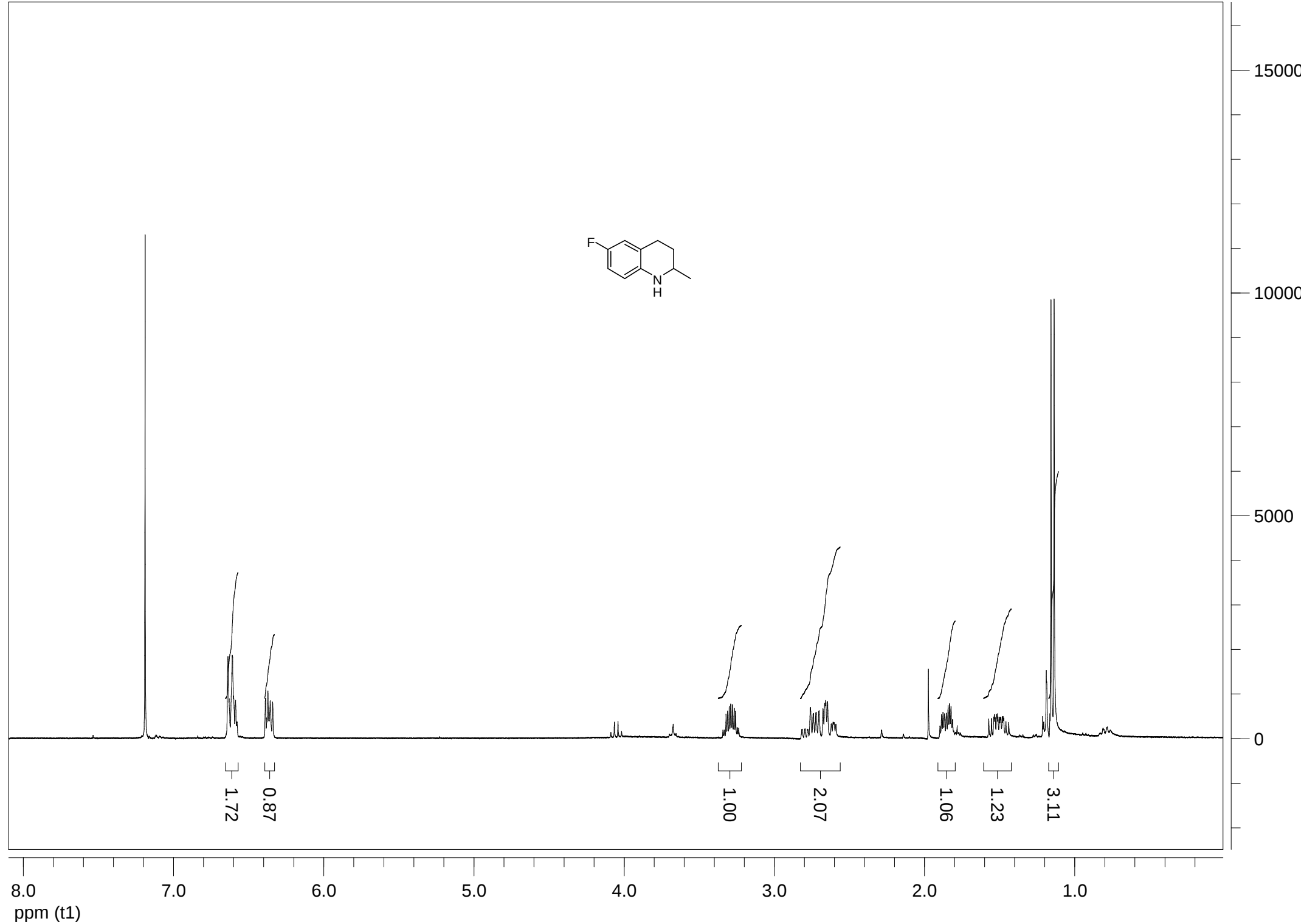
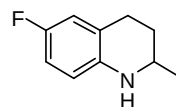
Analysis Date & Time : 10.09.2009 14:43:34
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 Vial# : 4
 Sample Name : rk641-51c
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

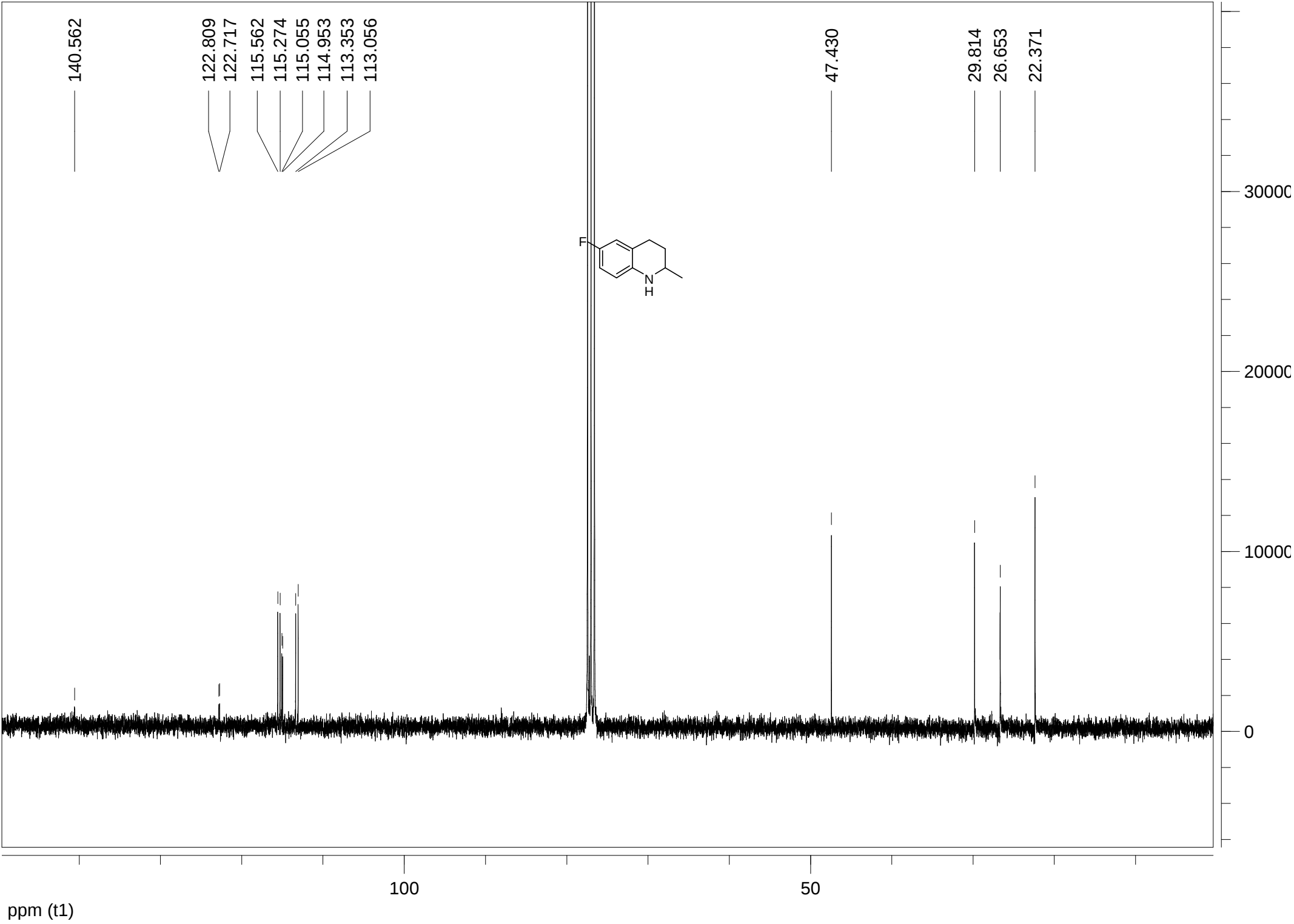
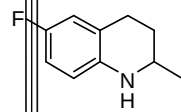


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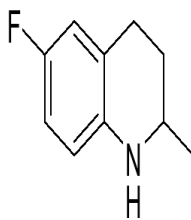


Peak#	Ret.Time	Area	Height	Area%
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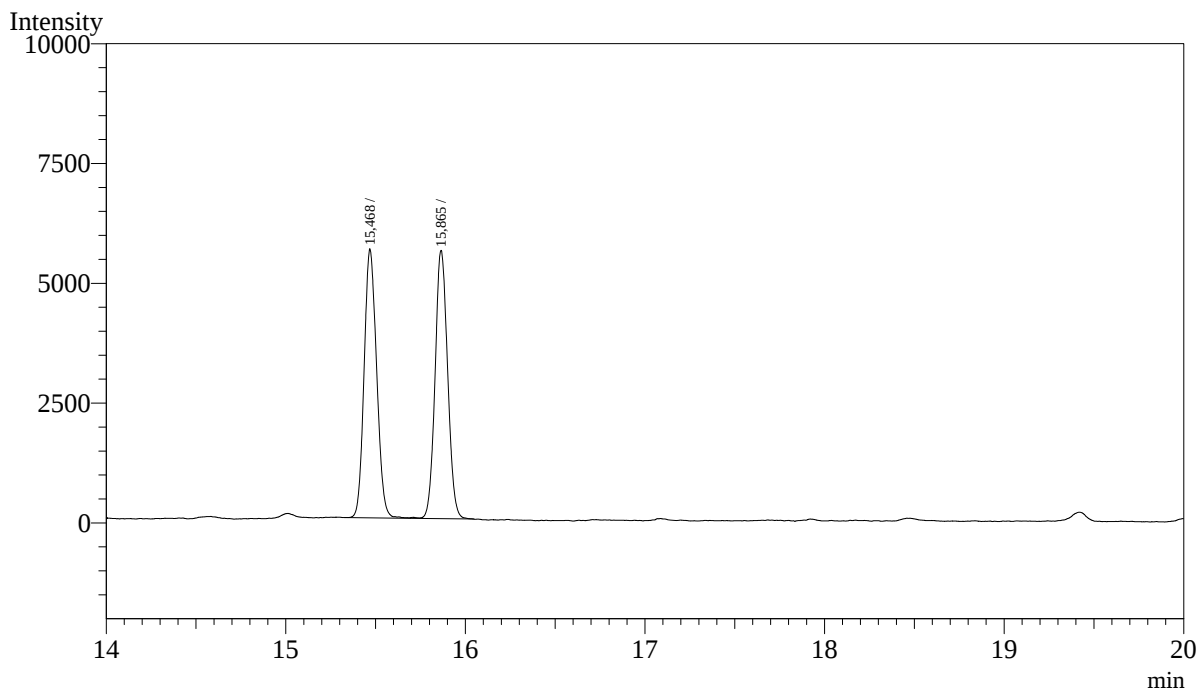




Analysis Date & Time : 28.08.2009 11:17:02
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 Sample Name : RK_6F_rac
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

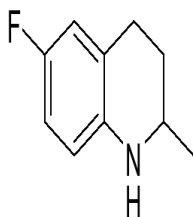


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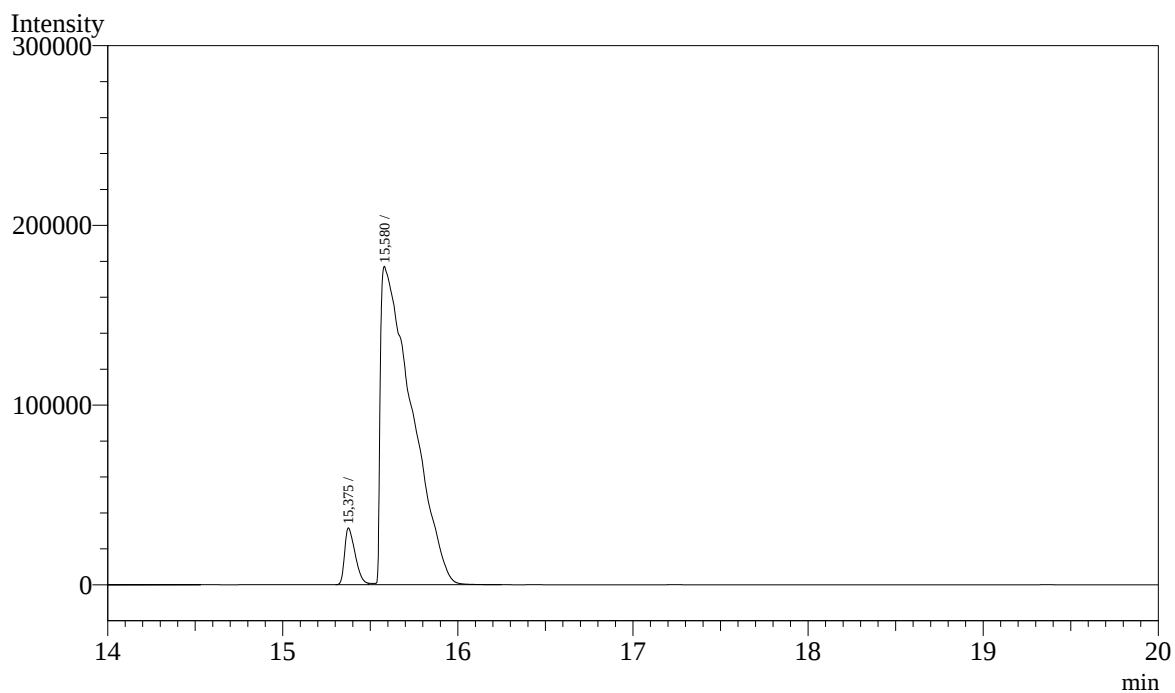


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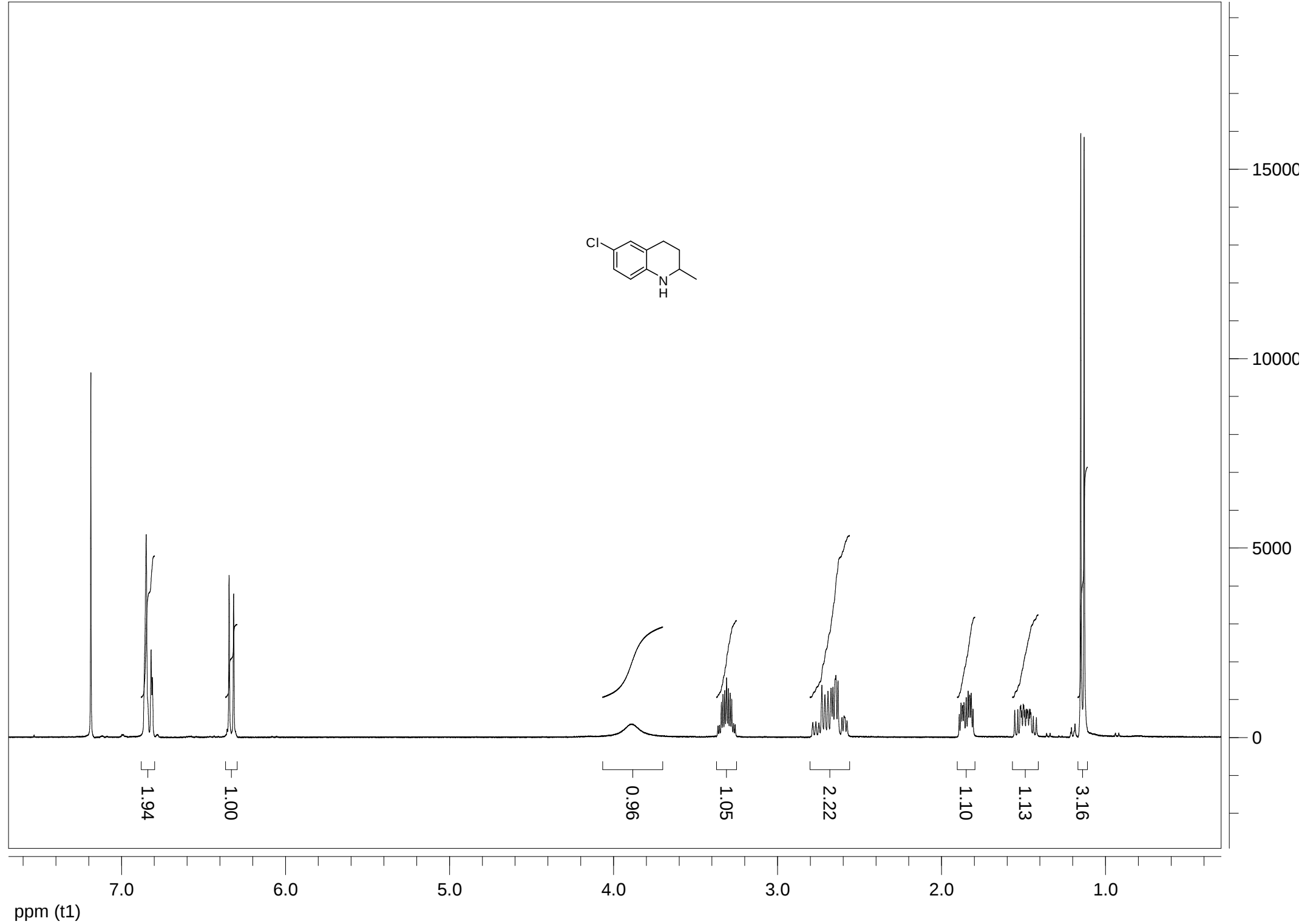
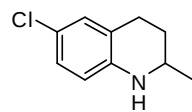
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 Sample ID :
 Sample Type : Unknown
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 ISTD Amount :

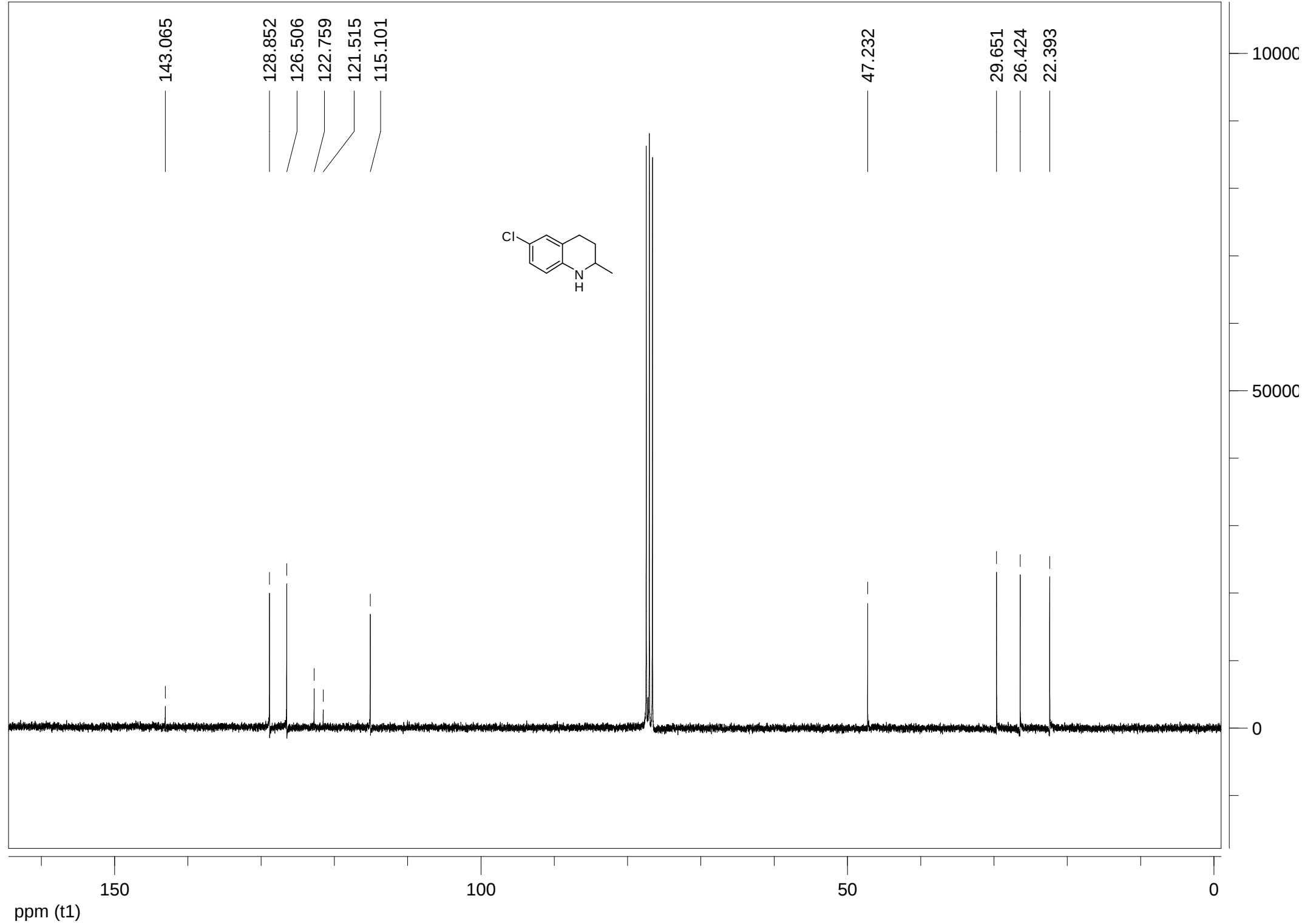


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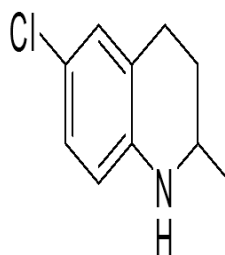


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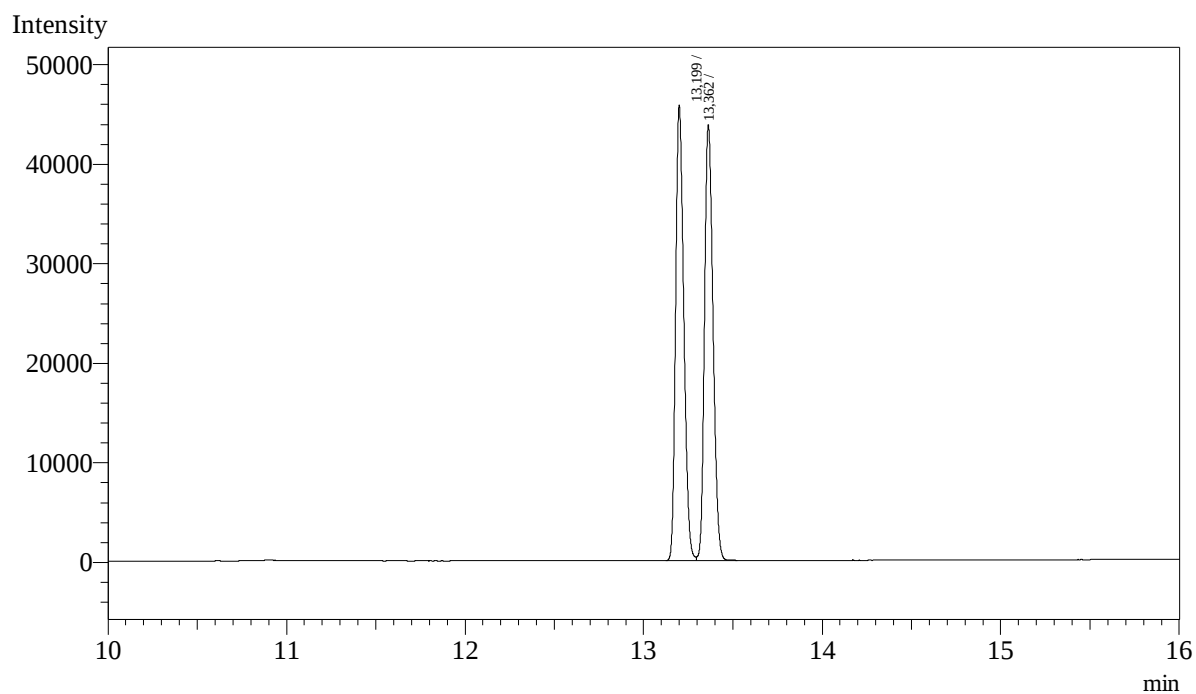




Analysis Date & Time : 26.01.2009 19:49:46
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 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

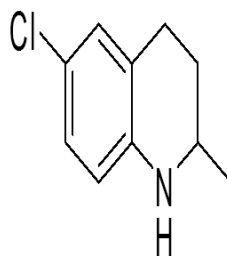


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 Method Name : D:\DATA_GC\rk\quinolines\rk-quinaldine-scope 1.gcm

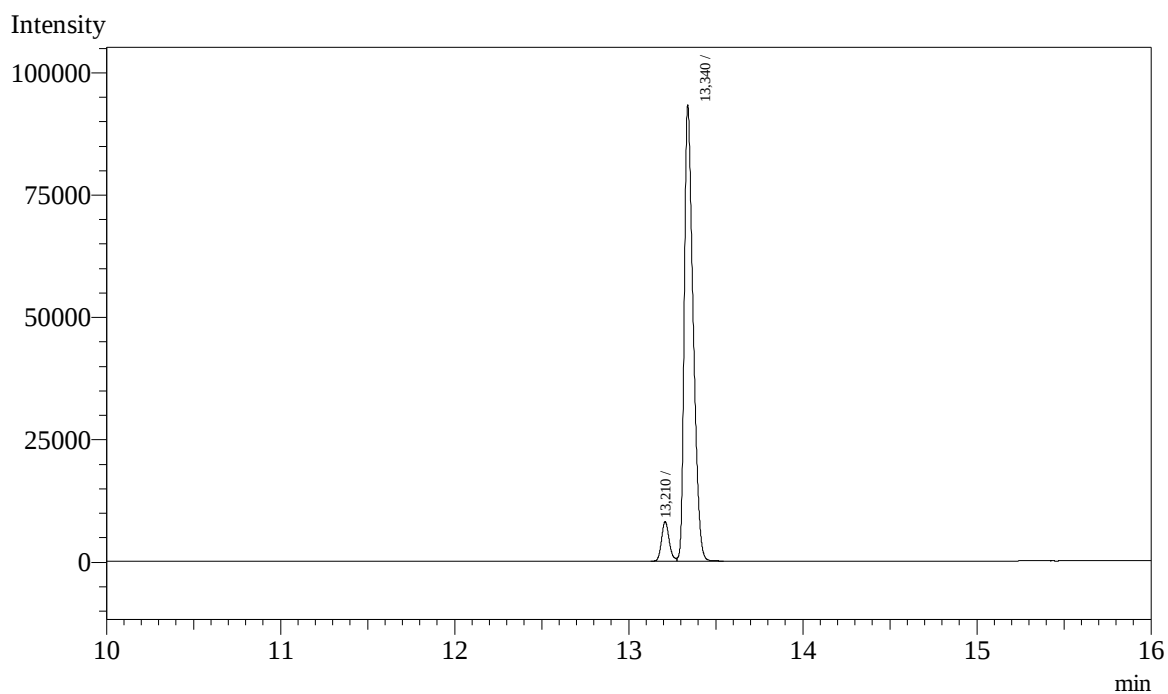


Peak#	Ret.Time	Area	Height	Conc.	Unit Mark	ID#	Cmpd Name
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2	13,362	145329	43634	49,984	V		
Total		290749	89287				

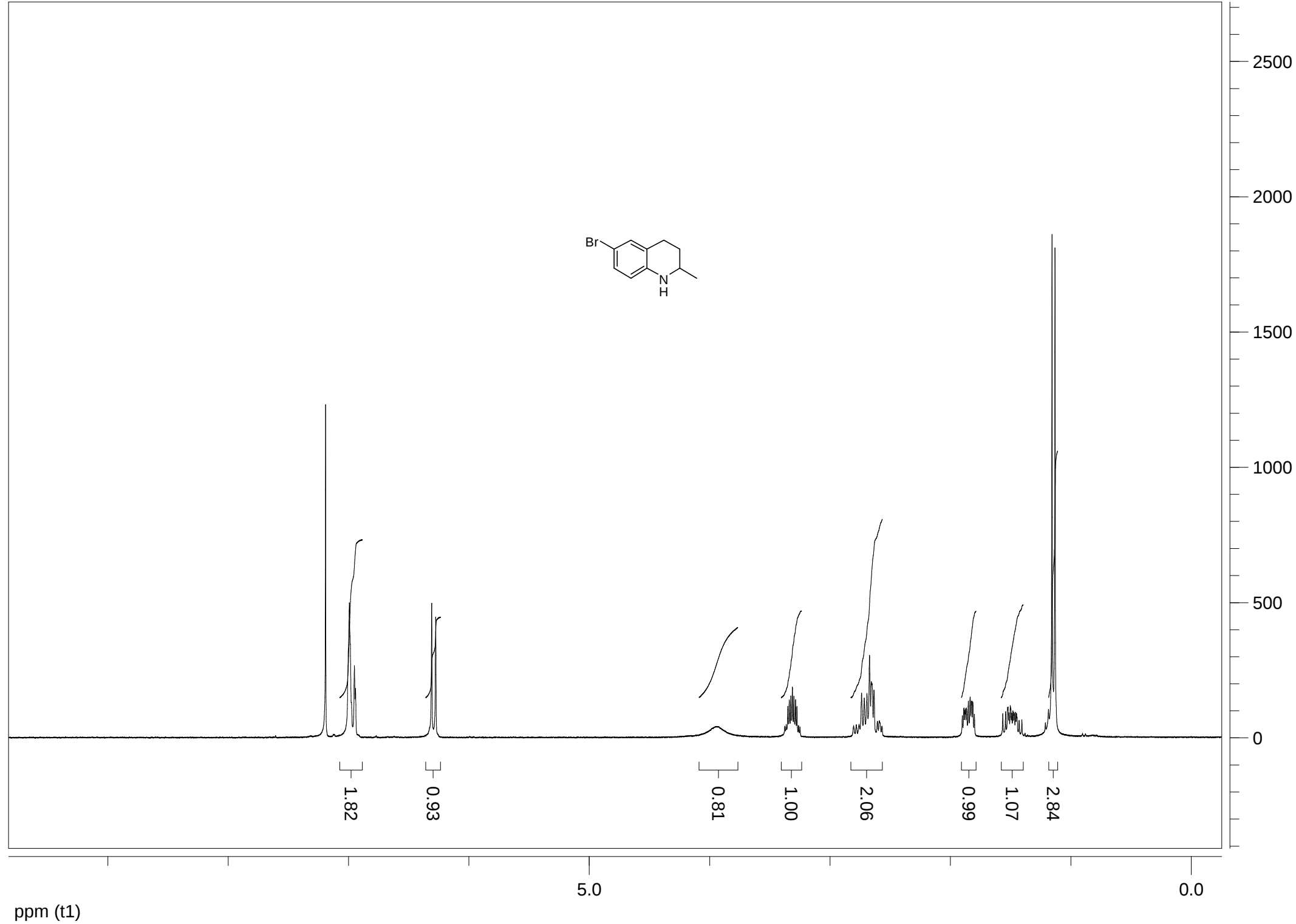
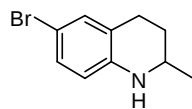
Analysis Date & Time : 10.02.2009 16:16:30
 User Name : Admin
 Vial# : 41
 Sample Name : rk537-5
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

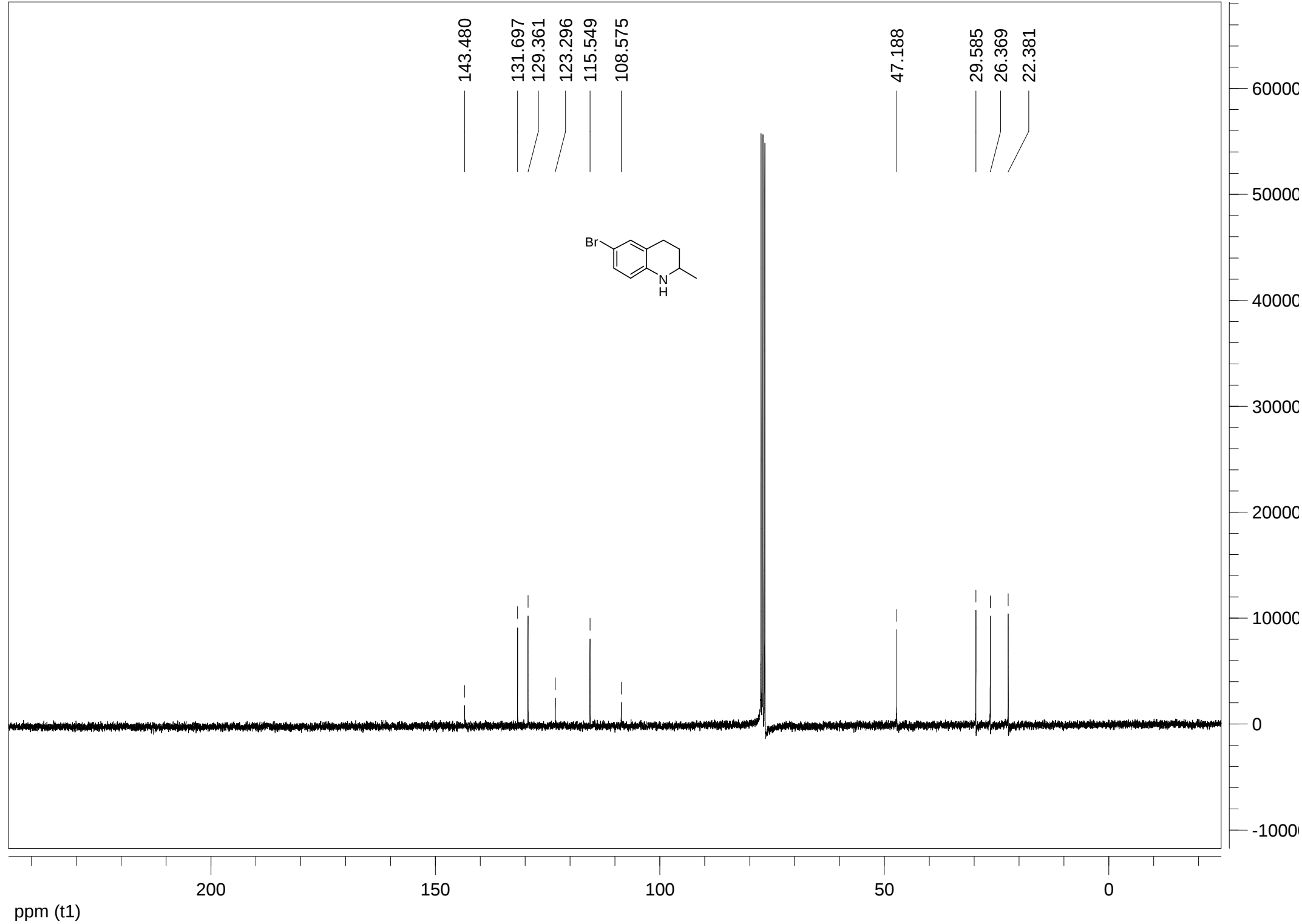


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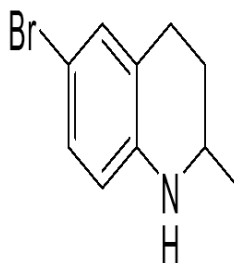


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2	13,340	329966	92984	93,096	V		
Total		354436	101023				

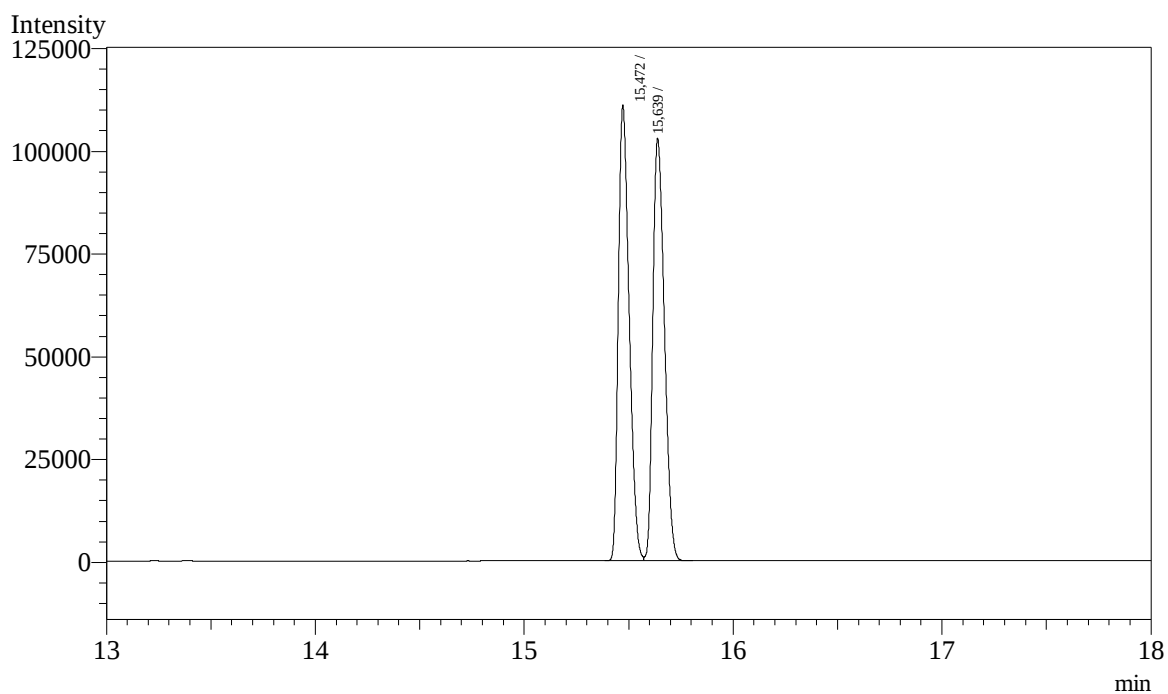




Analysis Date & Time : 26.01.2009 21:15:42
User Name : Admin
Vial# : 47
Sample Name : rk538-1
Sample ID :
Sample Type : Unknown
Injection Volume : 1,00
ISTD Amount :

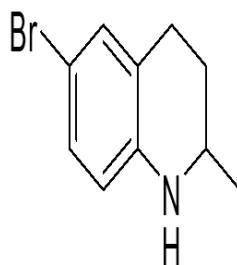


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Method Name : D:\DATA_GC\rk\quinolines\rk-quinaldine-scope 1.gcm



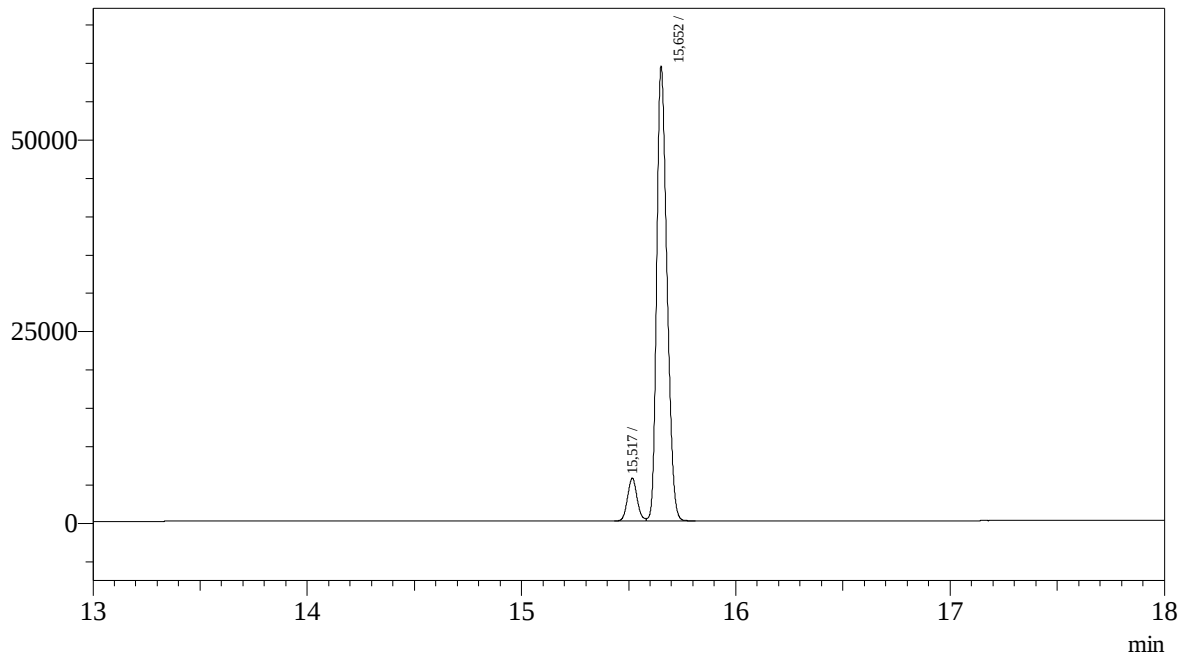
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1	15,472	399102	110642	50,182	V	
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Total		795303	212932			

Analysis Date & Time : 10.02.2009 16:42:27
 User Name : Admin
 Vial# : 42
 Sample Name : rk538-5
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

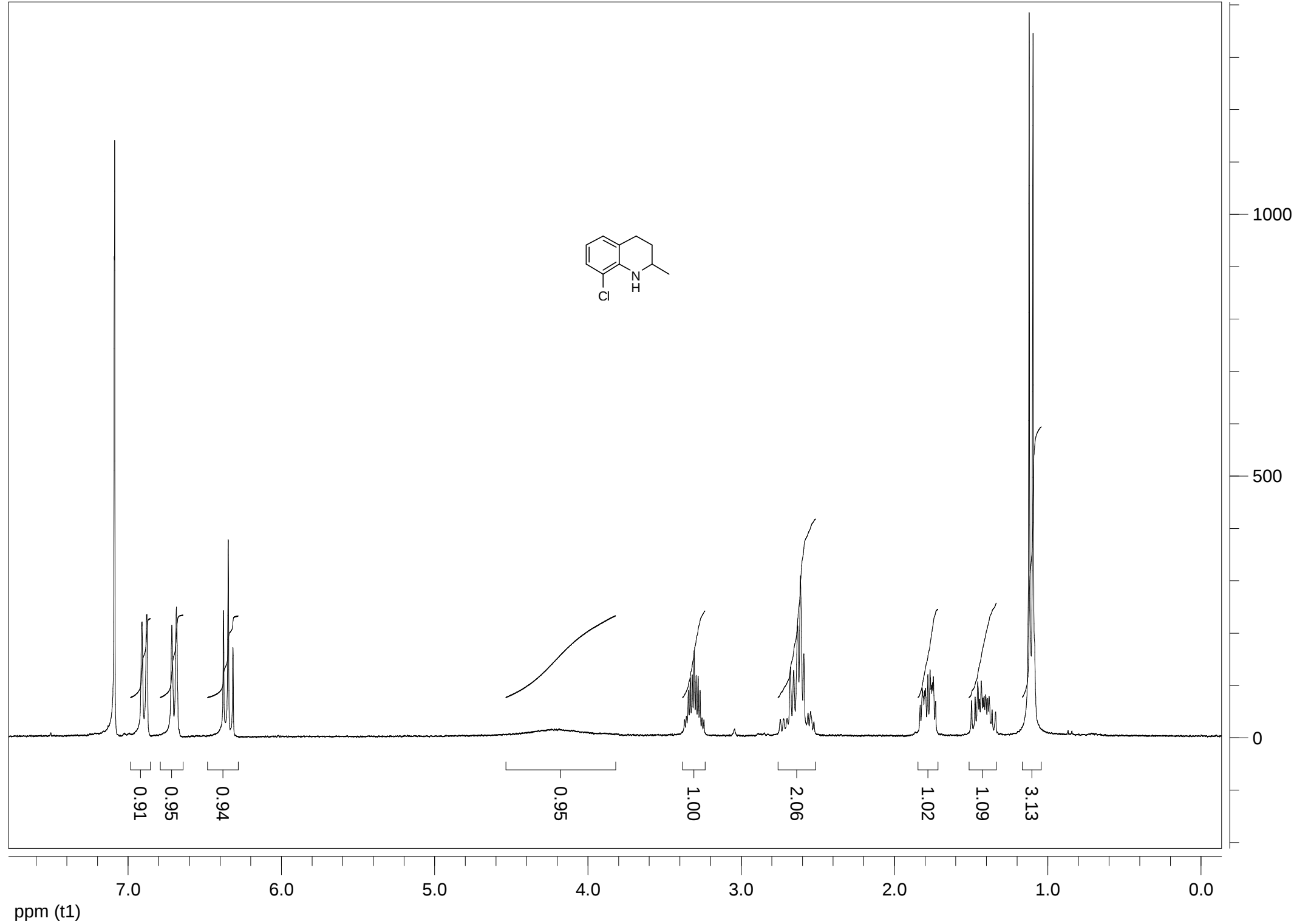
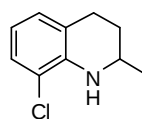


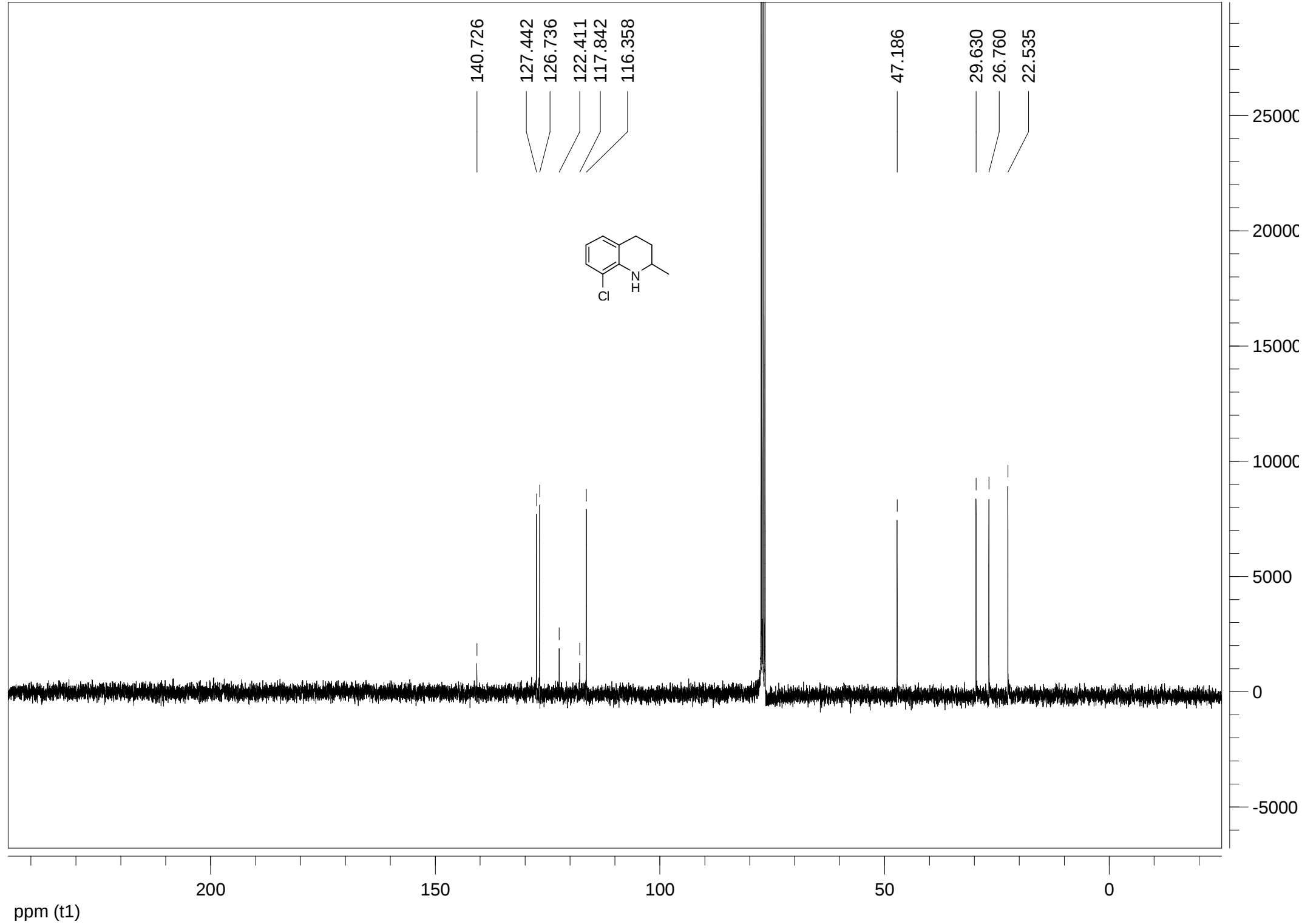
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Intensity

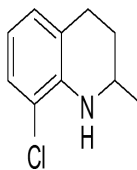


Peak#	Ret.Time	Area	Height	Conc.	Unit Mark	ID#	Cmpd Name
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2	15,652	197326	59067	91,982	V		
Total		214528	64604				

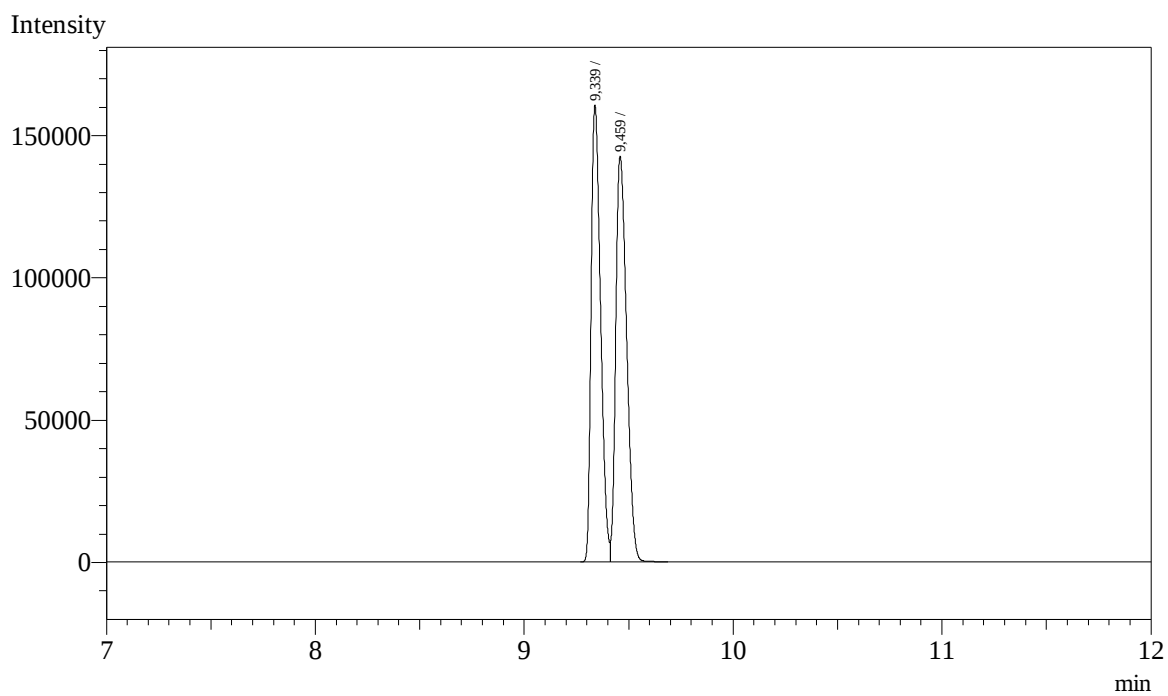




Analysis Date & Time : 26.01.2009 20:41:39
 User Name : Admin
 Vial# : 48
 Sample Name : rk540-1
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

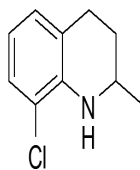


Data Name : D:\DATA_GC\rk\rk540-1.gcd
 Method Name : D:\DATA_GC\rk\quinolines\rk-quinaldine-scope 1.gcm

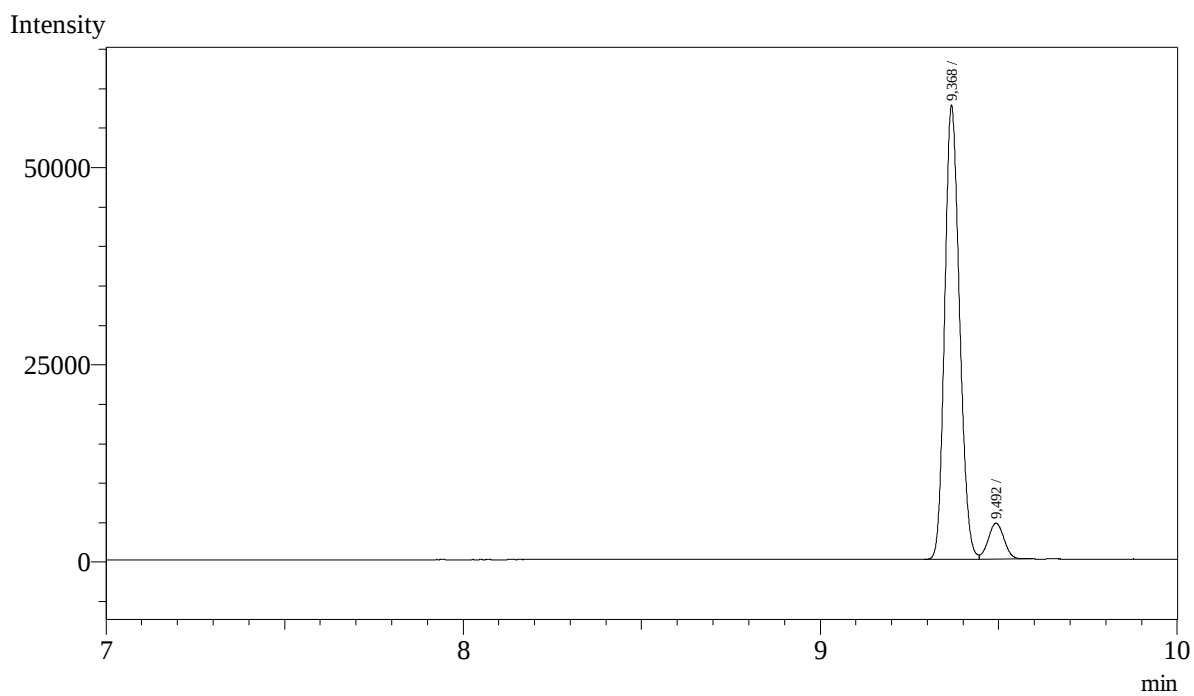


Peak#	Ret.Time	Area	Height	Conc.	Unit Mark ID#	Cmpd Name
1	9,339	505264	159822	49,548	V	
2	9,459	514483	141946	50,452		
Total		1019747	301768			

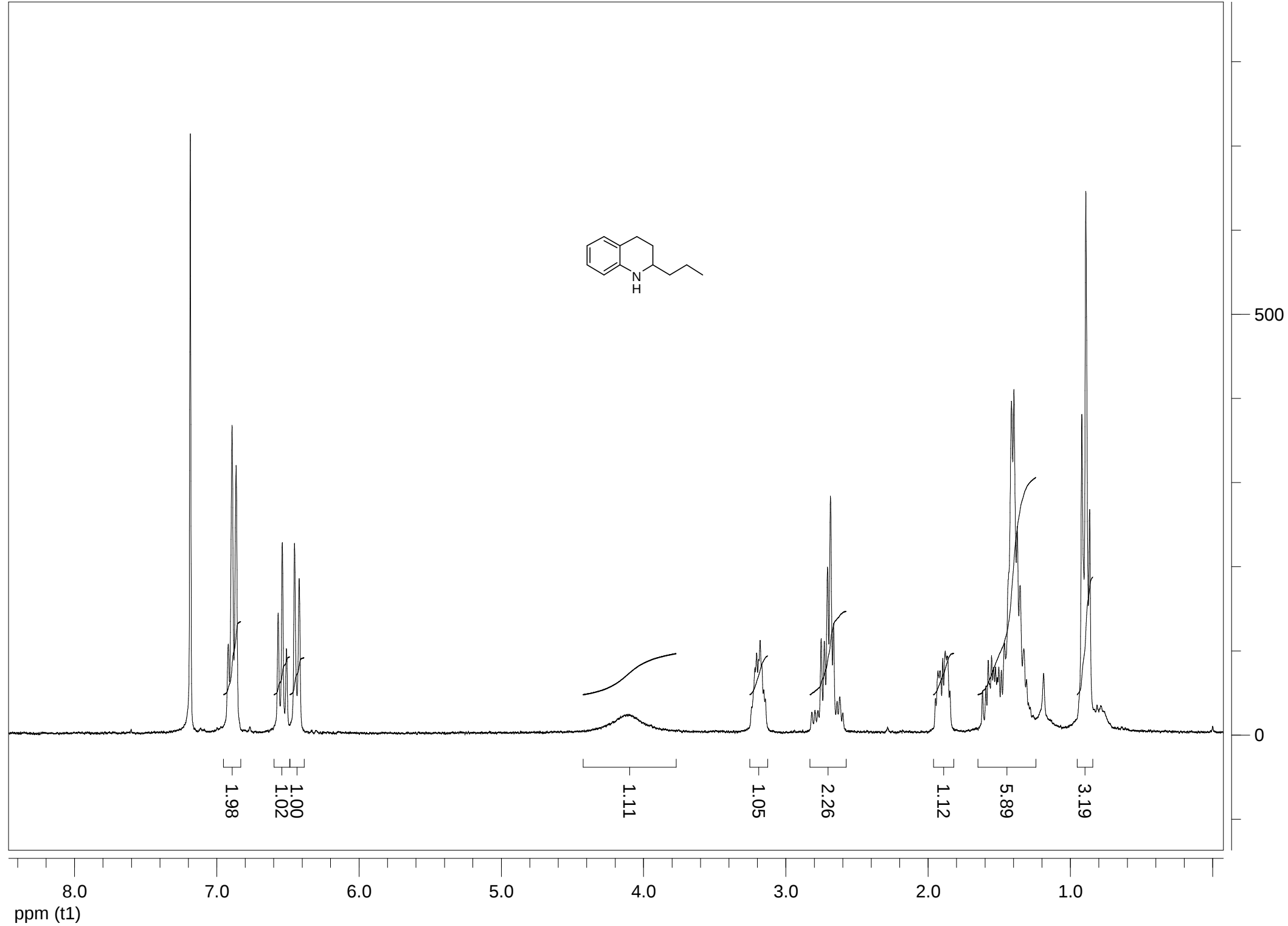
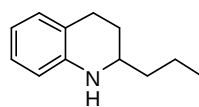
Analysis Date & Time : 10.02.2009 16:08:26
 User Name : Admin
 Vial# : 43
 Sample Name : rk540-4
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

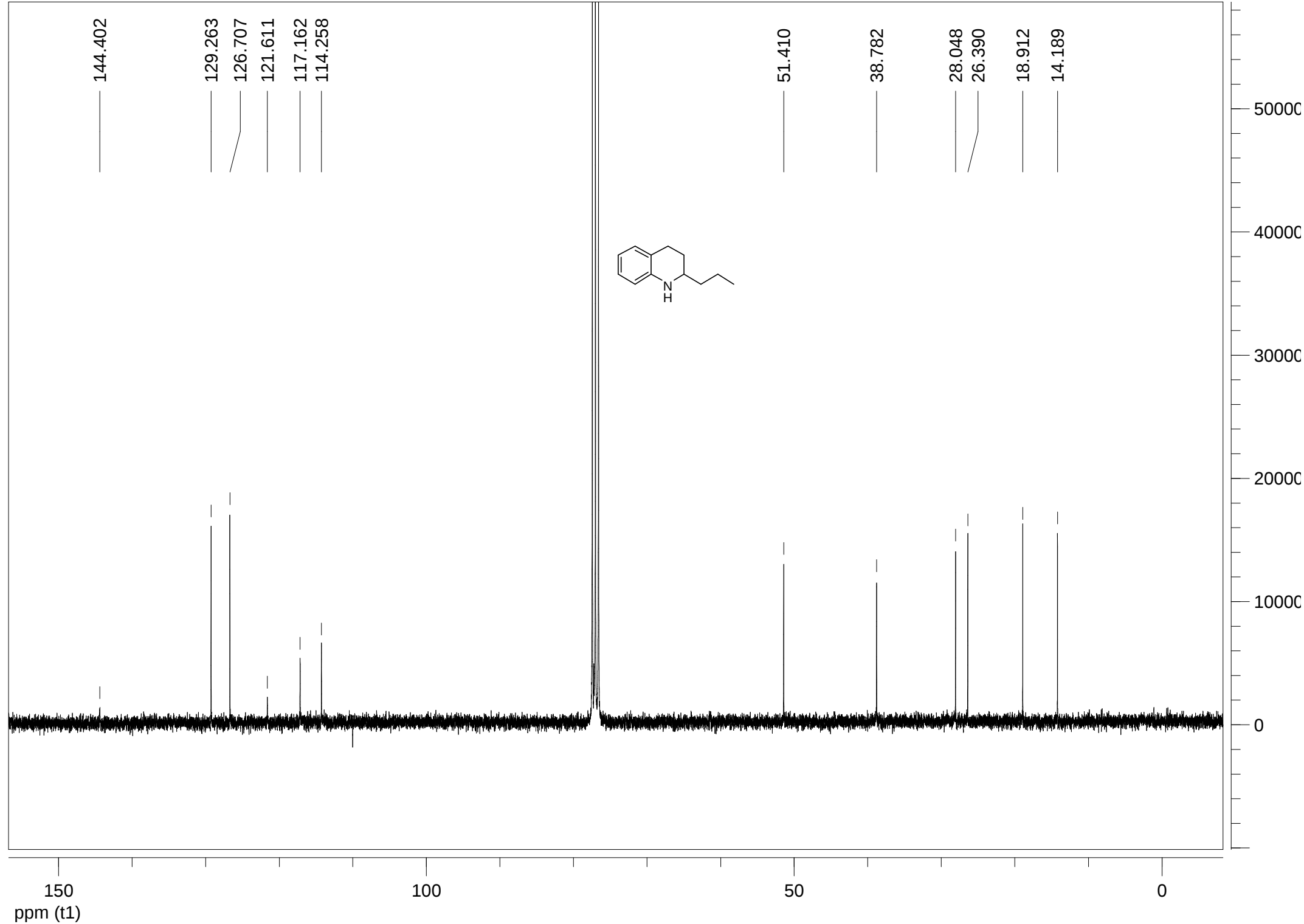
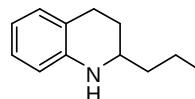


Data Name : D:\DATA_GC\rk\rk540-4.gcd
 Method Name : D:\DATA_GC\rk\quinolines\rk-quinaldine-scope 1.gcm

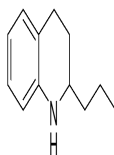


Peak#	Ret.Time	Area	Height	Conc.	Unit Mark	ID#	Cmpd Name
1	9,368	166135	57184	92,222			
2	9,492	14011	4534	7,778	V		
Total		180146	61718				

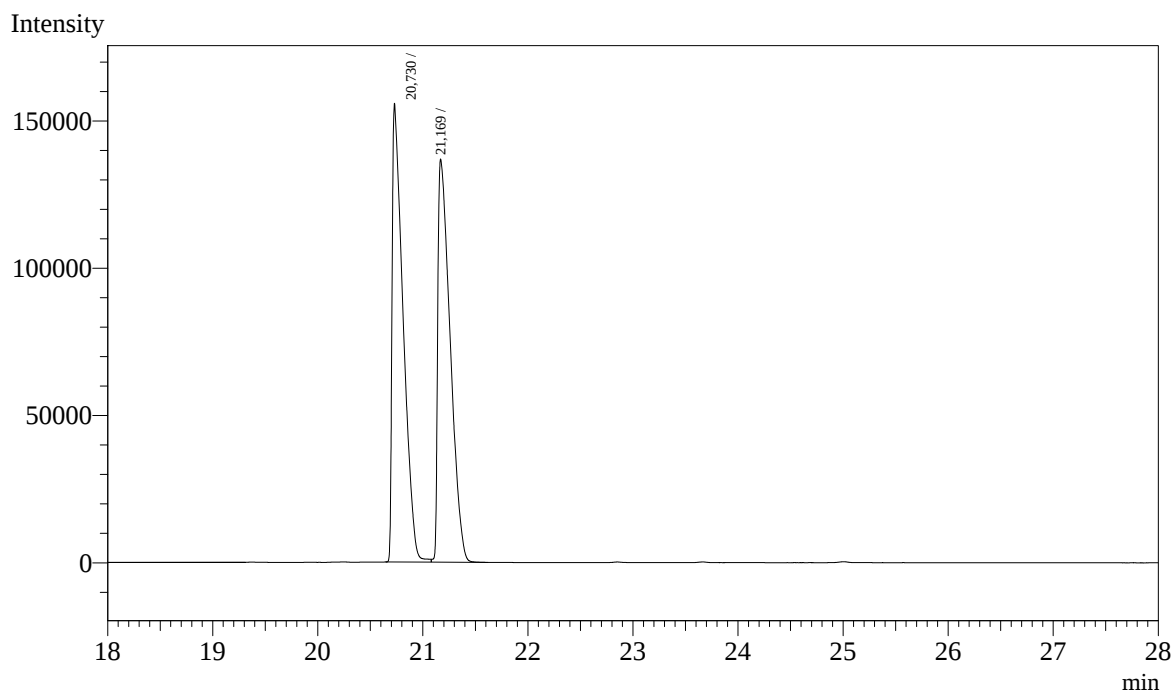




Analysis Date & Time : 17.09.2009 11:11:54
 User Name : Admin
 Vial# : 13
 Sample Name : rk_pr_rac
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

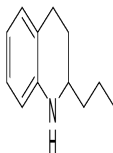


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk_npr_rac.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm

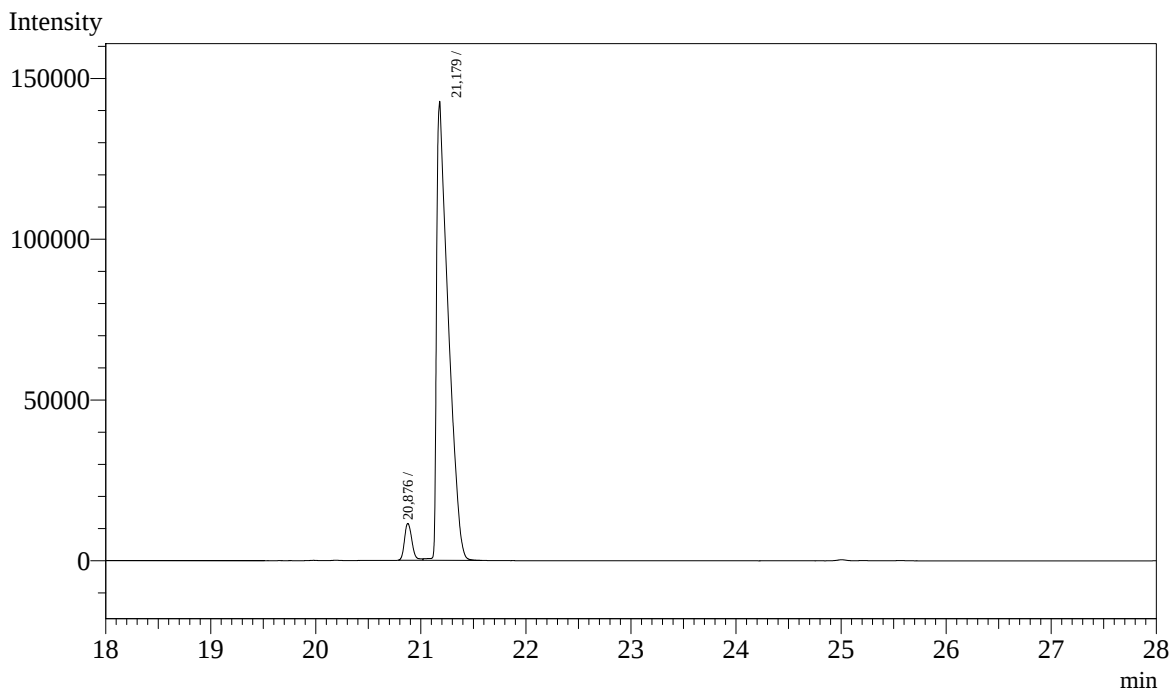


Peak#	Ret.Time	Area	Height	Area%
1	20,730	1138473	155243	50,3049
2	21,169	1124673	136647	49,6951
Total		2263146	291890	100,0000

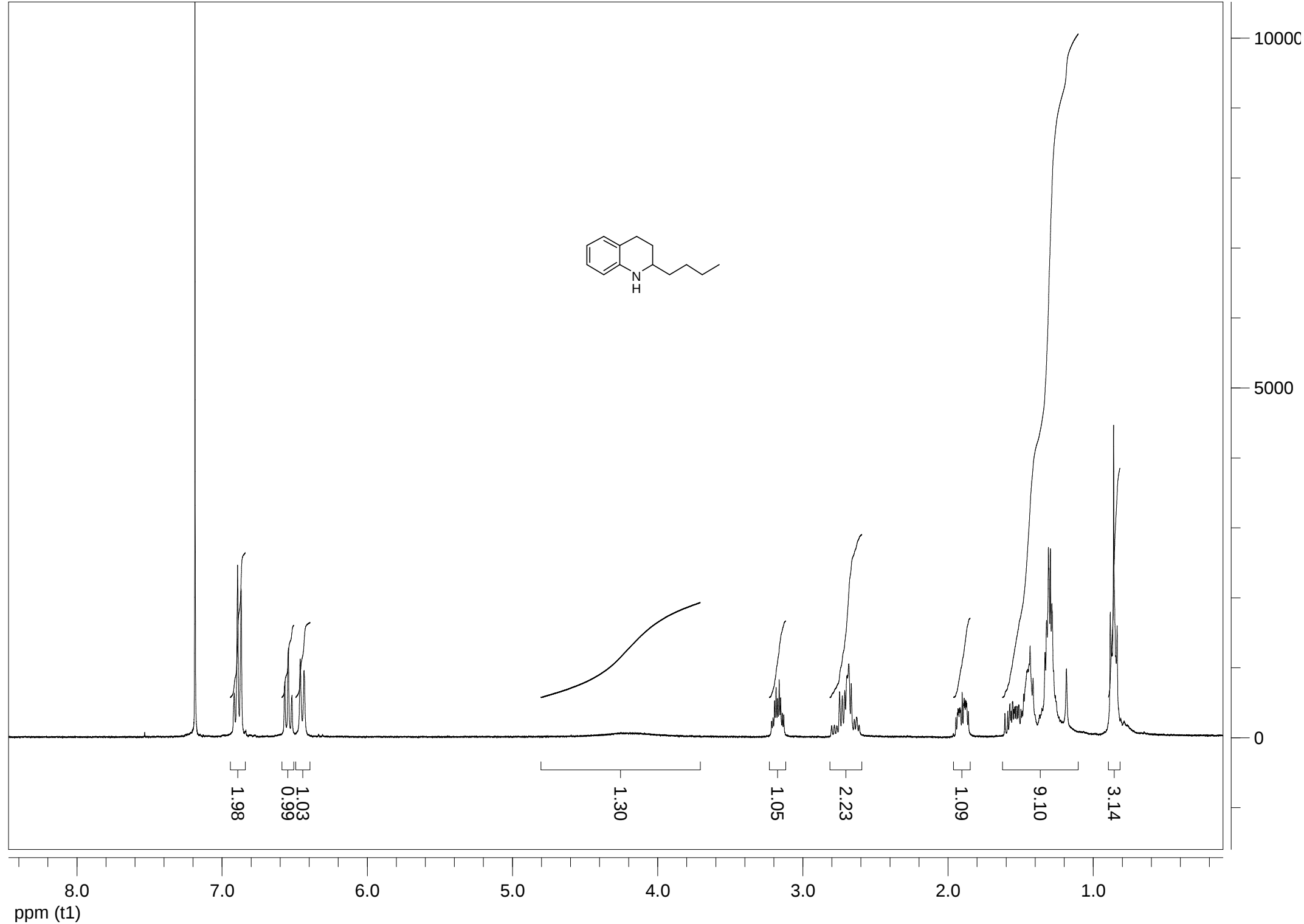
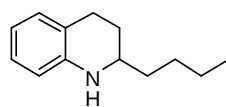
Analysis Date & Time : 17.09.2009 12:40:50
 User Name : Admin
 Vial# : 15
 Sample Name : rk_648_bu_
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

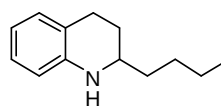


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk_648_nPr_.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm



Peak#	Ret.Time	Area	Height	Area%
1	20,876	56286	11456	4,8849
2	21,179	1095968	142648	95,1151
Total		1152254	154104	100,0000





129.268
126.716
121.639
117.179
114.273

51.689
36.297
28.041
27.923
26.387
22.830
14.077

20000

15000

10000

5000

0

200

150

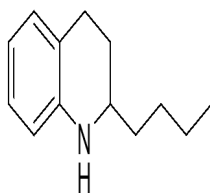
100

50

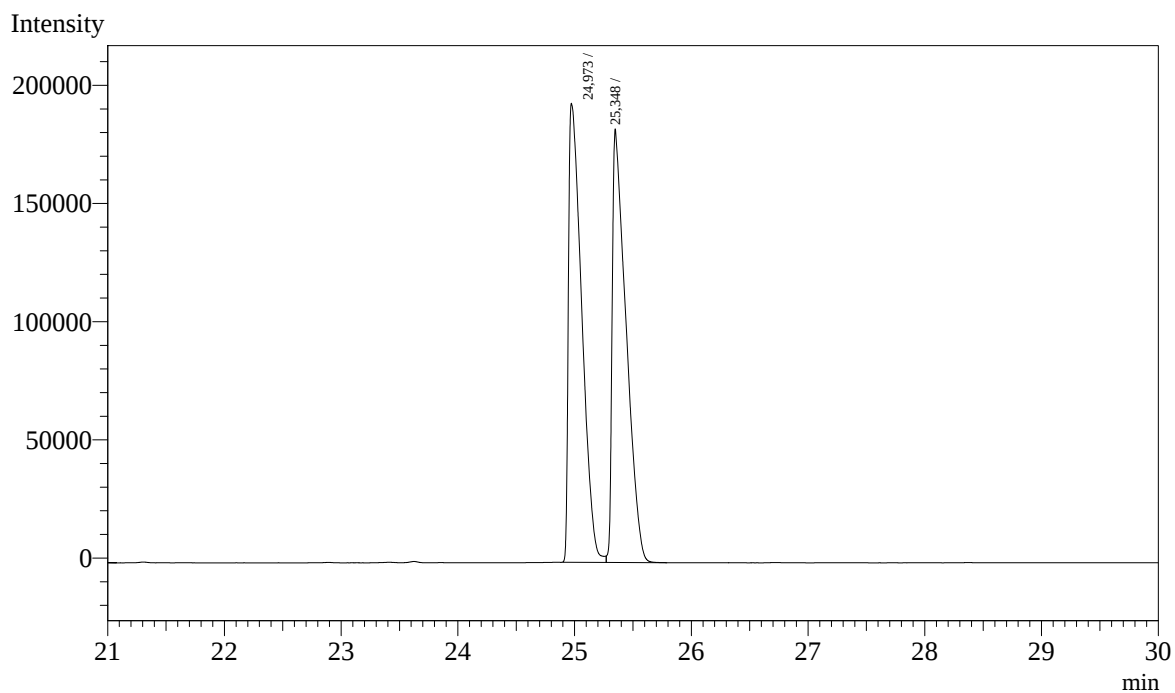
0

ppm (t1)

Analysis Date & Time : 22.09.2009 11:34:48
 User Name : Admin
 Vial# : 31
 Sample Name : rk_nBu_rac
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

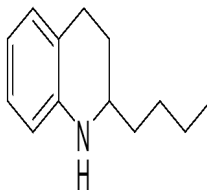


Data Name : F:\data shimadzu chiral gc\rk\rk_nBu_rac-dil.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm

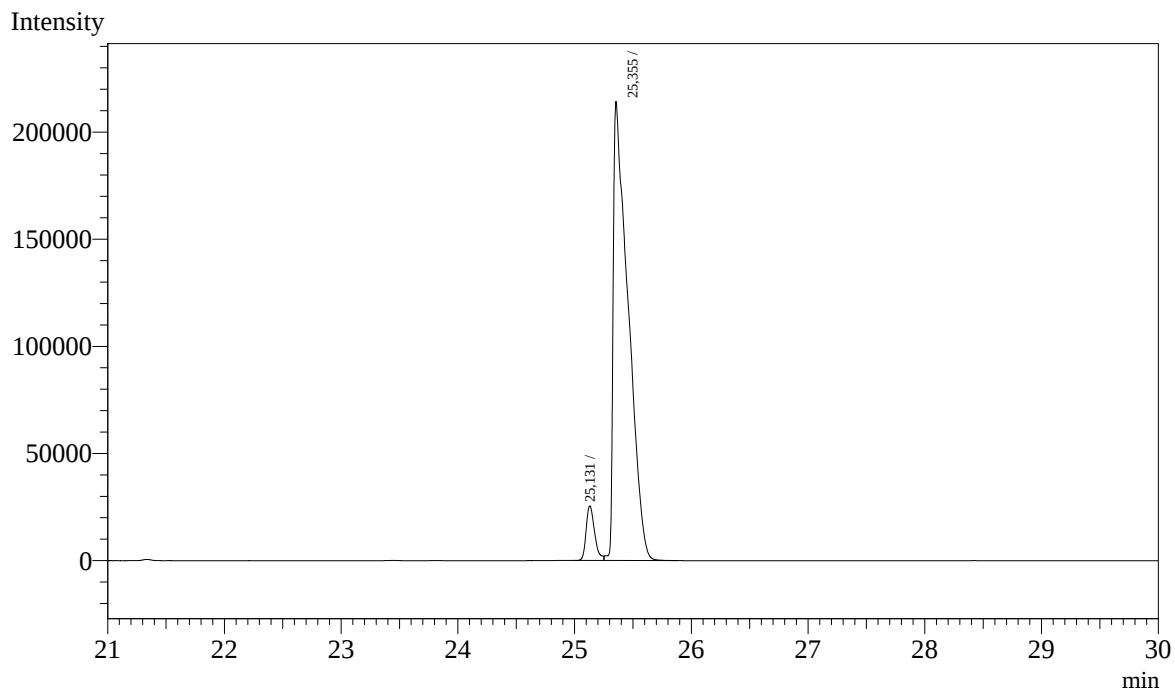


Peak#	Ret.Time	Area	Height	Area%
1	24,973	1540361	193895	50,3401
2	25,348	1519547	181681	49,6599
Total		3059908	375576	100,0000

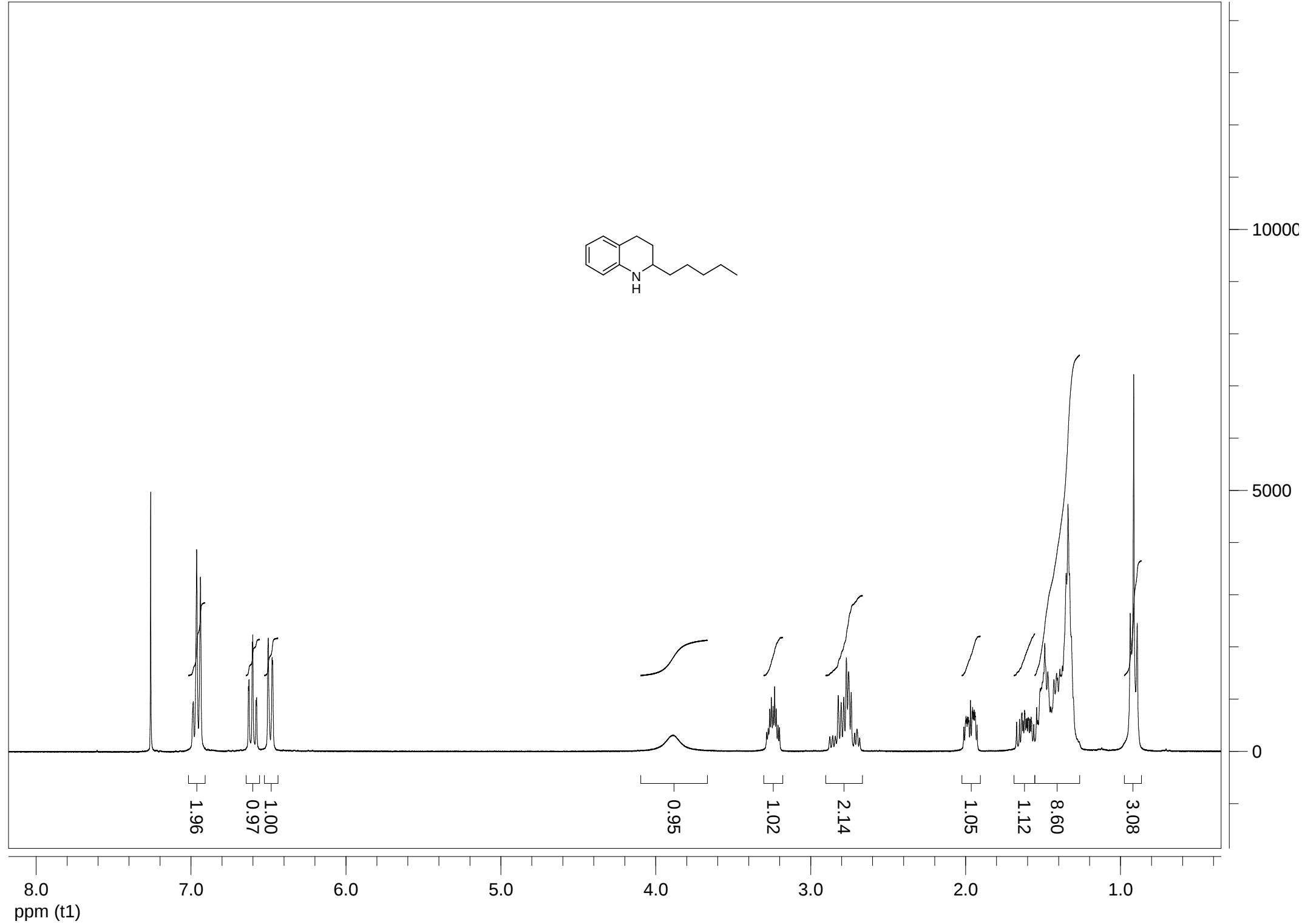
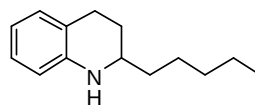
Analysis Date & Time : 17.09.2009 15:17:30
 User Name : Admin
 Vial# : 16
 Sample Name : rk_648_nBu_dil
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

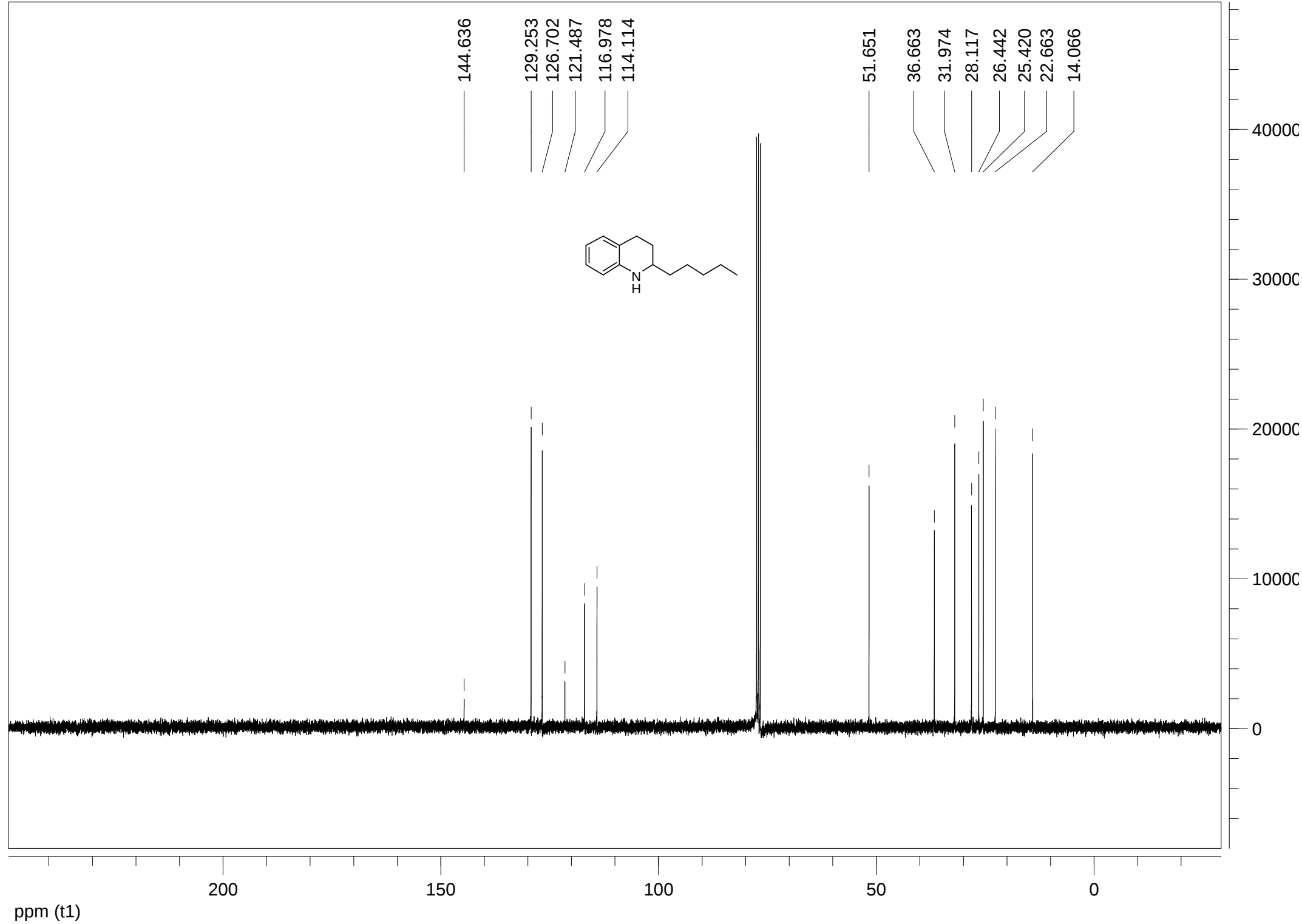


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk_648_nBu_dil.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm

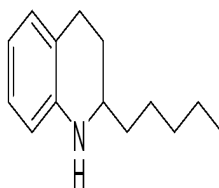


Peak#	Ret.Time	Area	Height	Area%
1	25,131	128615	25400	6,3042
2	25,355	1911542	213967	93,6958
Total		2040157	239367	100,0000

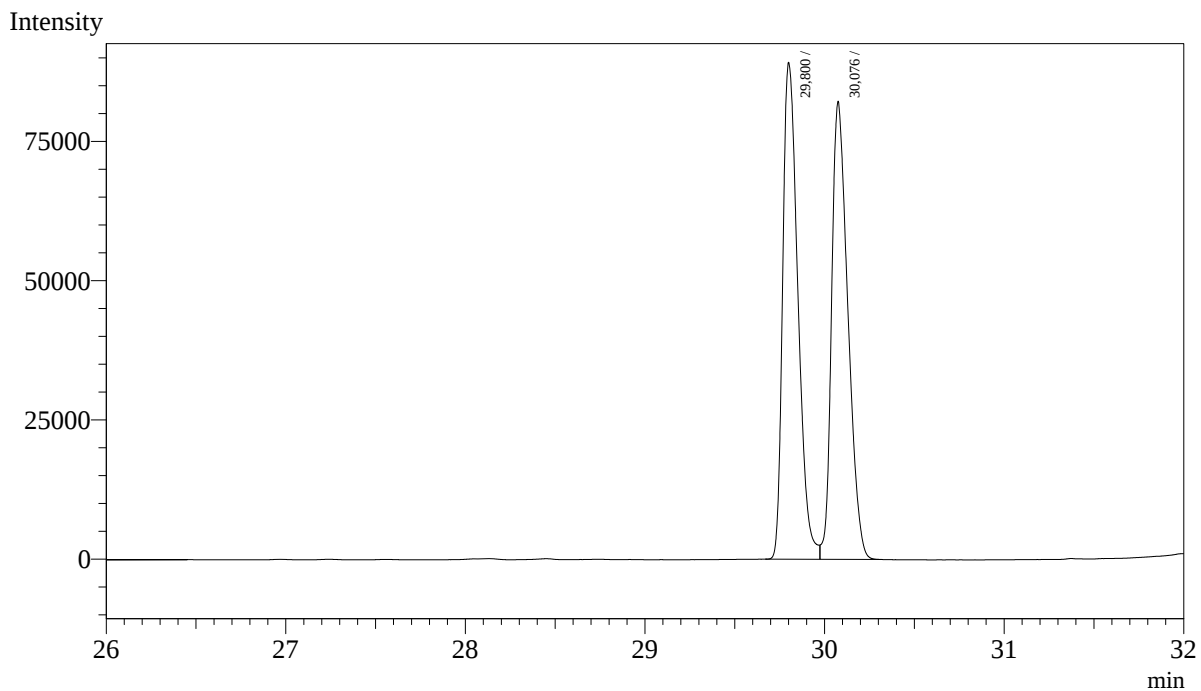




Analysis Date & Time : 17.09.2009 09:42:56
 User Name : Admin
 Vial# : 11
 Sample Name : rk_pent_rac
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

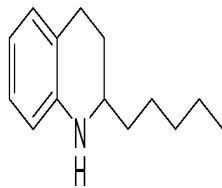


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk_npent_rac.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm

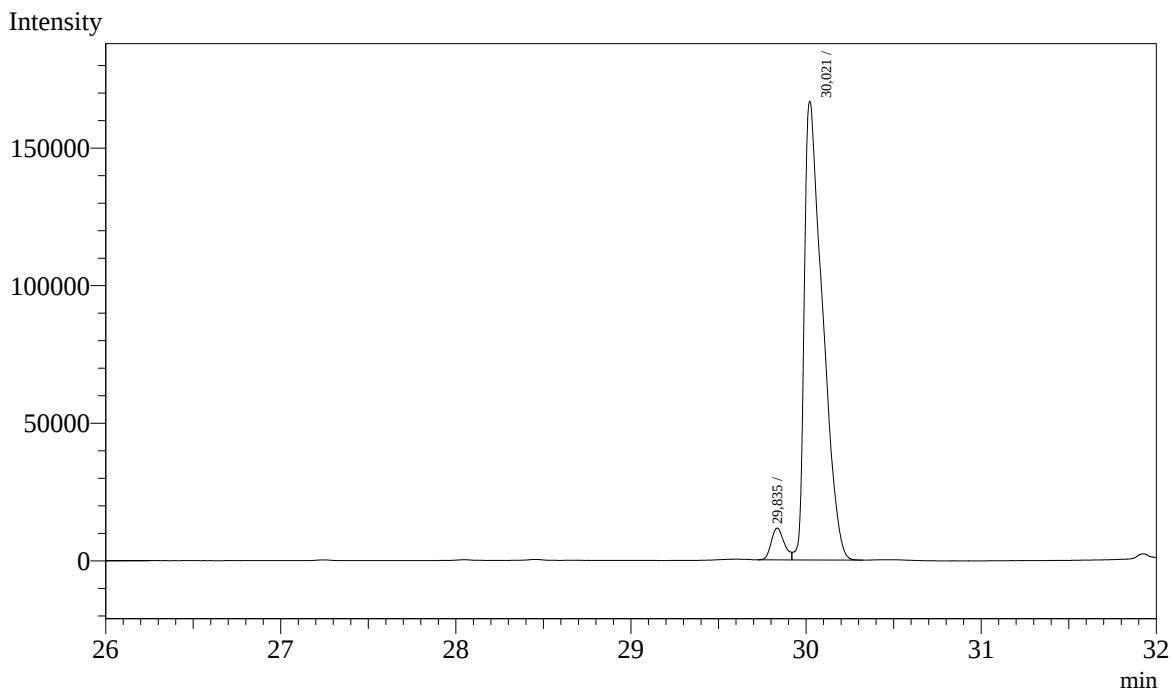


Peak#	Ret.Time	Area	Height	Area%
1	29,800	526644	89181	50,3596
2	30,076	519123	81643	49,6404
Total		1045767	170824	100,0000

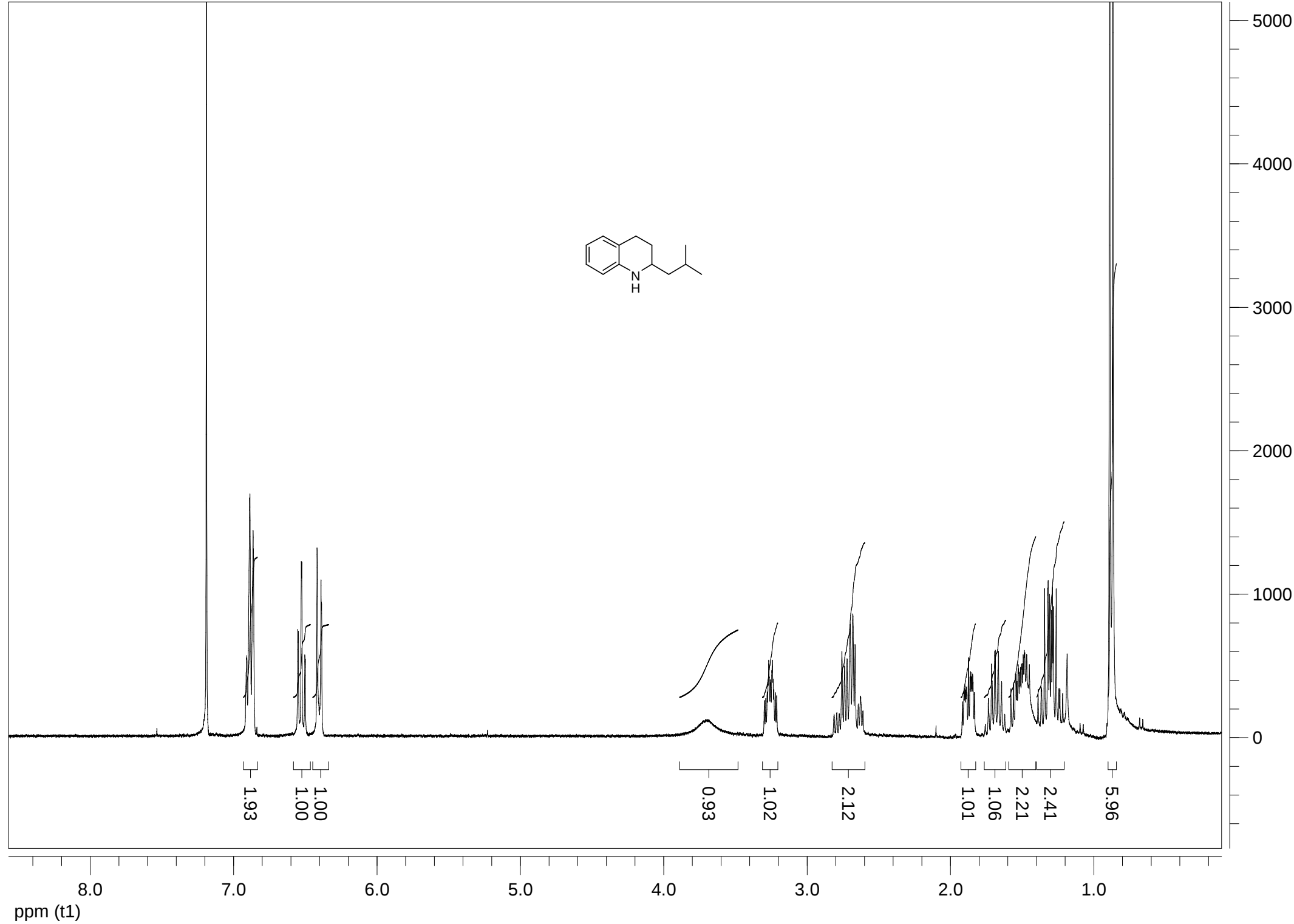
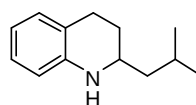
Analysis Date & Time : 17.09.2009 10:27:28
 User Name : Admin
 Vial# : 12
 Sample Name : rk_648_nPent_c
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

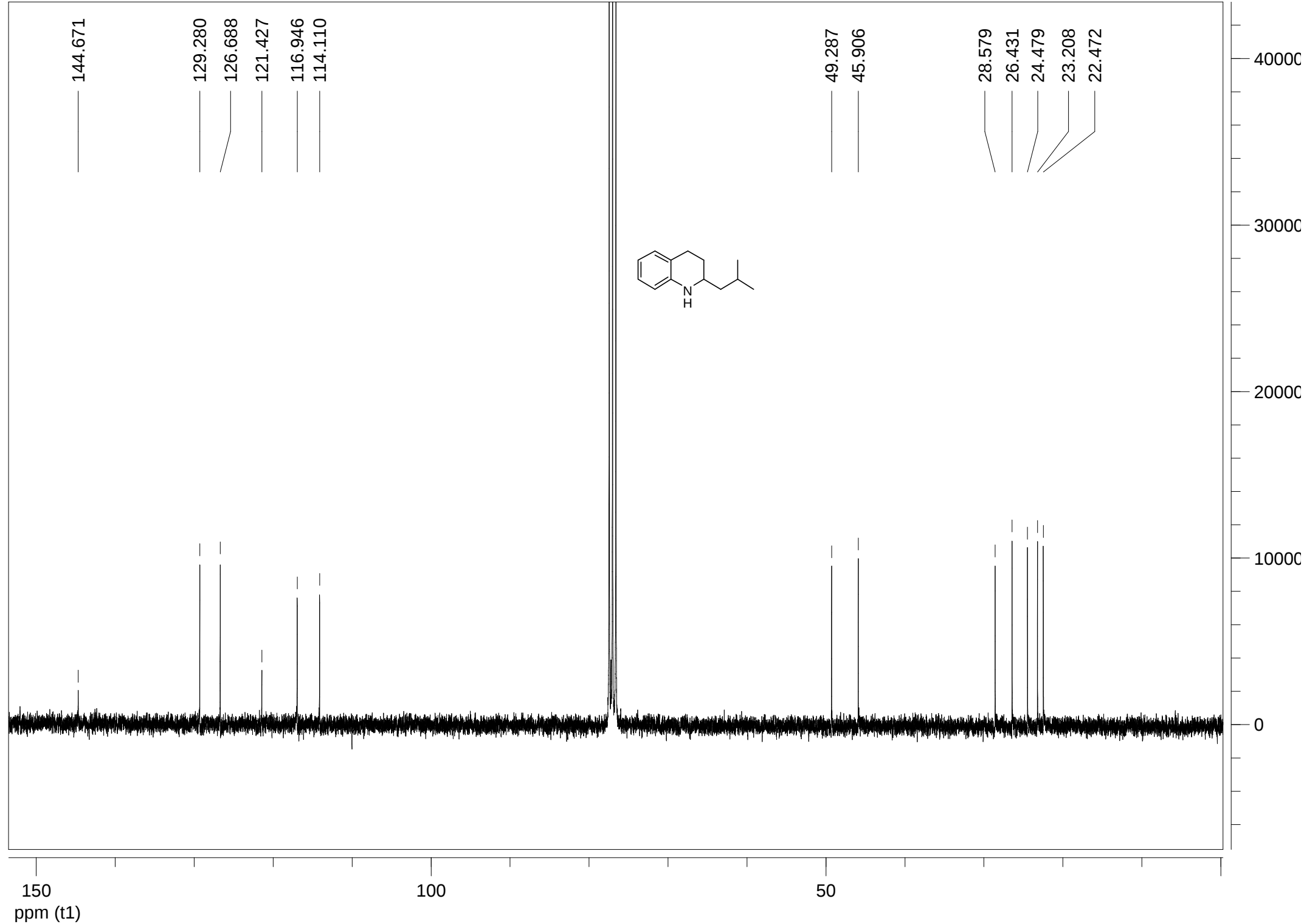
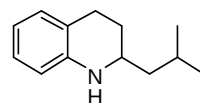


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk648_nPent_c.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6cl.gcm

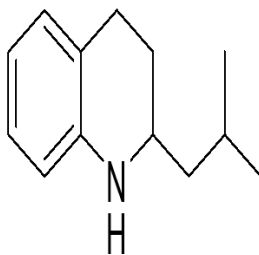


Peak#	Ret.Time	Area	Height	Area%
1	29,835	59575	11542	4,6893
2	30,021	1210859	166443	95,3107
Total		1270434	177985	100,0000

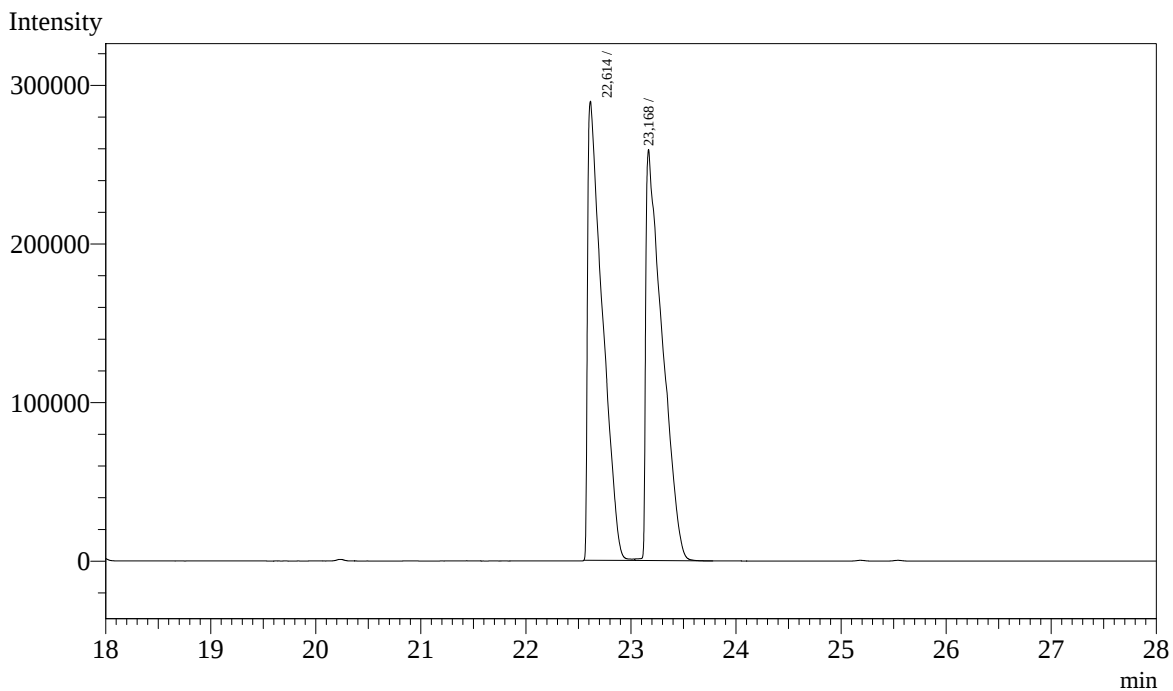




Analysis Date & Time : 20.09.2009 22:38:52
 User Name : Admin
 Vial# : 28
 Sample Name : rk_iBu_rac
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

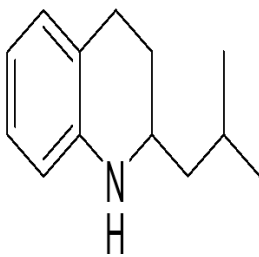


Data Name : F:\data shimadzu chiral gc\rk\quinoline project ac\rk_iBu_rac.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6br.gcm

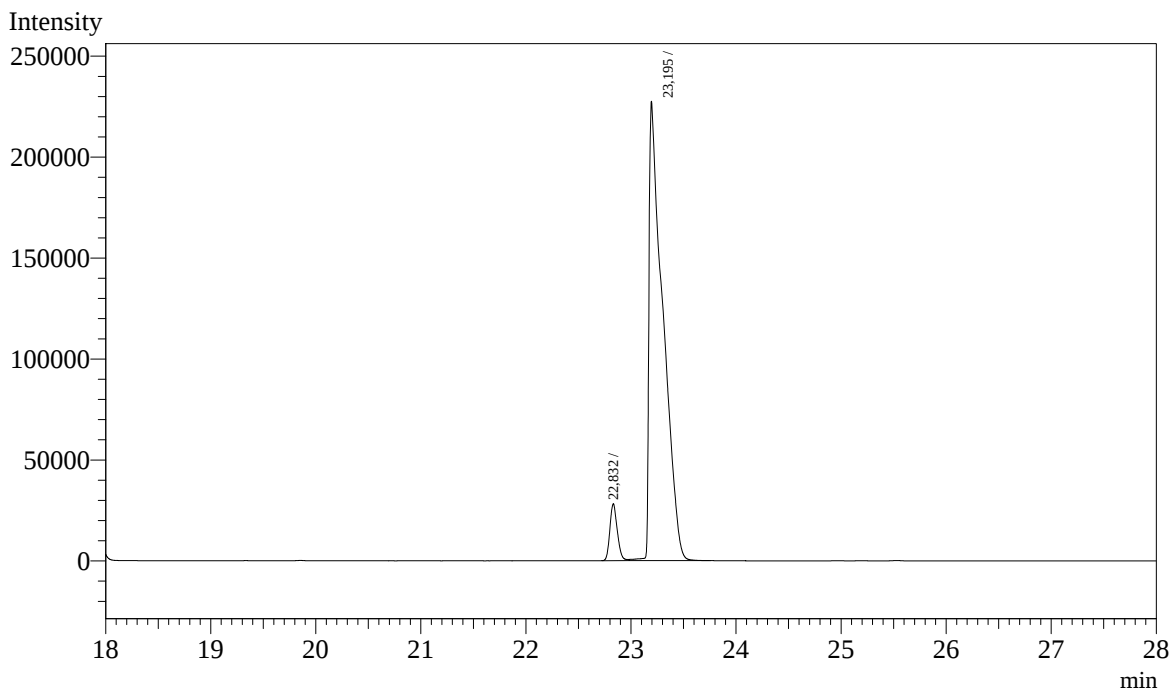


Peak#	Ret.Time	Area	Height	Area%
1	22,614	2821224	288566	50,0481
2	23,168	2815806	259063	49,9519
Total		5637030	547629	100,0000

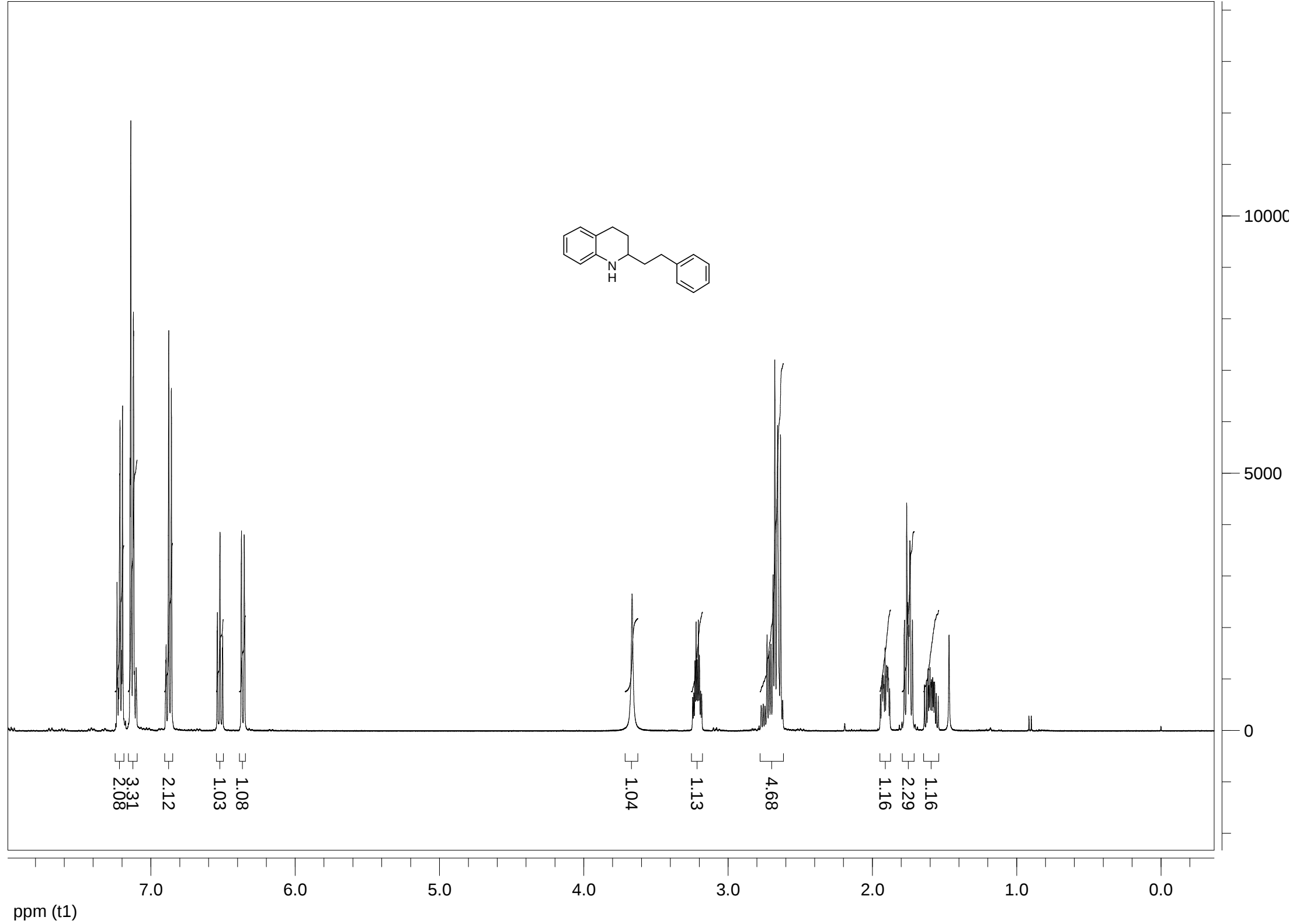
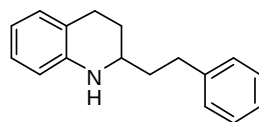
Analysis Date & Time : 23.09.2009 14:50:54
 User Name : Admin
 Vial# : 30
 Sample Name : rk651_iBu_dil
 Sample ID :
 Sample Type : Unknown
 Injection Volume : 1,00
 ISTD Amount :

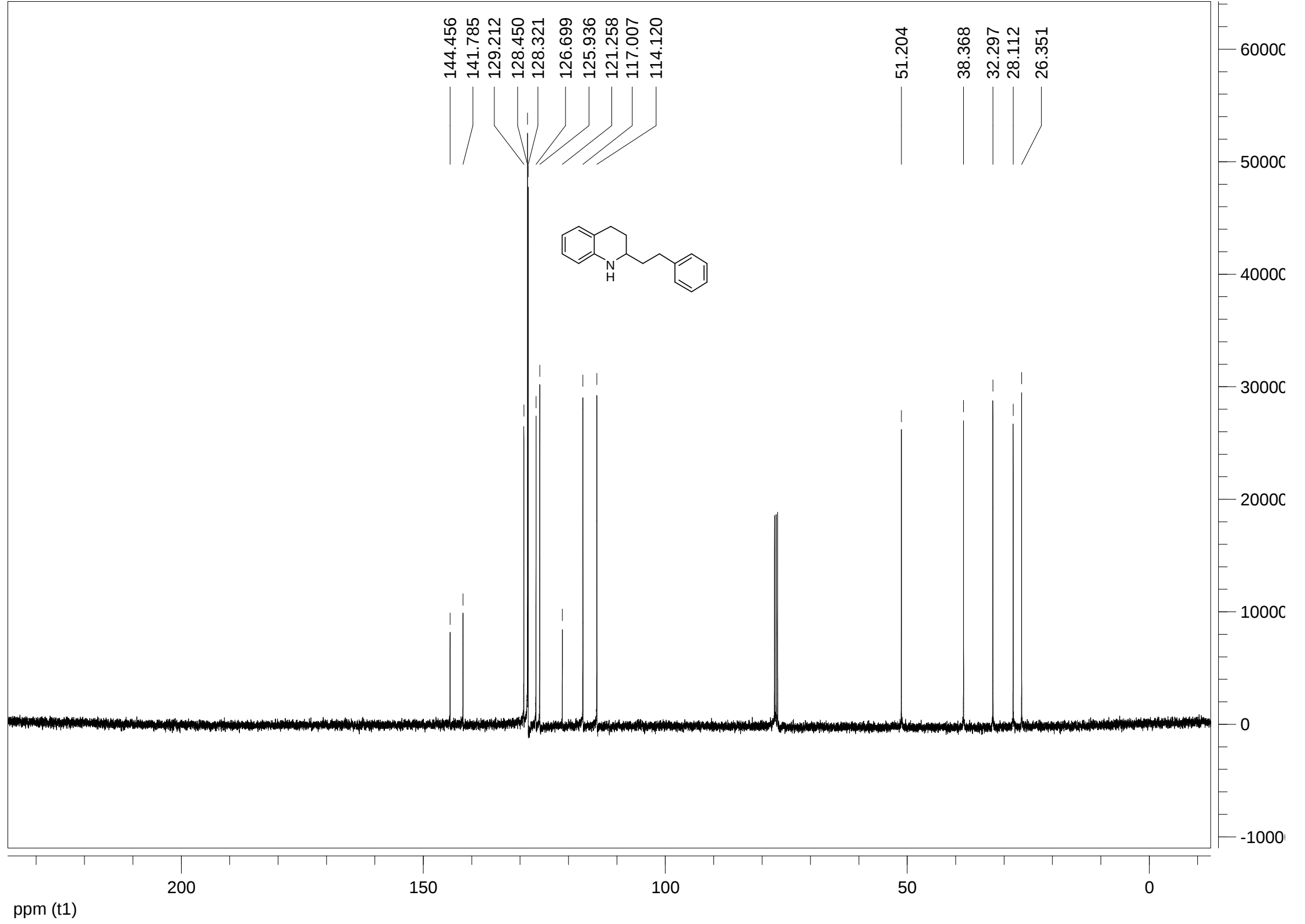


Data Name : F:\data shimadzu chiral gc\rk\rk651_iBu_dil.gcd
 Method Name : F:\data shimadzu chiral gc\rk\quinaldine_6br.gcm



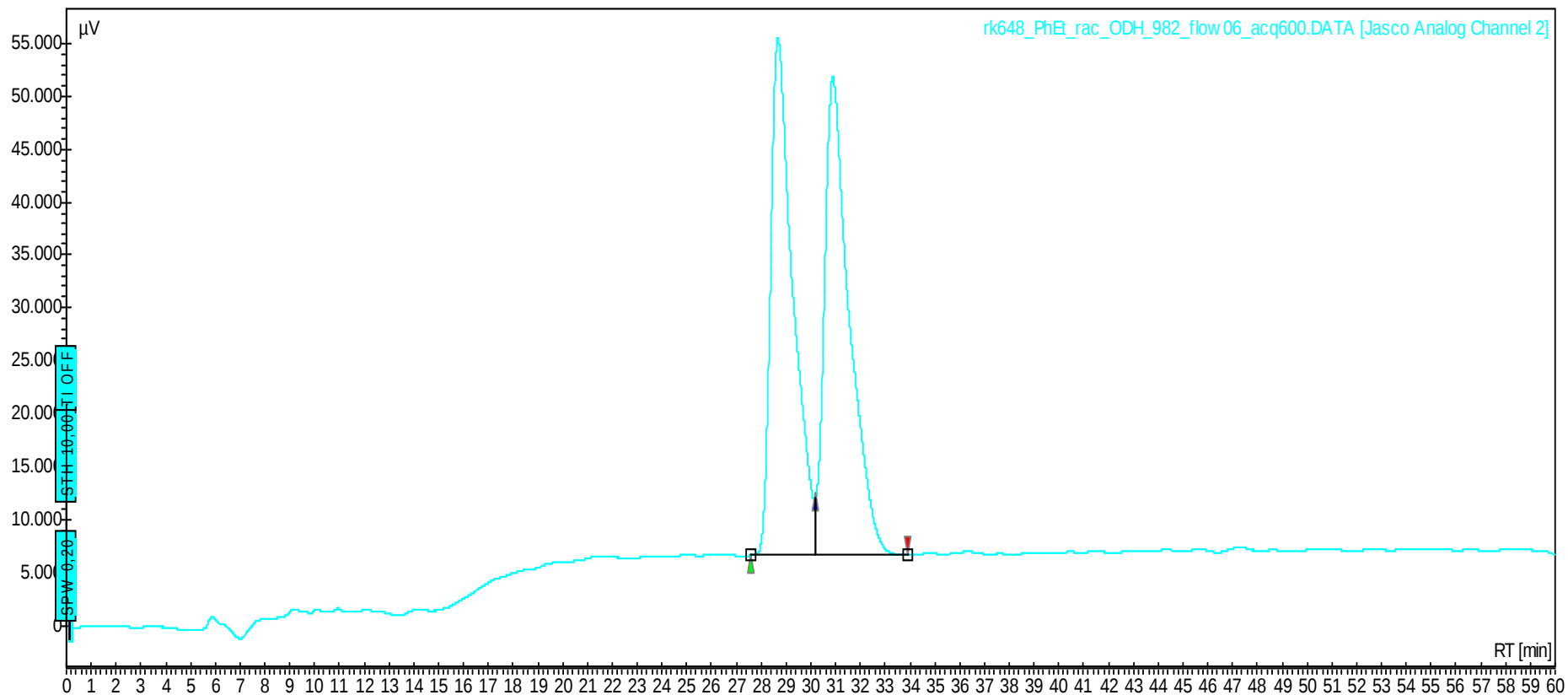
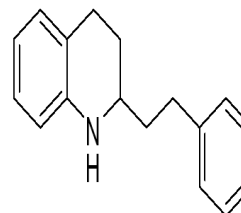
Peak#	Ret.Time	Area	Height	Area%
1	22,832	138433	28222	6,1960
2	23,195	2095801	227093	93,8040
Total		2234234	255315	100,0000





Chromatogram : rk648_PhEt_rac_ODH_982_flow06_acq600

Data file: rk648_PhEt_rac_ODH_982_flow06_acq600.DATA
Method: HPLC2_ODH_982_flow06_acq60
Date: 18.09.2009 10:05:12

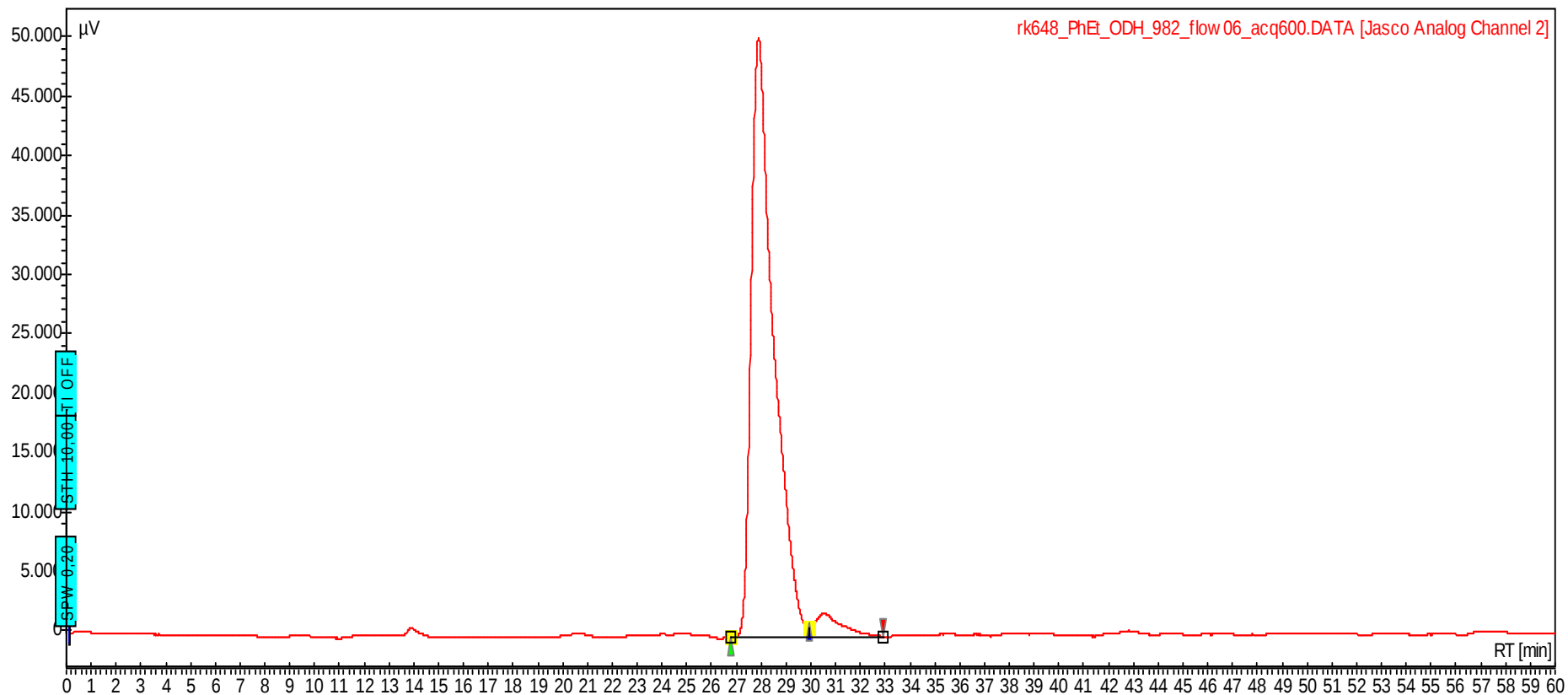
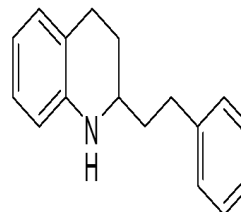


rk648_PhEt_rac_ODH_982_flow06_acq600.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	27,611	28,675	30,204	0,000	49,32	48728,0	50832,8	49,320
2	UNKNOWN	30,204	30,892	33,910	0,000	50,68	45091,2	52235,2	50,680
Total						100,00	93819,2	103068,1	100,000

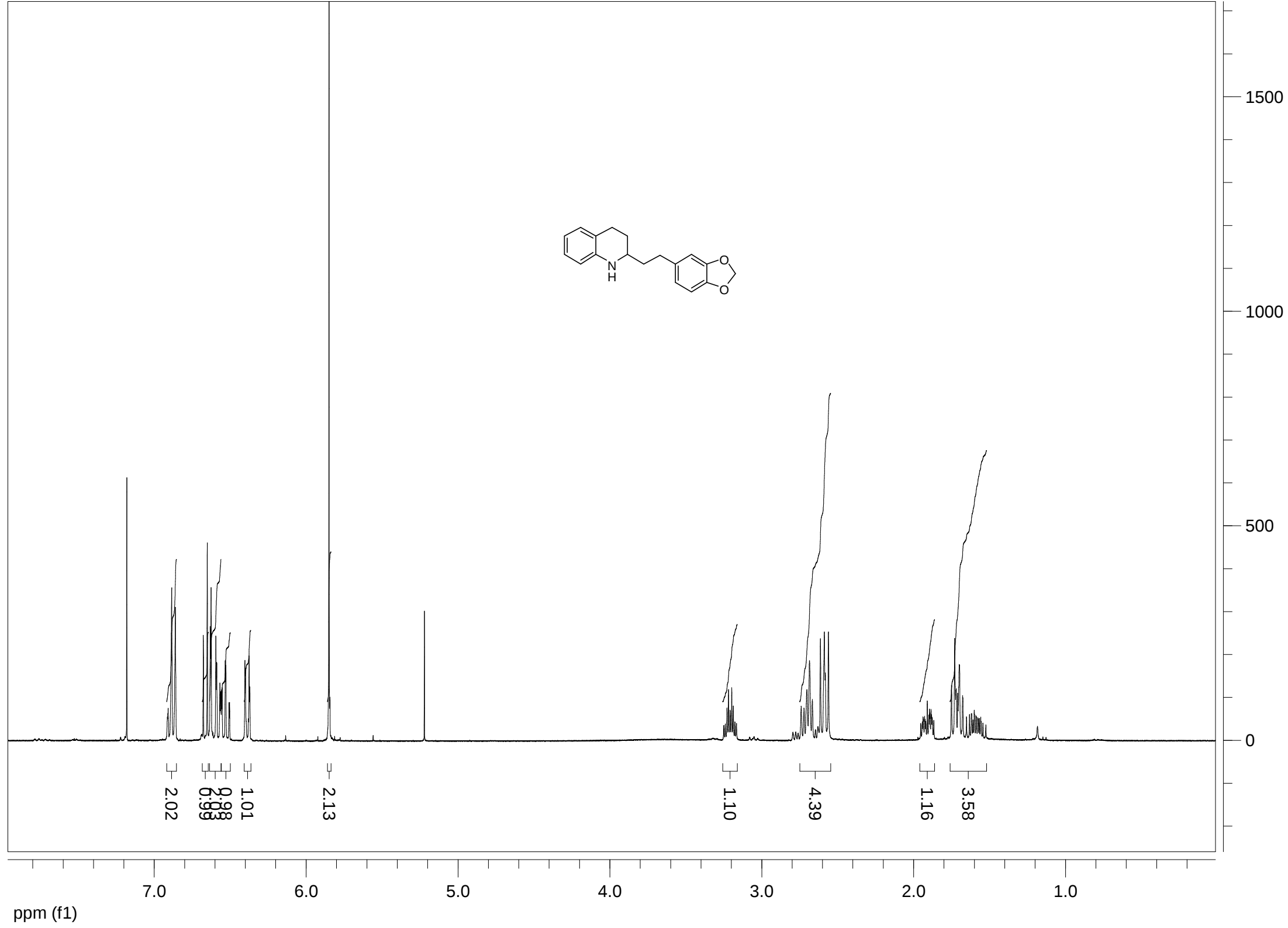
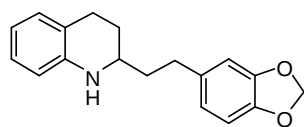
Chromatogram : rk648_PhEt_ODH_982_flow06_acq600

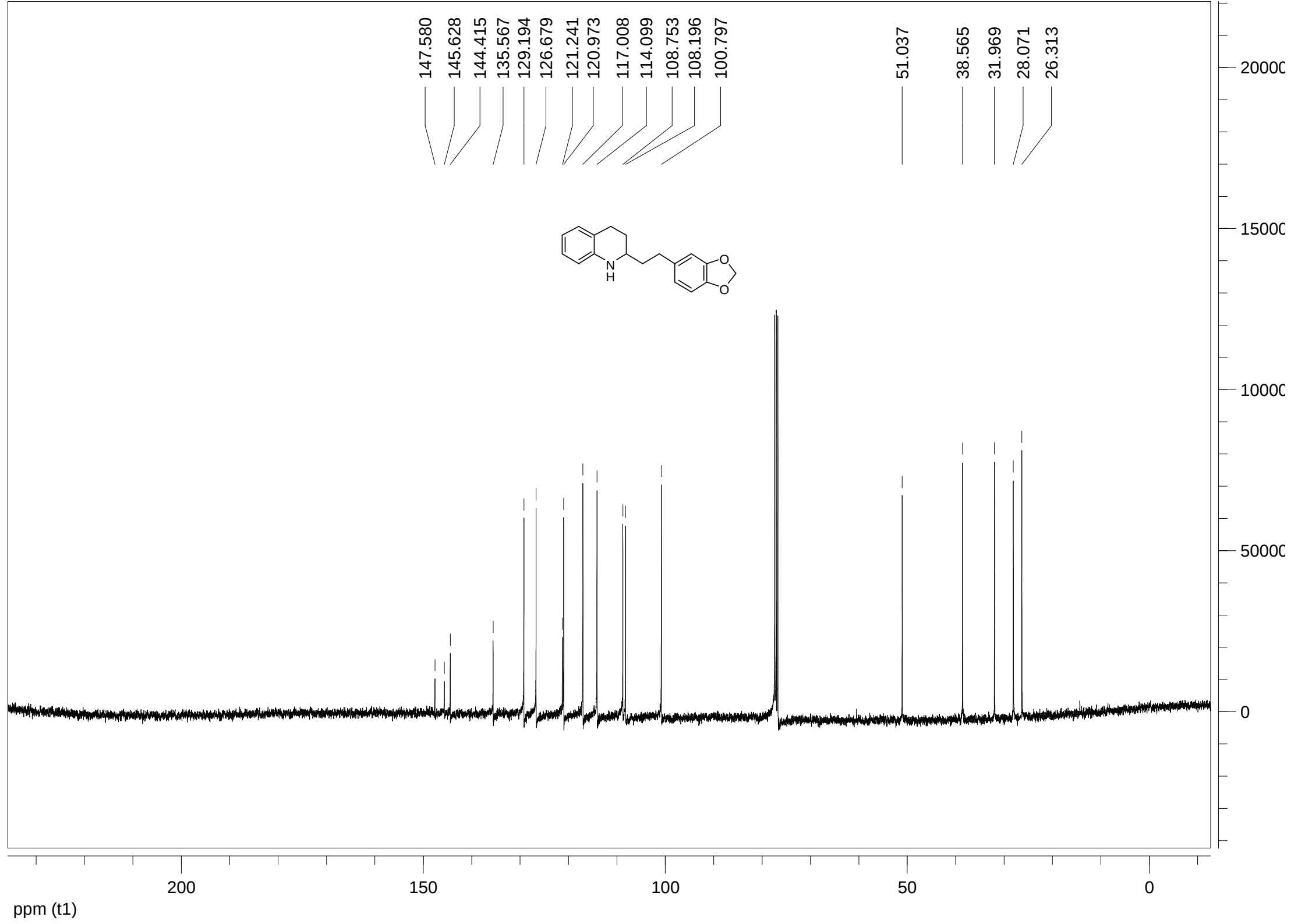
Data file: rk648_PhEt_ODH_982_flow06_acq600.DATA
Method: HPLC2_ODH_982_flow06_acq60
Date: 18.09.2009 11:07:51



rk648_PhEt_ODH_982_flow06_acq600.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[μV]	[$\mu\text{V}\cdot\text{Min}$]	[%]
1	UNKNOWN	26,800	27,892	29,952	0,000	95,55	50318,5	52754,0	95,551
2	UNKNOWN	29,952	30,517	32,921	0,000	4,45	1930,5	2456,1	4,449
Total						100,00	52249,0	55210,1	100,000



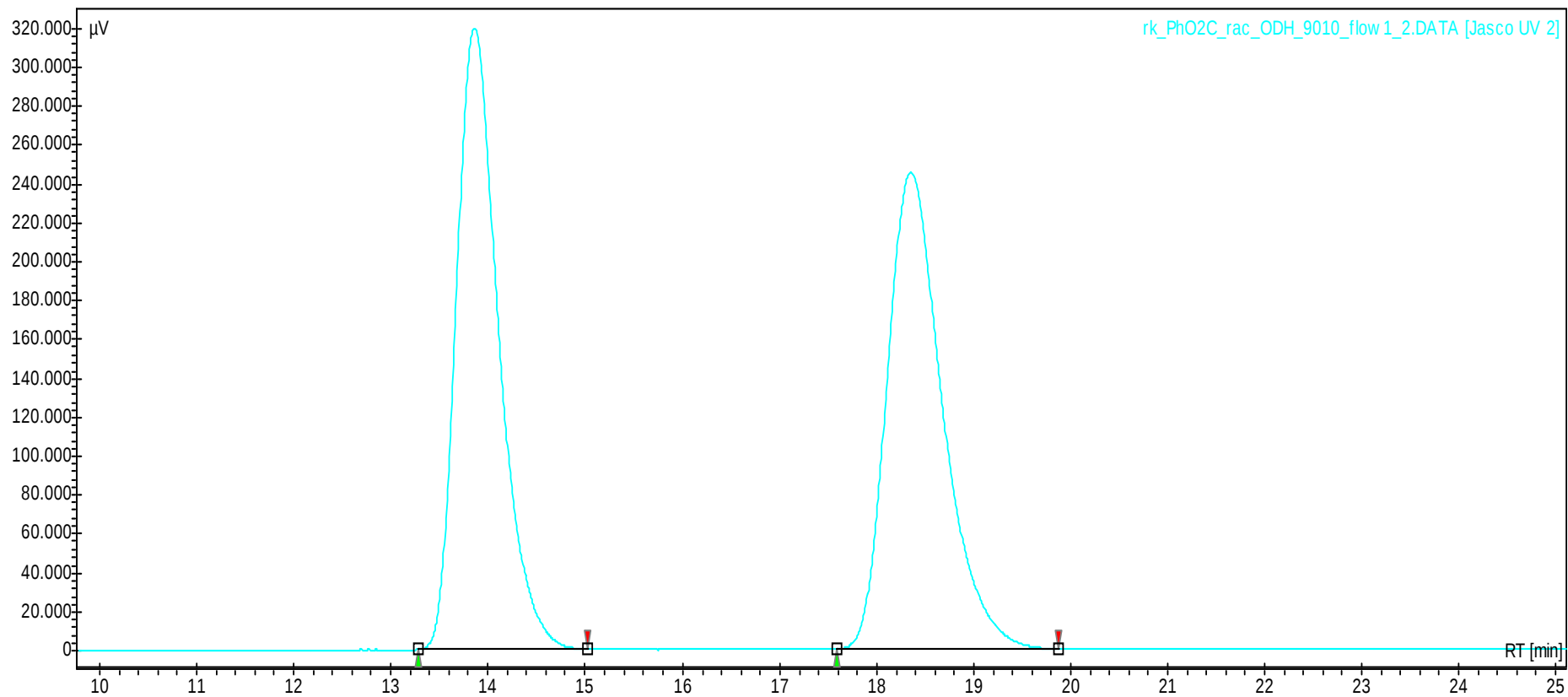
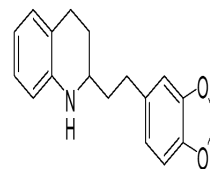


Chromatogram : rk_PhO2C_rac_ODH_9010_flow1_2

Data file: rk_PhO2C_rac_ODH_9010_flow1_2.DATA

Method: HPLC1_ODH_9010_flow1_acq_30

Date: 22.09.2009 09:00:43



rk_PhO2C_rac_ODH_9010_flow1_2.DATA [Jasco UV 2]

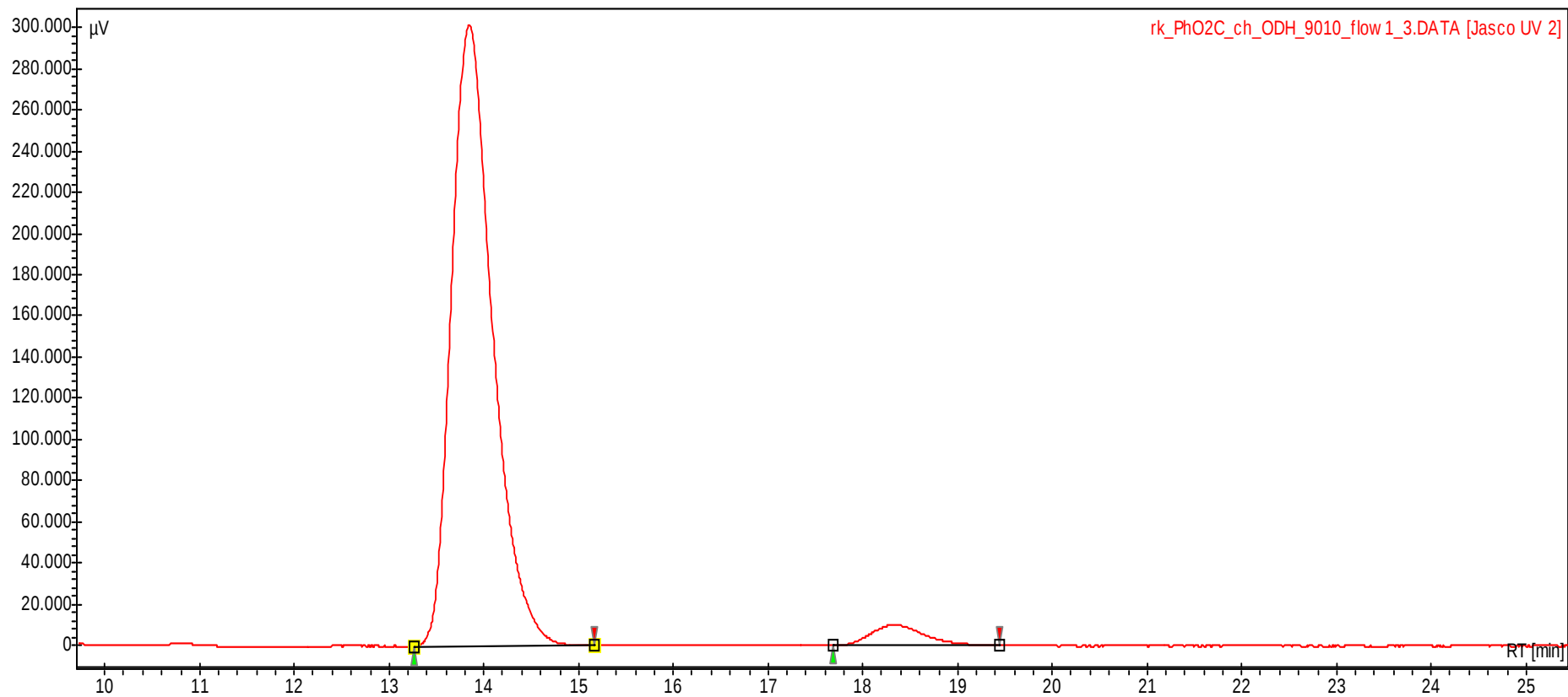
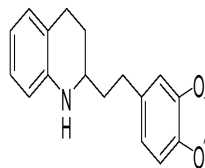
Index	Start	Time	End	Area %
	[Min]	[Min]	[Min]	[%]
1	13,274	13,858	15,018	49,878
2	17,591	18,350	19,866	50,122
Total				100,000

Chromatogram : rk_PhO2C_ch_ODH_9010_flow1_3

Data file: rk_PhO2C_ch_ODH_9010_flow1_3.DATA

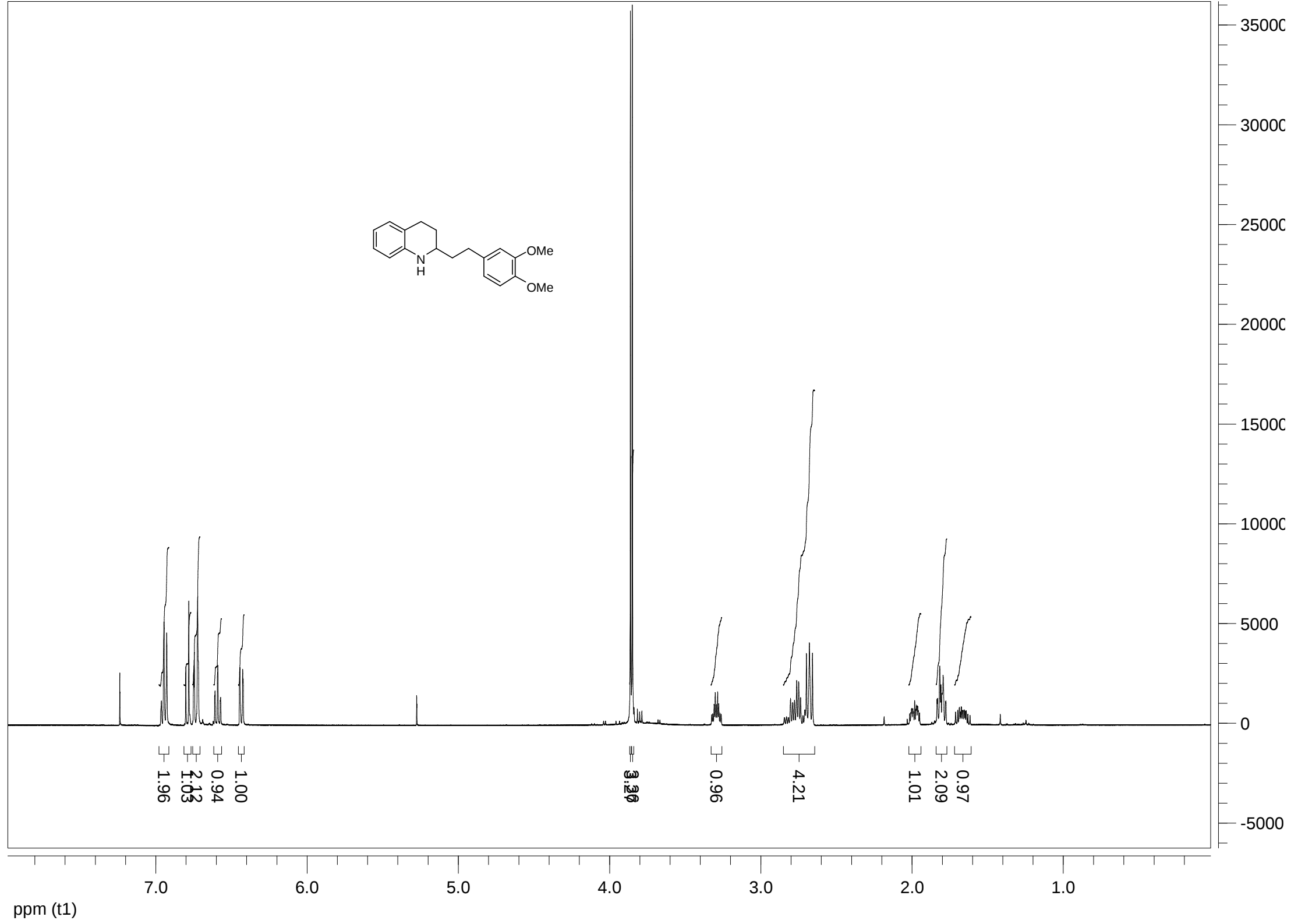
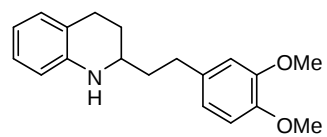
Method: HPLC1_ODH_9010_flow1_acq_30

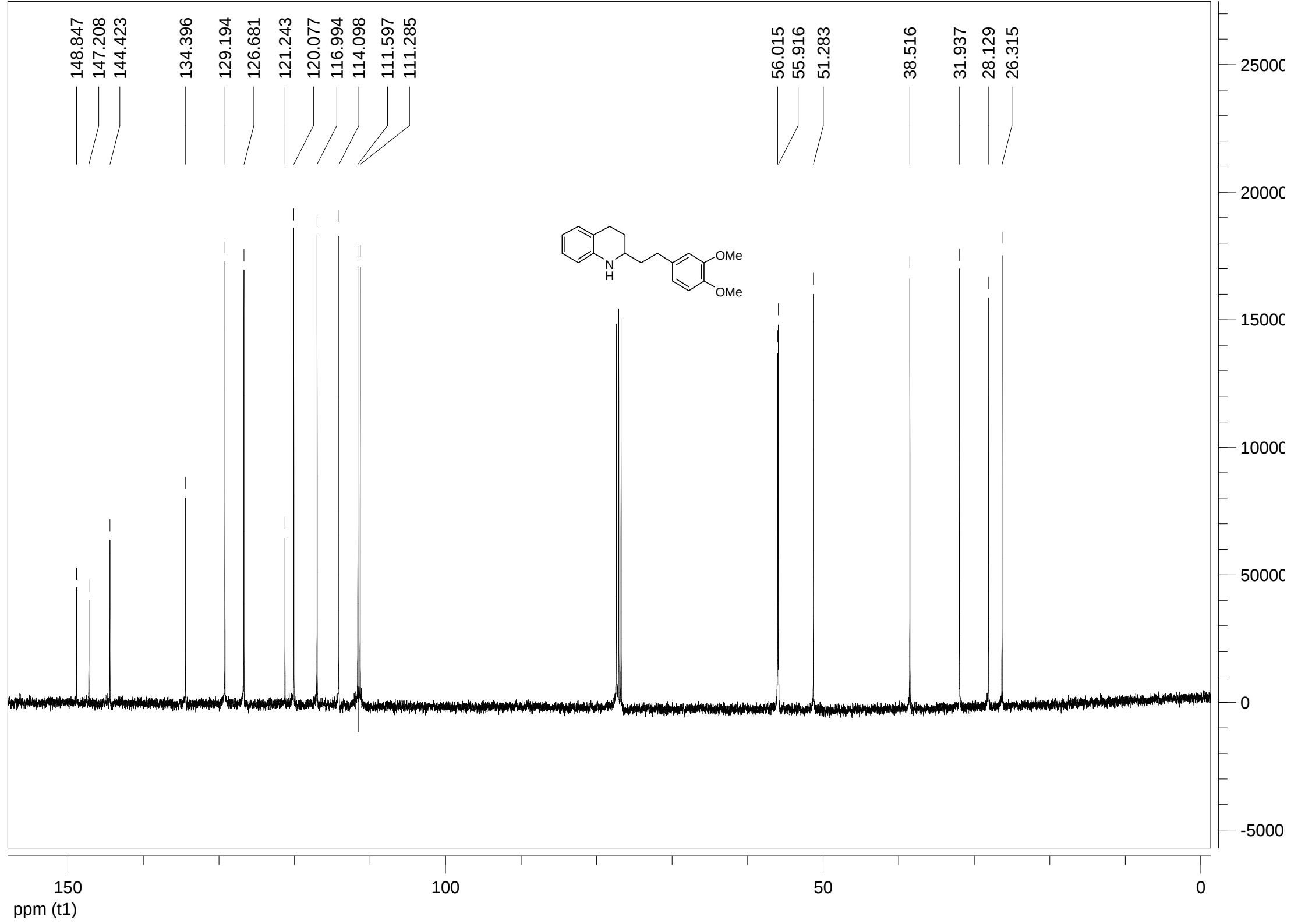
Date: 22.09.2009 09:33:28



rk_PhO2C_ch_ODH_9010_flow1_3.DATA [Jasco UV 2]

Index	Start	Time	End	Area %
	[Min]	[Min]	[Min]	[%]
1	13,257	13,842	15,157	95,814
2	17,691	18,333	19,440	4,186
Total				100,000



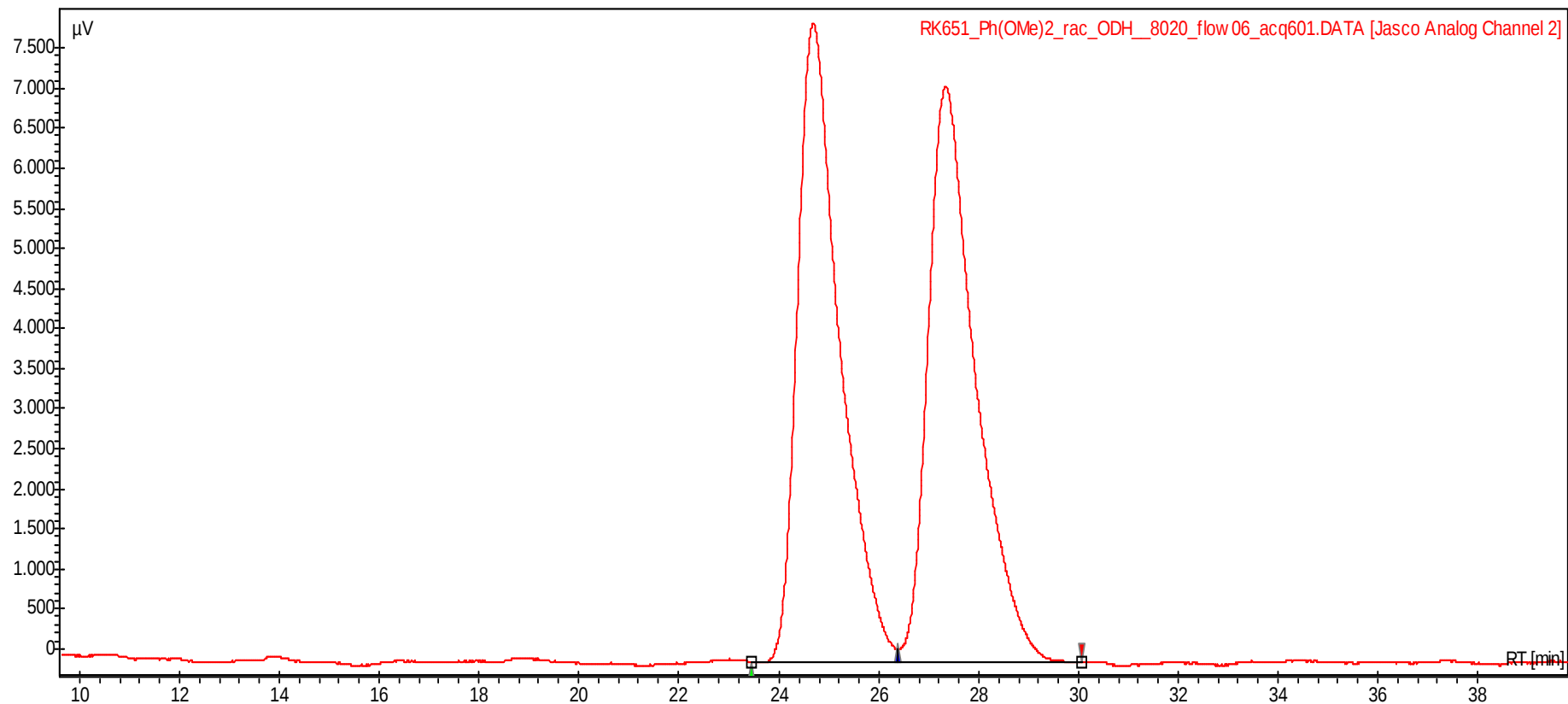
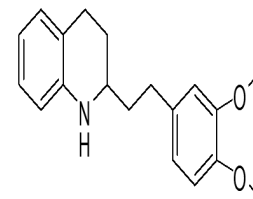


Chromatogram : RK651_Ph(OMe)2_rac_ODH__8020_flow06_acq601

Data file: RK651_Ph(OMe)2_rac_ODH__8020_flow06_acq601.DATA

Method: HPLC2_ODH_8020_flow06_acq60

Date: 19.09.2009 16:06:51

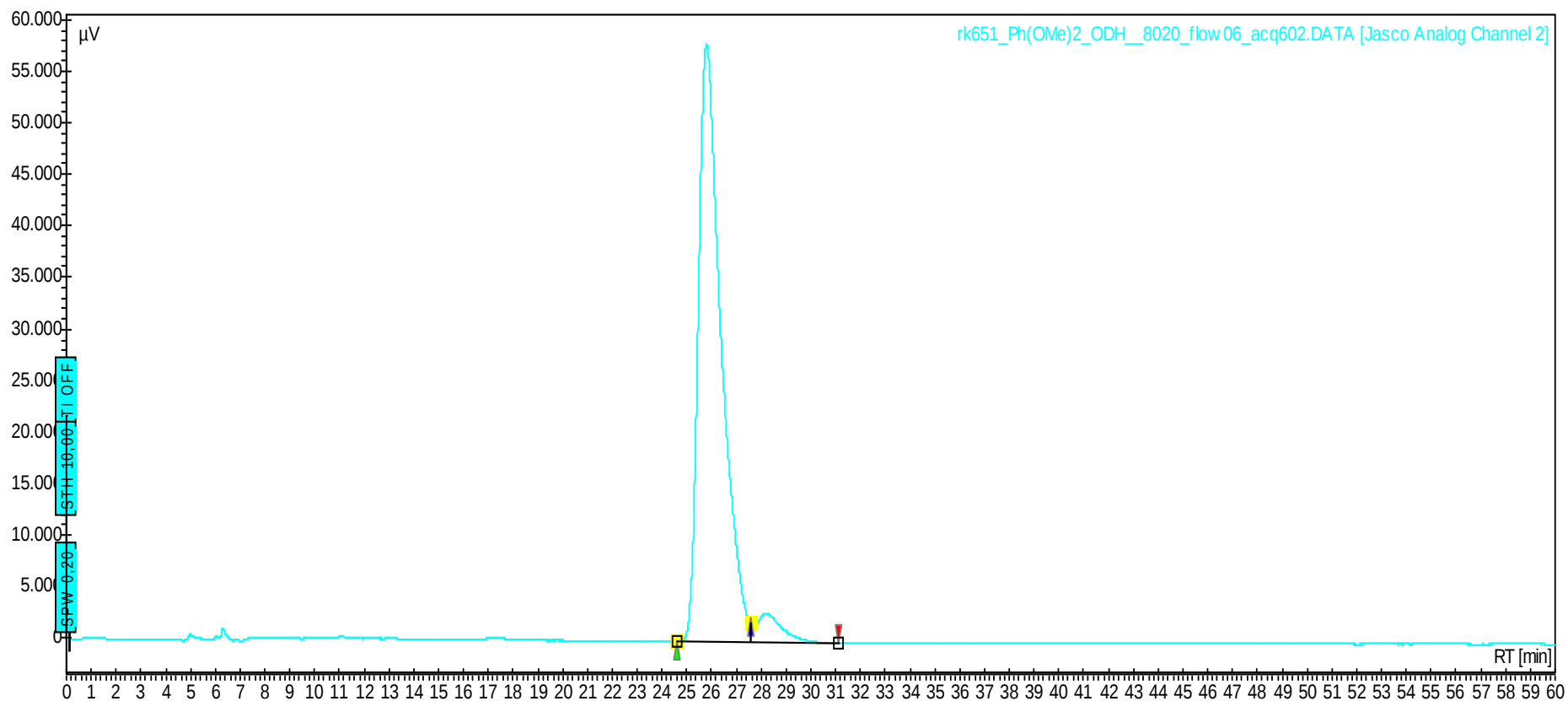
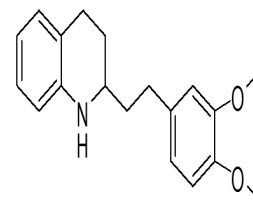


RK651_Ph(OMe)2_rac_ODH__8020_flow06_acq601.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	23,453	24,692	26,384	0,000	49,69	7964,0	7817,8	49,688
2	UNKNOWN	26,384	27,342	30,064	0,000	50,31	7178,1	7915,9	50,312
Total						100,00	15142,1	15733,7	100,000

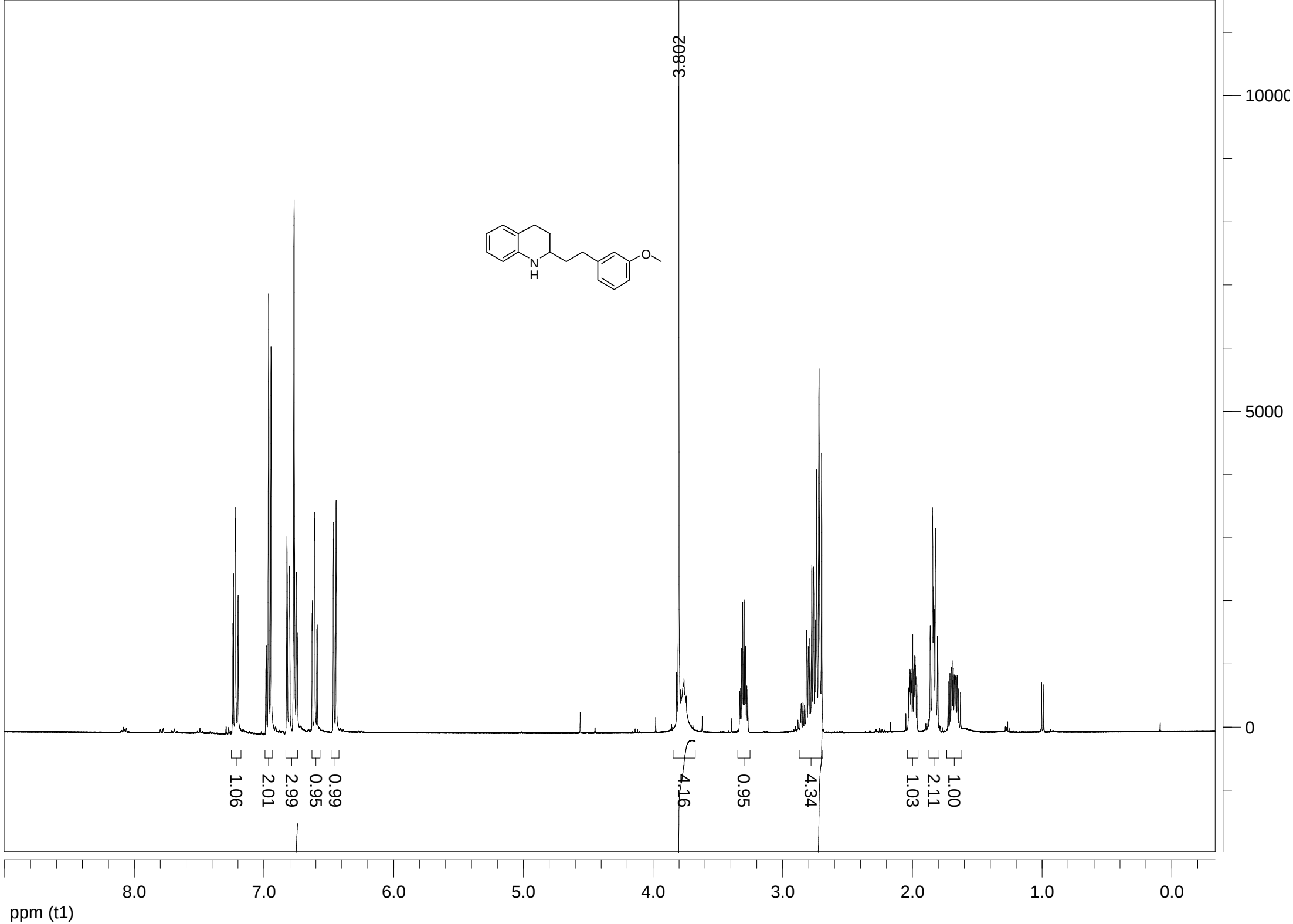
Chromatogram : rk651_Ph(OMe)2_ODH_8020_flow06_acq602

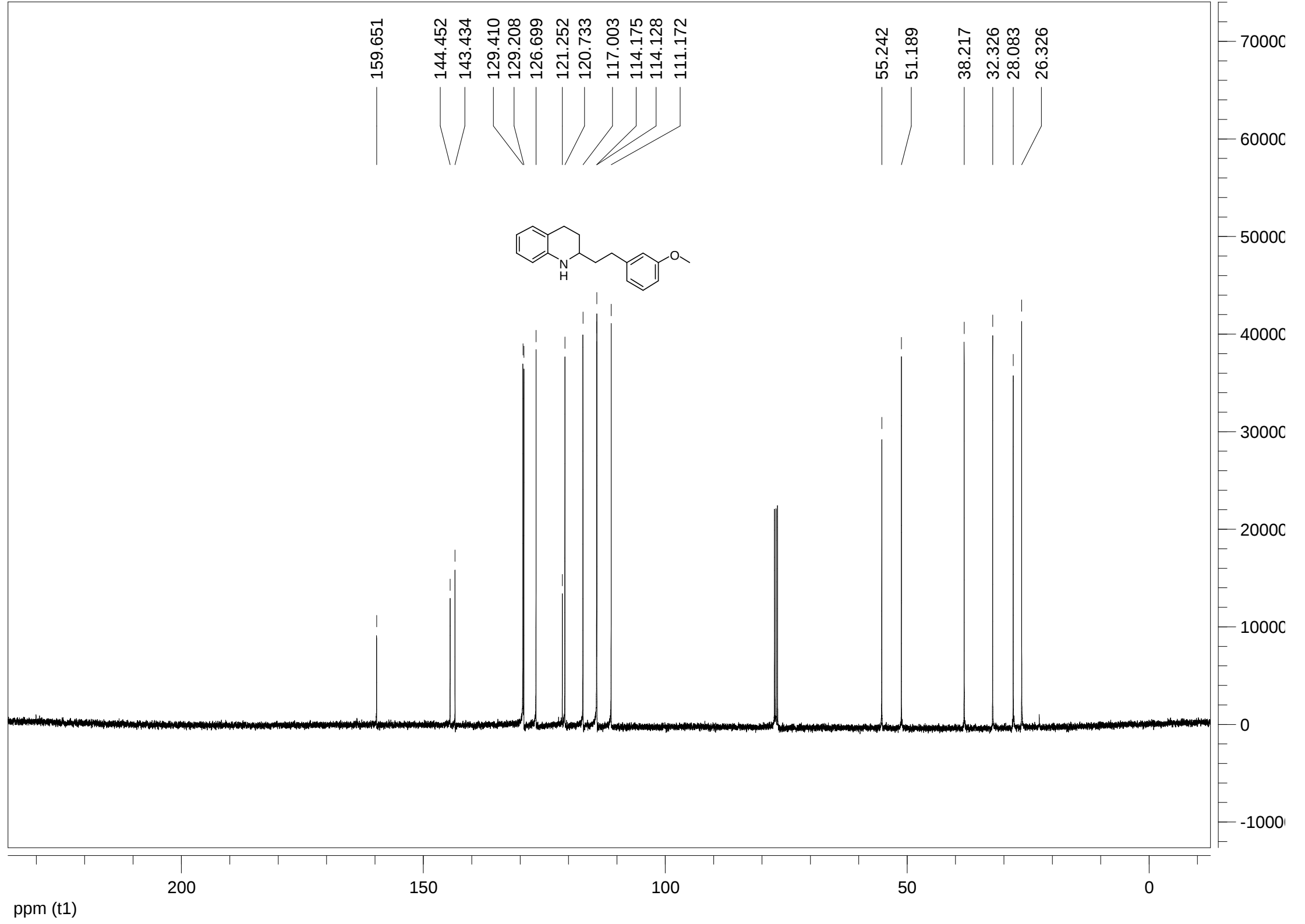
Data file: rk651_Ph(OMe)2_ODH_8020_flow06_acq602.DATA
Method: HPLC2_ODH_8020_flow06_acq60
Date: 20.09.2009 23:31:08



rk651_Ph(OMe)2_ODH_8020_flow06_acq602.DATA [Jasco Analog Channel 2]

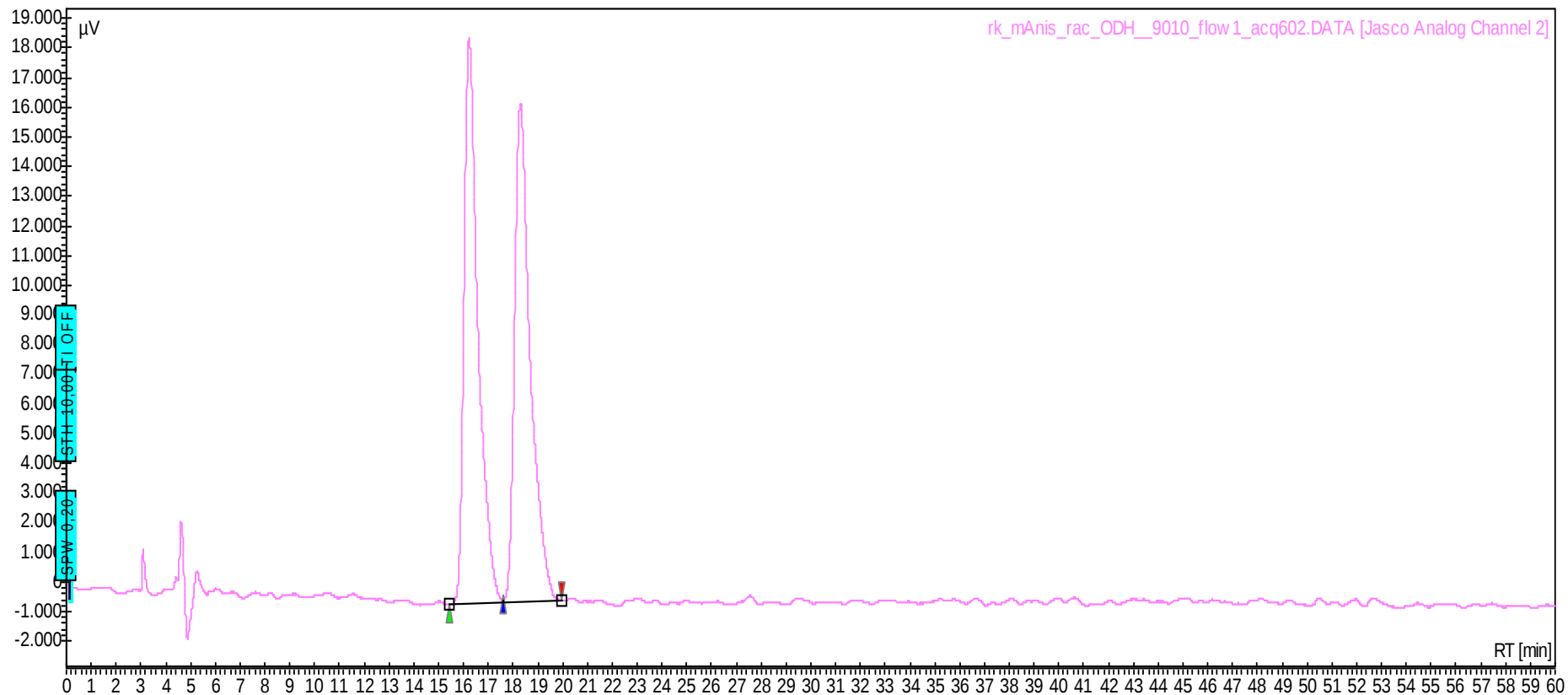
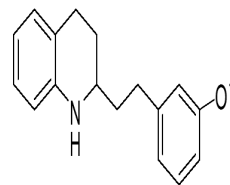
Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	24,632	25,800	27,559	0,000	94,76	58092,2	61974,0	94,764
2	UNKNOWN	27,559	28,217	31,130	0,000	5,24	2787,8	3424,3	5,236
Total						100,00	60879,9	65398,4	100,000





Chromatogram : rk_mAnis_rac_ODH__9010_flow1_acq602

Data file: rk_mAnis_rac_ODH__9010_flow1_acq602.DATA
Method: HPLC2_ODH_9010_flow1_acq60
Date: 24.09.2009 01:28:41

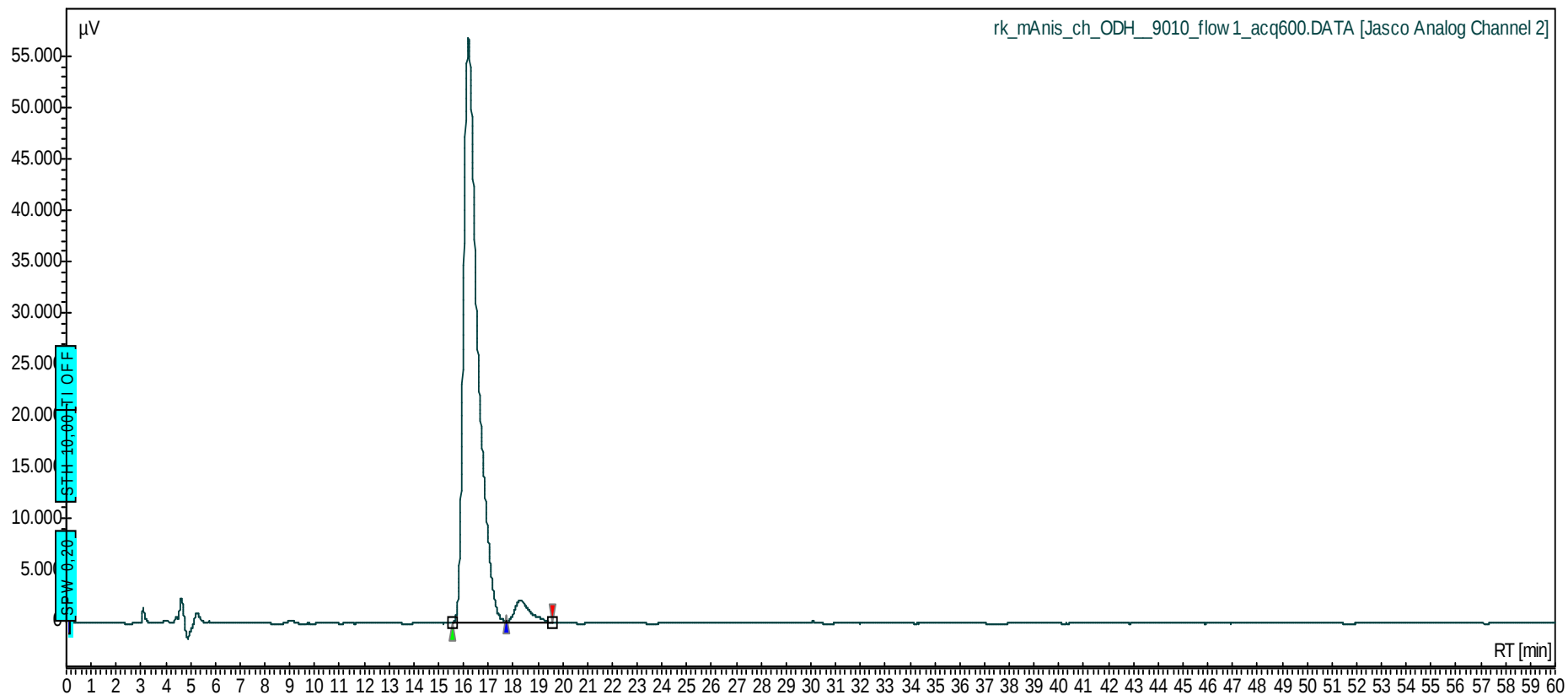
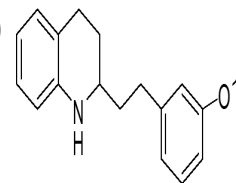


rk_mAnis_rac_ODH__9010_flow1_acq602.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	15,446	16,233	17,607	0,000	50,19	19060,0	12499,7	50,192
2	UNKNOWN	17,607	18,292	19,985	0,000	49,81	16833,5	12404,1	49,808
Total						100,00	35893,5	24903,8	100,000

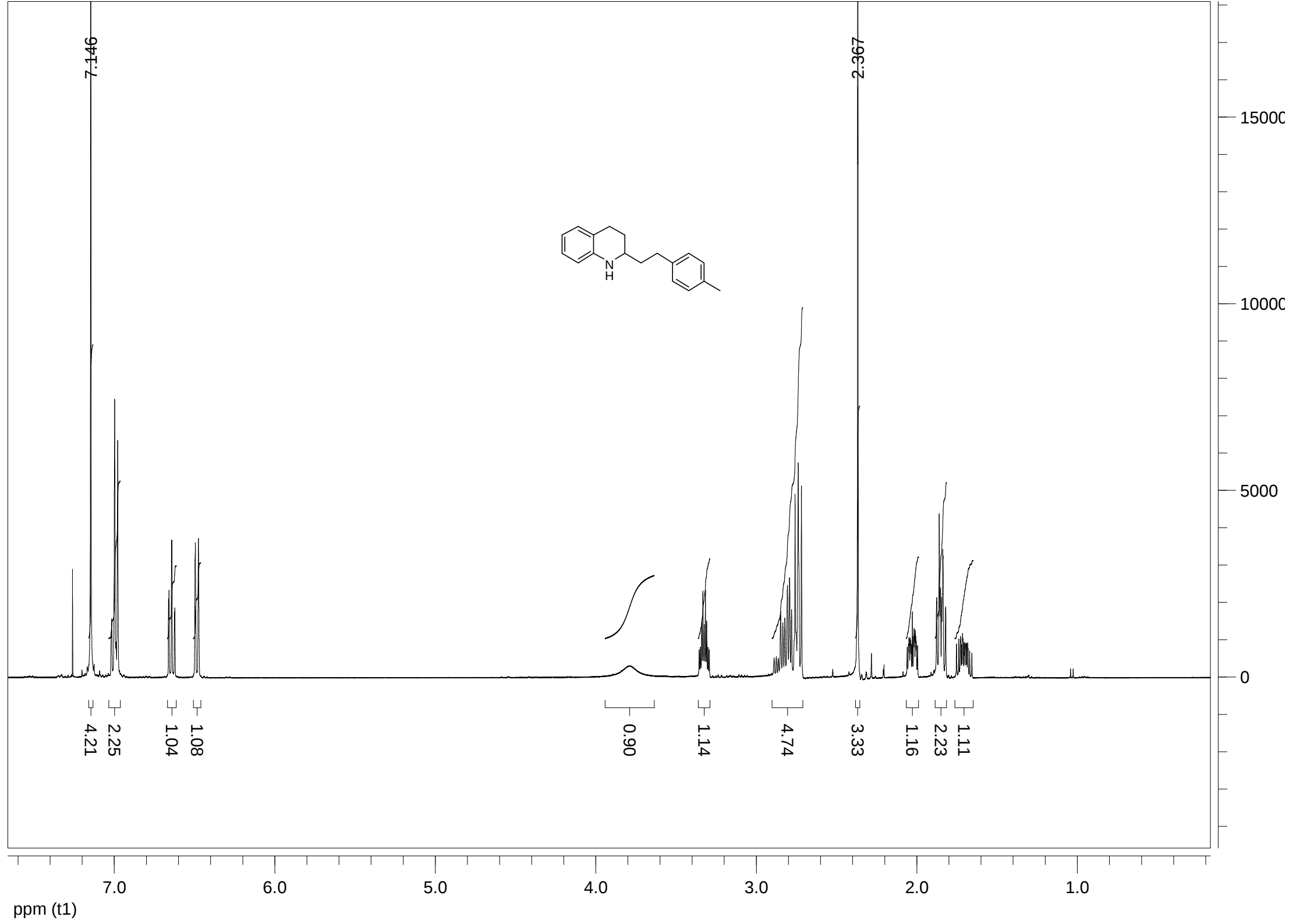
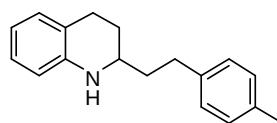
Chromatogram : rk_mAnis_ch_ODH__9010_flow1_acq600

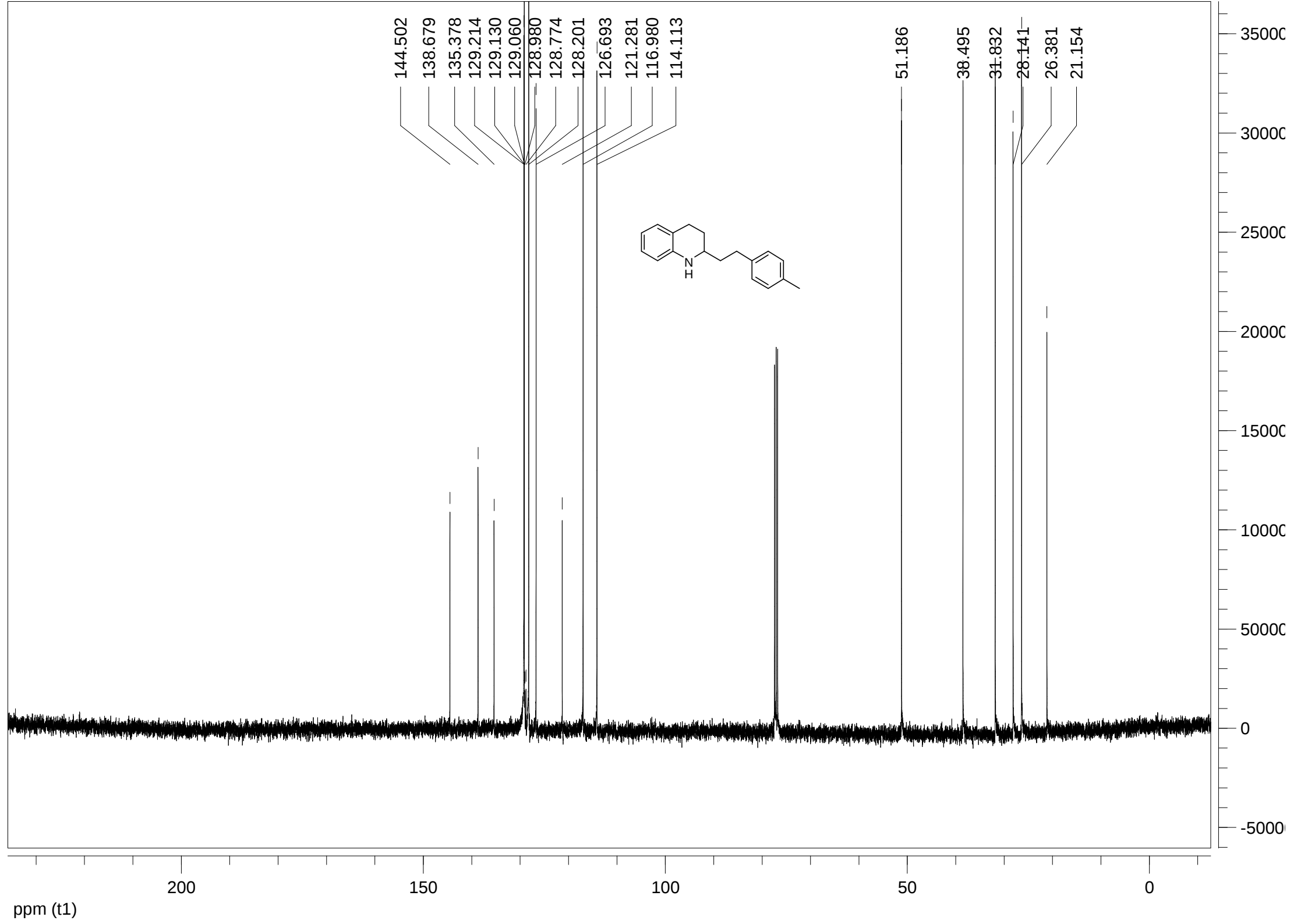
Data file: rk_mAnis_ch_ODH__9010_flow1_acq600.DATA
Method: HPLC2_ODH_9010_flow1_acq60
Date: 24.09.2009 02:31:22



rk_mAnis_ch_ODH__9010_flow1_acq600.DATA [Jasco Analog Channel 2]

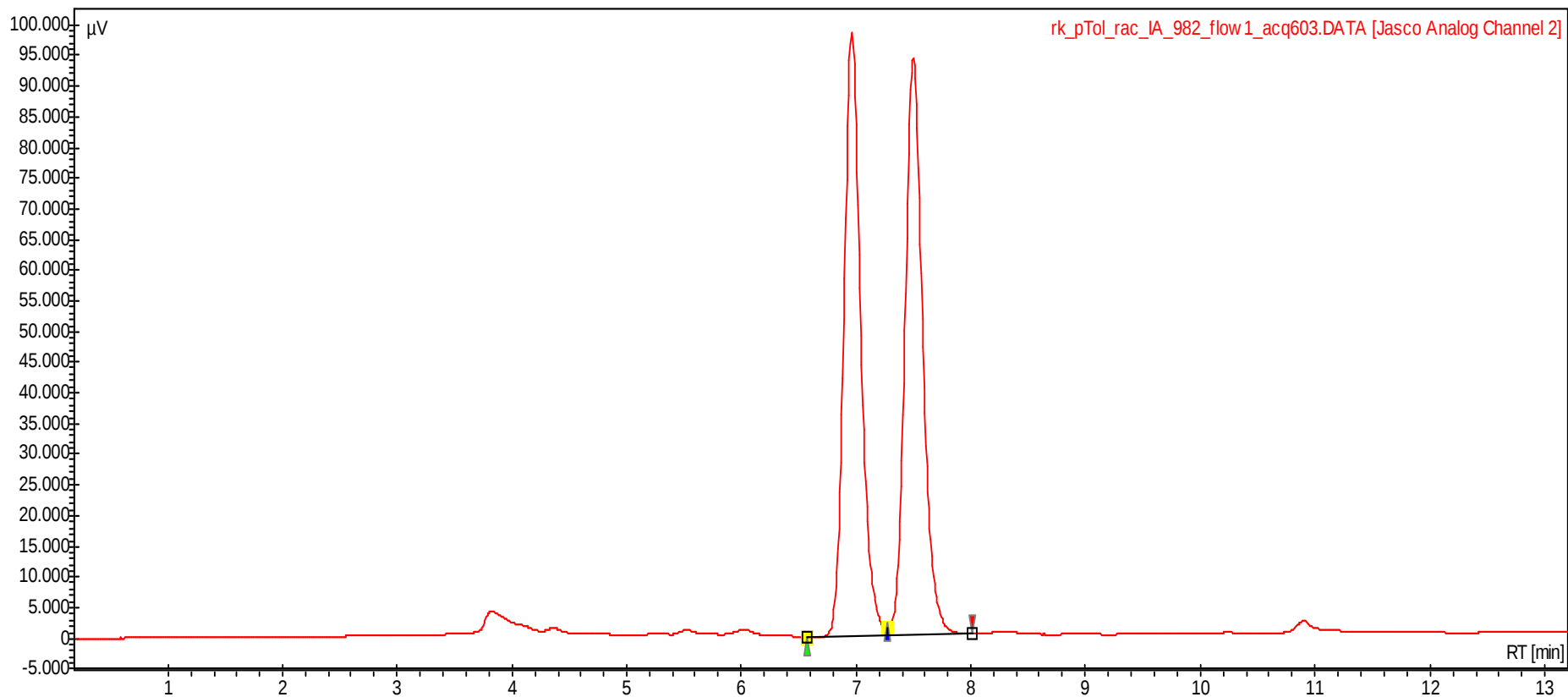
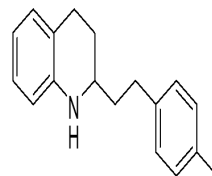
Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	15,532	16,200	17,716	0,000	95,72	56878,7	37458,7	95,716
2	UNKNOWN	17,716	18,300	19,588	0,000	4,28	2191,1	1676,6	4,284
Total						100,00	59069,8	39135,3	100,000





Chromatogram : rk_pTol_rac_IA_982_flow1_acq603

Data file: rk_pTol_rac_IA_982_flow1_acq603.DATA
Method: HPLC2_IA_982_flow1_acq60
Date: 24.09.2009 04:06:39

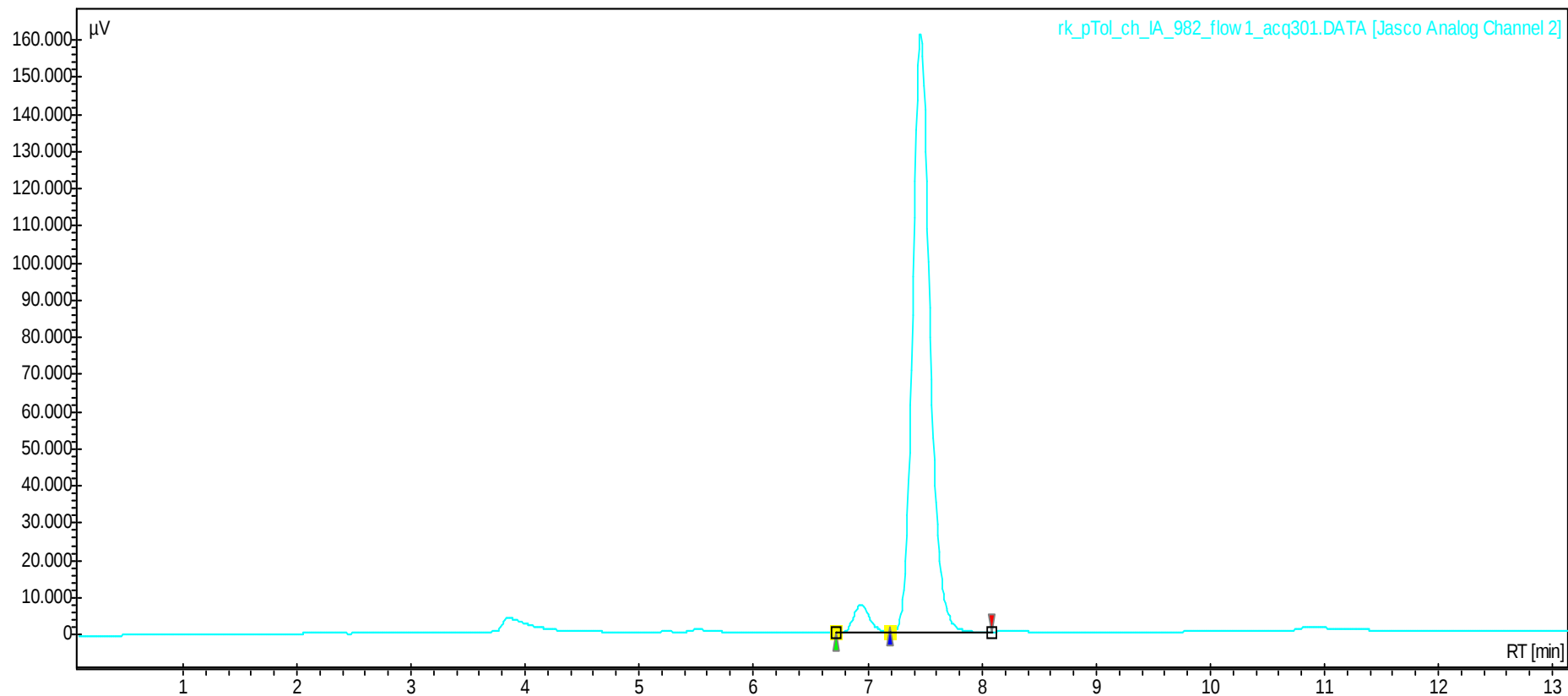
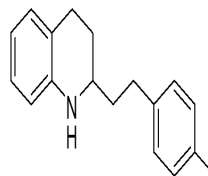


rk_pTol_rac_IA_982_flow1_acq603.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	6,572	6,958	7,275	0,000	49,93	98434,4	16686,8	49,926
2	UNKNOWN	7,275	7,500	8,014	0,000	50,07	94128,0	16736,5	50,074
Total						100,00	192562,4	33423,3	100,000

Chromatogram : rk_pTol_ch_IA_982_flow1_acq301

Data file: rk_pTol_ch_IA_982_flow1_acq301.DATA
Method: HPLC2_IA_982_flow1_acq30
Date: 24.09.2009 08:58:11



rk_pTol_ch_IA_982_flow1_acq301.DATA [Jasco Analog Channel 2]

Index	Name	Start	Time	End	Ret. time Offset	Quantity	Height	Area	Area %
		[Min]	[Min]	[Min]	[Min]	[% Area]	[µV]	[µV.Min]	[%]
1	UNKNOWN	6,727	6,942	7,197	0,000	4,31	7726,0	1288,0	4,314
2	UNKNOWN	7,197	7,467	8,084	0,000	95,69	161258,6	28569,5	95,686
Total						100,00	168984,6	29857,5	100,000