

Supporting information for:

(E,E)-1,5-cyclooctadiene: a small and fast click-chemistry multitalent

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Cell culture

Murine T-cell lymphoma (EL4, from ATCC, Teddington, UK) cells were grown in RPMI-1640 medium (Invitrogen, Paisley, UK); 2.0 g L⁻¹ glucose, 300 mg L⁻¹ L-glutamine) supplemented with 10% FBS (fetal bovine serum, PAA laboratories, Yeovil, UK) and maintained in a 5% CO₂, water-saturated atmosphere at 37 °C.

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Cell surface azidoglycan labeling and detection by flow cytometry

EL4 cells were incubated for 24 h in media containing 50 μ M Ac₄GalNAz. Control cells were grown in the absence of Ac₄GalNAz. The following procedure was followed for both azidosugar-pulsed and non-pulsed sets of cells. The medium was removed from the flasks and cells were washed with warm PBS (phosphate buffered saline; water, NaCl, KCl, Na₂HPO₄, KH₂PO₄; Fisher Scientific, Loughborough, UK). The contents of each flask were transferred to a centrifuge tube, centrifuged (600 g, 4 °C, 4 min), resuspended in cold FACS buffer (1% FBS in 20 mM HEPES, 150 mM NaCl, pH 7.4) and transferred to 1.5 mL Eppendorf tubes. Cells were centrifuged and resuspended in 100 μ L labeling buffer (100 μ M **11** in FACS buffer) or as a control 100 μ L FACS buffer. The Eppendorf tubes were incubated in a hot block with orbital shaking (500 rpm, 37 °C, 30 min). The cells were washed three times with 700 μ L ice cold FACS buffer, centrifuged and resuspended in 100 μ L labeling buffer (10 μ M **12** in FACS buffer containing 50 nM SYTOX Green, Invitrogen) or as a control 100 μ L 50 nM SYTOX Green in FACS buffer, followed by incubation as above for 20 min. Cells were then filtered through a 50 μ m cut-off membrane into flow cytometry tubes and kept on ice. Each sample was analysed by a flow cytometer (model LSRII, BD Oxford, UK) using 20,000 events.

Data analysis was performed using FlowJo flow cytometry analysis software (Tree Star, Ashland, OR). The viable cell population (population of interest) was determined by gating cells to exclude those with a low NADH auto-fluorescence and those with high levels of SYTOX Green (cell death marker). The far-red median fluorescence intensity (MFI, Alexa Fluor 647 fluorophore) of the viable cell population was then assessed (Figure S1).[†] Data points were collected in triplicate.

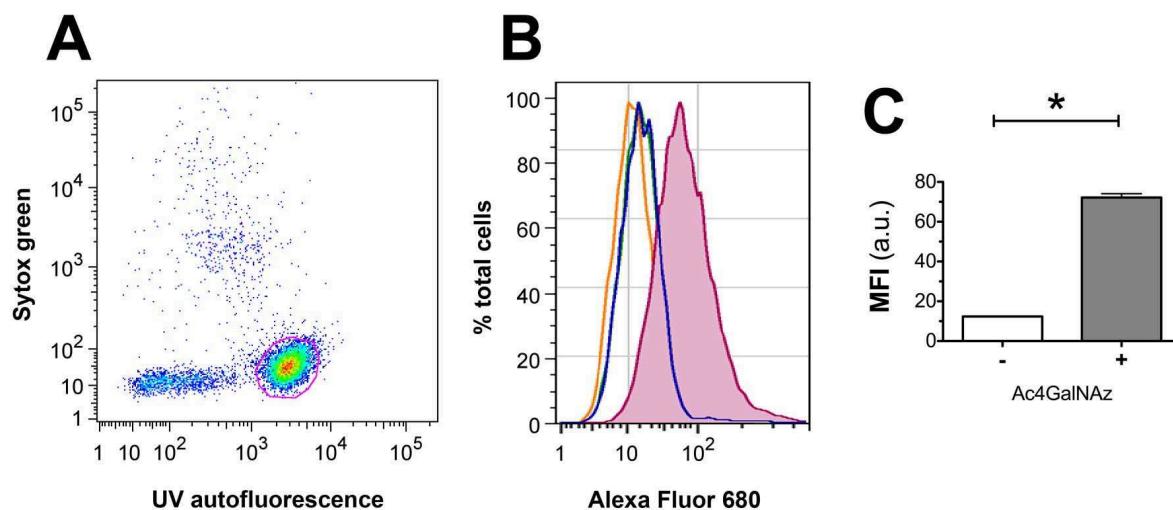
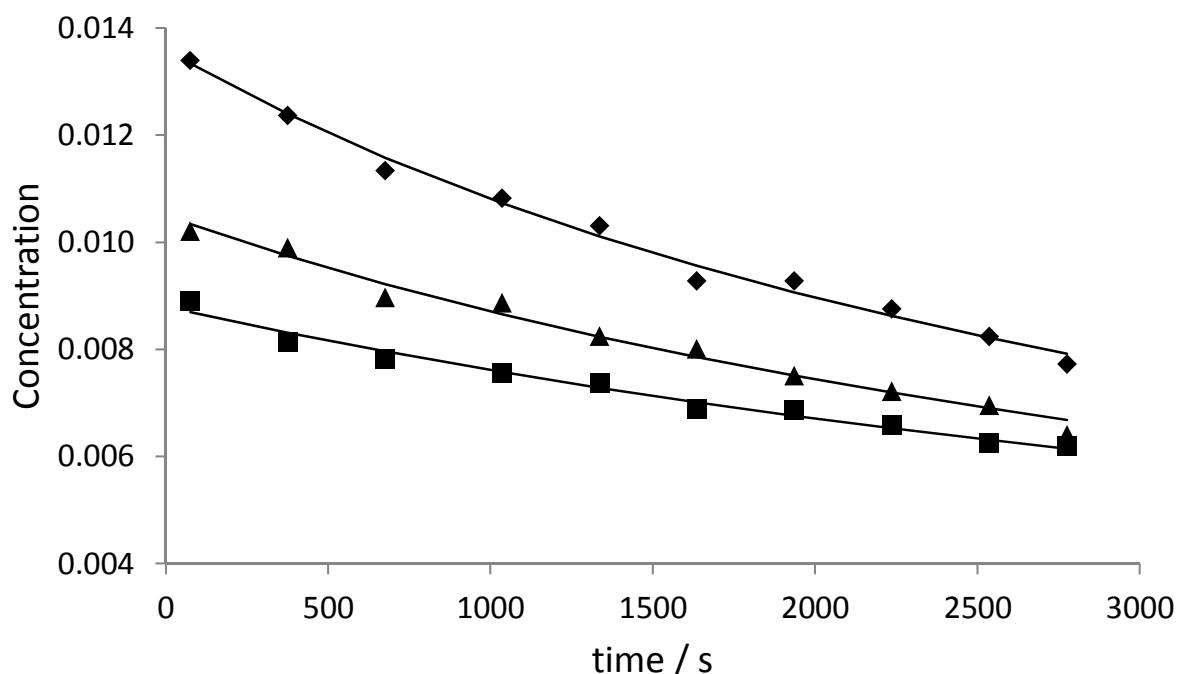


Figure S1: Flow cytometry analysis of EL4 cell labelling using **11** and tetrazine **12**. Cells were cultured in the presence or absence of Ac₄GalNAz (50 mM) for 24h, then incubated with a 100 μ M solution of **11** for 30 min, 37 °C, 500 r.p.m. After washing, cells were reacted with **12**, 10 μ M, for 20 min, 37 °C, 500 r.p.m., re-washed and analyzed by flow cytometry against controls. The population of viable cells (population of interest; pink circle in (A)) was identified from the high levels of UV autofluorescence, caused by intracellular NADH, and low levels of Sytox green (cell death marker); Histogram analysis (B) of EL4 cells revealed a ca. 6-fold difference (C) between Ac₄GalNAz-treated (+) and non-treated (-) cells. MFI, median fluorescence intensity. Histogram color scale: orange: cells with no dyes, blue: cells + **12**, green: cells+**11+12**, red: cells+Ac₄GalNAz+**11+12**. Data points (C) were collected in triplicate and are shown as mean \pm SD (error bars lie within the chart bars when not visible). * p < 0.05, one-tailed t-test with Mann Whitney correction.

Kinetic evaluation of the [3+2] cycloaddition of (*E,E*)-1,5-cyclooctadiene with benzyl azide

Kinetic experiments for the first [3+2] cycloaddition were performed under second-order conditions. (*E,E*)-1,5-cyclooctadiene was mixed in a 1:1 molar ratio with benzyl azide in CDCl₃ and the reaction was monitored by ¹H-NMR (500 MHz) at 25 °C probe temperature. The second-order rate constants were determined by fitting the experimental data to the second-order rate equation.

time / s	integral	concentration / M	fit	error
0	0.264	0.0136	0.0136	0.0E+00
75	0.260	0.0134	0.0133	3.7E-09
375	0.240	0.0124	0.0124	7.9E-10
675	0.220	0.0113	0.0116	5.8E-08
1035	0.210	0.0108	0.0107	8.6E-09
1335	0.200	0.0103	0.0101	3.8E-08
1635	0.180	0.0093	0.0096	8.1E-08
1935	0.180	0.0093	0.0091	4.4E-08
2235	0.170	0.0088	0.0086	1.9E-08
2535	0.160	0.0082	0.0082	7.4E-10
2775	0.150	0.0077	0.0079	3.7E-08
		Sum	2.9E-07	



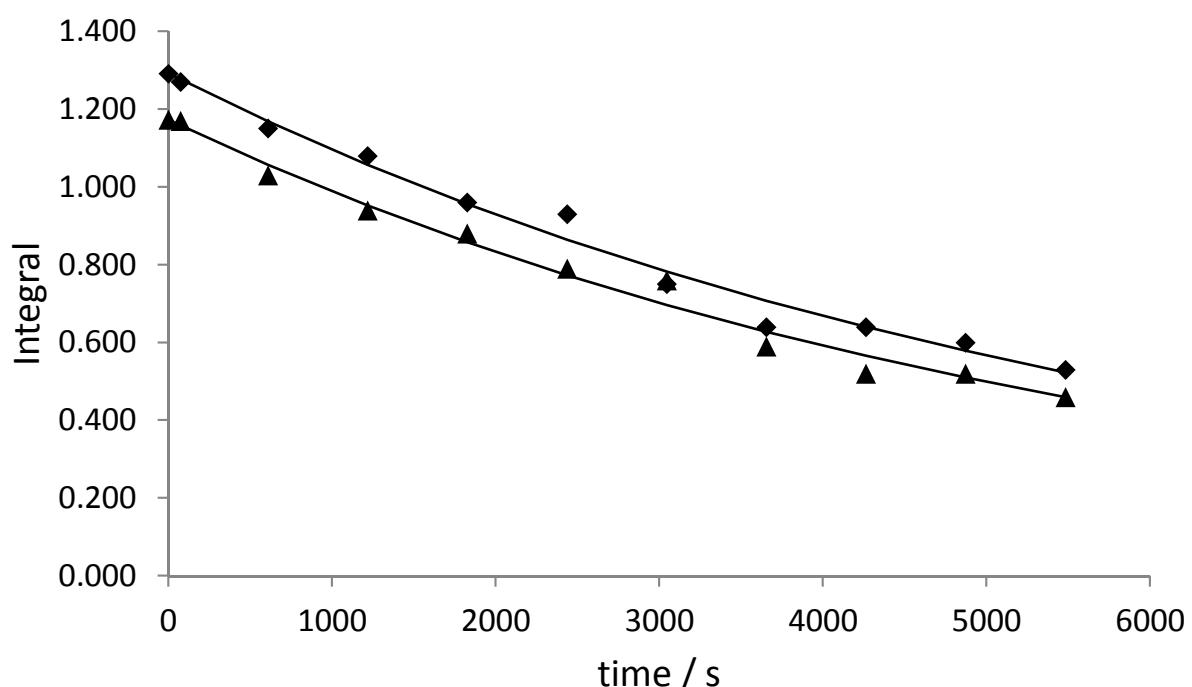
Entry	c_0 / M	$k / s^{-1} \times M^{-1}$
1	0.0136	0.0190
2	0.0105	0.0196
3	0.0177	0.0177
average	0.0187	
error	0.0010	

Figure S2: Kinetic evaluation of [3+2] cycloaddition of (*E,E*)-1,5-cyclooctadiene with benzyl azide. Top: Example experimental data and fitted data from one experiment. Center: Plots of benzyl azide concentration *vs.* time from three different experiments. Bottom: Summary of data (starting concentrations and rate constants) from three different experiments.

Kinetic evaluation of the second [3+2] cycloaddition of (*E,E*)-1,5-cyclooctadiene with benzyl azide

Kinetic experiments for the second [3+2] cycloaddition were performed under pseudo first order conditions. Triazoline **8** was mixed in a ca. 1:20 molar ratio with benzyl azide in CDCl_3 and the reaction was monitored by $^1\text{H-NMR}$ (500 MHz) at 25 °C probe temperature. The pseudo first order rate constants were determined by fitting the experimental data to the pseudo first order rate equation.

time / s	integral	fit	error
0	1.292	1.2919	0.0E+00
75	1.270	1.2761	3.7E-05
609	1.150	1.1686	3.4E-04
1218	1.080	1.0570	5.3E-04
1827	0.960	0.9560	1.6E-05
2436	0.930	0.8648	4.3E-03
3045	0.750	0.7822	1.0E-03
3654	0.640	0.7075	4.6E-03
4263	0.640	0.6399	5.1E-09
4872	0.600	0.5788	4.5E-04
5481	0.530	0.5235	4.2E-05
Sum			1.1E-02



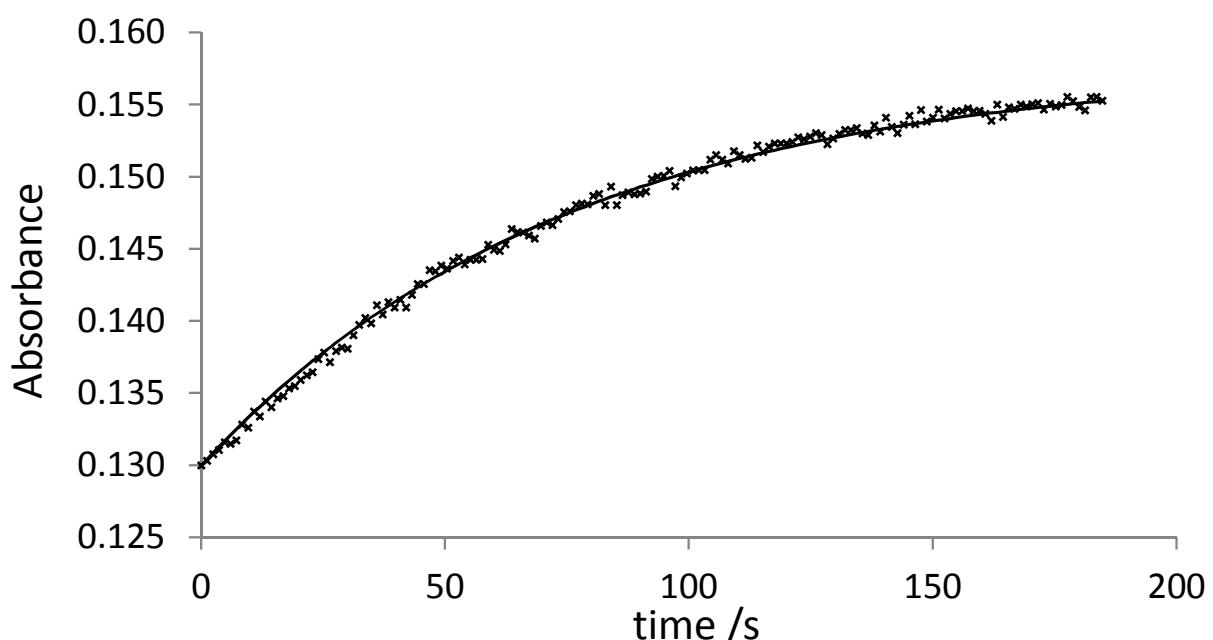
Entry	c ₀ / M	k' / s ⁻¹	k / s ⁻¹ x M ⁻¹
1	0.5380	1.65E-04	3.06E-04
2	0.5800	1.71E-04	2.95E-04
		average	3.0E-04
		error	8.1E-06

Figure S3: Kinetic evaluation of the second [3+2] cycloaddition of (*E,E*)-1,5-cyclooctadiene with benzyl azide. Top: Example experimental data and fitted data from one experiment. Center: Plots of benzyl azide concentration vs. time from three different experiments. Bottom: Summary of data (starting concentrations and rate constants) from two different experiments.

Kinetic evaluation of inverse-electron-demand Diels-Alder reactions with tetrazine 5

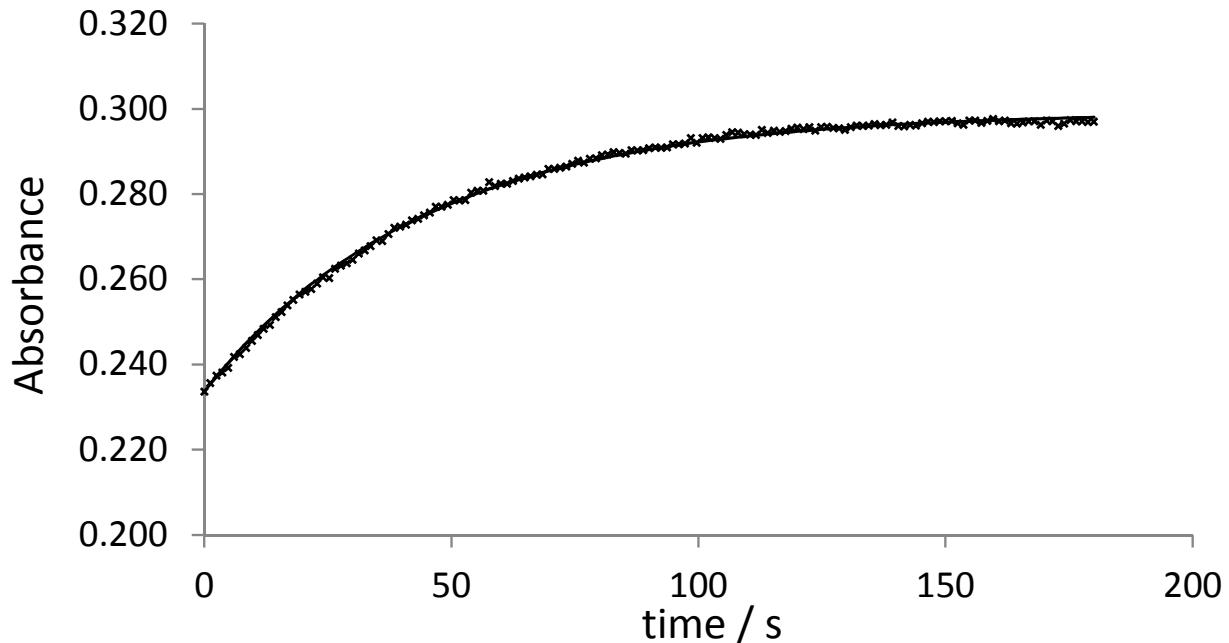
Kinetic experiments for inverse-electron-demand Diels-Alder reactions were performed under pseudo first order conditions by UV-Vis spectroscopy. The pseudo first order rate constants were determined by fitting the experimental data to the corresponding rate equation.

The reactions between trans-cyclooctenes **6** and **8** and 3,6-di-(2-pyridyl)-*s*-tetrazine (**5**) were monitored by UV-Vis spectroscopy at 340 nm using the reagents in a 1:10 molar ratio in MeOH at 25 °C. The concentrations of excess reagent are given in the tables.



Entry	k'	A_{∞}	c_0 / M	$k / s^{-1} \times M^{-1}$
1	0.0134	0.1576	5.00E-05	267.04
2	0.0159	0.1494	5.00E-05	318.02
3	0.0153	0.1598	5.00E-05	305.49
average				296.85
error				26.56

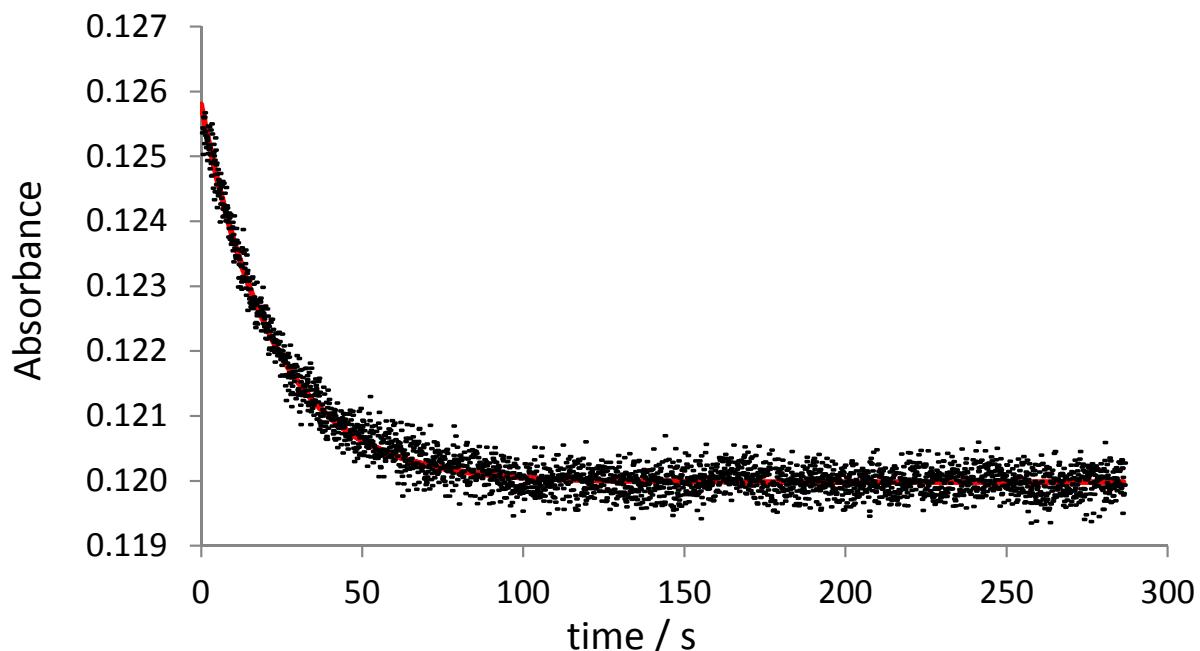
Figure S4: Kinetic evaluation of inverse-electron-demand Diels-Alder reaction of triazoline **8** with 3,6-di-(2-pyridyl)-*s*-tetrazine (**5**). Top: Plot of absorbance vs. time. Bottom: Summary of data (pseudo first order rate, final absorbance, concentration of excess substrate and second order rate constants) from three different experiments.



Entry	k'	A_{∞}	A_0	c_0 / M	$k / s^{-1} \times M^{-1}$
1	0.0224	0.2992	0.2336	5.00E-05	448.52
2	0.0221	0.2982	0.2363	5.00E-05	442.28
3	0.0235	0.2816	0.2219	5.00E-05	470.41
average					453.74
error					14.77

Figure S5: Kinetic evaluation of inverse-electron-demand Diels-Alder reaction of trans-cyclooctene **6** with 3,6-di-(2-pyridyl)-*s*-tetrazine (**5**). Top: Plot of absorbance vs. time. Bottom: Summary of data (pseudo first order rate, final absorbance, concentration of excess substrate and second order rate constants) from three different experiments.

The reaction between (*E,E*)-1,5-cyclooctadiene **4** and 3,6-di-(2-pyridyl)-*s*-tetrazine (**5**) was monitored by UV-Vis spectroscopy at 295 nm using (*E,E*)-1,5-cyclooctadiene and tetrazine in a 1:10 molar ratio in THF at 25 °C. The concentrations of excess reagent are given in the table.



Entry	k' / s^{-1}	A_{∞}	c_0 / M	$k / \text{s}^{-1} \times \text{M}^{-1}$
1	0.0441	0.1200	5.00E-06	8819.22
2	0.0506	0.1234	5.00E-06	10126.70
3	0.0385	0.1185	5.00E-06	7709.28
average				8885.07
error				1210.05

Figure S6: Kinetic evaluation of inverse-electron-demand Diels-Alder reaction of (*E,E*)-1,5-cyclooctadiene **4** with 3,6-di-(2-pyridyl)-*s*-tetrazine (**5**). Top: Plot of absorbance *vs.* time. Bottom: Summary of data (pseudo first order rate, final absorbance, concentration of excess substrate and second order rate constants) from three different experiments.

General methods and materials

NMR spectra were recorded on the following instruments: Bruker DRX500, Bruker Avance BB, Bruker Avance TCI, Bruker AM400 and Bruker DRX400. All chemical shifts are quoted in ppm, relative to tetramethylsilane, using the residual solvent peak as a reference standard. All coupling constants are quoted in Hz. Infrared spectra were recorded on a Perkin Elmer Spectrum One (FT-IR) spectrophotometer. High resolution mass spectra (HRMS) were recorded on a Waters LCT Premier TOF mass spectrometer with electrospray and modular Lockspray interface. Analytical thin layer chromatography (TLC) was carried out on Merck Kieselgel 60 F254 plates with visualisation by ultra violet light (254 nm), potassium permanganate or ninhydrin or phosphomolybdic acid /Ce(SO₄)₂ dip. Flash column chromatography was carried out using Merck Kieselgel 60 (230-400 mesh). HPLC was carried out on a Varian ProStar system, UV detection at 254 nm, Phenomenex Jupiter C18 column, particle size 5 µm. LC-MS analysis was performed on a Waters 2795 system, UV detection at 254 nm, Supelco ABZ+plus column, 3.3cm x 4.6mm, particle size 3 µm. All solvent mixtures are reported as % vol/vol unless otherwise stated. Reagents and solvents were purified using standard means. All other chemicals were used as received unless noted otherwise. Extractive procedures were performed using distilled solvents and evaporation of solvents was performed under reduced pressure. All aqueous solutions used were saturated. Unless otherwise stated, all non-aqueous reactions were carried out under an argon atmosphere using anhydrous conditions and oven-dried glassware. Standard techniques were employed for handling air-sensitive materials.

Synthetic procedures

cis,cis-5,10-Dioxatricyclo[7.1.0.0^{4,6}]decane (2)¹

Sodium carbonate (53.9 g, 508 mmol, 1.1 eq) was added to a stirred solution of (Z,Z)-1,5-cyclooctadiene (50 g, 462 mmol, 1.0 eq) in DCM (200 mL). The suspension was cooled to 0 °C and peracetic acid (36%, 179 mL, 971 mmol, 2.1 eq) was added. The mixture was warmed to r.t. and

stirred until the reaction had gone to completion (typically 2 d). The pH was adjusted to ca. 11 with 40% NaOH at 0 °C, followed by extraction with Et₂O (4 x 150 mL). The combined organic layers were dried over MgSO₄, concentrated and distilled to give 5,10-dioxatricyclo[7.1.0.0^{4,6}]decane **2** (45.8 g, 327 mmol, 70.7% yield), b.p. 69-70 °C at 0.3 mm Hg, which crystallized on standing. The spectroscopic data were in accordance with the literature.

β-hydroxy-diphenylphosphine oxides

***rel*-((1S,2S,5S,6S)-2,6-dihydroxyclooctane-1,5-diyl)bis(diphenylphosphine oxide) (3a)** and

***rel*-((1R,4S,5S,8R)-5,8-dihydroxyclooctane-1,4-diyl)bis(diphenylphosphine oxide) (3b)**

To a solution of diphenylphosphine (8.4 g, 44.9 mmol, 2.1 eq.) and cis,cis-5,10-dioxatricyclo-[7.1.0.0^{4,6}]decane (**2**) (3.0 g, 21.4 mmol, 1.0 eq.) in THF (86 mL), BuLi (1.5 M solution in hexanes, 30.0 mL, 44.9 mmol, 2.1 eq.) was added dropwise at -78 °C. The mixture was stirred for 1 h, allowed to warm to room temperature and stirred for 12 h. The brown solution was cooled to 0 °C, diluted with THF (86 mL) and quenched by the addition of AcOH (3.7 mL, 3.86 g, 64.2 mmol, 3 eq) and H₂O₂ (30%, 6.6 mL, 64.2 mmol, 3 eq). The mixture became clear, followed by the formation of a white precipitate. The mixture was removed from the ice bath and stirred vigorously at 25 °C for 2 h whereupon the white precipitate dissolved. The clear solution was then transferred to a separatory funnel, Et₂O (86 mL) added, the aqueous layer removed, and the organic layer washed with brine (2 x 20 mL). The combined organic layers were dried over MgSO₄, filtered and concentrated to afford a white solid. The remaining AcOH was azeotropically removed with toluene to yield an isomeric mixture of phosphine oxides (11.07 g, 20.3 mmol, 95%) as a white solid, mp 112-120 °C.

R_f (10% MeOH in DCM) 0.61 and 0.71;

δ_H(500 MHz, MeOD) 7.93-7.40 (20H, m), 4.16 (1H, app q, J = 8.0), 4.06 (1H, app q, J = 7.1), 3.19 (1H, app q, J = 8.3), 3.05 (1H, app q, J = 9.2), 2.12-1.94 (3H, m), 1.94-1.83 (1H, m), 1.83-1.71 (1H, m), 1.71-1.48 (3H, m);

δ_C(125 MHz, MeOD) 135.01 (d, J = 58.2), 134.22 (d, J = 54.6), 133.96 (d, J = 54.6), 133.19 (d, J

= 54.5), 133.03 (d, J = 2.4), 132.90 (d, J = 2.4), 132.76 (d, J = 2.6), 132.62 (d, J = 2.7), 132.34 (d, J = 8.8), 132.07 (d, J = 9.4), 131.96 (d, J = 8.8), 131.83 (d, J = 9.2), 129.95 (d, J = 11.4), 129.86 (d, J = 11.4), 129.60 (d, J = 11.8), 129.58 (d, J = 12.0), 71.17 (d, J = 3.2), 70.62 (d, J = 2.4), 44.01 (d, J = 69.8), 42.20 (d, J = 71.2), 34.74 (dd, J = 9.6, 9.6), 32.42 (d, J = 9.2), 24.29 (d, J = 13.9), 21.59;

ν_{max} (solid)/cm⁻¹ 3319.37, 3057.03, 2923.32, 1590.98, 1483.93, 1437.23, 1313.20;
 m/z (EI) 545.2021 ($M+H^+$. C₃₂H₃₅O₄P₂ requires 545.2011).

(E,E)-1,5-cyclooctadiene (4)

The mixture of phosphine oxides **3a** and **3b** (3.00 g, 5.5 mmol, 1.0 eq) was dissolved in DMF (30 mL) with stirring under an Ar atmosphere. The clear solution became cloudy and NaH (60% in mineral oil, 0.72 g, 17.1 mmol, 3.1 eq) was added in portions over 5 min at r.t. under a positive Ar stream. The reaction mixture was stirred for 12 h at room temperature, cooled to 0 °C, diluted with pentane (25 mL) and quenched with half-saturated NH₄Cl (11 mL). The aqueous layer was extracted with pentane (25 mL) and the combined organic layers were washed with water (5 x 22 mL) and brine (1 x 22 mL) to give (E,E)-1,5-cyclooctadiene (50 mL of a 25 mM solution in pentane, 1.3 mmol, 23% yield).

The concentration of (E,E)-1,5-cyclooctadiene was determined by titration with 3,6-Di-2-pyridyl-1,2,4,5-tetrazine, color change from pink to yellow.

Transfer of the product into other organic solvents: The 10 mL of the pentane solution was extracted with an equal volume of aqueous 0.1 M AgNO₃. The pentane layer was discarded, the desired volume of organic solvent was added, followed by the addition of 10 mL of 35 % aqueous NH₃ and vigorous shaking. The organic layer was washed with water (10 mL) and brine (10 mL). R_f (pentane) 0.43;

δ_H (500 MHz, D₂O) 5.40-5.27 (4H, m), 2.83-2.70 (4H, m), 2.58-2.41 (4H, m);
 δ_C (125 MHz, D₂O) 126.63, 31.85.

Tetrazine monoadduct 6

rel-(4aS,10aS,pR)-1,4-di(pyridin-2-yl)-4a,5,6,9,10,10a-hexahydrocycloocta[d]pyridazine

To (*E,E*)-1,5-cyclooctadiene (2 mL, 0.02 molar in pentane, 40 μ mol, 1.0 eq) was added 3,6-di(pyridin-2-yl)-1,2,4,5-tetrazine (4.7 mg, 20 μ mol, 0.5 eq) with stirring at room temperature. A small amount of solid that formed during the reaction (bis-adduct) was removed by filtration and the filtrate concentrated to give the desired product (10.8 mg, 34 μ mol, 85% yield) as a yellow solid, mp 198-202 °C.

Rf=0.27 (5% MeOH in /DCM);

δ_H (500 MHz, CDCl₃) 8.64 (2H, qd, *J* = 0.88, 4.78), 8.28 (2H, td, *J* = 0.99, 7.98), 7.74 (2H, dt, *J* = 1.78, 7.73), 7.31 (2H, ddd, *J* = 1.16, 4.79, 7.49), 5.92-5.82 (2H, m), 3.56 (2H, d, *J* = 7.65), 2.64 (2H, d, *J* = 11.80), 2.58-2.42 (2H, m), 1.84 (2H, dd, *J* = 5.18, 13.03), 1.72-1.60 (2H, m);

δ_C (125 MHz, CDCl₃) 163.26, 154.53, 148.87, 136.33, 135.22, 124.64, 122.83, 36.81, 36.54, 36.28;

ν_{max} (solid)/cm⁻¹ 3045.75, 2999.87, 2950.88, 2862.98, 1596.41, 1560.02, 1546.85, 1463.16, 1426.46, 1364.35;

m/z (EI) 317.1757 (M+H⁺. C₂₀H₂₁N₄ requires 317.1761).

Tetrazine bisadduct 7

rel-(4aS,6aS,10aS,12aS)-1,4,7,10-tetra(pyridin-2-yl)-4a,5,6,6a,10a,11,12,12a-octahydrocycloocta-[1,2-d:5,6-d']dipyridazine

To (*E,E*)-1,5-cyclooctadiene (2 mL, 0.02 molar in pentane, 40 μ mol, 1.0 eq) was added 3,6-di(pyridin-2-yl)-1,2,4,5-tetrazine (19.0 mg, 80 μ mol, 2.0 eq) with stirring at room temperature. The solid that formed during the reaction was isolated by filtration, washed with hexane and dried to give the desired product (19.0 mg, 36 μ mol, 89% yield) as a yellow solid, mp 250-252 °C. Impurities are presumably due to tautomerization of the product.

δ_H (500 MHz, CDCl₃) 8.63 (2H, qd, *J* = 0.85, 4.75), 8.36 (2H, td, *J* = 0.96, 8.03), 7.75 (2H, dt, *J* = 1.77, 7.76), 7.30 (2H, ddd, *J* = 1.13, 4.80, 7.43), 4.51 (4H, s), 2.18-1.99 (8H, m);

δ_C (125 MHz, CDCl₃) 162.16, 154.55, 148.95, 136.26, 124.59, 123.05, 36.43, 35.66;
 ν_{max} (solid)/cm⁻¹ 2941.10, 1595.37, 1561.35, 1545.55, 1461.80, 1429.72, 1388.14, 1355.43.
 m/z (EI) 525.2512 (M+H⁺. C₃₂H₂₉N₈ requires 525.2510).

Benzyl azide monoadduct 8

rel-(3a*S*,9a*S*, p*R*)-1-benzyl-3a,4,5,8,9,9a-hexahydro-1*H*-cycloocta[*d*][1,2,3]triazole

To (*E,E*)-1,5-cyclooctadiene (470 μ l, 0.2 molar in DCM, 94 μ mol, 1.1 eq) was added benzyl azide (11.4 mg, 86 μ mol, 1.0 eq) with stirring at r.t. No precautions were taken to exclude moisture or air from the reaction mixture. When the reaction had finished (typically 2 h, TLC), the solvent and excess (*E,E*)-1,5-cyclooctadiene were removed by rotary evaporation giving the desired product (20.5 mg, 85 μ mol, 99% yield) as a white crystalline solid, mp 113-115 °C.

R_f (50% Et₂O in hexane) 0.34;

δ_H (500 MHz, CDCl₃) 7.36-7.25 (3H, m), 7.21-7.16 (2H, m), 5.51-5.38 (1H, ddd, *J* = 3.4, 11.1, 15.5), 5.34-5.24 (1H, ddd, *J* = 3.4, 11.1, 15.5), 4.92 (1H, d, *J* = 15.3), 4.41 (1H, d, *J* = 15.3), 3.87 (1H, td, *J* = 1.5, 11.3), 2.68-2.63 (1H, m), 2.61 (1H, td, *J* = 1.5, 11.3), 2.42-2.34 (2H, m), 2.34-2.25 (1H, m), 2.13-2.00 (2H, m), 1.64-1.52 (1H, m), 1.45-1.34 (1H, m);
 δ_C (125 MHz, CDCl₃) 136.36, 135.89, 134.28, 128.86, 128.55, 127.97, 87.22, 65.55, 52.76, 37.85, 35.74, 33.52, 32.60;

ν_{max} (solid)/cm⁻¹ 2934.65, 2919.09, 2854.46, 1497.02, 1454.83, 1438.84;

m/z (EI) 242.1662 (M+H⁺. C₁₅H₂₀N₃ requires 242.1652).

PEG adduct 9a

rel-2-(2-(2-((3a*R*,9a*S*,p*R*)-3a,4,5,8,9,9a-hexahydro-1*H*-cycloocta[*d*][1,2,3]triazol-1-yl)ethoxy)-ethoxy)ethoxyethanamine

To (*E,E*)-1,5-cyclooctadiene (345 μ L, 0.2 molar in DCM, 69 μ mol, 1.1 eq) was added 2-(2-(2-azidoethoxy)ethoxy)ethoxyethanamine (13.7 mg, 63 μ mol, 1.0 eq) with stirring at r.t. When the reaction had finished (typically 2 h, TLC), the solvent and excess (*E,E*)-1,5-cyclooctadiene were

removed by rotary evaporation giving the desired product (20.5 mg, 63 μ mol, 100% yield) as a clear oil.

Rf=0.20 (10% MeOH/1% TEA/DCM);

δ_H (500 MHz, CDCl₃) 5.55-5.40 (2H, m), 3.84 (1H, t, *J* = 11.1), 3.77-3.55 (11H, m), 3.55-3.44 (1H, m), 3.49 (2H, t, *J* = 5.3), 2.97 (1H, app t, *J* = 11.1), 2.85 (2H, t, *J* = 5.3), 2.70 (1H, dd, *J* = 4.6, 13.8), 2.50-2.39 (2H, m), 2.39-2.29 (1H, m), 2.26-2.13 (2H, m), 1.72-1.54 (4H, m);
 δ_C (125 MHz, CDCl₃) 135.69, 134.45, 86.77, 73.45, 70.74, 70.72, 70.51, 70.39, 69.56, 67.39, 48.09, 41.84, 37.83, 36.07, 33.51, 32.65;

ν_{max} (film)/cm⁻¹ 3428.61, 2940.31, 1725.74;

m/z (EI) 327.2401(M+H⁺. C₁₆H₃₁N₄O₃ requires 327.2391).

PEG adduct 9b

***rel*-32-((3aS,9aS,pR)-3a,4,5,8,9,9a-hexahydro-1*H*-cycloocta[*d*][1,2,3]triazol-1-yl)-3,6,9,12,15,-18,21,24,27,30-decaoxadotriacontan-1-amine**

To (*E,E*)-1,5-cyclooctadiene (300 μ L, 0.2 M in DCM, 60 μ mol, 1.2 eq) was added 32-azido-3,6,9,12,15,18,21,24,27,30-decaoxadotriacontan-1-amine (26.3 mg, 50 μ mol, 1 eq) with stirring at room temperature. When the reaction had finished (typically 2 h, TLC), the solvent and excess (*E,E*)-1,5-cyclooctadiene were removed by rotary evaporation giving the desired product (31.5 mg, 50 μ mol, 99% yield) as a clear oil.

Rf=0.26 (10% MeOH/1% TEA/DCM);

δ_H (500 MHz, CDCl₃) 5.53-5.38 (2H, m), 3.83 (1H, td, *J* = 1.5, 11.3), 3.66-3.57 (40H, m), 3.55 (2H, t, *J* = 5.2), 3.53-3.44 (1H, m), 2.97 (1H, td, *J* = 1.3, 11.0), 2.88 (2H, t, *J* = 1.5, 11.3), 2.76-2.62 (4H, m), 2.49-2.38 (2H, m), 2.38-2.28 (1H, m), 2.24-2.12 (2H, m), 1.70-1.50 (2H, m);

δ_C (125 MHz, CDCl₃) 135.69, 134.45, 86.78, 72.40, 70.71, 70.66, 70.65, 70.62, 70.61, 70.57, 70.55, 70.50, 70.32, 69.59, 67.39, 48.09, 41.66, 37.82, 36.06, 33.51, 32.65;

ν_{max} (film)/cm⁻¹ 3428.61, 2940.31, 1725.74;

m/z (EI) 635.4244 (M+Na⁺. C₃₀H₅₉N₄O₁₀ requires 635.4226).

TMDIBO-PEG adduct 11

To Carbonic acid-2',3',2'',3''-tetramethoxy-7,8-didehydro-1,2:5,6-dibenzocycloocta-1,5,7-triene-3-yl 4-nitrophenyl ester² (20.7 mg, 41 μmol, 1.0 eq) and PEG adduct **9b** (26.0 mg, 41 μmol, 1.0 eq) in CH₂Cl₂ (5 mL) was added NEt₃ (29 μL, 205 μmol, 5.0 eq) with stirring at room temperature. When the reaction had finished (typically 4 h, TLC), the solvent was removed by rotary evaporation followed by column chromatography (1% MeOH/DCM) giving the desired product (35.0 mg, 35 μmol, 85% yield) as a clear oil.

Rf=0.15 (10% MeOH/DCM);

δ_H (500 MHz, CDCl₃) 7.03 (1H, s), 6.87 (1H, s), 6.80 (1H, s), 6.79 (1H, s), 5.62-5.38 (3H, m), 3.90 (6H, s), 3.86 (3H, s), 3.85 (3H, s), 3.84 (1H, td, *J* = 1.5, 11.3), 3.73-3.55 (42H, m), 3.52-3.47 (1H, m), 3.41-3.37 (1H, m), 3.06 (1H, dd, *J* = 2.0, 15.0), 2.97 (1H, td, *J* = 1.3, 11.0), 2.80 (1H, dd, *J* = 3.6, 15.0), 2.73-2.65 (1H, m), 2.49-2.38 (2H, m), 2.38-2.28 (1H, m), 2.24-2.12 (2H, m), 2.01-1.90 (1H, br s), 1.70-1.50 (2H, m);

δ_C (125 MHz, CDCl₃) 155.56, 149.06, 148.82, 147.86, 147.82, 145.78, 144.57, 135.69, 134.45, 115.67, 113.99, 113.26, 112.52, 109.41, 109.39, 109.16, 108.81, 108.20, 86.78, 70.71, 70.65, 70.50, 70.36, 70.21, 69.59, 67.39, 56.20, 56.15, 48.09, 46.53, 41.05, 37.83, 36.06, 33.50, 32.65;

ν_{max} (film)/cm⁻¹ 2862.81, 2335.41, 1721.16, 1601.90, 1561.42, 1504.90;

m/z (EI) 1023.5143 (M+Na⁺. C₅₁H₇₆O₁₆N₄ requires 1023.5149).

Benzyl azide-tetrazine bisadduct 10

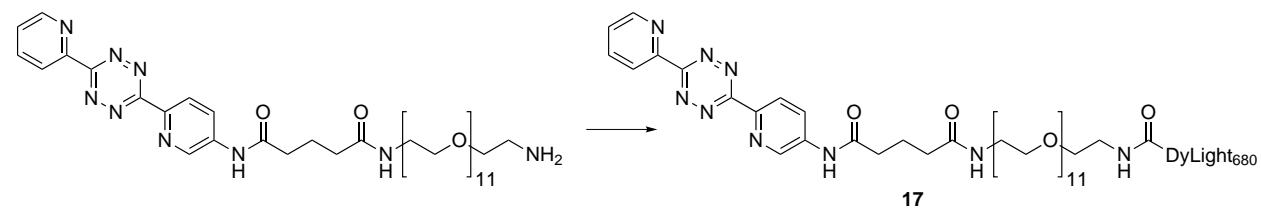
rel-(3aS,5aS,9aS,11aS)-1-benzyl-6,9-di(pyridin-2-yl)-3a,4,5,5a,9a,10,11,11a-octahydro-1*H*-[1,2,3]-triazolo[4',5':5,6]cycloocta[1,2-*d*]pyridazine

Benzyl azide monoadduct (**8**) (20.0 mg, 83 μmol, 1.0 eq) was dissolved in DCM (800 μl) and 3,6-di(pyridin-2-yl)-1,2,4,5-tetrazine (19.6 mg, 83 μmol, 1.0 eq) was added with stirring. The solvent was removed by rotary evaporation to give the desired product (37.0 mg, 82 μmol, 99% yield) as a yellow solid, mp 185-187 °C.

R_f (10% MeOH in DCM) 0.54;

δ_H (500 MHz, CDCl₃) 8.56 (1H, app d, J = 4.7), 8.49 (1H, app d, J = 4.7), 8.27 (1H, app d, J = 8.0), 8.25 (1H, app d, J = 8.0), 7.69 (2H, app tt, J = 2.1, 7.7), 7.32-7.22 (7H, m), 4.73 (1H, d, J = 15.4), 4.61 (1H, d, J = 15.4), 4.61 (1H, app t, J = 11.5), 3.65 (1H, dd, J = 3.4, 8.5), 3.47 (1H, dd, J = 3.4, 8.5), 3.35 (1H, app td, J = 2.1, 11.5), 2.74-2.66 (1H, app dd, J = 4.1, 15.2), 2.11-1.95 (2H, m), 1.91-1.80 (3H, m), 1.64-1.52 (1H, m), 1.44-1.32 (1H, m);
 δ_C (125 MHz, CDCl₃) 161.72, 161.71, 154.13, 154.12, 149.08, 149.01, 136.66, 136.57, 128.77, 128.48, 127.77, 125.02, 124.97, 123.03, 123.01, 83.76, 65.16, 53.39, 35.69, 35.27, 34.63, 33.97, 33.78;
 ν_{max} (solid)/cm⁻¹ 3062.08, 2956.11, 2929.45, 2850.02, 1599.69, 1562.51, 1549.56, 1489.16, 1464.54, 1455.94, 1440.16, 1429.25;
 m/z (EI) 450.2417 ($M+H^+$. C₂₇H₂₈N₇ requires 450.2401).

Tetrazine **12**



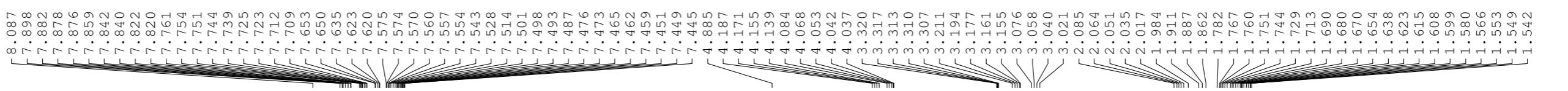
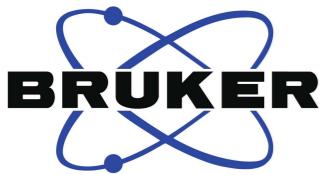
The precursor to tetrazine **12** was prepared as previously described.³

*N*1-(35-amino-3,6,9,12,15,18,21,24,27,30,33-undecaoxapentatriacontyl)-*N*5-(6-(6-(pyridin-2-yl)-1,2,4,5-tetrazin-3-yl)pyridin-3-yl)glutaramide (1.9 mg, 2 μ mol, 2.0 eq) was dissolved in DMF (200 μ L) followed by the addition of DyLight680 NHS ester (1 mg, 1 μ mol, 1.0 eq) and Hunig's Base (1.0 μ L, 5 μ mol, 5.0 eq). The solution was stirred overnight at room temperature, concentrated and purified by HPLC to give the desired product (analysis by LC-MS).

m/z (ES) 1683.6 ($M+H^+$).

References

- (1) Newton, P. F.; Whitham, G. H. *J.C.S. Perkin I* **1979**, 3067–3071.
- (2) Stöckmann, H.; Neves, A. A.; Stairs, S.; Ireland-Zecchini, H.; Brindle, K. M.; Leeper, F. J. *Chem. Sci.* **2011**, DOI: 10.1039/C0SC00631A.
- (3) Rossin, R.; Verkerk, P. R.; van den Bosch, S. M.; Vulders, R. C. M.; Verel, I.; Lub, J.; Robillard, M. S. *Angew Chem Int Ed Engl* **2010**, 49, 3375–3378.

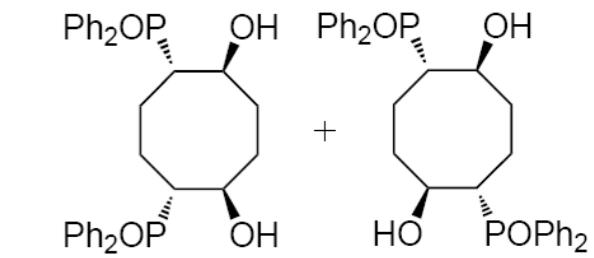
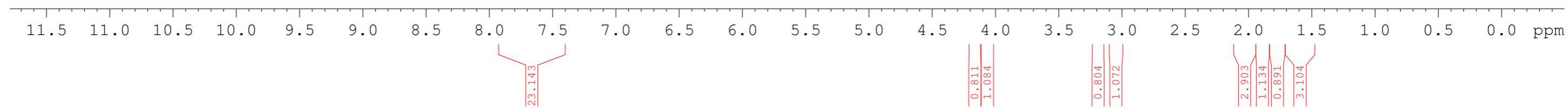


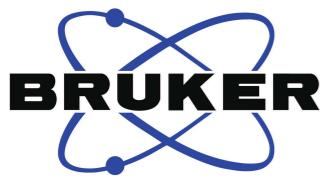
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PULPROG zg30
TD 65536
SOLVENT MeOD
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0500124 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00





Current Data Parameters

NAME fjl-had8

EXPNO 11

PROCNO 1

F2 - Acquisition Parameters

Date_ 20110207

Time 14.39

INSTRUM drx500

PROBHD 5 mm CPDUL Z-

PULPROG zgpg30

TD 65536

SOLVENT MeOD

NS 256

DS 8

SWH 34013.605 Hz

FIDRES 0.519006 Hz

AQ 0.9634292 sec

RG 4096

DW 14.700 usec

DE 6.00 usec

TE 300.0 K

D1 3.00000000 sec

d11 0.03000000 sec

DELTA 2.90000010 sec

TD0 1

===== CHANNEL f1 =====

NUC1 13C

P1 9.80 usec

PL1 -6.00 dB

SFO1 125.7520828 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16

NUC2 1H

PCPD2 100.00 usec

PL2 -6.00 dB

PL12 10.62 dB

PL13 15.00 dB

SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536

SF 125.7374972 MHz

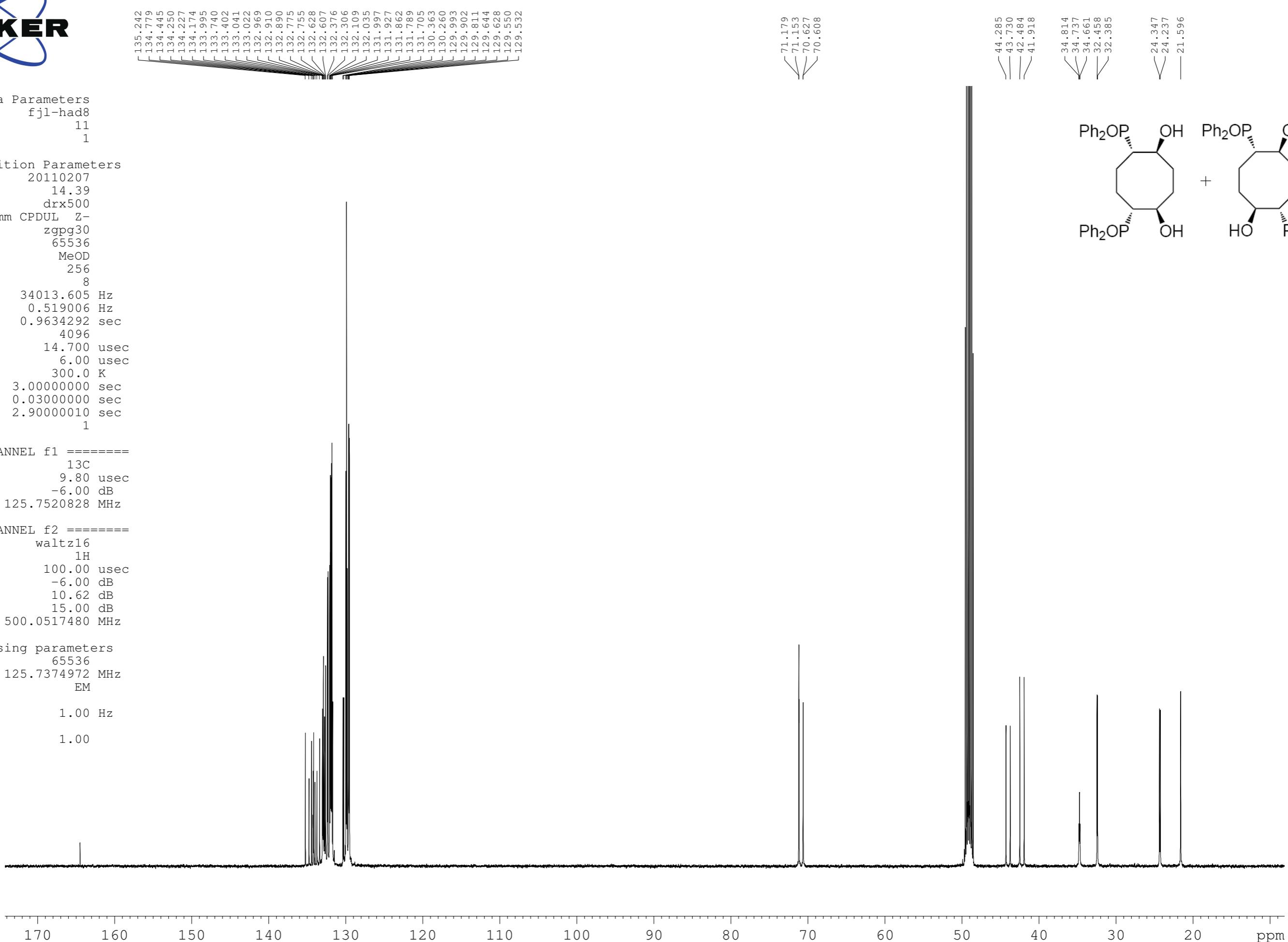
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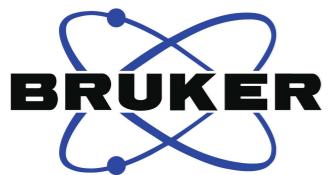
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LB 1.00 Hz

GB 0

PC 1.00



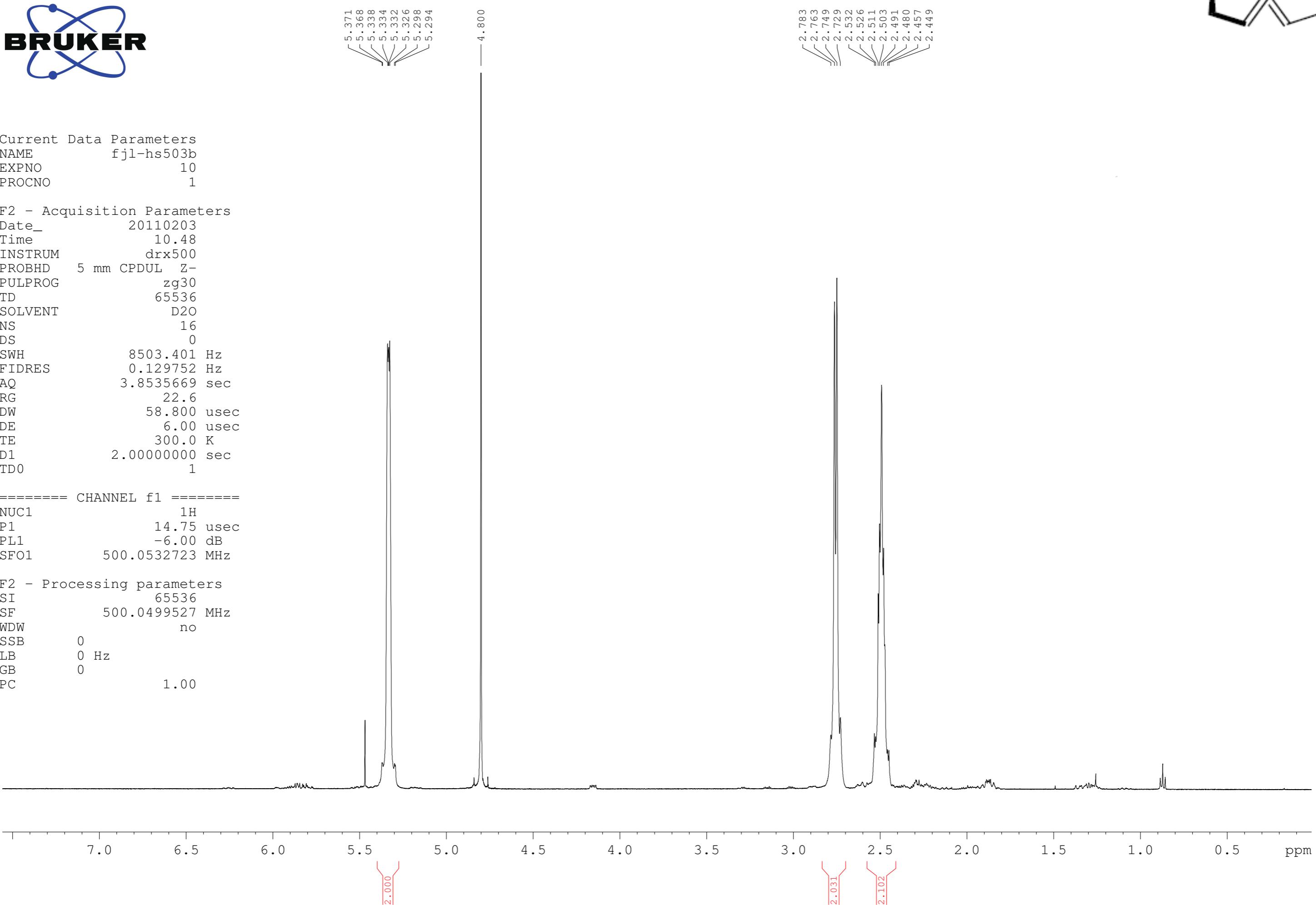


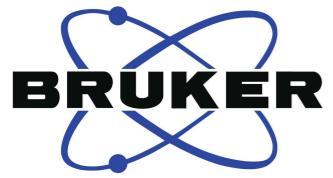
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PULPROG zg30
TD 65536
SOLVENT D2O
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 22.6
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0499527 MHz
WDW no
SSB 0
LB 0 Hz
GB 0
PC 1.00





Current Data Parameters

NAME fjl-hs503b
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110203
Time 11.06
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT D2O
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.0000000 sec
d11 0.0300000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

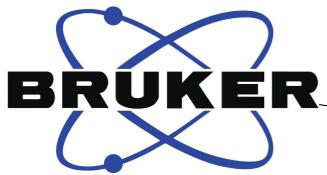
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LB 0 Hz
GB 0
PC 1.00

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— 31.848 —





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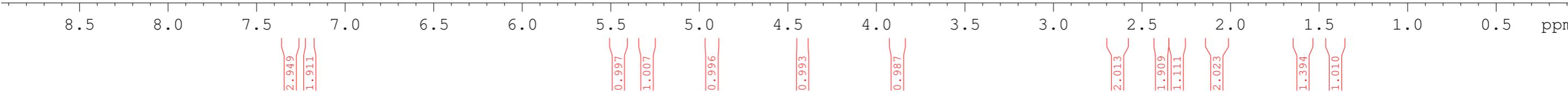
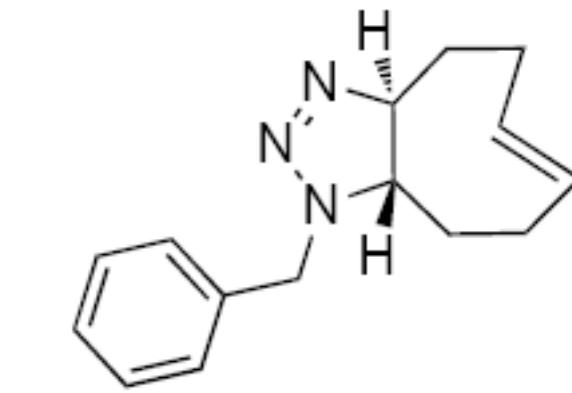
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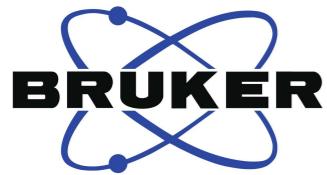
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EXPNO 20
PROCNO 1

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PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 ======
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0500257 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.50





Current Data Parameters
NAME fjl-hs511
EXPNO 21
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110202
Time 11.17
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.0000000 sec
d11 0.0300000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

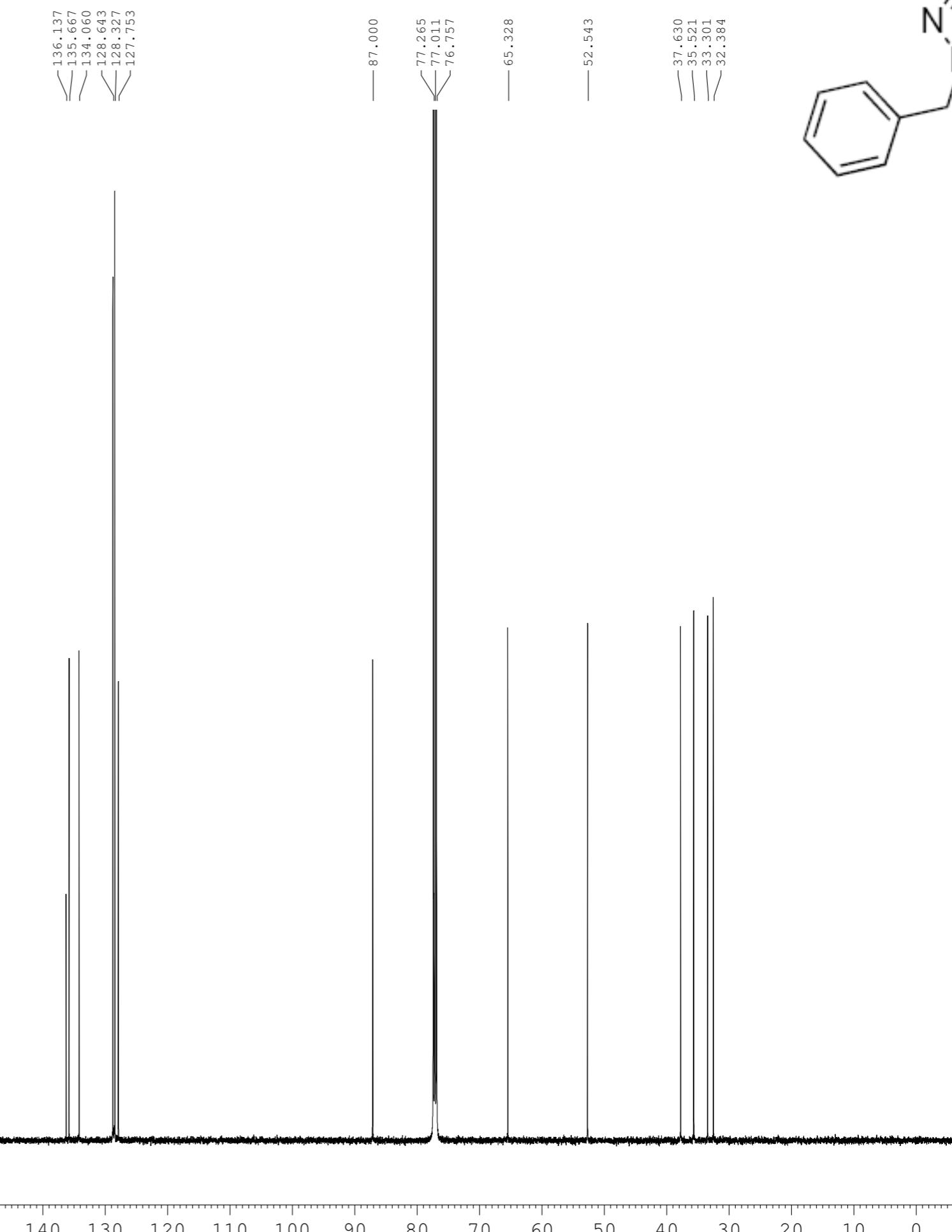
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

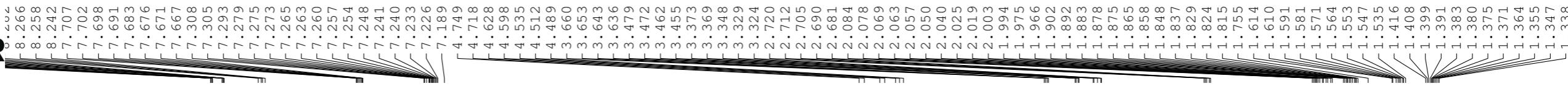
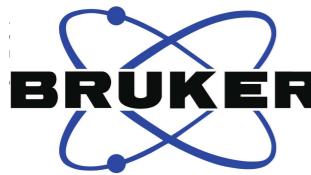
===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536
SF 125.7376590 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 2.00



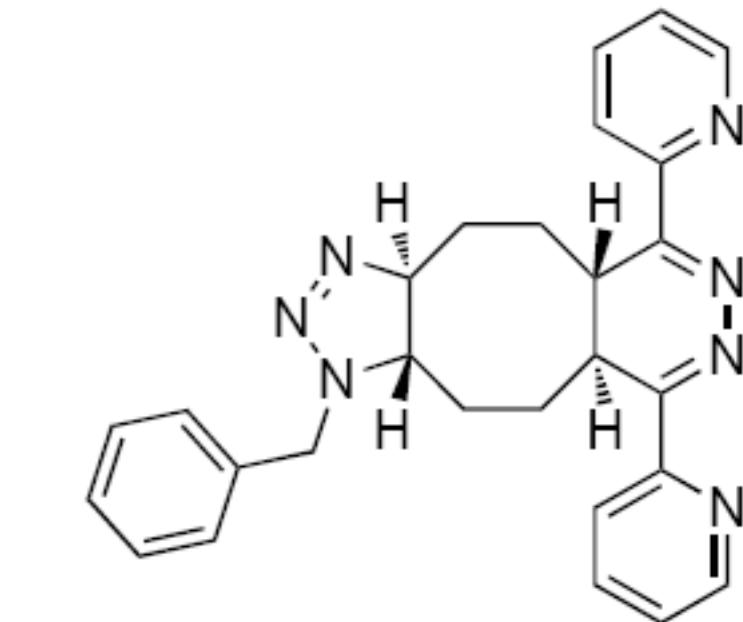
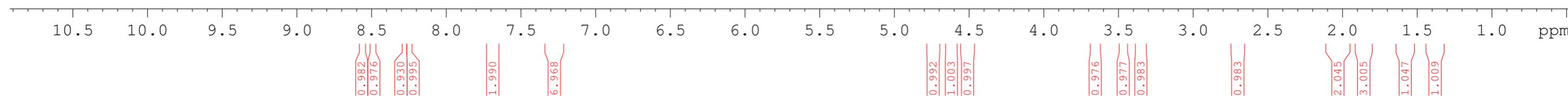


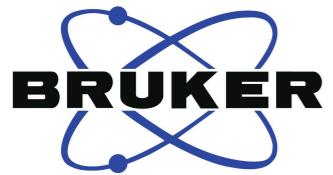
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NAME fjl-hs515
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110203
Time 11.34
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0500512 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00





Current Data Parameters
NAME fjl-hs515
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110203
Time 11.52
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.0000000 sec
d11 0.0300000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

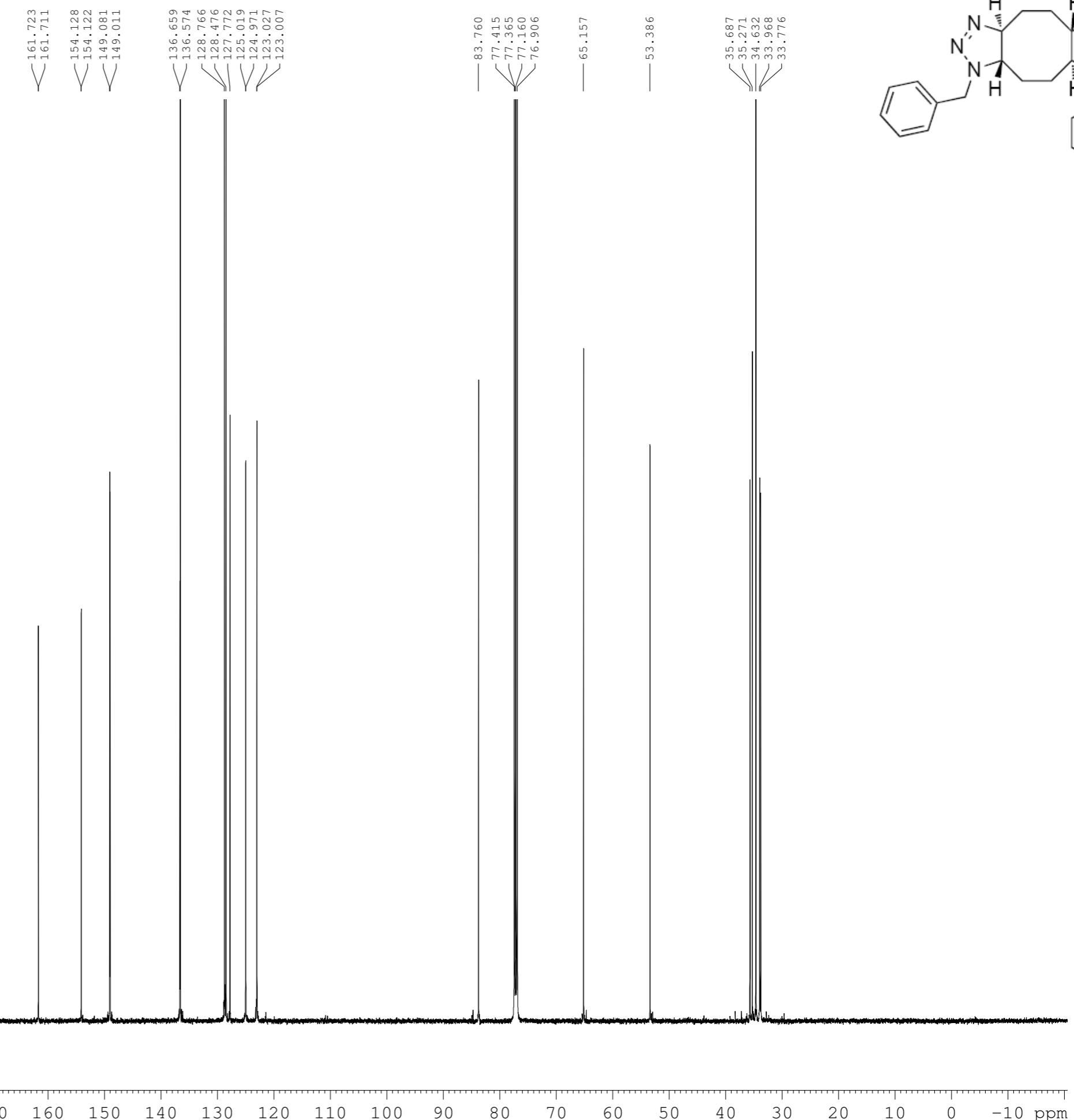
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

===== CHANNEL f2 =====

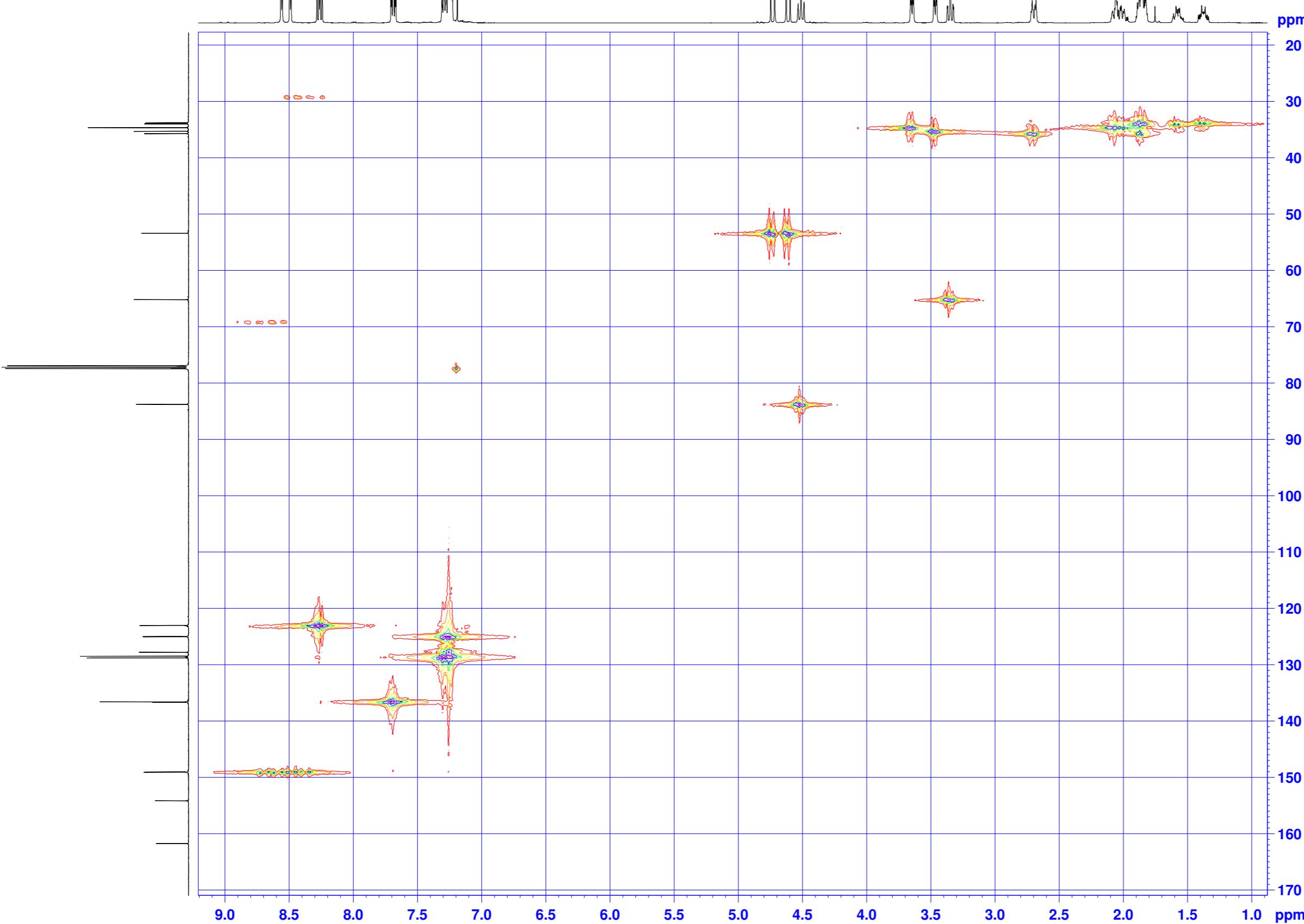
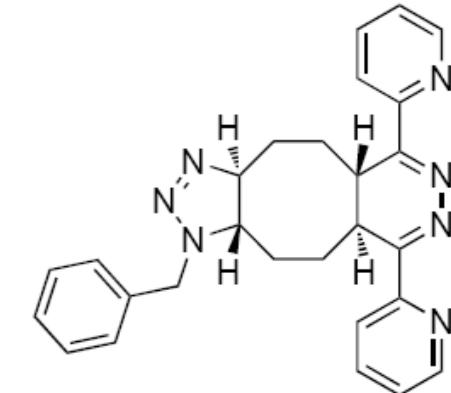
CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536
SF 125.7376619 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



HS515



Time 12.03
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG hmqcgpf
TD 2048
SOLVENT CDCl3
NS 2
DS 16
SWH 6009.615 Hz
FIDRES 2.934382 Hz
AQ 0.1704436 sec
RG 2048
DW 83.200 usec
DE 6.00 usec
TE 300.0 K
CNST2 145.000000
d0 0.0000300 sec
D1 2.0000000 sec
d2 0.00344828 sec
d12 0.00002000 sec
d13 0.00000400 sec
D16 0.00010000 sec
DELTA1 0.00232428 sec
IN0 0.00001990 sec

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
p2 29.50 usec
PL1 -6.00 dB
SFO1 500.0525002 MHz

===== CHANNEL f2 =====
CPDPRG2 garp
NUC2 13C
P3 9.25 usec
PCPD2 80.00 usec
PL2 -6.00 dB
PL12 12.74 dB
SFO2 125.7476820 MHz

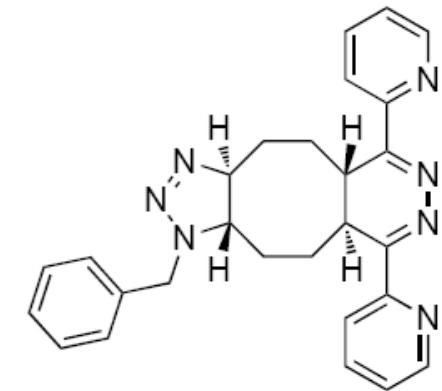
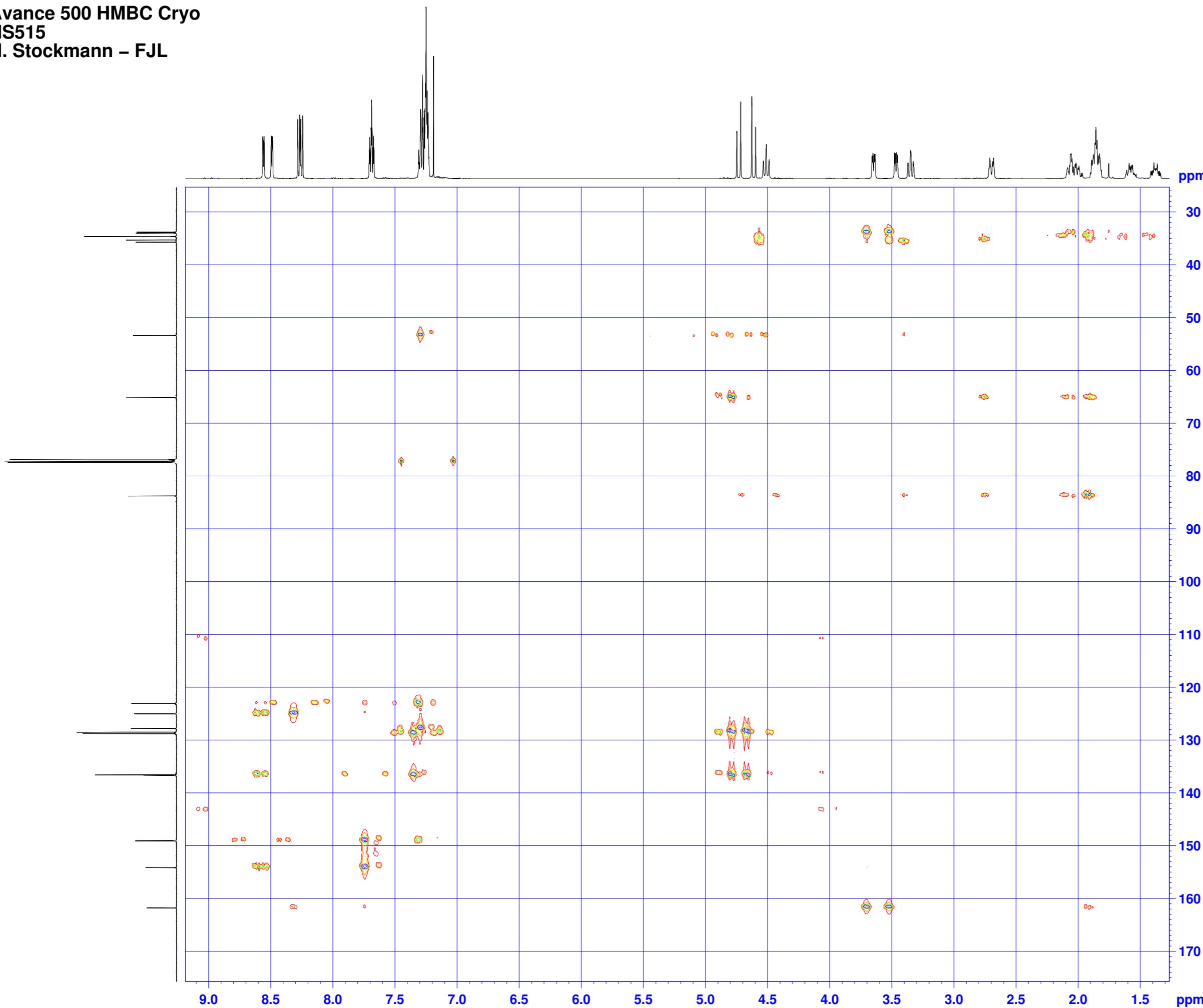
===== GRADIENT CHANNEL =====
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GPNAM2 SINE.100
GPNAM3 SINE.100
GPZ1 50.00 %
GPZ2 30.00 %
GPZ3 40.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 125.7477 MHz
FIDRES 98.146988 Hz
SW 199.810 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 500.0500440 MHz
WDW SINE
SSB 4
LB 0 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 QF
SF 125.7376451 MHz
WDW States
SSB 4
LB 0 Hz
GB 0

Avance 500 HMBC Cryo
HS515
H. Stockmann – FJL



Time 21.42
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG hmbcgplndqf
TD 2048
SOLVENT CDCl₃
NS 2
DS 8
SWH 5482.456 Hz
FIDRES 2.676980 Hz
AQ 0.1868276 sec
RG 4096
DW 91.200 usec
DE 6.00 usec
TE 300.0 K
CNST2 145.000000
CNST13 10.000000
d0 0.00000300 sec
D1 2.0000000 sec
d2 0.00344828 sec
d6 0.05000000 sec
D16 0.00010000 sec
INO 0.00001730 sec

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
p2 29.50 usec
PL1 -6.00 dB
SFO1 500.0525002 MHz

===== CHANNEL f2 =====
NUC2 13C
P3 9.25 usec
PL2 -6.00 dB
SFO2 125.7508255 MHz

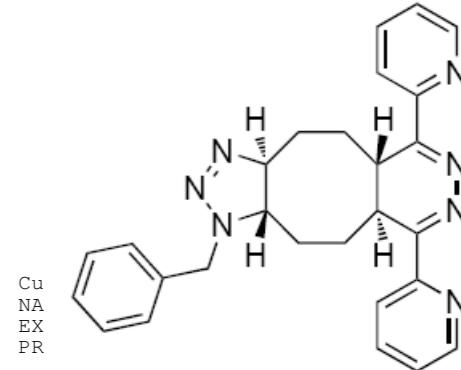
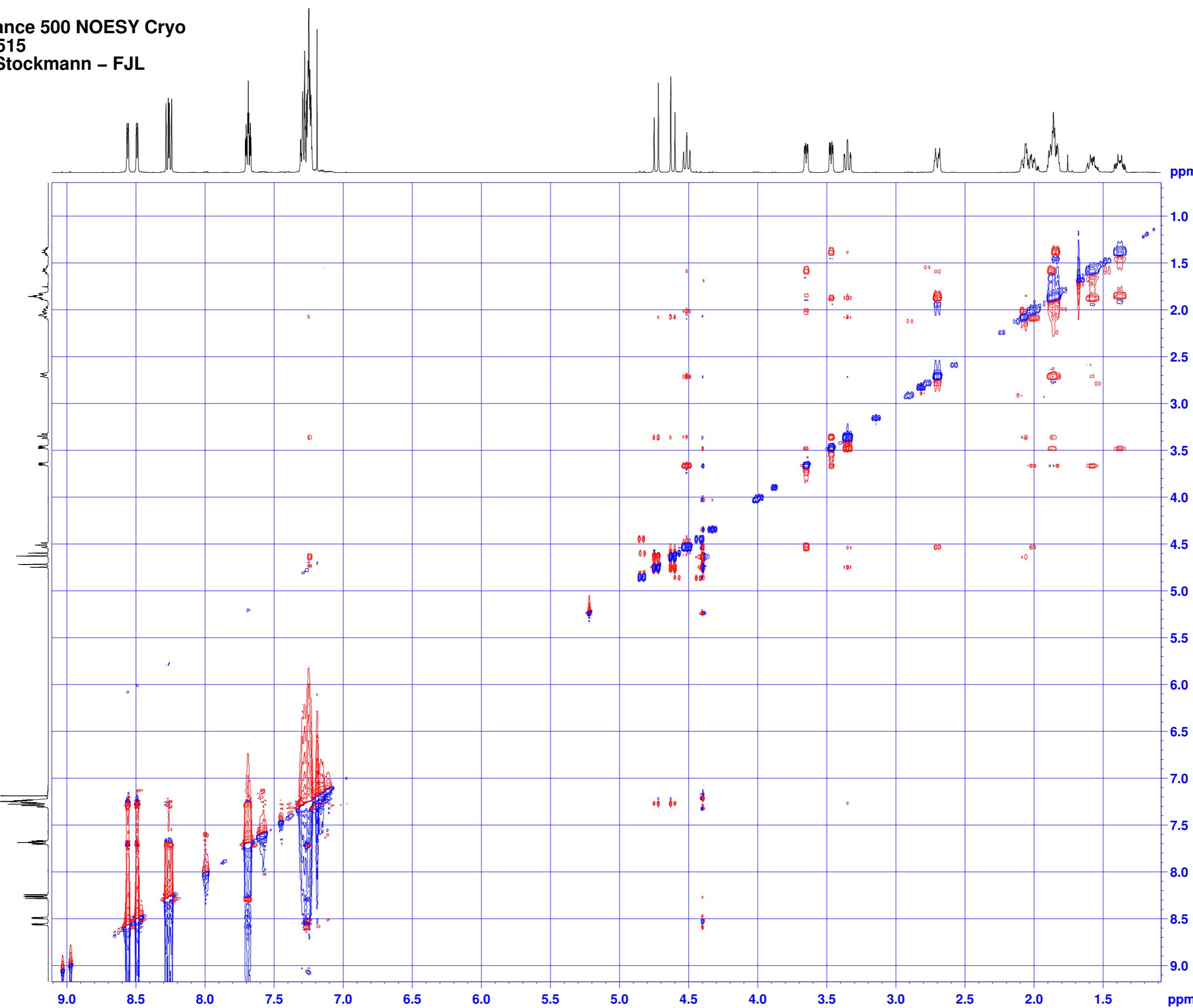
===== GRADIENT CHANNEL =====
GPNAM1 SINE.100
GPNAM2 SINE.100
GPNAM3 SINE.100
GPZ1 50.00 %
GPZ2 30.00 %
GPZ3 40.10 %
P16 1000.00 usec

F1 - Acquisition parameters
TD 256
SFO1 125.7508 MHz
FIDRES 112.897400 Hz
SW 229.833 ppm
FnMODE QF

F2 - Processing parameters
SI 1024
SF 500.0500219 MHz
WDW SINE
SSB 4
LB 0 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 QF
SF 125.7376786 MHz
WDW States
SSB 4
LB 0 Hz
GB 0

Avance 500 NOESY Cryo
HS515
H. Stockmann – FJL



Cu
NA
EX
PR

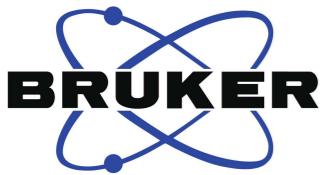
F2
Date_ 20110208
Time 22.03
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG noesyph
TD 2048
SOLVENT CDCl3
NS 8
DS 4
SWH 5000.000 Hz
FIDRES 2.441406 Hz
AQ 0.2048500 sec
RG 10.1
DW 100.000 usec
DE 6.00 usec
TE 300.0 K
d0 0.00008122 sec
D1 2.0000000 sec
D8 1.2000005 sec
INO 0.0002000 sec

===== CHANNEL f1 ======
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0522502 MHz

F1 - Acquisition parameters
TD 320
SFO1 500.0523 MHz
FIDRES 15.625000 Hz
SW 9.999 ppm
FnMODE TPPI

F2 - Processing parameters
SI 2048
SF 500.0500512 MHz
WDW QSINE
SSB 2.5
LB 0 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 2048
MC2 TPPI
SF 500.0500418 MHz
WDW States-TPPI
SSB 2.5
LB 0 Hz
GB 0



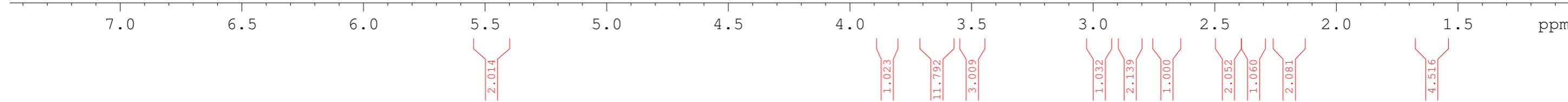
Current Data Parameters
NAME fjl-hs516-2
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110208
Time 12.19
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 ======

NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

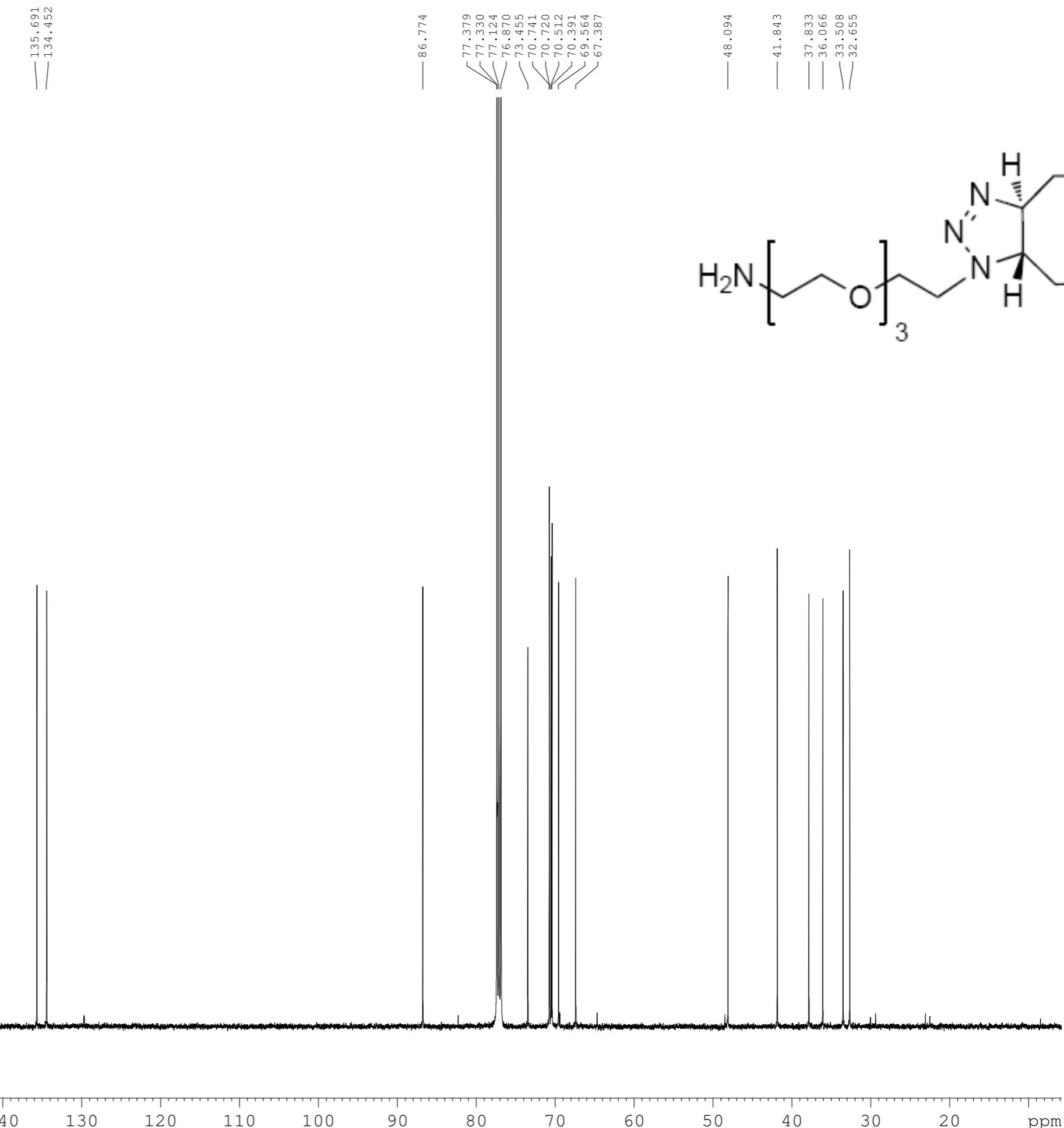
F2 - Processing parameters
SI 65536
SF 500.0500219 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00

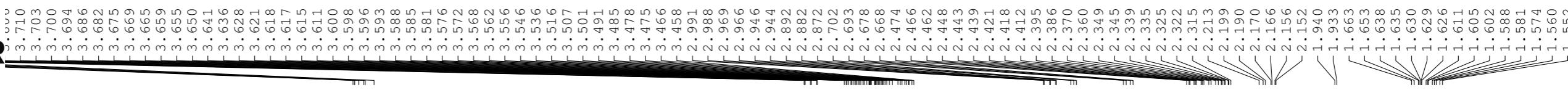
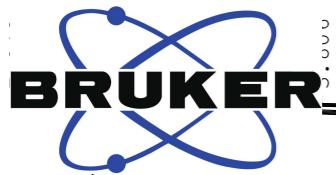


rance 500 13C Cryo

HS516-2

. Stockmann - FJL





Current Data Parameters

NAME fj1-hs519

EXPNO 10

PROCNO 1

F2 - Acquisition Parameters

Date 20110215

Time 9.42

INSTRUM drx500

PROBHD 5 mm CPDUL Z-

PULPROG zg30

TD 65536

SOLVENT CDCl₃

NS 16

DS 0

SWH 8503.401 Hz

FIDRES 0.129752 Hz

AQ 3.8535669 sec

RG 11.3

DW 58.800 usec

DE 6.00 usec

TE 300.0 K

D1 2.0000000 sec

TD0 1

===== CHANNEL f1 =====

NUC1 1H

P1 14.75 usec

PL1 -6.00 dB

SFO1 500.0532723 MHz

F2 - Processing parameters

SI 65536

SF 500.0500219 MHz

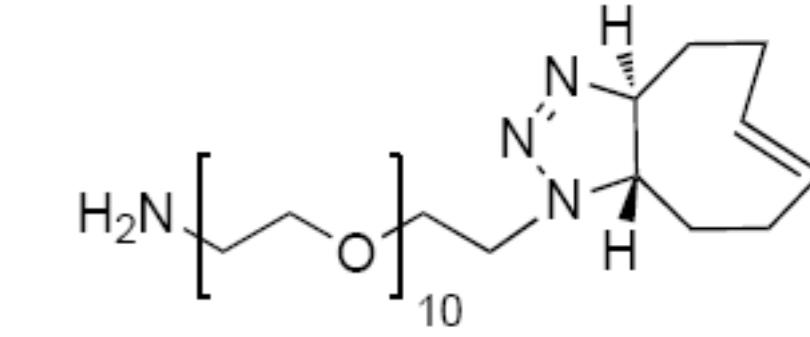
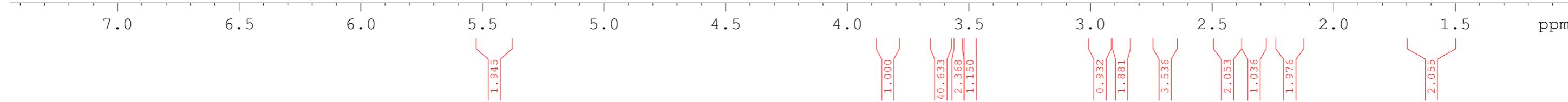
WDW EM

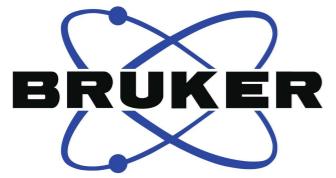
SSB 0

LB 0.10 Hz

GB 0

PC 1.00





Current Data Parameters

NAME fjl-hs519
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110215
Time 10.00
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

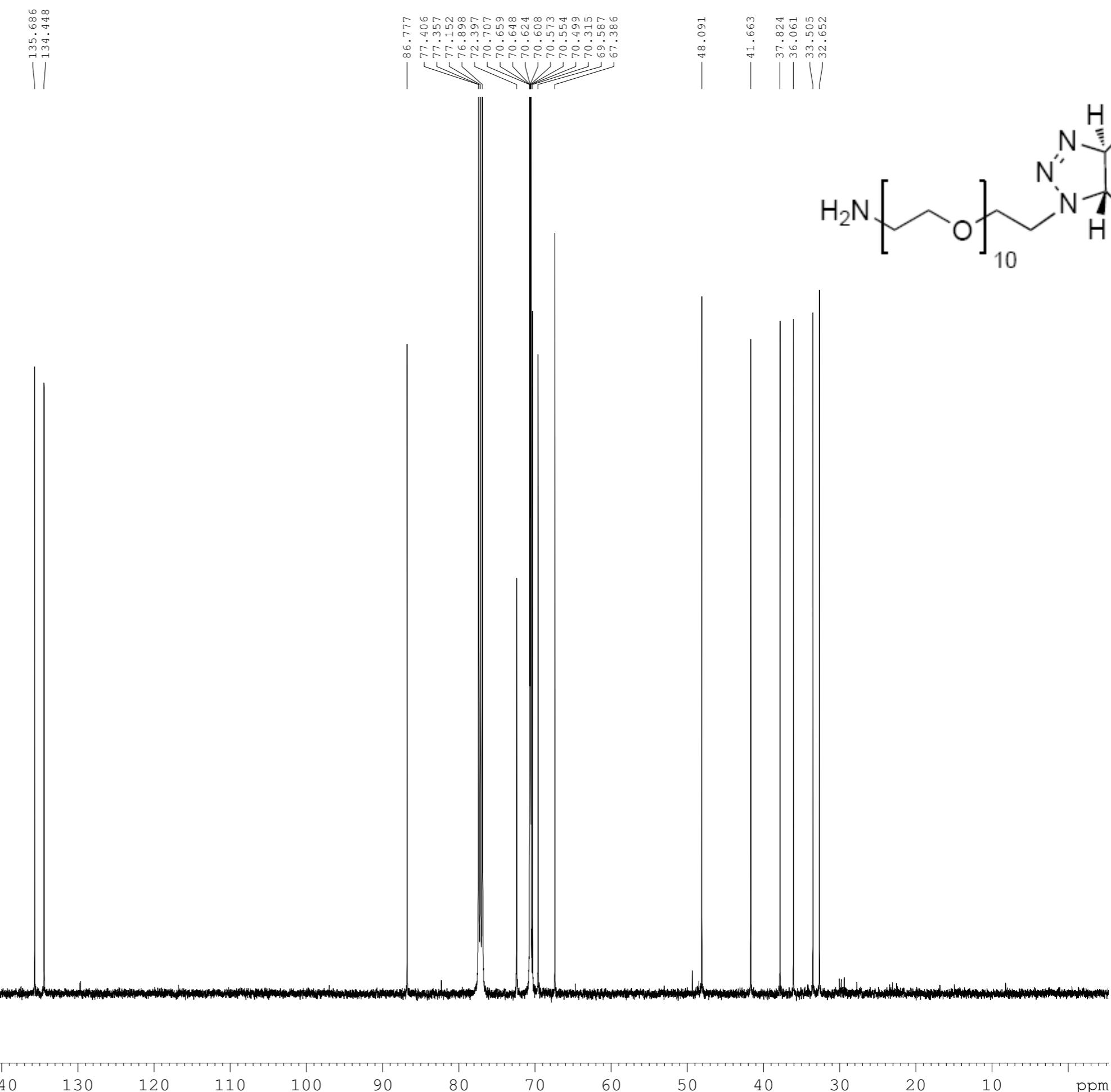
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

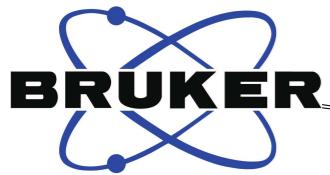
===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536
SF 125.7376627 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



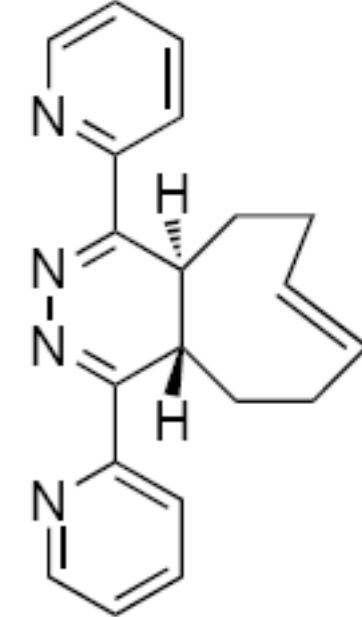
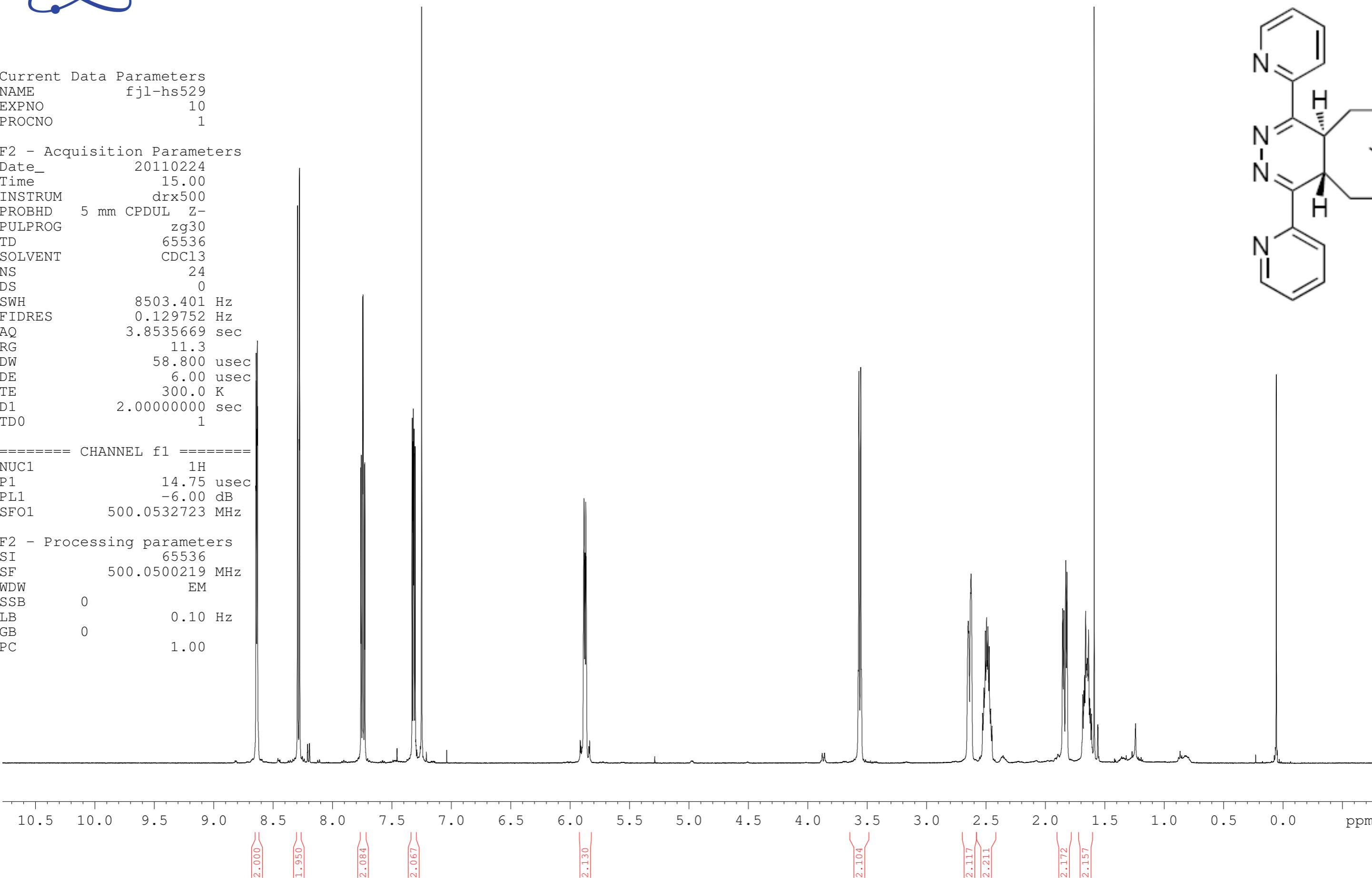


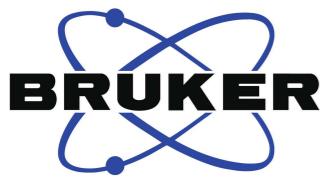
Current Data Parameters
NAME fj1-hs529
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110224
Time 15.00
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 24
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0500219 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00





Current Data Parameters

NAME fjl-hs529
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110224
Time 15.36
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 512
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

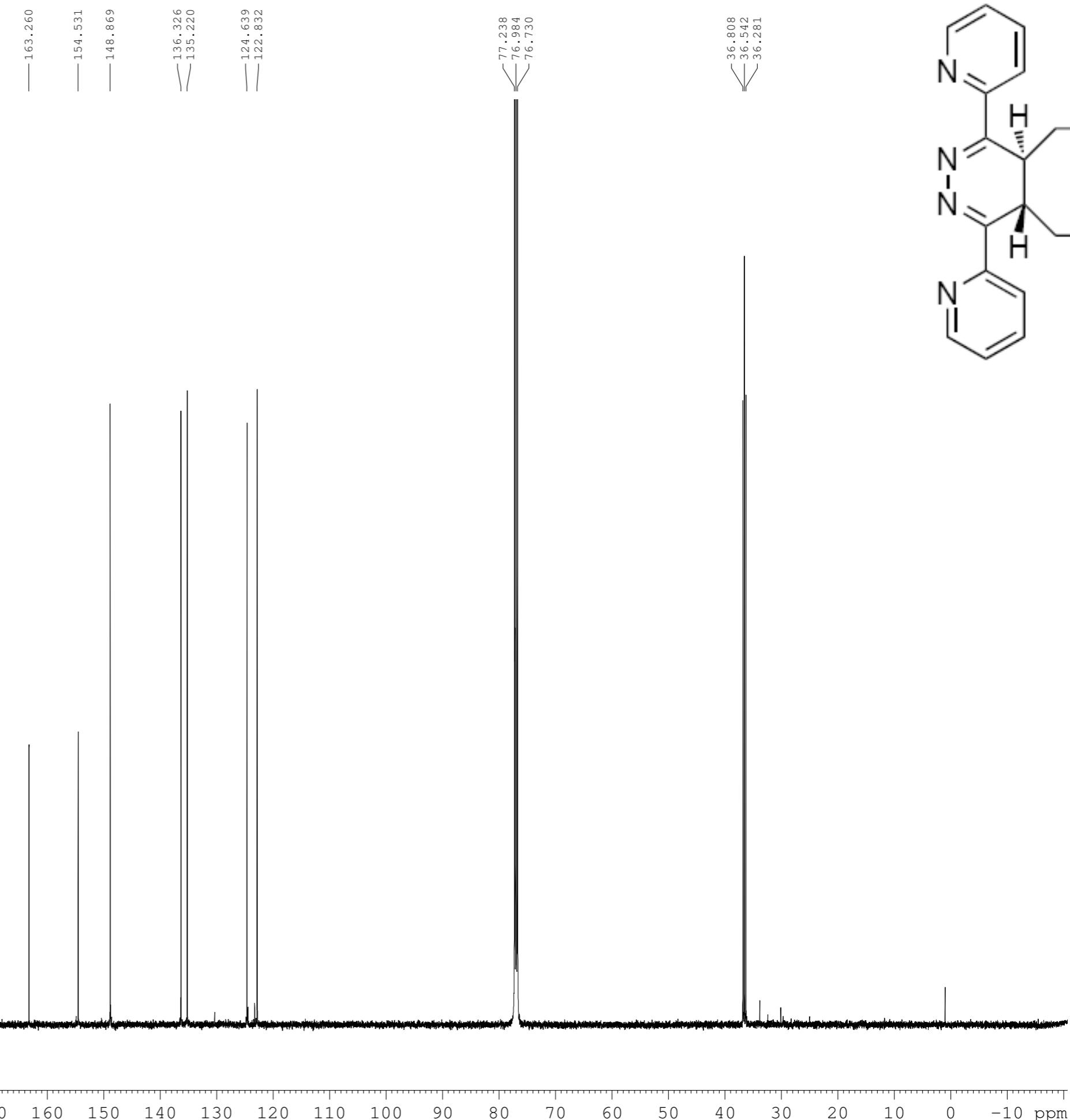
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

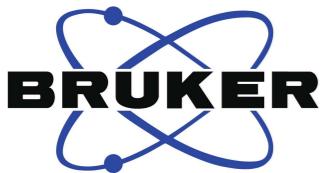
===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536
SF 125.7376786 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



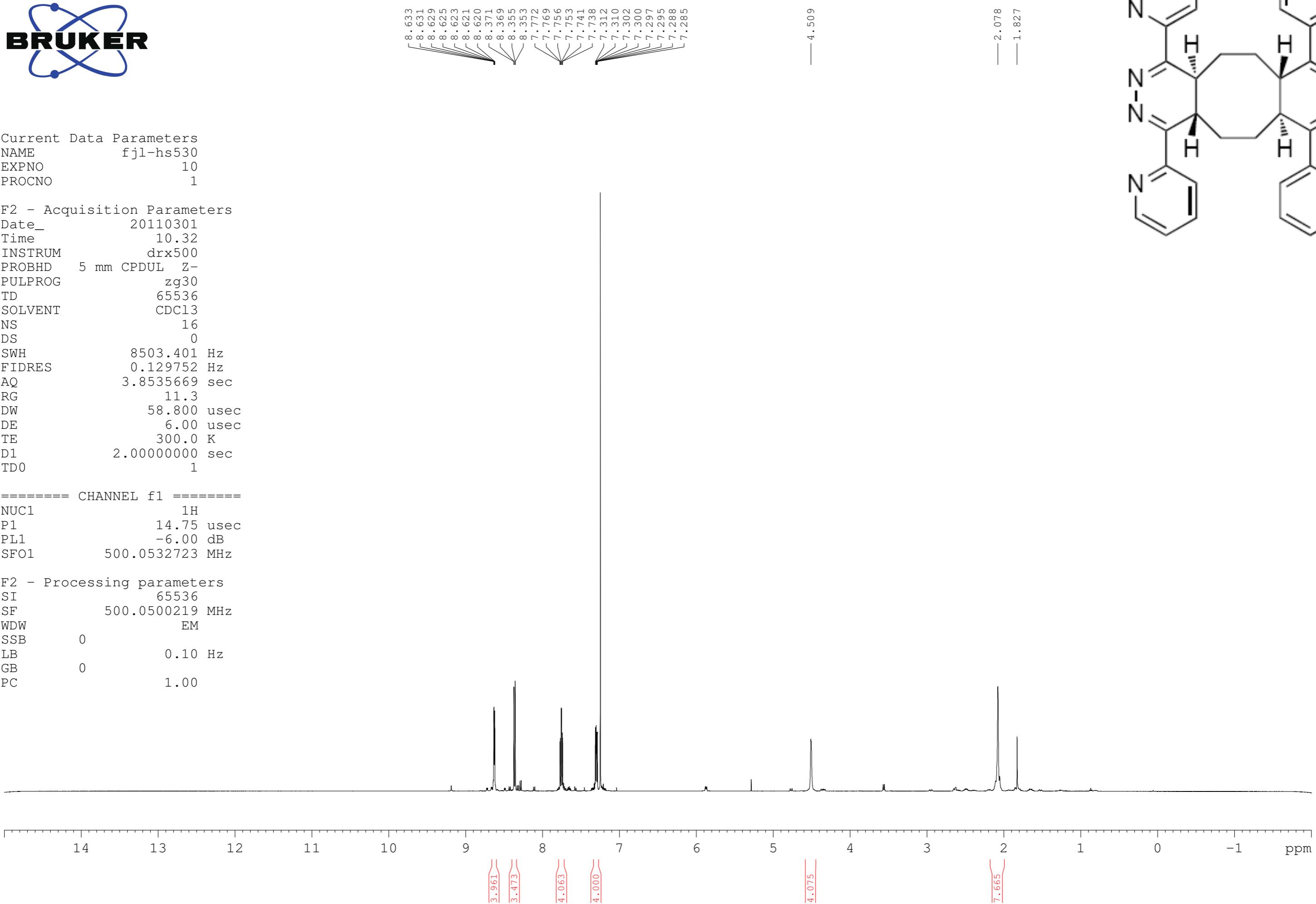


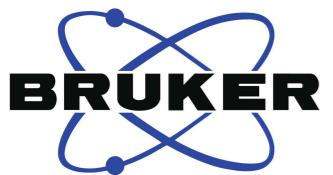
Current Data Parameters
NAME fj1-hs530
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters
Date_ 20110301
Time 10.32
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

===== CHANNEL f1 =====
NUC1 1H
P1 14.75 usec
PL1 -6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters
SI 65536
SF 500.0500219 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00





Current Data Parameters

NAME fjl-hs530
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110301
Time 10.50
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.0000000 sec
d11 0.0300000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

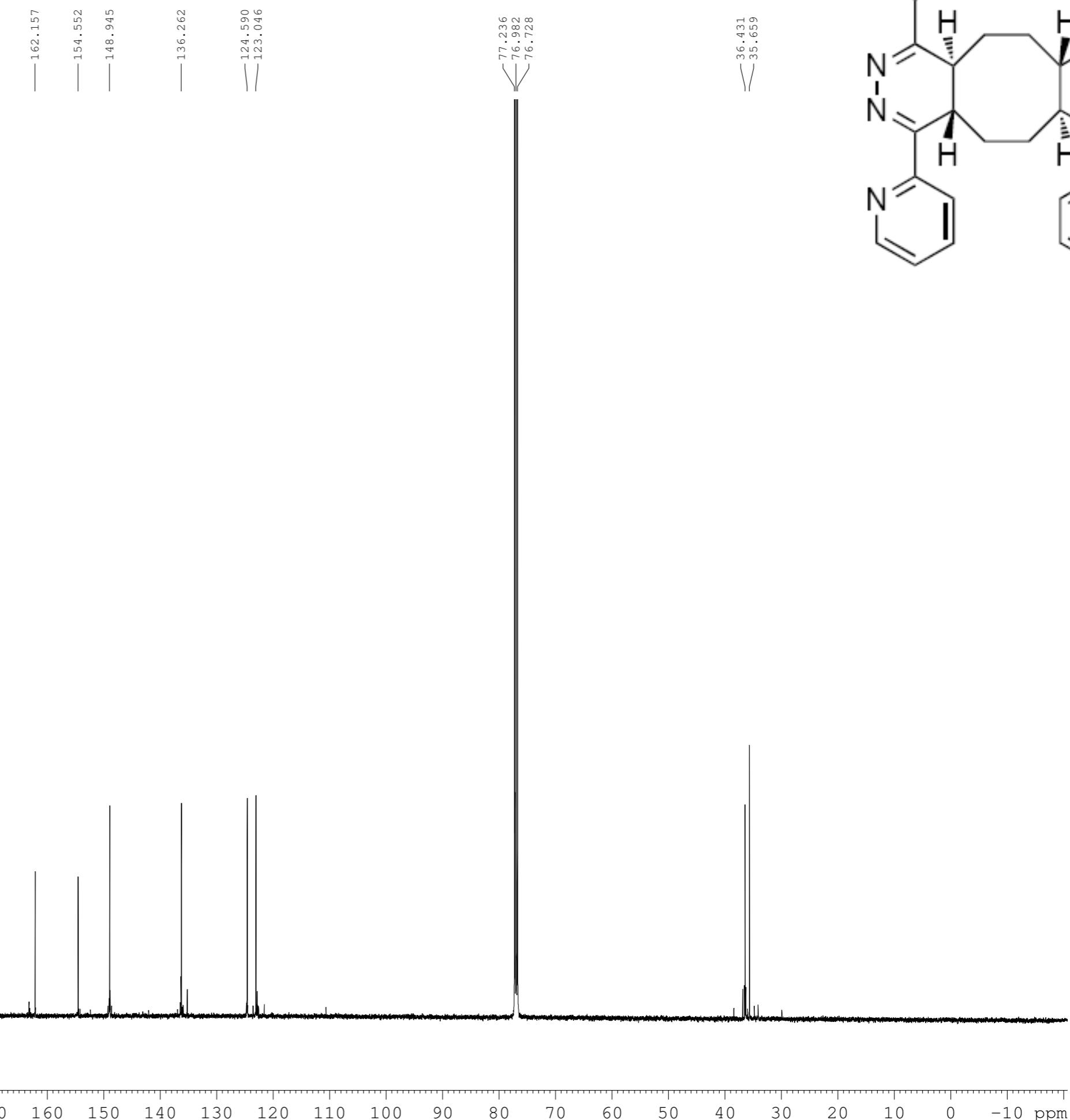
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

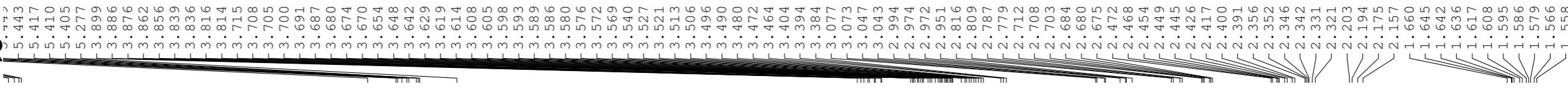
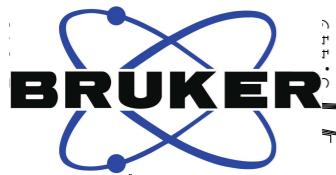
===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

SI 65536
SF 125.7376786 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00





Current Data Parameters
NAME fjl-hs525
EXPNO 10
PROCNO 1

F2 - Acquisition Parameters

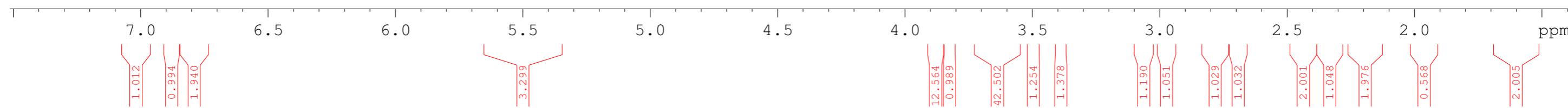
Date_ 20110216
Time 19.38
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zg30
TD 65536
SOLVENT CDCl3
NS 16
DS 0
SWH 8503.401 Hz
FIDRES 0.129752 Hz
AQ 3.8535669 sec
RG 11.3
DW 58.800 usec
DE 6.00 usec
TE 300.0 K
D1 2.0000000 sec
TD0 1

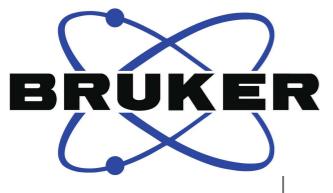
===== CHANNEL f1 =====

NUC1 1H
P1 14.75 usec
PL1 +6.00 dB
SFO1 500.0532723 MHz

F2 - Processing parameters

SI 65536
SF 500.0500219 MHz
WDW EM
SSB 0
LB 0.10 Hz
GB 0
PC 1.00





Current Data Parameters

NAME fjl-hs525
EXPNO 11
PROCNO 1

F2 - Acquisition Parameters

Date_ 20110216
Time 19.56
INSTRUM drx500
PROBHD 5 mm CPDUL Z-
PULPROG zgpg30
TD 65536
SOLVENT CDCl3
NS 256
DS 8
SWH 34013.605 Hz
FIDRES 0.519006 Hz
AQ 0.9634292 sec
RG 4096
DW 14.700 usec
DE 6.00 usec
TE 300.0 K
D1 3.00000000 sec
d11 0.03000000 sec
DELTA 2.90000010 sec
TD0 1

===== CHANNEL f1 =====

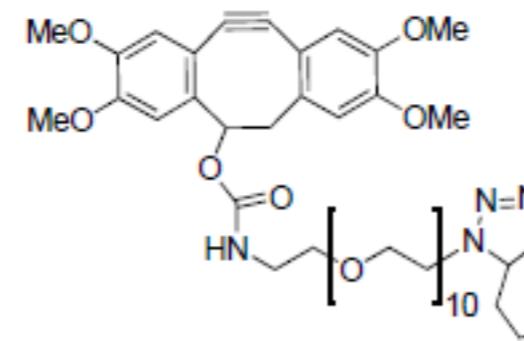
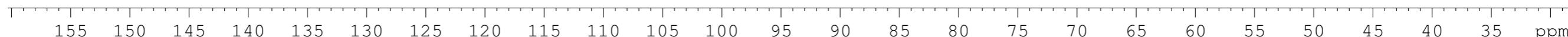
NUC1 13C
P1 9.80 usec
PL1 -6.00 dB
SFO1 125.7520828 MHz

===== CHANNEL f2 =====

CPDPRG2 waltz16
NUC2 1H
PCPD2 100.00 usec
PL2 -6.00 dB
PL12 10.62 dB
PL13 15.00 dB
SFO2 500.0517480 MHz

F2 - Processing parameters

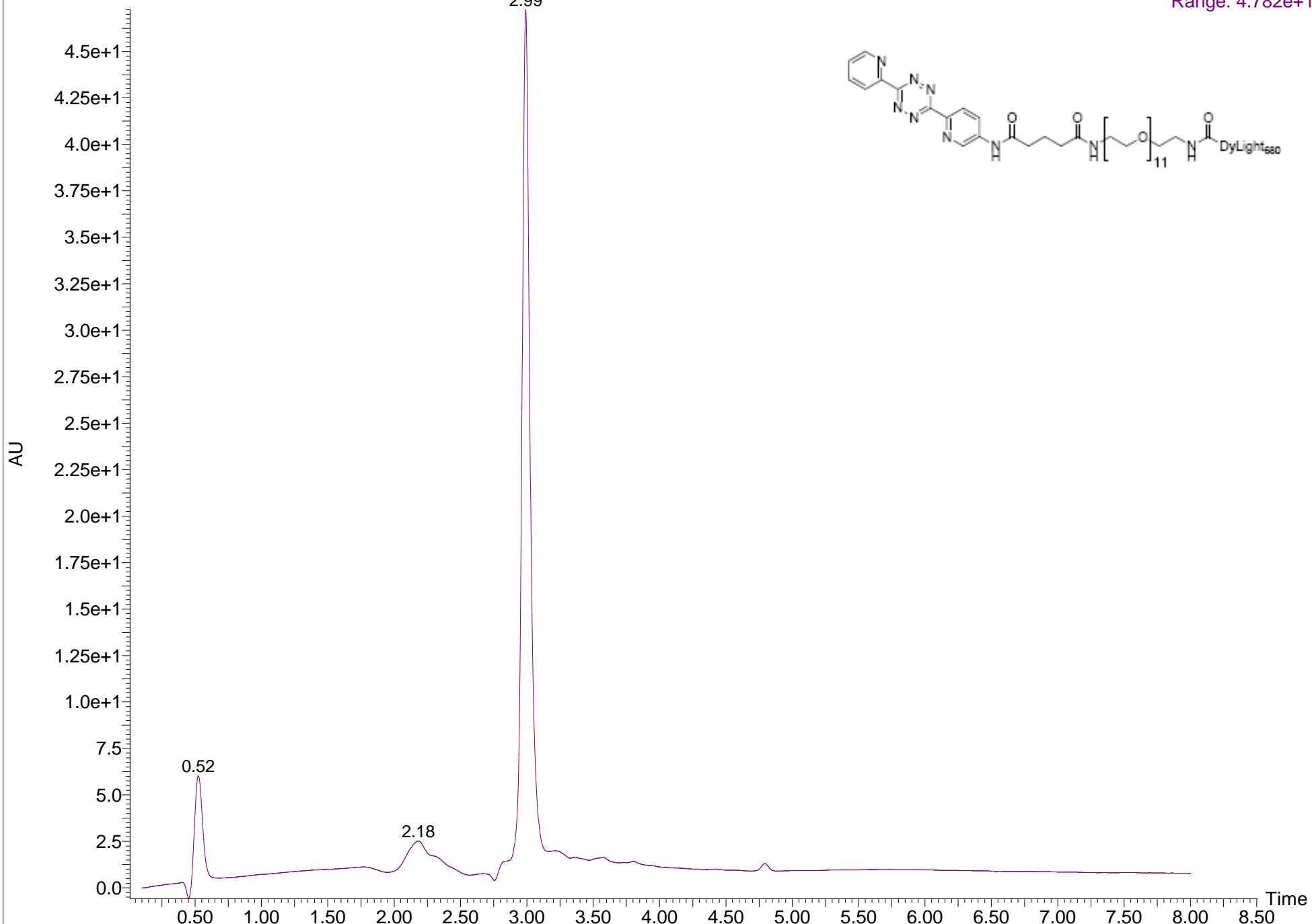
SI 65536
SF 125.7376638 MHz
WDW EM
SSB 0
LB 1.00 Hz
GB 0
PC 1.00



hs-509

F_Leeper2804-1

3: Diode Array



hs-509

F_Leeper2804-1 129 (3.023) Cm (127:132-(118:120+140:142))

2: Scan ES+
3.69e4

