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Supplementary information

Experimental Section

5 Materials

Synthesis of the redox polymers was achieved by adapting literature procedures¹ using $(NH_4)_2OsCl_6$ (Aldrich) as starting material to prepare the *cis*-Os(4,4'-dimethoxy-2,2'-bipyridine)₂Cl₂ and *cis*-Os(4,4'-dichloro-2,2'-bipyridine)₂Cl₂ complexes, which

- ¹⁰ were then complexed, via ligand substitution reaction in ethanol/water solvent, to a previously pre-synthesised polyvinylimidazole (PVI) polymer.² Glucose oxidase (GOx) from (A. Niger) was obtained from Sigma – Aldrich and ThLacc was donated by VTT Technology, Finland (K. Kruus). Unless ¹⁵ otherwise stated all other chemicals were obtained from Sigma-
- Aldrich. All buffers were prepared from solutions of the selected base then adjusted to the desired pH using solutions of the acid.

Apparatus

- 20 Graphite disc electrodes (3 mm diameter), formed by shrouding graphite rods (Goodfellow) in glass tubes using heatshrinkable tubing and establishing an electrical connection to copper rods (Farnell) at the rear with silver epoxy resin (Farnell), were used as working electrodes. Cyclic voltammetry was carried
- ²⁵ out with a CHI 650 potentiostat, using a graphite electrode, Ag/AgCl (3 M KCl) and platinum wire as working, reference and counter electrodes, respectively (IJ Cambria). EFCs were assembled by insertion of anode and cathode into a compartmentless electrochemical cell containing 5 mL of electrolyte solution.

³⁰ The anode and cathode were externally connected through a resistance box (IET Labs) over a resistance range of 5 M Ω to1 k Ω , and the voltage between the electrodes measured with a multimeter (Keithley) for each load.

Film assembly was monitored at each step with a quartz 25 crystal microbalance (QCM, USI Japan) with 9 MHz QCM resonators (AT-cut, International Crystal Mfg). The gold resonators were first treated with 4 mM 3-mercaptopropionic acid in ethanol overnight to form negative monolayer surface to mimic graphite surface. Films were assembled onto negatively charged 40 gold resonator surface and were dried in a stream of nitrogen

before measuring the frequency change (ΔF) .³

Enzyme activity

Glucose oxidase was obtained from Sigma – Aldrich (10 ⁴⁵ mg/ml stock solution was prepared in phosphate buffer pH 7.4) and *Trametes hirsuta* (*Th*Lacc) was provided by VTT (Finland) as (3.6 mg/mL) stock solution in citrate buffer pH 5. Enzyme activity was calculated using spectrophotometric assays (Agilent 8453). Glucose oxidase activity was monitored at 460 nm, where

- ⁵⁰ the increase in absorbance is a result of the oxidation of dianisidine through a peroxidase coupled system, using an extinction coefficient of 11300 M⁻¹ cm⁻¹, in phosphate buffer pH 6.⁴ The enzymatic activity of the laccase was measured by monitoring the oxidation of 5 mM ABTS in 50 mM acetate buffer
- $_{55}$ pH 4.5 at 420 nm over a period of 10 minutes using extinction coefficient of 36 000 $M^{-1}\,cm^{-1}.^{5}$

Methods

Electrodes were prepared, based on LBL self assembly of ⁶⁰ redox polymer and enzyme.⁶ For anode and cathode 10 μ L redox polymers (8–10 mg/ml solution/suspension in water) were adsorbed over graphite electrode for 20 minutes, then rinsed with milli Q water. Further enzymes 10 μ L GOx of 10 mg/ml (1500 U/ml), or *Th*Lacc (390 U/ml) was adsorbed for 20 minutes, then ⁶⁵ washed with Milli Q water, further repeating these steps to form (polymer /enzyme)_n films. Prepared films were dried for 12 h before testing electrochemically or EFC assembly. Unless otherwise stated, current and power densities were measured at 37 °C in 0.1M potassium phosphate buffered solutions containing 70 0.15 M NaCl, 0.1 M glucose and saturated O₂.

Supplementary figures

Figure S1. QCM frequency shifts for alternate redox polymer – 75 enzyme a) polymer I/GOx and b) polymer II/*Th*Lacc adsorbed on gold resonators first coated with a monolayer of 3-mercaptopropionic acid rendering a negatively charged surface that mimics the self assembly at graphite electrode.



Figure S2. Power versus current density curve of an EFC composed (polymer I /GOx)₂ and (polymer II / *Th*Lacc)₂ films in ⁹⁵ oxygen-saturated 0.1 M potassium phosphate buffer containing 0.15 M NaCl at 37 °C, in the presence of 0.1 M glucose at pH 7.4 (closed circles) and 5.5 (open circles).



ARTICLE TYPE

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