

Supporting Information

Is the Special Pair Structure a Good Strategy for the Kinetics During the Last Step of the Energy Transfer With the Nearest Antenna?

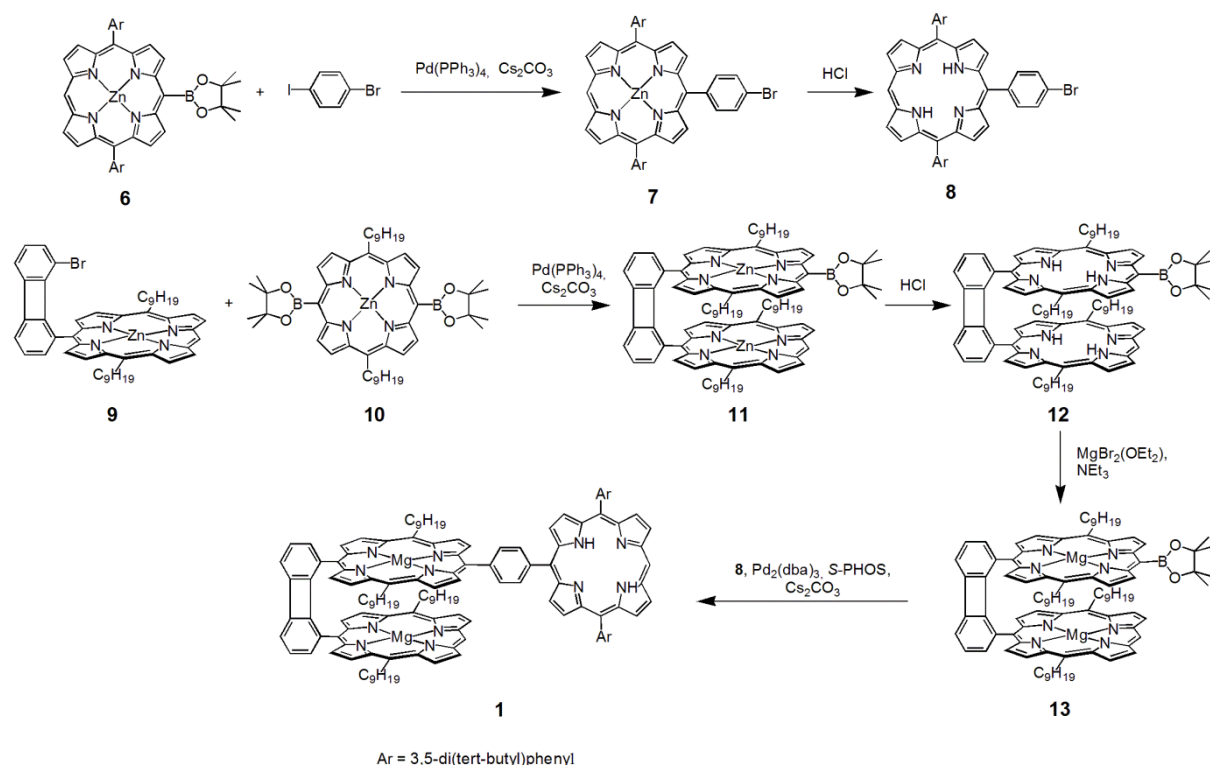
A Chemical Model Approach.

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Table of Content

1. Experimental Section	2
2. References	7
3. NMR and High Resolution Mass Spectra	8
¹ H NMR spectrum of 7	8
MALDI-TOF spectrum of 7	9
ESI HRMS spectrum of 7	10
¹ H NMR spectrum of 8	11
MALDI-TOF spectrum of 8	12
ESI HRMS spectrum of 8	13
¹ H NMR spectrum of 12	14
MALDI-TOF spectrum of 12	16
ESI HRMS spectrum of 12	16
¹ H NMR spectrum of 13	17
MALDI-TOF spectrum of 13	18
ESI HRMS spectrum of 13	18
¹ H NMR spectrum of 1	19
MALDI-TOF spectrum of 1	21
ESI HRMS spectrum of 1	21
4. Optical spectra	22
5. Computed energy barriers	24
6. Photophysical spectra and decays for 1 , 8 and 13	25

1. Experimental Section



Materials. Zinc(II) 5-(8-bromobiphenylene-1-yl)-10,20-bis(nonyl)porphyrin **9**,¹ zinc(II) 5,15-bis-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-10,20-bis(nonyl)porphyrin **10**,¹ zinc(II) 5-{8-[zinc(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-dinonyl-porphyrin **11**,¹ and zinc(II) 5-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-bis(3,5-di-*tert*-butylphenyl)porphyrin **6**² were synthesized as previously described. The handling of all air/water sensitive materials was carried out using standard high vacuum techniques. Dried toluene was obtained by passing through alumina under nitrogen in the solvent purification systems and then further dried over activated molecular sieves; extra dry DMF was purchased from Aldrich. Triethylamine, dichloromethane and 1,2-dichloroethane were distilled from CaH₂; THF and toluene were distilled from sodium benzophenone ketyl. Unless specified otherwise all other solvents were used as commercially supplied. Silica gel (Merck; 70-120 mm) was used for column chromatography. Analytical thin layer chromatography was performed using Merck 60 F254 silica gel (precoated sheets, 0.2 mm thick). Size exclusion chromatography was carried out under gravity using cross-linked polystyrene Bio-Beads® SX-1 (200 – 400 mesh) in DCM. Reactions were monitored by thin-layer chromatography, UV-vis spectroscopy, and MALDI/TOF mass spectrometry.

Zinc(II) 5-(4-bromophenyl)-10,20-bis-(3,5-di-tert-butyl-phenyl)-porphyrin 7

Under an inert atmosphere, a mixture of zinc (II) 5-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-bis(3,5-di-tert-butyl phenyl)porphyrin (155 mg, 0.18 mmol), 1-bromo,4-iodobenzene (76 mg, 0.27 mmol), cesium carbonate (91 mg, 0.28 mmol), Pd(PPh₃)₄ (21 mg, 0.0018 mmol) in 10 mL of anhydrous toluene and 5 mL of DMF was stirred at 100 °C overnight. The reaction was quenched with an aqueous saturated NH₄Cl solution (20 mL). The organic layer was separated, extracted from aqueous phase. The combined organic fractions were washed with water (2 x 20 mL) and dried over MgSO₄. The solvent was evaporated and under vacuum and the residue was purified by silica gel (chloroform/petroleum ether 1/4) affording the title compound as a purple solid (0.15 g, 92%). ¹H NMR (300.16 MHz, CDCl₃): δ 10.30 (s, 1H, *meso*), 9.44 (d, 2H, *J* = 4.5 Hz, β), 9.18 (d, 2H, *J* = 4.5 Hz, β), 9.08 (d, 2H, *J* = 4.7 Hz, β), 8.96 (d, 2H, *J* = 4.7 Hz, β), 8.15 (d, 4H, *J* = 1.8 Hz, C₆H₃), 8.12 (d, 2H, *J* = 8.4 Hz, C₆H₄), 7.90 (d, 2H, *J* = 8.4 Hz, C₆H₄), 7.84 (t, 2H, *J* = 1.8 Hz, C₆H₃) and 1.58 (s, 36H, tBu) ppm. MALDI-TOF MS : *m/z* 902.2 calcd for C₅₄H₅₅BrN₄Zn, 902.2 (M⁺). ESI HRMS : *m/z* 902.2896 calcd for C₅₄H₅₅BrN₄Zn, 902.2895 (M⁺). UV-vis (THF) λ_{max} (log ε) = 414 (5.73), 542 (4.31), 579 (3.30) nm.

5-(4-bromophenyl)-10,20-bis-(3,5-di-tert-butyl-phenyl)-porphyrin 8

20 mL of HCl aqueous solution (4M) was added onto a solution of zinc(II) 5-(4-bromophenyl)-10,20-bis-(3,5-di-tert-butyl-phenyl)porphyrin (0.15 g, 0.166 mmol) in 50 mL of dichloromethane. The reaction mixture was vigorously stirred for 1 hour. The organic layer was separated, washed with a saturated NaHCO₃ aqueous solution (2x20 mL) and dried over magnesium sulfate. After evaporation until dryness, the residue was recrystallized in dichloromethane/methanol affording a microcrystalline violet powder (0.14 g, quant.). ¹H NMR (300.16 MHz, CDCl₃): δ 10.23 (s, 1H, *meso*), 9.35 (d, 2H, *J* = 4.7 Hz, β), 9.07 (d, 2H, *J* = 4.6 Hz, β), 8.96 (d, 2H, *J* = 4.8 Hz, β), 8.84 (d, 2H, *J* = 4.8 Hz, β), 8.11 (m, 6H, C₆H₄+C₆H₃), 7.87 (d, 2H, *J* = 8.3 Hz, C₆H₄), 7.83 (m, 2H, C₆H₃), 1.56 (s, 36H, tBu) and -2.94 (s, 2H, NH) ppm. MALDI-TOF MS : *m/z* 840.3 calcd for C₅₄H₅₇BrN₄, 840.3 (M⁺). ESI HRMS : *m/z* 841.3839 calcd for C₅₄H₅₈BrN₄, 841.3844 (M+H)⁺. UV-vis (THF) λ_{max} (log ε) = 412 (5.43), 508 (4.09), 542 (3.69), 584 (3.59), 641 (3.29) nm.

5-{8-[10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-dinonylporphyrin 12

10 mL of HCl aqueous solution (4M) was added onto a solution of zinc(II) 5-{8-[zinc(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-dinonylporphyrin (58 mg, 0.038 mmol) in 15 mL of dichloromethane. The reaction mixture was vigorously stirred for 30 minutes. The acidic layer was removed and 10 mL of HCl aqueous solution (4M) was again added. After stirring for 30 minutes, the organic layer was separated, washed with a saturated NaHCO₃ aqueous solution (2x20 mL) and dried over magnesium sulfate. After evaporation until dryness, the residue was purified by silica gel (chloroform/petroleum ether 1/2-2/1) affording the title compound as a purple solid (33 mg, 62%). ¹H NMR (300.16 MHz, CD₂Cl₂): δ 9.34 (d, 2H, *J* = 4.6 Hz, β), 9.21 (s, 1H, *meso*), 8.77 (d, 2H, *J* = 4.9 Hz, β), 8.74 (d, 2H, *J* = 5.0 Hz, β), 8.66 (d, 2H, *J* = 4.7 Hz, β), 8.57 (br d, 2H, *J* = 4.9 Hz, β), 8.50 (d, 2H, *J* = 4.9 Hz, β), 8.47 (br d, 2H, *J* = 4.2 Hz, β), 8.37 (d, 2H, *J* = 4.8 Hz, β), 7.35-7.28 (m, 3H, DPB), 7.22-7.12 (m, 3H, DPB), 3.92 (m, 8H, CH₂), 2.13 (s, 12H, CH₃), 1.99 (m, 8H, CH₂), 1.62 (m, 8H, CH₂), 1.45 (m, 8H, CH₂), 1.32 (m, 32H, CH₂), 0.89 (t, 12H, *J* = 7.3 Hz, CH₃), -7.13 (s, 2H, NH) and -7.21 (s, 2H, NH) ppm. MALDI-TOF MS: *m/z* 1398.9 calcd for C₉₄H₁₁₅BN₈O₂, 1399.9 (M+H⁺). ESI HRMS : *m/z* 1399.9323 calcd for C₉₄H₁₁₆BN₈O₂, 1399.9263 (M+H⁺). UV-vis (THF) λ_{max} (log ε) = 397 (5.54), 517 (4.13), 551 (3.81), 598 (3.68), 652 (3.43) nm.

Magnesium(II) 5-{8-[magnesium(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-dinonylporphyrin 13

5-{8-[10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2]-dioxaborolan-2-yl)-10,20-dinonylporphyrin (25 mg, 0.018 mmol) was dissolved in 10 mL of dichloromethane and 0.42 mL of NEt₃ (3 mmol) and 0.38 g of MgBr₂.OEt₂ (1.5 mmol) were successively added. The reaction mixture was stirred for 30 minutes. 10 mL of dichloromethane was added and the organic phase was washed by a diluted NaHCO₃ aqueous solution (2 x 20mL). The organic layer was dried over magnesium sulfate and evaporated until dryness. Chromatographic column on alumina eluting with CH₂Cl₂ and with acetone-CH₂Cl₂ (20%) afforded **3** as purple powder (20 mg, 80%). ¹H NMR (300.16 MHz, CD₂Cl₂): δ 9.33 (br s, 2H, β), 9.22(s, 1H, *meso*), 8.74 (m, 4H, β), 8.63 (br s, 2H, β), 8.54 (br s, 2H, β), 8.43 (br s, 2H, β), 8.30 (br s, 2H, β), 8.17 (br s, 2H, β), 7.30-7.23(m, 3H, DPB), 7.16-7.07 (m, 3H, DPB), 3.60 (br m, 8H, CH₂), 2.04 (s, 12H, CH₃), 1.95 (br m, 8H, CH₂), 1.62 (br m, 8H, CH₂), 1.49-1.21 (m, masked by heptane, CH₂), 0.91 (m, masked by heptane, CH₃) ppm. MALDI-TOF MS : *m/z* 1442.8 calcd for C₉₄H₁₁₁BMg₂N₈O₂, 1442.8 (M⁺). ESI HRMS : *m/z* 1443.8654 calcd for C₉₄H₁₁₁BMg₂N₈O₂, 1443.8746 (M+H⁺). UV-vis (THF) λ_{max} (log ε)= 408 (5.58), 569 (4.04), 614 (3.85) nm.

Magnesium(II) 5-{8-[magnesium(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-[4-{10,20-bis-3,5-di-tert-butyl-phenyl)-porphyrin-5-yl}phenyl]-10,20-dinonylporphyrin 1

Under an inert atmosphere, a solution of Pd₂(dba)₃ (0.9 mg, 0.001 mmol) and *S*-PHOS (2.5 mg, 0.006 mmol) in 6 mL of freshly distilled toluene was added onto a mixture of magnesium(II) 5-{8-[magnesium(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2] dioxaborolan-2-yl)-10,20-dinonylporphyrin (15 mg, 0.010 mmol), 5-(4-bromophenyl)-10,20-bis-(3,5-di-tert-butyl-phenyl)-porphyrin (13 mg, 0.015 mmol) and cesium carbonate (13 mg, 0.04 mmol). 3 mL of anhydrous DMF was added and the reaction mixture was stirred at 90 °C for 16 h. The reaction was quenched with 10 ml of water. The organic layer was separated, extracted from aqueous phase. The combined organic fractions were washed with brine (2 x 20 mL) and dried over MgSO₄. The solvents were removed under vacuum and the purple residue was purified over SEC chromatography affording the title compound as a dark purple solid (6 mg, 28%). ¹H NMR (300.16 MHz, THF-d⁸): δ 10.41 (s, 1H, *meso*), 9.83 (dd, 1H, *J* = 7.0 Hz, *J* = 1.7 Hz, C₆H₄), 9.56 (d, 2H, *J* = 4.6 Hz, β), 9.50 (d, 2H, *J* = 4.1 Hz, β), 9.39 (s, 1H, *meso*), 9.24 (d, 2H, *J* = 4.4 Hz, β), 9.14 (d, 2H, *J* = 4.2 Hz, β), 9.11 (dd, 1H, *J* = 7.0 Hz, *J* = 1.9 Hz, C₆H₄), 8.99 (d, 2H, *J* = 4.2 Hz, β), 8.90 (d, 2H, *J* = 3.9 Hz, β), 8.79 (d, 2H, *J* = 3.9 Hz, β), 8.62 (d, 2H, *J* = 4.7 Hz, β), 8.52 (d, 3H, *J* = 4.0 Hz, β + C₆H₃), 8.46 (dd, 1H, *J* = 7.0 Hz, *J* = 1.9 Hz, C₆H₄), 8.39 (s, 2H, C₆H₃), 8.31 (s, 2H, C₆H₃), 8.24 (d, 2H, *J* = 4.7 Hz, β), 8.03 (t, 2H, *J* = 1.8 Hz, C₆H₃), 7.97 (d, 2H, *J* = 5.4 Hz, β), 7.44 (d, 1H, *J* = 7.2 Hz, DPB), 7.30 (d, 1H, *J* = 6.6 Hz, DPB), 7.24 (dd, 1H, *J* = 2.8 Hz, *J* = 7.8 Hz, DPB), 7.21 (dd, 1H, *J* = 2.1 Hz, *J* = 8.1 Hz, DPB), 7.05 (m, 2H, DPB), 3.58 (masked by THF-d⁸), 3.46 (m, 8H, CH₂), 2.01 (m, 8H, CH₂), 1.66 (s, 18H, tBu), 1.34 (m, masked by water and heptane, CH₂), 1.29 (s, 18H, tBu), 0.90 (m, masked by heptane, CH₃) and -2.60 (s, 2H, NH) ppm. MALDI-TOF MS : *m/z* 2079.2 calcd for C₁₄₂H₁₅₆Mg₂N₁₂, 2079.2 (M⁺). ESI HRMS : *m/z* 2079.2318 calcd for C₁₄₂H₁₅₆Mg₂N₁₂, 2079.2344 (M⁺). UV-vis (THF) λ_{max} (log ε) = 417 (5.65), 509 (4.32), 544 (4.10), 572 (4.10), 583 (4.13), 615 (3.99), 640 (3.92) nm.

Instruments. ¹H NMR spectra were recorded on a Bruker Avance II 300 (300 MHz) spectrometer at the "Plateforme d'Analyse Chimique et de Synthèse Moléculaire de l'Université de Bourgogne (PACSMUB)"; chemical shifts are expressed in ppm relative to chloroform (7.26 ppm), methylene chloride (5.30 ppm) or THF-d⁸ (1.73 and 3.58 ppm). UV-vis spectra were recorded on a Varian Cary 1 spectrophotometer. Mass spectra and accurate

mass measurements (HR-MS) were obtained respectively on a Bruker Daltonics Ultraflex II spectrometer in the MALDI/TOF reflectron mode using dithranol as a matrix and on a Bruker MicroQToF instrument or on a Thermo LTQ Orbitrap XL mass spectrometer in the positive ESI mode. Both measurements were performed at the “Plateforme d’Analyse Chimique et de Synthèse Moléculaire de l’Université de Bourgogne (PACSMUB)”. UV-visible spectra were recorded on a Hewlett-Packard diode array model 8452A and the emission spectra were obtained using a double monochromator Fluorolog 2 instrument from Spex. The fluorescence lifetimes were measured using a TimeMaster Model TM-3/2003 apparatus from PTI. The source was nitrogen laser with high-resolution dye laser (fwhm ~1600ps) and fluorescence lifetimes were obtained from deconvolution or distribution lifetimes analysis.

Transient absorption spectroscopy. The fs transient absorption spectra (at 1.5 nm of spectra resolution) along with the decay traces were acquired with a Helios spectrometer from Ultrafast Systems in the 450-800 nm range and 150 fs to 3.0 ns time window. This Helios instruments used an all-reflective supercontinuum generation setup resulting in a temporal chirp of the probe pulse of ~200 fs. Fiber optics were used to couple the multichannel spectrometer with a CMOS sensor with 1.5 nm intrinsic resolution. The instrument control and the data acquisition were made with a LabVIEW-based software. The data analysis was made with the Surface ExplorerTM software package. The laser excitation was performed using the Solstice (Spectra-Physics), a one box ultrafast regenerative Ti-sapphire amplifier. The system produced pulses centered at 795nm with bandwidth of ~15nm, energy of 3.75mJ, ~75 fs duration and 1 kHz repetition rate. The frequency was doubled with a second harmonic generator crystal (BBO) to obtain an excitation at 400 nm.

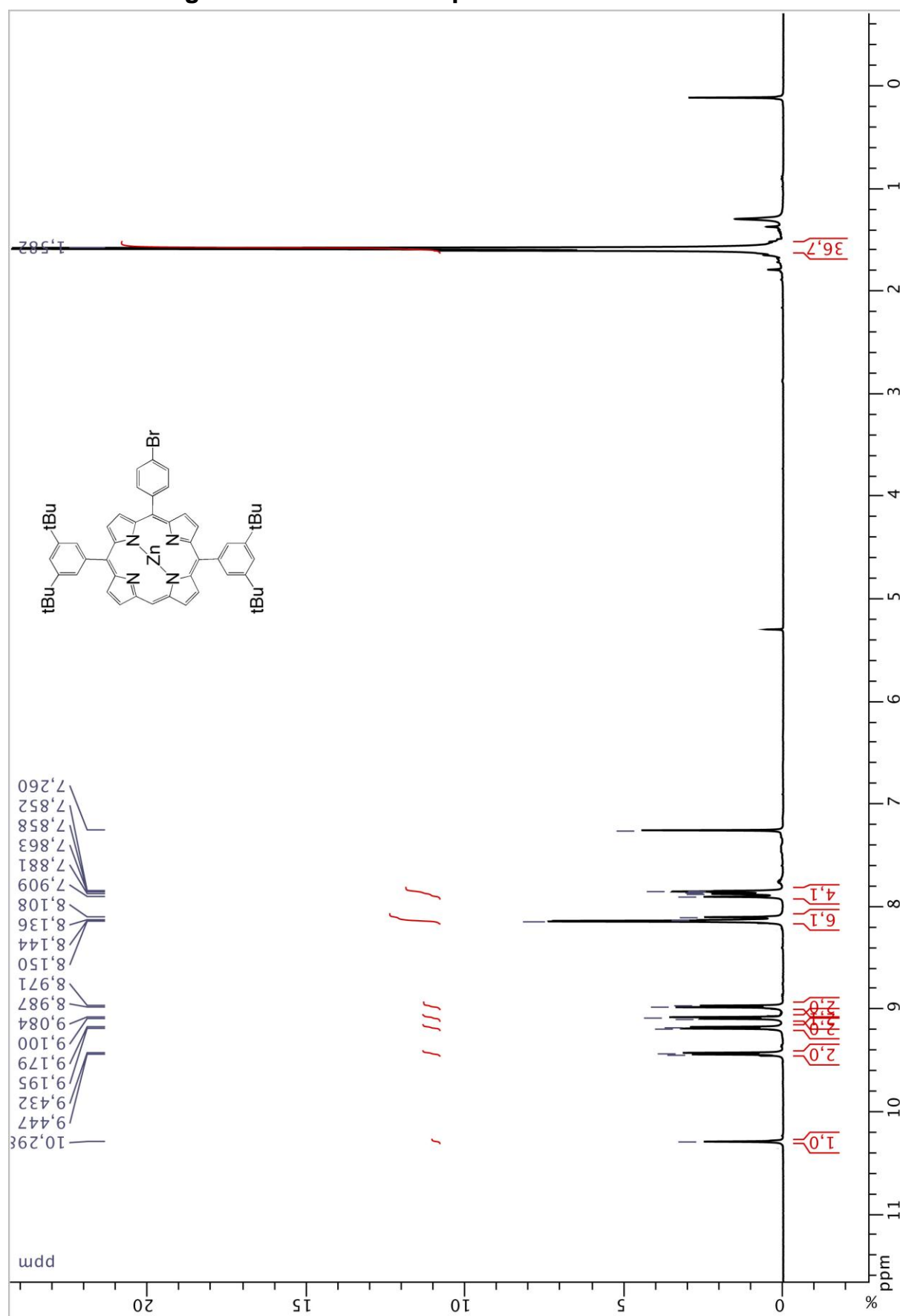
Computations. All density functional theory (DFT) and time dependent density functional theory (TD-DFT) calculations were performed with Gaussian 09³ at the Université de Sherbrooke with the Mammouth supercomputer supported by *Le Réseau Québécois De Calculs Hautes Performances*. The DFT geometry optimisations as well as TD-DFT calculations⁴⁻¹³ were carried out using the B3LYP method. 6-31g* basis sets for all porphyrin macrocycles, the biphenylene linkage as well as the bridging benzene were used. 3-21g* basis sets for solubilising side groups¹⁴⁻¹⁸ were employed. 6-31g* basis sets were used for magnesium atoms.¹⁴⁻¹⁹

Quantum yields. The fluorescence quantum yields were measured against H₂TPP tetraphenylporphyrin in 2MeTHF ($\Phi_F = 0.11$ in 2MeTHF at 296K).²⁰ Each measurement was performed 3 times with fresh solutions.

2. References

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3. NMR and High Resolution Mass Spectra



Comment 1

Comment 2

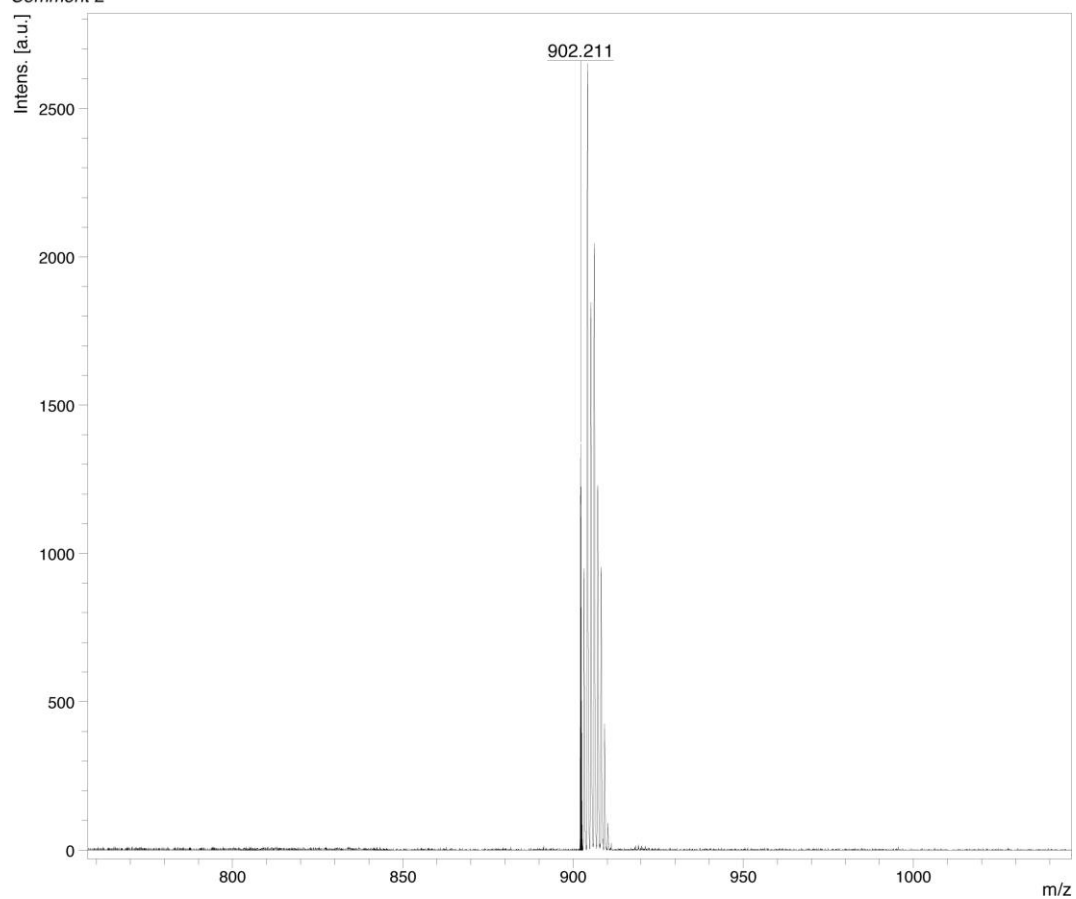


Figure S2. MALDI-TOF spectrum of **7** (positive mode)

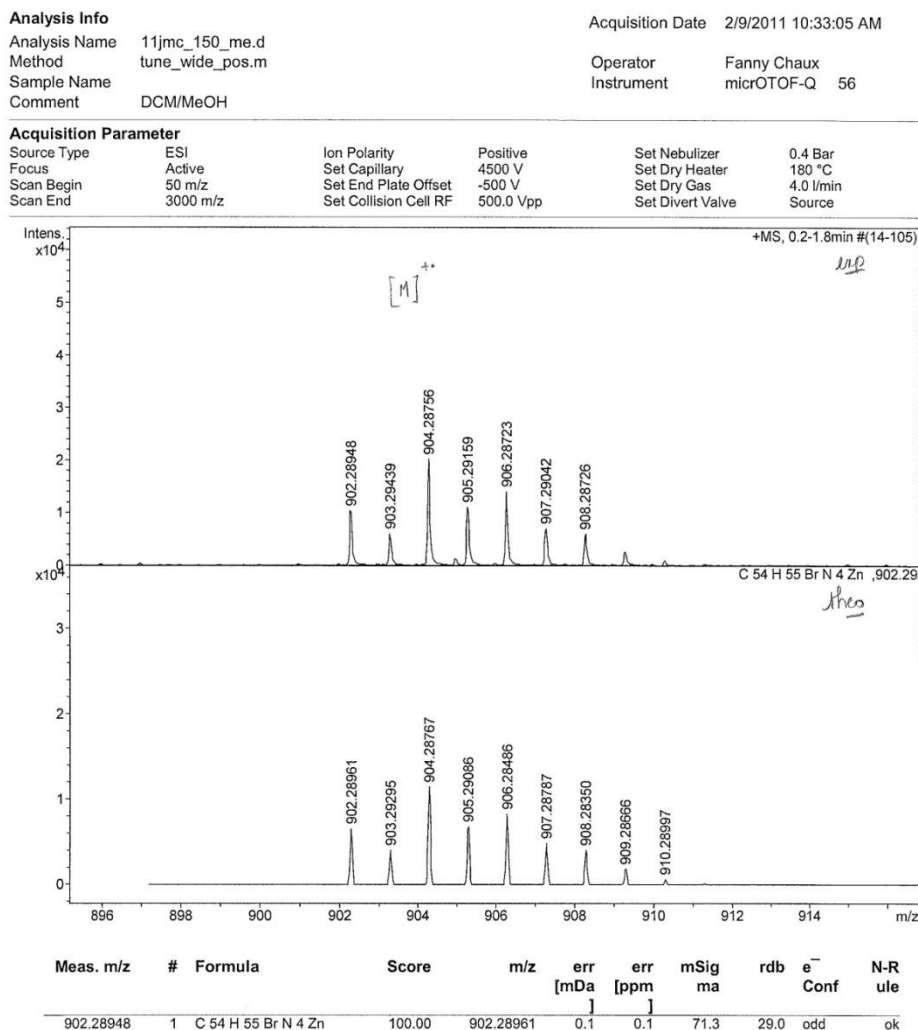


Figure S3. ESI HRMS spectrum of **7** (positive mode)

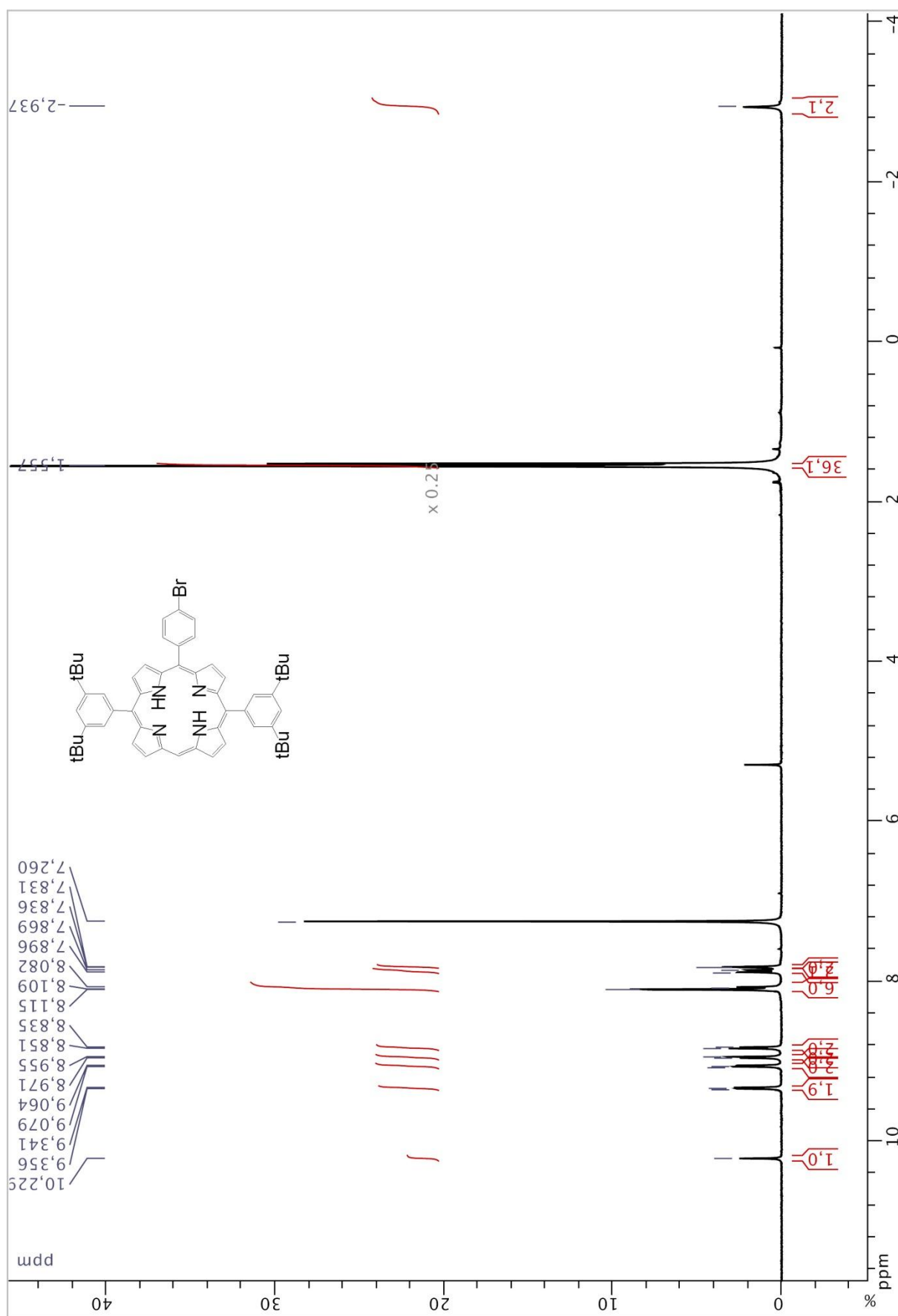


Figure S4. ^1H NMR of 5-(4-bromophenyl)-10,20-bis-(3,5-di-tert-butylphenyl)-porphyrin **8** (300 MHz, CDCl_3).

Comment 1

Comment 2

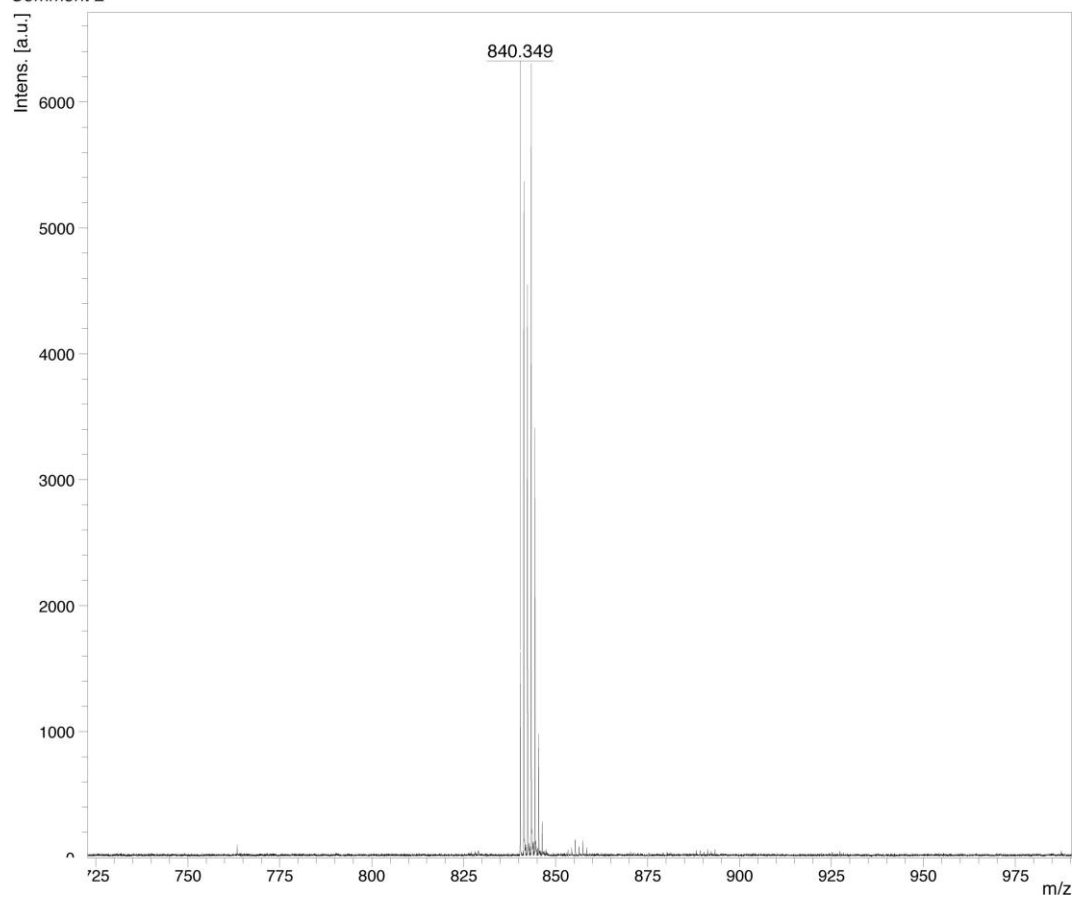


Figure S5. MALDI-TOF spectrum of **8** (positive mode)

Analysis Info

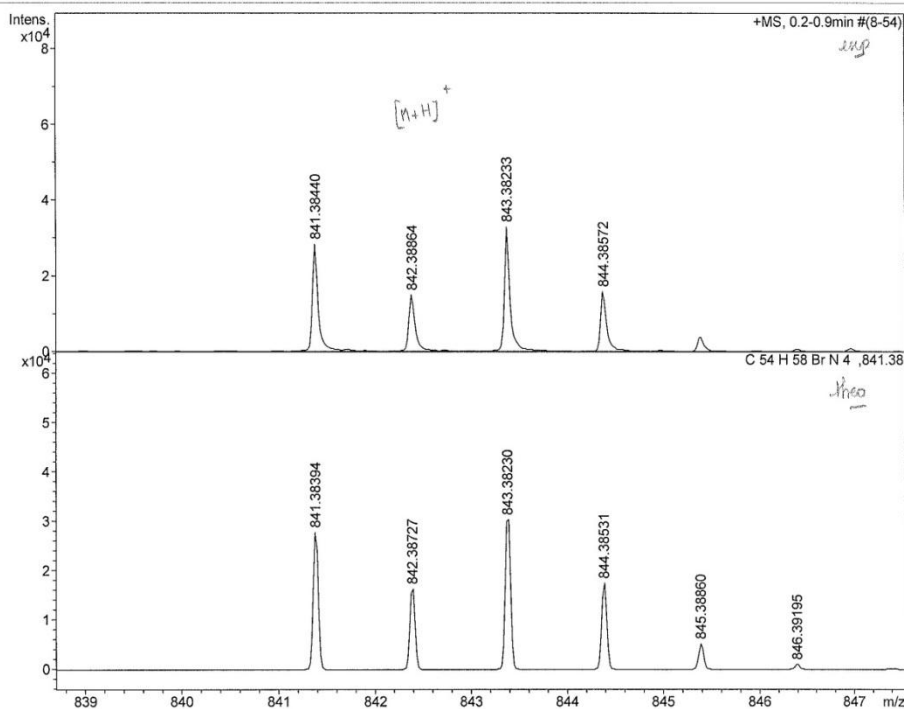
Analysis Name 11jmc_153_me.d
 Method tune_wide_pos.m
 Sample Name
 Comment DCM/MeOH

Acquisition Date 2/9/2011 10:24:09 AM

Operator Fanny Chaux
 Instrument micrOTOF-Q 56

Acquisition Parameter

Source Type	ESI	Ion Polarity	Positive	Set Nebulizer	0.4 Bar
Focus	Active	Set Capillary	4500 V	Set Dry Heater	180 °C
Scan Begin	50 m/z	Set End Plate Offset	-500 V	Set Dry Gas	4.0 l/min
Scan End	3000 m/z	Set Collision Cell RF	500.0 Vpp	Set Divert Valve	Source



Meas. m/z	#	Formula	Score	m/z	err [mDa]	err [ppm]	mSigma	rdb	e ⁻ Conf	N-Rule
841.38440	1	C 54 H 58 Br N 4	100.00	841.38394	-0.5	-0.6	70.0	27.5	even	ok

Figure S6. ESI HRMS spectrum of **8** (positive mode)

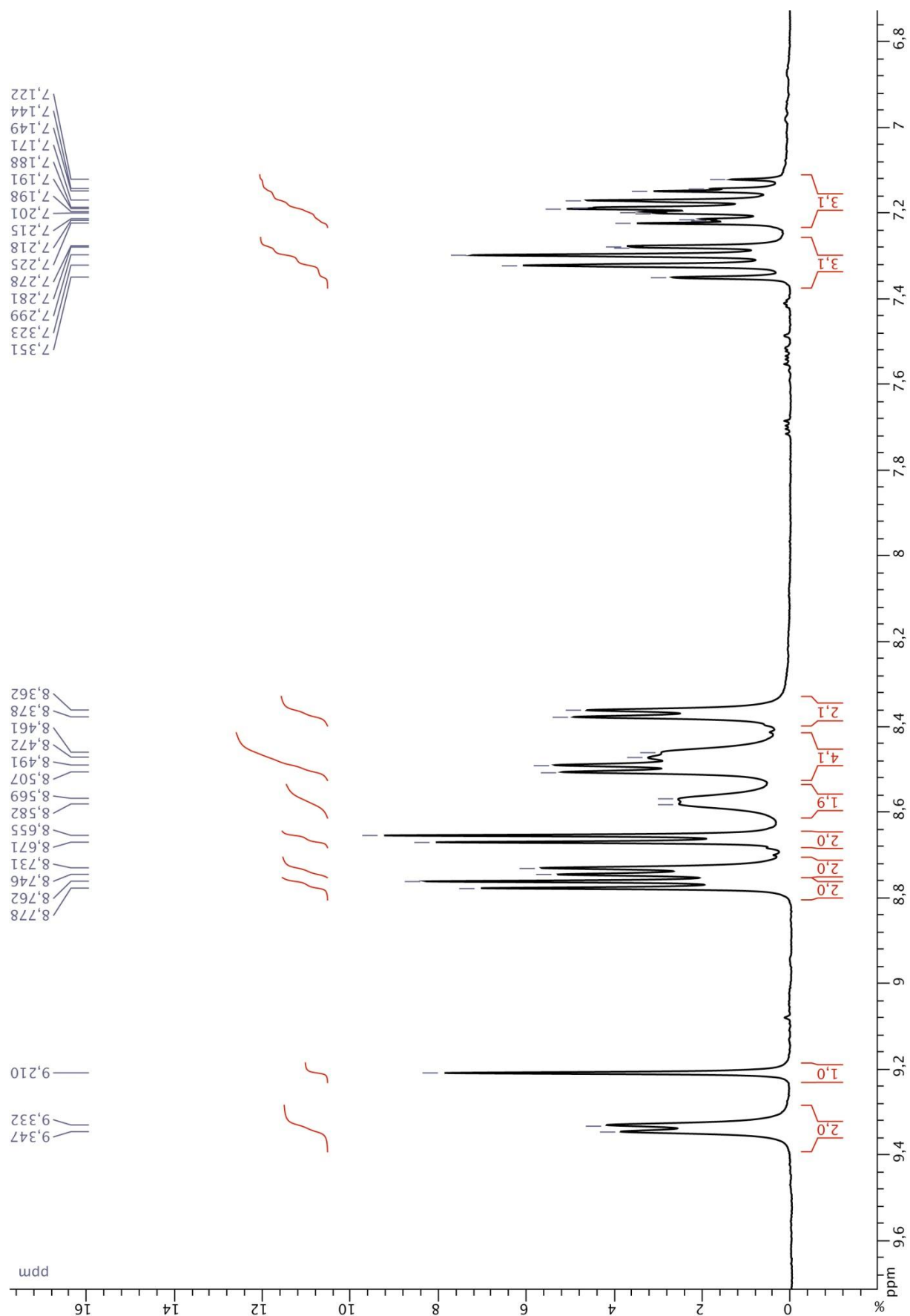


Figure S8. Aromatic part of ^1H NMR of **12** (300 MHz, CD_2Cl_2)

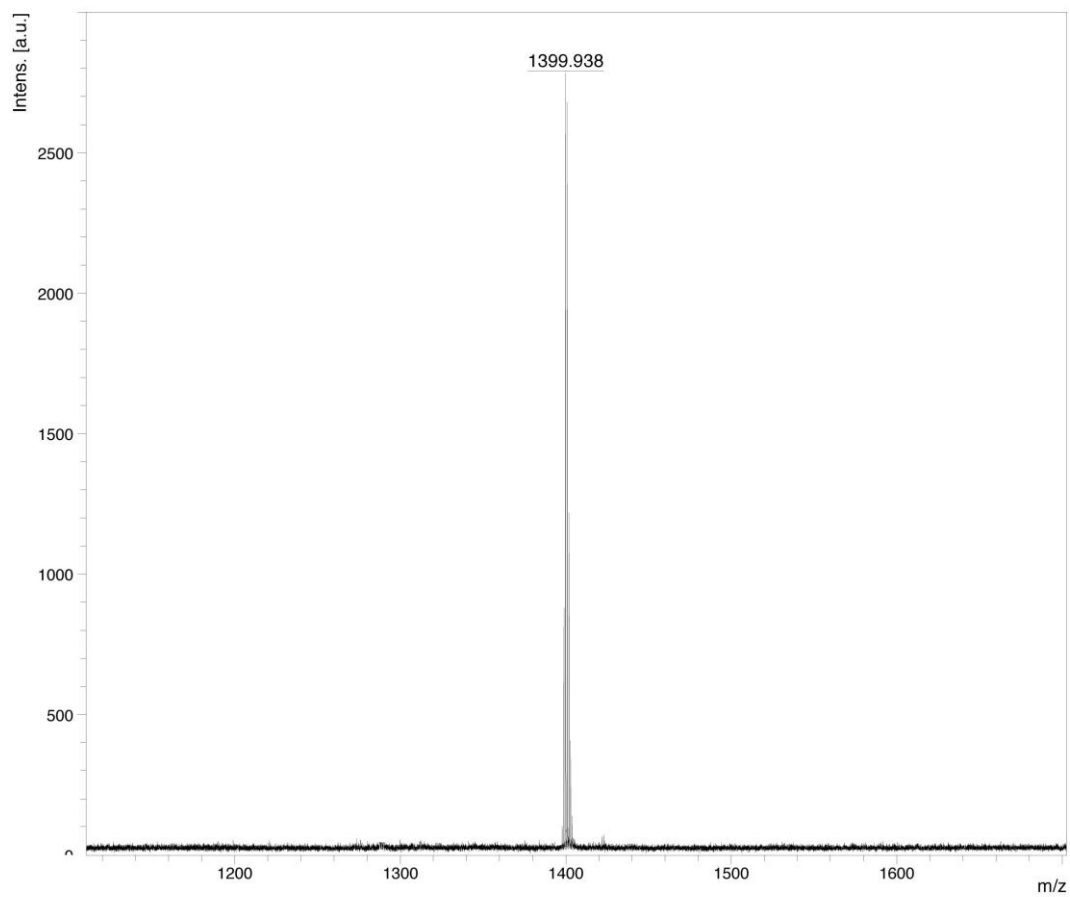


Figure S9. MALDI-TOF spectrum of **12** (positive mode)

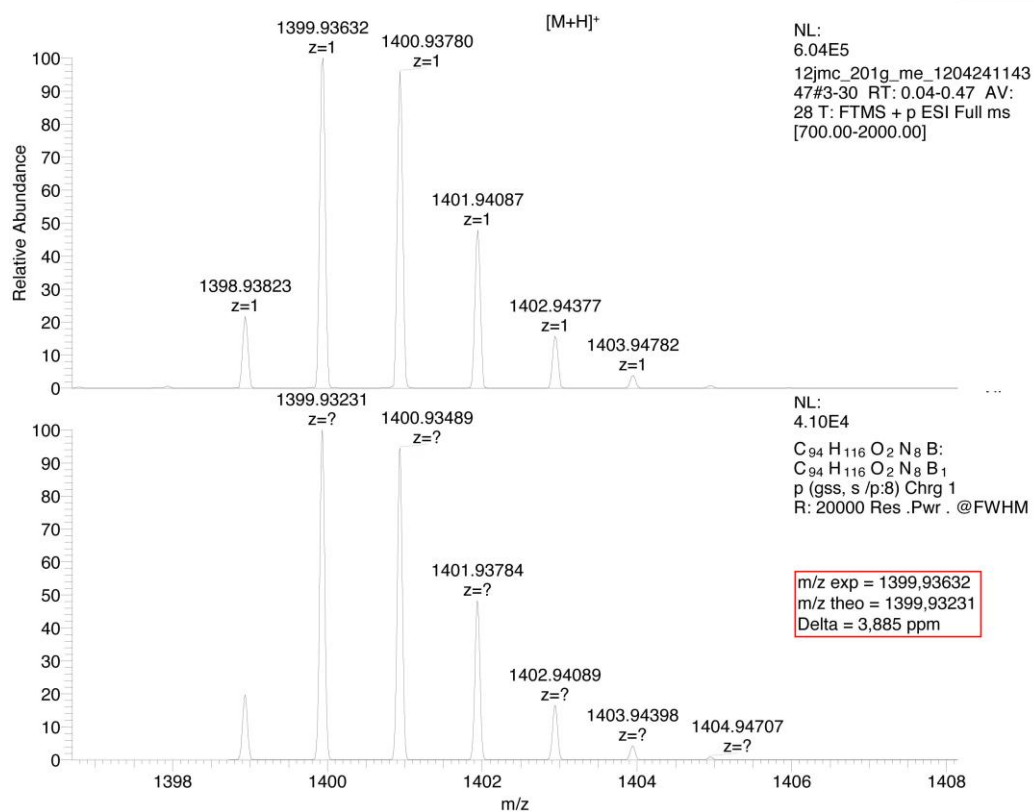


Figure S10. ESI HRMS spectrum of **12** (positive mode)

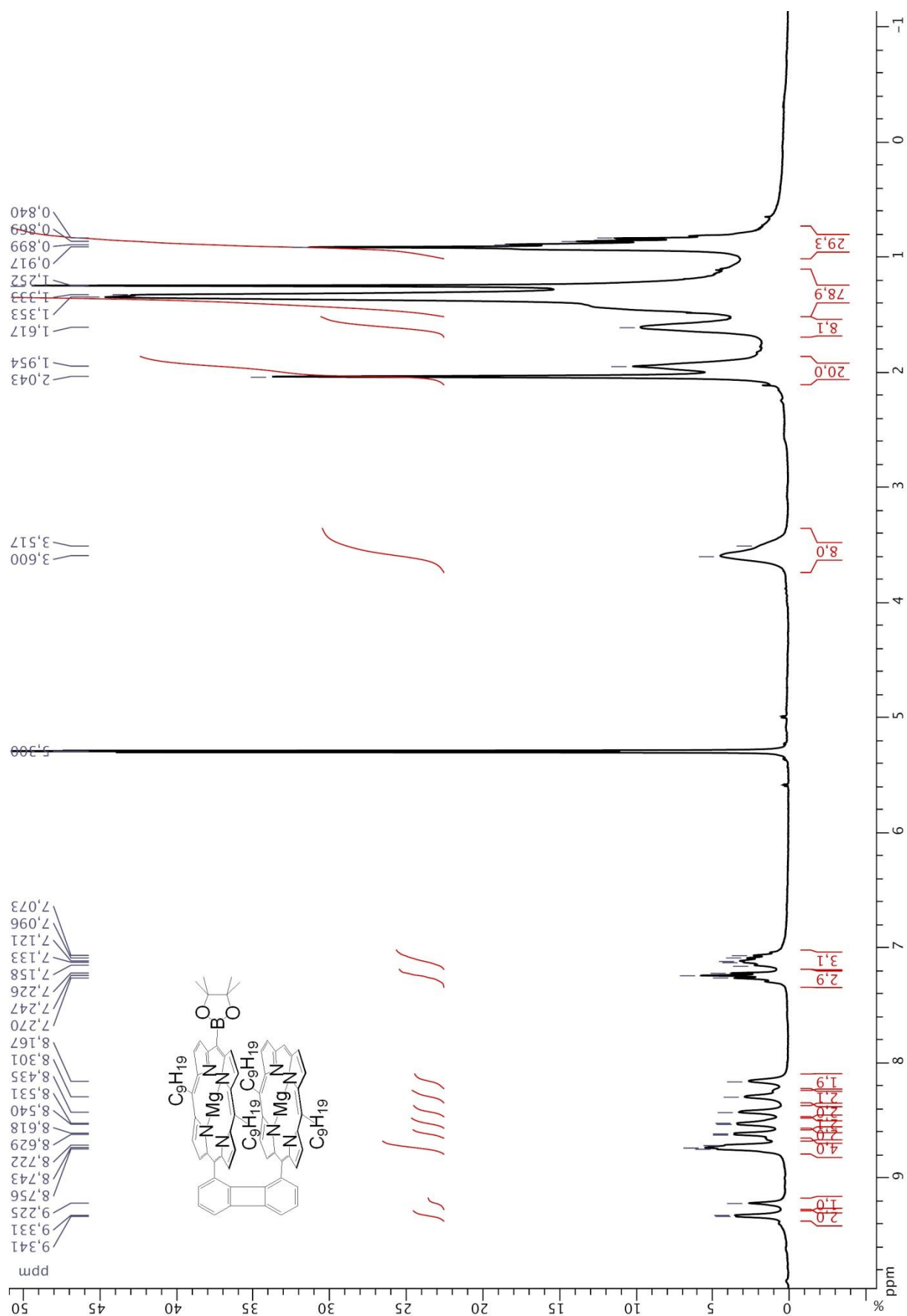


Figure S11. 1H NMR of magnesium(II) 5-{8-[magnesium(II)-10,20-dinonylporphyrin-5-yl]-biphenylene-1-yl}-15-(4,4,5,5-tetramethyl-[1,3,2] dioxaborolan-2-yl)-10,20-dinonylporphyrin **13** (300 MHz, CD_2Cl_2)

Comment 1

Comment 2

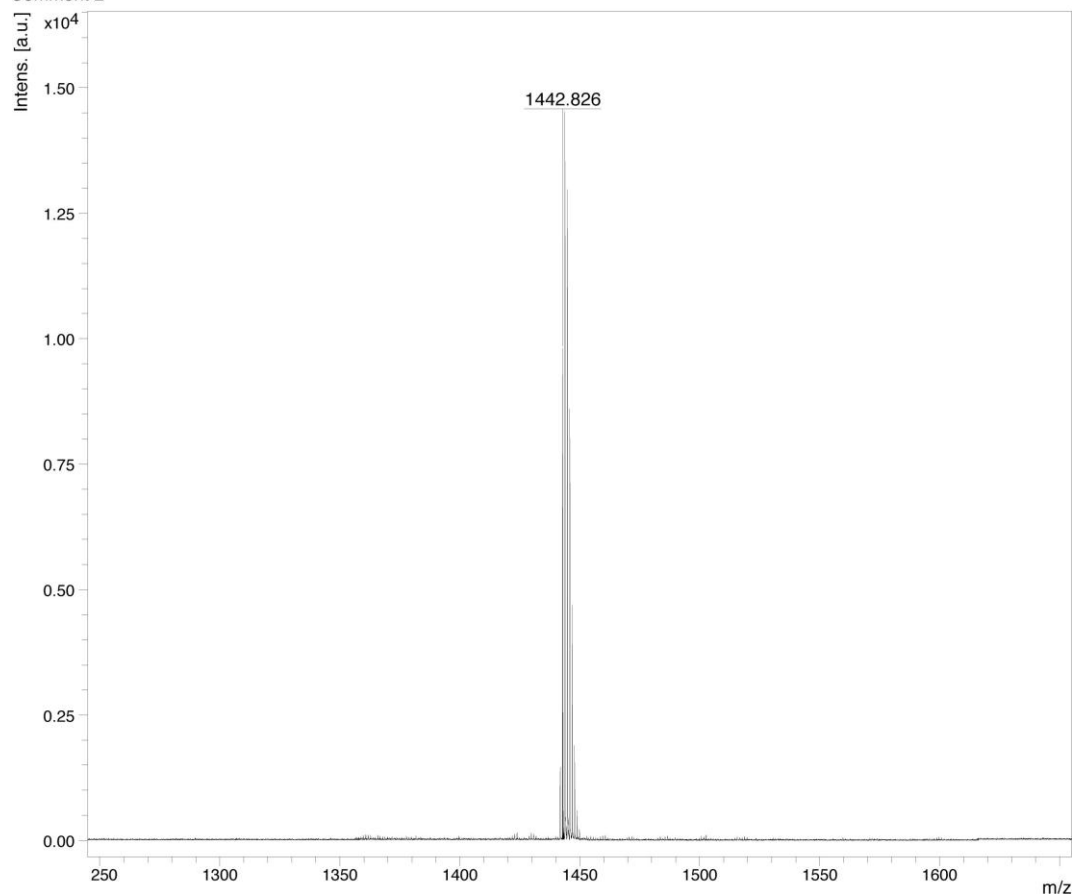


Figure S12. MALDI-TOF spectrum of **13** (positive mode)

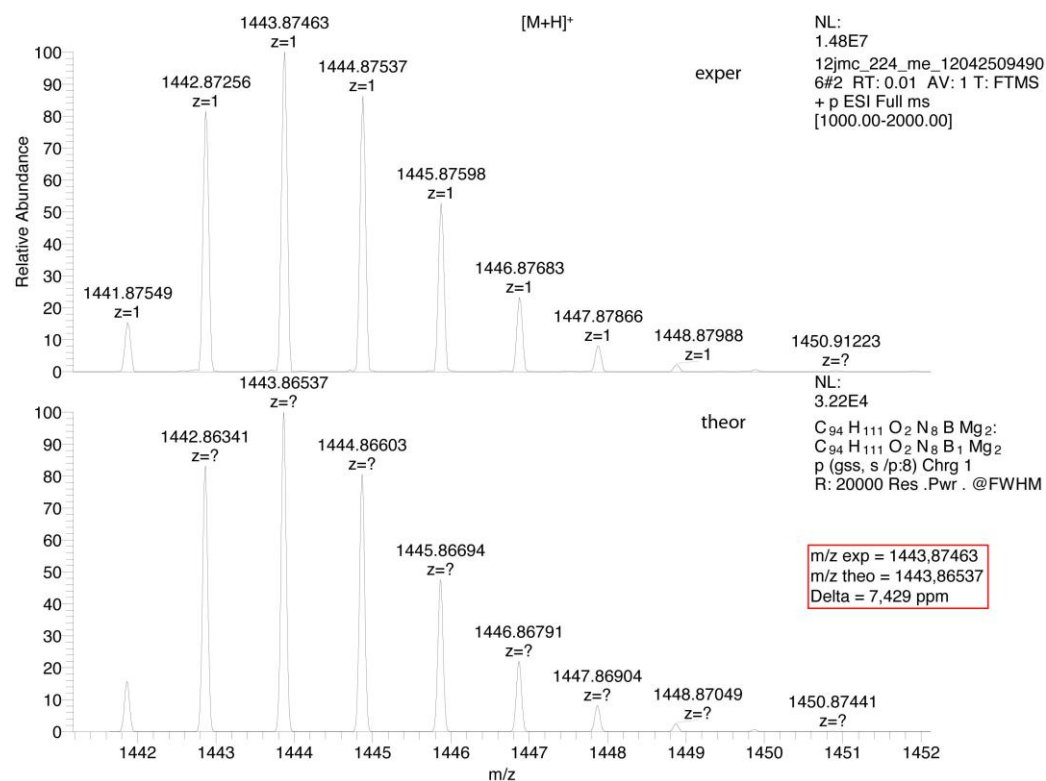


Figure S13. ESI HRMS spectrum of **13** (positive mode)

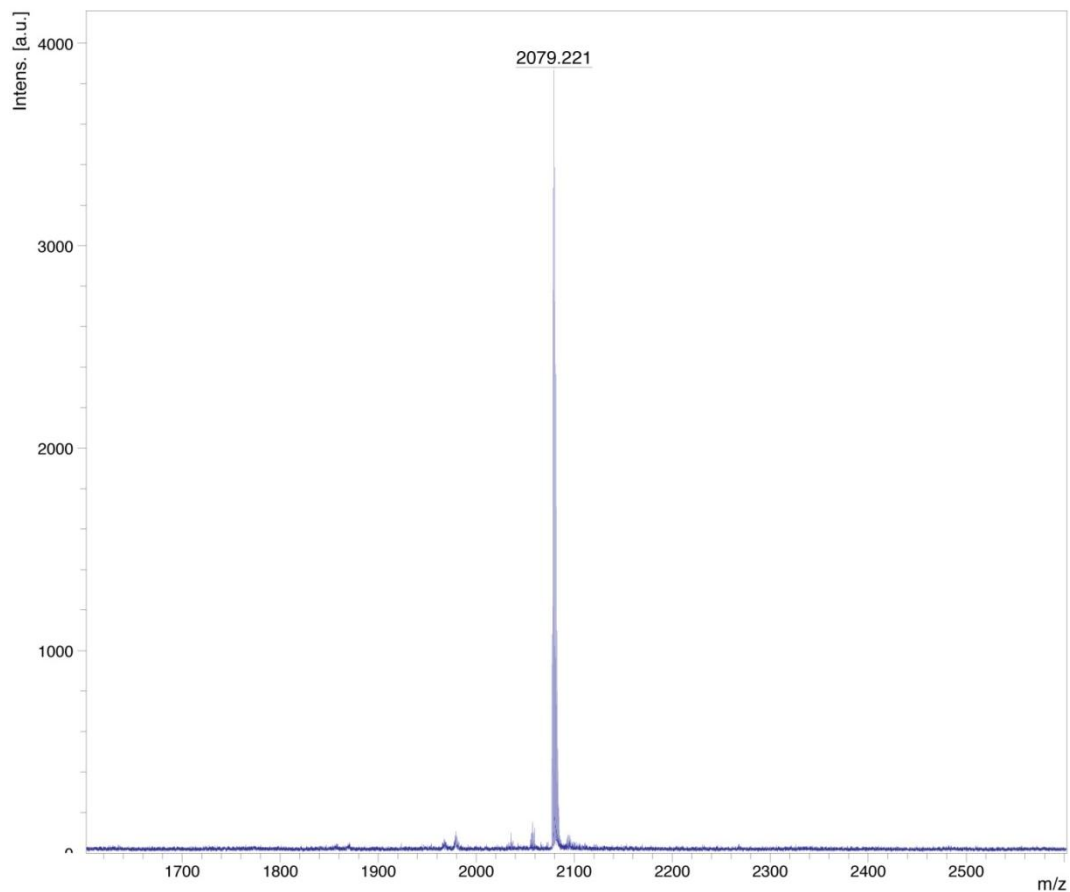


Figure S 16. MALDI-TOF spectrum of **1** (positive mode)

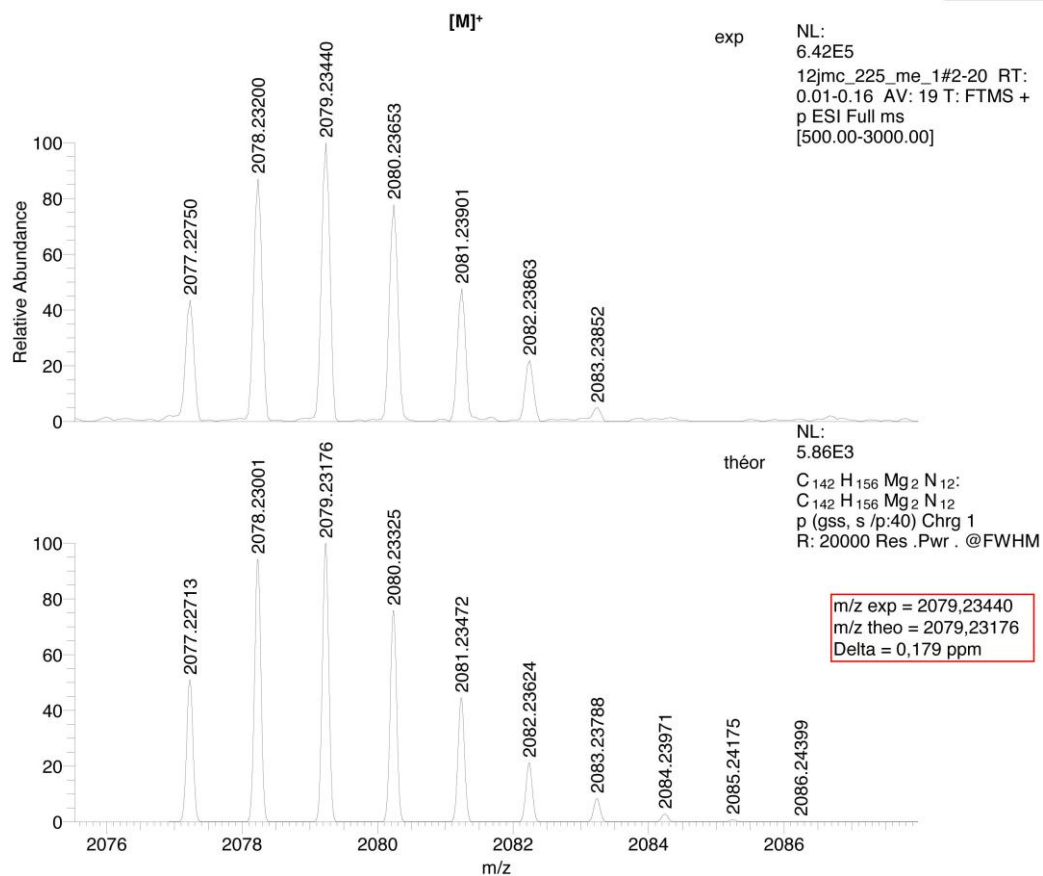


Figure S17. ESI HRMS spectrum of **1** (positive mode)

4. Optical spectra

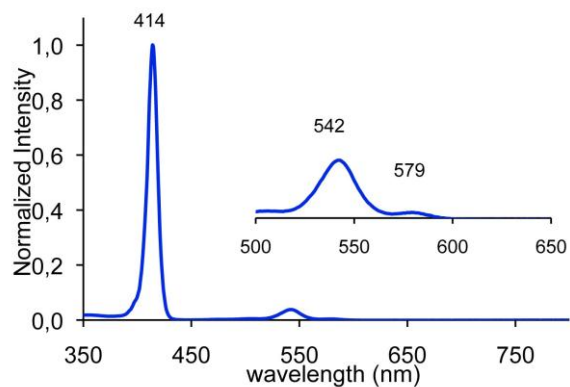


Figure S 20. Absorption spectrum of **7** in THF

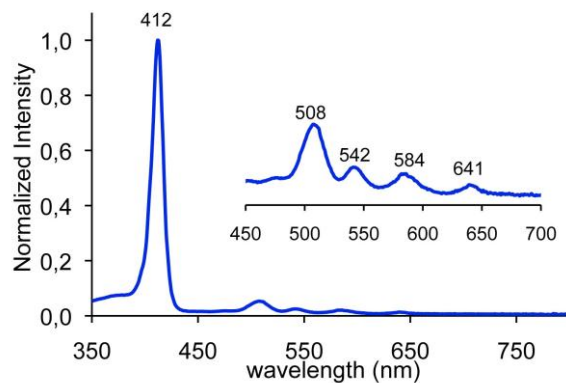


Figure S 18. Absorption spectrum of **8** in THF

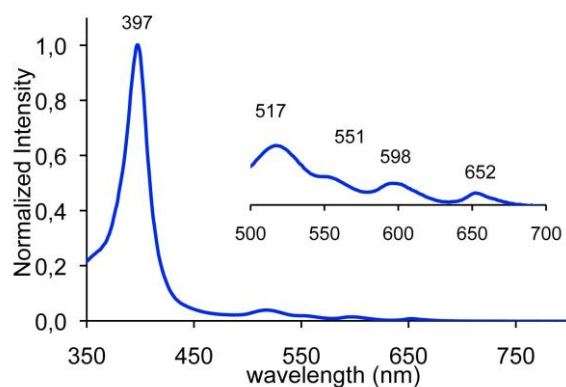


Figure S19. Absorption spectrum of **12** in THF

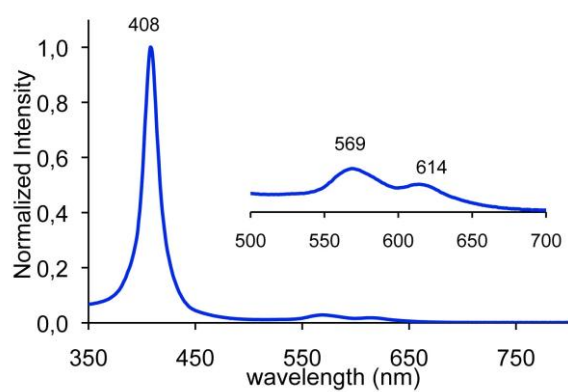


Figure S 20. Absorption spectrum of **13** in THF

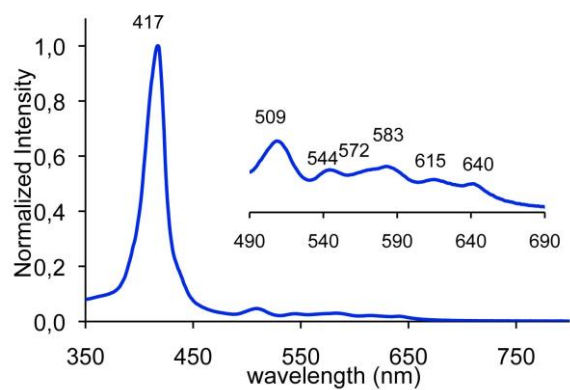


Figure S 21. Absorption spectrum of **1** in THF

5. Computed energy barriers

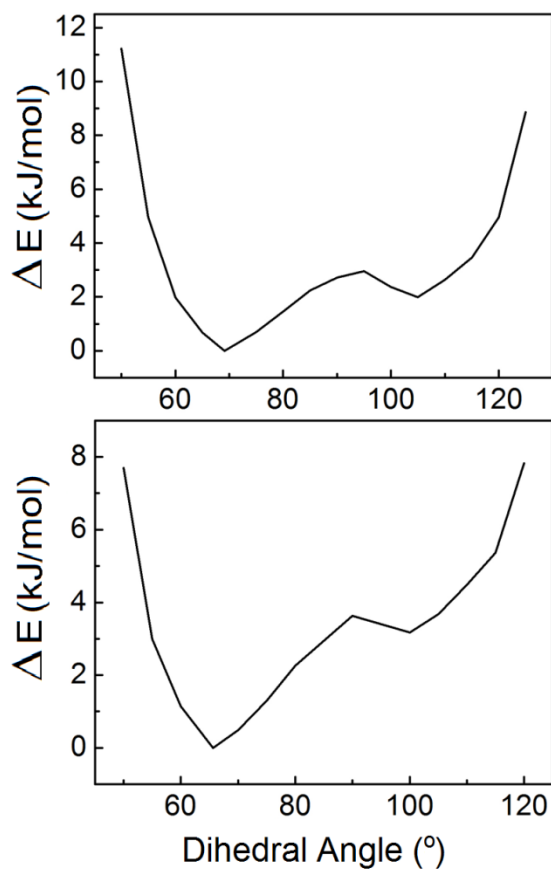


Figure S23. Change in total energy of **1** upon the change in dihedral angle formed by the Mg(II)porphyrin and C₆H₄ (up) and between the free base and C₆H₄ average planes (bottom). The minima are respectively, 71.5 and 67.5°.

6. Photophysical spectra and decays

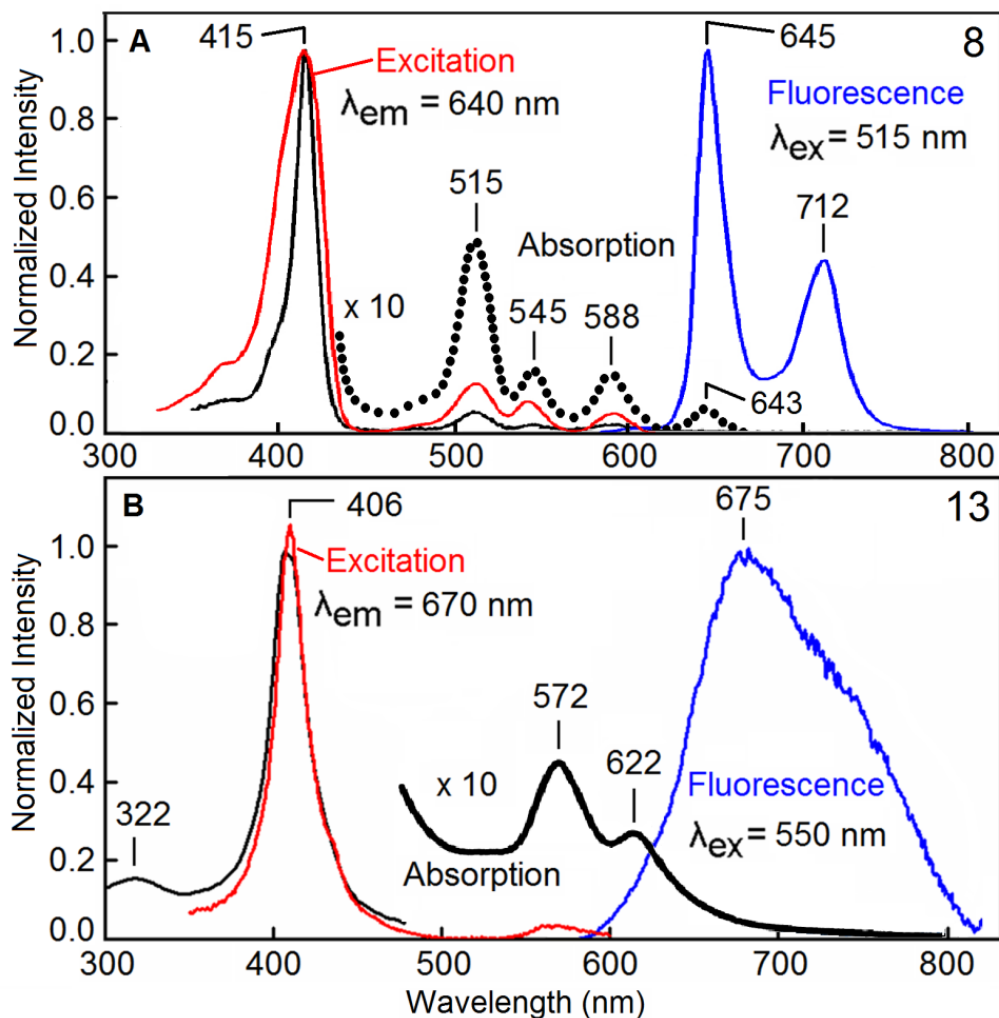


Figure S24 : Absorption (black), fluorescence (blue) and excitation (red) spectra of **8** (up) and **13** (down) in 2MeTHF at 298K.

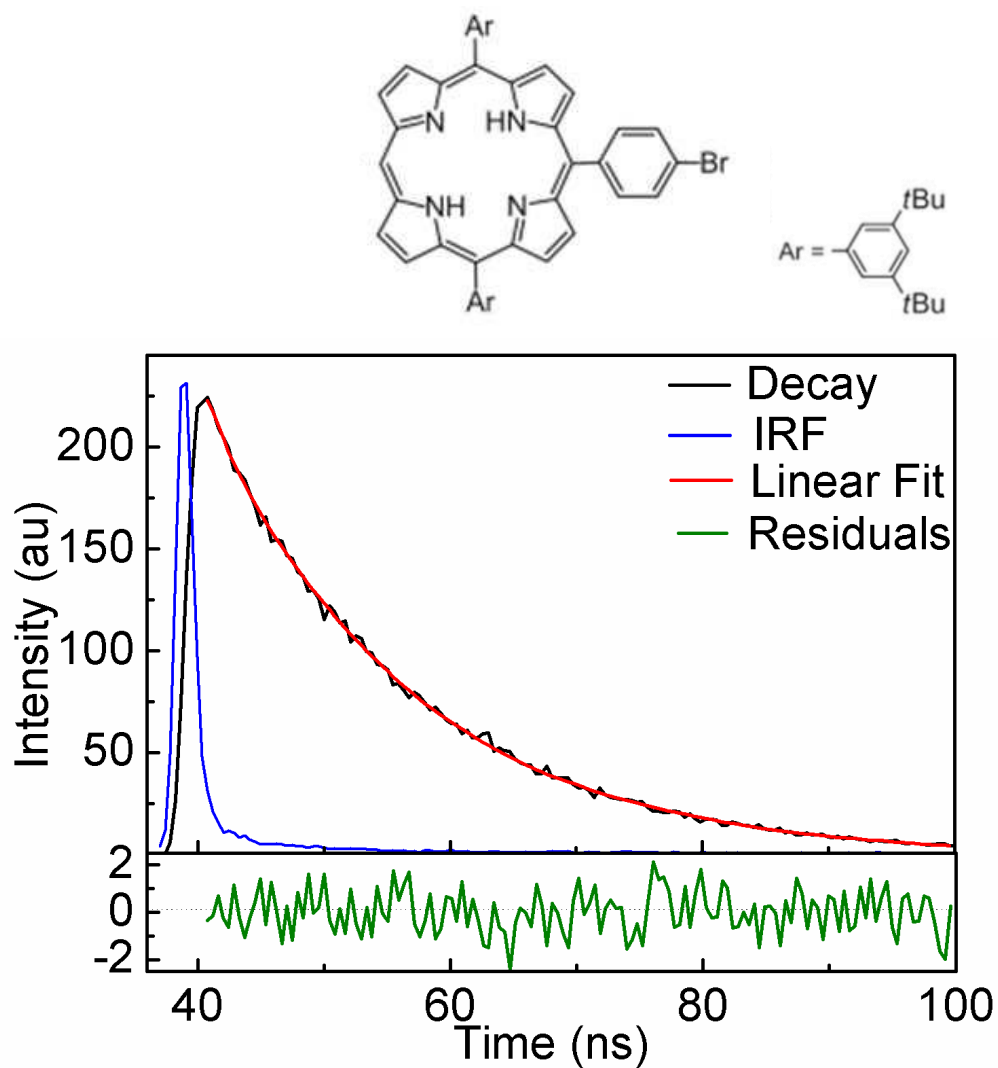


Figure S24 : Fluorescence decay curve for **8** in 2MeTHF at 298K. $\lambda_{\text{exc}} = 510 \text{ nm}$, $\lambda_{\text{em}} = 645 \text{ nm}$. Pre-exp = 0.537 ± 0.005 ; lifetime = $13.87 \pm 0.11 \text{ ns}$; $\chi^2 = 0.850$.

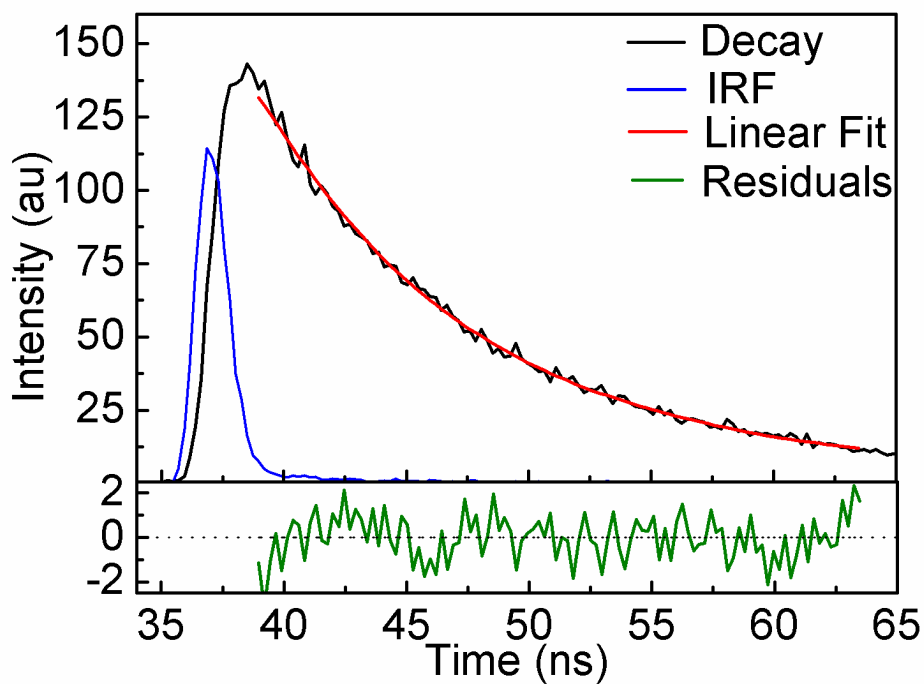
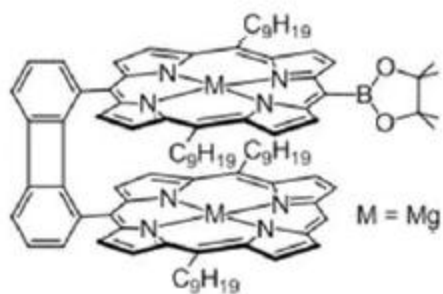


Figure S25 : Log (top) and linear scale (bottom) of the fluorescence decay curve for **13** in 2MeTHF at 298K. $\lambda_{\text{exc}} = 595 \text{ nm}$, $\lambda_{\text{em}} = 667 \text{ nm}$. Pre-exp = 0.846 ± 0.007 ; lifetime = $7.58 \pm 0.11 \text{ ns}$; $\chi^2 = 1.049$.

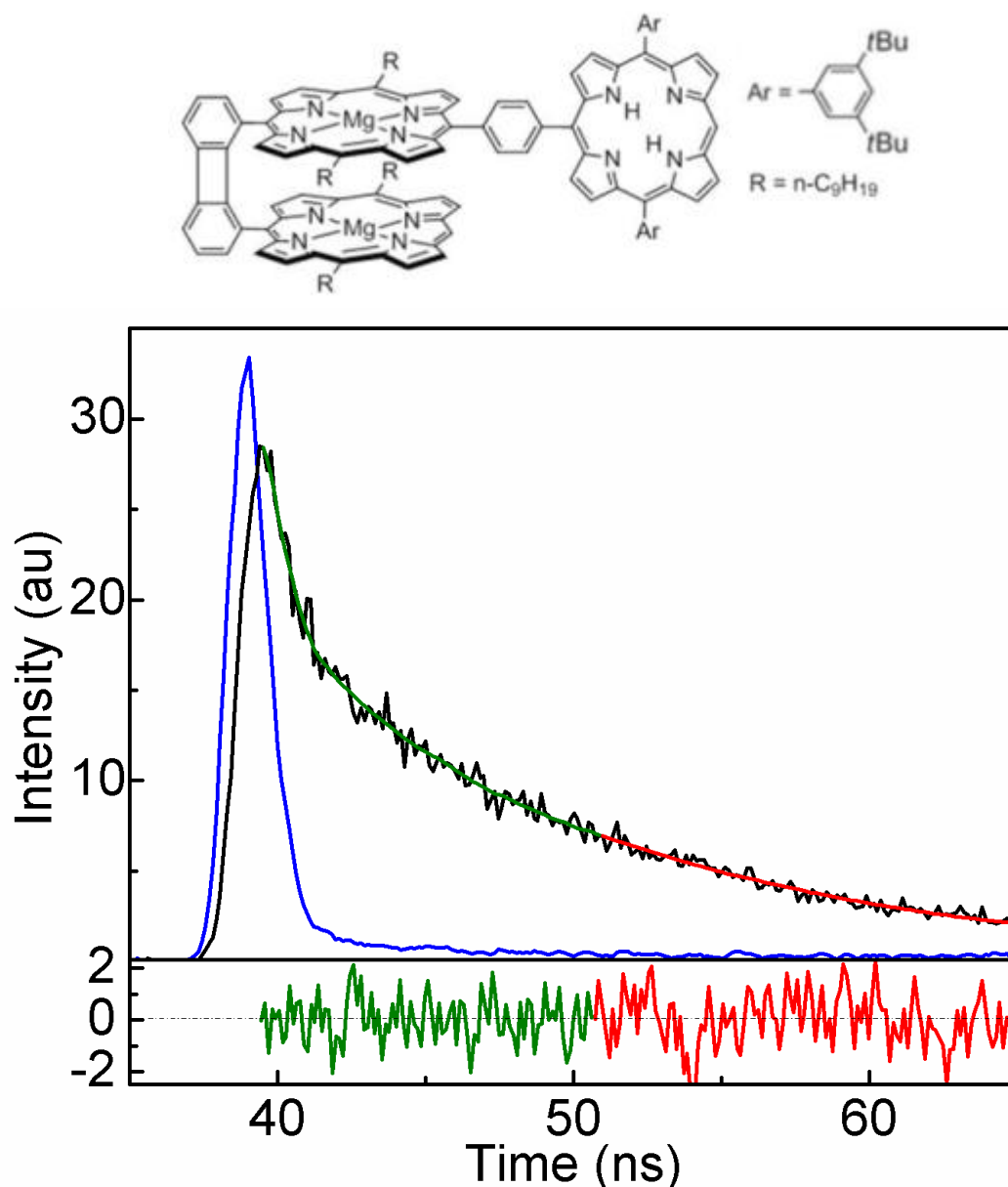


Figure S26. Fluorescence decay curve for **1** in 2MeTHF at 298K. $\lambda_{\text{exc}} = 510$ nm, $\lambda_{\text{em}} = 720$ nm. Fit for the first half (in green): Pre-exp. 1 = 2.45 ± 0.79 , lifetime 1 = 0.21 ± 0.06 ; Pre-exp. 2 = 0.329 ± 0.001 , lifetime 2 = 7.11 ± 0.85 ; $\chi^2 = 0.865$. Fit for the second half (in red): Pre-exp. = 0.335 ± 0.018 ; lifetime = 11.2 ± 1.20 ; $\chi^2 = 1.127$. The relative low counts of the decays are due to the sensibility limit of the Strobe detection.

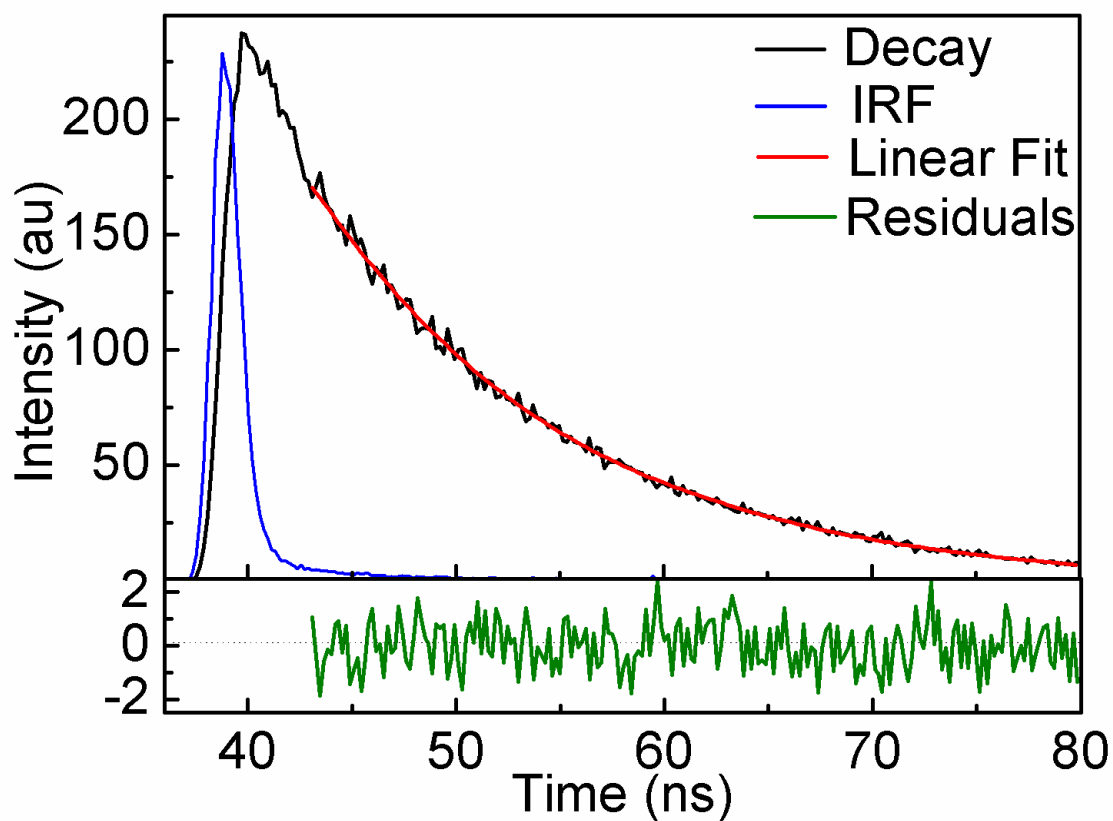


Figure S27. Fluorescence decay curve for **1** in 2MeTHF at 298K. $\lambda_{\text{exc}} = 510$ nm, $\lambda_{\text{em}} = 670$ nm. Pre-exp. = 0.601 ± 0.004 , lifetime = 10.99 ± 0.08 ; $\chi^2 = 0.8043$. The slower components of 0.21 and 7.1 ns associated with the bis(porphyrin unit) are not accurately determined in this decay due to their lower relative intensity.

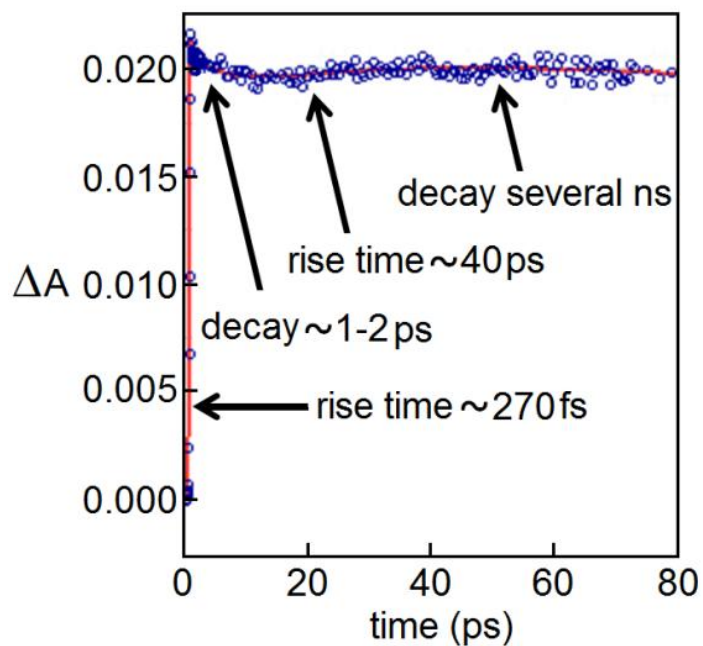


Figure 28. Rise and decay traces for the transient absorption spectra of **1** in 2MeTHF at 298 K monitored at 480 nm. Laser excitation = 400 nm, pulse width = 0.12 ps, laser power = 0.5 mJ/pulse, abs at 400 nm = 0.1. Note that the shape of the traces remains the same for wavelength > 480 nm. Because the time scale cannot exceed more than 3 ns on this apparatus, the long-lived components cannot be accurately analyzed.