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Rapid and sensitive LC-MS approach to quantify non-radioactive transition metal impurities in metal radionuclides

Supporting information:

Reagents. All solvents and reagents were purchased from commercial sources and used without additional purifications. The stable-isotope enriched *DL*-Phenylalanine-2,3-¹³C₂ (50 atom% ¹³C) and ethylene-*d*₄-diamine (98 atom % D) were purchased from CDN Isotopes (Quebec, Canada). Fmoc-Cl and formic acid (LC-MS Grade) were obtained from Fisher Scientific (Hanover Park IL). 4-Methyltrityl chloride resin and Fmoc-DL-Phe-OH were obtained from Chem-Impex International (Wood Dale, IL). LC-MS grade acetonitrile and water were obtained from Sigma Aldrich (St. Louis, MO). DOTA-NHS ester was obtained from Macrocyclics (Dallas, TX). Chelex@100 resins were obtained from Bio-Rad (Hercules, CA). Ultrapure Water was produced using a Thermo Scientific Barnstead Nanopure® Ultrapure Water Systems. ⁶⁴Cu²⁺ in 0.1 M HCl was obtained from Sigma Aldrich (St. Louis, MO). The reactions were conducted under N₂ atmosphere, unless described otherwise.

Instrumentation. ¹H NMR spectra were recorded on Brucker DRX 300 MHz. Waters LCT Premier mass spectrometer and 2695 Alliance HPLC were used for all the LC-MS experiments, with isocratic elution (Acetonitrile: $H_2O = 12$: 88, with 0.1% formic acid). Waters 1525 binary pump HPLC system was used for the EdF-DOTA agent purification using two elution buffers (0.1 v% TFA in de-ionized water as elution buffer A and 0.1 v%

TFA in acetonitrile as elution buffer B). Two Phenomenex HPLC columns, C18(2) Luna (250*10 mm) and C18(2) Luna (150 * 4.6 mm), were used for the HPLC and LC-MS experiments respectively.

Removal of trace amount of metal contamination from the ligand chelation. Before experiments of the EdF-DOTA ligand chelation, reagents and vessels used must be free from trace amount of metal contamination according to a previously reported protocol.¹ To remove trace amount of metal contaminants from reaction tubes, caps and pipette tips, the following procedure was used: 1) soak the tubes, caps and/or tips in 2.0 M nitric acid (diluted from concentrated nitric acid with ultrapure H₂O) overnight, and then drained; 2) wash with absolute ethanol and then drained; 3) wash with diethyl ether and then drain; 4) dry at room temperature under moderate nitrogen flow overnight. To remove trace amount of metal contaminants from reaction buffers, HPLC elution buffer and other solutions, the procedure is as follows: 1) add Chelex resin (10 g/L) to the buffers and solutions; 2) stir over night at room temperature; 3) filter through a Corning 1-liter filter system (pore size 0.2 mm).

Synthesis of Fmoc-($^{13}C_2$)Phe-OH. The title compound was synthesized through a procedure slightly modified from previously reported one² by using the $^{13}C_2$ -encoded phenylalanine instead of native phenylalanine: *DL*-phenylalanine-2,3- $^{13}C_2$ (50 atom% ^{13}C) (660 mg, 3.97 mmol) was stirred vigorously in 20 mL 5% K₂CO₃ with ice bath cooling. A solution of Fmoc-Cl (1.2 g, 4.2 mmol) in 10 mL tetrahydrofuran (THF) was added slowly, and then the reaction mixture was stirred for 3 h while it was allowed to warm up to room temperature gradually. After the reaction was complete,

THF was removed in *vacuo*. The residue was poured into 80 mL 1 N HCl, and then extracted with 70 mL dichloromethane (DCM) three times. The combined organic solvent was washed with 100 mL brine, and dried over anhydrous Na₂SO₄. The product was purified by silica gel chromatography with ethyl acetate/hexane (1:1) to obtain the title compound (1.17 g, 76%) as a white powder. The ¹H NMR matched that of the previously synthesized Fmoc-DL-Phenylalanine.³ ¹H NMR (400 MHz, CDCl3) δ 3.09-3.14 (m, 1H), 3.20-3.24 (m, 1H), 4.18-4.22 (t, 1H), 4.33-4.38 (m, 1H), 4.43-4.47 (m, 1H), 4.70-4.74 (m, 1H), 5.17-5.21(d, 1H), 7.14-7.16 (d, 2H), 7.22-7.42 (m, 7H), 7.53-7.56 (m, 2H), 7.75-7.77 (d, 2H); ESI-MS: (M+H)⁺ = 389.51.



Synthesis of the ¹³C/²H labeled EdF-DOTA. The synthesis is shown in Scheme S1. 4-Methyltrityl chloride resin (20 μ mol) was incubated with ethylenediamine (100 μ mol) and triethylamine (TEA, 200 μ mol) in 1 mL dimethylformamide (DMF) for 2 h, and was then washed thoroughly with DMF and DCM respectively. Fmoc- $(^{13}C_2)$ Phe-OH (100 µmol) was then coupled to the resin by mixing with PyBrOP (100 µmol) and DIPEA (200 µmol) in 1 mL DMF for 2 h at room temperature. Afterwards, the Fmoc group was removed with 20% piperidine in DMF (1.0 mL*3). After washing with DMF and DCM, the resin was incubated with the mixture of DOTA-NHS (60 µmol) and DIPEA (250 µmol). Finally, ¹³C/²H labeled EdF-DOTA was cleaved from the resin with 20% TFA in DCM. The purification of EdF-DOTA was performed using a Phenomenex Luna C18(2) (250*10 mm) column with the following elution conditions: 4.0 mL/minute flow rate; 0-5 minutes 100% A, 5-25 minutes 100% A to 70% A, 25-30 minutes 70% A to 10% A. Under these condition, the EdF-DOTA agent was eluted out at ~18.4 min. The identity and purity of the purified ${}^{13}C/{}^{2}H$ EdF-DOTA was confirmed by LC-MS using a Phenomenex C18(2) Luna (150*4.6 mm) column with the following elution conditions: 1.5 mL/minute flow rate; 0-5 minutes 100% A, 5-20 minutes 100% A to 60% A, 25-30 minutes 10% A. The peak at ~10.1 min corresponded to pure ${}^{13}C/{}^{2}H$ labeled EdF-DOTA (calculated, m/z (M+H)⁺, 599.67; observed, $m/z (M+H)^+ = 599.66$).

Synthesis of native EdF-DOTA. The light-tagged agent was synthesized and purified in a similar manner using native ethylenediamine and phenylalanine. Its identity and purity were confirmed under the same LC-MS conditions, and the peak at ~10.1 min corresponded to pure native EdF-DOTA. (Calculated, m/z (M+H)⁺, 594.68; observed, m/z (M+H)⁺ = 594.69) **Preparation of internal standard solutions:** Native Fe(III)DOTA-EdF (10 μ M) was prepared by incubating 500 μ M native EdF-DOTA ligand with 100 μ M Fe³⁺ solution in 0.1 M NH₄OAc buffer (pH = 6.80) at 60 °C for 30 minutes and diluted 5-fold with 0.1 M NH₄OAc buffer (pH = 6.80). Other four internal standards were prepared in a similar manner using Ni²⁺, Cu²⁺, Co²⁺, Zn²⁺, respectively. The internal standard solutions were stored at -80 °C.

Quantification of the ¹³C/²H labeled EdF-DOTA with Cu-64. Copper-64 (in 0.1 N HCl) obtained from Wisconsin was immediately neutralized and buffered with 0.1 M NH₄OAc (pH = 6.8), to a final concentration of 20 μ Ci/ μ L (decay corrected to the time of receipt). After one week when the Cu-64 was completely decayed, 10 μ L of the stock solution (200 μ Ci at the time of receiving activity) was mixed with 10 μ L of 0.5 mM ¹³C/²H labeled EdF-DOTA and 55 μ L of 0.1 M NH₄OAc buffer (pH = 6.80). The mixture was incubated at 22 °C for 10 (or 30) min or at 60 °C for 30 min, and then the five internal standards (10 μ M) prepared earlier were added (5.0 μ L for each standard, 25 μ L in total) and mixed thoroughly. The entire mixture was transferred and loaded for LC-MS analysis. As described in **Fig. 2**, the relative amount of a particular heavy complex can be quantified by comparing its total ion counts with the corresponding native complexes.





References:

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- 2. H. Ishiyama, K. Yoshizawa and J. Kobayashi, *Tetrahedron*, 2012, **68**, 6186-6192.
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