Electronic Supplementary Information for

A Simple and Highly Selective 2,2-diferrocenylpropane-based Multichannel Ion Pair Receptor for Pb²⁺ and HSO₄-

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1. Materials and instruments

All of the starting materials were obtained from commercial suppliers and used without further purification. All reactions were performed under the nitrogen atmosphere, the solvents were dried by standard methods and then distilled prior to use. NMR spectra were recorded on a Bruker Avance III 400 NMR apparatus. Highresolution mass spectra (HRMS) were obtained on a Bruker En Apex ultra 7.0 FT-MS spectrometer. Melting points were determined with a 4X microscopic melting point instrument. The UV-visible spectra were measured on a Perkin Elmer lambda 750UV/VIS/NIR spectrometer. Fluorescence spectra were collected on an Edinburgh FL-FS 920 TCSPC. Fluorescence images were obtained on a CEL-M 500 high-pressure mercury lamp. X-ray structural measurements were carried out with a Rigaku RAXISIV CCD diffract meter with a graphitemonochromator Mo-Ka radiation (λ = 0.071073 nm) at 293(2) K. All diffraction data were collected by scanning in a certain mode and refined in Lp factor. Theoretical calculation for the geometrical optimization of receptor 1 and complex 1·Pb²⁺ was performed at the density functional theory (DFT) level using the B3LYP hybrid functional. The 6-31G and 6-31G(d) basis sets were employed for H and C atoms, aug-cc-pVTZ for N atom and LANL2DZ with effective core potential for Fe and Pb atom, respectively. In addition, the energy of the receptor 1 and complex 1·Pb²⁺ were -41493.6 eV and -41575.2 eV, respectively. After obtaining the optimized structure, the further time-dependent density functional theory (TD-DFT) calculation was performed to investigate the excited states of receptor **1**. All calculations were performed using the Gaussian 09 package.¹

Cyclic voltammetry (CV) and Differential pulse voltammetry (DPV) studies were performed by CHI620C electrochemical workstation with a conventional three-electrode configuration, glassy carbon as the working electrode, platinum electrode as counter electrode, and Ag/AgCl (3.0 M KCl) as reference electrode at room temperature (scan rate: 100 mV/ s). The experiments were carried out with a 2 \times 10⁻⁴ M solution of receptor **1** in CH₃CN containing 0.1 M (*n*-C₄H₉)₄ClO₄ (TBAP) as supporting electrolyte for cations, [*n*-Bu₄N]PF₆ (TBAPF₆) (c = 0.10 M) for anions.

2. Synthesis details and characteristic data

2.1 The Synthetic routine of 1

In a 100 mL round bottom flask 2,2-Diferrocenylpropanecarbaldehyde **2** (510 mg, 1.2 mmol) and 2,3diaminopyridine **3** (200 mg, 1.9 mmol) were dissolved in MeOH (60 mL). Iodine was added into the solution for initiating the reaction. The mixture was stirred at reflux 24 h under nitrogen atomosphere to complete the reaction which was monitored by TLC. The reaction mixture obtained was allowed to cool to room temperature and solvent was evaporated under reduced pressure to give the residue. The residue was redissolved in dichloromethane and washed with saturated NaS₂O₄ until colorless in the aqueous layer. Then the organic layer was washed with saturated NaCl, dried over Na₂SO₄ and concentrated to give the crude product, which was further purified by column chromatography over basic alumina using petroleum ether/ethyl acetate (2:1) mixture as eluent to give receptor **1** as a yellow solid (235mg, Yield = 38.6%). mp: >200 °C. Anal. Found: C, 65.69, H, 5.39, N, 7.88. Calc. for C₂₉H₂₇Fe₂N₃ (529.09): C, 65.81; H, 5.14; N, 7.94%. IR (KBr) v_{max} /cm⁻¹: 3096, 2968, 1562, 1404, 1330, 1271, 1032, 818, 777; ¹H NMR $\delta_{\rm H}$ (400 MHz, DMSO-d₆.) 12.98 (1H, s , NH), 8.25 (1H, d, *J* = 3.8 Hz, Py-H), 7.88 (1H, d, *J* = 7.2 Hz, Py-H), 7.17 (1H, dd, *J* = 7.7, 4.8 Hz, Py-H), 5.07-3.94 (m, 17H, Cp-H), 1.55 (s, 6H, Me). ¹³C NMR δ (101 MHz, DMSO-d₆, Me₄Si) 103.10, 101.37, 73.77, 71.28, 69.10, 68.54, 68.41, 67.52. 66.89, 65.93, 33.04, 30.14. HRMS (ESI): Found: 530.0972, Calc. for C₂₉H₂₇Fe₂N₃ [M+H]⁺: 530.0982.

2.2 Characteristic data



Fig. S1 ¹H NMR spectrum of 1 in DMSO-d₆



Fig. S2 ¹³C NMR spectrum of 1 in DMSO-d₆



Fig. S3 HRMS spectrum (ESI positive ion) of 1



Fig. S4 ORTEP of 1 (thermal ellipsoids were set at 50% probability).

3. Optical Spectroscopy Data



Fig. S5 Fluorescent response of 1 to metal cations, $(F-F_0)/F_0$ was calculated according to the emission at 420 nm.



Fig. S6 Visual features observed in CH₃CN solutions of 1 (1 \times 10⁻⁴ M) towards metal cations.



Fig. S7 Job's plots for the binding of 1 with Pb²⁺ using fluorescence method, $\lambda_{ex} = 350$ nm and $\lambda_{em} = 420$ nm.



Fig. S8 Job's plots for the binding of 1 with Zn²⁺ using Absorption spectra, absorption at 311 nm.



Fig. S9 Determination of association constant of 1 for Pb²⁺ in CH₃CN, $\lambda_{ex} = 350$ nm, $\lambda_{em} = 420$ nm.



Fig. S10 Determination of association constant of 1 for Zn²⁺ in CH₃CN, $\lambda_{ex} = 350$ nm, $\lambda_{em} = 420$ nm, $K_a = 1.21 \times 10^5$ M⁻¹.



Fig. S11 Fluorescence spectra of 1, $1 \cdot \text{HSO}_{4^-}$, $1 \cdot \text{Zn}^{2+}$ and $1 \cdot \text{Zn}^{2+} \cdot \text{HSO}_{4^-}$ (10 μ M), excited at 350 nm.



Fig. S12 Color changes of complex $1 \cdot Pb^{2+}$ (1 × 10⁻⁴ M) in the presence of 2 equiv. corresponding anions: naked eye (top); under mercury lamp (bottom) in CH₃CN.



Fig. S13 Changes in the absorption spectra of 1 (2.5 \times 10⁻⁵ M) in CH₃CN upon addition of HSO₄⁻, Pb²⁺, Pb²⁺ and HSO₄⁻.



Fig. S14 Stepwise complexation/decomplexation cycles of receptor 1 (2.5×10^{-5} M in CH₃CN) and Pb²⁺ using [*n*-Bu₄N]F as decomplexation agent; carried out by UV/Vis analysis.



Fig. S15 Evolution of the CV (left) and DPV (right) of 1 (2×10^{-4} M) in CH₂Cl₂ when Zn²⁺ is added: from 0 to 2 equiv.



Fig. S16 Evolution of the CV (left) and DPV (right) of 1 (2×10^{-4} M) in CH₂Cl₂ when 8 equiv. HSO₄⁻ is added.



Fig. S17 Evolution of the CV (left) and DPV (right) of $1 \cdot Pb^{2+}$ in CH_2Cl_2 by gradually addition of HSO_4^- up to 10 equiv.



Fig. S18 Fluorescence spectra of 1, $1 \cdot Pb^{2+} 1 \cdot Pb \cdot HSO_4^-$, $1 \cdot Zn^{2+} \cdot HSO_4^-$ (10 µM) in CHCl₃, excited at 350 nm.



Fig. S19 Job's plots for receptor 1 with HSO₄⁻ in CDCl₃ by monitoring the H2 proton of pyridine ring.



Fig. S20 ¹H NMR titration curves of receptor 1 in CDCl₃ by monitoring the H2 proton of pyridine ring.



Fig. S21 Complexation/decomplexation process for receptor 1 (6.61 \times 10⁻³ M) using [*n*-Bu₄N]F as decomplexation agent, carried out by ¹H NMR analysis in CDCl₃.



Fig. S22 Mass spectra of receptor 1 upon addition of Pb²⁺ and HSO₄⁻ in CH₃CN.

Table S1. Selected data of electronic transitions in 1 by TD-DFT calculations using B3LYP/Gen method.

$\lambda [nm/(eV)]$	fa	Composition of bands and CI ^b coefficients
495 (2.50)	0.0037	H-3→L+1, 0.16
488 (2.54)	0.0011	H-2→L+1, 0.14
311 (3.98)	0.1104	H-2→L, 0.21; H-2→L+2, 0.18
288 (4.29)	0.4602	H-4→L, 0.40
286 (4.33)	0.018	H→L+1, 0.40

^a *f*- oscillator strength; ^b CI- configurational intergration coefficient, H - HOMO, L - LUMO.





Fig. S23 Frontier molecular orbitals of 1 contributing to UV-visible absorption bands.

Table S2. Evolution of ¹ HNMR data for marked H in the presence of several anions and cation	ons
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	H1	H2	Н3	Ηα	Hβ
	(Δδ)	(Δδ)	(Δδ)	(Δδ)	(Δδ)
1	8.478	7.310	8.060	5.093	4.553
$1 \cdot Zn^{2+}$	8.515	7.285	8.094	4.722	4.392
	(+0.037)	(-0.025)	(+0.034)	(-0.371)	(-0.161)
1.Pb ²⁺	8.359	7.452	8.332	5.269	4.782
	(-0.119)	(+0.142)	(+0.272)	(+0.176)	(+0.229)
1·HSO ₄ -	8.230	7.181	8.039	5.112	4.413
	(-0.248)	(-0.129)	(-0.021)	(+0.019)	(-0.140)
1·Pb ²⁺ ·HSO ₄ -	8.344	7.285	8.344	5.246	4.469
	(-0.134)	(-0.025)	(+0.284)	(+0.153)	(-0.084)

C ₃₁ H ₃₃ Fe ₂ N ₃ OS 607.36 293(2)
607.36 293(2)
293(2)
0.71073
Monoclinic
P21/n
11.1271(7)
19.1069(12)
13.0946(10)
90.00
94.223(5)
90.00
2776.4(3)
4
1.453
1264
$0.50\times0.40\times0.20$
$-14 \le h \le 14$,
$-24 \le k \le 24$,
-17 51517
21944
$5282 R_{All} = 0.0429$
0.987
Full-matrix least-squares on F^2
$R_I = 0.1268,$
$wR_2 = 0.3388$
$R_I = 0.1422,$
$wR_2 = 0.3465$ 2.217

 Table S3.Crystallographic data and structure refinement parameters.

bond	(Å)	bond	(Å)
N1—C25	1.363(10)	C10—C11	1.533(8)
N1—C24	1.391(10)	C11—C13	1.535(8)
N2—C24	1.316(11)	C11—C12	1.540(9)
N2—C29	1.346(11)	C11—C14	1.540(9)
N3—C28	1.308(18)	C22—C23	1.433(13)
N3—C29	1.392(15)	C23—C24	1.467(11)
A			
Angles	(°)	Angles	(⁰)
Angles C25—N1—C24	(°) 104.8(7)	Angles C13—C11—	(°) 110.3(5)
Angles C25—N1—C24 C24—N2—C29	(°) 104.8(7) 105.4(8)	Angles C13—C11— C14—C11—	(°) 110.3(5) 110.7(5)
Angles C25—N1—C24 C24—N2—C29 C28—N3—C29	(°) 104.8(7) 105.4(8) 118.6(11)	Angles C13—C11— C14—C11— C10—C11—	(°) 110.3(5) 110.7(5) 104.7(5)
Angles C25—N1—C24 C24—N2—C29 C28—N3—C29 C13—C11—	(°) 104.8(7) 105.4(8) 118.6(11) 110.2(6)	Angles C13—C11— C14—C11— C10—C11— N2—C24—N1	(°) 110.3(5) 110.7(5) 104.7(5) 113.2(8)
Angles C25—N1—C24 C24—N2—C29 C28—N3—C29 C13—C11— C10—C11—	(°) 104.8(7) 105.4(8) 118.6(11) 110.2(6) 111.4(5)	Angles C13—C11— C14—C11— C10—C11— N2—C24—N1 N2—C24—C23	(°) 110.3(5) 110.7(5) 104.7(5) 113.2(8) 127.3(8)

Table S4. Selected bond lengths (Å) and angles (°) for 1.

References

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