

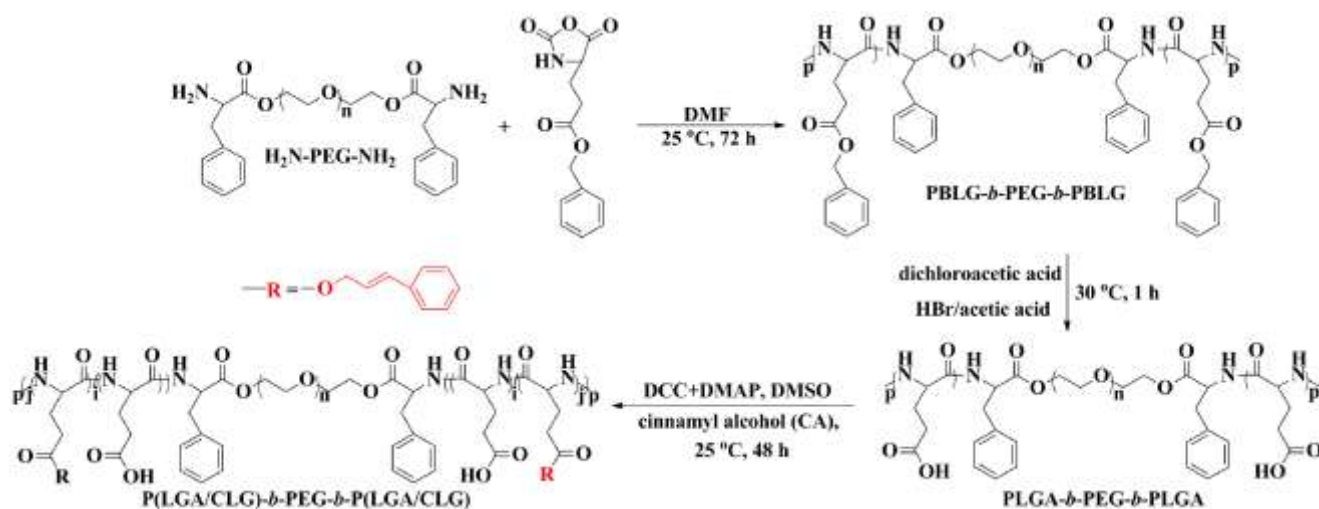
Supporting Information

Preparation of photo-cross-linked pH-responsive polypeptide nanogels as potential carriers for controlled drug delivery

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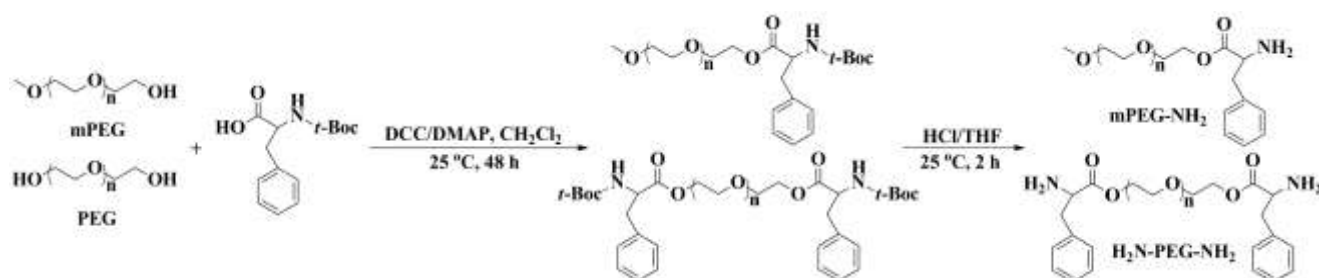
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Scheme S1 Synthesis procedure for P(LGA/CLG)-*b*-PEG-*b*-P(LGA/CLG).

Synthesis of amino group terminated poly(ethylene glycol) monomethyl ether (mPEG-NH₂) and poly(ethylene glycol) (H₂N-PEG-NH₂) macroinitiator

mPEG₁₁₃-NH₂ and H₂N-PEG₁₀₄-NH₂ were synthesized by condensation reaction with dicyclohexylcarbodiimide (DCC)/4-*N,N*-dimethylaminopyridine (DMAP) as the coupling reagents and then the deprotection of *tert*-butoxycarbonyl (*t*-Boc) group as depicted in Scheme S2. Briefly, mPEG ($M_n = 5000$, 10.0 g, 2.0 mmol), *N*-(*tert*-Butoxycarbonyl)-L-phenylalanine (0.53 g, 2.0 mmol) and DMAP (24.43 mg, 0.2 mmol) were dissolved in 50 mL dry methylene chloride (CH₂Cl₂) in a flame-dry flask, then DCC (2.06 g, 10.0 mmol) in CH₂Cl₂ was added slowly to the solution with stirring and the reaction was conducted at room temperature for 48 h. The by-product dicyclohexylurea (DCU) precipitate was removed by filtration. Then, the solution was precipitated into excessive diethyl ether. The obtained product was further washed twice with diethyl ether and dried under vacuum at room temperature for 24 h (Yield: 96%). Subsequently, mPEG-NH-*t*-Boc (9.0 g, 1.72 mmol) was dissolved in 50 mL tetrahydrofuran (THF) at 25 °C in a flask. After 10 mL HCl/THF saturated solution was added, the solution was slowly stirred at 25 °C for 2 h and then the final product was precipitated into excessive diethyl ether and washed twice with diethyl ether. The precipitate was collected and dried under vacuum to a constant weight at room temperature. The yield was approximately 97%. The chemical structure of mPEG₁₁₃-NH₂ and H₂N-PEG₁₀₄-NH₂ were characterized by ¹H NMR and ¹³C NMR, as shown in Fig. S1 and S2.



Scheme S2 Synthesis procedures for mPEG-NH₂ and H₂N-PEG-NH₂.

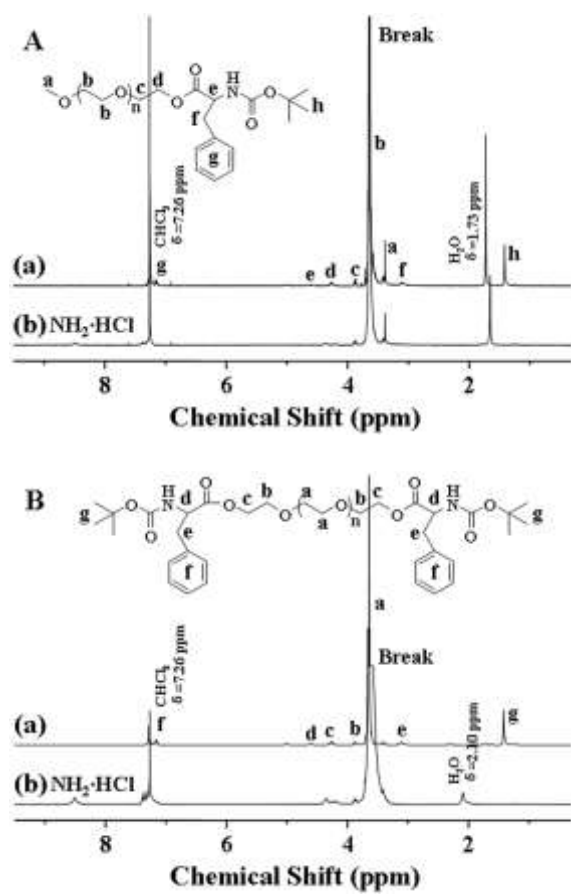


Fig. S1 ^1H NMR of macroinitiators before (a) and after (b) deprotection, mPEG₁₁₃ (A), and PEG₁₀₄ (B) (in CDCl_3).

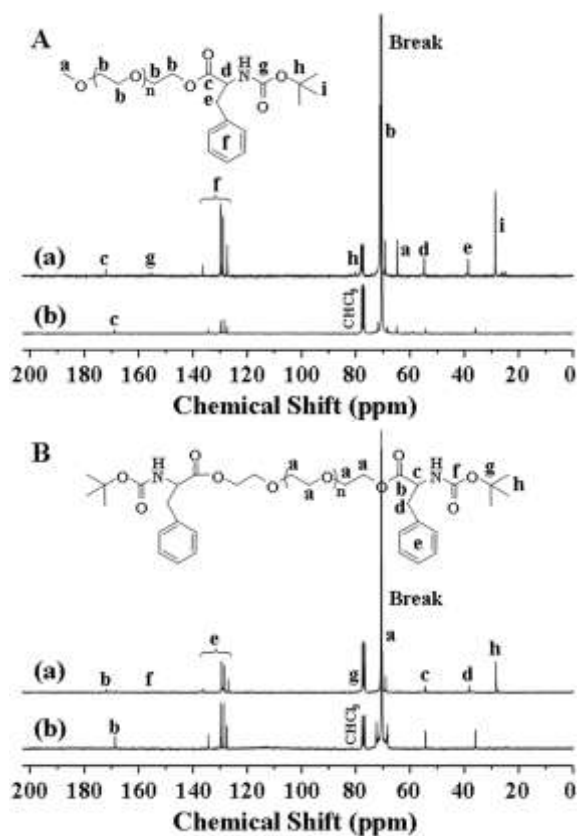


Fig. S2 ^{13}C NMR of macroinitiators before (a) and after (b) deprotection, mPEG_{113} (A), and PEG_{104} (B) (in CDCl_3).

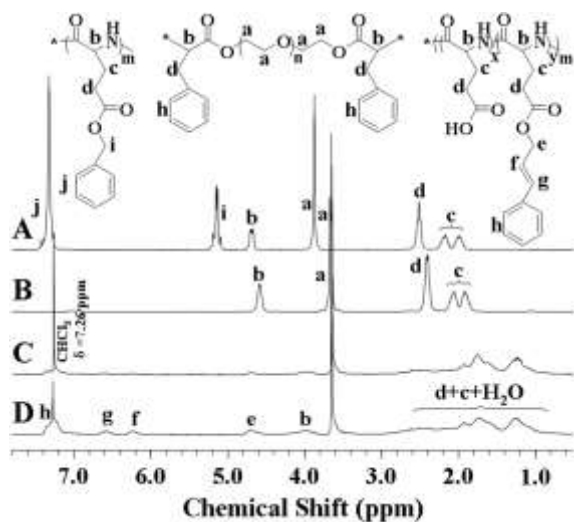


Fig. S3 ^1H NMR spectra of $\text{PBLG}_{106}\text{-}b\text{-PEG}_{104}\text{-}b\text{-PBLG}_{106}$ (A), $\text{PLG}_{106}\text{-}b\text{-PEG}_{104}\text{-}b\text{-PLG}_{106}$ (B), $\text{P(LGA}_{87}\text{/CLG}_{19})\text{-}b\text{-PEG}_{104}\text{-}b\text{-P(LGA}_{87}\text{/CLG}_{19})$ **2** (C) and $\text{P(LGA}_{68}\text{/CLG}_{38})\text{-}b\text{-PEG}_{104}\text{-}b\text{-P(LGA}_{68}\text{/CLG}_{38})$ **3** (D) (in $\text{CF}_3\text{COOD}+\text{CDCl}_3$ (1/1, v/v) for (A), CF_3COOD for (B), and CDCl_3 for (C) and (D)).

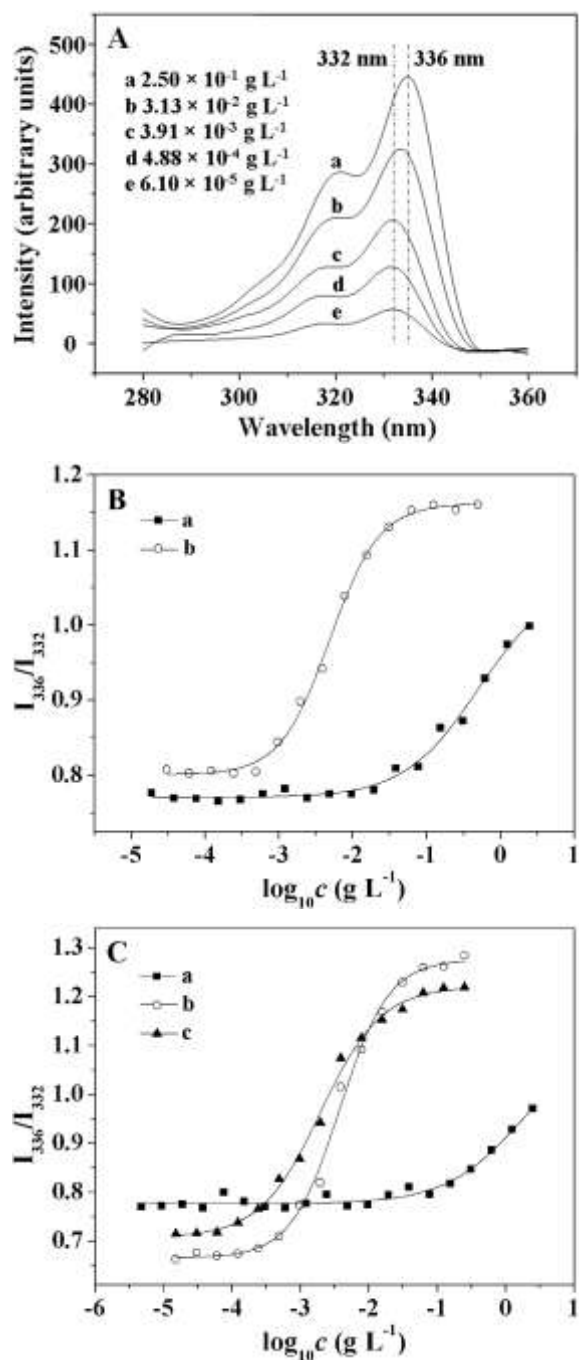


Fig. S4 Excitation spectra of pyrene in aqueous solution of mPEG₁₁₃-b-P(LGA₇₃/CLG₃₇) at different concentrations ($\lambda_{\text{em}} = 390 \text{ nm}$) (A), the intensity ratio (I_{336}/I_{332}) as a function of concentration of mPEG₁₁₃-b-PLGA₁₁₀ (a) and mPEG₁₁₃-b-P(LGA₇₃/CLG₃₇) (b) (B), and PLGA₁₀₆-b-PEG₁₀₄-b-PLGA₁₀₆ (a), P(LGA₈₇/CLG₁₉)-b-PEG₁₀₄-b-P(LGA₈₇/CLG₁₉) (b) and P(LGA₆₈/CLG₃₈)-b-PEG₁₀₄-b-P(LGA₆₈/CLG₃₈) (c) (C).

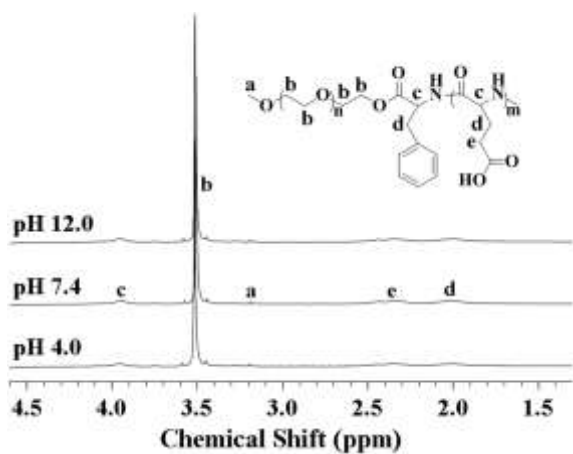


Fig. S5 ^1H NMR spectra of $\text{mPEG}_{113}\text{-}b\text{-PLGA}_{110}$ at pH 4.0, 7.4, and 12.0 (in D_2O).

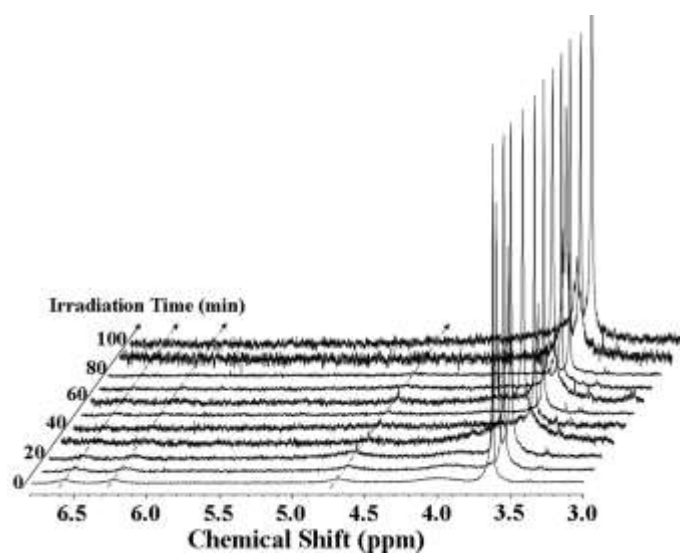


Fig. S6 Change in the ^1H NMR spectra of $\text{mPEG}_{113}\text{-}b\text{-P(LGA}_{73}\text{/CLG}_{37})$ **1** as a function of irradiation time (in CDCl_3) (nanogels were prepared in aqueous solution at pH 7.4).

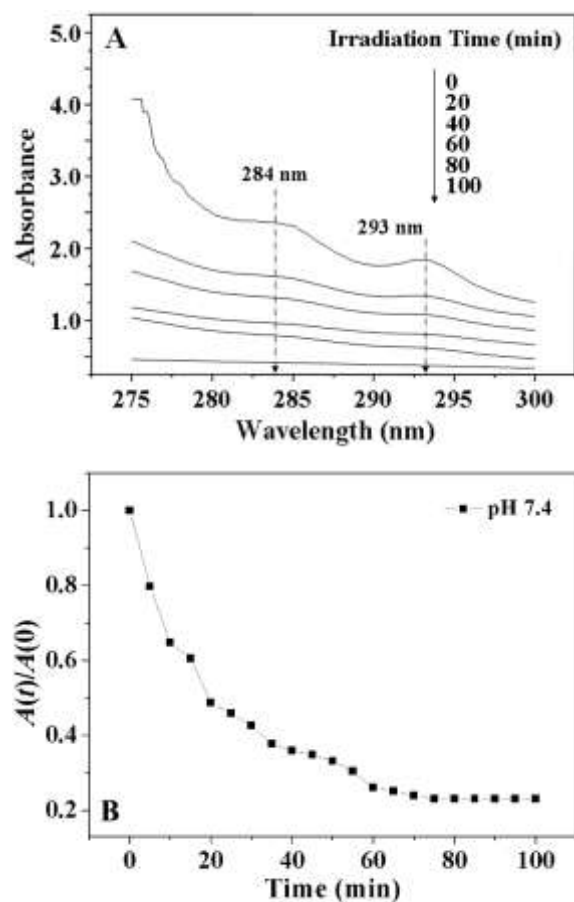


Fig. S7 UV-vis spectra of P(LGA₈₇/CLG₁₉)-*b*-PEG₁₀₄-*b*-P(LGA₈₇/CLG₁₉) aqueous solution of 0.5 g L⁻¹ at pH 7.4 subjected to UV irradiation at $\lambda = 254$ nm as a function of irradiation time (A), decrease in $A(t)/A(0)$ for the cinnamyl residues determined from the UV-vis absorption spectra at pH 7.4 plotted as a function of irradiation time (B).

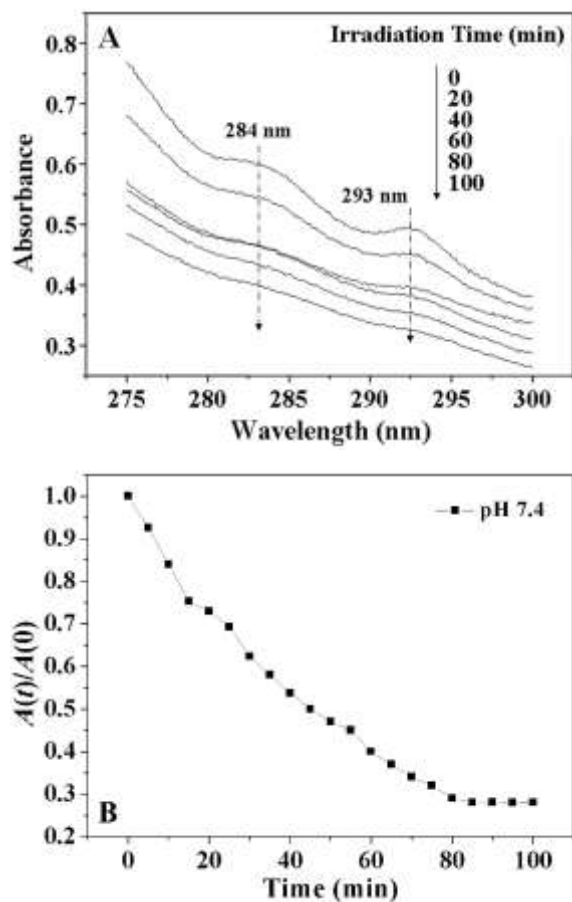
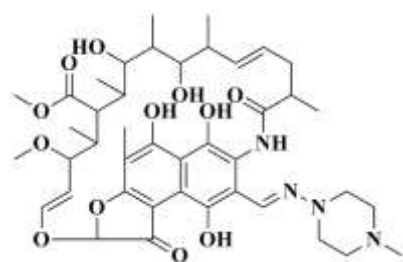


Fig. S8 UV-vis spectra of $P(LGA_{68}/CLG_{38})-b-PEG_{104}-b-P(LGA_{68}/CLG_{38})$ aqueous solution of 0.5 g L^{-1} at pH 7.4 subjected to UV irradiation at $\lambda = 254 \text{ nm}$ as a function of irradiation time (A), decrease in $A(t)/A(0)$ for the cinnamyl residues determined from the UV-vis absorption spectra at pH 7.4 plotted as a function of irradiation time (B).



Scheme S3 Chemical structure of rifampin.

The rifampin chemical structure was used to explain the release behavior as an ancillary factor.