

Electronic Supplementary Information for the manuscript

Dendritic polyglycerolamine as a functional antifouling coating of gold surfaces

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Spectroscopic Characterization of starting PG

¹H NMR (400 MHz, MeOD, δ): 4.05-3.45 (m, CH₂ and CH, PG backbone), 1.35 (m, CH₂ core), 0.86 (m, CH₃ core). **IR** (KBr) ν_{max} = 3362s (OH), 2913m and 2877m (CH), 1651w, 1456w, 1106s and 1076s (C-OH).

Synthesis of polyglycerol derivatives

Synthesis of mesylpolyglycerol, PG-OMs (example given for 9% of conversion)

In a three-necked flask equipped with magnetic stirring, PG-OH (2.957g, 39.92 mmol -OH) was dissolved in pyridine (45 mL) and cooled to 0 °C. Then, MsCl (0.34 mL, 4.39 mmol, 0.11 eq.) was dissolved in pyridine (2 mL) and added dropwise through syringe. After stirring the solution for 1 h at 0 °C, it was allowed to reach room temperature and was stirred for 16 h. A yellowish-brown mixture was obtained. The solvent was removed under vacuum and the resulting mixture was dialyzed in MeOH to afford 3.13 g of PG-OMs as a honey-like product. Conversion = 9%, quantified by ¹H RMN. Yield= 97%. **¹H NMR** (400 MHz, MeOD, δ): 4.55-3.40 (m, CH₂ and CH, PG scaffold); 3.30-3.17 (m, CH₃ mesyl), 1.35 (m, CH₂ core); 0.87 (m, CH₃ core). **IR** (KBr) ν_{max} = 3376s (OH), 2929m and 2877m (CH), 1651w, 1456w, 1352w and 1173w (S=O mesyl), 1106s and 1076s (C-OH).

Synthesis of polyglycerolazide, PG-N₃

In a Schlenk tube equipped with magnetic stirring, PG-OMs (3.130 g, 3.82 mmol -OMs) was dissolved in DMF (20 mL). Then, NaN₃ (1.24 g, 19 mmol, 5 eq.) was added and the suspension was stirred for 72 h at 60 °C. The resulting reddish-brown mixture was filtered,

and the supernatant was dialyzed in MeOH to afford 2.53 g of PG-N₃. Conversion: quantitative (determined by ¹H RMN and IR), yield = 86%. **¹H NMR** (400 MHz, MeOD, δ): 4.00-3.40 (m, CH₂ and CH, PG scaffold), 1.43 (m, CH₂ core), 0.91 (m, CH₃ core). **IR** (KBr) $\nu_{\text{max}} = 3375s$ (OH), 2915*m* and 2877*m* (CH), 2102*m* (-N₃), 1109*s* and 1078*s* (C-OH).

Synthesis of polyglycerolamine, PG-NH₂

In a one-necked flask equipped with magnetic stirring, PG-N₃ (0.700 g, 0.92 mmol -N₃) was dissolved in 50 mL of THF/water (70:30) and PPh₃ (0.72 g, 2.76 mmol, 3 eq.) was added. While stirring , the proportion of water was continuously increased in order to adjust the solubility of the partially reduced product. The resulting mixture was filtered, concentrated in vacuum, extracted three times with CHCl₃ and finally dialyzed in MeOH. 0.62 g of a pale yellow product, PG-NH₂, was obtained. Conversion: quantitative (determined by IR), yield = 92%. **¹H NMR** (400 MHz, MeOD, δ): 4.55-3.40 (m, CH₂ and CH, PG scaffold), 3.16-2.60 (m, OCH-NH₂ and CH₂-NH₂), 1.41 (m, CH₂ core), 0.91 (m, CH₃ core). **IR** (KBr) $\nu_{\text{max}} = 3364s$ (OH), 2915*m* and 2876*m* (CH), 1119*s* and 1088*s* (C-OH), 742*m* and 695*m*.

Synthesis of thioctic-polyglycerolamine, TA-PG-NH₂

In a two-necked flask equipped with magnetic stirring, PG-NH₂ (0.300 g, 0.40 mmol -NH₂) was dissolved in DMF (10 mL) and cooled to 0 °C. Then, DMAP (1 mg, 8 μmol dissolved in 1 mL of DMF), thioctic acid (6.8 mg dissolved in 3 mL of DMF, 33 μmol, 0.08 eq.) and DCC (6.8 mg, 33 μmol, 0.08 eq.) were added. After stirring the solution for 1 h at 0 °C, it was allowed to reach room temperature and was stirred for 59 h. The resulting yellow solution was concentrated in vacuum and then dialyzed in MeOH to afford 0.295 g of TA-PG-NH₂ as a pale yellow product. **¹H NMR** (400 MHz, MeOD, δ): 4.55-3.40 (m, CH₂ and CH, PG scaffold), 3.16-2.60 (m, OCH-NH₂ and CH₂-NH₂), 2.55-1.15 (m, overlaid signals

corresponding to AT and CH₂ core), 0.92 (m, CH₃ core). **IR** (KBr) $\nu_{\max} = 3364s$ (OH), 2918*m* and 2876*m* (CH), 1651*w* and 1570*w*, 1103*s* and 1076*s* (C-OH).

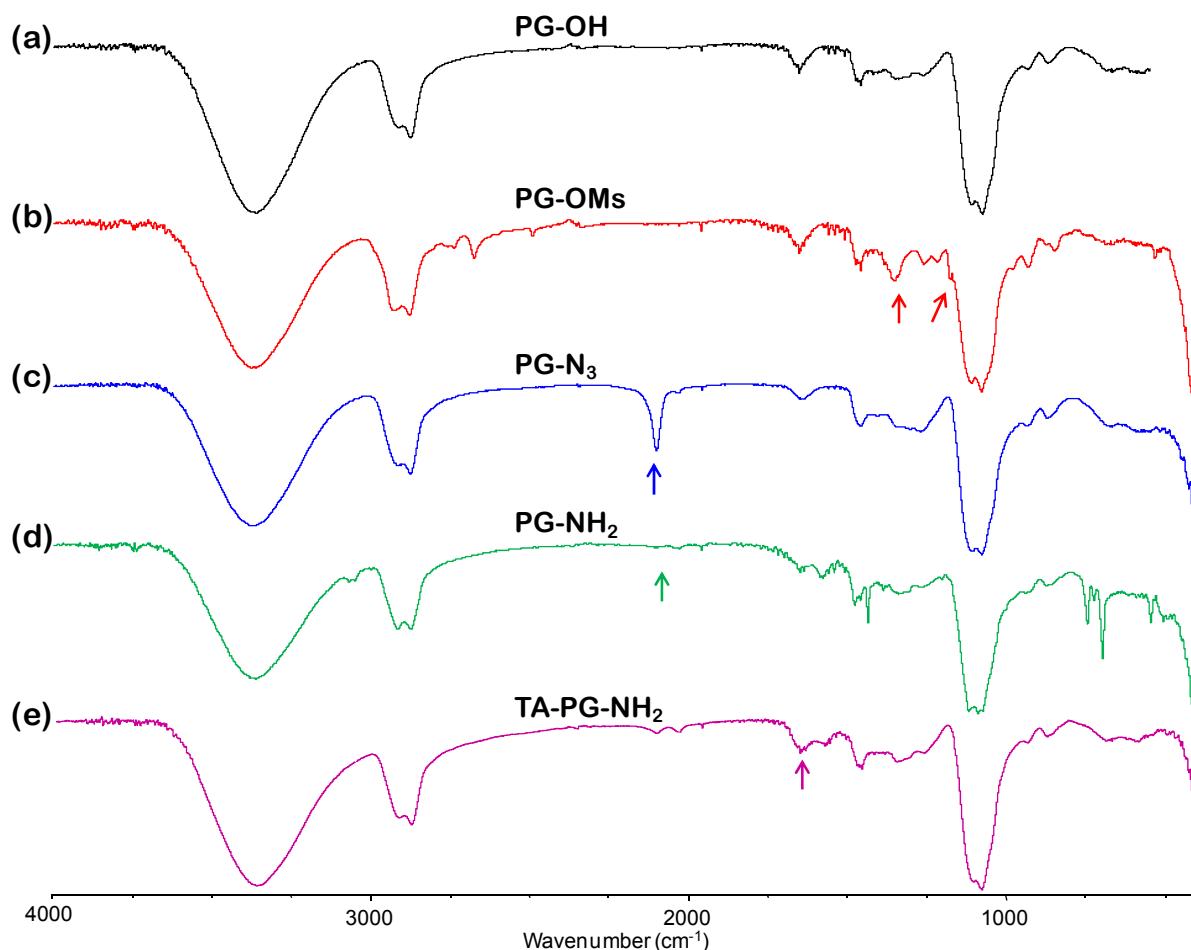


Figure S1. IR spectra of: (a) starting PG, (b) mesylpolyglycerol, (c) polyglycerolazide, (d) polyglycerolamine and (e) thioctic-polyglycerolamine, corresponding to a 9% of conversion of -OH groups into -NH₂ groups. The colored arrows indicate the appearance or disappearance of relevant signals.

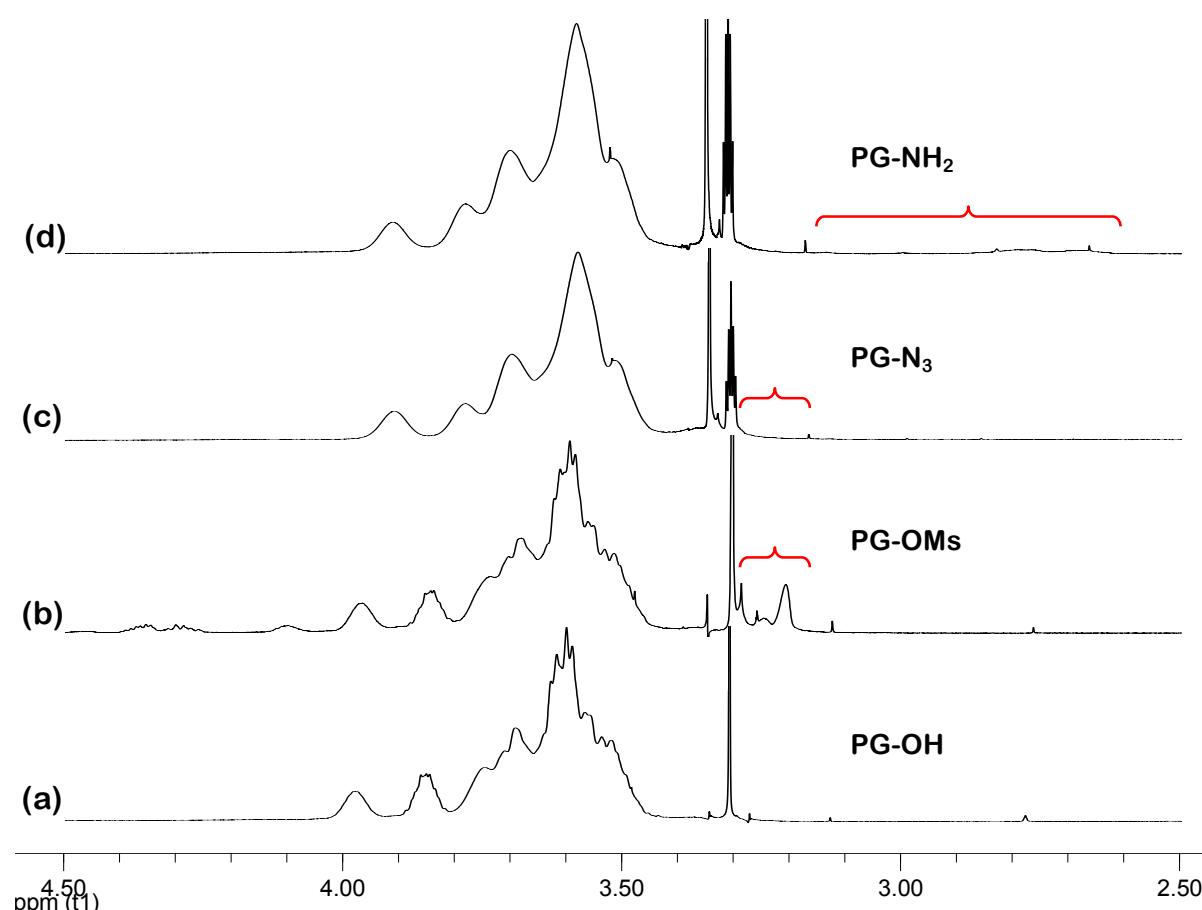


Figure S2. ^1H NMR spectra of: (a) starting PG, (b) mesylpolyglycerol, (c) polyglycerolazide, and (d) polyglycerolamine, corresponding to a 9% of conversion of -OH groups into -NH₂ groups. The colored brackets indicate the appearance/disappearance of relevant signals.

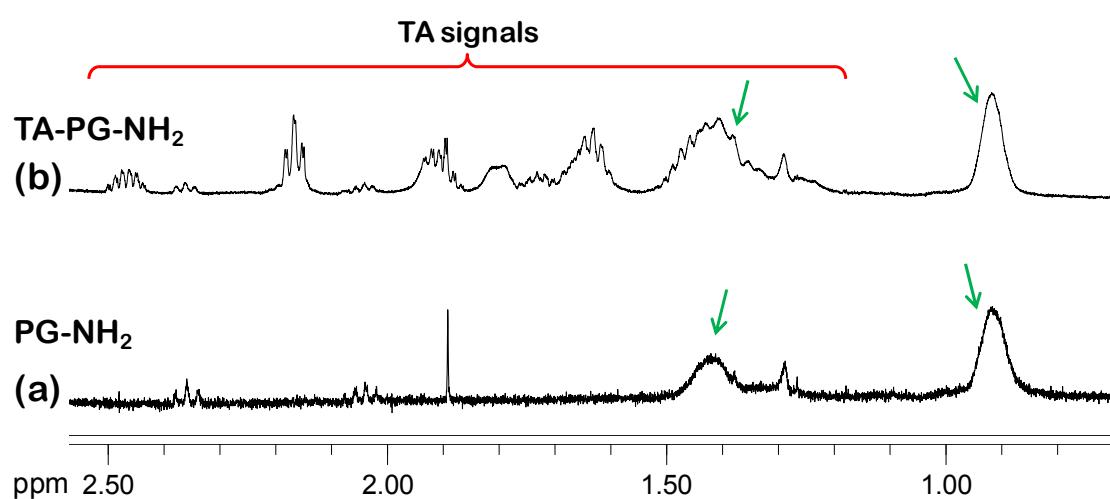


Figure S3. ^1H NMR spectra in the 2.6-0.7 ppm region of: (a) polyglycerolamine and (b) thioctic-polyglycerolamine corresponding to a 9% of conversion of -OH groups into -NH₂ groups. The relevant signals are indicated: PG core (TMP, green arrows) and signals corresponding to 10H of the incorporated TA (red bracket).

AFM characterization of Au/Ta-PG-NH₂

Experimental part

The AFM measurements were performed using a Nanoscope Multimode IIIA (Veeco, now Bruker AXS, Mannheim). Measurements were conducted in laboratory air at room temperature. The microscope was operated in the tapping mode (TM-AFM) using silicone probes NST-NCLF (Nascatec, Stuttgart) with a size of $250 \pm 10 \mu\text{m}$ and a tip radius of $7 \pm 1 \text{ nm}$ at resonance frequencies of 165 kHz under ambient conditions. The force constant was 30 N/m. The cantilever was forced to oscillate near its resonance frequency.

Pretreatment of the gold surface and functionalization by self-assembly

Before each experiment, the 12x12x0.7 mm gold substrate on borosilicate glass (Arrandee, Germany) was annealed (hydrogen flame length *ca.* 2-4 cm) by sweeping the substrate back and forth for about 30-60 s, keeping the small flame spot on the film a dim orange color. After annealing, the surface was cooled down under a stream of N₂. Then, it was functionalized by immersing the substrate overnight into a 1 mM solution of the corresponding PG derivative in MeOH. The modified gold surface was removed and rinsed copiously with MeOH for 1 min, and then with EtOH for another min before drying with N₂. The functionalized surface was used immediately for AFM measurements.

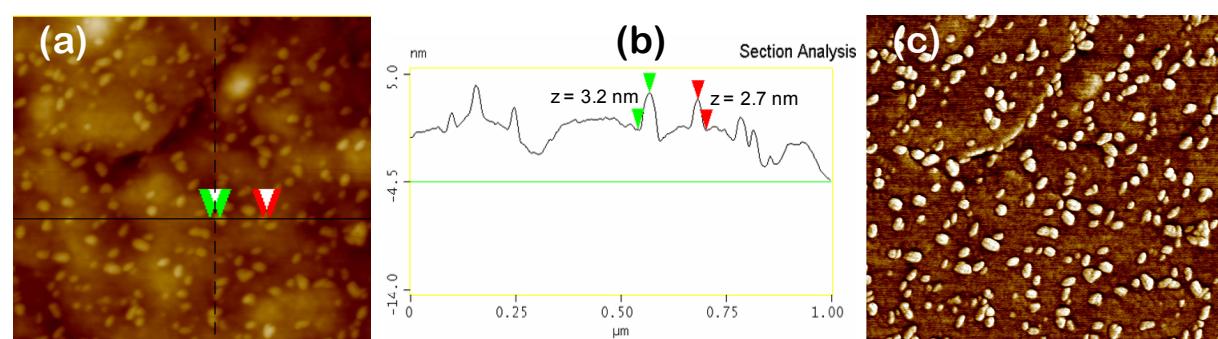


Figure S4. AFM measurements of Au/Ta-PG-NH₂ 9 %: (a) 1000 nm x 1000 nm topographical image; (b) cross-section profiles corresponding to the black solid line in (a); (c) 1000 nm x 1000 nm phase image showing an incomplete monolayer.