(2Z,7Z)-Cycloocta-2,7-dienone 4



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(R,Z)-7-Methylcyclooct-2-enone 5





(3R,7S)-Methyl 8-hydroxy-3,7-dimethyloctanoate 3



## (3*R*,7*S*)-Methyl 8-((*tert*-butyldiphenylsilyl)oxy)-3,7-dimethyloctanoate 7

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(3R,7S)-8-((tert-Butyldiphenylsilyl)oxy)-3,7-dimethyloctan-1-ol 8



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5-(((3*R*,7*S*)-8-((*tert*-Butyldiphenylsilyl)oxy)-3,7-dimethyloctyl)sulfonyl)-1-phenyl-1H-tetrazole 9



(3R,7S)-Methyl 3,7-dimethyl-8-oxooctanoate 10



(3*R*,7*S*,11*R*,15*S*)-Methyl 16-((*tert*-butyldiphenylsilyl)oxy)-3,7,11,15-tetramethylhexadec-8-enoate 11



(3R,7S,11R,15S)-16-((tert-Butyldiphenylsilyl)oxy)-3,7,11,15-tetramethylhexadec-8-en-1-ol 12



(3R,7R,11S,15S)-16-((tert-Butyldiphenylsilyl)oxy)-3,7,11,15-tetramethylhexadecan-1-ol 13



(4*S*,9*R*,13*R*,17*S*,21*S*)-9,13,17,21,25,25-Hexamethyl-1,24,24-triphenyl-2,6,23-trioxa-24-silahexacosan-4-ol 14



tert-Butyl(((2S,6S,10R,14R)-16-iodo-2,6,10,14-tetramethylhexadecyl)oxy)diphenylsilane 15



1-O-Benzyl-2,3-bis-O-[(3*R*,7*R*,11*S*,15*S*)-16-((*tert*-butyldiphenylsilyl)oxy)-3,7,11,15-tetramethylhexadecanyl]-*sn*-glycerol 16



1-O-Benzyl-2,3-bis-O-[(3R,7R,11S,15S)-16-hydroxy-3,7,11,15-tetramethylhexadecanyl]-sn-glycerol 17



1-O-Benzyl-2,3-bis-O-[(3R,7R,11S,15S)-16-formyl-3,7,11,15-tetramethylhexadecanyl]-sn-glycerol 18



1-O-Benzyl-2,3-bis-O-[(3R,7R,11S,15S)-3,7,11,15-tetramethylheptadec-16-enyl]-sn-glycerol 19





(2*S*,7*R*,11*R*,15*S*,19*S*,22*S*,26*S*,30*R*,34*R*,*E*)-2-((Benzyloxy)methyl)-7,11,15,19,22,26,30,34-octamethyl-1,4-dioxacyclohexatriacont-20-ene 20



(2*S*,7*R*,11*R*,15*S*,19*S*,22*S*,26*S*,30*R*,34*R*)-2-((Benzyloxy)methyl)-7,11,15,19,22,26,30,34-octamethyl-1,4-dioxacyclohexatriacontane 21



((2*R*,7*R*,11*R*,15*S*,19*S*,22*S*,26*S*,30*R*,34*R*)-7,11,15,19,22,26,30,34-Octamethyl-1,4-dioxacyclohexatriacontan-2-yl)methanol 2





Carbon NMR after Pt reduction of the double bond but still with the benzyl group on:

## (2*R*,3*R*,4*S*,5*R*,6*R*)-2-(((7*R*,11*R*,15*S*,19*S*,22*S*,26*S*,30*R*,34*R*)-7,11,15,19,22,26,30,34-Octamethyl-1,4-dioxacyclohexatriacontan-2-yl)methoxy)-6-((pivaloyloxy)methyl)tetrahydro-2H-pyran-3,4,5-triyl tris(2,2-dimethylpropanoate) 22







## Sample description and analysis for the comparison with natural 2 and 23

The sample was collected from the Rainbow hydrothermal vent field (RHF) located on the Mid-Atlantic Ridge. RHF has been characterized as an ultra-mafic hydrothermal system with fluids that are known to reach up to  $360^{\circ}$ C and contain abundant H<sub>2</sub>, CH<sub>4</sub> and CO<sub>2</sub>.<sup>1</sup> Samples were collected during a sampling campaign in 2008 using the ROV Jason.

The sample analysed is composed of material from the interior of a vent chimney collected at a depth of 2293 m below sea level. The location of the vent is 57°39'N, 32°65'W.

Intact Polar Lipids were extracted from freeze-dried, powdered (approximately 5 g) vent deposit using a modified Bligh and Dyer method described by Pitcher *et al.*<sup>2</sup> The extract was analysed by high performance liquid chromatography electrospray ionisation mass spectrometry (HPLC-ESI-MS<sup>n</sup>) using modified literature methods.<sup>3</sup> The analysis was conducted on an Agilent 1200 series LC equipped with a Lichrosphere diol column (250 x 2.1  $\mu$ m, 5  $\mu$ m particles; Alltech Associates Inc.) coupled to a Thermo LTQ XL linear ion trap with Ion Max source with electrospray ionization (ESI) probe (Thermo Scientific, Waltham, MA).

The extract was acid hydrolysed to release **2**.<sup>4</sup> The hydrolyzed extract was reacted with diazomethane to convert free acids to methyl esters then separated over a short  $Al_2O_3$  column using hexane/DCM (9:1; v/v) and DCM/methanol (1:1; v/v). Part of the latter fraction was subsequently silylated using bis(trimethylsilyl)trifluoroacetamide (BSTFA) in pyridine at 60°C for 15 min. The fatty acid methyl ester and silylated alcohol fraction were analyzed by gas chromatography (GC) and GC-mass spectrometry (MS). GC was performed on a Hewlett-Packard 6890 series GC system with an on-column injector and flame ionization detector. A fused silica capillary column (length 25 m, internal diameter 0.32 µm) coated with CP Sil-5 (film thickness, df = 0.12 µm) was used with helium as the carrier gas. Samples were injected at 70 °C and oven temperature was increased to 130 °C at 20°C/min and then increased to 320°C at a rate of 4°C/min, where it remained stable for 10 min. GC-MS was performed on a Thermo Trace DSQ mass spectrometer fitted with a Thermo Trace GC Ultra. The mass spectrometer was operated at 70 eV with a mass range of m/z 50 – 800 and a cycle time of 0.33 s. The GC was equipped with a fused silica capillary column as described for GC analysis above and the same temperature program was used.



Gas chromatogram of synthetic  $\mathbf{2}$ ,  $R_t = 48.54$  min



Gas chromatogram of TMSi-derivatized acid hydrolysed extract of the hydrothermal vent sample,. First peak corresponds to archaeol (1), second peak corresponds to 2 with  $R_t = 48.56$  min.



Co-injection of the vent sample with synthetic  $\mathbf{2}$ , showing the same  $R_t = 48.56$  min



HPLC/MS selected ion chromatograms of mass m/z 830.5, the  $NH_4^+$  ion of the molecular mass of **23.** *Top* chromatogram shows synthetic **23** the *lower* chromatogram shows the hydrothermal vent sample at Rainbow hydrothermal field with natural **23**, both eluting at 10.2 min.

<sup>&</sup>lt;sup>1</sup>C. Konn, J. L. Charlou, J. P. Donval, N. G. Holm, F. Dehairs, and S. Bouillon, *Chemical Geology*, 2009, 258, 299 – <sup>2</sup> A. Pitcher, E.C. Hopmans, S. Schouten, and J.S. Sinninghe Damsté, *Organic Geochemistry* 2009, 40, 12-19.
 <sup>3</sup> H. F. Sturt, R. E. Summons, K. Smith, M. Elvert, and K.-H. Hinrichs, *Rapid Comm in Mass Spectr.*, 2004, 18, 617 –

<sup>&</sup>lt;sup>628.</sup>
<sup>4</sup> J. S. Sinninghe Damsté, W. I. C. Rijpstra, E. C. Hopmans, J. W. H. Weijers, B. U. Foesel, J. Overmann, and S. N. Dedysh, *Appl. Environm. Microbiol.*, 2011, **77**, 4147 - 4154.