

Activated Phosphonated Trifunctional Chelates for Highly Sensitive Lanthanide-based FRET Immunoassays applied to Total Prostate Specific Antigen Detection

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Supplementary Information (8 pages including this one)

Figure S1. ^1H -NMR spectrum of \mathbf{L}^* (25°C , 200 MHz, D_2O)

Figure S2. UV-Vis absorption spectrum of \mathbf{L}^* labelled streptavidin in TRIS/HCl 0.01 M, pH 7.4 buffer.

Figure S3. Excitation spectrum ($\lambda_{\text{em}} = 545$ nm) of TbL^* labelled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer.

Figure S4. Time-resolved (delay = 10 μs) emission spectrum of the TbL^* labelled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer ($\lambda_{\text{exc}} = 330$ nm).

Figure S5. Tb based emission decay profile of the TbL^* labelled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer ($\lambda_{\text{exc}} = 330$ nm and $\lambda_{\text{em}} = 545$ nm).

Figure S6. MALDI-TOF mass spectra of AB1 (A) and AB1 labelled with \mathbf{L}^* (B). A shift of *ca* 2500 Da is observed for labelled antibody in the expanded region.

Figure S7. UV-Vis absorption spectra of $\mathbf{L}3$ (blue) and TbL3 (violet) and luminescence spectra of TbL3 (green, $\lambda_{\text{exc}} = 328$ nm) in TRIS/HCl 0.01M at pH 7.4

Scheme S1. Full ^1H -NMR assignment for $\mathbf{L}3$ and \mathbf{L}^* .

PL decay time analysis

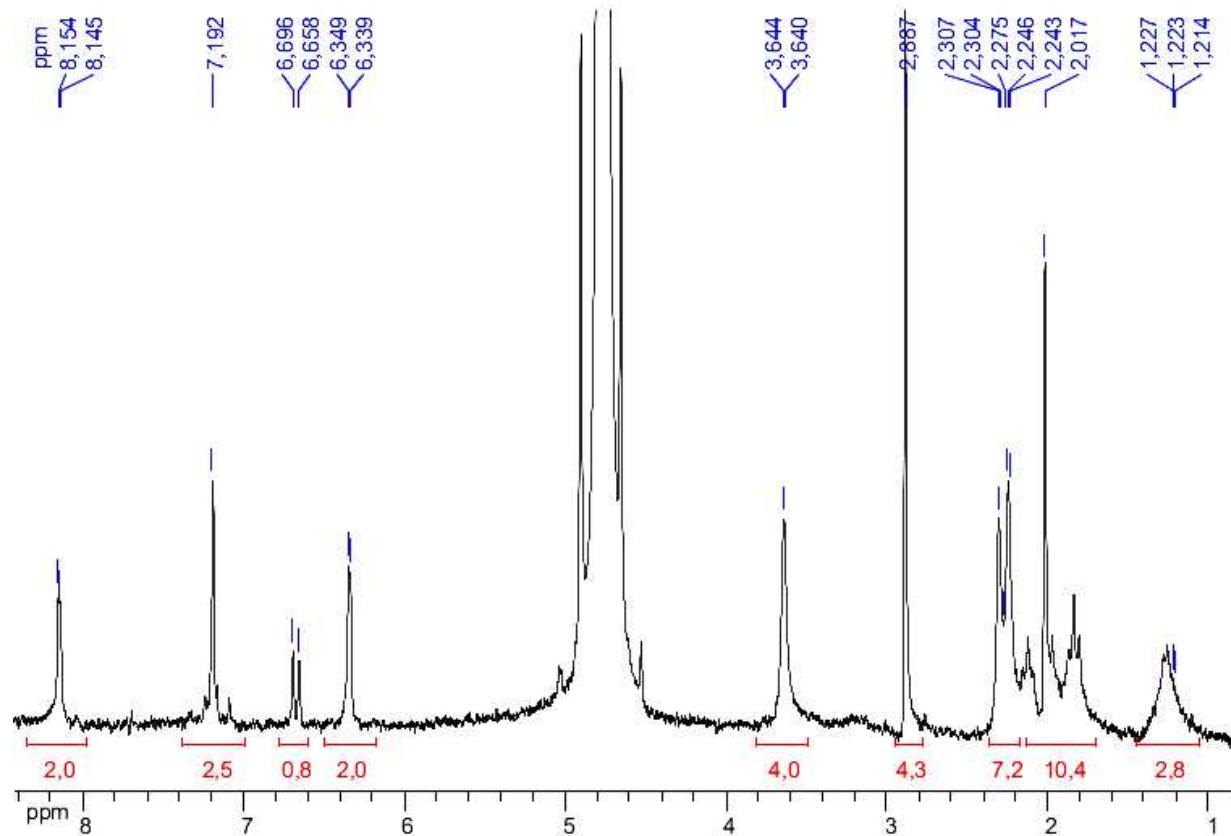


Figure S1. ¹H-NMR spectrum of L* (25°C, 200 MHz, D₂O)

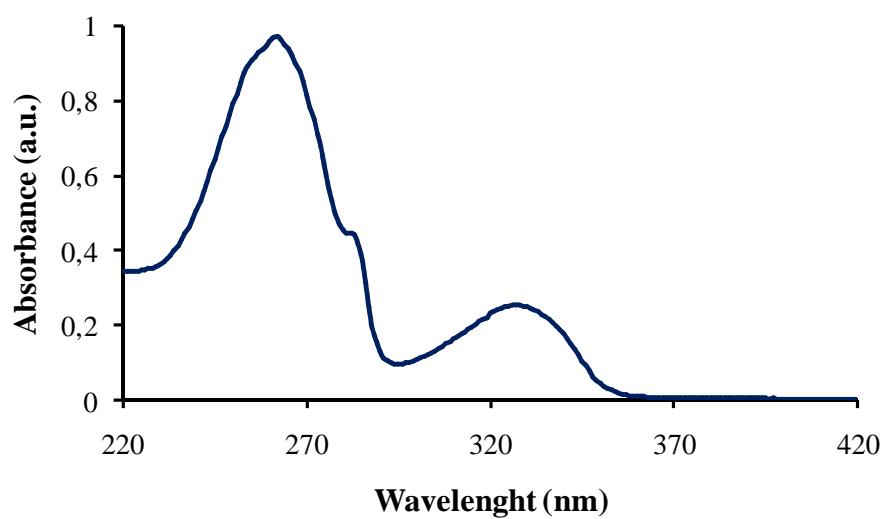


Figure S2. UV-Vis absorption spectrum of TbL* labelled streptavidin in TRIS/HCl 0.01 M, pH 7.4 buffer.

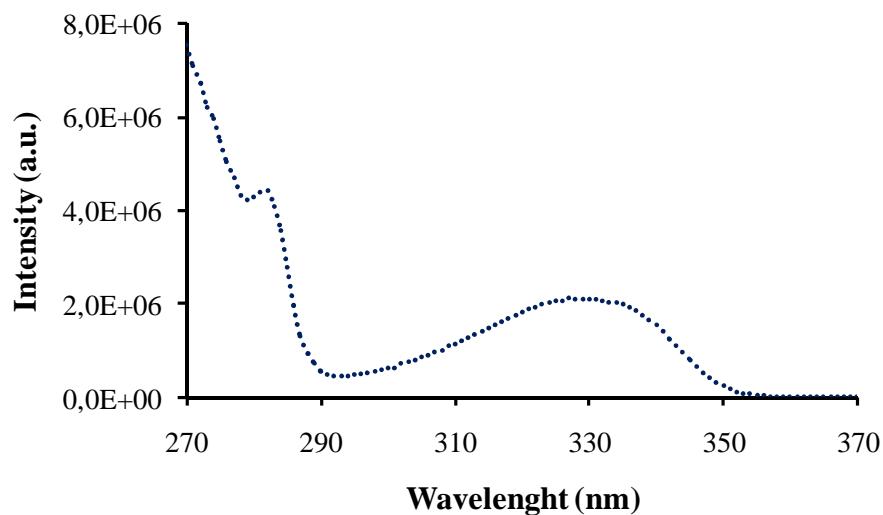


Figure S3. Excitation spectrum ($\lambda_{\text{em}} = 545 \text{ nm}$) of TbL* labelled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer.

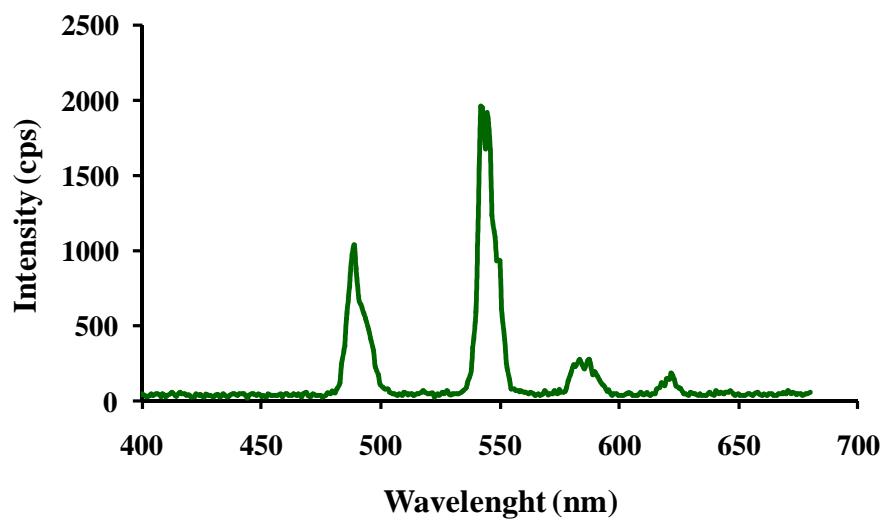


Figure S4. Time-resolved (delay = 10 μs) emission spectrum of the TbL* labeled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer.

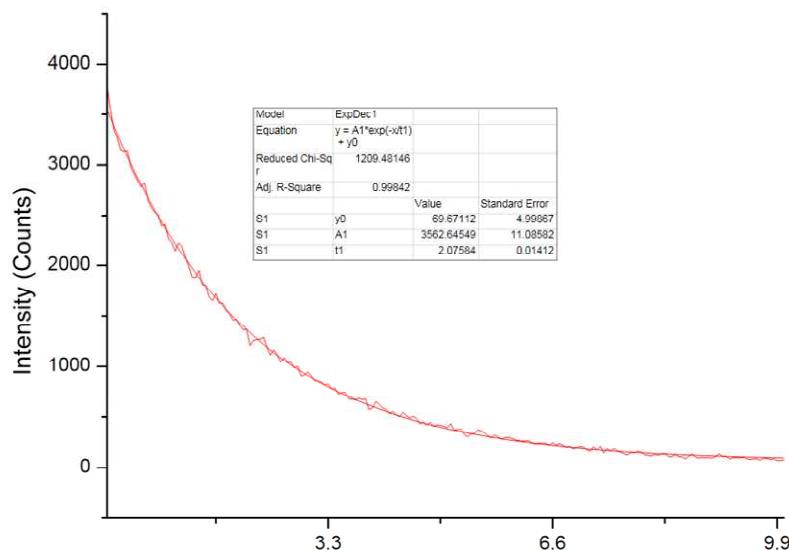
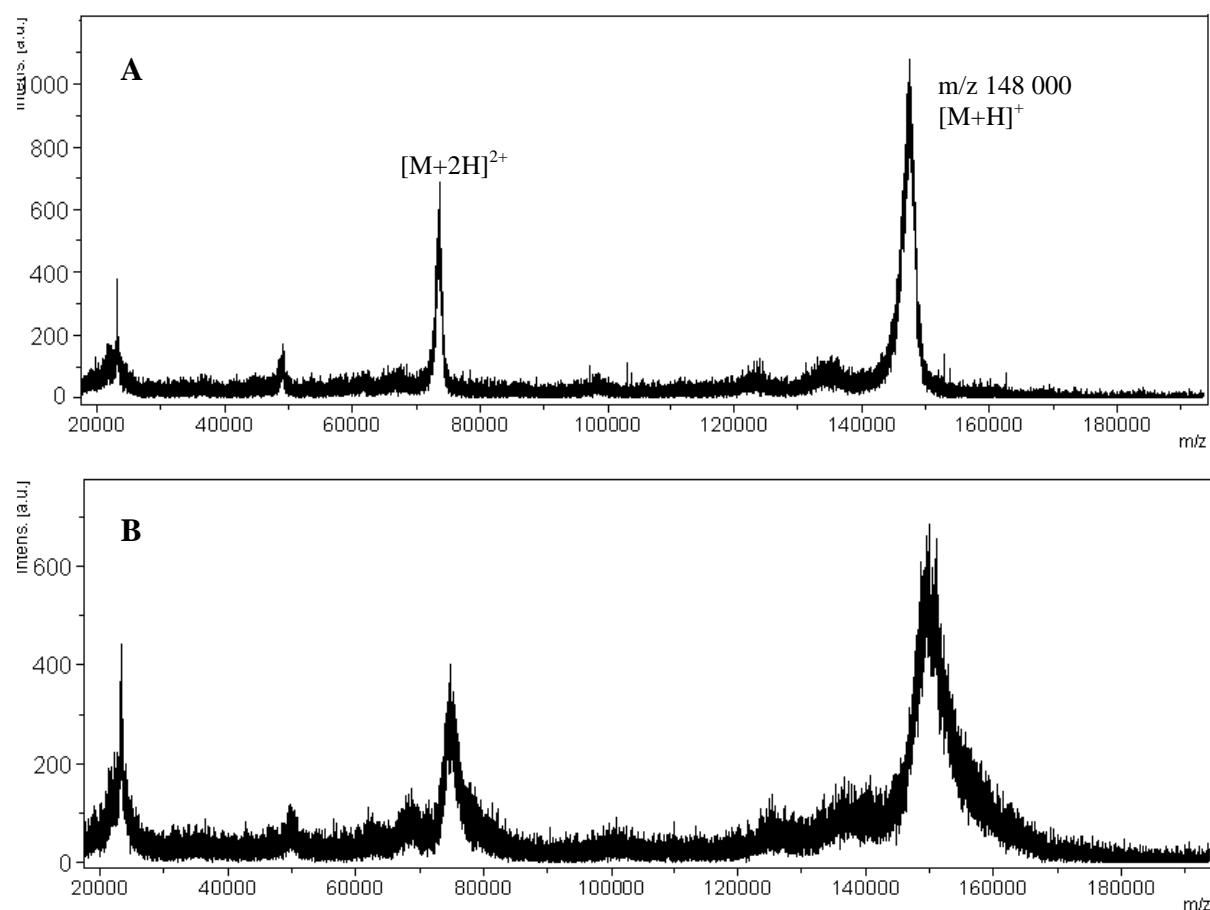


Figure S5. Tb based emission decay profile of the TbL* labelled streptavidine in TRIS/HCl 0.01 M, pH 7.4 buffer ($\lambda_{\text{exc}} = 330$ nm and $\lambda_{\text{em}} = 545$ nm).



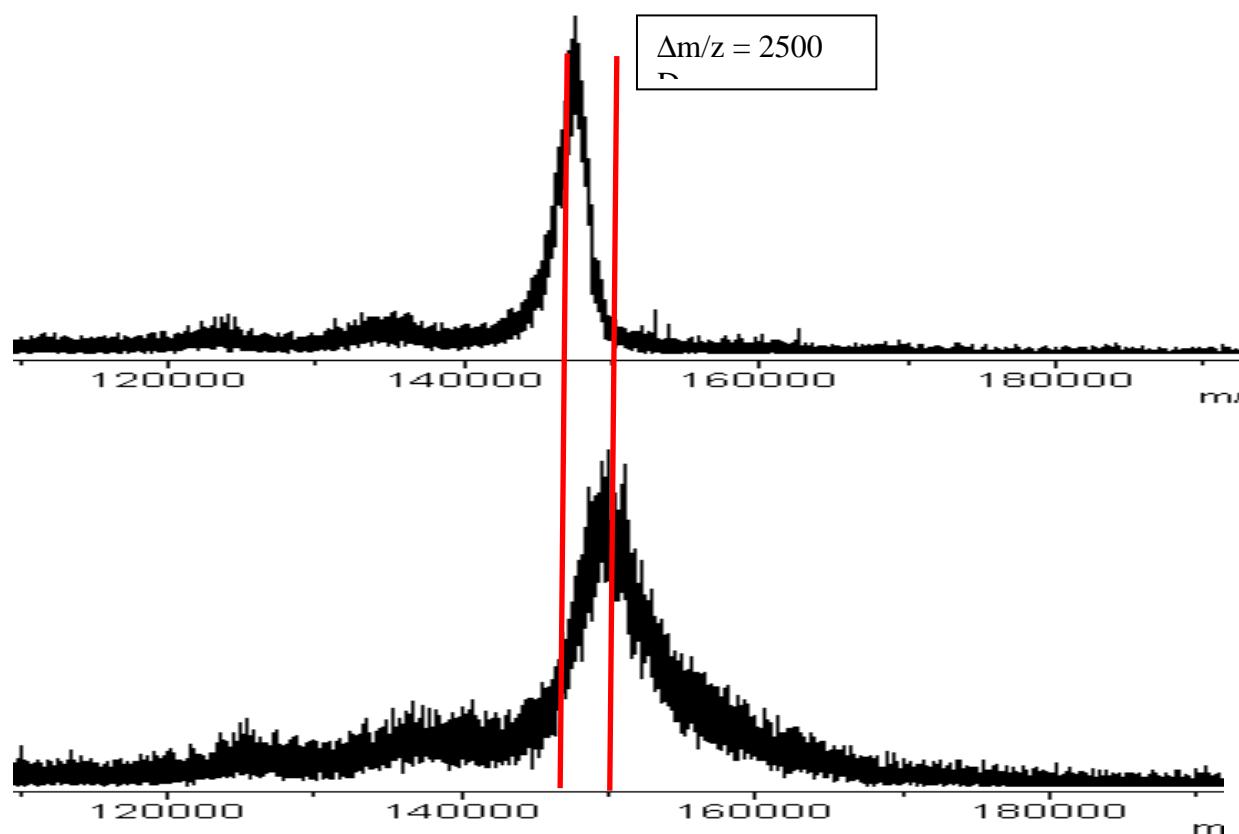


Figure S6. MALDI-TOF mass spectra of AB1 (A) and AB1 labelled with L* (B). A shift of *ca* 2500 Da is observed for labelled antibody in the expanded region.

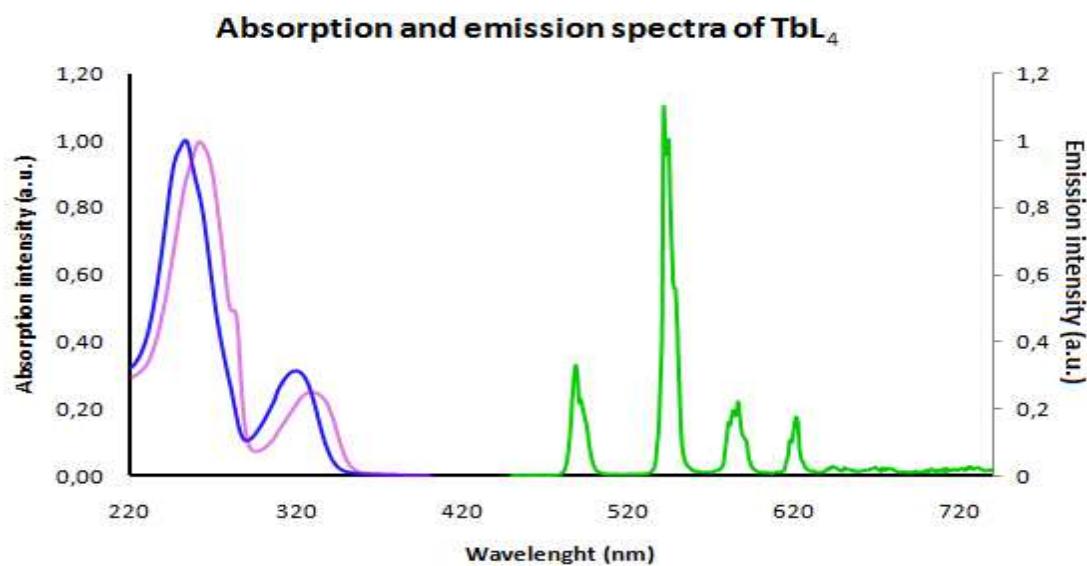
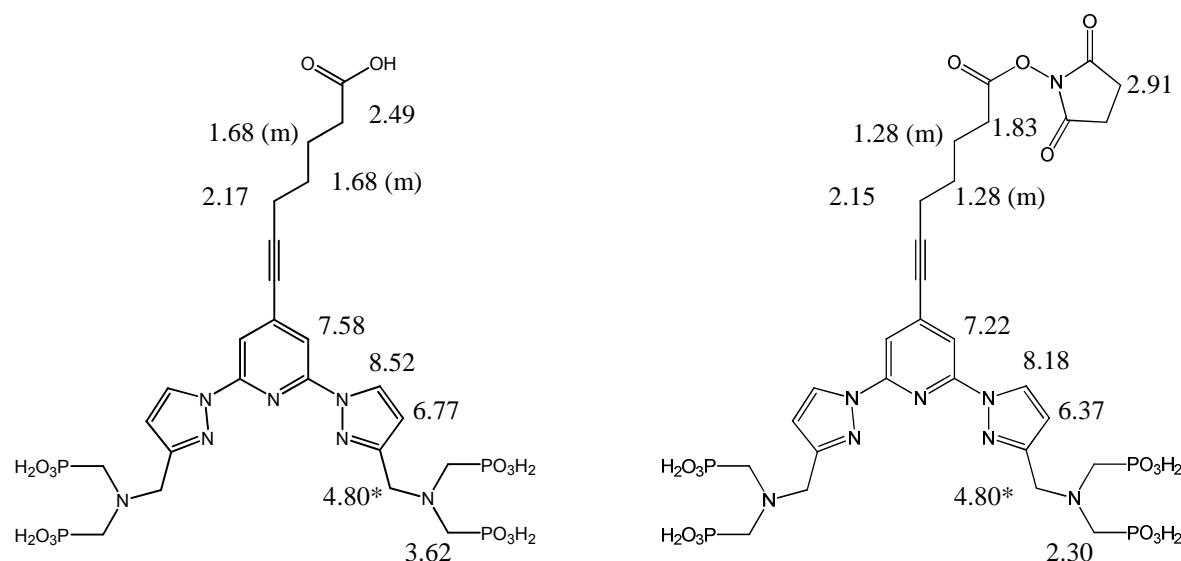


Figure S7. UV-Vis absorption spectra of L3 (blue) and TbL3 (violet) and luminescence spectra of TbL3 (green, $\lambda_{\text{exc}} = 328$ nm) in TRIS/HCl 0.01M at pH 7.4



Scheme S1. Full ^1H -NMR assignment for **L3** and **L***. The asterisks denote the fact that the signals can be hidden by the peak of the water, m = multiplet.

PL decay time analysis. The decay time analysis in the donor and acceptor channel was performed as described in reference 41 (from the manuscript). All decay curves were fit using a multi-exponential PL intensity decay function:

$$I = \sum A_i \exp(-t/\tau_i) = A \sum \alpha_i \exp(-t/\tau_i) \quad (1)$$

where A is the total amplitude and α_i are the amplitude fractions ($\sum \alpha_i = 1$). All PL lifetime averaging for the dynamic FRET quenching process was performed using amplitude weighted average lifetimes:

$$\langle \tau \rangle = \sum \alpha_i \tau_i \quad (2)$$

First the decay curve of the pure Tb donor was fit using a double-exponential decay function, which led to the amplitude fractions α_{D1} and α_{D2} , the PL decay times τ_{D1} and τ_{D2} (with $\tau_{D2} > \tau_{D1}$) and the average PL decay time of the pure donor (in the absence of the acceptor) $\langle \tau_D \rangle$. The FRET-quenched decay curves in the donor detection channel were fit using a triple-exponential decay function, leading to the amplitude fractions α_{DA*1} , α_{DA*2} and α_{DA*3} and the PL decay times τ_{DA1} , τ_{DA2} and τ_{DA3} , for which the third decay time component was fixed to $\tau_{DA3} = \tau_{D2}$ in order to take into account the emission of unquenched donors. For the calculation of the average donor decay time in the presence of the acceptor $\langle \tau_{DA} \rangle$ only the first two amplitudes and decay times were used (as the third component represents unquenched donors). Therefore the amplitude fractions must be redefined for these two decay times τ_{DA1} and τ_{DA2} :

$$\alpha_{DA1} = \frac{\alpha_{DA*1}}{\alpha_{DA*1} + \alpha_{DA*2}} \quad \text{and} \quad \alpha_{DA2} = \frac{\alpha_{DA*2}}{\alpha_{DA*1} + \alpha_{DA*2}} \quad (3)$$

As the unquenched donor possesses two decay time components (τ_{D1} and τ_{D2}), $\langle \tau_{DA} \rangle$ must be corrected for the shorter time component (τ_{D1}) by a factor z_D (the fraction of unquenched

donors in the short time components), which is determined by comparing the amplitude fractions of τ_{D2} and τ_{DA3} ($\tau_{DA3} = \tau_{D2}$) multiplied by the amplitude fraction α_{D1} :

$$z_D = \alpha_{D1}(\alpha_{DA*3}/\alpha_{D2}) \quad (4)$$

The average FRET-quenched decay time is then:

$$\langle \tau_{DA} \rangle = \frac{\alpha_{DA1}\tau_{DA1} + \alpha_{DA2}\tau_{DA2} - z_D\tau_{D1}}{1-z_D} \quad (5)$$

and the average FRET-efficiency is:

$$\langle \eta_{FRET} \rangle = 1 - \frac{\langle \tau_{DA} \rangle}{\langle \tau_D \rangle} \quad (6)$$

The FRET-sensitized decay curves in the acceptor detection channel were fit using a quadruple-exponential decay function, leading to the amplitude fractions α_{AD*0} , α_{AD*1} , α_{AD*2} and α_{AD*3} and the PL decay times τ_{AD0} , τ_{AD1} , τ_{AD2} and τ_{AD3} , for which the fourth decay time component was fixed to $\tau_{AD3} = \tau_{D2}$ in order to take into account the emission of unquenched donors, which is much less intense compared to the donor channel but still present due to spectral crosstalk of the Tb emission in the dye acceptor detection channel. The correction factor z_A (the fraction of unquenched donors in the short time components) is almost negligible but is still taken into account for a correct treatment:

$$z_A = \alpha_{D1}(\alpha_{AD*3}/\alpha_{D2}) \quad (7)$$

In order to calculate the average FRET decay time $\langle \tau_{AD} \rangle$ only the amplitudes and lifetimes with $i = 0$ to 2 are taken into account ($i = 3$ represents the unquenched donor emission). Moreover, the amplitudes α_{AD*i} must be corrected by the FRET rates $k_i = 1/\tau_{ADi} - 1/\langle \tau_D \rangle$, to take into account the dependence of the excitation of the acceptors (and therefore the amplitude fractions) on the different FRET efficiencies for the different distances (corresponding to the decay times τ_{ADi}). The corrected amplitude fractions are (for $i = 0$ to 2):

$$\alpha_{\text{ADI}} = \frac{\alpha_{\text{Ai}} / k_i}{\sum \alpha_{\text{Ai}} / k_i} \quad (8)$$

The average FRET decay time is then calculated by:

$$\langle \tau_{\text{AD}} \rangle = \frac{\alpha_{\text{AD1}} \tau_{\text{AD1}} + \alpha_{\text{AD2}} \tau_{\text{AD2}} + \alpha_{\text{AD3}} \tau_{\text{AD3}} - z_A \tau_{\text{D1}}}{1 - z_A} \quad (9)$$

and the average FRET-efficiency is:

$$\langle \eta_{\text{FRET}} \rangle = 1 - \frac{\langle \tau_{\text{AD}} \rangle}{\langle \tau_D \rangle} \quad (10)$$

For each FRET decay time the donor-acceptor distance r_x can be calculated by:

$$r_x = \left(\frac{\tau_x R_0^6}{\tau_D - \tau_x} \right)^{1/6} \quad (11)$$

where τ_x represents the different lifetimes τ_{DAi} , τ_{ADI} , $\langle \tau_{\text{DA}} \rangle$ or $\langle \tau_{\text{AD}} \rangle$. The fractions of FRET-pairs found at the different distances corresponding to τ_{DAi} and τ_{ADI} are given by the amplitude fractions of these decay times.

Complete decay time fit data and resulting FRET data an distances

Donor Channel		R_d (nm)		Decay time fit results and calculated FRET parameters for AF647 as acceptor																			
D or DA	TPSA	τ_1	A_1	α_{d0^+1}	α_{d0^+}	τ_2	A_2	α_{d0^+2}	α_{d0^+}	$\langle \tau_{d0} \rangle$	τ_3	A_3	α_{d0^+3}	$\langle \tau_{d0} \rangle$	$z(D)$	$\langle \tau_{dA} \rangle$	η_{FRET}	fixed					
Channel 25: 399 nm fixed decay background																							
D or DA		τ_{TPA} [fm]	τ_0	A_0	α_{d0^+0}	k_{FRET0}	α_{d0^+0}	τ_1	A_1	α_{d0^+1}	$\langle \tau_{d0} \rangle$	τ_2	A_2	α_{d0^+2}	$\langle \tau_{d0} \rangle$	τ_3	A_3	α_{d0^+3}	$\langle \tau_{d0} \rangle$	fixed			
D		2.90	289	15	0.65	0.37	541	568	268	0.09	255	2679	0.91	2738					0.000	0.119	0.38		
DA		4.80	263	36	0.13	0.44	924	421	0.16	0.63	427	250	2382	0.85	2567	0.038	436	0.84	0.006	0.020	0.37		
DA		6.00	278	50	0.19	0.45	107	566	0.22	0.53	748	250	1516	0.59	2039	0.026	778	0.73	0.027	0.120	0.35		
DA		8.00	251	648	0.25	0.47	1107	726	0.28	0.53	704	250	2242	0.47	1666	0.025	711	0.71	0.027	0.120	0.35		
DA		10.00	263	738	0.29	0.48	1165	797	0.31	0.52	731	250	1004	0.48	1008	0.024	738	0.73	0.027	0.120	0.35		
DA		12.00	253	688	0.28	0.48	1115	736	0.30	0.52	704	250	1054	0.48	1056	0.024	704	0.74	0.027	0.120	0.35		
DA		14.00	265	663	0.26	0.48	1096	705	0.28	0.52	693	250	1142	0.46	1720	0.025	699	0.74	0.027	0.120	0.35		
DA		16.00	255	657	0.26	0.47	1056	709	0.28	0.53	671	250	1220	0.46	1725	0.025	705	0.74	0.027	0.120	0.35		
DA		20.00	236	552	0.22	0.46	1021	633	0.27	0.55	660	250	1299	0.53	1835	0.025	665	0.76	0.028	0.147	0.32		
DA		25.00	259	557	0.22	0.46	1022	634	0.25	0.54	665	250	1386	0.53	1832	0.025	670	0.76	0.028	0.147	0.32		
distance r		255	635	0.25	0.47	1094	207	0.28	0.53	697	250	1204	0.47	1762	0.025	703	0.74	0.027	0.120	0.35			
Accepter Channel		255	635	0.25	0.47	1094	207	0.28	0.53	697	250	1204	0.47	1762	0.025	703	0.74	0.027	0.120	0.35			
distance r		2.8																	4.4				
Accepter Channel		2.8																	5.0				
distance r		2.8																	5.0				
Donor Channel		R_d (nm)		Decay time fit results and calculated FRET parameters for XL665 as acceptor																			
Donor Channel																							
Channel 25: 399 nm fixed decay background																							
D or DA	TPSA	τ_1	A_1	α_{d0^+1}	α_{d0^+}	τ_2	A_2	α_{d0^+2}	α_{d0^+}	$\langle \tau_{d0} \rangle$	τ_3	A_3	α_{d0^+3}	$\langle \tau_{d0} \rangle$	$z(A)$	$\langle \tau_{dA} \rangle$	η_{FRET}	fixed					
D or DA		τ_{TPA} [fm]	τ_0	A_0	α_{d0^+0}	k_{FRET0}	α_{d0^+0}	τ_1	A_1	α_{d0^+1}	$\langle \tau_{d0} \rangle$	τ_2	A_2	α_{d0^+2}	$\langle \tau_{d0} \rangle$	τ_3	A_3	α_{d0^+3}	$\langle \tau_{d0} \rangle$	fixed			
D		2.80	200	49	0.36	0.022	0.058	232	274	0.37	0.0394	0.30	858	118	0.16	0.0080	0.64	2950	75	0.103	0.033		
D		2.78	4.00	57	0.57	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	622	2950	75	0.103	0.033	
D		2.78	6.00	54	0.57	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	603	2950	74	0.102	0.032	
D		2.78	8.00	56	1264	0.40	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	593	2950	74	0.102	0.032
D		2.78	10.00	60	1365	0.43	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	575	2950	74	0.102	0.032
D		2.78	12.00	59	1229	0.43	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	557	2950	74	0.102	0.032
D		2.78	14.00	59	1225	0.43	0.022	0.072	230	274	0.37	0.0394	0.30	858	233	0.16	0.0080	0.64	537	2950	74	0.102	0.032
D		2.78	16.00	59	1175	0.39	0.022	0.072	226	266	0.37	0.0394	0.30	851	245	0.15	0.0081	0.57	527	2950	75	0.102	0.032
D		2.78	18.00	59	1178	0.42	0.022	0.072	226	266	0.37	0.0394	0.30	954	270	0.15	0.0080	0.55	509	2950	61	0.022	0.022
D		2.78	20.00	61	1020	0.39	0.022	0.072	226	268	0.37	0.0394	0.30	857	305	0.15	0.0080	0.57	521	2950	61	0.022	0.022
D		2.78	22.00	61	1020	0.39	0.022	0.072	226	268	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	24.00	183	266	0.39	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	26.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	28.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	30.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	32.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	34.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	36.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	38.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	40.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	42.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	44.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	46.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	48.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	50.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	52.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	54.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	56.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	58.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	60.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15	0.0080	0.57	581	2950	67	0.022	0.022
D		2.78	62.00	183	266	0.43	0.022	0.072	226	274	0.37	0.0394	0.30	859	305	0.15							