

C₃-Symmetric Chiral Trisimidazoline-Catalyzed Friedel-Crafts (FC)-Type Reaction

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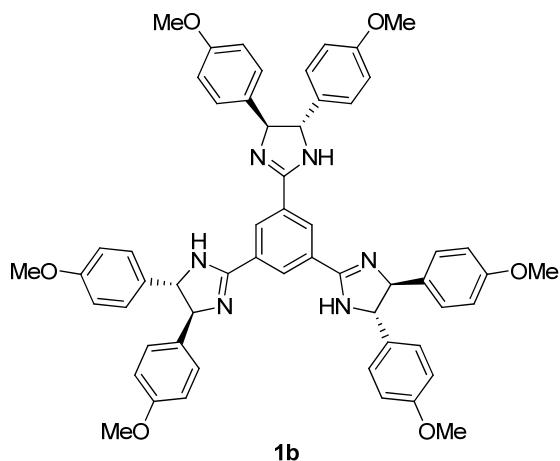
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General information

¹H-, ¹³C- and ³¹P-NMR spectra were recorded with JEOL JMN ECS400 FT NMR, JNM ECA600 FT NMR or Bruker AVANCE II (¹H-NMR 400, 500, 600 or 700 MHz, ¹³C-NMR 100, 126, 150 or 175 MHz). ¹H-NMR spectra are reported as follows: chemical shift in ppm relative to the chemical shift of CHCl₃ at 7.26 ppm, ACETONE-D₆ at 2.09 ppm or DMSO-D₆ at 2.49 ppm, integration, multiplicities (s = singlet, d = doublet, q = quartet, t = triplet, m = multiplet), and coupling constants (Hz). ¹³C-NMR spectra reported in ppm relative to the central line of triplet for CDCl₃ at 77 ppm, septet for ACETONE-D₆ at 30 ppm or septet for DMSO-D₆ at 40 ppm. FT-MS spectra were obtained with LTQ Orbitrap XL (Thermo Fisher Scientific). ESI-MS spectra were obtained with JMS-T100LC (JEOL). FAB-MS spectra were obtained with JMS-700 (JEOL). Optical rotations were measured with JASCO P-1030 polarimeter. HPLC analyses were performed on a JASCO HPLC system (JASCO PU 980 pump and UV-975 UV/Vis detector) using a mixture of hexane and 2-propanol as eluents. FT-IR spectra were recorded on a JASCO FT-IR system (FT/IR4100). Mp was measured with SHIMADZU DSC-60. Column chromatography on SiO₂ was performed with Kanto Silica Gel 60 (40-100 μm). Commercially available organic and inorganic compounds were used without further purification except for the solvent, which was distilled from sodium/benzophenone or CaH₂. *N*-4-Nosyl imines **2** were prepared following the reported procedures.¹ The products **4b**, **4d**, **4o** and **8** were identical in all respects with reported in the literature.²⁻⁶ Absolute configurations were assigned by comparison of optical rotation reported in the literature.²⁻⁶

Preparation of trisimidazoline **1b**

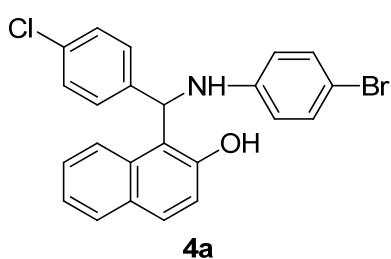


1,3,5-Triformylbenzene (35.6 mg, 0.219 mmol) and (1*S*,2*S*)-1,2-bis(4-methoxyphenyl)ethane-1,2-diamine (185 mg, 0.680 mmol) was dissolved in dioxane (6.8 mL) and stirred for 2 h at rt under N₂. The resulting solution was added NBS (121 mg, 0.679 mmol) at 0 °C and stirred for 24 h at rt. After the reaction was completed, sat. Na₂S₂O₅ aq. and 5% NaOH aq. was added to the reaction mixture and the solution was extracted with CH₂Cl₂. Organic layer was dried over Na₂SO₄, and evaporated in vacuo. After the purification *via* SiO₂ column chromatography

(hexane/AcOEt/TEA = 1/2/0.3), the desired product **1b** was obtained as pale yellow solid. Mp: >260 °C; [α]_D²⁴ = -96.9 (c 1.2, CHCl₃); ¹H-NMR (CDCl₃) δ: 8.61 (s, 3H), 7.16 (2H, d, *J* = 8.8 Hz), 6.82 (2H, d, *J* = 8.8 Hz), 4.80 (brs, 6H), 3.78 (s, 18H); ¹³C-NMR (CDCl₃) δ: 161.6, 159.0, 135.1, 130.6, 128.5, 127.7, 114.0, 55.2; IR (KBr): ν 3130, 3001, 2955, 2835, 1612, 1514, 1248, 1173, 1034 cm⁻¹; HRMS (FAB) calcd for C₅₇H₅₅N₆O₆ m/z = 919.4183 [(M+H⁺)], found m/z = 919.4175.

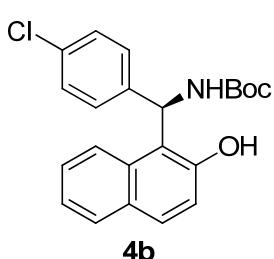
General procedure for enantioselective Friedel–Crafts (FC)-type reactions of aldimines (2) with 2-naphthols (3)

Under N₂ atmosphere, a test tube was charged with aldimine **2** (0.10 mmol), 2-naphthols **3** (0.15 mmol), and the catalyst **1** (0.005 mmol, 5 mol %) in toluene (0.4 mL). The reaction mixture was stirred at -5 °C. After the purification *via* SiO₂ column chromatography, the desired product **4** was obtained.

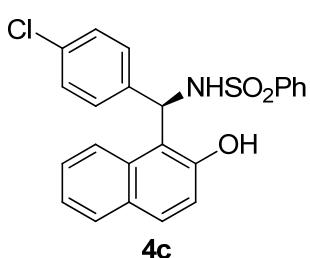


4a: 82% yield; ¹H-NMR (CDCl₃) δ: 7.76 (2H, d, *J* = 8.5 Hz), 7.76 (2H, d, *J* = 8.5 Hz), 7.68 (2H, d, *J* = 7.3 Hz), 7.45-7.41 (2H, m), 7.35-7.31 (2H, m), 7.15 (2H, d, *J* = 2.3 Hz), 7.10 (2H, dd, *J* = 8.7, 2.3 Hz), 6.57 (1H, d, *J* = 8.7 Hz), 4.97 (2H, bs); ¹³C-NMR (CDCl₃) δ: 155.7, 145.4, 139.0, 132.3, 132.0, 131.2, 130.4, 129.6, 129.4, 129.2, 128.8, 127.0, 123.1, 121.1, 119.9, 117.8, 114.2, 113.0, 61.5;

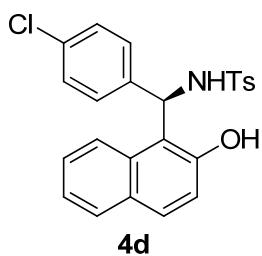
HRMS (ESI) calcd for C₂₃H₁₆BrClNO, m/z = 436.0104 [(M-H)⁺], found m/z = 436.0094; IR (KBr): ν 3569, 3339, 2345, 1719, 1625, 1490, 1233, 913, 815 cm⁻¹; enantiomeric excess: *Rac*, determined by HPLC (Chiralpak AS-H, hexane/2-propanol = 4/1, flow rate 1.0 mL/min, 25 °C, 330 nm) first peak: t_R = 6.7 min, second peak: t_R = 9.3 min.



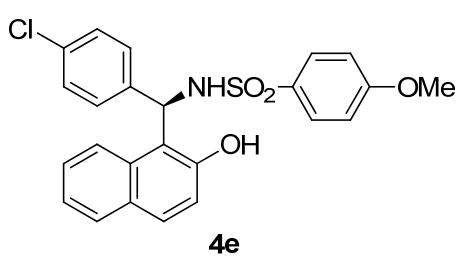
4b^{2b}: 17% yield; ¹H-NMR (ACETONE-D₆) δ: 9.30 (1H, bs), 8.14 (1H, d, *J* = 8.2 Hz), 7.89 (1H, d, *J* = 8.2 Hz), 7.85 (1H, d, *J* = 8.2 Hz), 7.53 (1H, t, *J* = 8.2 Hz), 7.39-7.30 (6H, m), 7.00 (1H, s), 3.27 (1H, bs), 1.47 (9H, s); ¹³C-NMR (CDCl₃) δ: 156.5, 153.6, 143.0, 133.4, 132.5, 130.6, 130.0, 129.7, 128.9, 128.8, 128.0, 124.0, 123.4, 120.2, 119.3, 79.6, 50.8, 28.6; enantiomeric excess: 17%, determined by HPLC (Chiralpak IB, hexane/2-propanol = 95/5, flow rate 1.0 mL/min, 25 °C, 230 nm) minor peak: t_R = 7.6 min, major peak: t_R = 7.0 min.



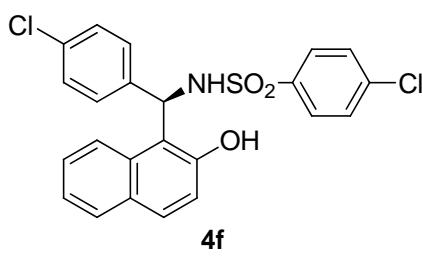
4c: 88% yield; ¹H-NMR (CDCl₃) δ: 7.71 (1H, d, *J* = 8.2 Hz), 7.65 (1H, d, *J* = 8.2 Hz), 7.50 (1H, d, *J* = 8.7 Hz), 7.45-7.40 (3H, m), 7.30 (1H, t, *J* = 7.3 Hz), 7.22 (2H, d, *J* = 8.7 Hz), 7.15 (2H, t, *J* = 4.4 Hz), 7.07 (1H, t, *J* = 7.3 Hz), 6.89-6.82 (4H, m), 6.66 (1H, bs), 6.40 (1H, d, *J* = 10.1 Hz); ¹³C-NMR (CDCl₃) δ: 150.8, 139.2, 138.6, 133.1, 132.1, 130.2, 128.9, 128.6, 128.4 x 2, 128.1 x 2, 127.4, 126.4, 123.5, 121.6, 117.8, 116.9, 53.7; HRMS (ESI) calcd for C₂₃H₁₈CINO₃SNa, m/z = 446.0594 [(M+Na)⁺], found m/z = 446.0584; IR (KBr): ν 3319, 1701, 1630, 1518, 1490, 1439, 1328, 1156, 1091, 1014, 751 cm⁻¹; enantiomeric excess: 22%, determined by HPLC (Chiralcel OD-3, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 12.0 min, major peak: t_R = 16.3 min.



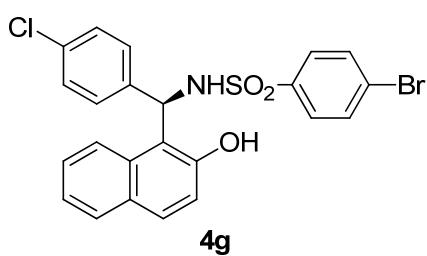
4d^{2a,3-5}: 70% yield; ¹H-NMR (CDCl₃) δ: 7.70 (1H, d, *J* = 9.2 Hz), 7.66 (1H, d, *J* = 8.7 Hz), 7.55 (1H, d, *J* = 8.7 Hz), 7.41 (1H, t, *J* = 7.8 Hz), 7.31 (2H, d, *J* = 8.2 Hz), 7.23 (2H, d, *J* = 8.7 Hz), 7.18 (2H, d, *J* = 8.2 Hz), 6.83 (1H, d, *J* = 8.7 Hz), 6.64 (2H, d, *J* = 7.8 Hz), 6.60 (1H, bs), 6.35 (1H, d, *J* = 10.1 Hz), 6.28 (1H, bs), 2.10 (3H, s); ¹³C-NMR (CDCl₃) δ: 151.1, 142.8, 138.7, 135.9, 132.9, 132.1, 129.8, 128.7, 128.6, 128.4, 128.3, 128.2, 127.2, 126.4, 123.3, 121.6, 117.9, 116.9, 53.7, 21.1; enantiomeric excess: 27%, determined by HPLC (Chiralpak IC, hexane/2-propanol = 4/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 14.1 min, major peak: t_R = 16.1 min.



4e: 65% yield; ¹H-NMR (CDCl₃) δ: 7.72 (1H, d, *J* = 8.7 Hz), 7.68 (1H, d, *J* = 8.1 Hz), 7.57 (1H, d, *J* = 8.7 Hz), 7.43 (1H, t, *J* = 7.3 Hz), 7.36-7.31 (3H, m), 7.23-7.18 (4H, m), 6.83 (1H, d, *J* = 8.7 Hz), 6.33 (3H, t, *J* = 8.1 Hz), 3.62 (3H, s); ¹³C-NMR (CDCl₃) δ: 162.3, 150.8, 138.7, 133.1, 132.3, 130.8, 130.0, 129.0, 128.7, 128.5, 128.4, 128.2, 127.4, 123.6, 121.8, 118.0, 117.4, 113.3, 55.3, 53.7; HRMS (ESI) calcd for C₂₄H₂₀ClNO₄SNa, m/z = 476.0699 [(M+Na)⁺], found m/z = 476.0689; IR (KBr): ν 3310, 2930, 1597, 1496, 1257, 1155, 1092, 910, 815, 731 cm⁻¹; enantiomeric excess: 30%, determined by HPLC (Chiralpak IA, hexane/2-propanol = 9/1, flow rate 1.0 mL/min, 25 °C, 335 nm) minor peak: t_R = 17.7 min, major peak: t_R = 22.6 min.

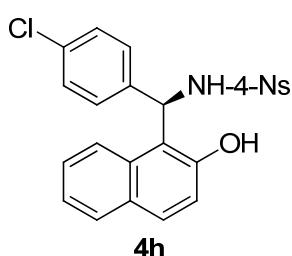


4f: 100% yield; ¹H-NMR (CDCl₃) δ: 7.74-7.71 (2H, m), 7.60 (1H, d, *J* = 8.7 Hz), 7.46 (1H, t, *J* = 7.5 Hz), 7.37 (1H, t, *J* = 7.5 Hz), 7.30-7.20 (8H, m), 6.77 (3H, d, *J* = 8.7 Hz), 6.40 (1H, s); ¹³C-NMR (CDCl₃) δ: 150.7, 138.4, 138.3, 137.7, 133.2, 130.2, 129.4, 128.7, 128.5, 128.1 x 2, 127.9, 127.8, 127.6, 123.8, 121.5, 117.7, 117.0, 53.8; HRMS (ESI) calcd for C₂₃H₁₇Cl₂NO₃SNa, m/z = 480.0204 [(M+Na)⁺], found m/z = 480.0193; IR (KBr): ν 3414, 3318, 2345, 1718, 1629, 1578, 1509, 1323, 1273, 1152, 1091, 812, 752 cm⁻¹; enantiomeric excess: 48%, determined by HPLC (Chiralcel OD-H, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 21.1 min, major peak: t_R = 27.7 min.

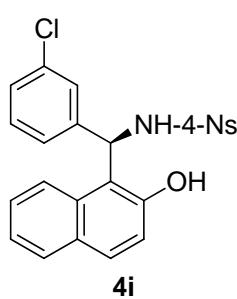


4g: 72% yield; ¹H-NMR (CDCl₃) δ: 7.73 (2H, d, *J* = 8.7 Hz), 7.61 (1H, d, *J* = 8.7 Hz), 7.46 (1H, t, *J* = 7.9 Hz), 7.37 (1H, t, *J* = 7.9 Hz), 7.27-7.20 (6H, m), 6.93 (2H, d, *J* = 8.7 Hz), 6.79 (1H, d, *J* = 8.7 Hz), 6.54 (1H, bs), 6.39 (1H, bs), 5.73 (1H, bs); ¹³C-NMR (CDCl₃) δ: 150.5, 138.3 x 2, 133.3, 132.4, 131.1, 130.2, 128.8, 128.5, 128.1, 128.0, 127.9 x 2, 127.7, 126.9, 123.9, 117.6, 117.1, 53.7; HRMS (ESI) calcd for C₂₃H₁₇BrClNO₃SNa, m/z = 523.9699 [(M+Na)⁺], found

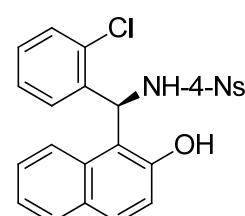
m/z = 523.9687; IR (KBr): ν 3403, 3317, 3089, 2925, 2853, 2345, 1903, 1719, 1629, 1575, 1544, 1326, 1273, 1153, 1089 cm^{-1} ; enantiomeric excess: 63%, determined by HPLC (Chiralcel OD-H, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 16.2 min, major peak: t_R = 22.2 min.



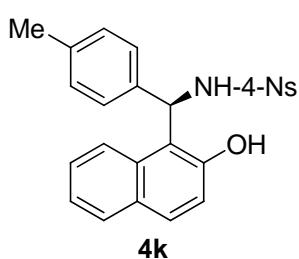
4h: 89% yield; $^1\text{H-NMR}$ (DMSO-D₆) δ : 9.99 (1H, br s), 8.81 (1H, br s), 7.88 (2H, d, J = 8.7 Hz), 7.63 (2H, d, J = 8.7 Hz), 7.58 (2H, d, J = 8.2 Hz), 7.55 (1H, d, J = 9.2 Hz), 7.31 (2H, d, J = 8.7 Hz), 7.28-7.20 (3H, m), 7.16 (1H, t, J = 7.3 Hz), 6.96 (1H, d, J = 8.7 Hz), 6.52 (1H, s); $^{13}\text{C-NMR}$ (DMSO-D₆) δ : 153.0, 148.6, 145.9, 139.9, 131.3, 131.2, 129.7, 128.2, 128.1, 128.0, 127.4, 126.3, 123.1, 122.4, 117.6, 116.1, 51.9; HRMS (ESI) calcd for C₂₃H₁₇ClN₂O₅SNa, *m/z* = 491.0444 [(M+Na)⁺], found *m/z* = 491.0432; IR (KBr): ν 3432, 3315, 3112, 2548, 2375, 1730, 1628, 1586, 1521, 1320, 1280, 1154, 1080 cm^{-1} ; enantiomeric excess: 96%, determined by HPLC (Chiralcel OD-3, hexane/2-propanol = 9/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 8.9 min, major peak: t_R = 10.9 min; $[\alpha]_D^{22}$ = +16.9 (*c* 1.8, CHCl₃).



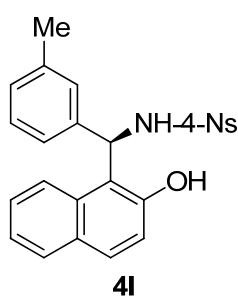
4i: 92% yield; $^1\text{H-NMR}$ (DMSO-D₆) δ : 7.89 (2H, d, J = 9.2 Hz), 7.65 (2H, d, J = 9.2 Hz), 7.60 (2H, d, J = 7.8 Hz), 7.57 (1H, d, J = 8.7 Hz), 7.31-7.21 (4H, m), 7.17 (1H, t, J = 7.3 Hz), 7.13-7.09 (1H, m), 6.96 (1H, d, J = 8.7 Hz), 6.53 (1H, s); $^{13}\text{C-NMR}$ (DMSO-D₆) δ : 153.1, 148.7, 146.0, 143.5, 132.8, 131.3, 130.0, 129.8, 128.3, 128.1, 127.5, 126.6, 126.4, 125.9, 125.0, 123.2, 122.9, 122.4, 117.7, 116.1, 52.1; HRMS (ESI) calcd for C₂₃H₁₇ClN₂O₅SNa, *m/z* = 491.0444 [(M+Na)⁺], found *m/z* = 491.0436; IR (KBr): ν 3443, 3310, 3108, 2377, 1629, 1581, 1522, 1432, 1323, 1275, 1156, 1081 cm^{-1} ; enantiomeric excess: 98%, determined by HPLC (Chiraldak AD-H, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 56.1 min, major peak: t_R = 44.9 min; $[\alpha]_D^{21}$ = +17.7 (*c* 1.5, CHCl₃).



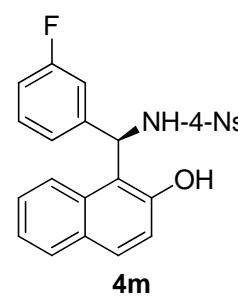
4j: 90% yield; $^1\text{H-NMR}$ (DMSO-D₆) δ : 10.30 (1H, br s), 7.92 (2H, d, J = 8.7 Hz), 7.77 (1H, d, J = 8.7 Hz), 7.65-7.50 (5H, m), 7.30-7.22 (2H, m), 7.18-7.11 (3H, m), 6.91 (1H, d, J = 8.7 Hz), 6.60 (1H, s); $^{13}\text{C-NMR}$ (DMSO-D₆) δ : 154.8, 148.2, 139.3, 132.0, 131.9, 129.4, 129.1, 128.5, 128.3, 127.8, 127.4, 126.6, 126.1, 123.0, 122.2, 122.0, 118.8, 115.9, 53.1; HRMS (ESI) calcd for C₂₃H₁₇ClN₂O₅SNa, *m/z* = 491.0444 [(M+Na)⁺], found *m/z* = 491.0435; IR (KBr): ν 3277, 3107, 3071, 2372, 1685, 1628, 1597, 1527, 1446, 1344, 1270, 1171, 1092 cm^{-1} ; enantiomeric excess: 88%, determined by HPLC (Chiraldak IC-3, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 254 nm) minor peak: t_R = 77.6 min, major peak: t_R = 61.2 min; $[\alpha]_D^{22}$ = +92.7 (*c* 1.8, MeOH).



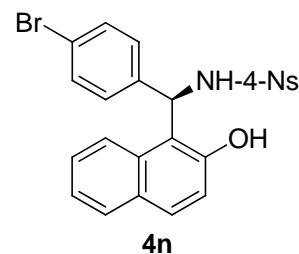
4k: 100% yield; $^1\text{H-NMR}$ (ACETONE-D₆) δ : 7.85 (1H, d, J = 9.2 Hz), 7.78 (2H, d, J = 8.7 Hz), 7.69-7.66 (3H, m), 7.60 (1H, d, J = 8.7 Hz), 7.43 (1H, t, J = 7.6 Hz), 7.29 (3H, m), 7.11 (2H, d, J = 7.8 Hz), 7.04 (1H, d, J = 9.2 Hz), 6.58 (1H, s), 2.29 (3H, s); $^{13}\text{C-NMR}$ (ACETONE-D₆) δ : 153.8, 150.4, 147.5, 138.9, 137.7, 133.6, 131.0, 130.1, 130.0, 129.7, 129.0, 128.1, 127.9, 125.5, 124.3, 124.2, 119.2, 118.4, 55.1, 21.4; HRMS (ESI) calcd for C₂₄H₂₀N₂O₅SNa, m/z = 471.0991 [(M+Na)⁺], found m/z = 471.0984; IR (KBr): ν 3432, 3313, 3111, 2375, 1629, 1520, 1431, 1321, 1277, 1153, 1078 cm⁻¹; enantiomeric excess: 90%, determined by HPLC (Chiralpak AD-H, hexane/2-propanol = 4/1, flow rate 1.0 mL/min, 25 °C, 266 nm) minor peak: t_R = 16.2 min, major peak: t_R = 21.1 min; $[\alpha]_D^{23} = +42.4$ (*c* 1.2, CHCl₃).



4l: 95% yield; $^1\text{H-NMR}$ (ACETONE-D₆) δ : 7.86 (1H, d, J = 8.2 Hz), 7.79 (2H, d, J = 8.7 Hz), 7.70-7.66 (3H, m), 7.61 (1H, d, J = 8.7 Hz), 7.44 (1H, t, J = 8.0 Hz), 7.32-7.26 (2H, m), 7.20-7.14 (2H, m), 7.09-7.01 (2H, m), 6.59 (1H, s), 2.27 (3H, s); $^{13}\text{C-NMR}$ (ACETONE-D₆) δ : 153.3, 150.0, 147.1, 141.5, 138.4, 133.1, 130.6, 129.6, 129.2, 128.9, 128.6, 128.5, 128.0, 127.7, 124.6, 123.8, 123.8, 123.3, 118.7, 118.0, 54.7; HRMS (ESI) calcd for C₂₄H₂₀N₂O₅SNa, m/z = 471.0991 [(M+Na)⁺], found m/z = 471.0980; IR (KBr): ν 3452, 3316, 3107, 1620, 1523, 1434, 1323, 1281, 1156, 1079 cm⁻¹; enantiomeric excess: 99%, determined by HPLC (Chiralpak IC-3, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 231 nm) minor peak: t_R = 36.9 min, major peak: t_R = 41.0 min; $[\alpha]_D^{24} = +29.5$ (*c* 2.0, CHCl₃).

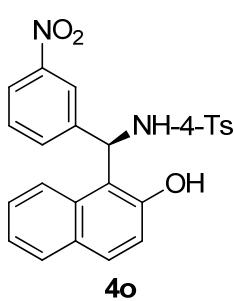


4m: 98% yield; $^1\text{H-NMR}$ (CDCl₃) δ : 7.75 (2H, d, J = 8.7 Hz), 7.65-7.47 (2H, m), 7.35 (2H, t, J = 7.8 Hz), 7.25 (1H, m), 7.12 (2H, d, J = 7.8 Hz), 7.03 (1H, d, J = 10.1 Hz), 6.94 (2H, t, J = 7.8 Hz), 6.76 (2H, d, J = 8.7 Hz), 6.70 (1H, s), 6.43 (1H, d, J = 10.1 Hz), 5.73 (1H, s); $^{13}\text{C-NMR}$ (ACETONE-D₆) δ : 164.8, 162.4, 153.4, 150.0, 147.0, 144.8, 133.0, 131.0, 130.9, 130.8, 129.6, 129.3, 128.7, 127.9, 123.9, 123.3, 118.6, 117.5, 114.6, 114.3, 114.1, 54.2; HRMS (ESI) calcd for C₂₃H₁₇FN₂O₅SNa, m/z = 475.0740 [(M+Na)⁺], found m/z = 475.0736; IR (KBr): ν 3419, 3304, 2375, 1696, 1619, 1526, 1439, 1346, 1256, 1165, 1086 cm⁻¹; enantiomeric excess: 85%, determined by HPLC (Chiralpak IE, hexane/2-propanol = 7/1, flow rate 1.0 mL/min, 25 °C, 266 nm) minor peak: t_R = 11.4 min, major peak: t_R = 8.6 min; $[\alpha]_D^{14} = +12.6$ (*c* 1.5, CHCl₃).

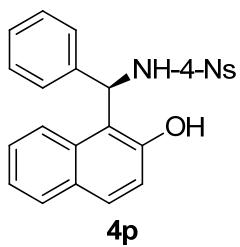


4n: 90% yield; $^1\text{H-NMR}$ (DMSO-D₆) δ : 9.96 (1H, br s), 8.81 (1H, br s), 7.88 (2H, d, J = 8.7 Hz), 7.64 (2H, d, J = 8.7 Hz), 7.58 (2H, d, J = 7.3 Hz), 7.55 (1H, d, J = 9.2 Hz), 7.44 (2H, d, J = 8.7 Hz), 7.25 (1H, t, J = 8.2 Hz), 7.19-7.13 (3H, m), 6.95 (1H, d, J = 9.2 Hz), 6.49 (1H, s); $^{13}\text{C-NMR}$ (DMSO-D₆) δ : 153.0, 148.6, 146.0, 140.4, 131.3, 131.0, 129.7, 128.4,

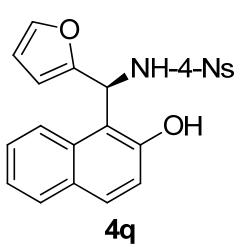
128.2, 127.4, 126.3, 123.1, 122.4, 119.7, 117.6, 116.1, 52.0; HRMS (ESI) calcd for $C_{23}H_{17}BrN_2O_5SNa$, m/z = 534.9939 [(M+Na) $^+$], found m/z = 534.9926; IR (KBr): ν 3426, 3315, 3110, 2544, 2465, 2374, 1618, 1522, 1318, 1280, 1157, 1112 cm^{-1} ; enantiomeric excess: 90%, determined by HPLC (Chiralcel OD-H, hexane/2-propanol = 19/1, flow rate 1.0 mL/min, 25 °C, 250 nm) minor peak: t_R = 47.5 min, major peak: t_R = 61.2 min; $[\alpha]_D^{20}$ = +21.8 (c 1.6, CHCl₃).



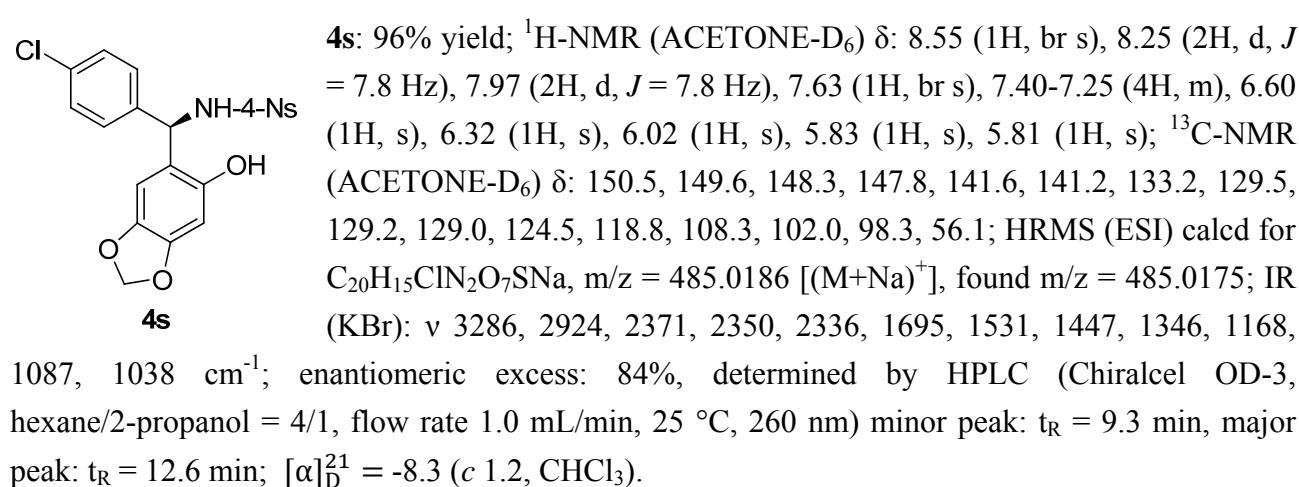
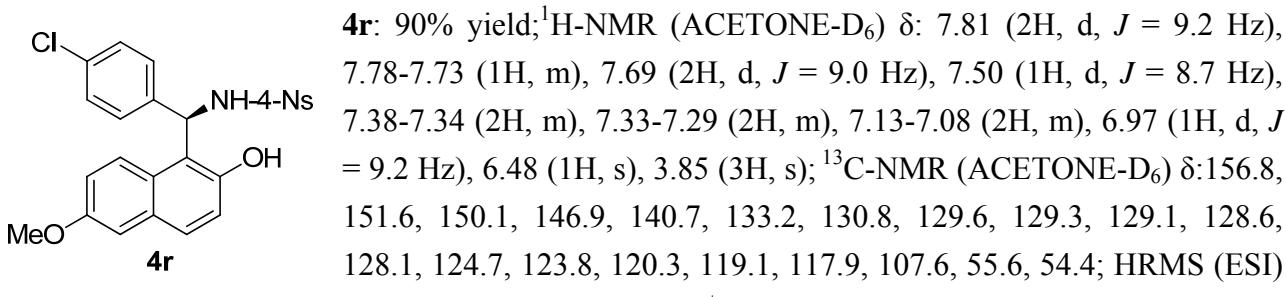
4o^{2a}: 80% yield; ¹H-NMR (CDCl₃) δ : 8.12 (1H, s), 8.06 (1H, d, J = 7.8 Hz), 7.75-7.67 (3H, m), 7.59 (1H, d, J = 8.6 Hz), 7.45 (1H, t, J = 7.1 Hz), 7.39 (1H, t, J = 7.1 Hz), 7.34 (3H, d, J = 8.6 Hz), 6.85 (1H, d, J = 8.6 Hz), 6.68 (3H, d, J = 7.8 Hz), 6.45 (1H, d, J = 10.1 Hz), 5.99 (1H, s), 2.11 (3H, s); ¹³C-NMR (CDCl₃) δ : 151.2, 148.1, 143.0, 142.8, 135.8, 133.1, 132.0, 130.2, 129.1, 128.8, 128.7, 128.5, 127.5, 126.4, 123.5, 122.2, 121.6, 121.3, 117.8, 116.4, 53.5, 21.1; enantiomeric excess: 73%, determined by HPLC (Chiraldak IC-3, hexane/2-propanol = 4/1, flow rate 1.0 mL/min, 25 °C, 335 nm) minor peak: t_R = 12.4 min, major peak: t_R = 15.4 min.



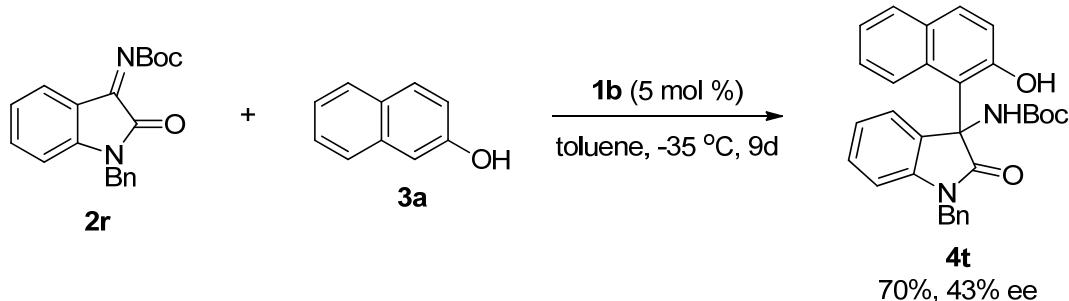
4p: 100% yield; ¹H-NMR (ACETONE-D₆) δ : 7.87 (1H, d, J = 8.7 Hz), 7.81 (2H, d, J = 8.7 Hz), 7.70 (2H, d, J = 8.7 Hz), 7.69 (1H, d, J = 8.7 Hz), 7.63 (1H, d, J = 8.7 Hz), 7.46-7.41 (3H, m), 7.34-7.23 (4H, m), 7.04 (1H, d, J = 8.7 Hz), 6.64 (1H, s); ¹³C-NMR (ACETONE-D₆) δ : 153.3, 150.0, 147.0, 141.5, 133.1, 130.7, 129.6, 129.2, 129.0 x 2, 128.6, 127.8, 127.4, 123.8, 123.3, 118.6, 117.9, 54.7; HRMS (ESI) calcd for $C_{23}H_{18}N_2O_5SNa$, m/z = 457.0834 [(M+Na) $^+$], found m/z = 457.0827; IR (KBr): ν 3305, 2359, 1698, 1528, 1430, 1348, 1312, 1162, 1090, 1028 cm^{-1} ; enantiomeric excess: 73%, determined by HPLC (Chiraldak AD-H, hexane/2-propanol = 7/1, flow rate 1.0 mL/min, 25 °C, 268 nm) minor peak: t_R = 18.3 min, major peak: t_R = 21.4 min; $[\alpha]_D^{20}$ = +22.4 (c 2.3, CHCl₃).



4q: 100% yield; ¹H-NMR (ACETONE-D₆) δ : 7.99 (1H, d, J = 8.7 Hz), 7.86 (2H, d, J = 8.7 Hz), 7.72 (2H, d, J = 8.7 Hz), 7.68 (1H, d, J = 8.2 Hz), 7.62 (1H, d, J = 8.7 Hz), 7.48-7.42 (2H, m), 7.29 (1H, t, J = 7.5 Hz), 7.05 (1H, d, J = 8.7 Hz), 6.69 (1H, s), 6.34 (1H, dd, J = 3.2, 1.8 Hz), 6.23 (1H, d, J = 3.2 Hz); ¹³C-NMR (ACETONE-D₆) δ : 153.8, 153.7, 150.0, 147.1, 143.0, 132.9, 130.9, 129.6, 129.2, 128.6, 127.6, 123.8, 123.2, 118.6, 115.7, 111.2, 108.4, 50.0; HRMS (ESI) calcd for $C_{21}H_{16}N_2O_6SNa$, m/z = 447.0627 [(M+Na) $^+$], found m/z = 447.0617; IR (KBr): ν 3396, 3324, 3162, 3104, 3067, 1702, 1614, 1525, 1411, 1345, 1310, 1281, 1160, 1078 cm^{-1} ; enantiomeric excess: 72%, determined by HPLC (Chiraldak AD-H, hexane/2-propanol = 7/1, flow rate 1.0 mL/min, 25 °C, 268 nm) minor peak: t_R = 45.3 min, major peak: t_R = 25.6 min; $[\alpha]_D^{20}$ = +37.0 (c 3.9, CHCl₃).



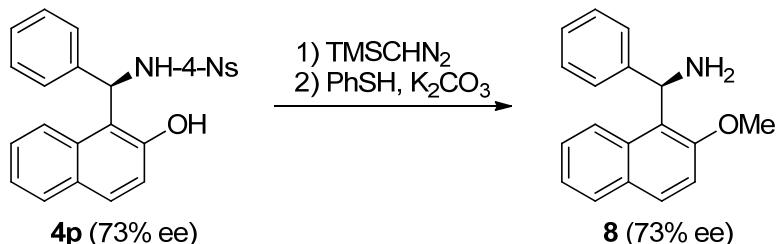
Procedure for enantioselective Friedel–Crafts (FC)-type reaction of ketimine (**2r**) with 2-naphthol (**3a**)



Under N₂ atmosphere, a test tube was charged with imine **2r** (0.10 mmol), 2-naphthol **3a** (0.15 mmol), and the catalyst **1b** (0.005 mmol, 5 mol %) in toluene (0.4 mL). The reaction mixture was stirred at -35 °C. After the purification *via* SiO₂ column chromatography (hexane/AcOEt = 3/1), the desired product **4** was obtained in 70% yield (33.6mg, 0.07 mmol) with 43% ee. **4t:** 70% yield; $^1\text{H-NMR}$ (CDCl₃) δ : 10.02 (1H, s), 7.68 (2H, dd, J = 7.8, 3.7 Hz), 7.34-7.16 (9H, m), 7.05 (1H, t, J = 7.8 Hz), 6.97 (1H, t, J = 7.8 Hz), 6.80 (2H, d, J = 7.8 Hz), 5.87 (1H, s), 5.18 (1H, d, J = 15.8 Hz), 4.87 (1H, d, J = 15.8 Hz), 1.31 (9H, s); $^{13}\text{C-NMR}$ (CDCl₃) δ : 179.7, 153.8, 143.0, 138.6, 138.2, 135.0, 131.9, 130.7, 130.2, 129.6, 128.8 x 2, 127.6, 127.1, 125.5 x 2, 124.3, 123.5, 122.8, 121.5,

114.2, 110.0, 80.7, 65.5, 44.6, 28.1; HRMS (ESI) calcd for $C_{30}H_{28}N_2O_4Na$, m/z = 503.1947 [(M+Na)⁺], found m/z = 503.1932; IR (KBr): ν 3269, 1696, 1610, 1490, 1349, 1274, 1161, 1048, 818, 738 cm⁻¹; enantiomeric excess: 43%, determined by HPLC (Chiralpak AD-H, hexane/2-propanol = 4/1, flow rate 1.0 mL/min, 25 °C, 231 nm) minor peak: t_R = 17.7 min, major peak: t_R = 36.9 min; $[\alpha]_D^{24} = +8.0$ (*c* 1.1, CHCl₃).

Procedures for the deprotection of sulfonamide group

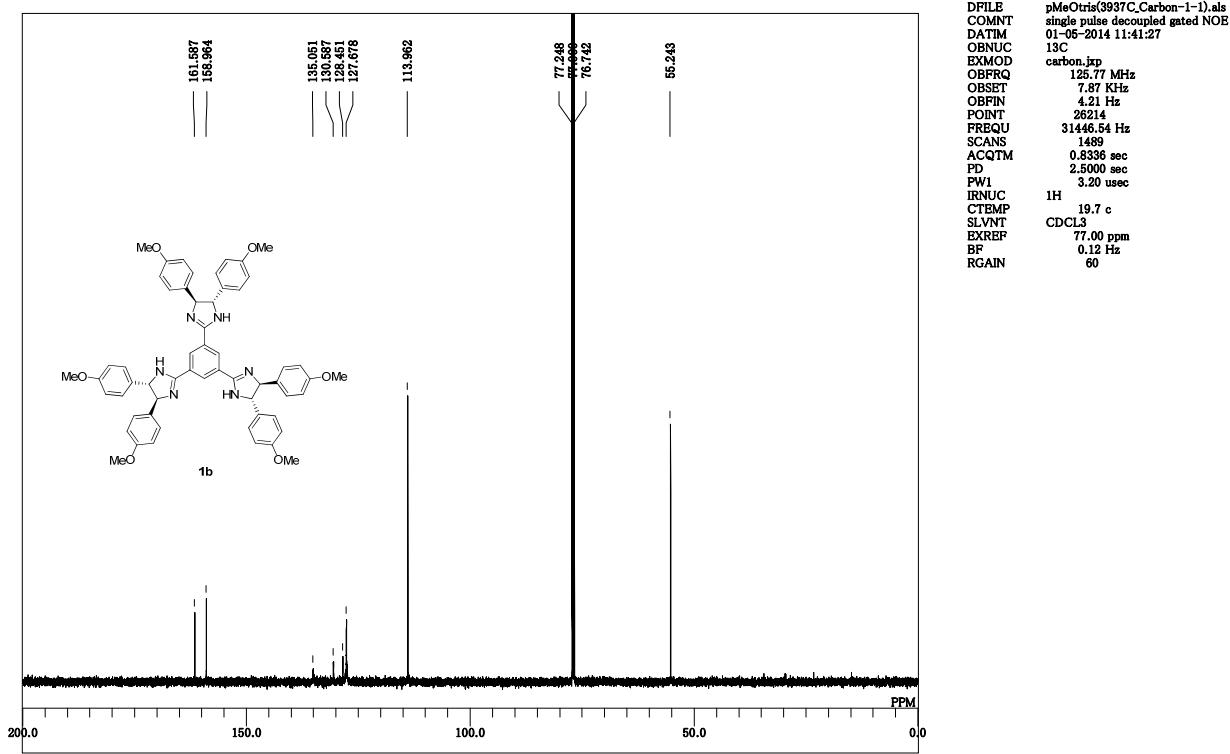
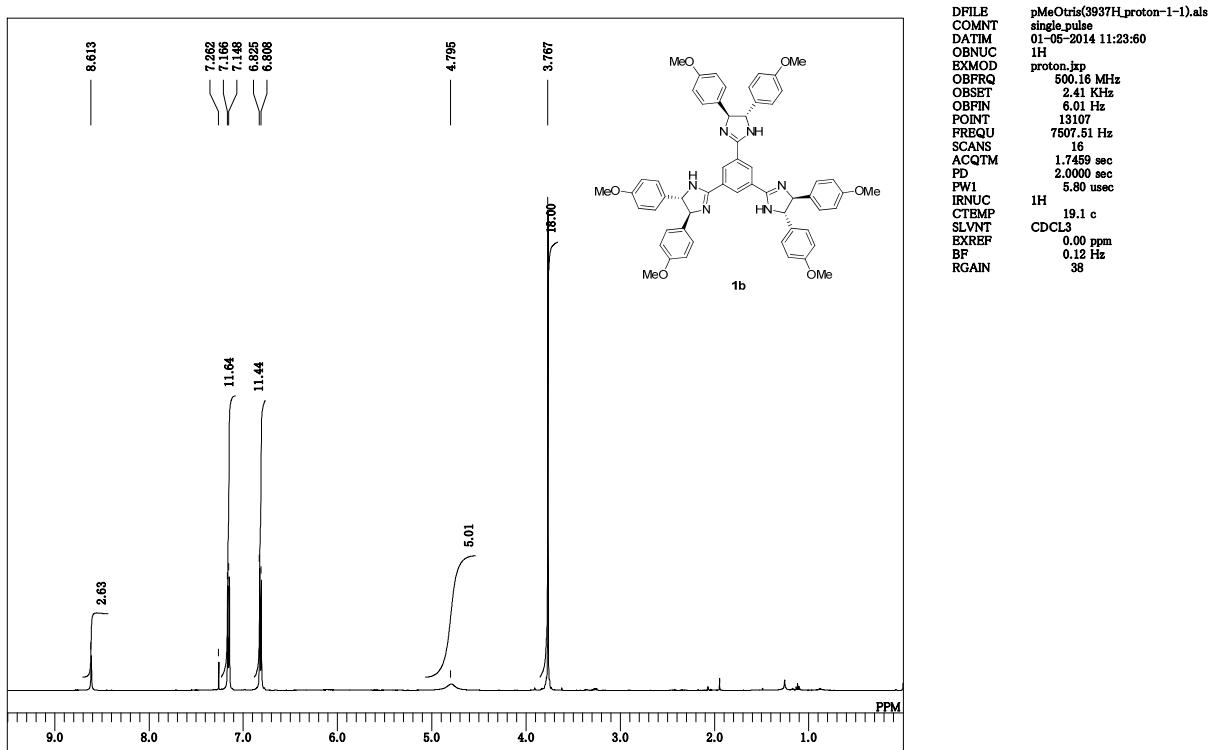


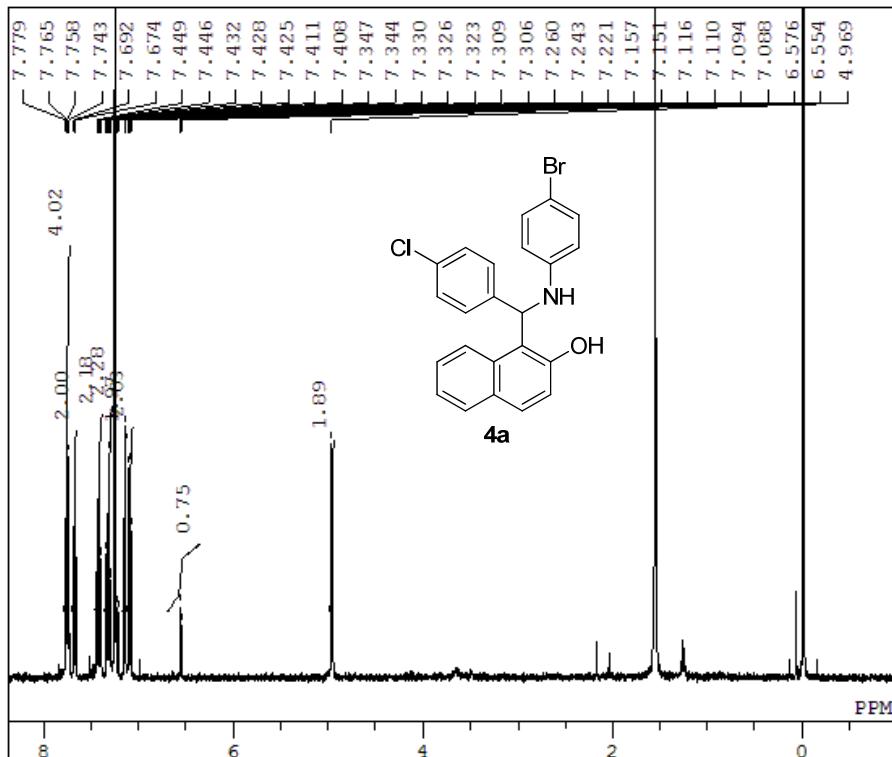
To a solution of compound **4p** (50 mg, 0.12 mmol) in MeOH (0.2 mL) and DCM (0.2 mL), TMSCHN₂ (2N solution in Et₂O, 0.44 mL, 1.0 mmol) was added in 10 portions over 1 h at rt. The reaction mixture was then stirred at 45 °C for 0.5 h. The solvent was evaporated in vacuo. After the purification *via* SiO₂ column chromatography (hexane/AcOEt = 2/1), the methyl-capped product was obtained (18 mg, 0.04 mmol, 40% yield); ¹H-NMR (CDCl₃) δ : 7.78 (1H, d, *J* = 8.4 Hz), 7.64 (2H, d, *J* = 9.2 Hz), 7.56 (2H, d, *J* = 8.4 Hz), 7.49-7.42 (3H, m), 7.35 (1H, t, *J* = 7.5 Hz), 7.28-7.20 (4H, m), 6.99 (1H, d, *J* = 9.2 Hz), 6.62 (1H, bs), 6.47 (1H, d, *J* = 10.5 Hz), 3.67 (3H, s); ¹³C-NMR (CDCl₃) δ : 154.3, 148.9, 145.4, 139.6, 132.0, 130.3, 128.8, 128.5, 128.3, 127.8, 127.6, 127.3, 126.3, 124.3, 122.7, 121.8, 120.4, 113.4, 56.5, 54.1; HRMS (ESI) calcd for $C_{24}H_{20}N_2O_5SNa$, m/z = 471.0991 [(M+Na)⁺], found m/z = 471.0966; IR (KBr): ν 3435, 3302, 3104, 2917, 2352, 1523, 1348, 1156, 1022 cm⁻¹. Subsequently PhSH (44 μ L, 0.43 mmol) was added to a solution of the methyl-capped product (10 mg, 0.022 mmol) and K₂CO₃ (55.2 mg, 0.040 mmol) in DMF (0.2 mL) and CH₃CN (0.2 mL). The mixture was stirred at 50 °C for 4 h. Water was added to the reaction mixture and the solution was extracted with AcOEt. The combined organic layer was back extracted with 1N HCl and then the aqueous layer was basified with NaOH and extracted with AcOEt. Organic layer was dried over Na₂SO₄, and evaporated in vacuo. After the purification *via* SiO₂ column chromatography (AcOEt), the desired product **8** was obtained (5.9 mg, 0.022 mmol, quant) without racemization. $[\alpha]_D^{23} = -145$ (73% ee, *c* 0.85, CHCl₃) (lit.⁶ $[\alpha]_D^{25} = +196$ (> 99% ee, *c* 1.6, CHCl₃)); ¹H-NMR (CDCl₃) δ : 8.02 (1H, d, *J* = 8.7 Hz), 7.80 (2H, d, *J* = 8.7 Hz), 7.42-7.36 (3H, m), 7.34-7.22 (4H, m), 7.18-7.13 (1H, m), 6.13 (1H, s), 3.79 (3H, s), 2.15 (2H, br s); ¹³C-NMR (CDCl₃) δ : 154.8, 145.4, 132.0, 129.6, 129.2, 128.7, 127.9, 126.8, 126.6, 126.0, 125.9, 123.5, 123.4, 114.2, 56.5, 50.9.

References

- 1) (a) Z. Xu and X. Lu, *J. Org. Chem.*, 1998, **63**, 5031; (b) T. Hayashi and M. Ishigedani, *J. Am. Chem. Soc.*, 2000, **122**, 976; (c) M. Yamanaka, A. Nishida and M. Nakagawa, *J. Org. Chem.*, 2003, **68**, 3112; (d) I. T. Raheem and E. N. Jacobsen, *Adv. Synth. Catal.*, 2005, **347**, 1701; (e) Y. Otomaru, N. Tokunaga, R. Shintani and T. Hayashi, *Org. Lett.*, 2005, **7**, 307; (f) Y. -X. Jia, J. -H. Xie, H. -F. Duan, L. -X. Wang and Q. -L. Zhou, *Org. Lett.*, 2006, **8**, 1621; (g) J. Esquivias, R. G. Arrayás and J. C. Carretero, *Angew. Chem. Int. Ed.*, 2006, **45**, 629; (h) S. Castellano, H. D. G. Fiji, S. S. Kinderman, M. Watanabe, P. de Leon, F. Tamanoi, and O. Kwon, *J. Am. Chem. Soc.*, 2007, **129**, 5843; (i) K. M. Partridge, M. E. Anzovino and T. P. Yoon, *J. Am. Chem. Soc.*, 2008, **130**, 2920; (j) F. Tato, V. Reboul and P. Metzner, *J. Org. Chem.*, 2008, **73**, 7837; (k) S. M. DePorter, A. C. Jacobsen, K. M. Partridge, K. S. Williamson and T. P. Yoon, *Tetrahedron Lett.*, 2010, **51**, 5223. (l) D. -J. Dong, H. -H. Li and S. -K. Tian, *J. Am. Chem. Soc.*, 2010, **132**, 5018 (m) E. G. -Bengoa, M. Maestro, A. Mielgo, I. Otazo, C. Palomo and I. Velilla, *Chem. Eur. J.*, 2010, **16**, 5333; (n) A. S. Tsai, M. E. Tauchert, R. G. Bergman and J. A. Ellman, *J. Am. Chem. Soc.*, 2011, **133**, 1248.
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- 4) G. Liu, S. Zhang, H. Li, T. Zhang and W. Wang, *Org. Lett.*, 2011, **13**, 828.
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¹H-NMR and ¹³C-NMR charts

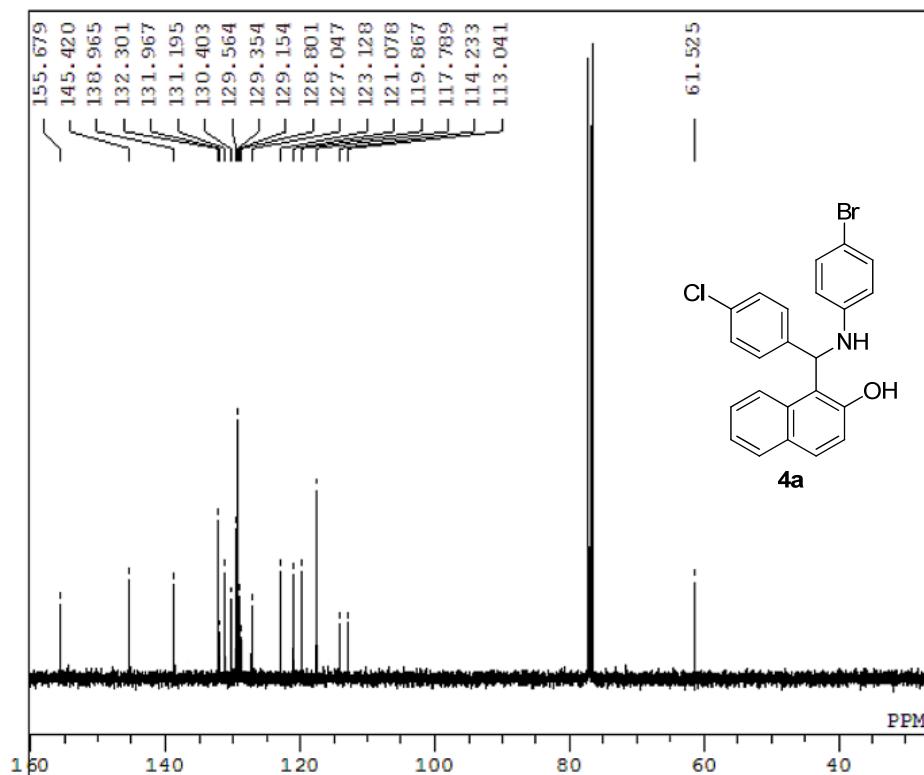




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PD          5.0000 sec
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RGAIN         58

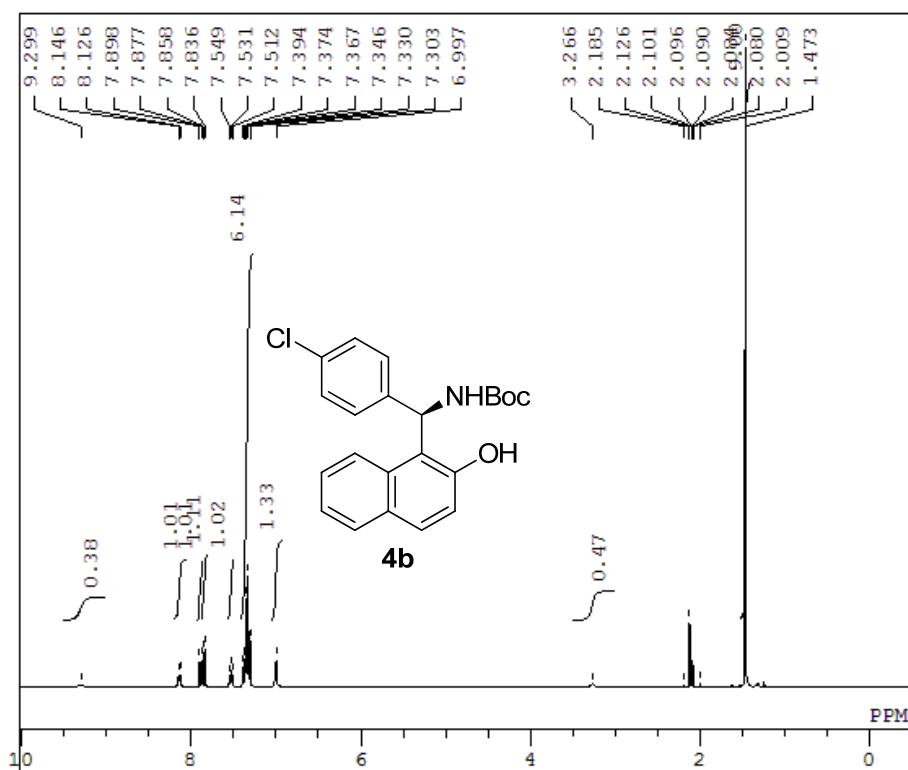
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PD          2.0000 sec
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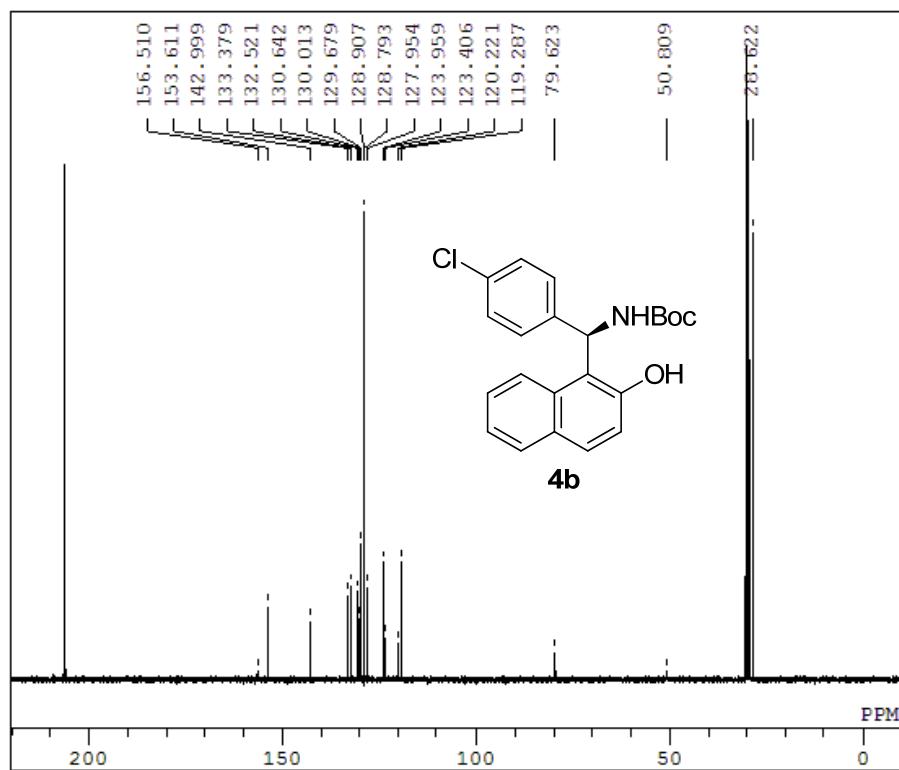
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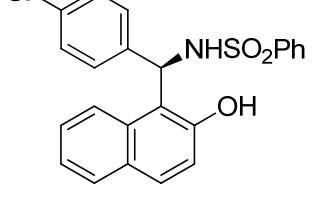
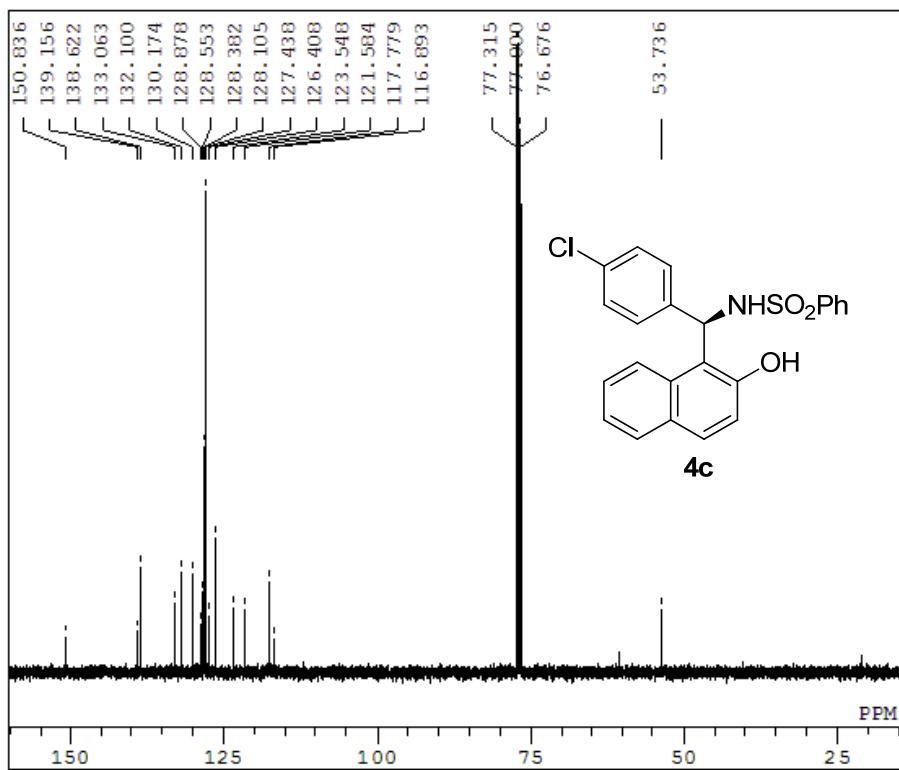
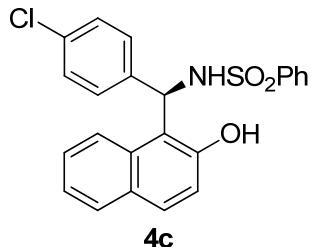
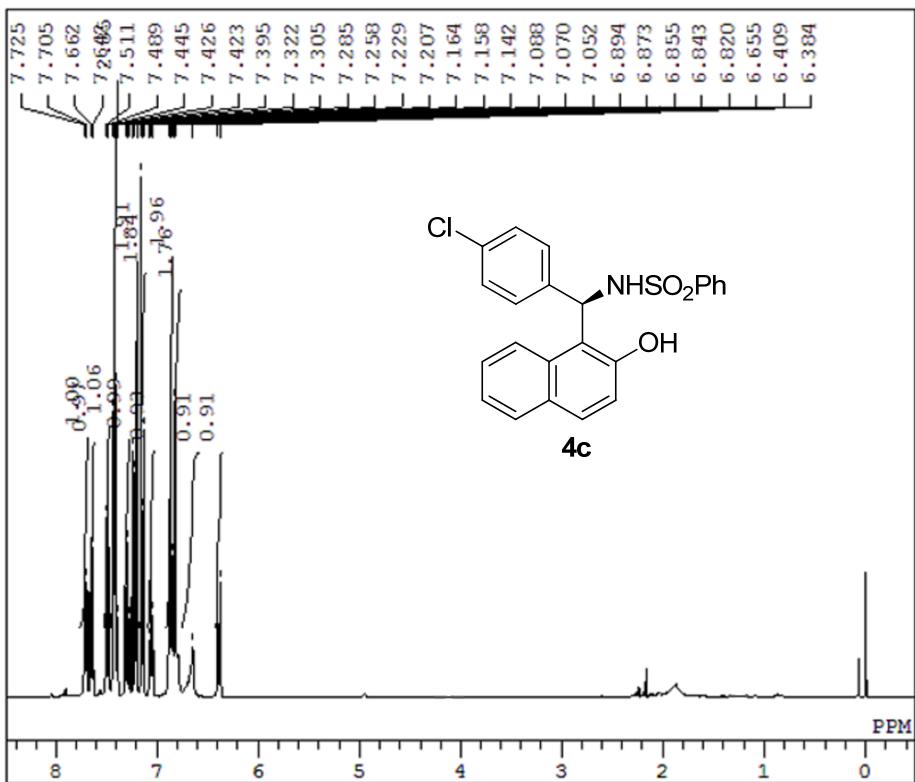
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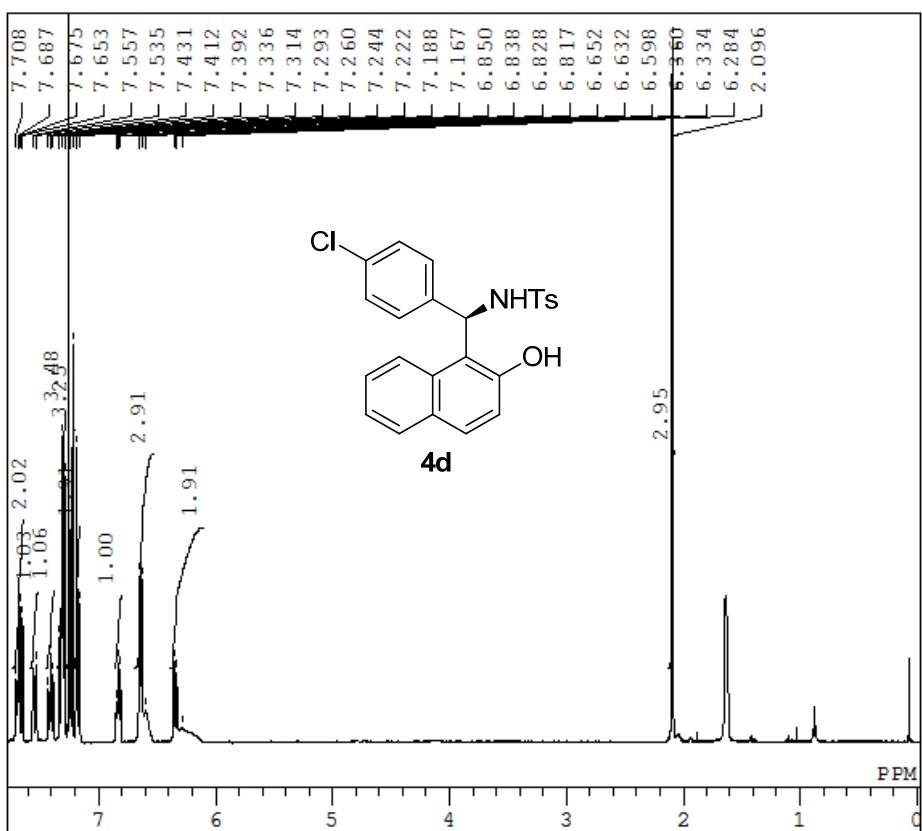
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RGAIN      40

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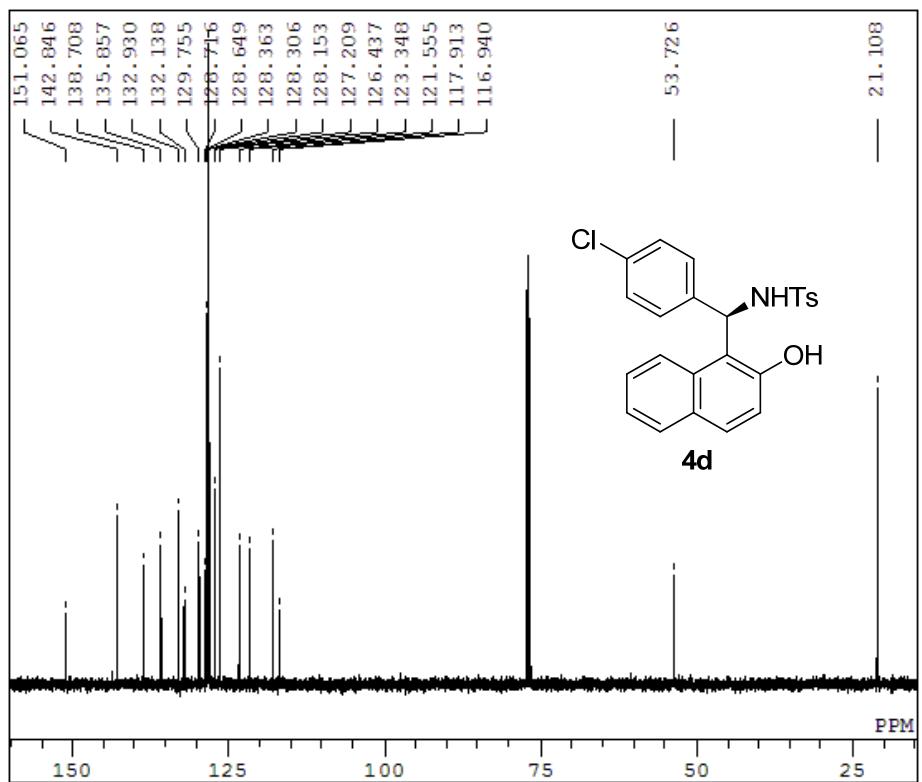
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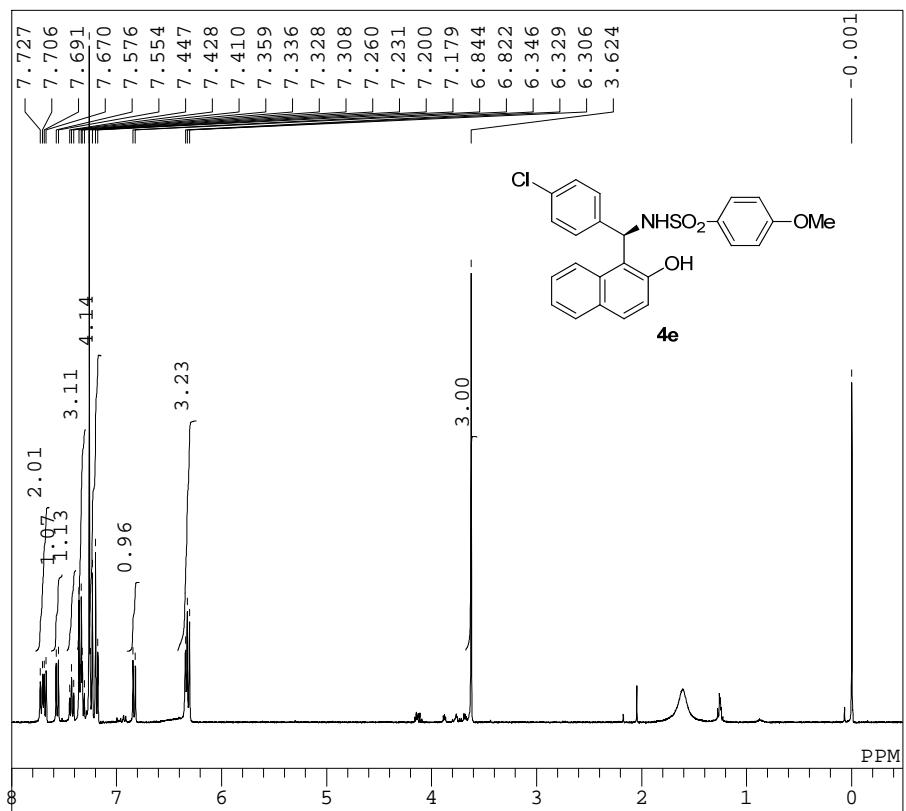


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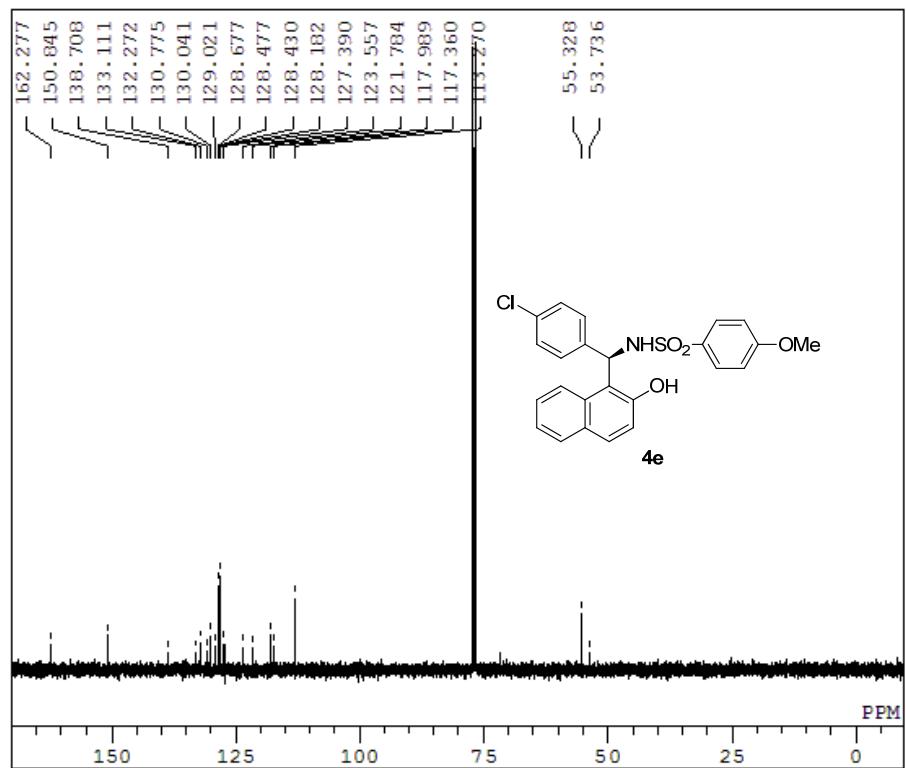
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SLVNT CDCL3
EXREF 7.26 ppm
BF 0.10 Hz
RGAIN 46

```

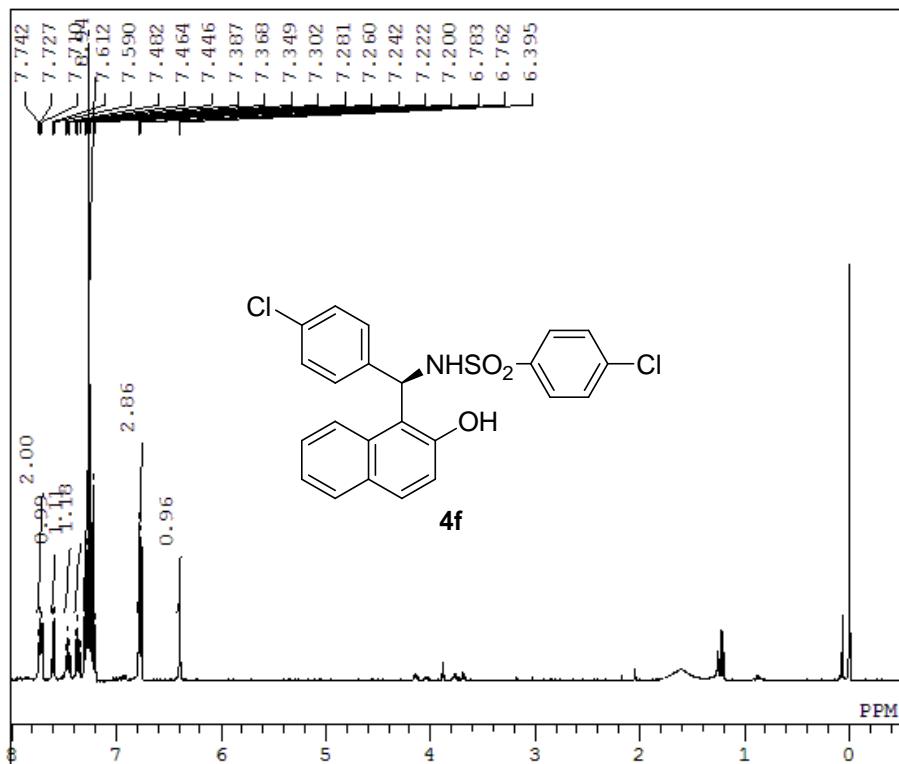




DFILE sh4032-gpc3_Protc
 COMNT single_pulse
 DATIM 2012-11-21 17:31:
 OBNUC 1H
 EXMOD proton.jxp
 OBFRQ 399.78 MHz
 OBSET 4.19 KHz
 OBFIN 7.29 Hz
 POINT 16384
 FREQU 7503.00 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 5.0000 sec
 PW1 5.50 usec
 IRNUC 1H
 CTEMP 20.3 c
 SLVNT CDCL3
 EXREF 7.26 ppm
 BF 0.10 Hz
 RGAIN 46



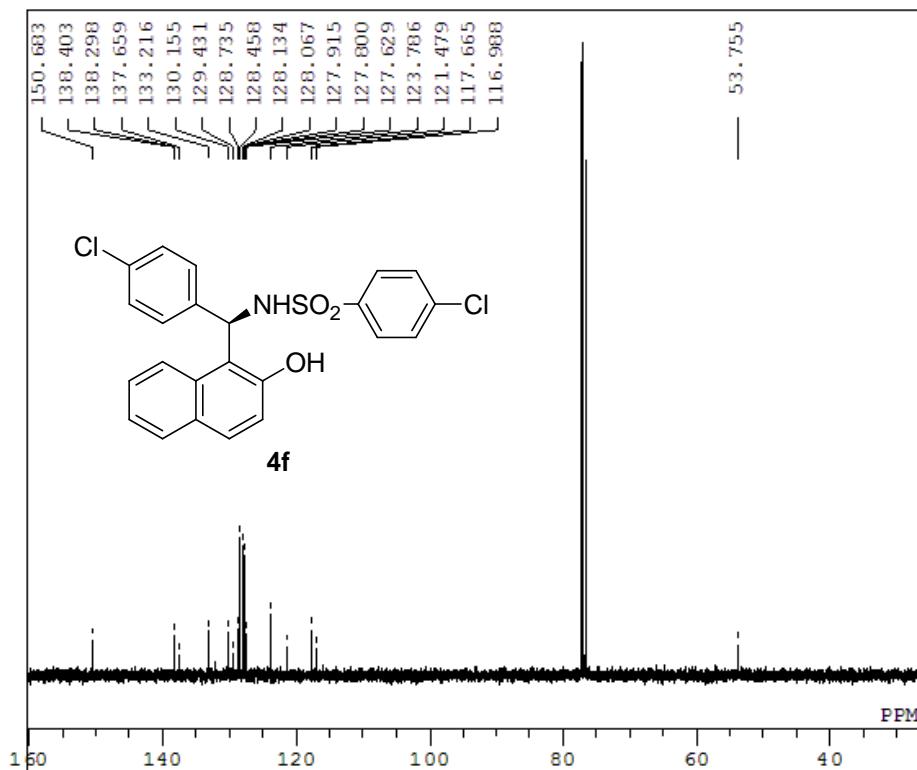
DFILE 8-54_Carbon-1-1.:
 COMNT single pulse dec:
 DATIM 2014-05-24 17:04:
 OBNUC 13C
 EXMOD carbon.jxp
 OBFRQ 100.53 MHz
 OBSET 5.35 KHz
 OBFIN 5.86 Hz
 POINT 32767
 FREQU 31407.04 Hz
 SCANS 300
 ACQTM 1.0433 sec
 PD 2.0000 sec
 PW1 3.20 usec
 IRNUC 1H
 CTEMP 24.0 c
 SLVNT CDCL3
 EXREF 77.00 ppm
 BF 0.10 Hz
 RGAIN 60



```

DFILE sh4030a-gpc_Proto
COMNT single_pulse
DATIM 2012-11-20 21:02:
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 16384
FREQU 7503.00 Hz
SCANS 8
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 5.50 usec
IRNUC 1H
CTEMP 20.6 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.10 Hz
RGAIN 48

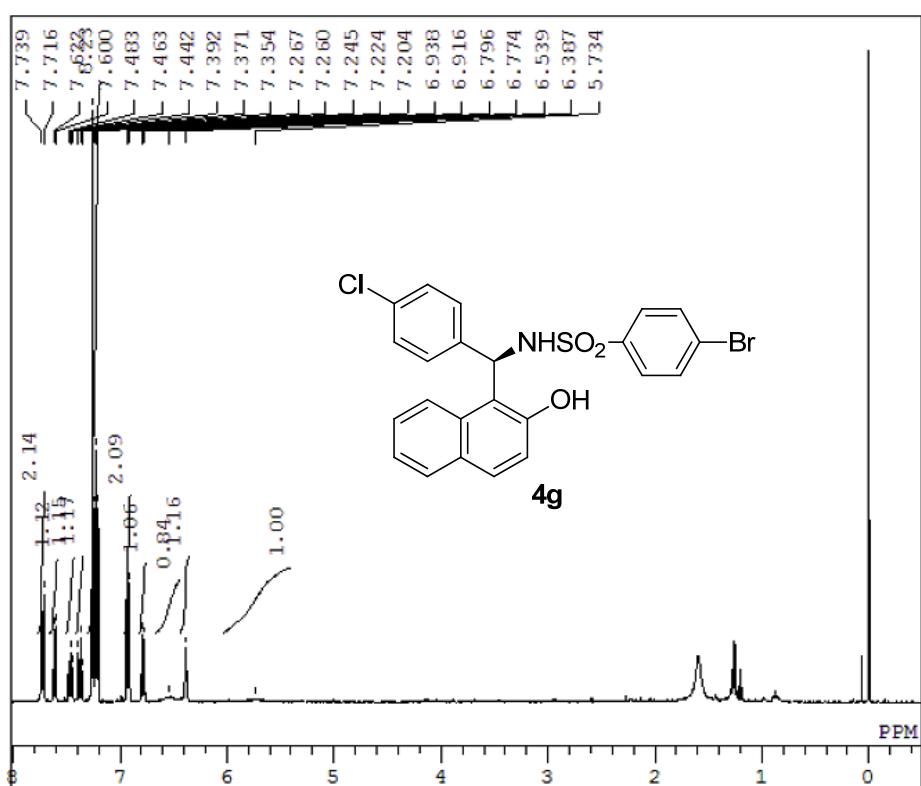
```



```

DFILE 8-53_Carbon-1-1.:
COMNT single pulse decoupled
DATIM 2014-05-26 10:44:
OBNUC 13C
EXMOD carbon.jxp
OBFRQ 100.53 MHz
OBSET 5.35 KHz
OBFIN 5.86 Hz
POINT 32767
FREQU 31407.04 Hz
SCANS 300
ACQTM 1.0433 sec
PD 2.0000 sec
PW1 3.20 usec
IRNUC 1H
CTEMP 19.0 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 60

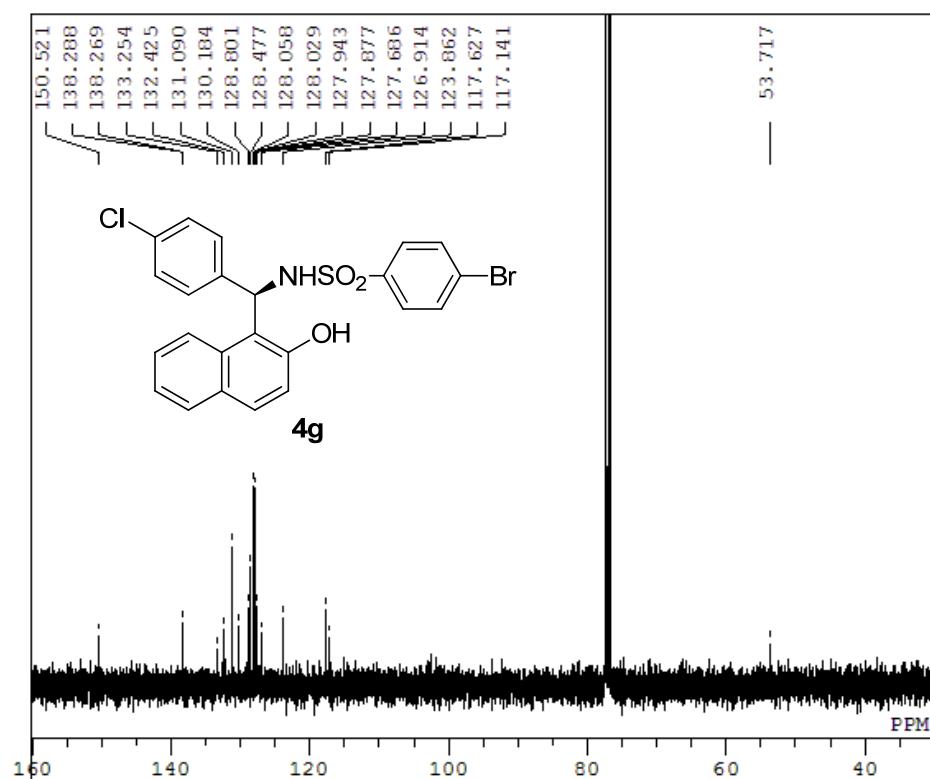
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```

DFILE sh4040-gpc22_Pro1
COMNT single_pulse
DATIM 2012-12-08 15:16:
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 16384
FREQU 7503.00 Hz
SCANS 8
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 6.05 usec
IRNUC 1H
CTEMP 23.0 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 50

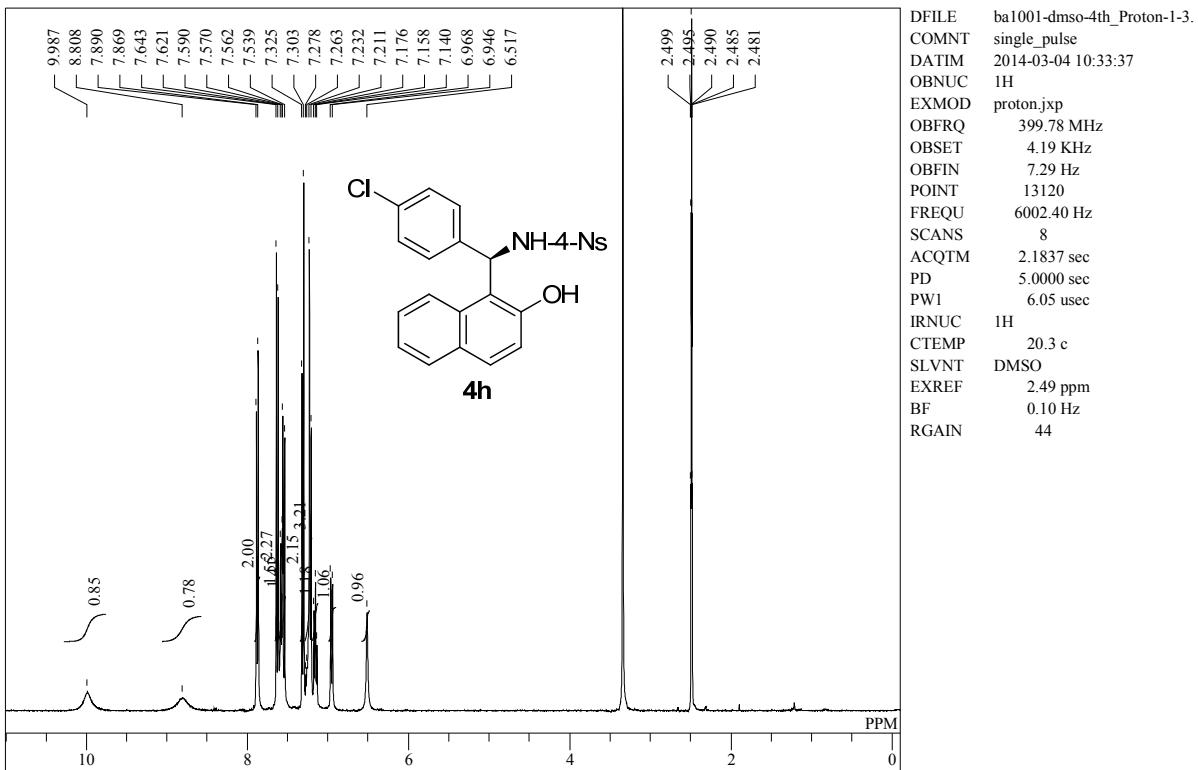
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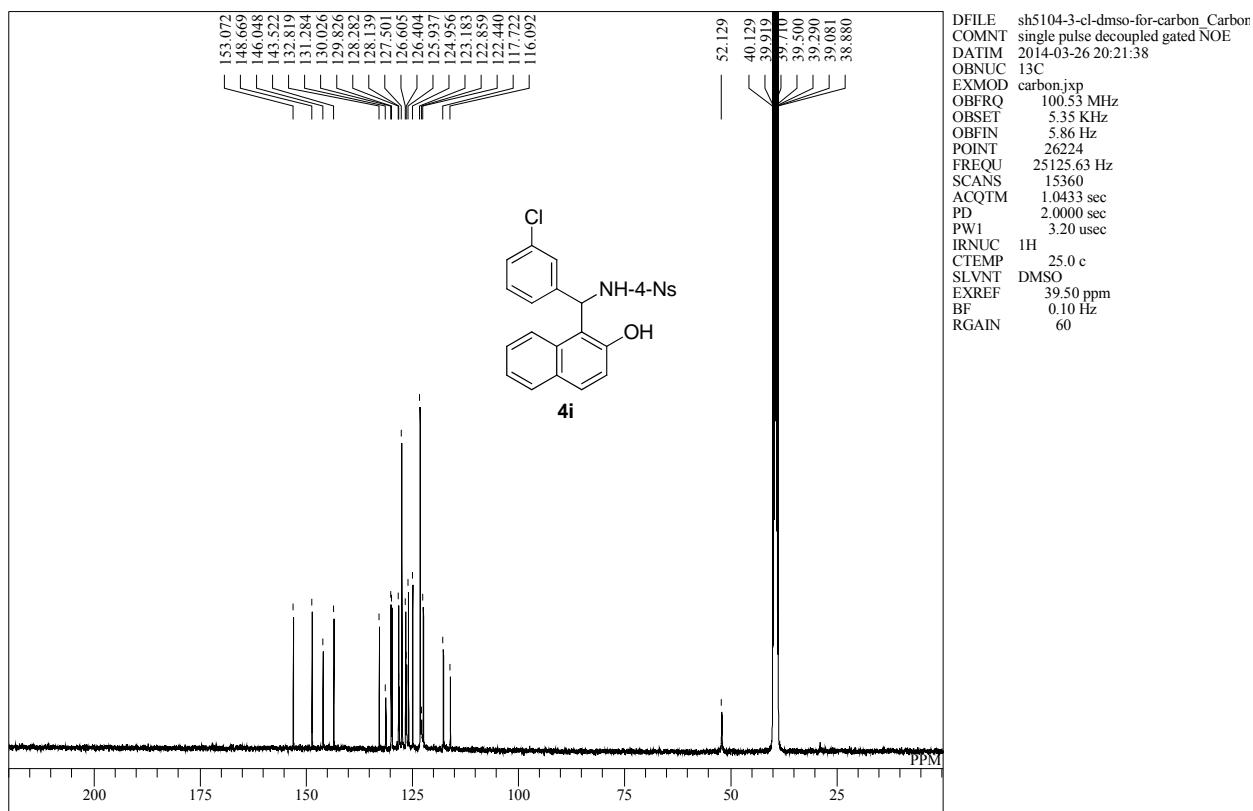
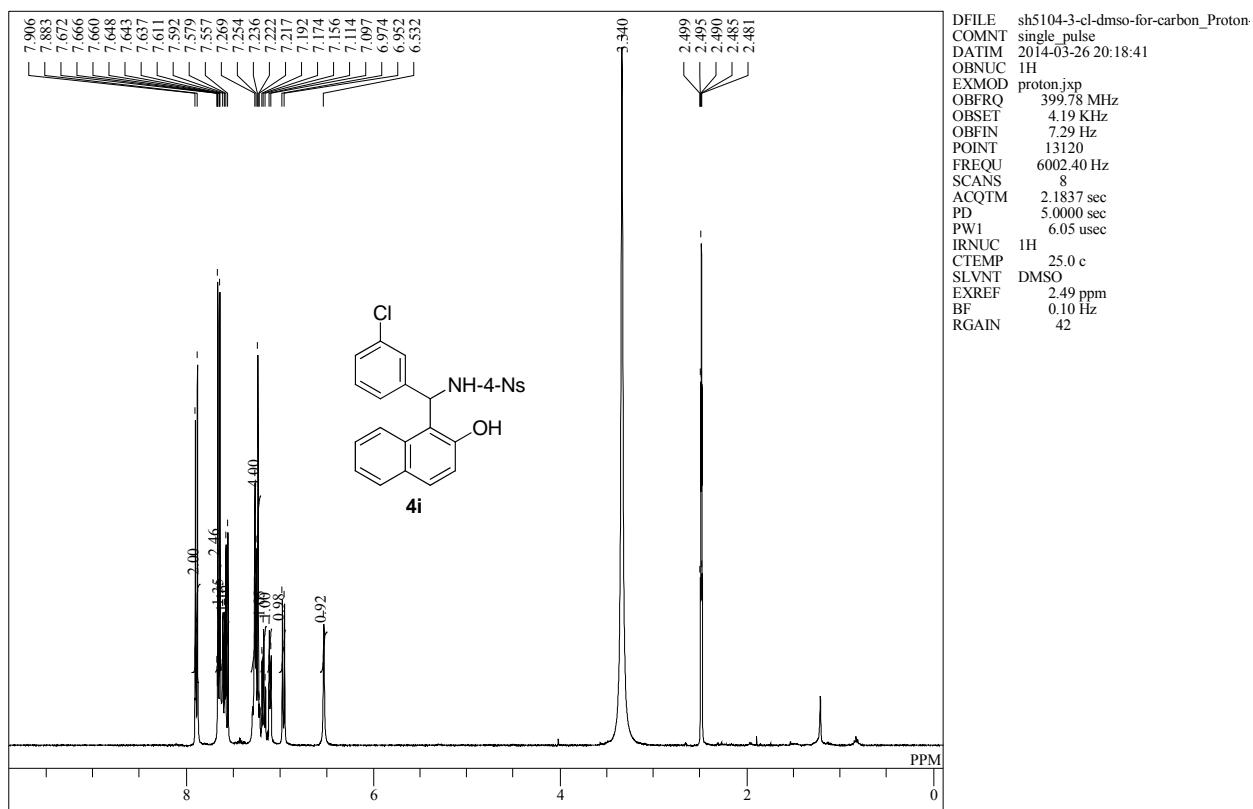


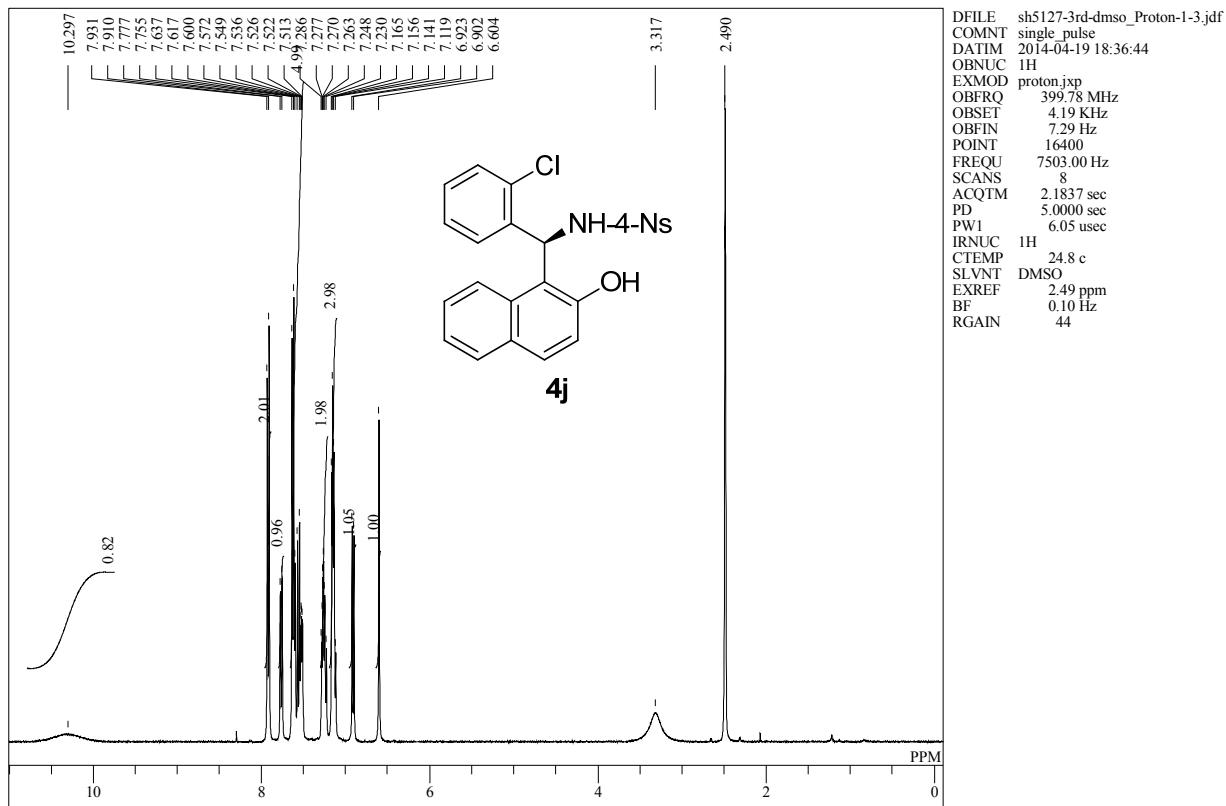
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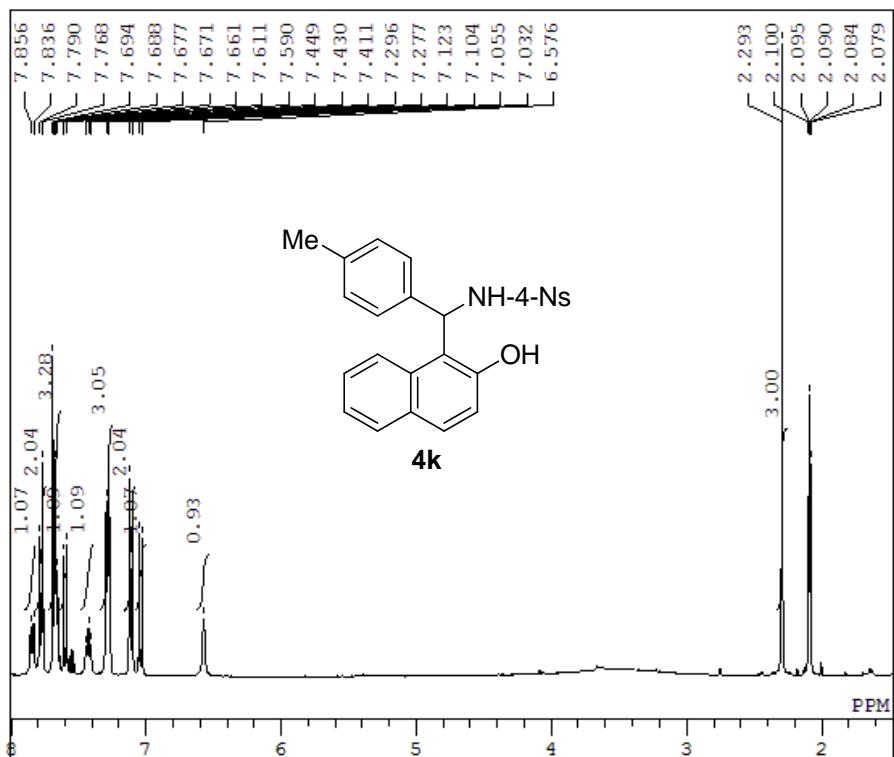
DFILE 8-58_Carbon-1-1.:
COMNT single pulse dec
DATIM 2014-05-26 11:09:
OBNUC 13C
EXMOD carbon.jxp
OBFRQ 100.53 MHz
OBSET 5.35 KHz
OBFIN 5.86 Hz
POINT 32767
FREQU 31407.04 Hz
SCANS 350
ACQTM 1.0433 sec
PD 2.0000 sec
PW1 3.20 usec
IRNUC 1H
CTEMP 19.4 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.12 Hz
RGAIN 50

```





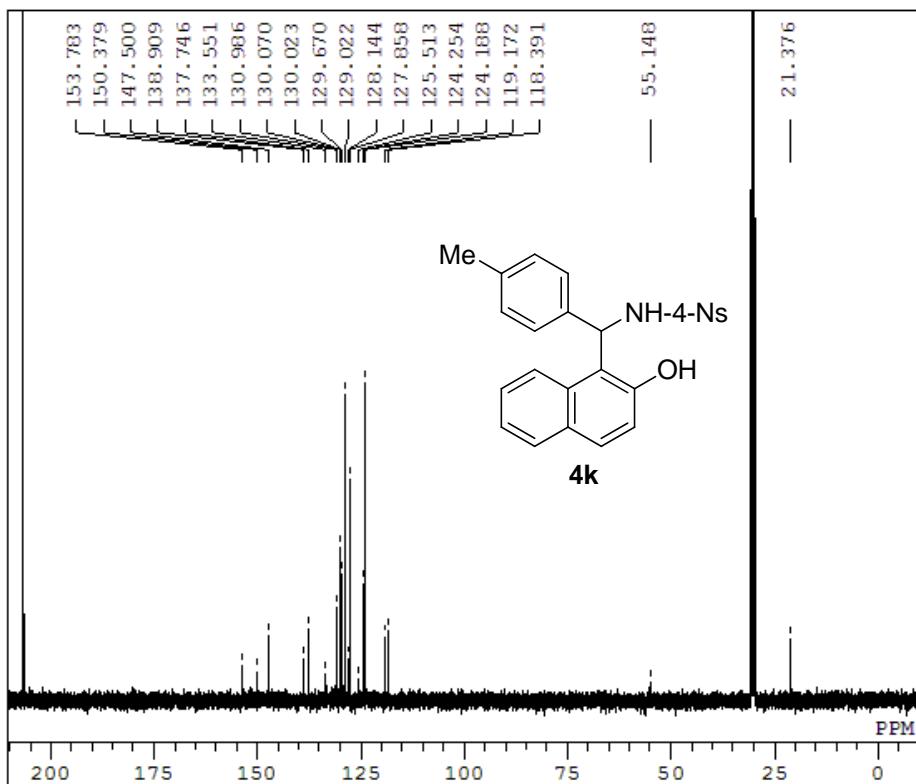


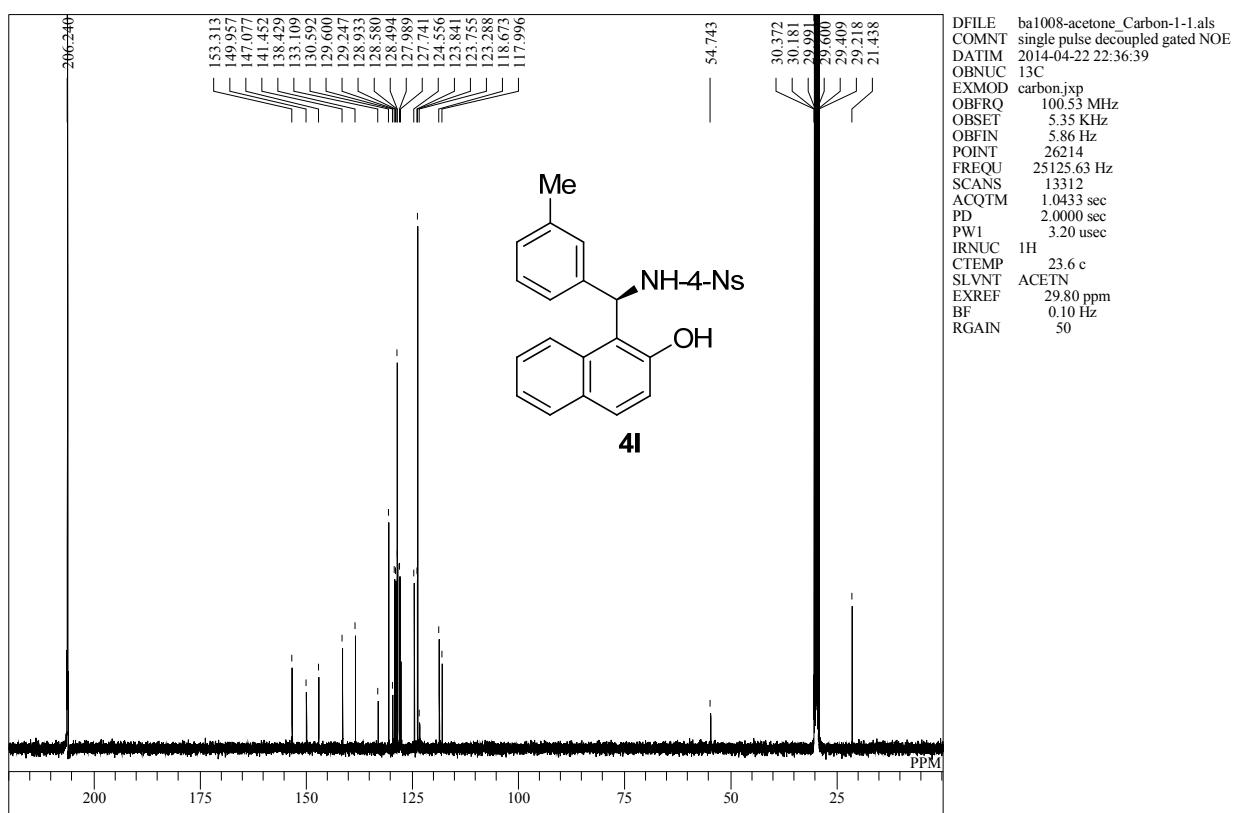
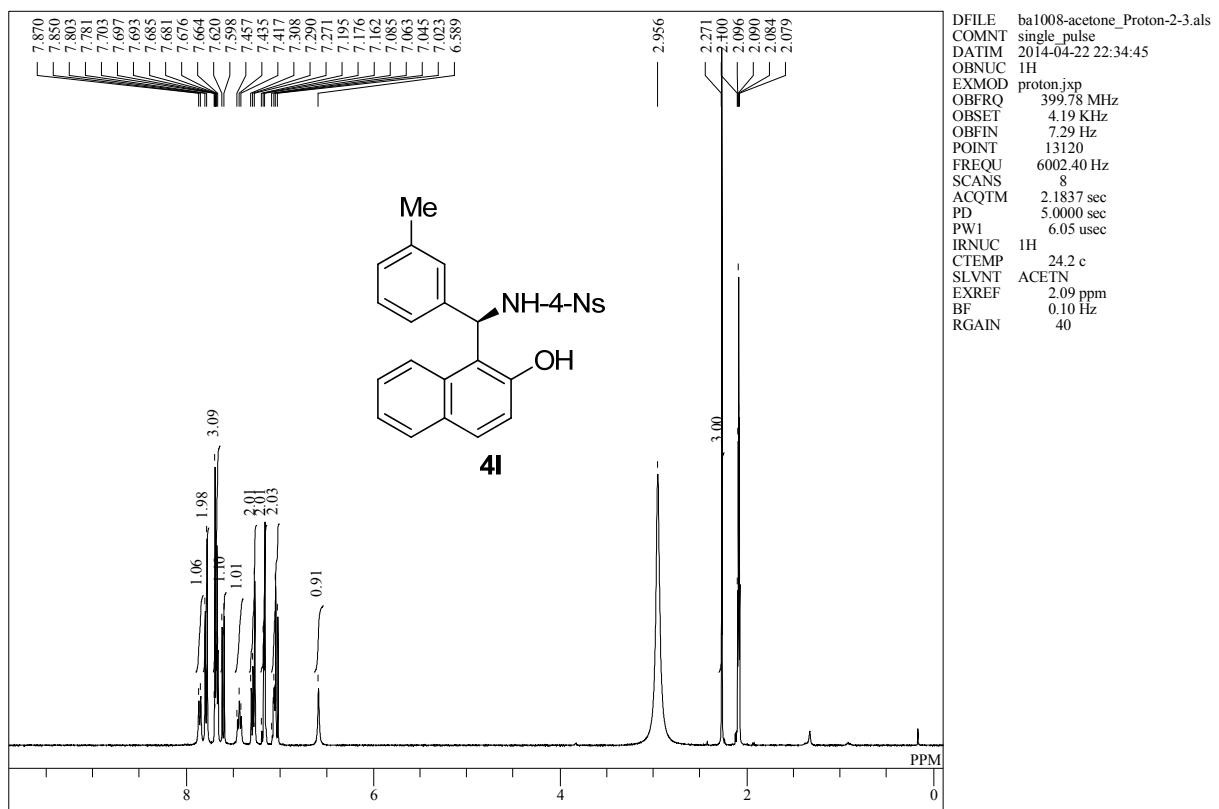


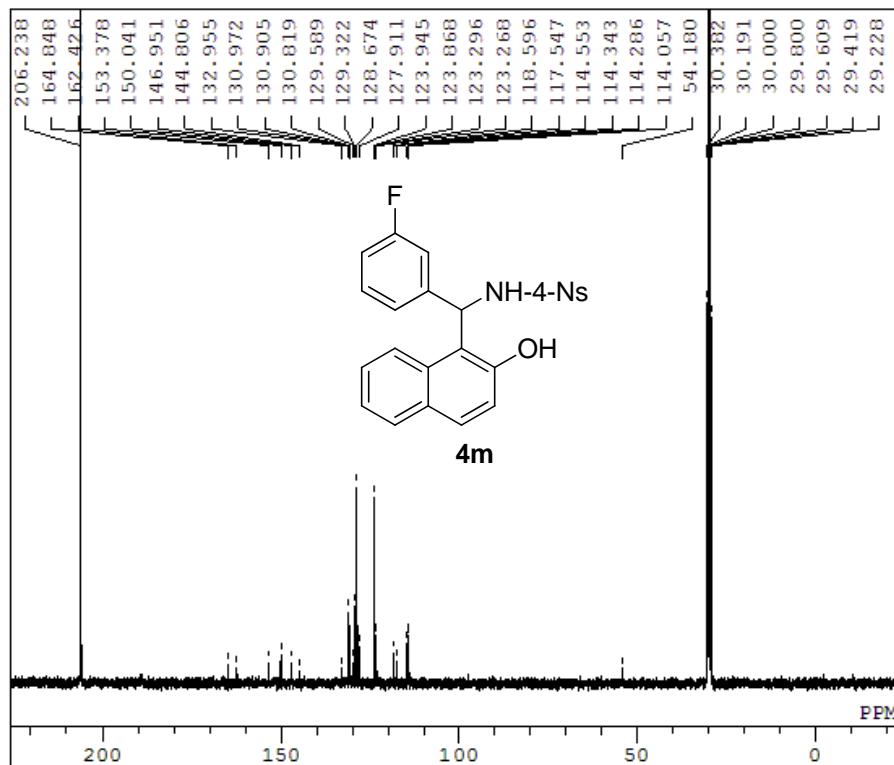
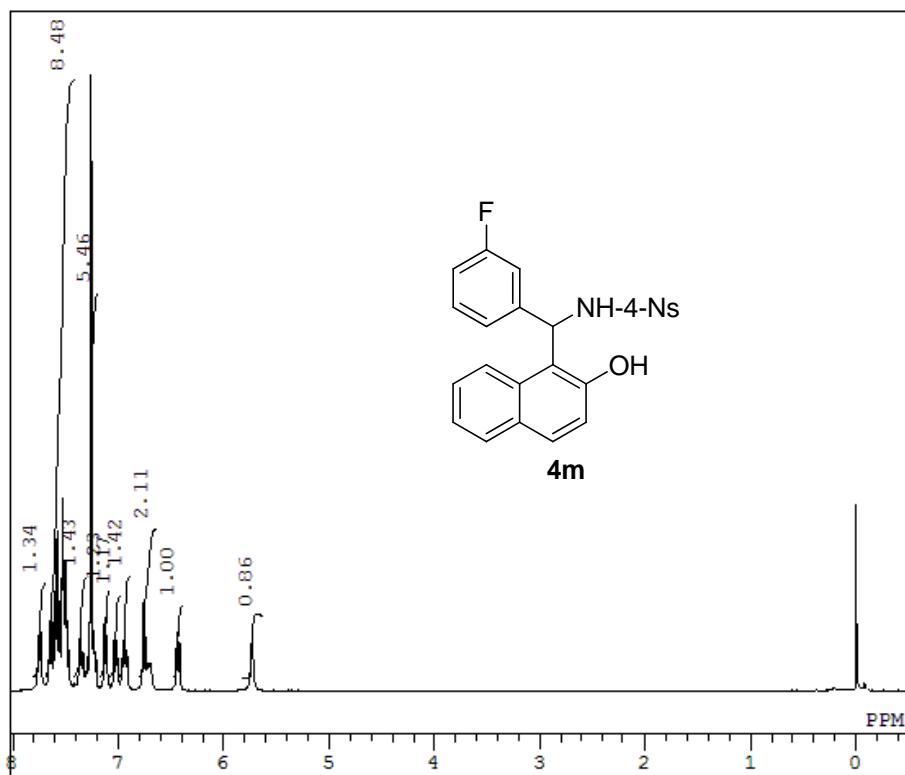
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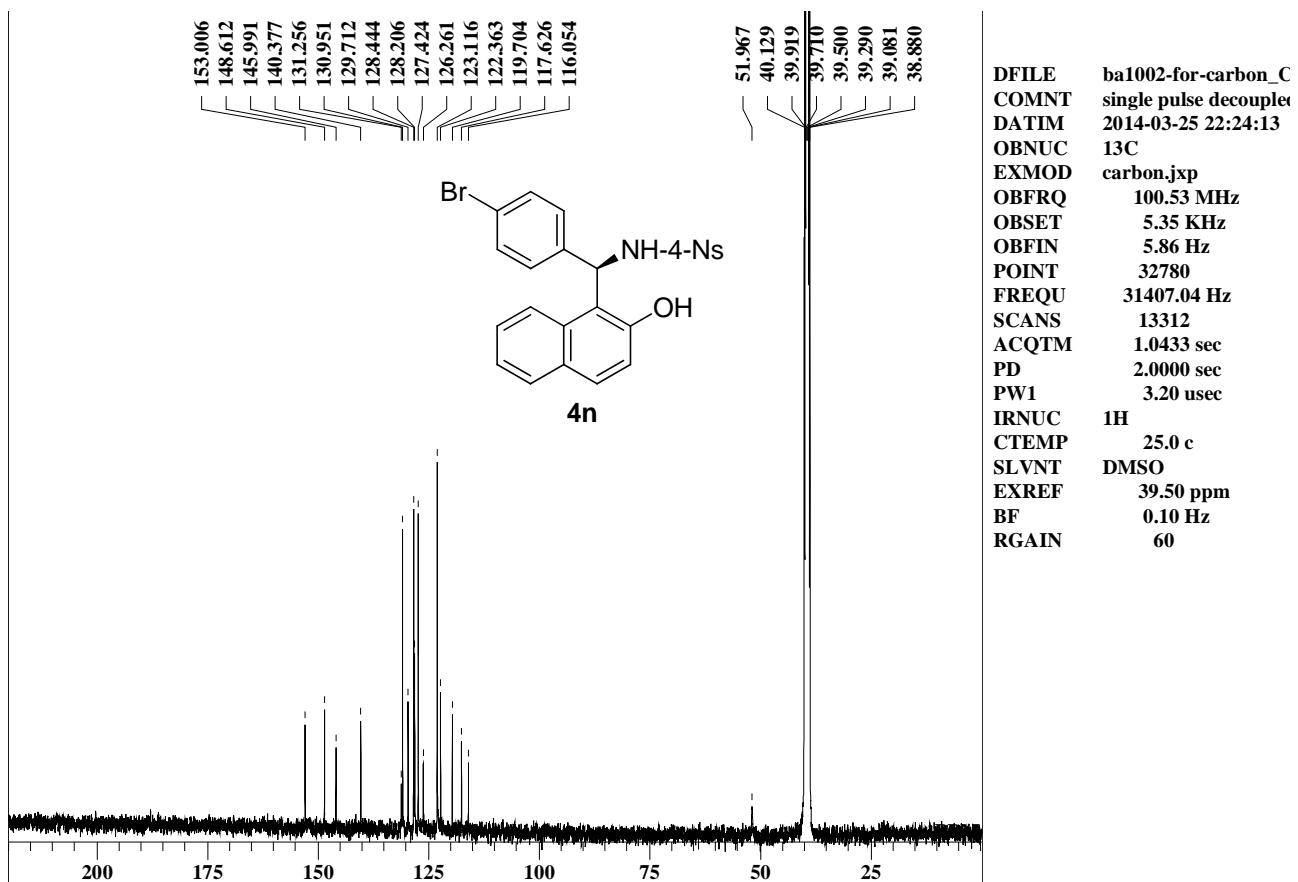
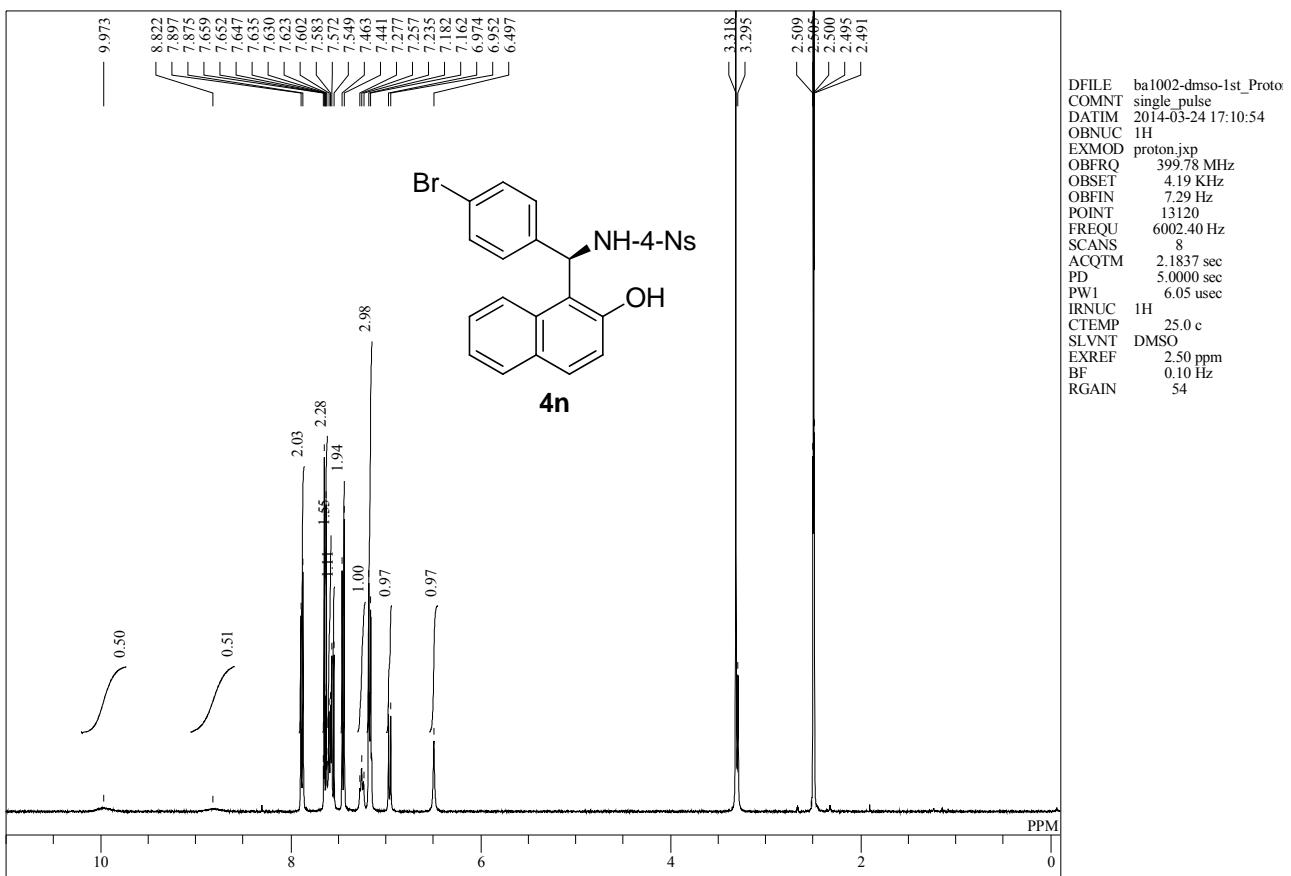
DFILE 8-141_Proton-1-1.
COMNT single_pulse
DATIM 2014-04-19 18:00:
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 16384
FREQU 7503.00 Hz
SCANS 8
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 6.05 usec
IRNUC 1H
CTEMP 24.9 c
SLVNT ACETN
EXREF 2.09 ppm
BF 0.12 Hz
RGAIN 36

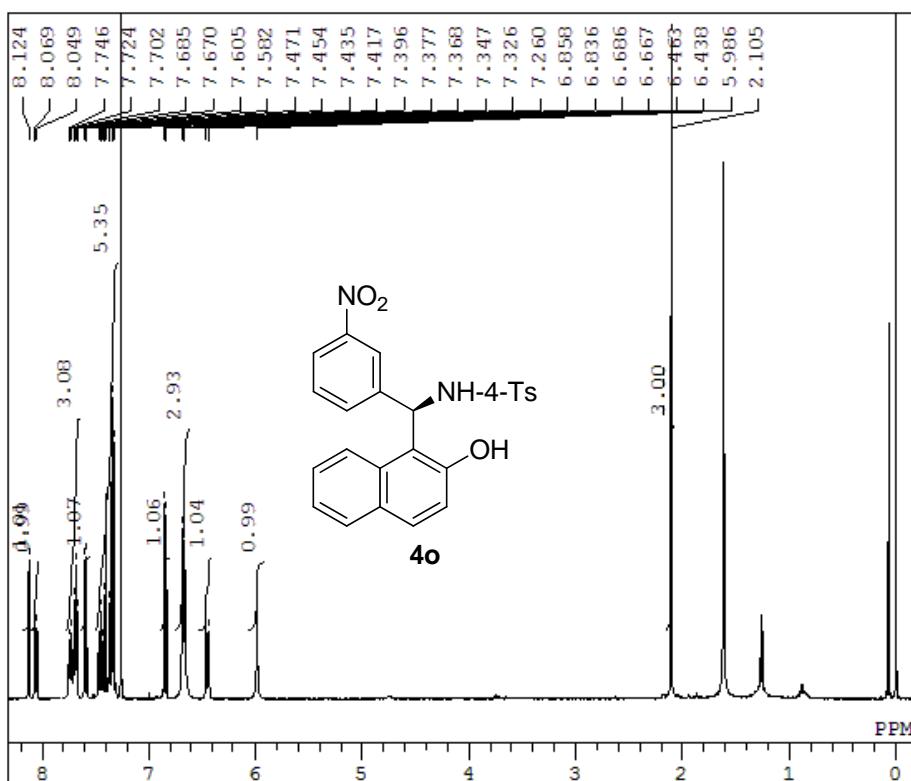
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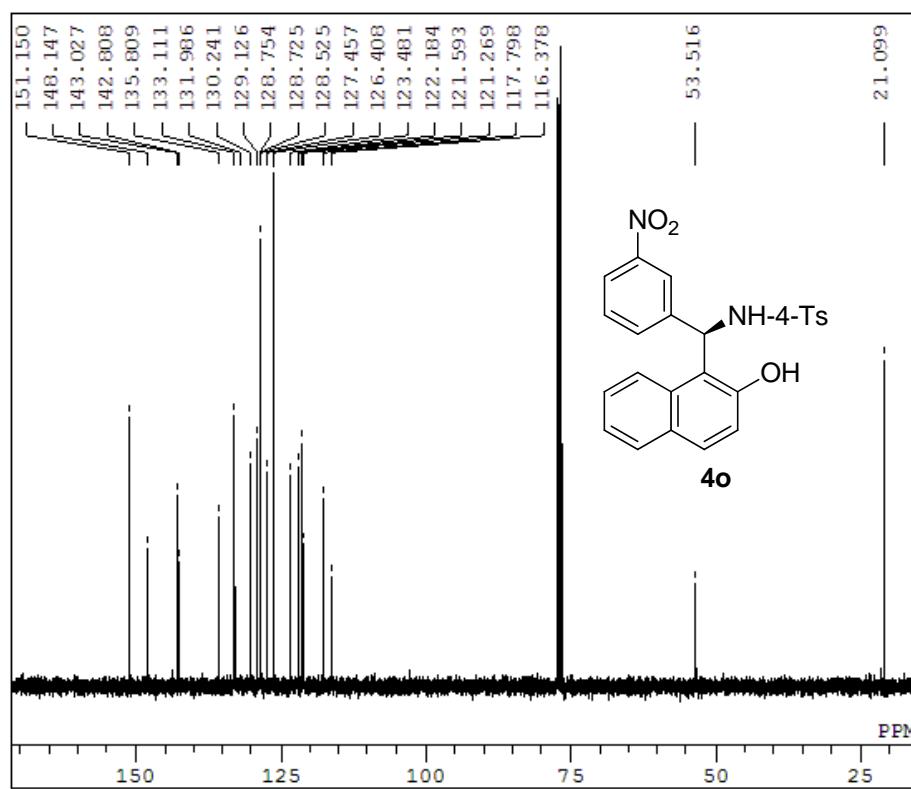




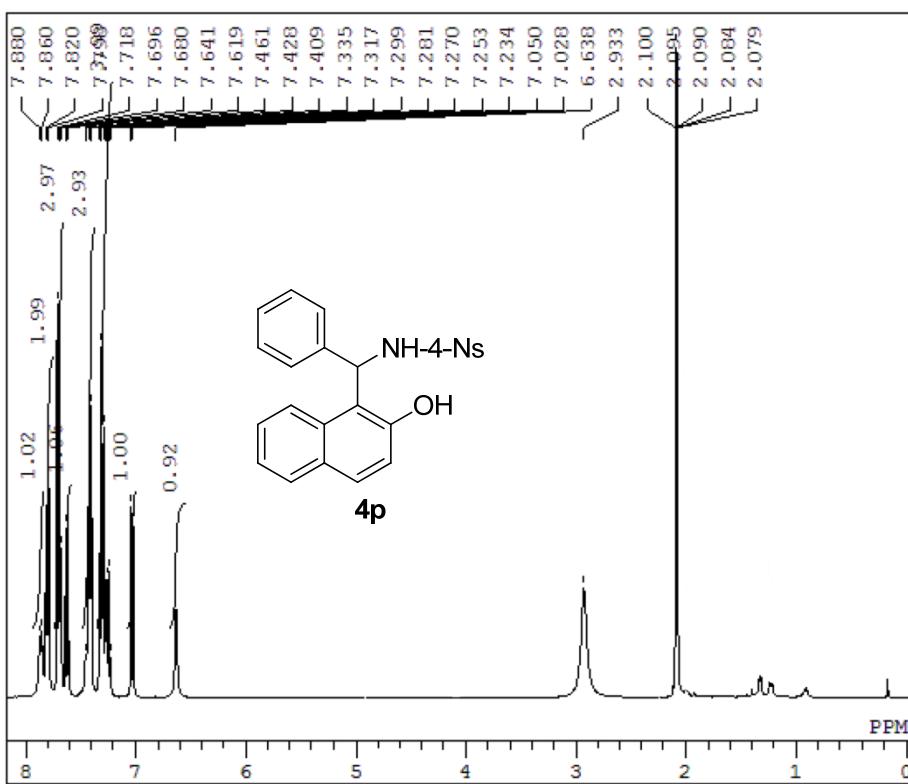




DFILE 8-100.als
 COMNT single_pulse
 DATIM 2013-06-25 15:28:
 OBNUC 1H
 EXMOD proton.jxp
 OBFRQ 399.78 MHz
 OBSET 4.19 KHz
 OBFIN 7.29 Hz
 POINT 13107
 FREQU 6002.40 Hz
 SCANS 8
 ACQTM 2.1837 sec
 PD 5.0000 sec
 PW1 6.05 usec
 IRNUC 1H
 CTEMP 22.1 c
 SLVNT CDCL₃
 EXREF 7.26 ppm
 BF 0.12 Hz
 RGAIN 50



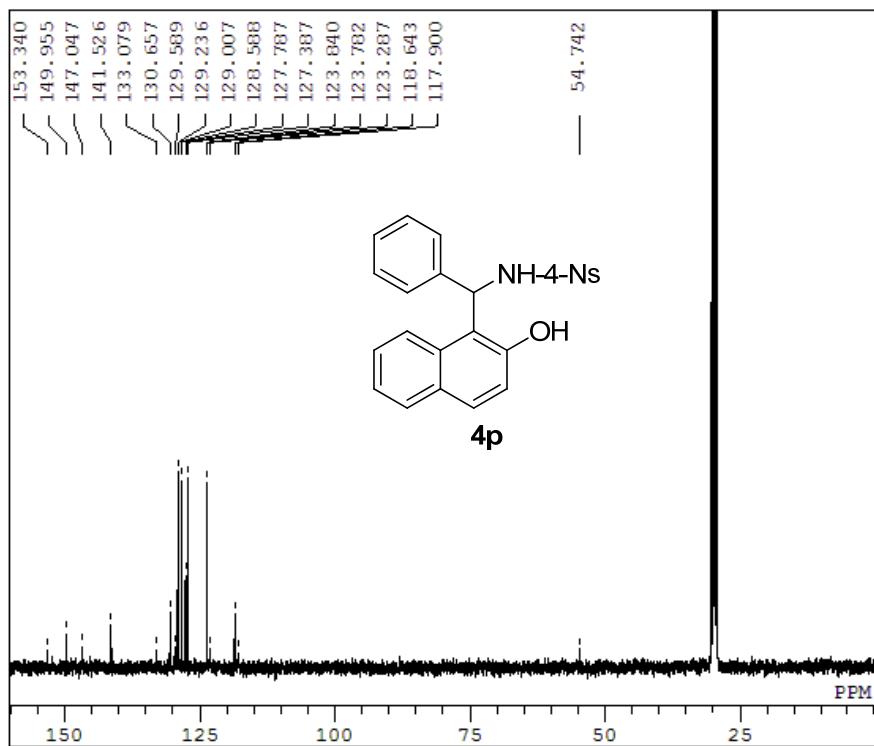
DFILE 8-166-1_Carbon-1-
 COMNT single pulse decoupled
 DATIM 2014-06-13 16:23:
 OBNUC 13C
 EXMOD carbon.jxp
 OBFRQ 100.53 MHz
 OBSET 5.35 KHz
 OBFIN 5.86 Hz
 POINT 32767
 FREQU 31407.04 Hz
 SCANS 200
 ACQTM 1.0433 sec
 PD 2.0000 sec
 PW1 3.20 usec
 IRNUC 1H
 CTEMP 20.7 c
 SLVNT CDCL₃
 EXREF 77.00 ppm
 BF 0.12 Hz
 RGAIN 50



```

DFILE Ph_Proton-1-1.jdi
COMNT single_pulse
DATIM 2014-03-05 19:05:
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 16384
FREQU 7503.00 Hz
SCANS 16
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 6.05 usec
IRNUC 1H
CTEMP 21.7 c
SLVNT ACETN
EXREF 2.09 ppm
BF 1.20 Hz
RGAIN 46

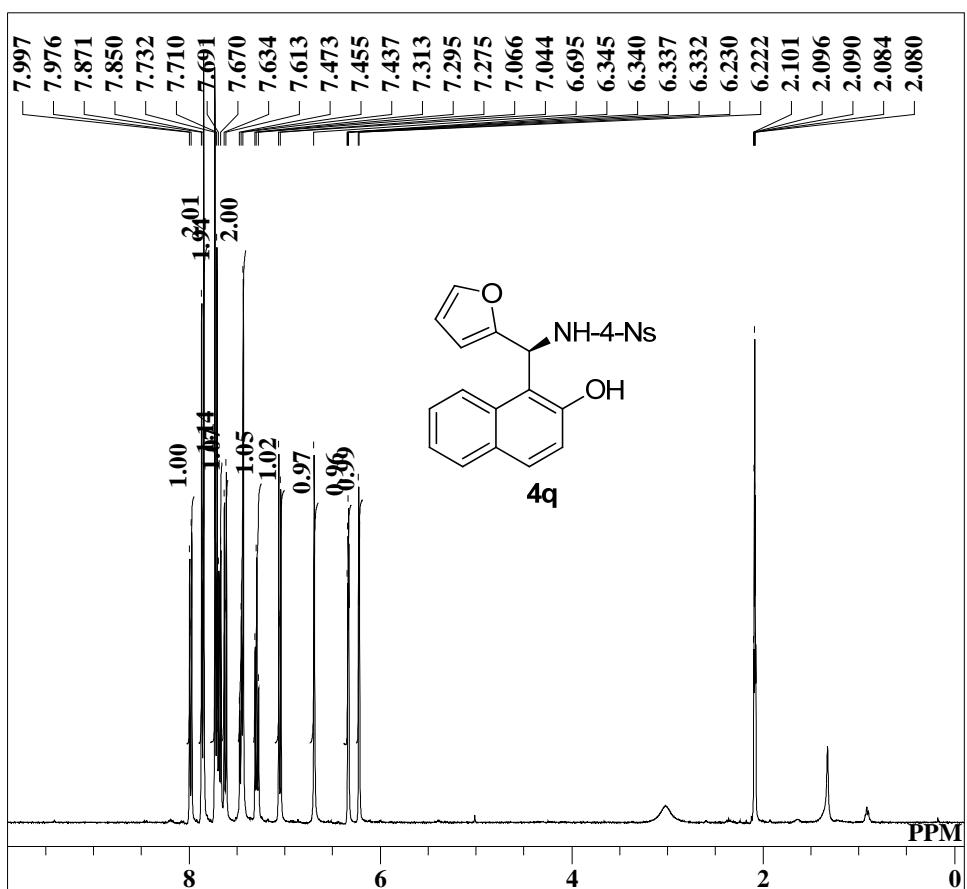
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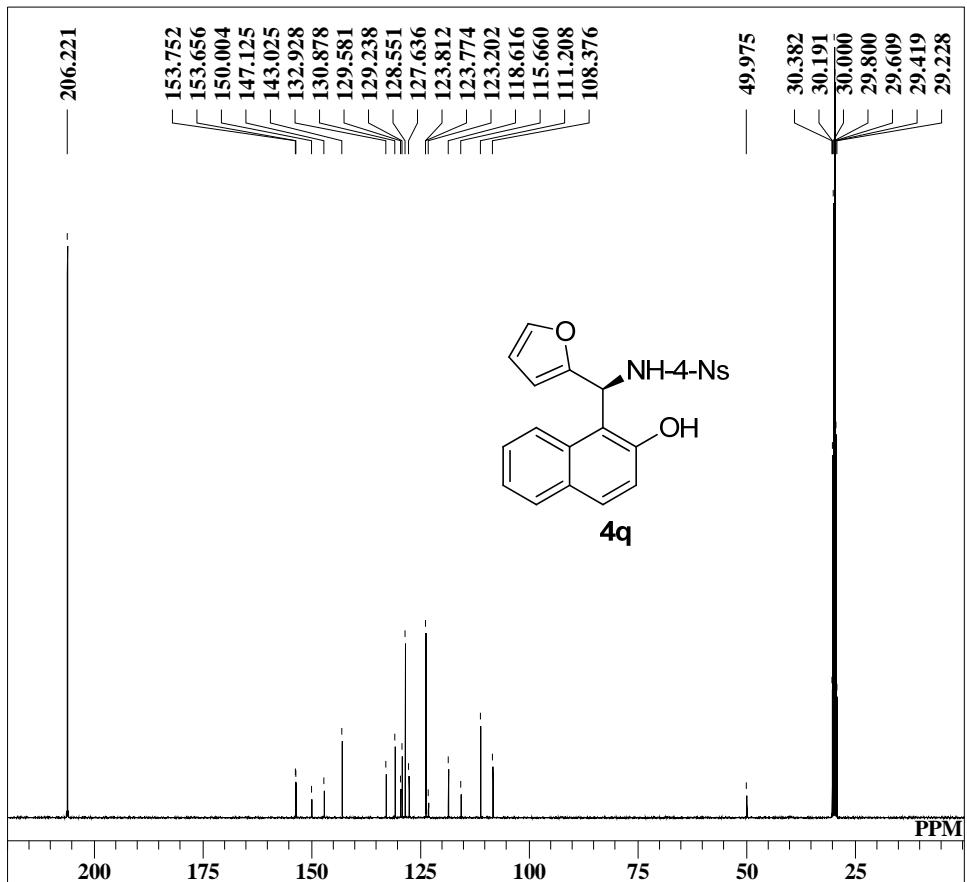
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DFILE 8-127_Carbon-1-1.jdi
COMNT single_pulse dec
DATIM 2014-03-14 11:56:
OBNUC 13C
EXMOD carbon.jxp
OBFRQ 100.53 MHz
OBSET 5.35 KHz
OBFIN 5.86 Hz
POINT 32767
FREQU 31407.04 Hz
SCANS 512
ACQTM 1.0433 sec
PD 2.0000 sec
PW1 3.20 usec
IRNUC 1H
CTEMP 21.9 c
SLVNT ACETN
EXREF 29.80 ppm
BF 1.20 Hz
RGAIN 60

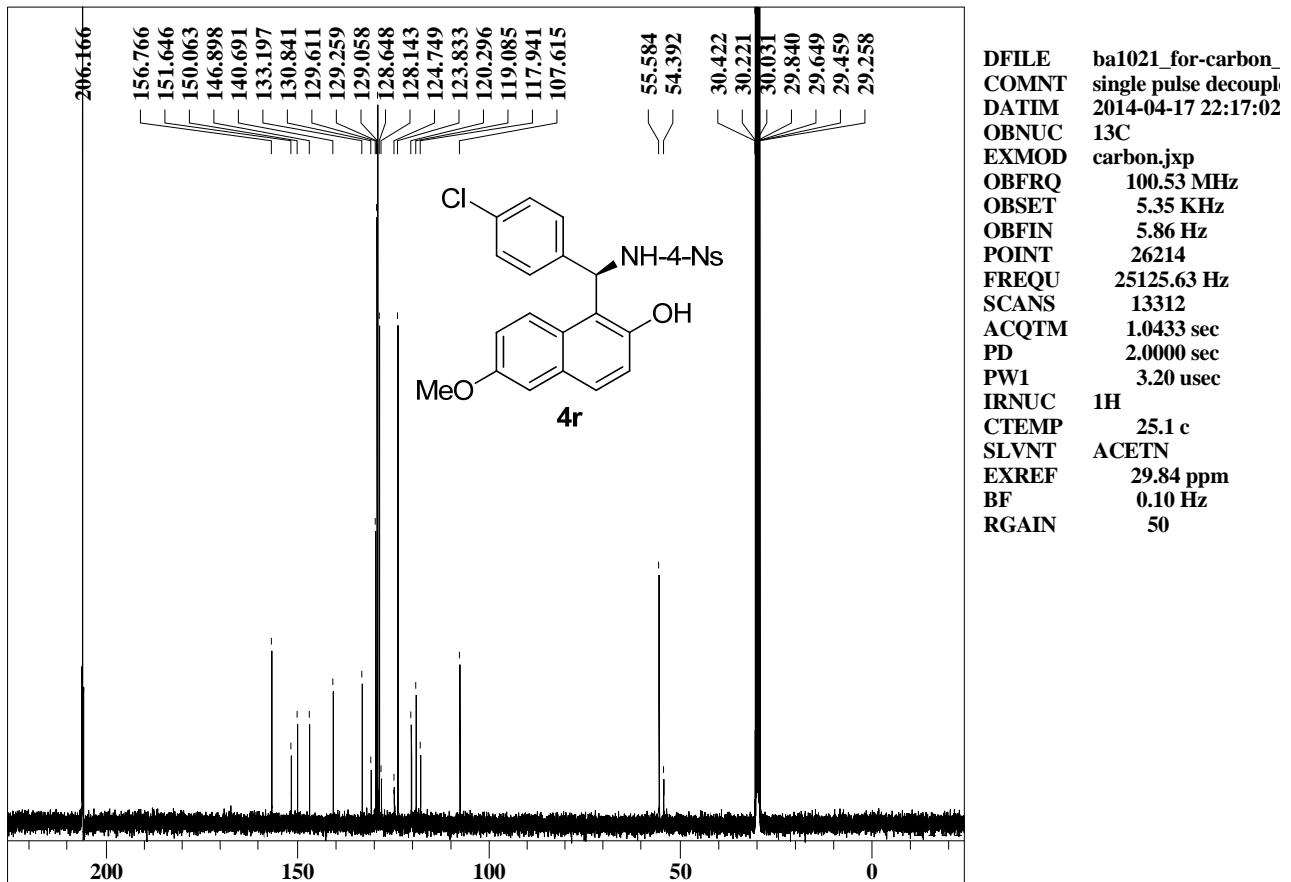
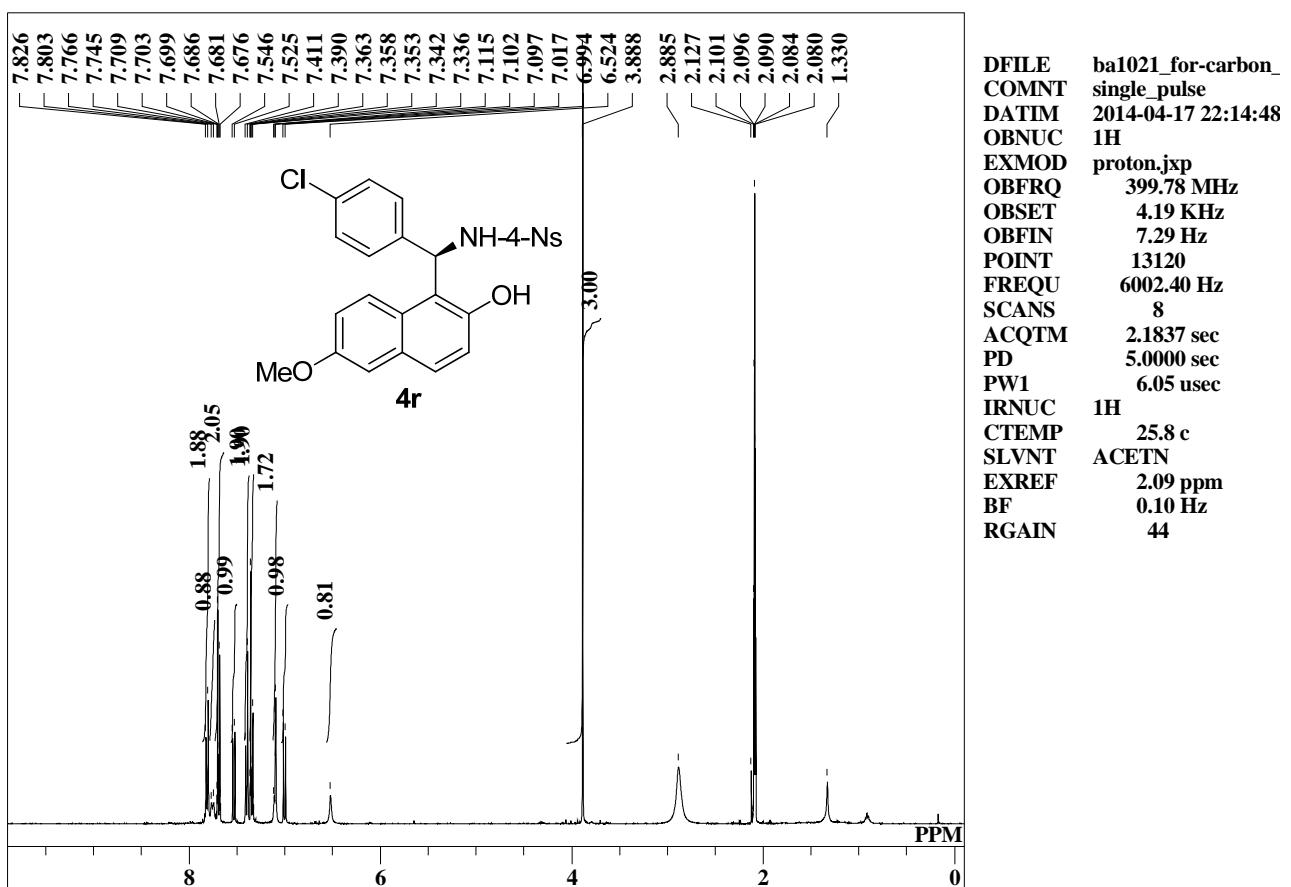
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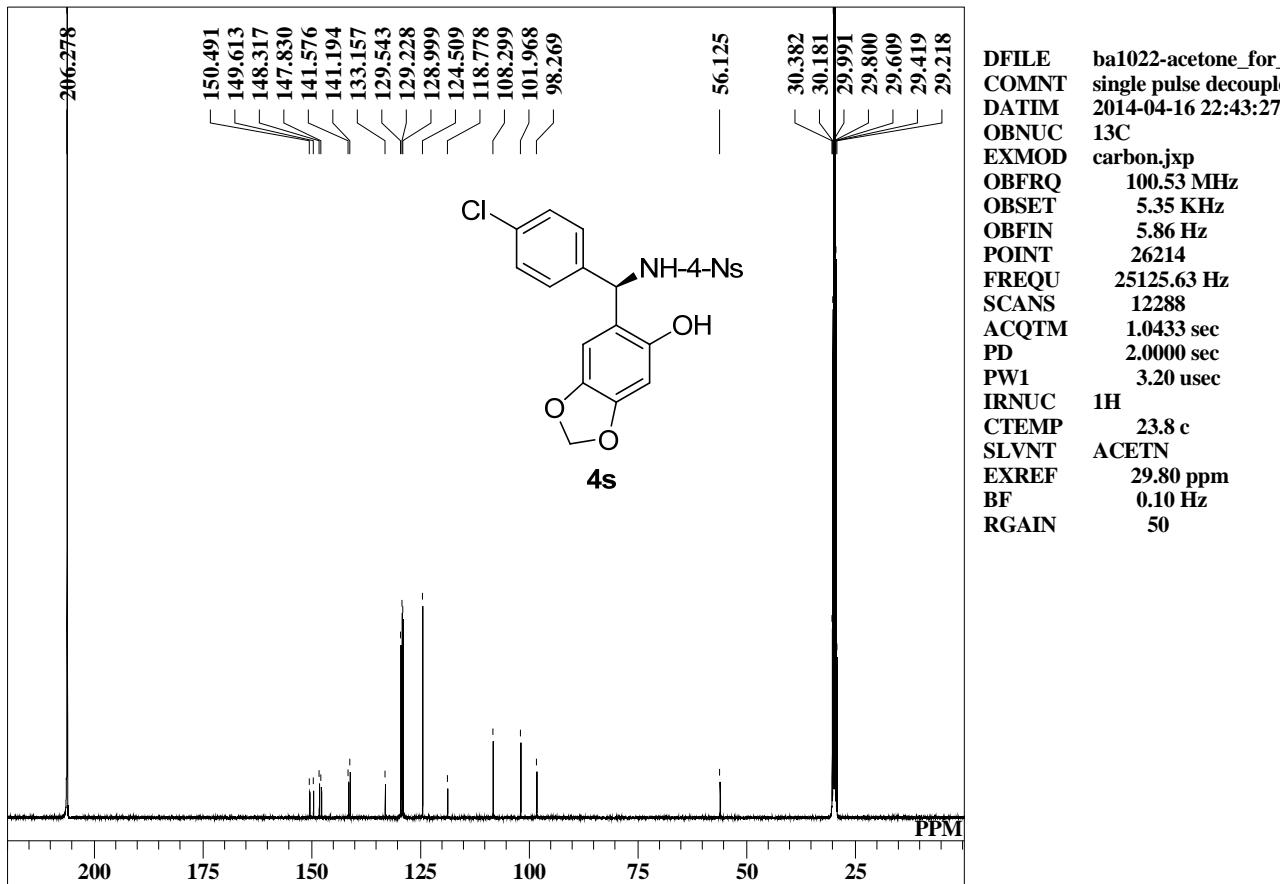
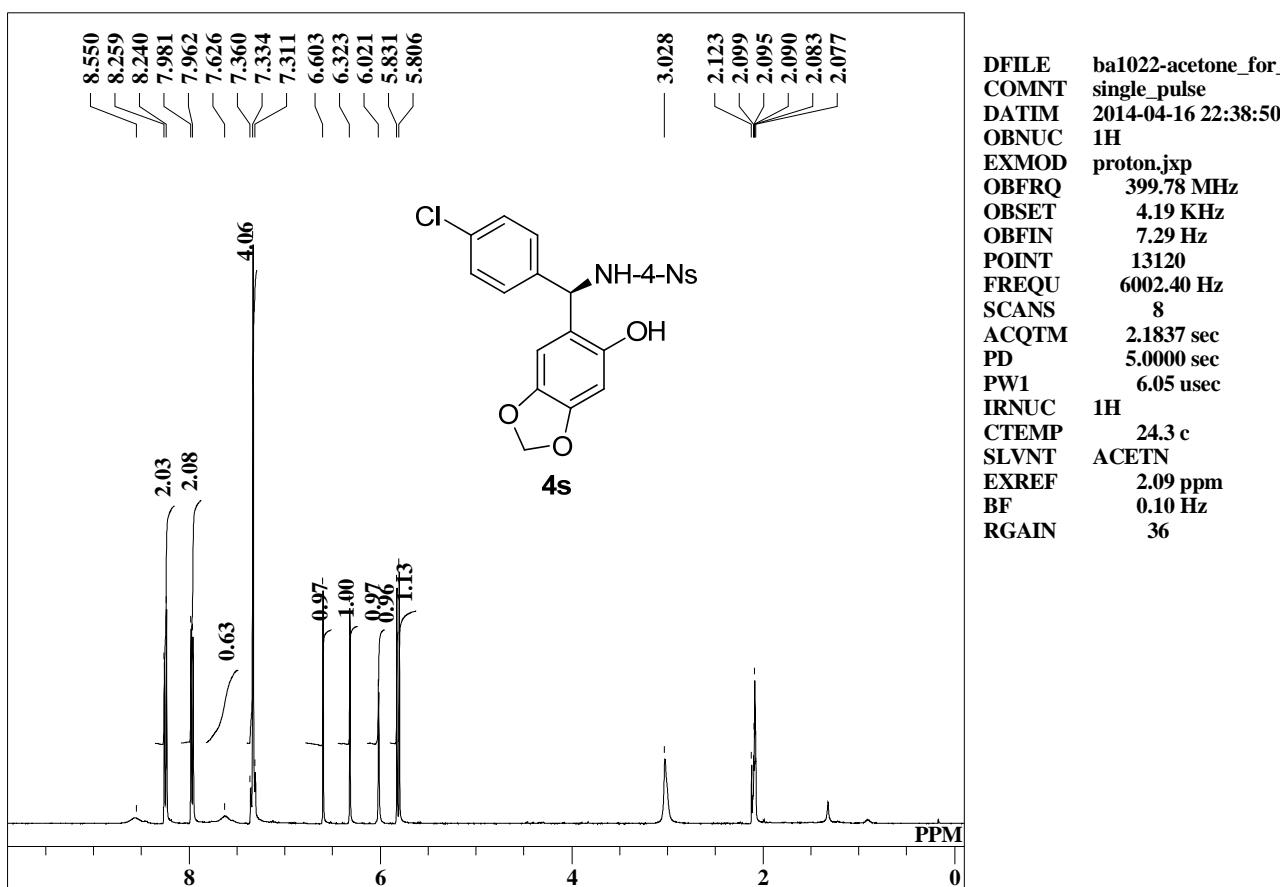


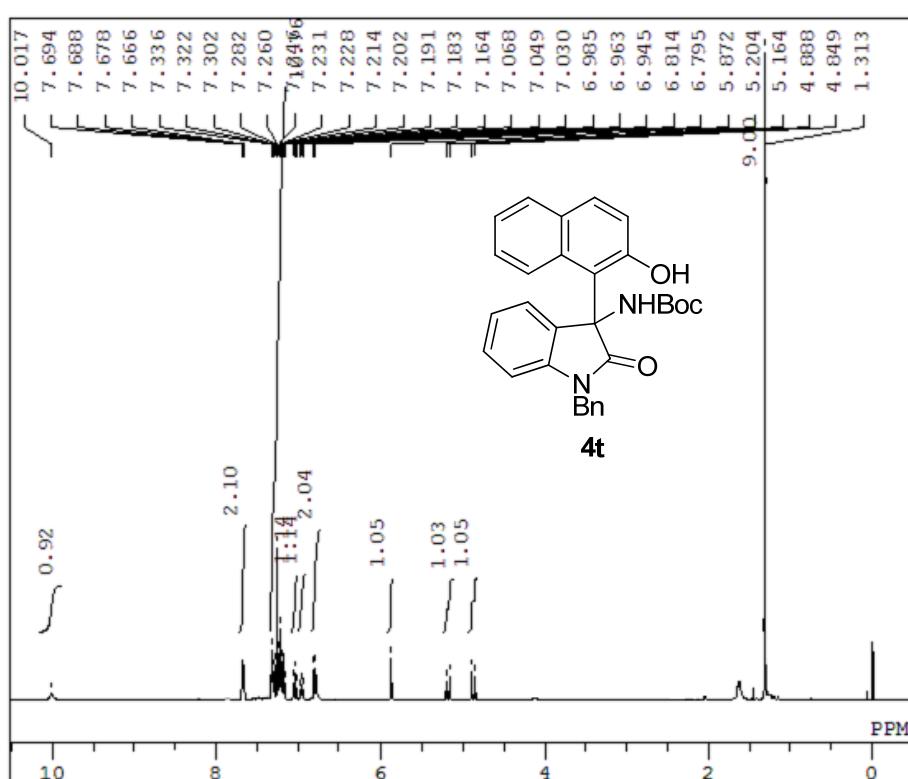
DFILE ba1007-acetone
COMNT single_pulse
DATIM 2014-04-14 21:5
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 13120
FREQU 6002.40 Hz
SCANS 8
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 6.05 usec
IRNUC 1H
CTEMP 23.3 c
SLVNT ACETN
EXREF 2.09 ppm
BF 0.10 Hz
RGAIN 40



DFILE ba1007-acetone3_C₂
COMNT single pulse decoupl
DATIM 2014-04-14 21:57:51
OBNUC 13C
EXMOD carbon.jxp
OBFRQ 100.53 MHz
OBSET 5.35 KHz
OBFIN 5.86 Hz
POINT 26214
FREQU 25125.63 Hz
SCANS 13312
ACQTM 1.0433 sec
PD 2.0000 sec
PW1 3.20 usec
IRNUC 1H
CTEMP 21.5 c
SLVNT ACETN
EXREF 29.80 ppm
BF 0.10 Hz
RGAIN 50



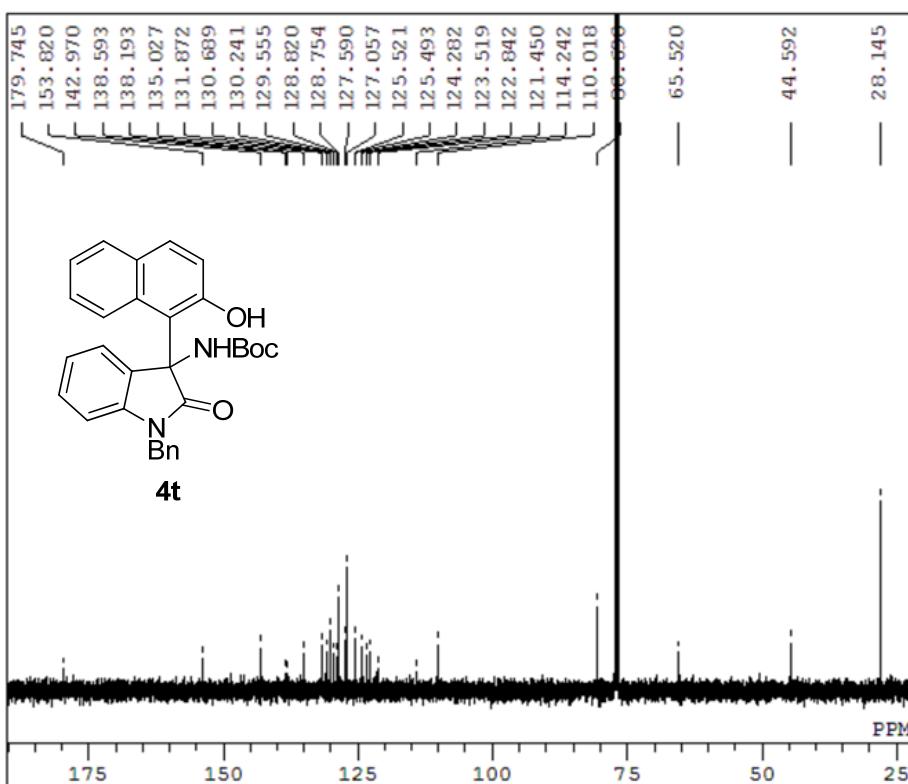


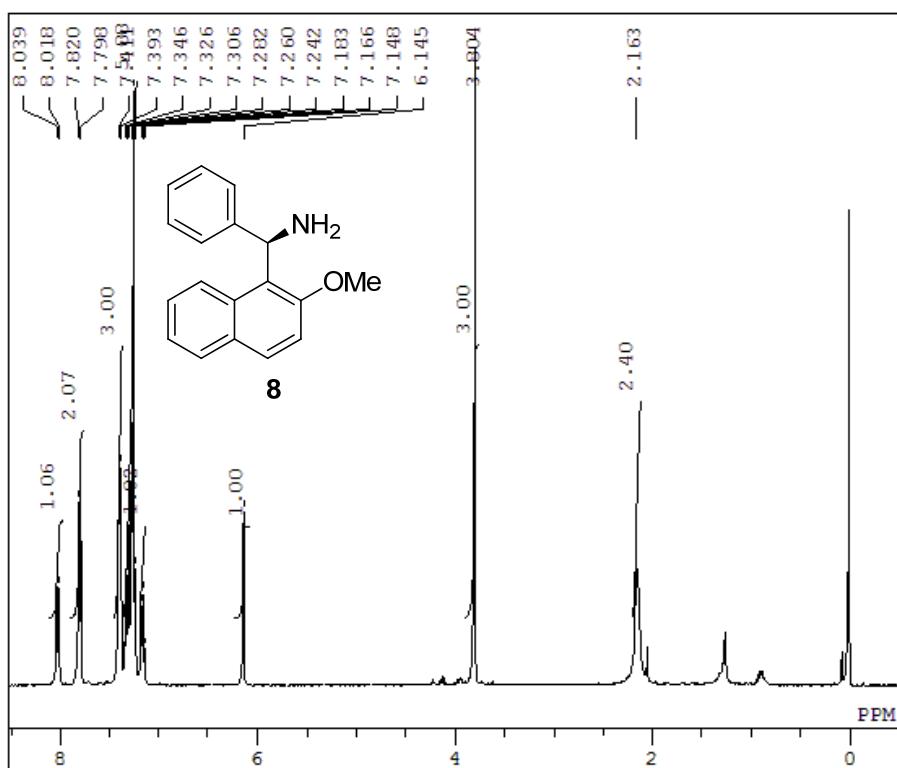


```

DFILE 8-158-A_Proton-1-
COMNT single_pulse
DATIM 2014-05-27 15:11:
OBNUC 1H
EXMOD proton.jxp
OBFRQ      399.78 MHz
OBSET       4.19 KHz
OBFIN        7.29 Hz
POINT      16384
FREQU     7503.00 Hz
SCANS         8
ACQTM      2.1837 sec
PD          5.0000 sec
PW1          6.05 usec
IRNUC    1H
CTEMP      19.0 c
SLVNT    CDCL3
EXREF      7.26 ppm
BF          0.12 Hz
RGAIN         46

```

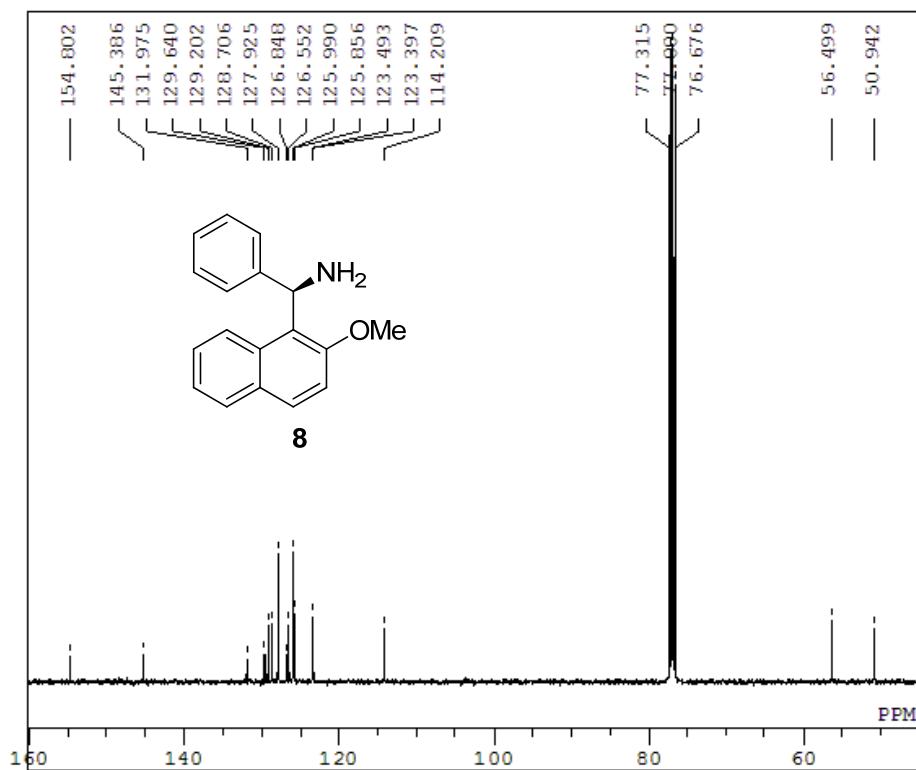




```

DFILE sh5166-column_Proton
COMNT single_pulse
DATIM 2014-06-14 15:56:
OBNUC 1H
EXMOD proton.jxp
OBFRQ 399.78 MHz
OBSET 4.19 KHz
OBFIN 7.29 Hz
POINT 16400
FREQU 7503.00 Hz
SCANS 8
ACQTM 2.1837 sec
PD 5.0000 sec
PW1 5.50 usec
IRNUC 1H
CTEMP 19.5 c
SLVNT CDCL3
EXREF 7.26 ppm
BF 0.12 Hz
RGAIN 46

```

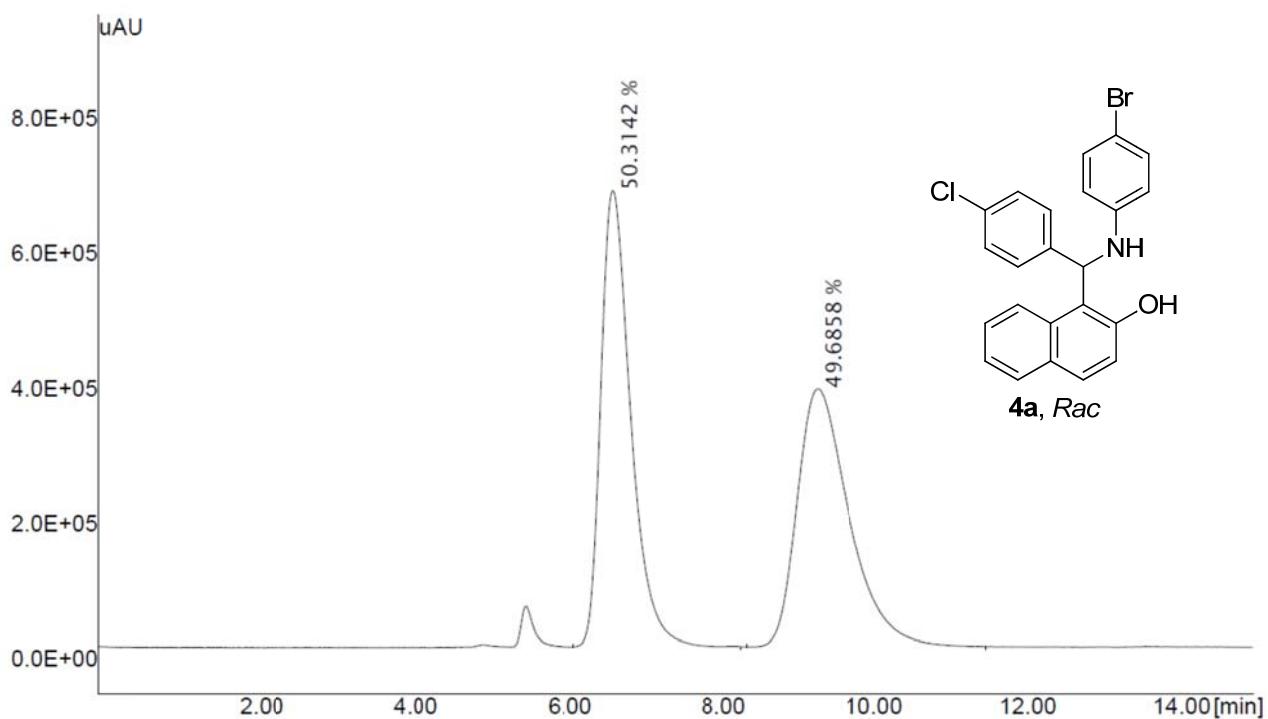


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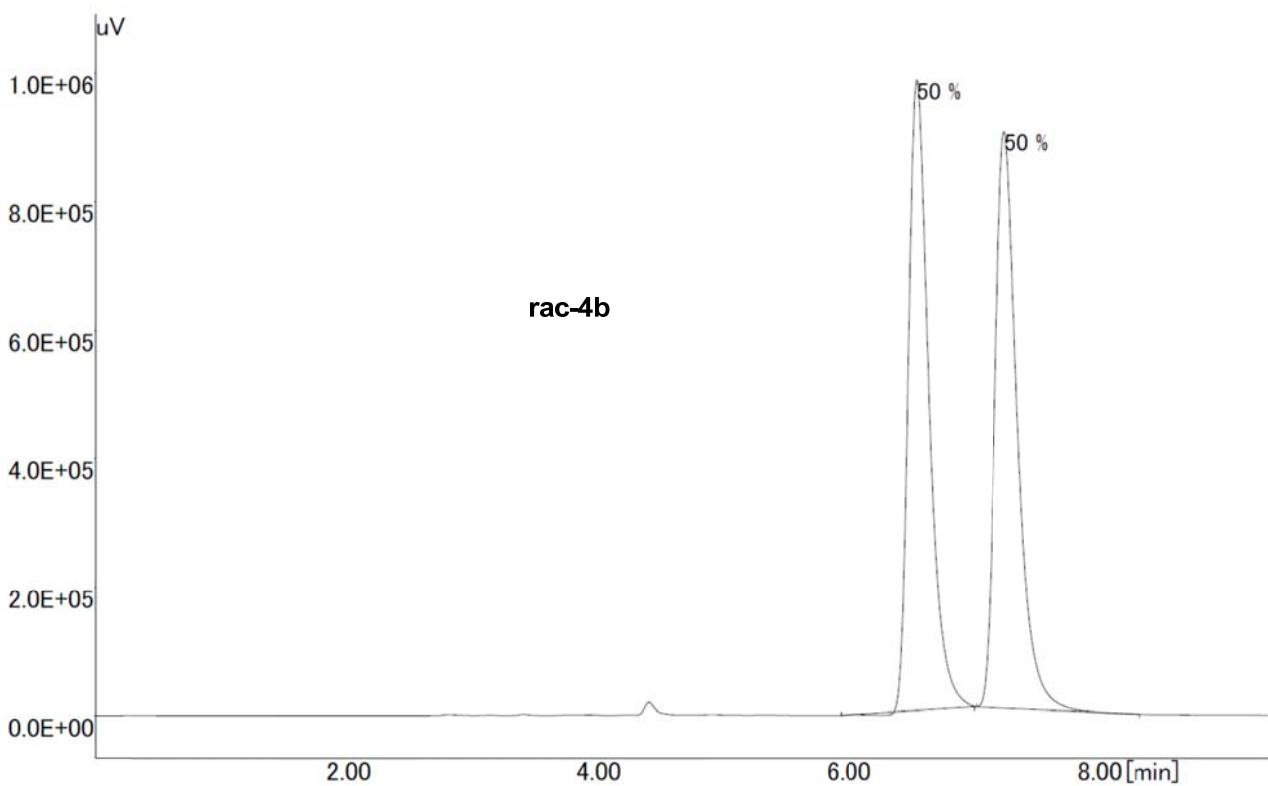
DFILE sh5166-column-2_Carbon
COMNT single_pulse_decc
DATIM 2014-06-16 10:54:
OBNUC 13C
EXMOD carbon.jxp
OBFRQ 100.53 MHz
OBSET 5.35 KHz
OBFIN 5.86 Hz
POINT 26224
FREQU 25125.63 Hz
SCANS 882
ACQTM 1.0433 sec
PD 2.0000 sec
PW1 3.20 usec
IRNUC 1H
CTEMP 20.9 c
SLVNT CDCL3
EXREF 77.00 ppm
BF 0.10 Hz
RGAIN 50

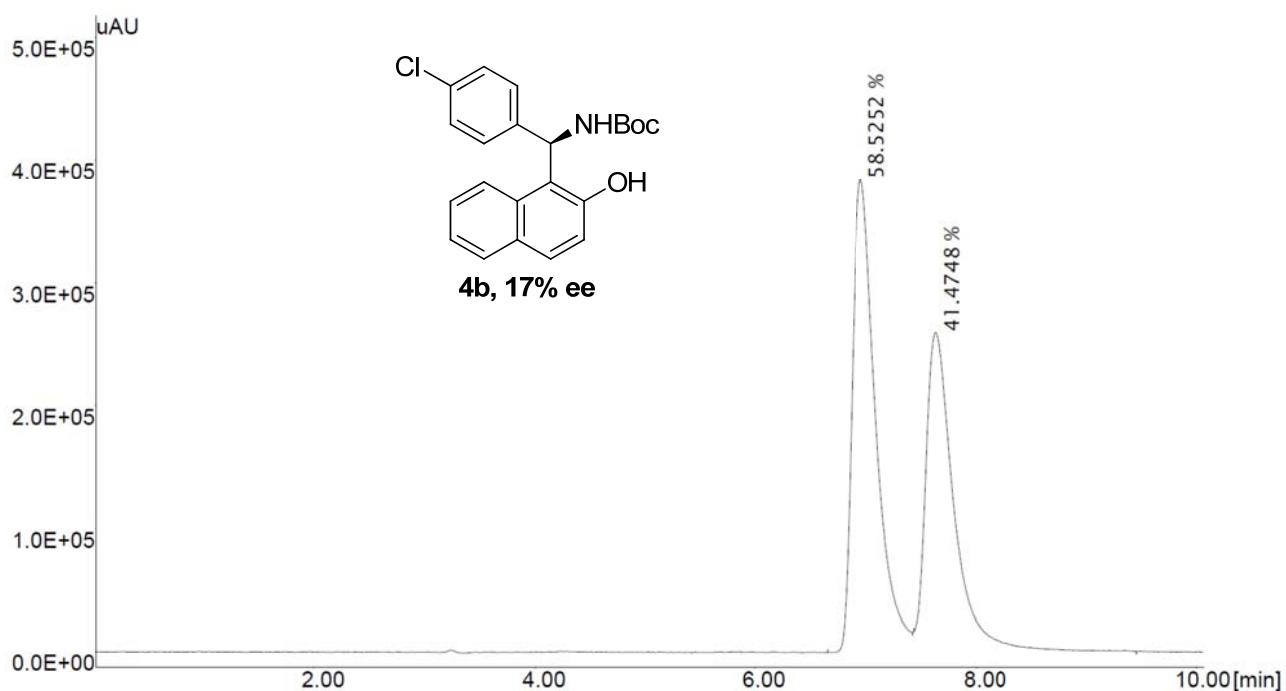
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HPLC Chart

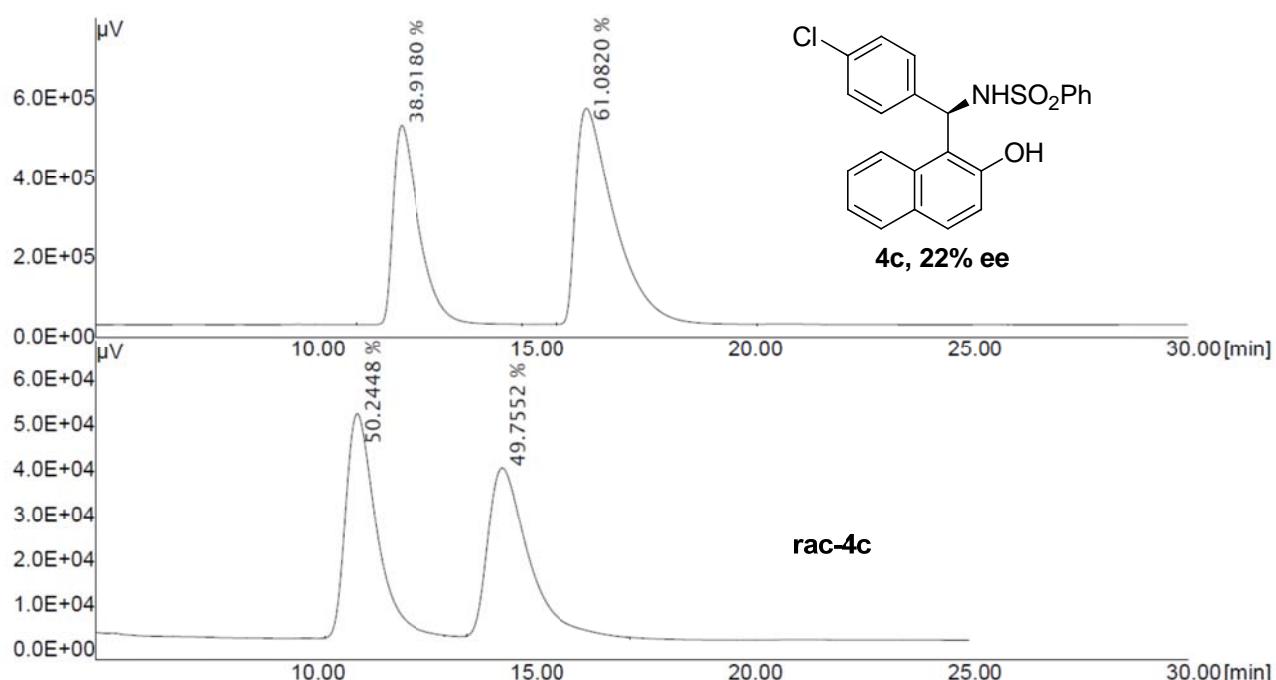


#	Name	RT	Height [uAU]	Area [uAU. Sec]	%Area
1		6.693	675953	18176759.071	50.31
2		9.347	381760	17949775.467	49.69

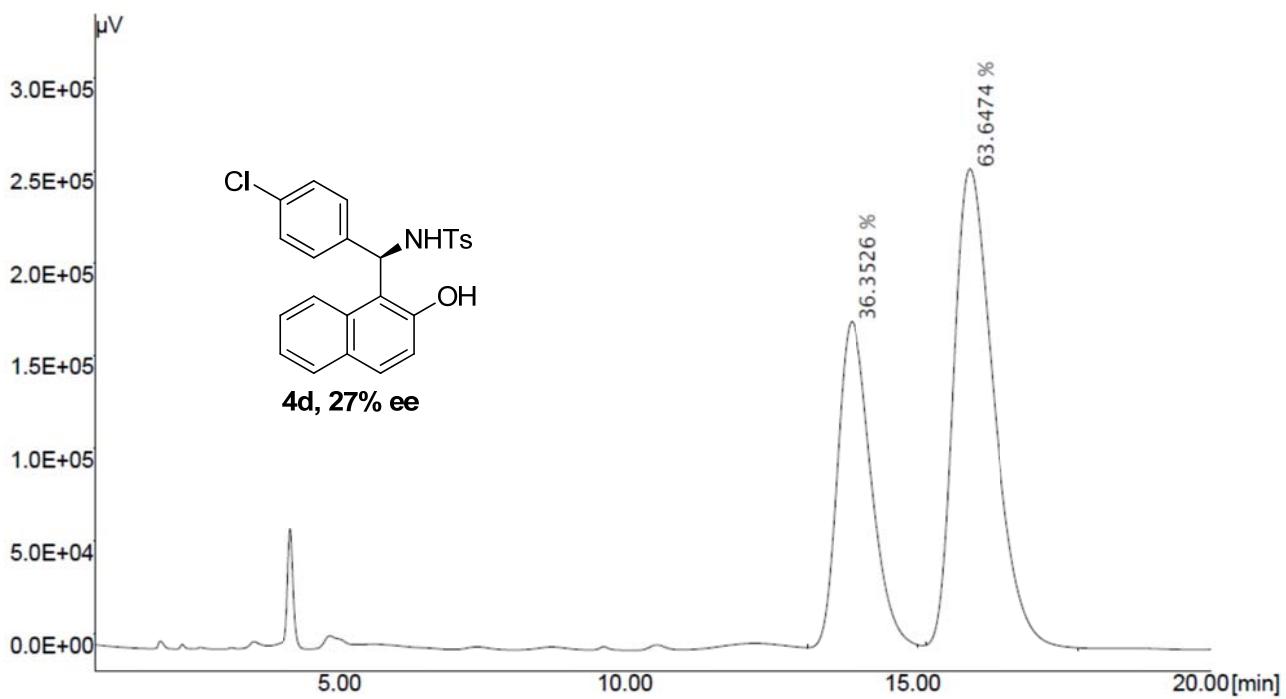
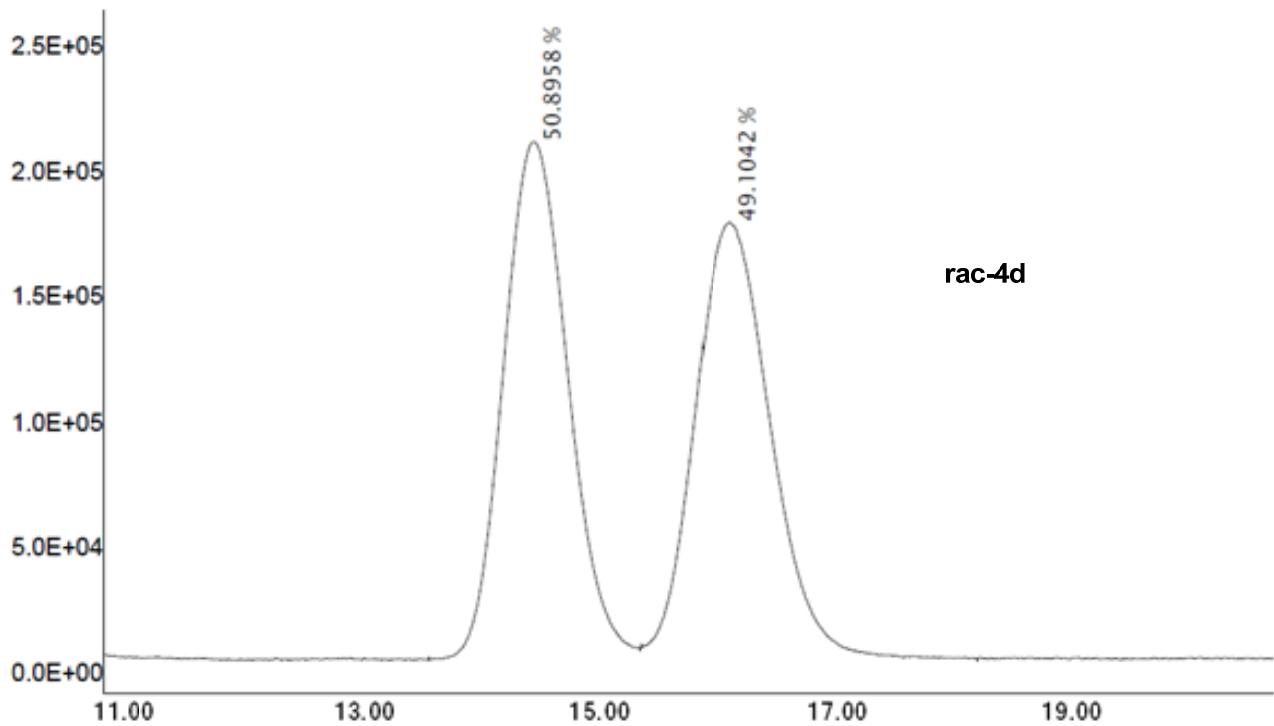




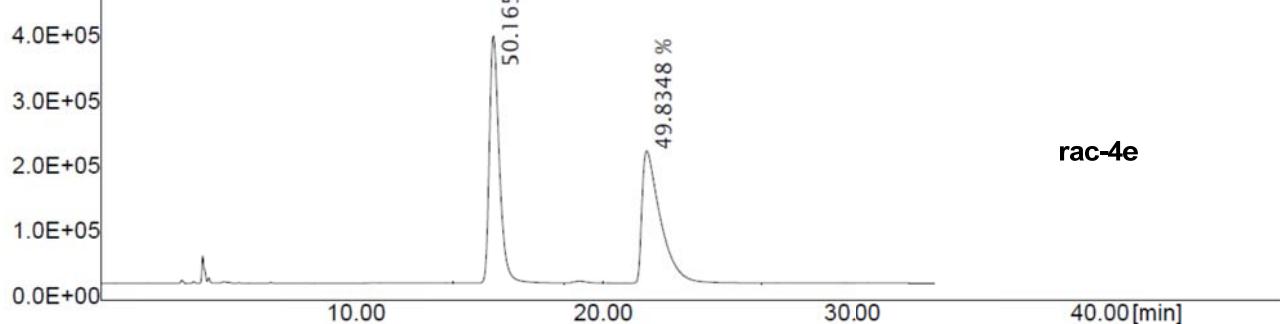
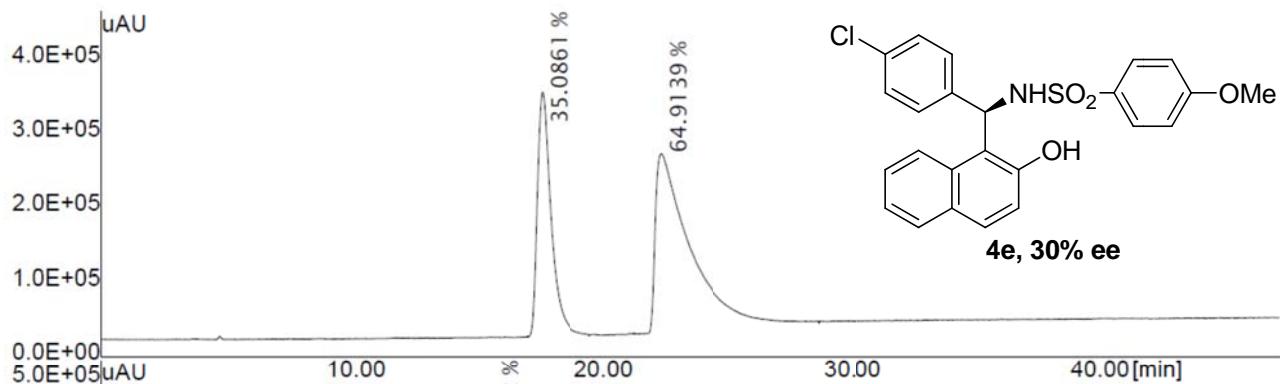
#	Name	RT	Area [uAU. Sec]	%Area
1		6.960	5357722.328	58.53
2		7.640	3796832.062	41.47



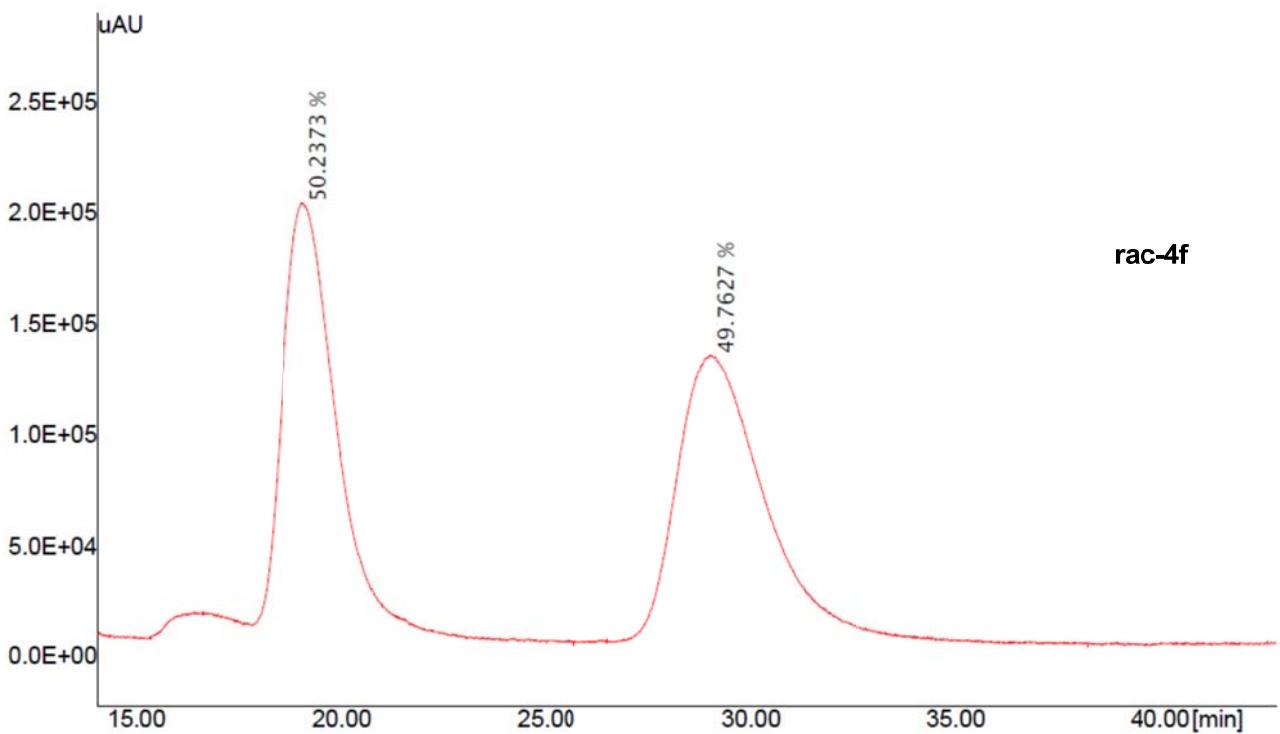
#	Name	RT	Area [$\mu\text{V} \cdot \text{Sec}$]	%Area
1		12.033	21077599.000	38.92
2		16.267	33081388.502	61.08

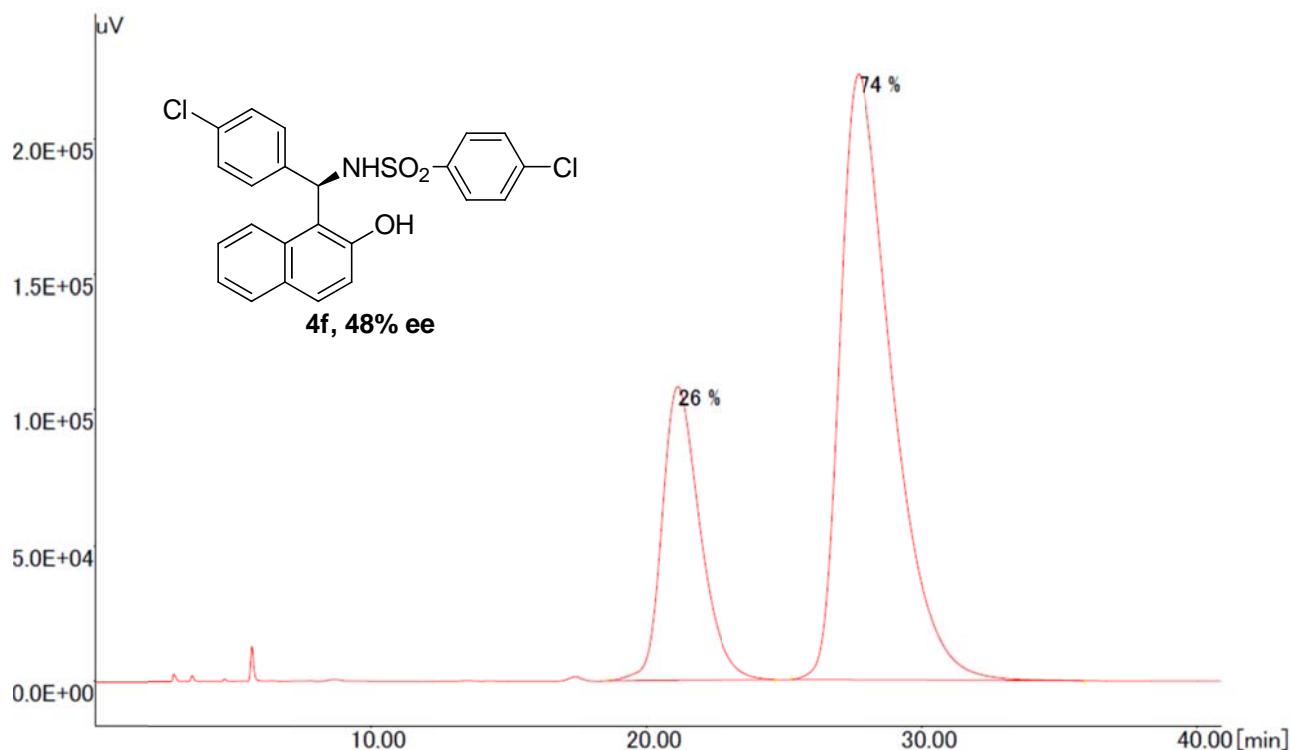


#	Name	RT	Area [fV.Sec]	%Area
1		14.058	7003414.437	36.35
2		16.092	12261830.500	63.65

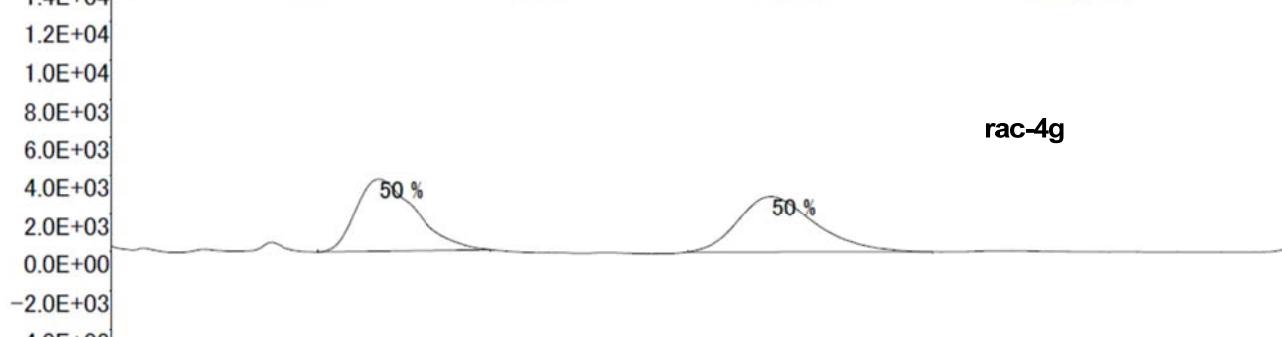
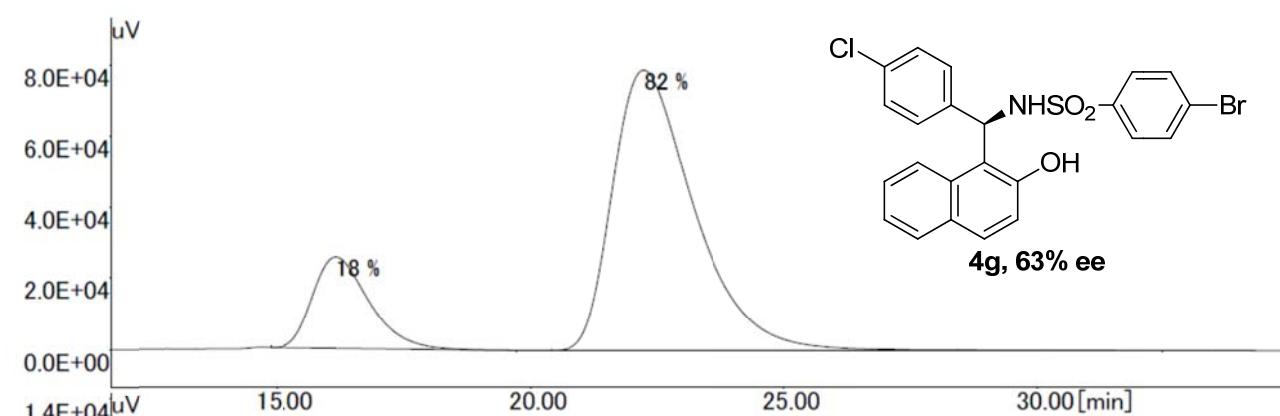


#	Name	RT	Area [uAU. Sec]	%Area
1		17.733	12585710.588	35.09
2		22.560	23285239.521	64.91

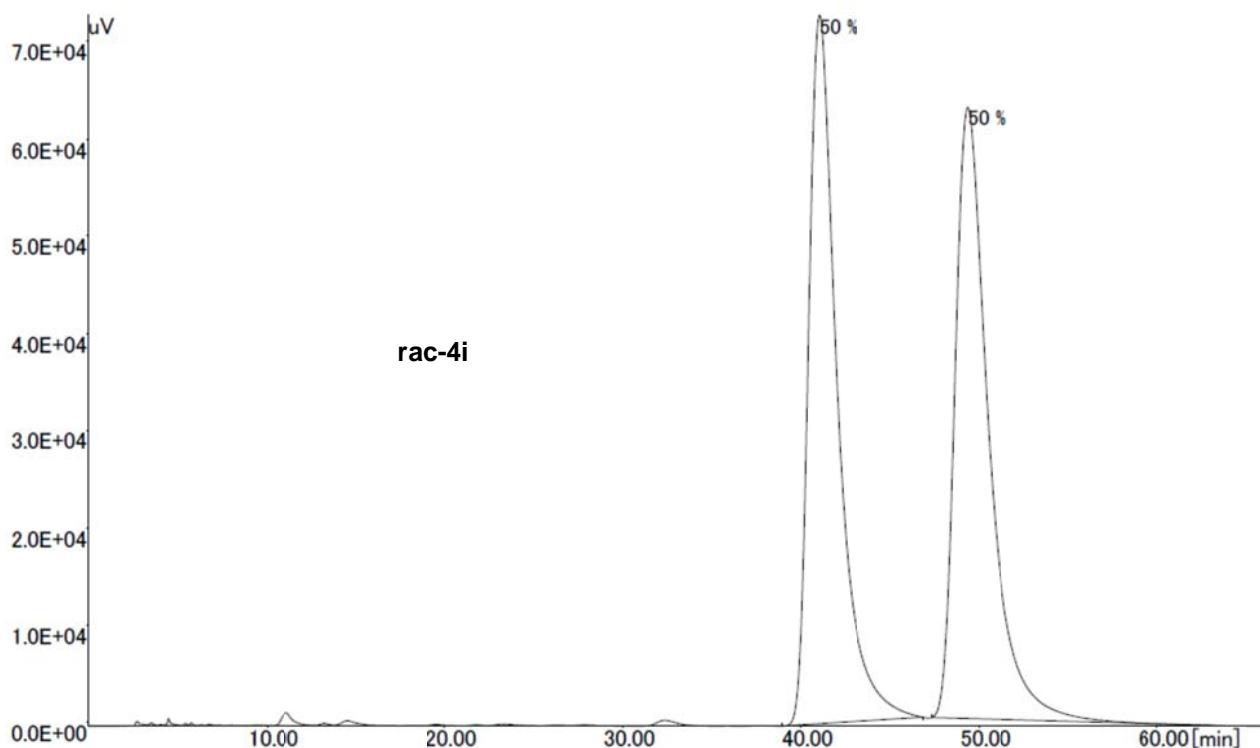
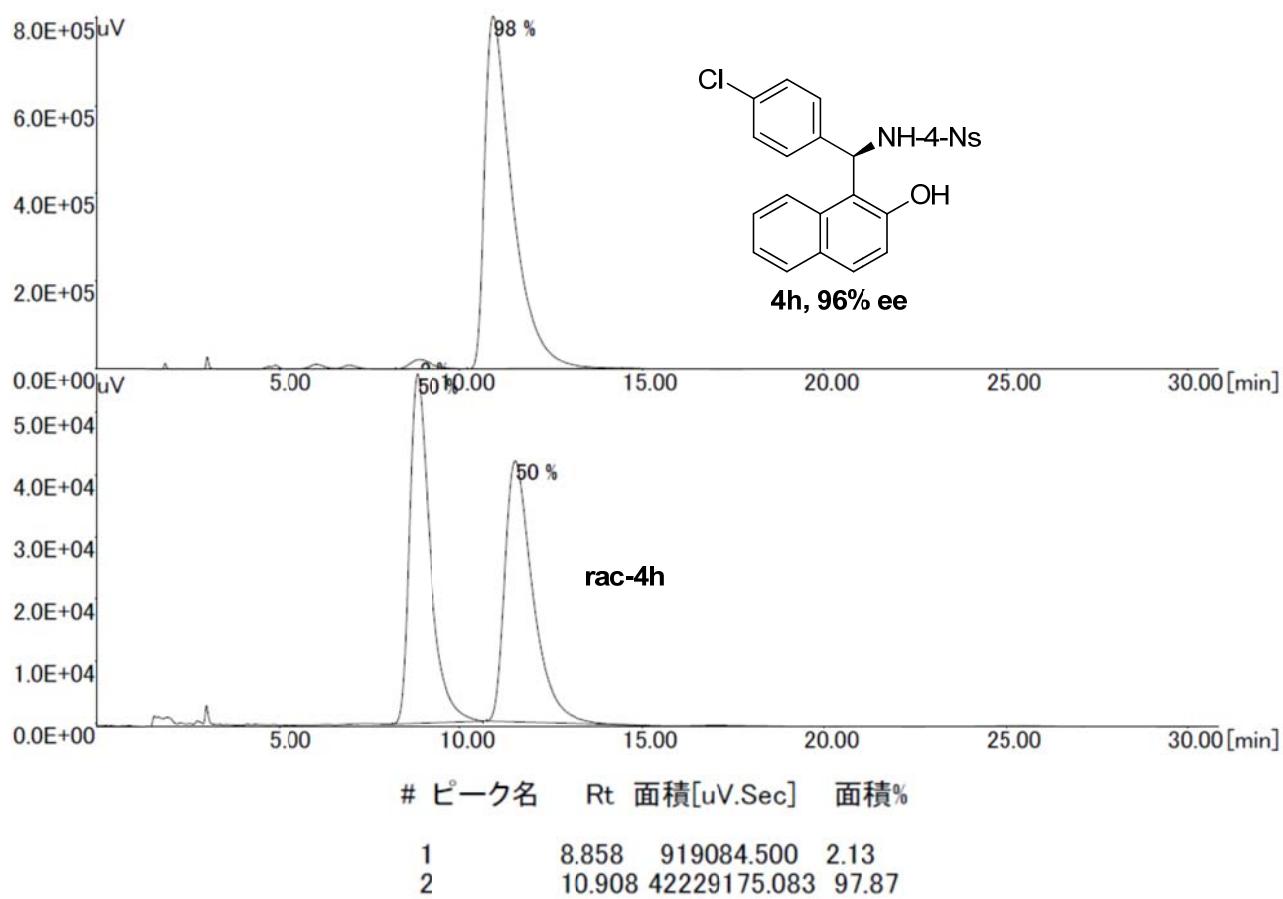


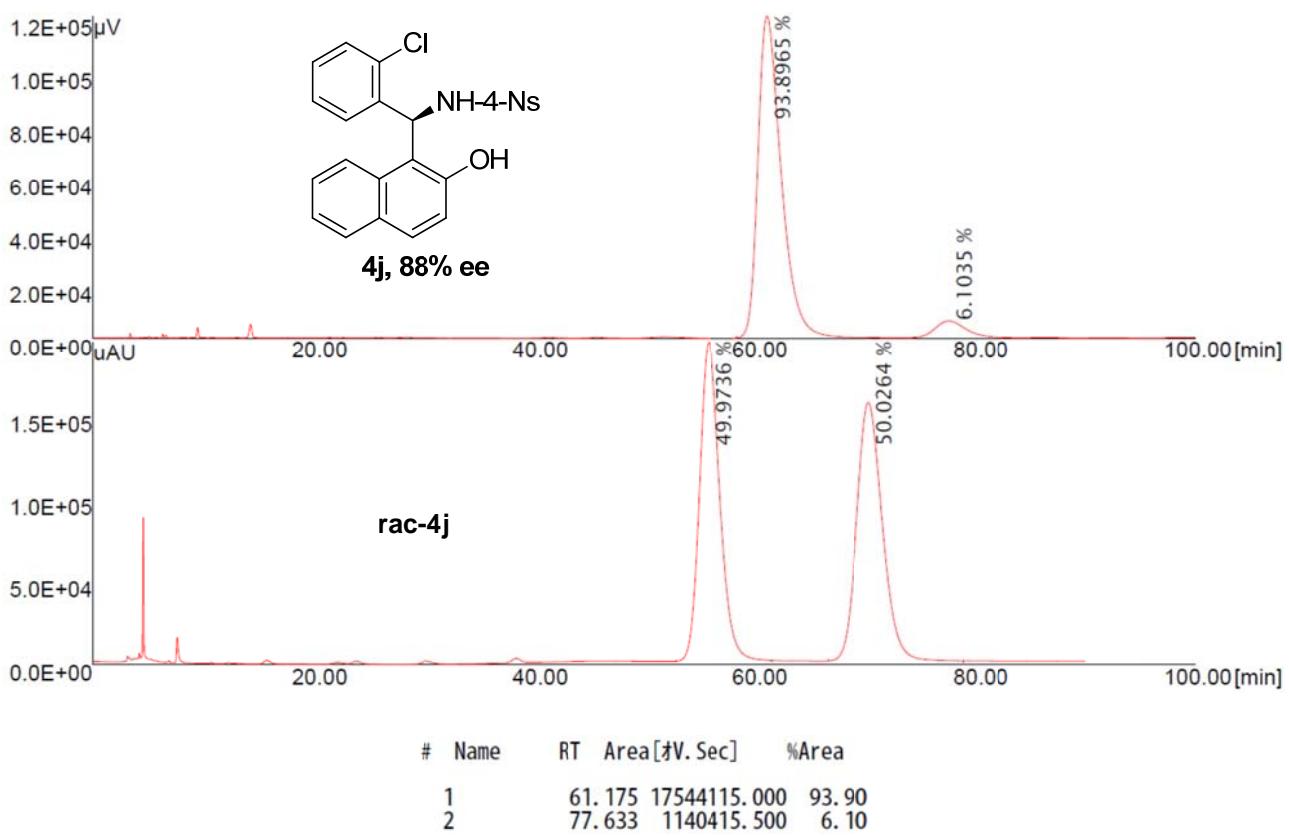
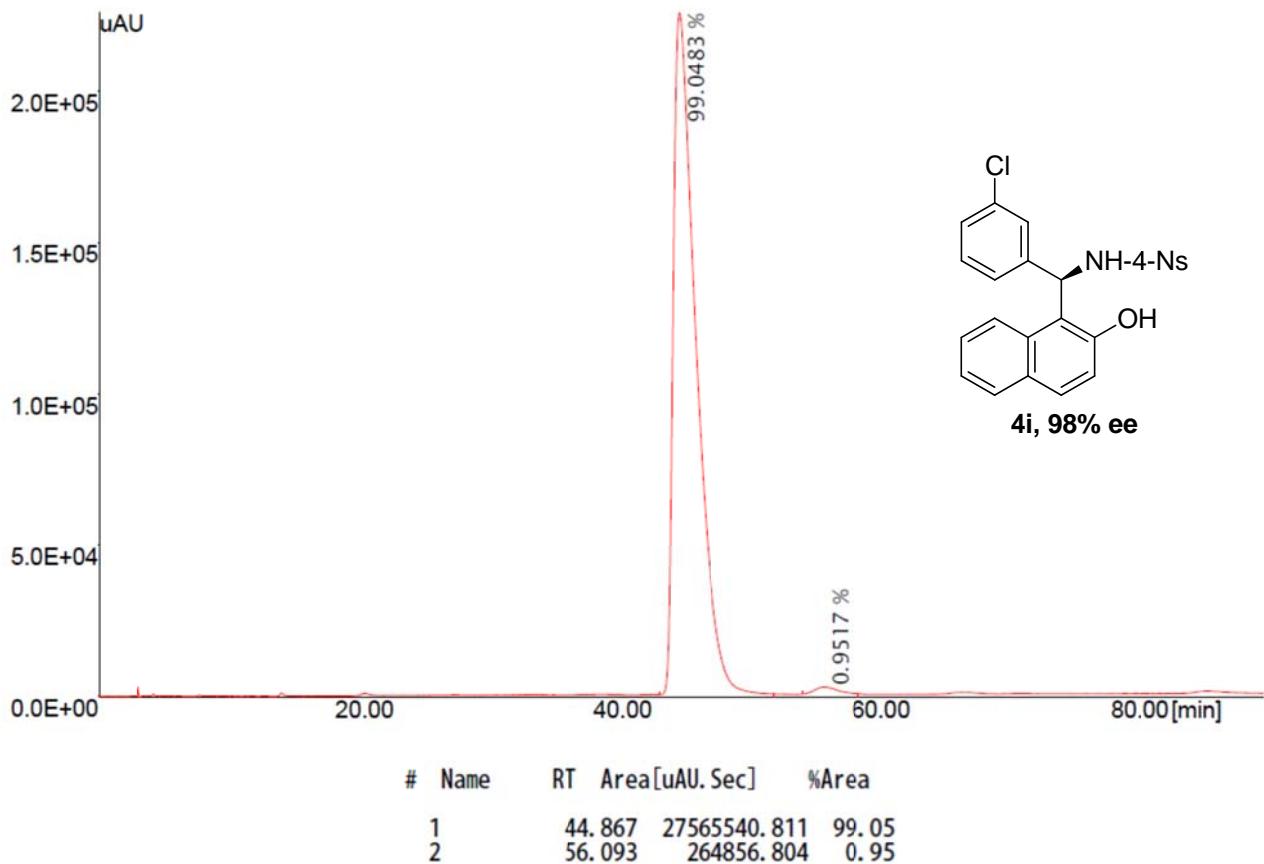


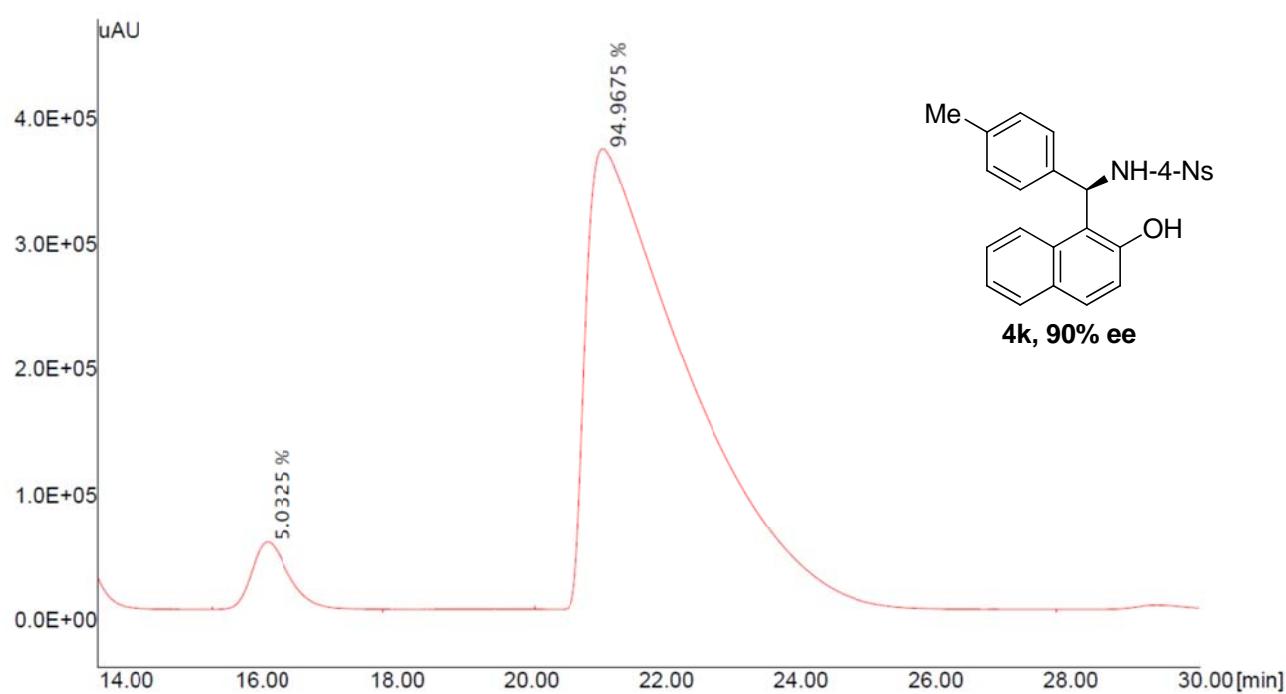
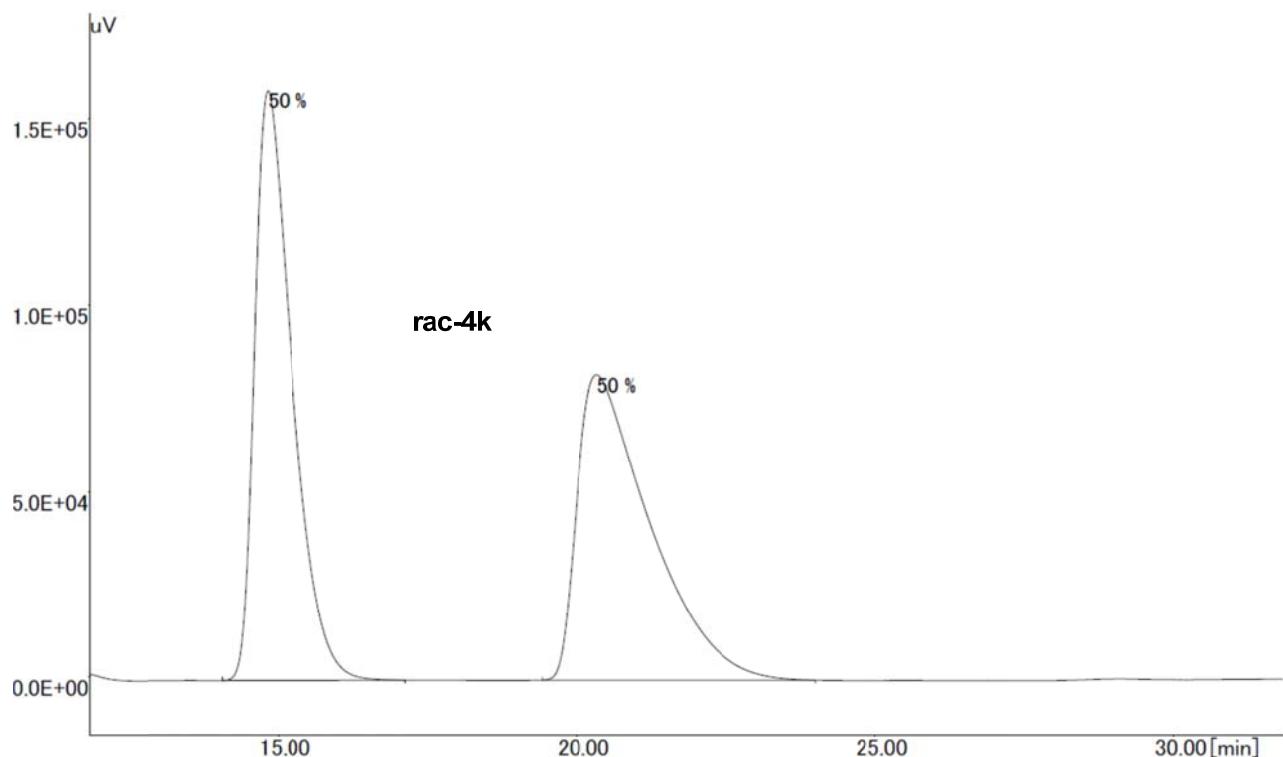
#	ピーク名	Rt ピーク名	面積%
1		21.142	26.13
2		27.717	73.87



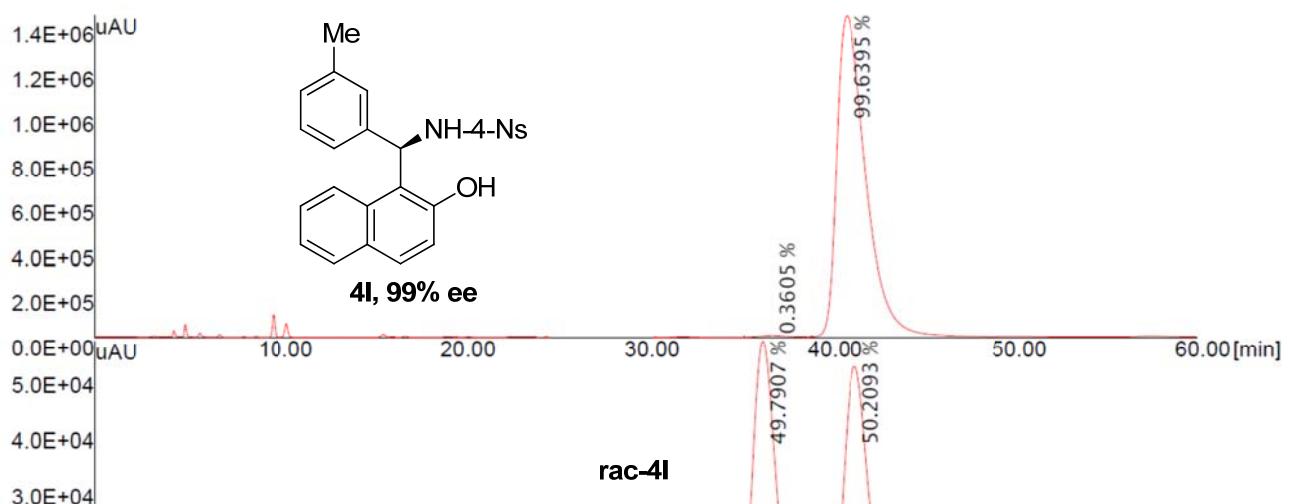
#	ピーク名	Rt ピーク名	面積%	定量値
1		16.175	18.37	0.000
2		22.242	81.63	0.000



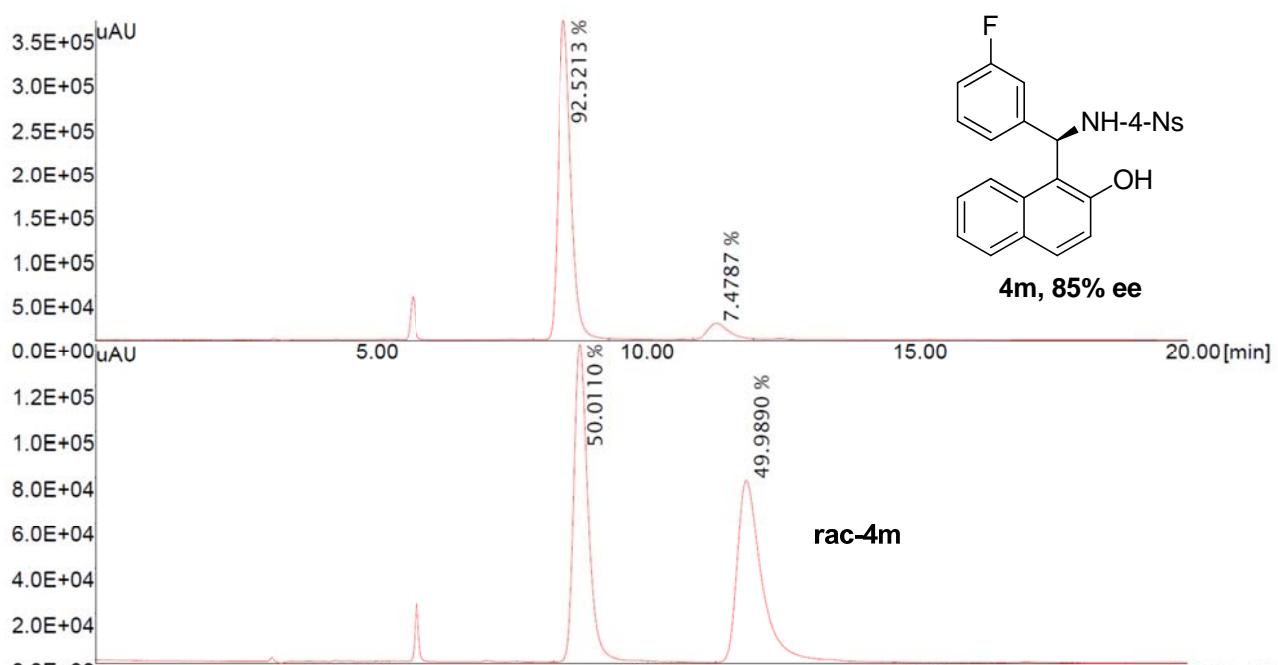




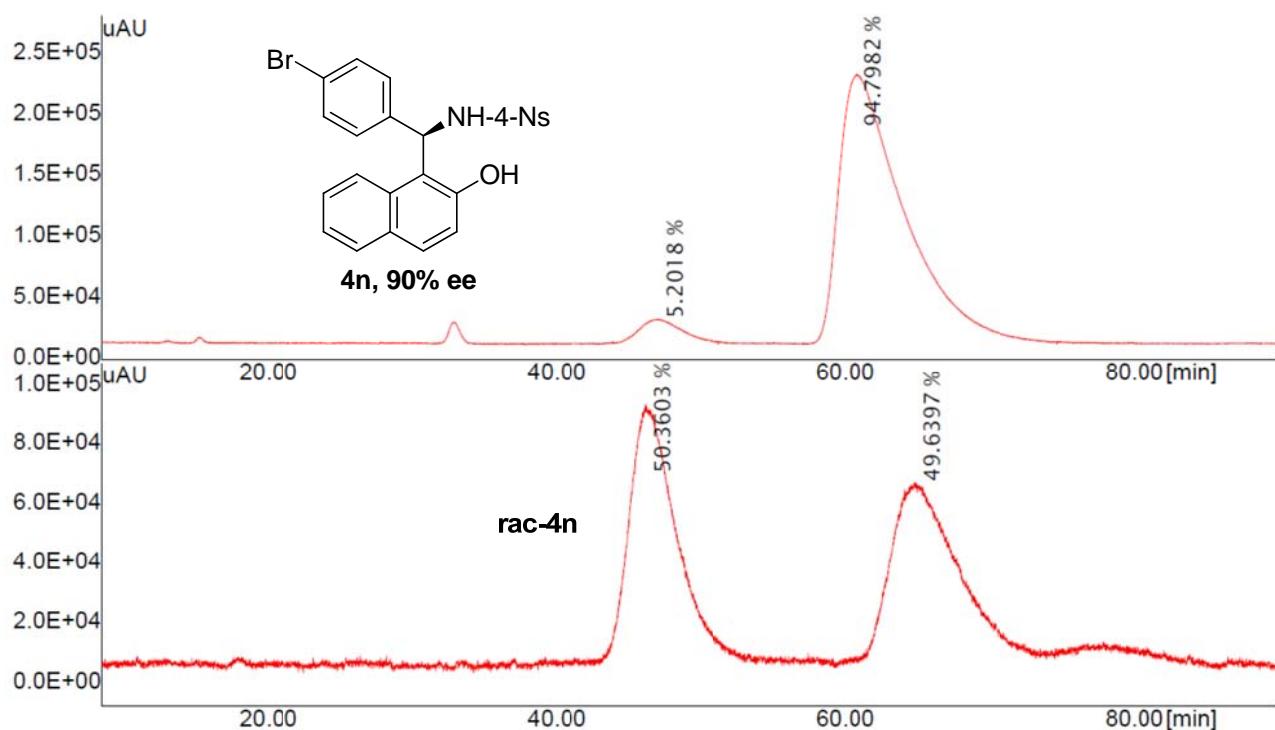
#	Name	RT	Area[uAU.Sec]	%Area
1		16.160	2062964.831	5.03
2		21.147	38929998.379	94.97



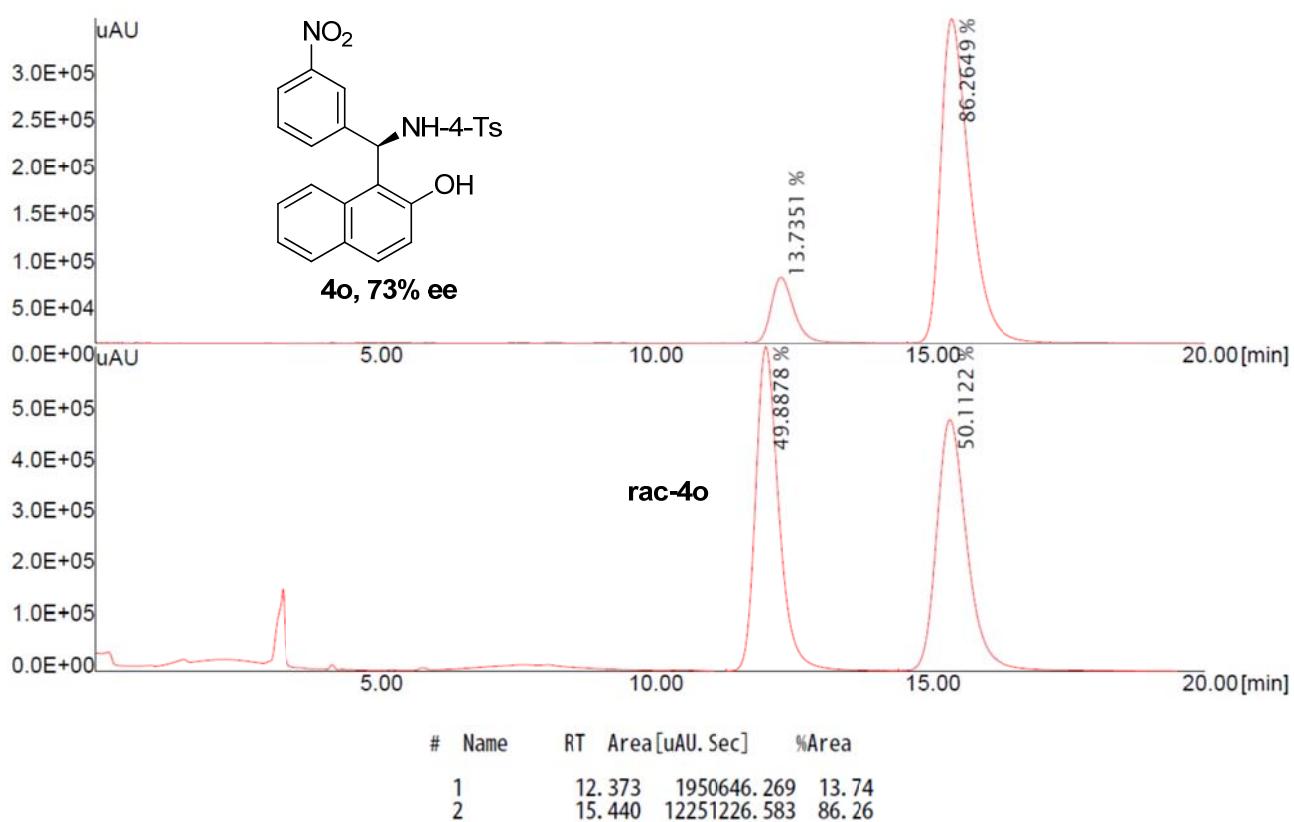
#	Name	RT	Area [uAU. Sec]	%Area
1		36.867	571956.009	0.36
2		40.973	158088692.760	99.64



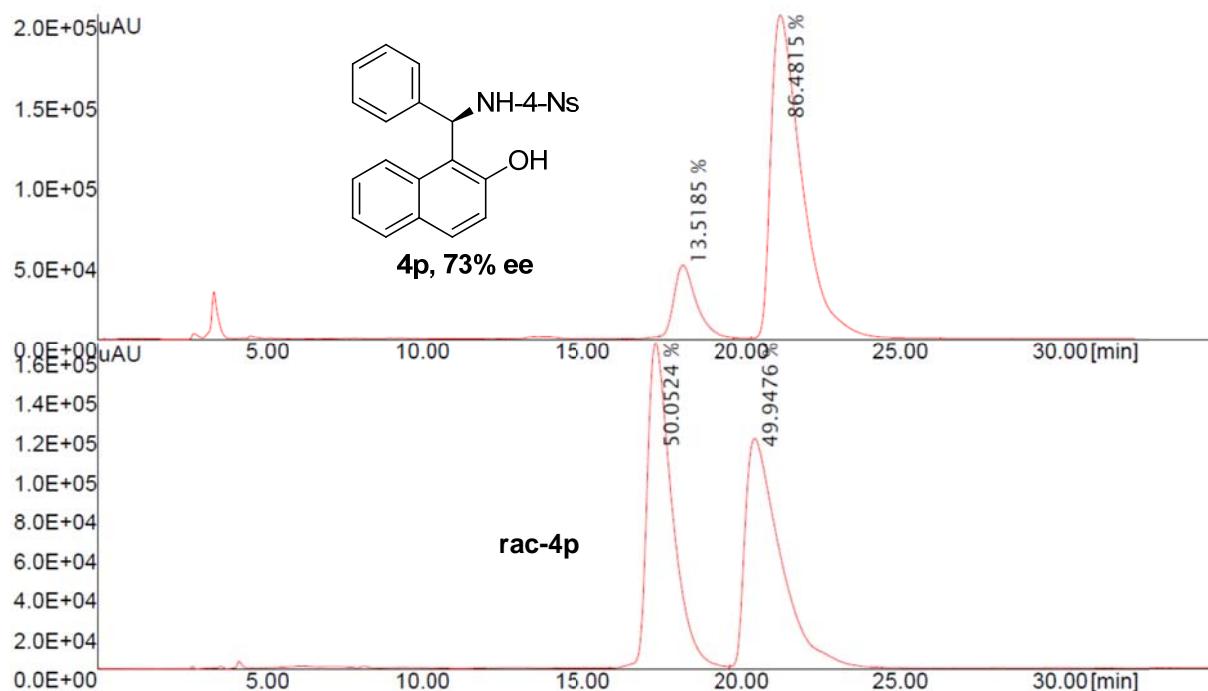
#	Name	RT	Area [uAU. Sec]	%Area
1		8.587	5839817.041	92.52
2		11.387	472047.207	7.48



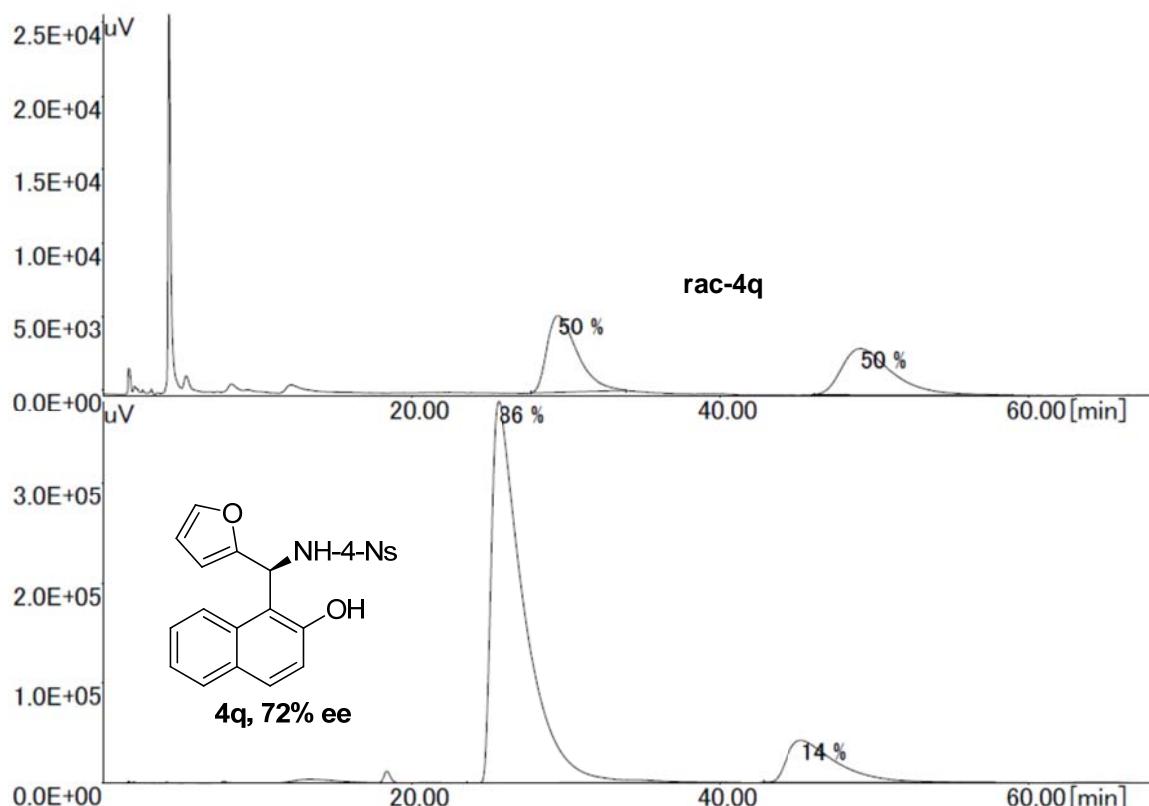
#	Name	RT	Area [uAU. Sec]	%Area
1		47.467	3798830.198	5.20
2		61.227	69230685.376	94.80



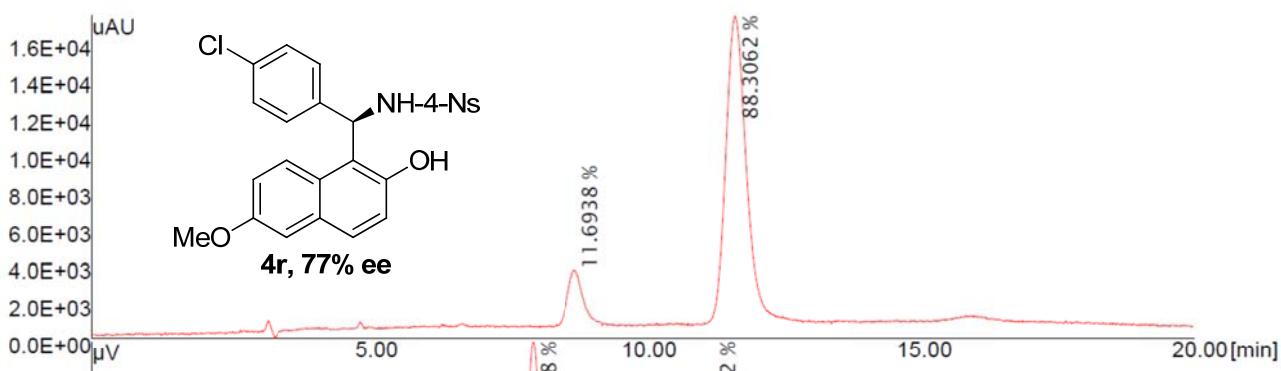
#	Name	RT	Area [uAU. Sec]	%Area
1		12.373	1950646.269	13.74
2		15.440	12251226.583	86.26



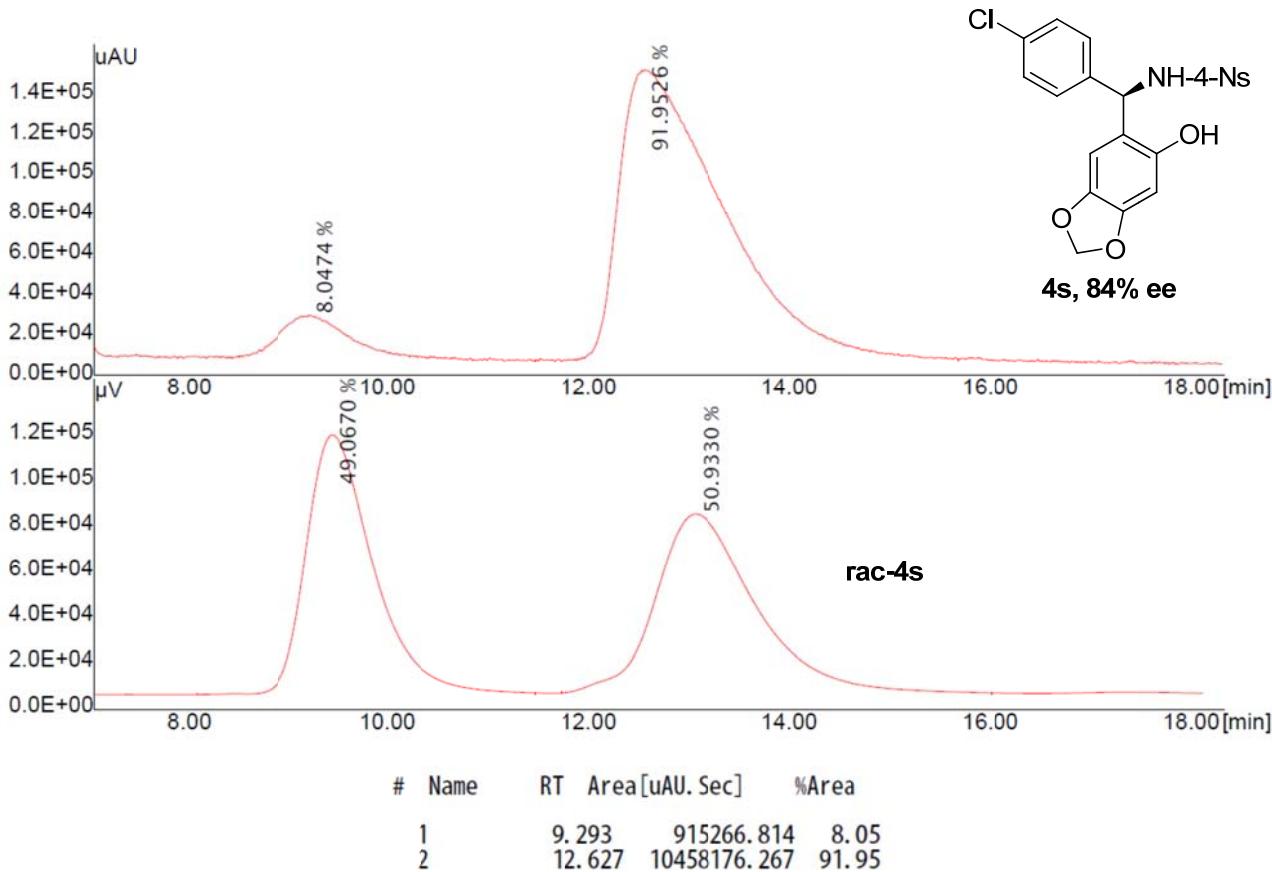
#	Name	RT	Area [uAU.Sec]	%Area
1		18.347	2100686.431	13.52
2		21.413	13438643.400	86.48



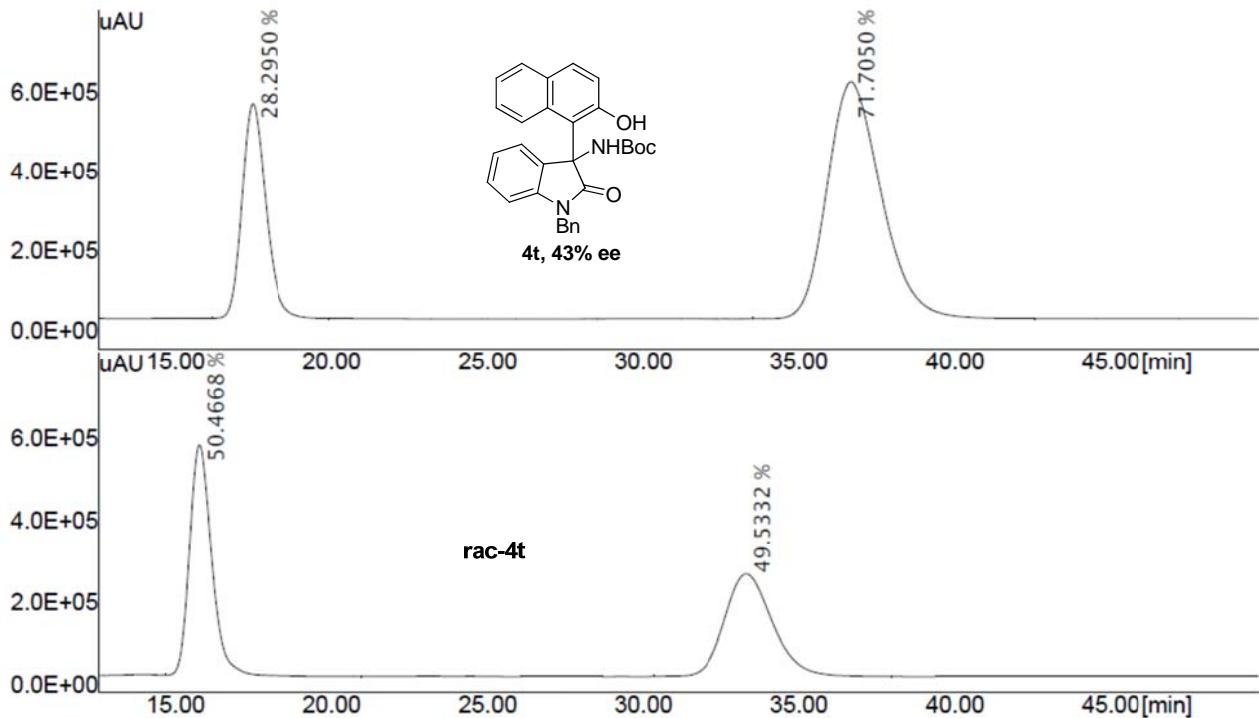
#	ピーク名	Rt	面積[uV.Sec]	面積%
1		25.633	56178788.909	85.88
2		45.292	9234136.780	14.12



#	Name	RT	Area [uAU.Sec]	%Area
1		8.760	57697.601	11.69
2		11.680	435706.770	88.31



#	Name	RT	Area [uAU.Sec]	%Area
1		9.293	915266.814	8.05
2		12.627	10458176.267	91.95



#	Name	RT	Area [uAU. Sec]	%Area
1		17.653	28493286.025	28.30
2		36.867	72207369.076	71.70