

Electronic Supplementary Information

Synthesis of a novel water-soluble cylindrical macrotricyclic host and its complexation with *N*-methylquinolinium and *N*-methylysoquinolinium salts: formation of 1:2 complexes in water

Fei Zeng^{a,b} and Chuan-Feng Chen^{*,a}

^aBeijing National Laboratory for Molecular Sciences, CAS Key Laboratory of Molecular Recognition and Function, Institute of Chemistry, Chinese Academy of Sciences, Beijing 100190, China. ^bUniversity of Chinese Academy of Sciences, Beijing 100049, China.

Email: cchen@iccas.ac.cn

Contents

1. Genaral information	S2
2. Complexation between host 1 and guest 3	S3
3. Complexation between host 1 and guest 4	S4
4. Photographs of complexation and decomplexation of complex 1·3 ₂	S5
5. Photographs of complexation and decomplexation of complex 1·4 ₂	S5
6. Determination of the average association constants of the complexes	S5
7. Partial ¹ H- ¹ H COSY and ROESY spectra of complexes 1·2 ₂ , 1·3 ₂ and 1·4 ₂	S11
8. ¹ H NMR, ¹³ C NMR and HRESI spectra of all new compounds	S13
9. UV/Vis spectral of complexes 1·2 ₂ , 1·3 ₂ and 1·4 ₂	S 错误！未定义书签。
10. The ¹ H-NMR titrations spetra of complexes 1·2 ₂ , 1·3 ₂ and 1·4 ₂	S 错误！未定义书签。
11. The Job-Plot for complexes 1·2 ₂ , 1·3 ₂ and 1·4 ₂	S20
12. Crystal data	S21
13. References	S22

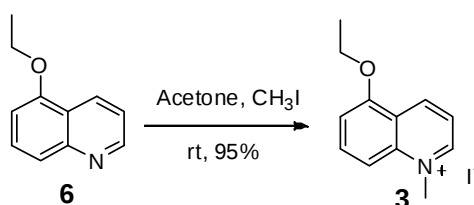
1. General information

Melting points, taken on an electrothermal melting point apparatus, are uncorrected.

^1H NMR, ^{13}C NMR spectra were recorded on a Bruker DMX300 NMR spectrometer.

2D-ROESY experiments were performed on a Bruker AVANCE 600 MHz spectrometer. High-resolution mass spectra (HRMS) were recorded on a Thermo Fisher Scientific ExactiveTM spectrometer. Compounds **2**,^{S1} **4**,^{S2} **5**^{S3} and **6**^{S4} were prepared according to the literature procedures.

Synthesis of host 1. A mixture of **5** (210 mg, 0.16 mmol) and NaOH (26 mg, 0.64 mmol) in $\text{CH}_3\text{OH}/\text{H}_2\text{O}$ (v/v = 9:1, 15 mL) was stirred for 12 hours at 60 °C under nitrogen atmosphere. Then, the solvent was removed under vacuum to give host **1** (211mg, 98%) as a dark red solid. Mp: >300 °C. ^1H NMR (300 MHz, D_2O , 295 K): δ 6.97 (s, 8H), 3.99-3.84 (m, 16H), 3.55-3.50 (m, 32H), 2.02 (s, 12H). ^{13}C NMR (75 MHz, D_2O , 295 K): δ 175.2, 147.5, 144.4, 143.6, 109.6, 69.9, 69.4, 69.3, 48.1, 13.2. HRMS cald. for $[\text{M}-2\text{Na}]^{2-}$: 633.1954. Found: 633.1966.



Synthesis of guest 3. A mixture of 5-ethoxyquinoline **6** (200 mg, 1.15 mmol) and CH_3I (0.2 mL) was in acetone (20 mL) stirred overnight at room temperature under nitrogen atmosphere. Then, the mixture was filtrated, washed with acetone, and dried in vacuo to give guest **3** (346 mg, 95%) as a yellow solid. Mp: 198-199 °C. ^1H NMR

(300 MHz, D₂O, 295 K): δ 9.37-9.34 (d, J = 8.7 Hz, 1H), 9.05-9.03 (d, J = 5.7 Hz, 1H), 8.09-8.04 (m, 1H), 7.85-7.74 (m, 2H), 7.33-7.30 (d, J = 8.1 Hz, 1H), 4.49 (s, 3H), 4.36-4.29 (m, 2H), 1.49-1.44 (m, 3H). ¹³C NMR (75 MHz, D₂O, 295 K): δ 155.7, 148.9, 142.0, 139.3, 137.1, 122.4, 119.9, 109.0, 65.8, 45.6, 13.7. HRMS calcd. for [M-I]⁺: 188.1070. Found: 188.1064.

2. Complexation between host 1 and guest 3

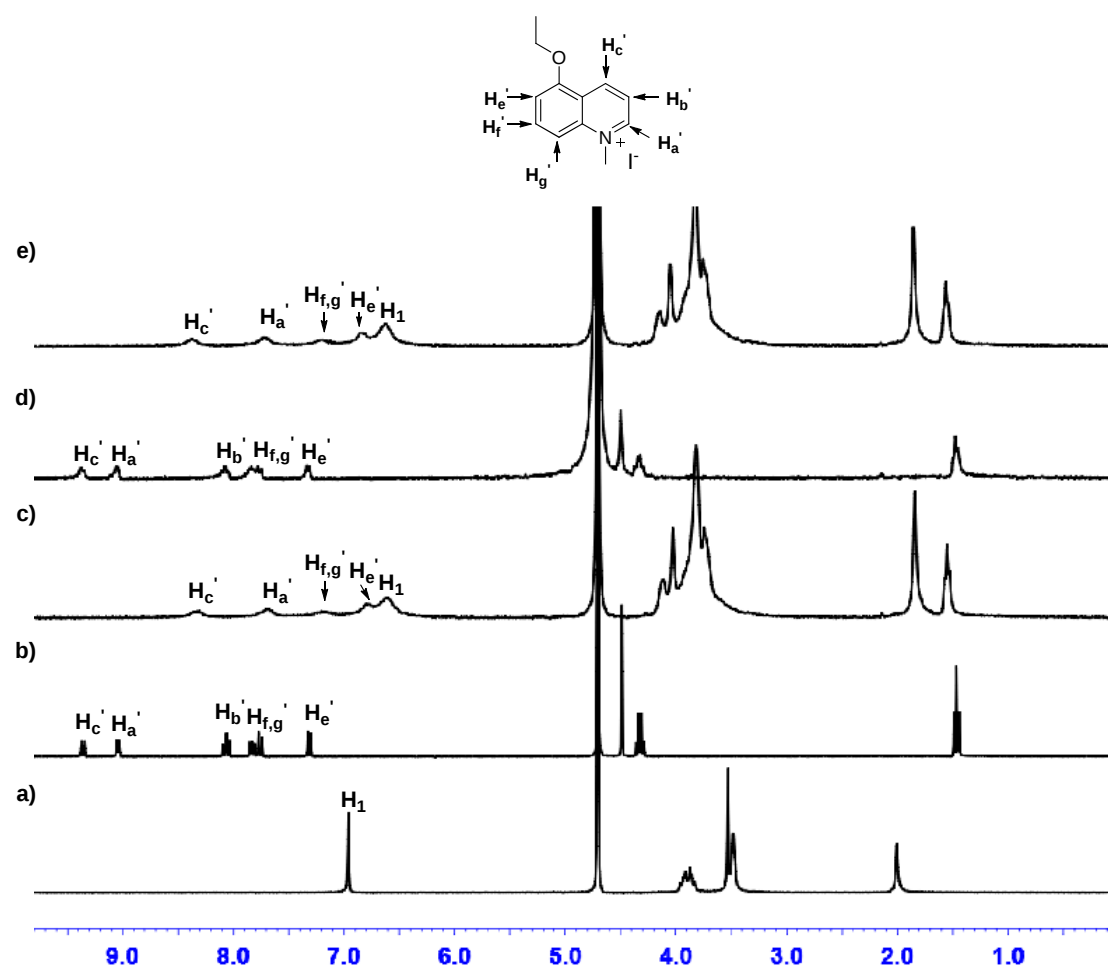


Fig. S1 Partial ¹H NMR spectra (300 MHz, D₂O, 295 K) of (a) free host **1**, (b) free guest **3**, (c) **1** and 2.0 equiv of **3**, (d) after addition of 2.0 μ L of aqueous DCl solution (20 wt%) to c, and (e) after addition of 1.0 μ L of aqueous NaOD solution (40 wt%) to d. [**1**]₀ = 4.0 mM.

3. Complexation between host 1 and guest 4

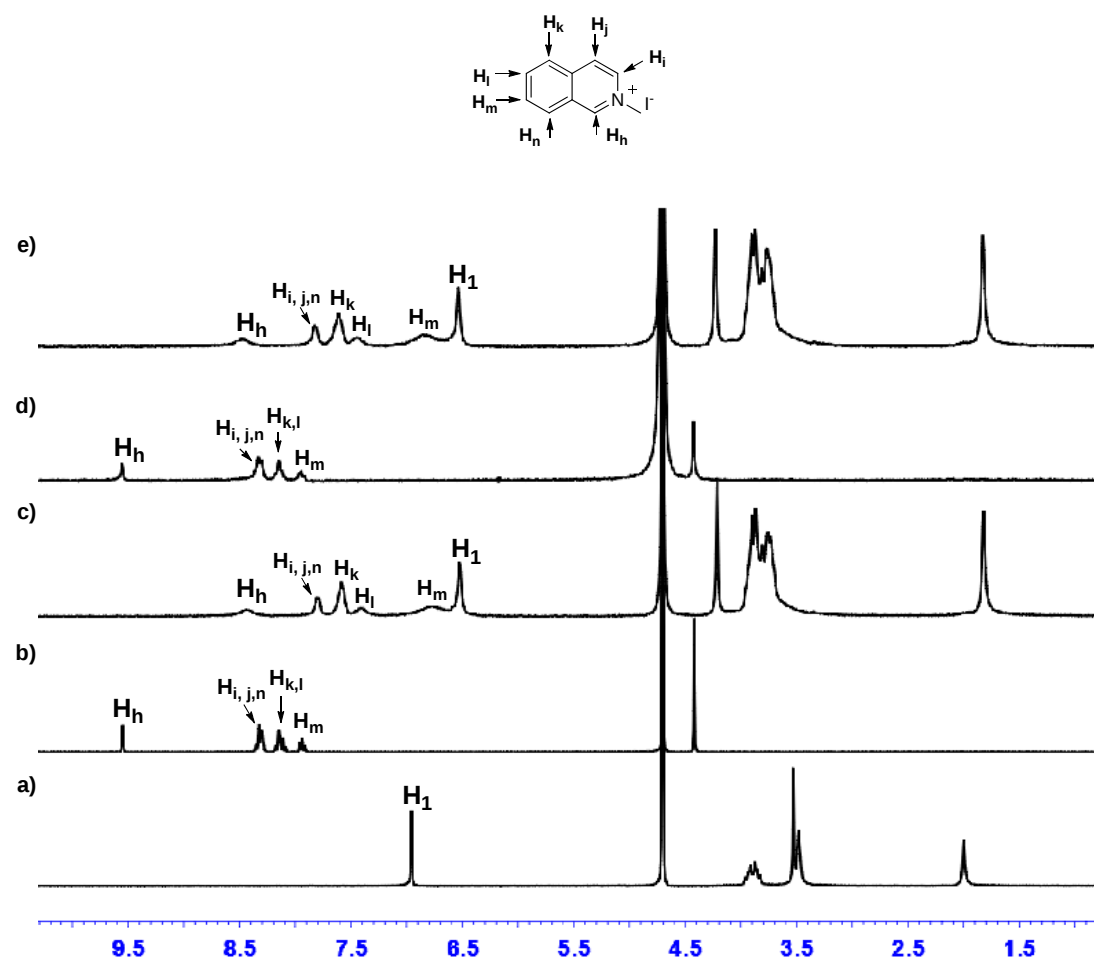


Fig. S2 Partial ^1H NMR spectra (300 MHz, D_2O , 295 K) of (a) free host **1**, (b) free guest **4**, (c) **1** and 2.0 equiv of **4**, (d) after addition of 2.0 μL of aqueous DCl solution (20 wt%) to c, and (e) after addition of 1.0 μL of aqueous NaOD solution (40 wt%) to d. $[\mathbf{1}]_0 = 4.0$ mM.

4. Photographs of complexation and decomplexation of complex 1·3₂

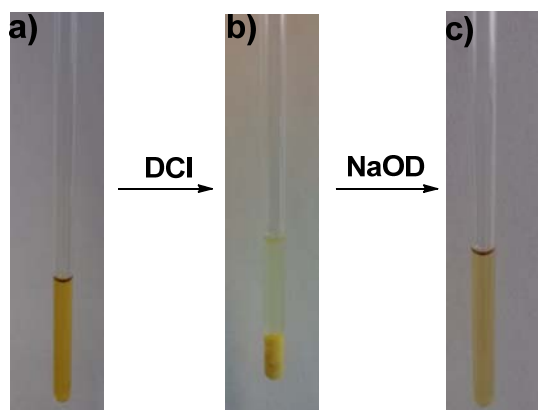


Fig. S3 Photographs of water solutions of (a) **1** and 2.0 equiv of **3**, (b) after the addition of 2.0 μL of aqueous DCl solution (20 wt%) to a, and (c) after the addition of 1.0 μL of aqueous NaOD solution (40 wt%) to b. $[\mathbf{1}]_0 = 4.0 \text{ mM}$.

5. Photographs of complexation and decomplexation of complex 1·4₂

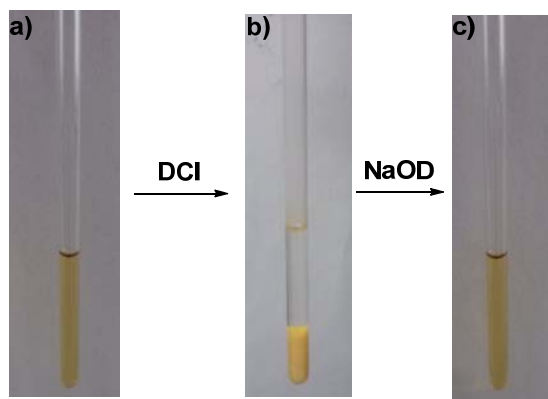


Fig. S4 Photographs of water solutions of (a) **1** and 2.0 equiv of **4**, (b) after the addition of 2.0 μL of aqueous DCl solution (20 wt%) to a, and (c) after the addition of 1.0 μL of aqueous NaOD solution (40 wt%) to b. $[\mathbf{1}]_0 = 4.0 \text{ mM}$.

6. Determination of the average association constants of the complexes

Δ is the observed chemical shift change relative to uncomplexed species, and Δ_0 is calculated chemical shift change of fully complexed species, and is determined by

extrapolation of a plot of Δ vs $1/[1]$ in the high initial concentration range of **1**. The complex fraction (p) value was calculated from $p = \Delta / \Delta_0$, and $[1]_{uc}$ values were calculated from $[1] - 0.5p[2]_0$. A plot of $p / [1]_{uc}$ vs p was used to determine the association constant K_{av} , which is the average value of the slope and the intercept. The method used for the determination of average association constants of complexes **1**·**3**₂ and **1**·**4**₂ is similar to that of **1**·**2**₂.

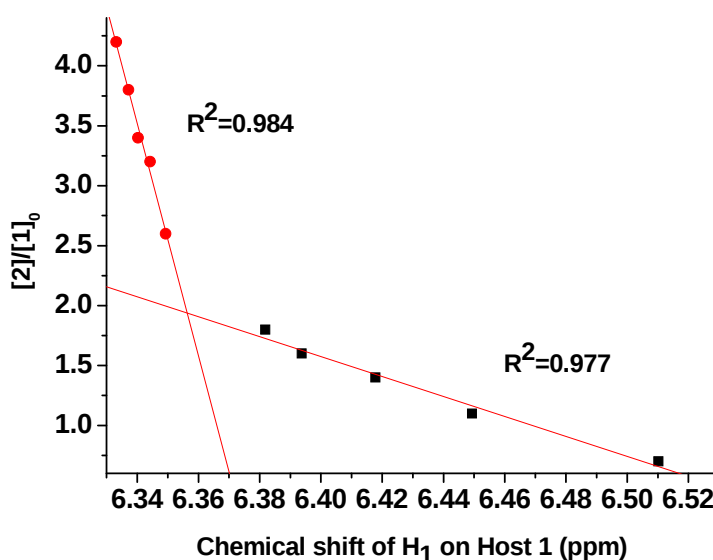


Fig. S5 Mole ratio plot for the complexation of **1** and **2** in D₂O at 295 K. $[1]_0 = 3.0$ mM .

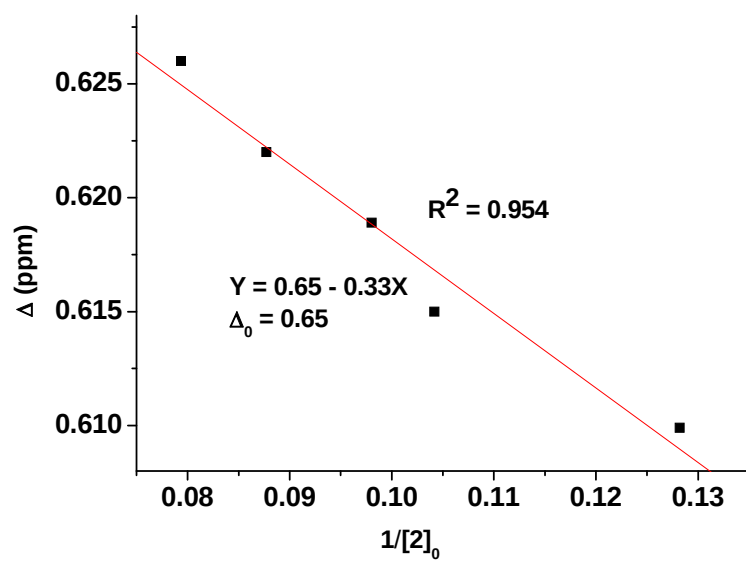


Fig. S6 Determination of Δ_0 of H_1 for the complexation between host **1** and guest **2** in D_2O at 295K.

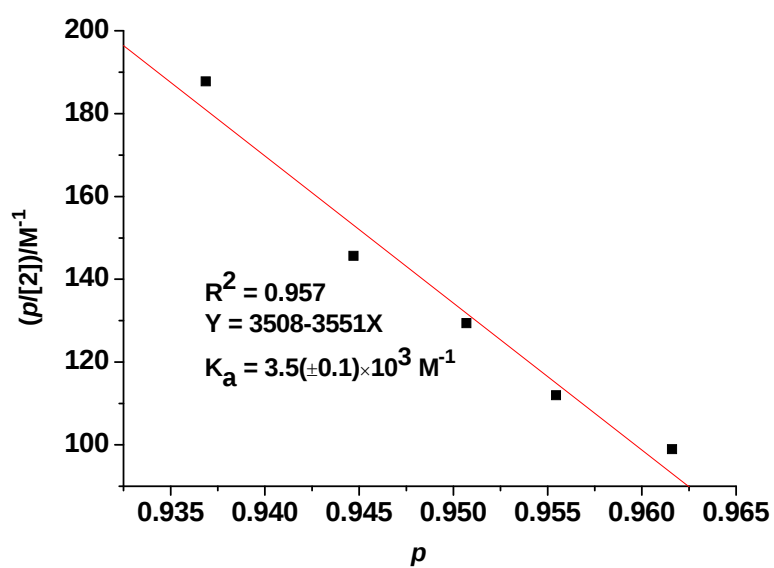


Fig. S7 Scatchard plot for the complexation of host **1** and guest **2** in D_2O at 295K.

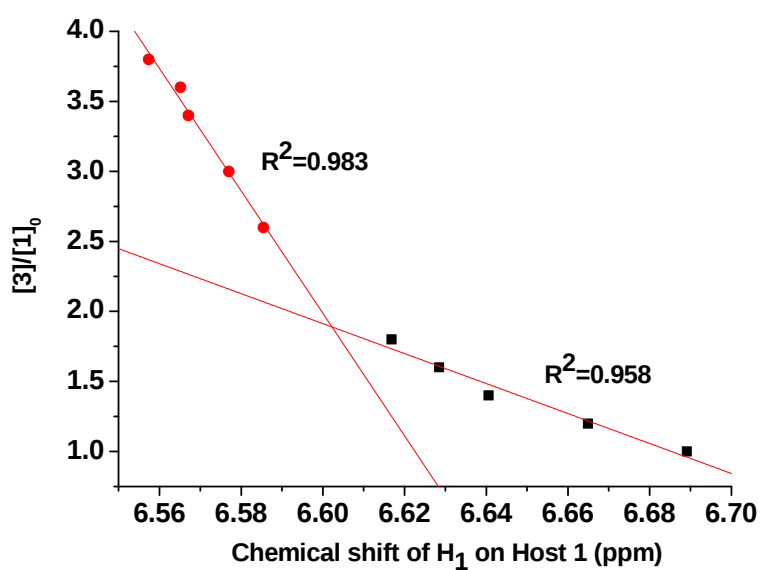


Fig. S8 Mole ratio plot for the complexation of **1** and **3** in D₂O at 295 K. $[1]_0 = 3.0$ mM .

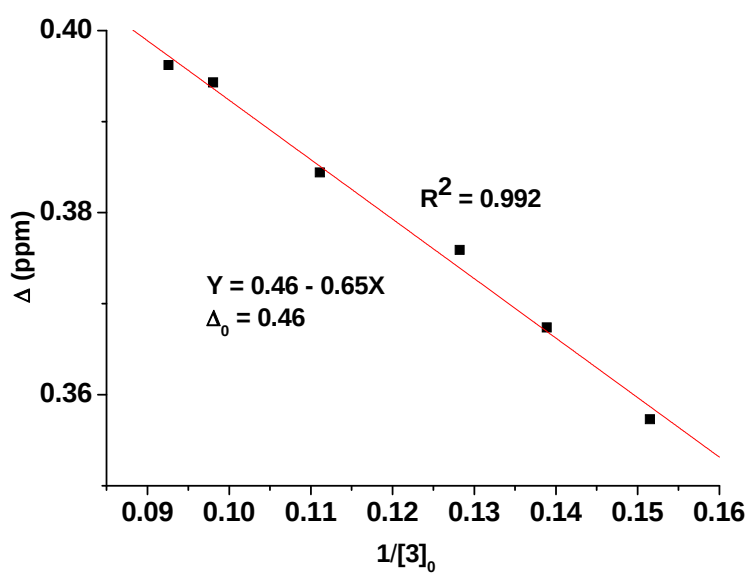


Fig. S9 Determination of Δ_0 of H₁ for the complexation between host **1** and guest **3** in D₂O at 295K.

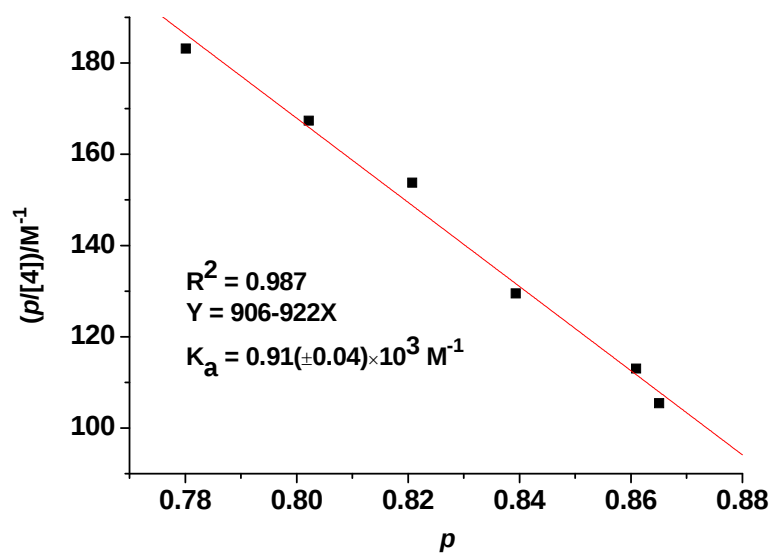


Fig. S10 Scatchard plot for the complexation of **1** and guest **3** in D₂O at 295K.

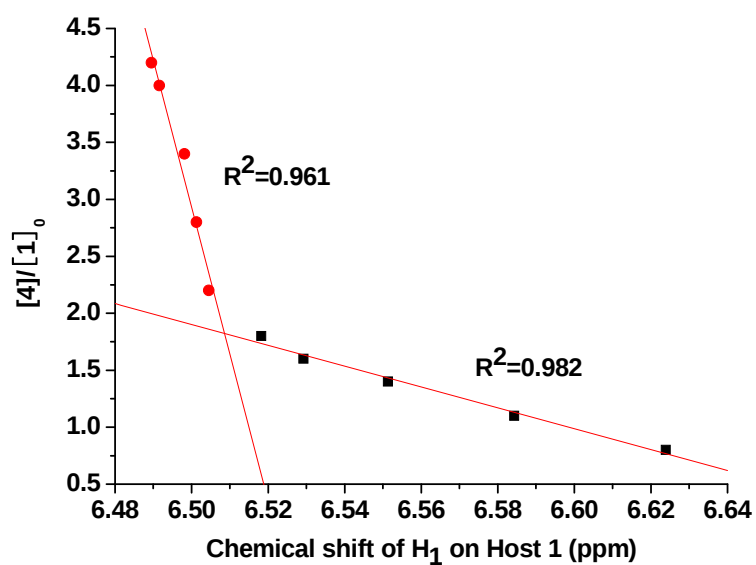


Fig. S11 Mole ratio plot for the complexation of **1** and **4** in D₂O at 295 K. $[1]_0 = 3.0$ mM .

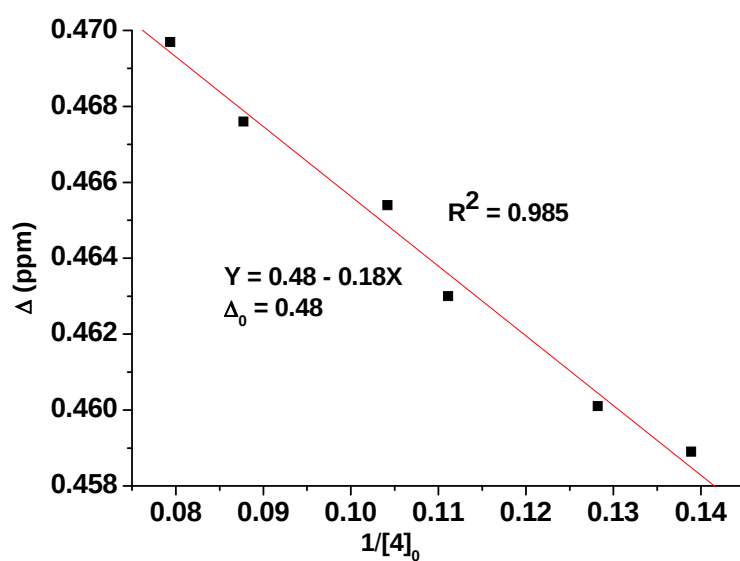


Fig. S12 Determination of Δ_0 of H_1 for the complexation between host **1** and guest **4** in D_2O at 295K.

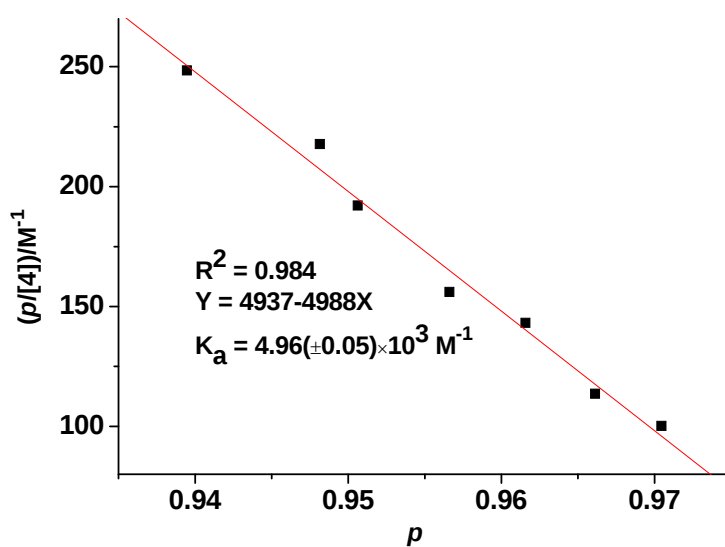


Fig. S13 Scatchard plot for the complexation of host **1** and guest **4** in D_2O at 295K.

7. Partial ^1H - ^1H COSY and ROESY spectra of complexes $1\cdot 2_2$, $1\cdot 3_2$ and $1\cdot 4_2$

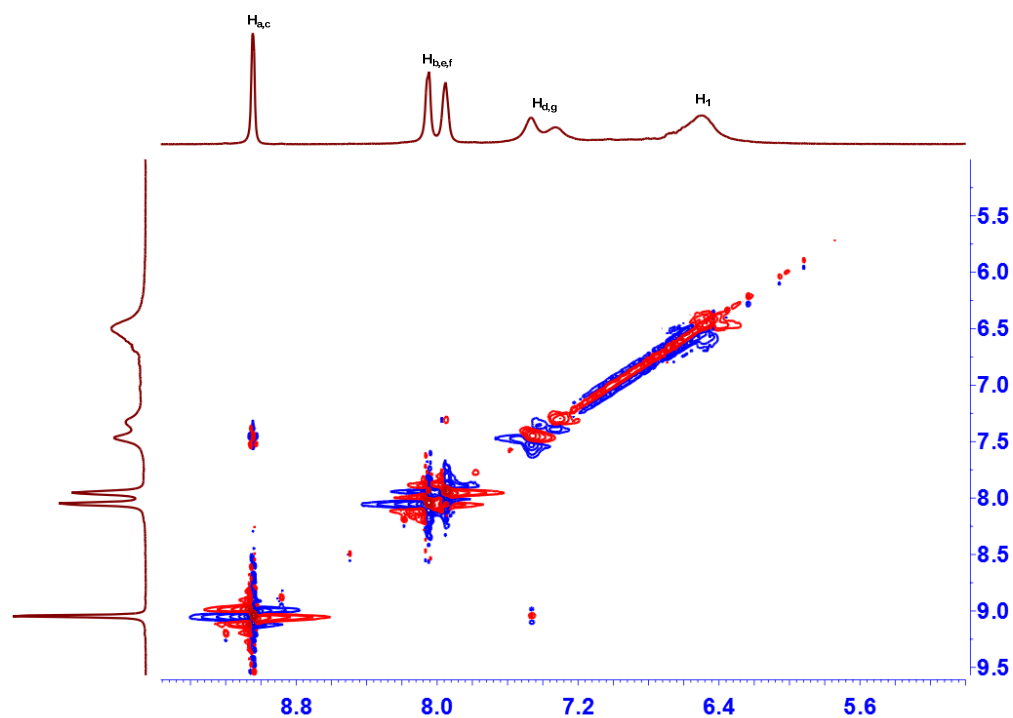


Fig. S14 Partial ^1H - ^1H COSY spectrum (600 MHz, D_2O , 295 K) of a solution of host **1** and 2.0 equiv of guest **2**. $[\mathbf{1}]_0 = 8.0$ mM.

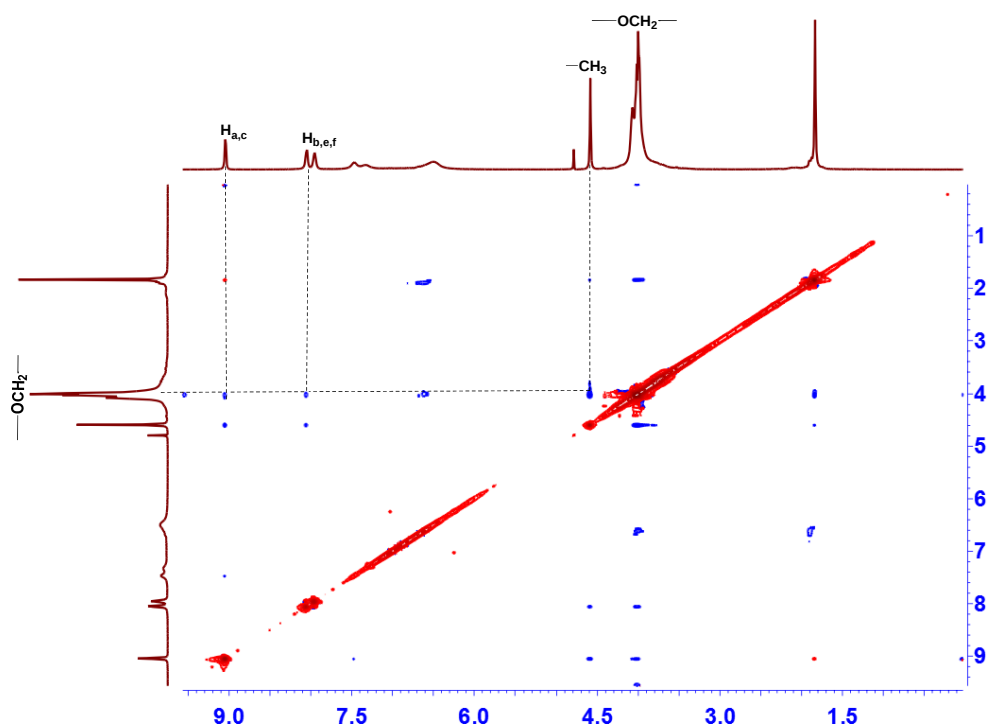


Fig. S15 Partial ^1H - ^1H ROESY spectrum (600 MHz, D_2O , 295 K) of a solution of host **1** and 2.0 equiv of guest **2**. $[\mathbf{1}]_0 = 8.0$ mM.

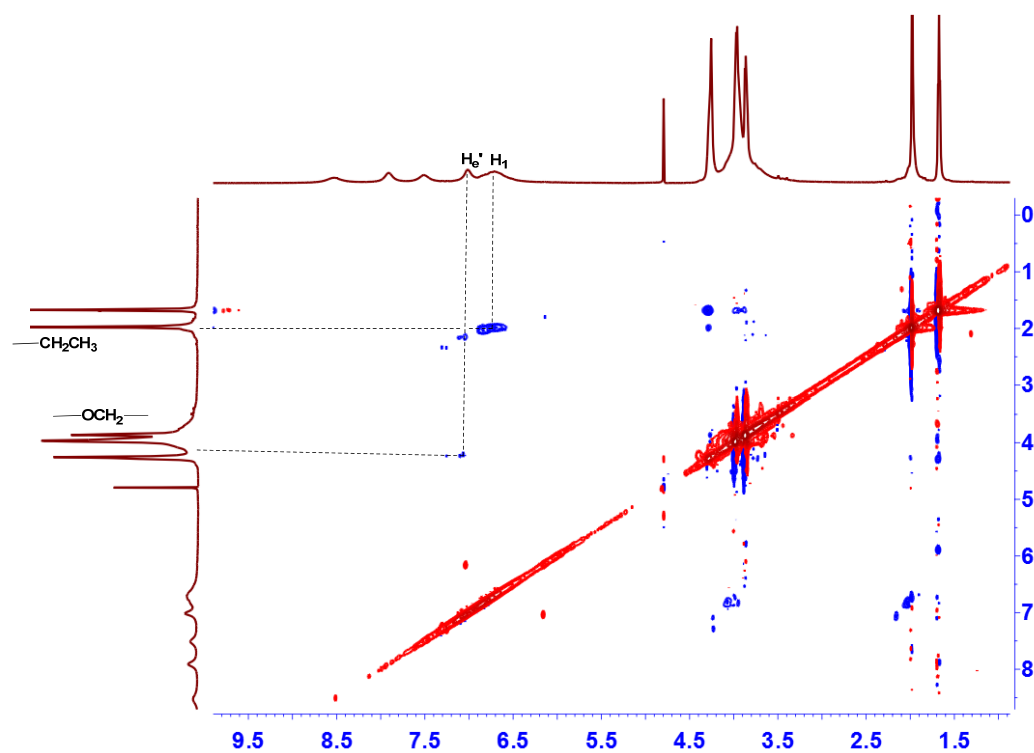


Fig. S16 Partial ^1H - ^1H ROESY spectrum (600 MHz, D_2O , 295 K) of a solution of host **1** and 2.0 equiv of guest **3**. $[\mathbf{1}]_0 = 8.0$ mM.

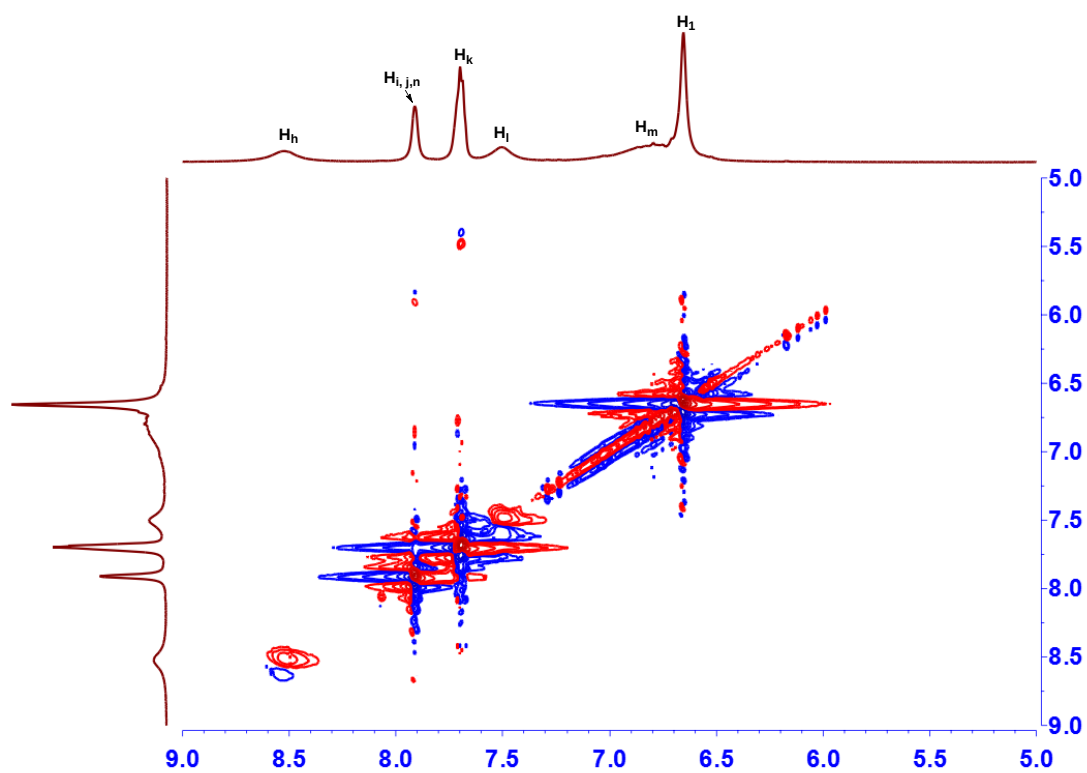


Fig. S17 Partial ^1H - ^1H COSY spectrum (600 MHz, D_2O , 295 K) of a solution of host **1** and 2.0 equiv of guest **4**. $[\mathbf{1}]_0 = 8.0$ mM.

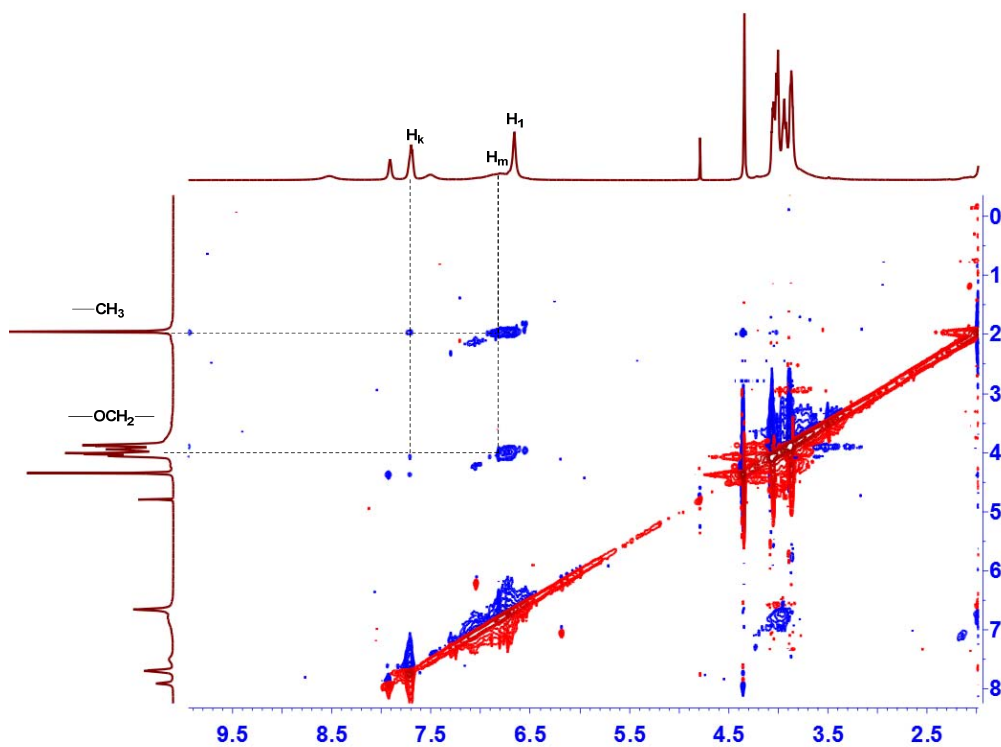


Fig. S18 Partial ^1H - ^1H ROESY spectrum (600 MHz, D_2O , 295 K) of a solution of host **1** and 2.0 equiv of guest **4**. $[\mathbf{1}]_0 = 8.0 \text{ mM}$.

8. ^1H NMR, ^{13}C NMR and HRMS spectra of new compounds

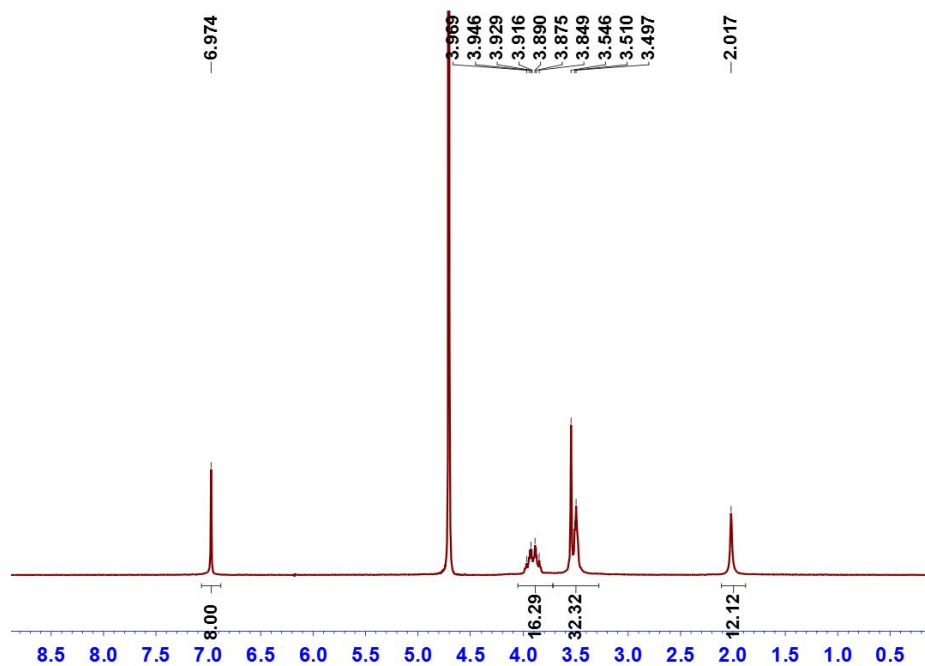


Fig. S19 ^1H NMR spectrum (300 MHz, D_2O , 295 K) of host **1**.

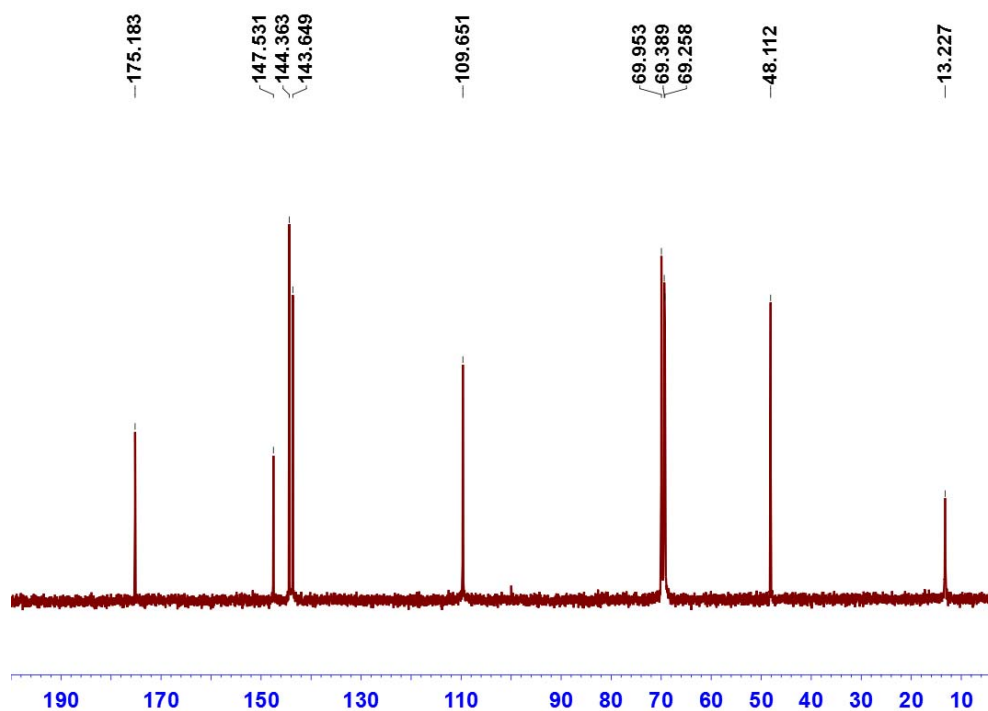


Fig. S20 ^{13}C NMR spectrum (75 MHz, D_2O , 295 K) of host **1**.

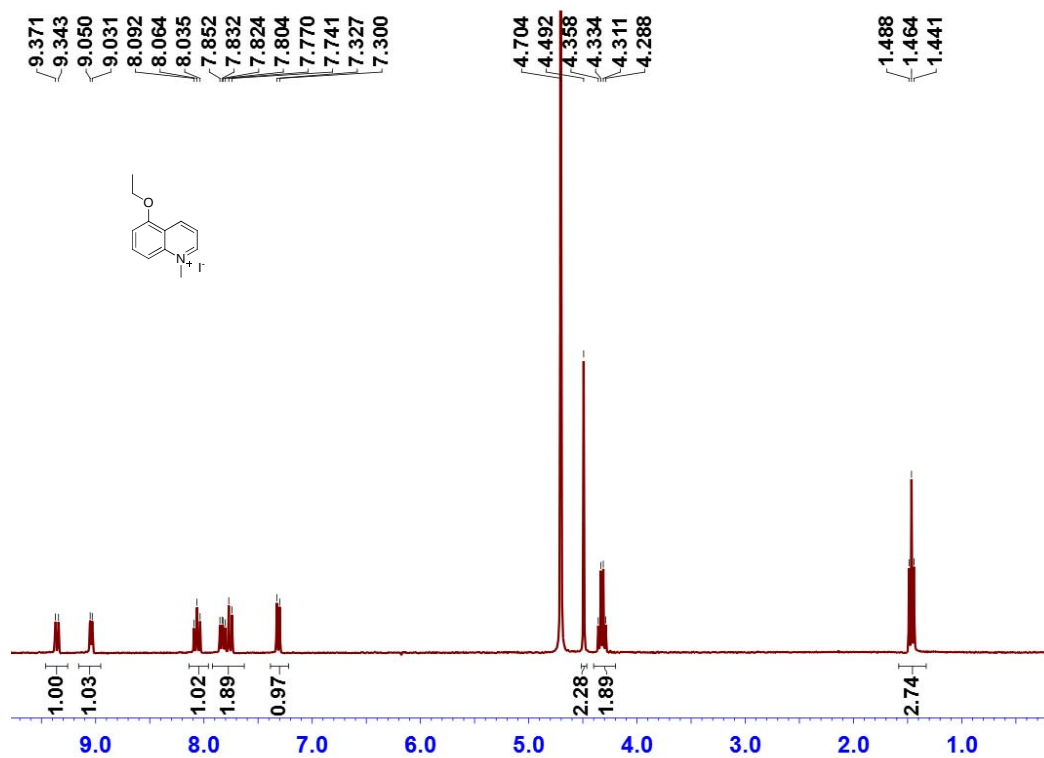


Fig. S21 ^1H NMR spectrum (300 MHz, D_2O , 295 K) of guest **3**.

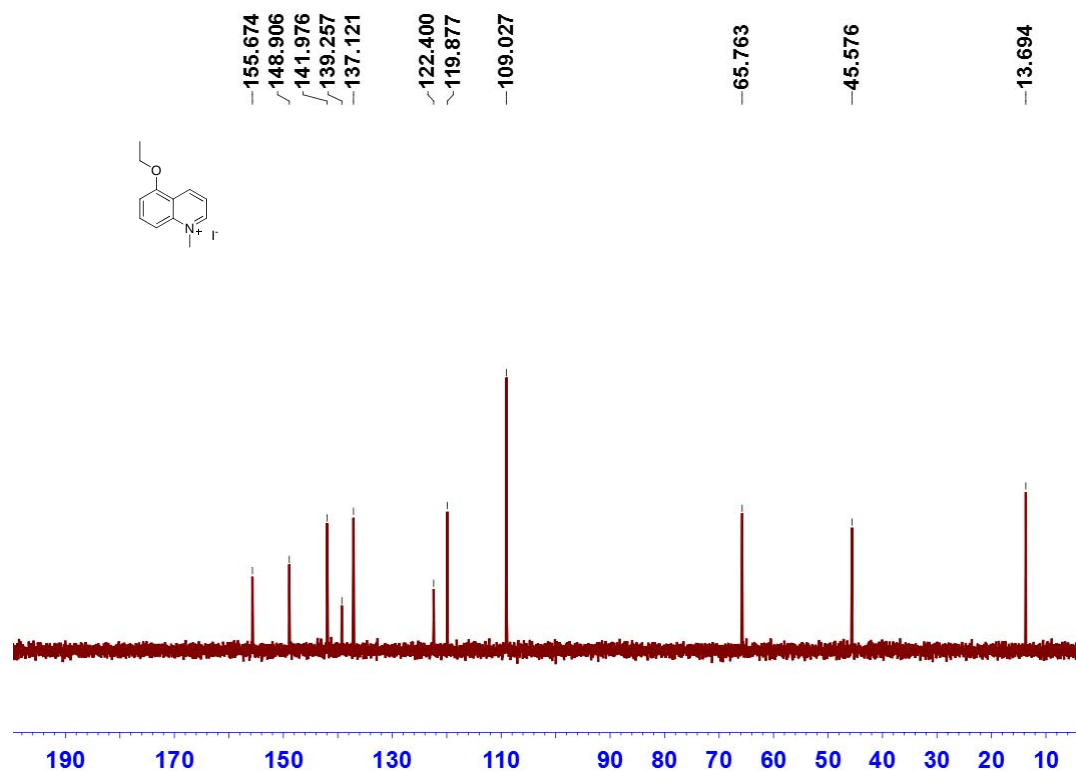


Fig. S22 ¹³C NMR spectrum (75 MHz, D₂O, 295 K) of guest **3**.

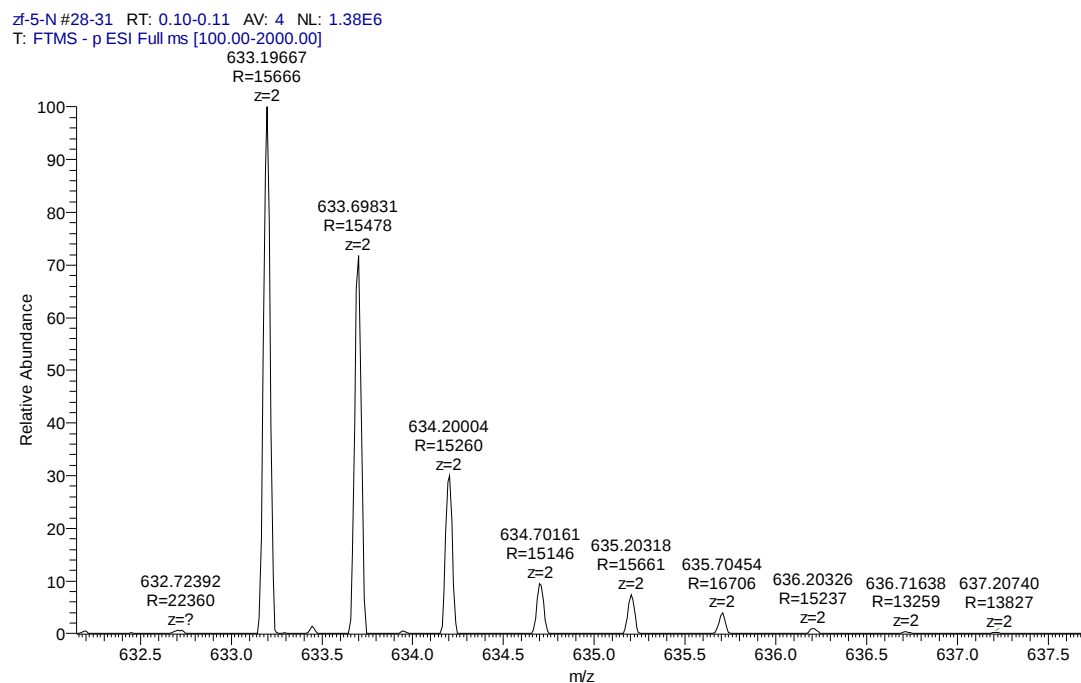


Fig. S23 HRESI mass spectrum of host **1**.

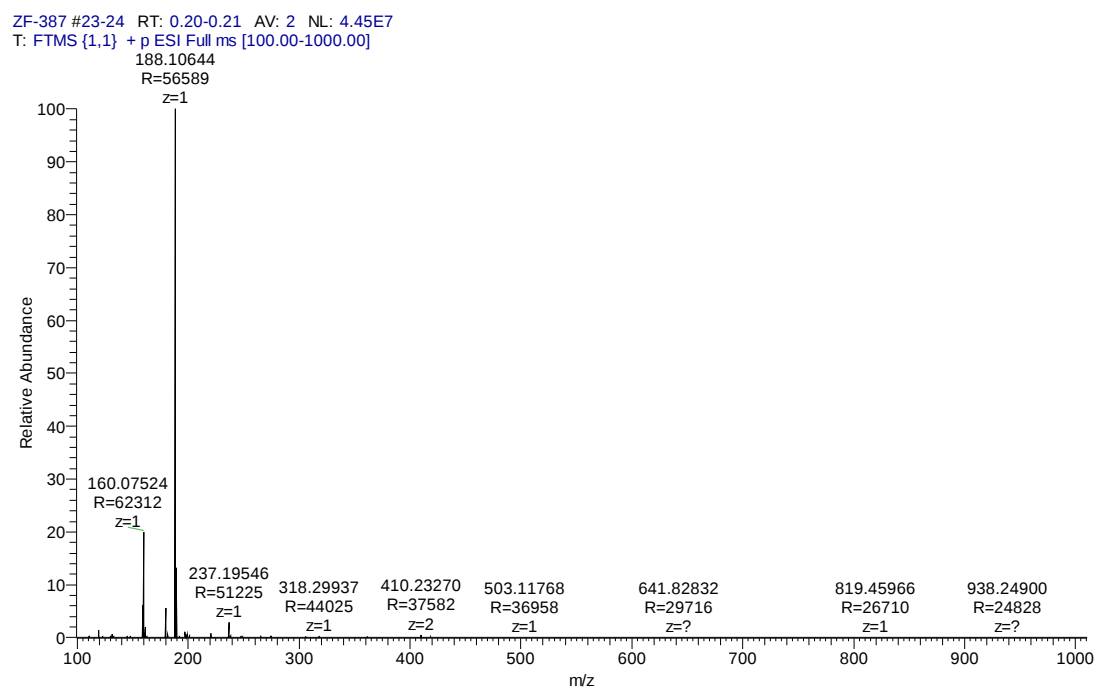


Fig. S24 HRESI mass spectrum of guest **3**.

9. UV/Vis spectra of complexes **1·2₂**, **1·3₂** and **1·4₂**

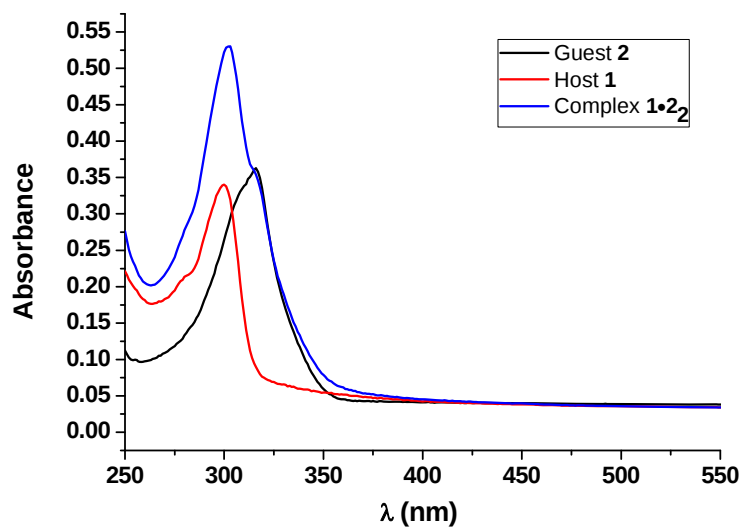


Fig. S25 UV/Vis absorption spectra of host **1**, guest **2** and complex **1·2₂**. $[1] = 2.0 \times 10^{-5}$ M.

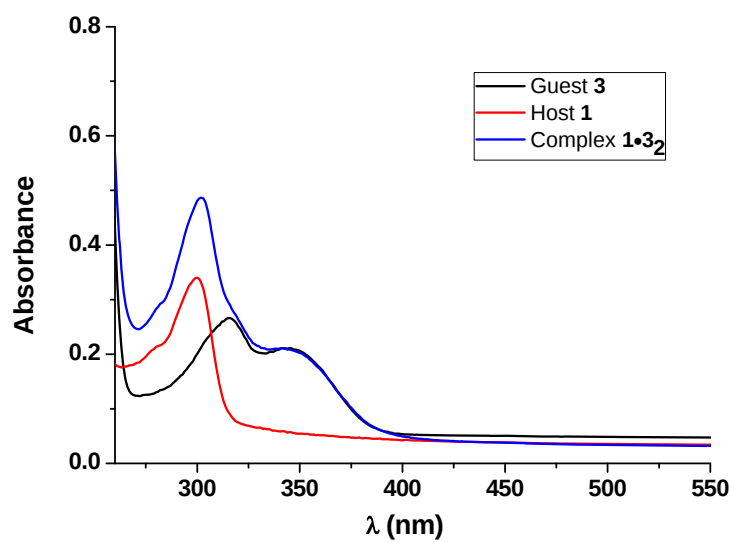


Fig. S26 UV/Vis absorption spectra of host **1**, guest **3** and complex **1·3₂**. [**1**]= 2.0×10^{-5} M.

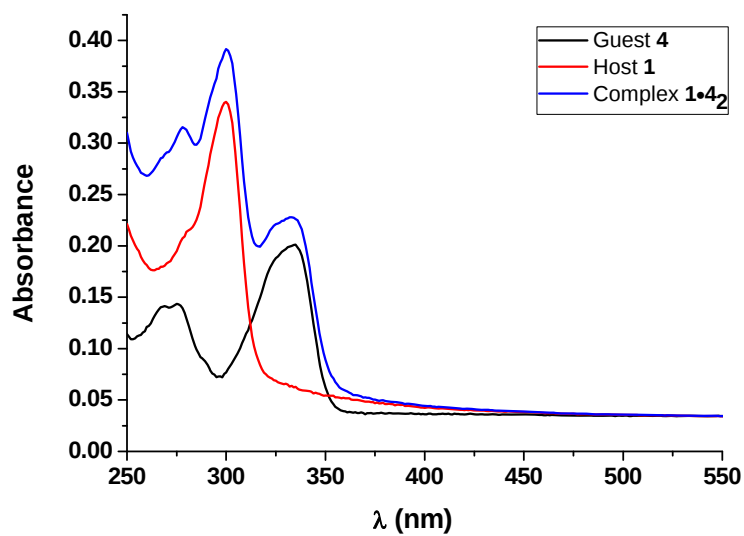


Fig. S27 UV/Vis absorption spectra of host **1**, guest **3** and complex **1·3₂**. [**1**]= 2.0×10^{-5} M.

10. The ^1H -NMR titrations spectra of complexes $1\cdot 2_2$, $1\cdot 3_2$ and $1\cdot 4_2$

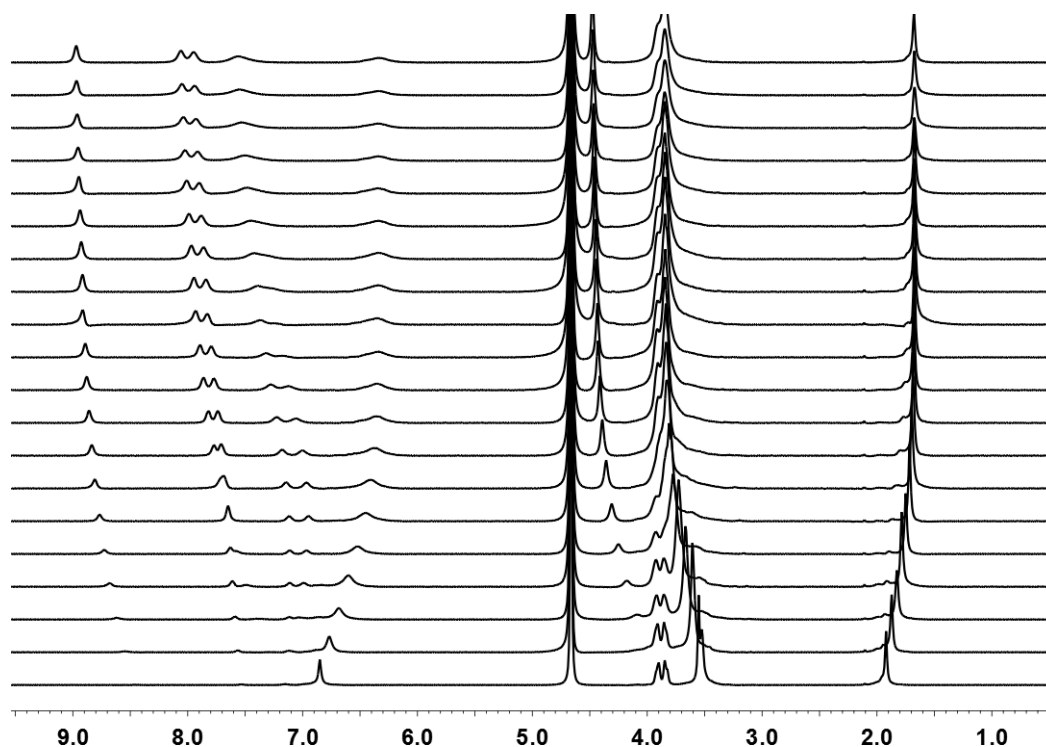


Fig. S28 ^1H -NMR (500 MHz, D_2O , 295 K) titrations spectra of guest **2** to host **1**, From bottom to top the addition of guest **2** are 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0, 4.2 equivalent of **1**. [**1**] = 3.0 mM.

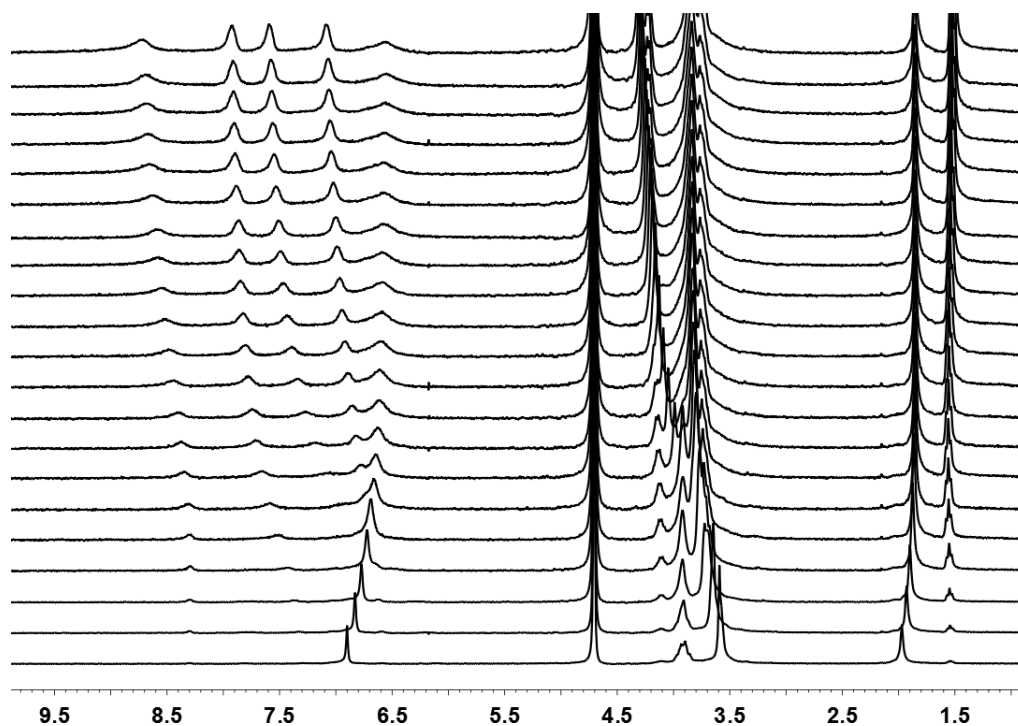


Fig. S29 ^1H -NMR (300 MHz, D_2O , 295 K) titrations spectra of guest **3** to host **1**, From bottom to top the addition of guest **3** are 0.2, 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0, 4.2 equivalent of **1**. [**1**] = 3.0 mM.

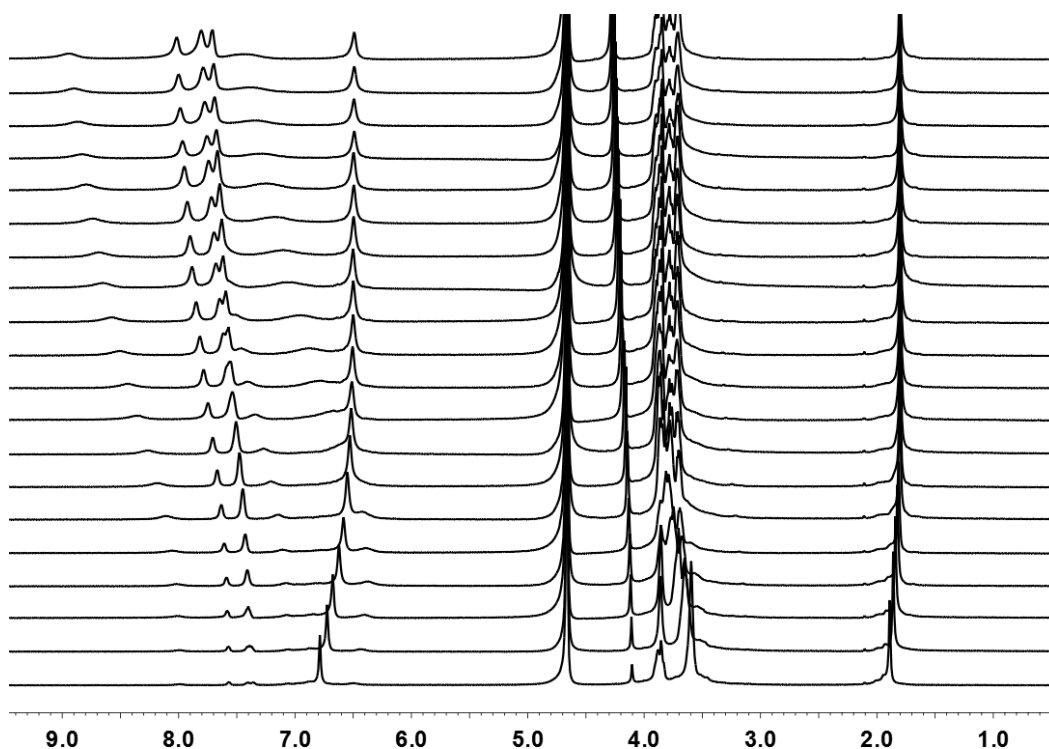


Fig. S30 ^1H -NMR (500 MHz, D_2O , 295 K) titrations spectra of guest **4** to host **1**, From bottom to top the addition of guest **4** are 0.4, 0.6, 0.8, 1.0, 1.2, 1.4, 1.6, 1.8, 2.0, 2.2, 2.4, 2.6, 2.8, 3.0, 3.2, 3.4, 3.6, 3.8, 4.0, 4.2 equivalent of **1**. $[\mathbf{1}] = 3.0 \text{ mM}$.

11. The Job-Plot for complexes $\mathbf{1}\cdot\mathbf{2}_2$, $\mathbf{1}\cdot\mathbf{3}_2$ and $\mathbf{1}\cdot\mathbf{4}_2$

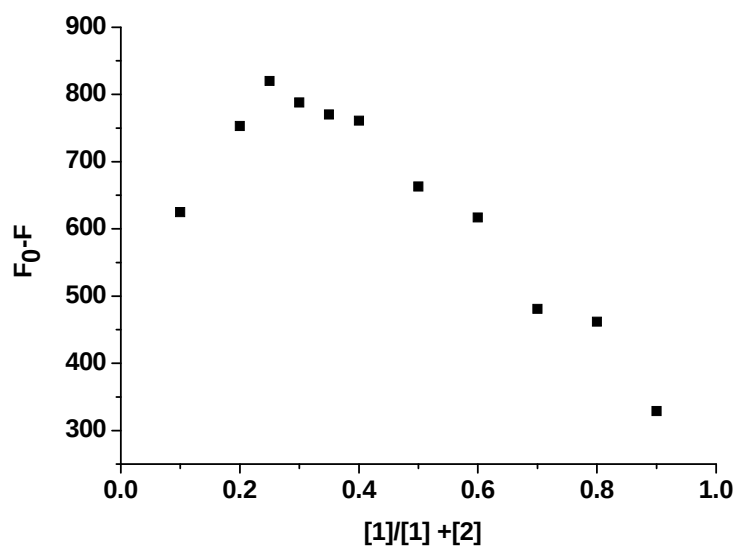


Fig. S31 The Job plot for $\mathbf{1}\cdot\mathbf{2}_2$ complex in water ($[\mathbf{1}] = [\mathbf{2}]$, $[\mathbf{1}] + [\mathbf{2}] = 8 \times 10^{-5} \text{ M}$, $\lambda_{\text{ex}} = 316 \text{ nm}$).

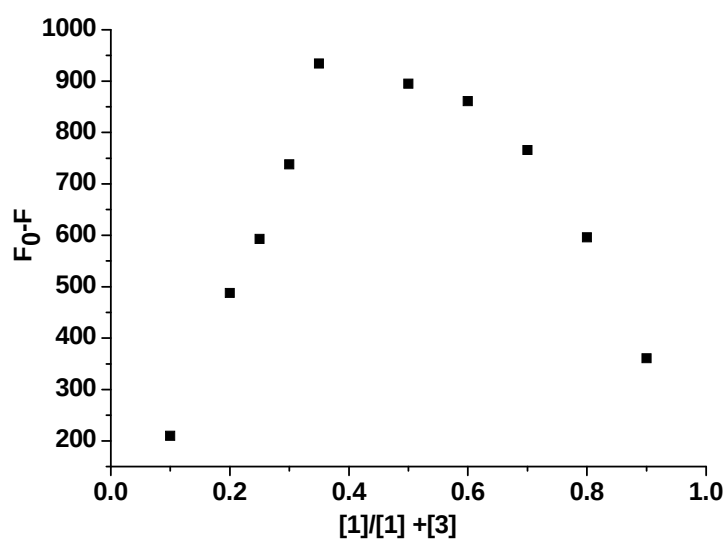


Fig. S32 The Job plot for $1 \cdot 3_2$ complex in water ($[1]=[3]$, $[1]+[3]=8 \times 10^{-5}$ M, $\lambda_{\text{ex}} = 316$ nm).

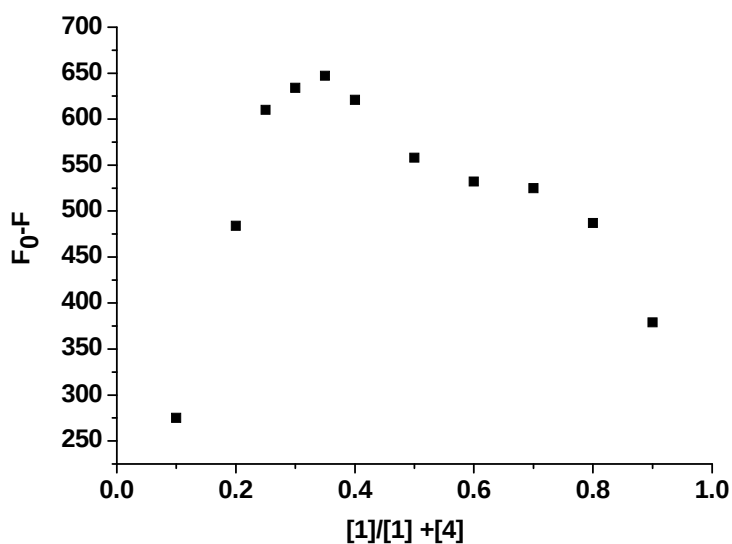


Fig. S33 The Job plot for $1 \cdot 4_2$ complex in water ($[1]=[4]$, $[1]+[4]=8 \times 10^{-5}$ M, $\lambda_{\text{ex}} = 320$ nm).

12. Crystal data

Empirical formula	C84 H90 N2 O24
Formula weight	1511.57
Temperature	173.1500 K
Wavelength	0.71073 Å
Crystal system	Monoclinic

Space group	P 1 21/n 1	
Unit cell dimensions	a = 15.814(4) Å	$\alpha = 90^\circ$.
	b = 14.779(3) Å	$\beta = 96.603(4)^\circ$.
	c = 21.657(5) Å	$\gamma = 90^\circ$.
Volume	5027.9(19) Å ³	
Z	2	
Density (calculated)	0.998 Mg/m ³	
Absorption coefficient	0.073 mm ⁻¹	
F(000)	1600	
Crystal size	0.26 x 0.17 x 0.15 mm ³	
Theta range for data collection	1.672 to 27.482°.	
Index ranges	-20 ≤ h ≤ 20, -19 ≤ k ≤ 19, -28 ≤ l ≤ 26	
Reflections collected	40163	
Independent reflections	11480 [R(int) = 0.0638]	
Completeness to theta = 26.000°	99.8 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	1.0000 and 0.6554	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	11480 / 402 / 580	
Goodness-of-fit on F ²	1.622	
Final R indices [I > 2sigma(I)]	R1 = 0.1131, wR2 = 0.2870	
R indices (all data)	R1 = 0.1401, wR2 = 0.3004	
Extinction coefficient	n/a	
Largest diff. peak and hole	1.069 and -0.600 e.Å ⁻³	

13. References

- [S1] U. Asseline, M. Chassignol, Y. Aubert and V. Roig, *Org. Biomol. Chem.*, 2006, **4**, 1949.
- [S2] J. Blagg, S. J. Coote, S. G. Davies and B. E. Mobbs, *J. Chem. Soc., Perkin Trans. I*, 1986, 2257.
- [S3] Y. S. Su, J. W. Liu, Y. Jiang and C.-F. Chen, *Chem. Eur. J.*, 2011, **17**, 2435.
- [S4] M.-H. Son, J. Y. Kim, E. J. Lim, D.-J. Baek, K. Choi, J. K. Lee, A. N. Pae, S.-J. Min and Y. S. Cho, *Bioorg. Med. Chem. Lett.*, 2013, **23**, 1472.