Supporting Information (SI)

Crocrassins A and B, two novel sesquiterpenoids with an unprecedented carbon skeleton from *Croton crassifolius*

Zhan-Xin Zhang,*^{*a*} Hui-Hong Li,^{*a*} Feng-Ming Qi,^{*b*} Le-Le Dong,^{*a*} Yang Hai,^{*a*} Gai-Xia Fan^{*a*} and Dong-Qing Fei*^{*a*}

^aSchool of Pharmacy, Lanzhou University, Lanzhou 730000, People's Republic of China. E-mail: <u>zhangzhx@lzu.edu.cn</u> (Z.X. Zhang); <u>feidq@lzu.edu.cn</u> (D.Q. Fei); Fax: +86 931 8915686; Tel: +86 931 8915686

^bState Key Laborratory of Applied Organic Chemistry, College of Chemistry and Chemical Engineering, Lanzhou University, Lanzhou 730000, People's Republic of China

TABLE OF CONTENTS

1. Detailed experimental procedures	S3
1.1 General Experimental Procedures	S3
1.2 Plant Material	S3
1.3 Extraction and isolation	S3
1.4 Chemical transformations	S3
1.4.1 Preparation of methyl ester derivative 2 of compound 1	S3
2. NMR, HRESIMS, and IR spectra of compounds 1–2	S4
Figure S1. ¹ H NMR spectrum of crocrassin A (1) in CDCl ₃	S4
Figure S2. ¹³ C NMR spectrum of crocrassin A (1) in CDCl ₃	S5
Figure S3. HMQC spectrum of crocrassin A (1) in CDCl ₃	S6
Figure S4. HMBC spectrum of crocrassin A (1) in CDCl ₃	S7
Figure S5. ¹ H- ¹ H COSY spectrum of crocrassin A (1) in CDCl ₃	S8
Figure S6. NOESY spectrum of crocrassin A (1) in CDCl ₃	S9
Figure S7. HRESIMS spectrum of crocrassin A (1)	S10
Figure S8. IR spectrum of crocrassin A (1)	S11
Figure S9. ¹ H NMR spectrum of crocrassin B (2) in CDCl ₃	S12
Figure S10. ¹³ C NMR spectrum of crocrassin B (2) in CDCl ₃	S13
Figure S11. HSQC spectrum of crocrassin B (2) in CDCl ₃	S14
Figure S12. HMBC spectrum of crocrassin B (2) in CDCl ₃	S15
Figure S13. ¹ H- ¹ H COSY spectrum of crocrassin B (2) in CDCl ₃	S16
Figure S14. NOESY spectrum of crocrassin B (2) in CDCl ₃	S17
Figure S15. HRESIMS spectrum of crocrassin B (2)	S18
Figure S16. IR spectrum of crocrassin B (2)	S19

1. Detailed experimental procedures

1.1 General Experimental Procedures

Melting points were determined on an X-4 digital display micromelting point apparatus, and are uncorrected. Optical rotations were measured on a Perkin Elmer 341 polarimeter. IR spectra were taken on a Nicolet NEXUS 670 FT-IR spectrometer. NMR spectra were recorded on a Varian INOVA-600 NMR spectrometer with TMS as internal standard. HRESIMS data were recorded on a Thermo LTQ Orbitrap Elite mass spectrometer. Sephadex LH-20 was supplied by Amersham Pharmacia Biotech. Silica gel (200-300 mesh) used for column chromatography and silica gel GF₂₅₄ (10-40 μ M) used for TLC were supplied by the Qingdao Marine Chemical Factory, Qingdao, China. Spots were detected on TLC under UV light or by heating after spraying with 5% H₂SO₄ in C₂H₅OH (v/v).

1.2 Plant Material

The roots of *C. crassifolius* were purchased from Hebei Anguo Medicine Market, and were originally collected from Fujian province of China in September 2012. The plant was identified by Dr. Jian-Yin Li (School of Pharmacy, Lanzhou University, Lanzhou, China). A voucher specimen (No. 201209CC) was deposited at the School of Pharmacy, Lanzhou University.

1.3 Extraction and isolation

The air-dried and powdered roots of *C. crassifolius* (9.5 kg) were extracted four times in 95% EtOH at room temperature. The filtrate was combined and concentrated under reduced pressure to afford a residue (962 g). The residue was suspended in H_2O and extracted with EtOAc, and n-BuOH, successively. The EtOAc extract (731 g) was subjected to CC over silica gel eluting with a petroleum ether-acetone step gradient system (40:1 to 0:1) to give fractions A-F. Fraction C was chromatographed over silica gel column, eluted with a gradient solvent system of increasing polarity (petroleum ether-acetone, 30:1 to 3:1), yielding four subfractions C1-C4. Subfraction C1 was subjected to repeated chromatography over silica gel (petroleum ether-EtOAc, 40:1 to 3:1) to give subfractions C1A-C1C. Subfraction C1B was purified by silica gel CC (CHCl₃-EtOAC, 80:1 to 20:1) followed by gel permeation chromatography (GPC) on Sephadex LH-20 in CHCl₃-MeOH (1:1) to furnish compound **2** (2 mg). Subfractions C2A-C2C. Subfarction C2C was further purified by silica gel CC (CHCl₃-acetone, 10:1 to 2:1) and Sephadex LH-20 (CHCl₃-MeOH, 1:1) to afford compound **1** (4 mg).

1.4 Chemical transformations

1.4.1 Preparation of methyl ester derivative 2 of compound 1

To a solution of 1 (2 mg) in diethyl ether (0.5 mL), an ethereal solution of diazomethane was added dropwise until persistence of yellow color. The solution was set aside for 10 min, the excess of diazomethane was destroyed by slowly adding dropwise a 5% ethereal solution of acetic acid (yellow color disappearance) and the solvent evaporated to yield 2 (2 mg).

2. NMR, HRESIMS, and IR spectra of compounds 1–2

Figure S1. ¹H NMR spectrum of crocrassin A (1) in CDCl₃













Figure S5. ¹H-¹H COSY spectrum of crocrassin A (1) in CDCl₃



Figure S6. NOESY spectrum of crocrassin A (1) in CDCl₃



Figure S7. HRESIMS spectrum of crocrassin A (1)



Figure S8. IR spectrum of crocrassin A (1)

Figure S9. ¹H NMR spectrum of crocrassin B (2) in CDCl₃



H₃COOC 200 Η ۰, ŌH 180 -172.059 160 147.217 140 120 __111.009 100 81.588 77.211 77.000 76.789 80 60 _55.613 ____51.224 ____48.061 41.786 38.488 40 29.697 29.071 28.344 28.170 25.143 25.016 18.496 16.242 20 bbw 蒹

Figure S10. ¹³C NMR spectrum of crocrassin B (2) in CDCl₃



Figure S11. HSQC spectrum of crocrassin B (2) in CDCl₃



Figure S12. HMBC spectrum of crocrassin B (2) in CDCl₃







Figure S16. IR spectrum of crocrassin B (2)



S19