

# Ecotoxicology of Nanomaterials - Finding the best way forward

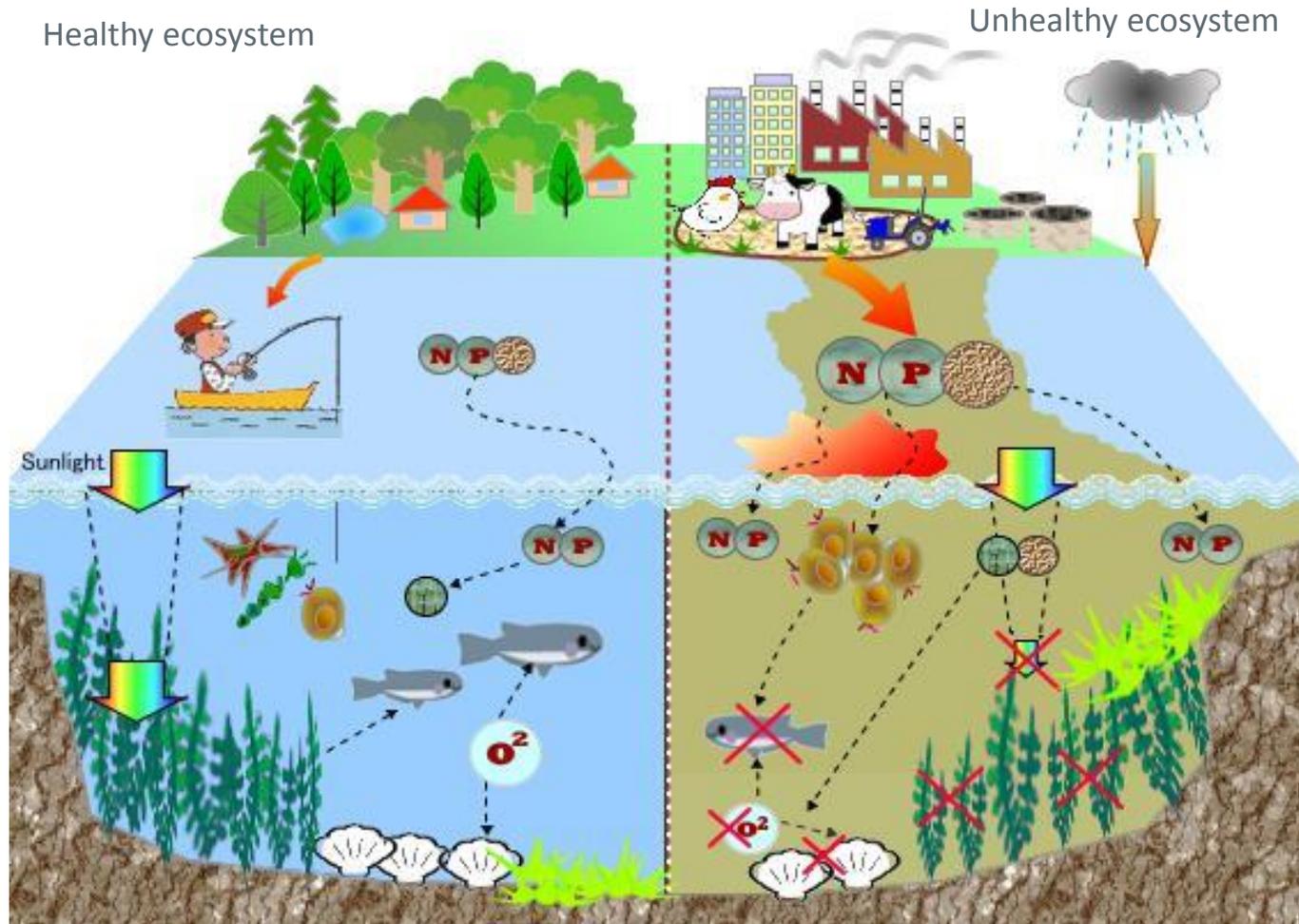
*Teresa F Fernandes*  
*t.fernandes@hw.ac.uk*



V. Stone Toxicology Award  
19<sup>th</sup> April 2016

- Why consider the environment
- Nanomaterials through their life cycle
- Early investigations of effects of nanomaterials in the environment
- Increasing realism – link to industry and providing the environmental context
- Cross species studies
- Safety by design
- Using toxicity information in decision making
- Acknowledgements

# Why should we care about the Environment?



# Nanomaterials – A risk in the environment?

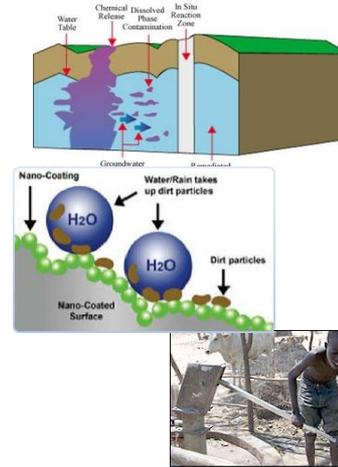
Risk = Hazard x Exposure  
(Toxicity)

# Nanotechnology Applications

## Energy



- More efficient and cost effective technologies for energy production
  - Solar cells
  - Fuel cells
  - Batteries
  - Bio fuels
- More energy efficient computers



## Environmental applications

- Energy efficiencies
- Waste remediation
- Removal of contaminants from water
- Desalination
- Reducing car emissions, NO<sub>x</sub> conversion

## Medicine



- Cancer treatment
- Bone treatment
- Drug delivery
- Appetite control
- Drug development
- Medical tools
- Diagnostic tests
- Imaging

## Consumer Goods

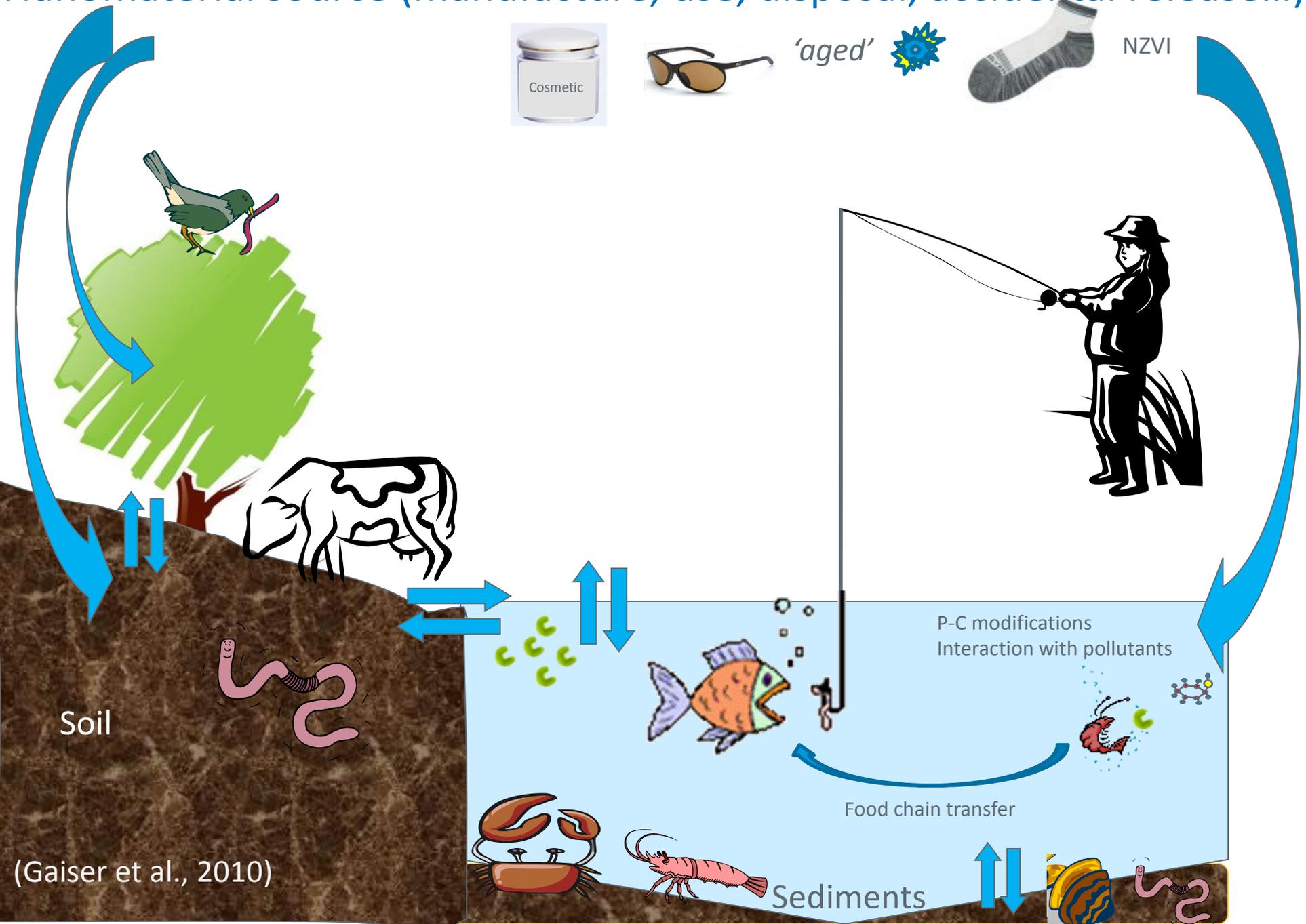


- Foods and beverages
  - Advanced packaging materials, sensors, and lab-on-chips for food quality testing
- Appliances and textiles
  - Stain proof, water proof and wrinkle free textiles
- Household and cosmetics
  - Self-cleaning and scratch free products, paints and cosmetics

# Exposure / fate



# Nanomaterial source (manufacture, use, disposal, accidental release...)



'aged'



NZVI

Soil

P-C modifications  
Interaction with pollutants

Food chain transfer

Sediments

(Gaiser et al., 2010)

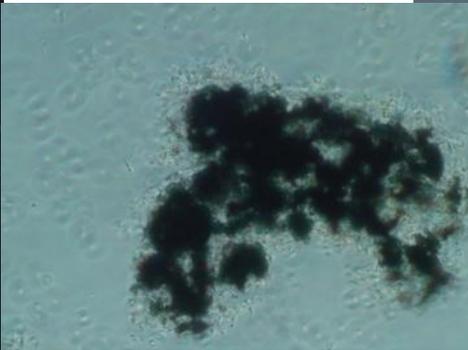
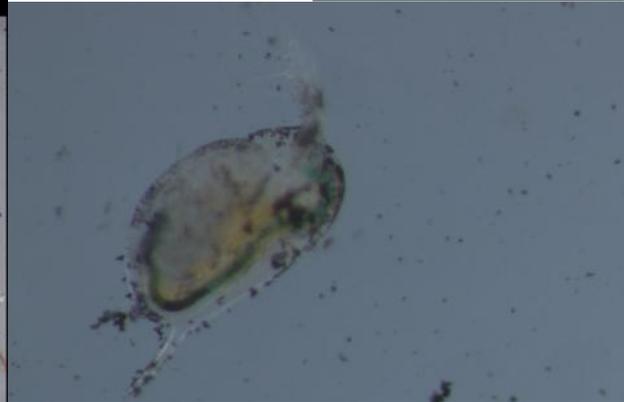
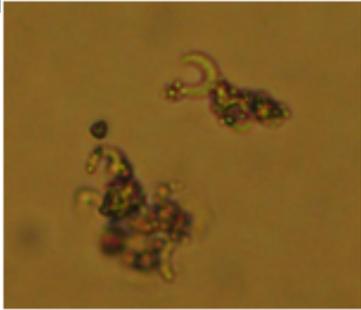
# Ecotoxicology- toxicology integration

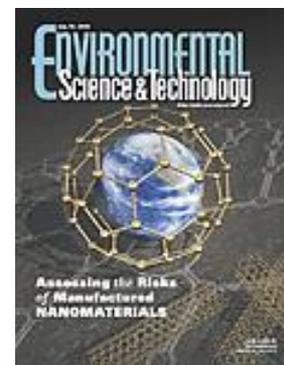
- Since more toxicological studies have been completed, the information gained from the toxicology can be used to inform ecotoxicology.
  
- The main findings of the toxicology can be broken down into two general areas:
  - (i) Physical and chemical characteristics
    - Size, surface area, dimensions,
    - solubility (biopersistence, durability),
    - aggregation/clumping, contaminants, composition.
  
  - (ii) Toxicological mechanisms
    - Free radical and reactive oxygen species production,
    - oxidative stress, inflammation, toxicokinetics (absorption, distribution, metabolism and excretion).

# Early nano ecotoxicology studies

- Back in the early 2000s it was becoming apparent that the nanotechnology sector was increasing at a very fast pace, with wide ranging applications
- The cross link between human nano toxicology and environmental nanotoxicology was obvious





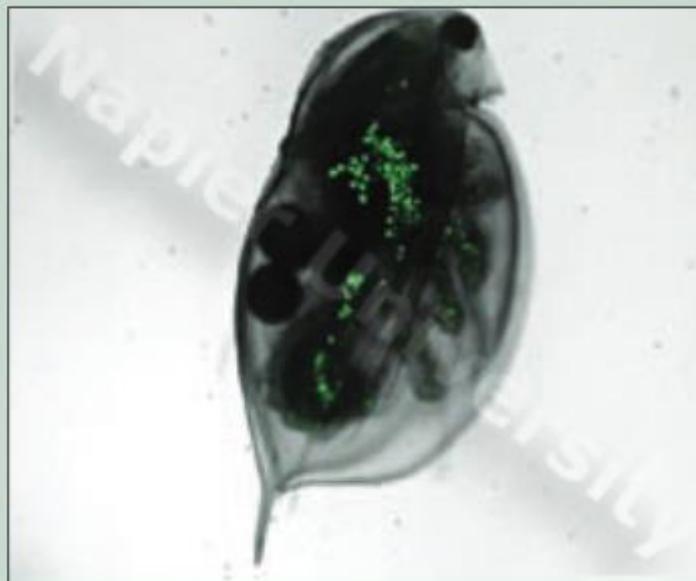


## Still life with nanoparticles

For the first time, researchers have captured an image of nanoparticles inside a whole, live organism. Nanoparticles have been photographed previously in cells *in vitro*, but this image, which was presented at the Society of Environmental Toxicology and Chemistry Europe meeting in May by Teresa Fernandes of Napier University (U.K.), captures the tiny particles inside a daphnid or water flea (*Daphnia magna*).

Fernandes and her team started with immature (neonates) and adult daphnids swimming in an environment containing nanoparticles of titanium dioxide and carbon black. Both the immature and adult water fleas ingested the particles immediately. Graduate student Philipp Rosenkranz captured images of the carboxylated, fluorescently tagged particles in an adult's gut as well as in its fatty-lipid storage droplets. A neonate, <1 d old, also ingested the particles, as documented in images that the team plans to publish later this year.

The researchers expected the particles to be readily seen in the adult animal's gut, but they wanted to determine whether the daphnid would excrete them immediately or store them elsewhere internally. "Within an hour of ingestion, one of the photographs indicated that the



TERESA FERNANDES, COPYRIGHT NAPIER UNIVERSITY (U.K.)

nanoparticles had been translocated to other parts of the body," Fernandes says. "They're going to use those storage deposits eventually," she says, "however, this requires further work to determine the relevance to nanoparticle safety." —NAOMI LUBICK



## **CORRIERE DELLA SERA / SALUTE**

» Corriere della Sera > Salute > *Dove vanno a finire le particelle?*



AMBIENTE ANCORA POCHE LE RICERCHE SULLA TOSSICITÀ DEI NANOMATERIALI PER GLI ORGANISMI VIVENTI

### **Dove vanno a finire le particelle?**

*Sono già usate nell'industria. Ma non si sa con quali conseguenze*

Nanoparticelle ben visibili in un organismo vivo. Le ha fotografate per la prima volta Teresa Fernandes della Napier University di Edimburgo all'interno di un crostaceo di acqua dolce molto impiegato negli studi sull'inquinamento delle acque, la *Daphnia magna*. La foto, presentata dalla ricercatrice all'incontro sui rischi delle nanotecnologie organizzato di recente a Bruxelles dalla Commissione europea, sembra testimoniare come la ricerca tossicologica oggi non riesca ad andare al di là di una «fotografia» del problema. Le nanoparticelle entrano all'interno delle cellule, questo è ormai dimostrato, ma che cosa significa in termini di rischio? «Non lo sappiamo — risponde Gunter Oberdorster, dell'università di Rochester, Stati Uniti, che al convegno belga ha tenuto una relazione in proposito —. Siamo a conoscenza del fatto che le nanoparticelle disperse nell'aria penetrano in profondità nell'albero respiratorio, possono venire assorbite, entrare in circolo e diffondersi in vari organi. Ma che cosa identifica la tossicità di una nanoparticella rispetto alla massa di quelle che vengono inalate?». Ma se la mancanza di test di laboratorio affidabili è ancora un problema per capire i

**F.P.**

**26 ottobre 2008**

## A COMPARISON OF NANOPARTICLE AND FINE PARTICLE UPTAKE BY *DAPHNIA MAGNA*

PHILIPP ROSENKRANZ,\*† QASIM CHAUDHRY,‡ VICKI STONE,† and TERESA F. FERNANDES†

†School of Life Sciences, Edinburgh Napier University, 10 Colinton Road, Edinburgh, EH10 5DT Scotland, United Kingdom

‡Central Science Laboratory, Sand Hutton, York YO41 1LZ, United Kingdom



**Abstract**—The use of nanoparticles in various applications is steadily on the rise, with use in a range of applications, including printer toner, sunscreen, medical imaging, and enhanced drug delivery. While research on human effects via, for example, inhalation is relatively well developed, the environmental assessment of nanoparticles is in its infancy. In the present study, we assessed the uptake and quantitative accumulation, as well as the depuration, of a model nanoparticle, a 20-nm fluorescent carboxylated polystyrene bead, in the aquatic invertebrate *Daphnia magna* and compared it to a larger, 1,000-nm particle. Using confocal microscopy, rapid accumulation in the gastrointestinal tract was observed within an hour of exposure to both particle sizes in both adults and neonates. Fluorescence could also be observed in the oil storage droplets, suggesting that both particle sizes have crossed the gut's epithelial barrier. Quantification of fluorescence of both sizes of particles showed that although uptake of the 20-nm particles was lower in terms of mass it was equal to or greater than 1,000-nm particle uptake when expressed as surface area or particle number. Depuration was relatively rapid for the 1,000-nm beads, decreasing by more than 90% over 4 h. In contrast, depuration of the 20-nm beads was less extensive, reaching 40% over 4 h. Transmission electron microscopy confirmed uptake of 1,000-nm beads, but uptake of 20-nm beads was inconclusive since similar-sized inclusions could be observed in control treatments.

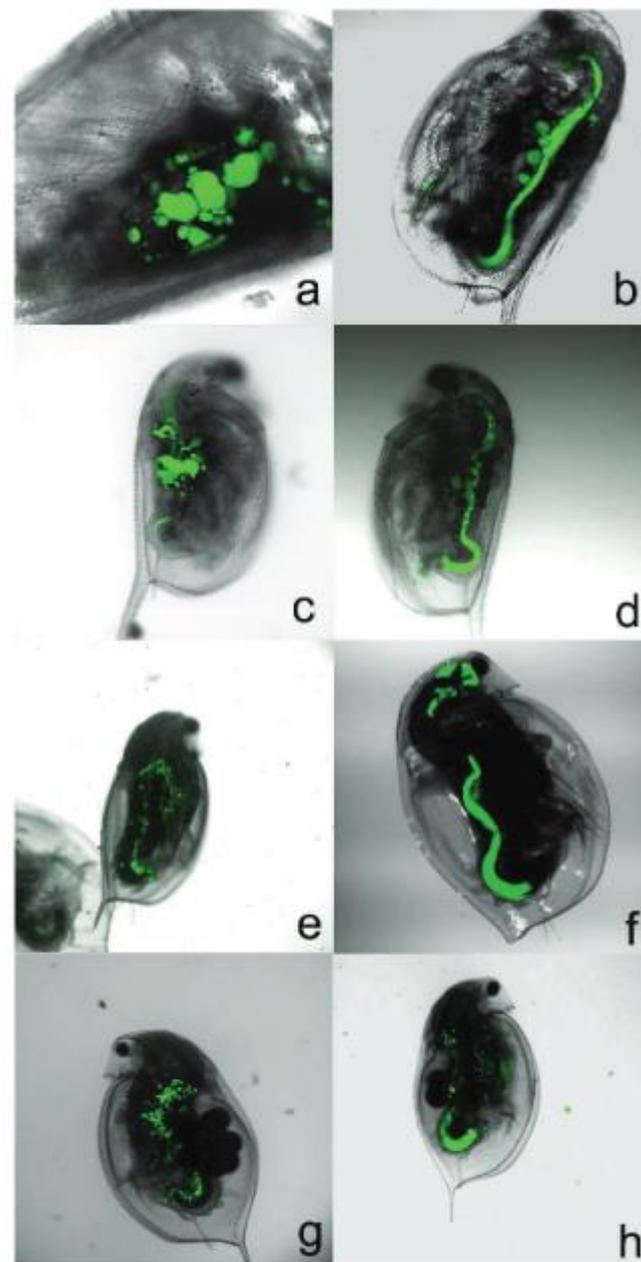
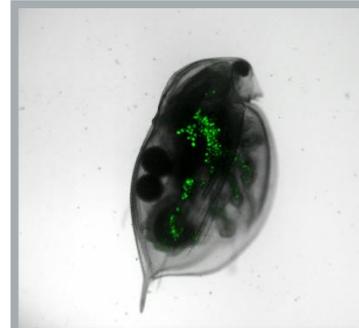
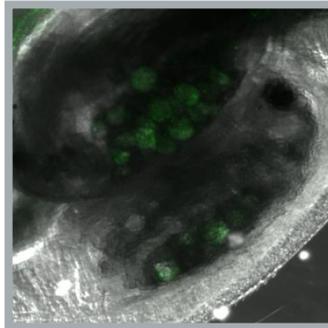
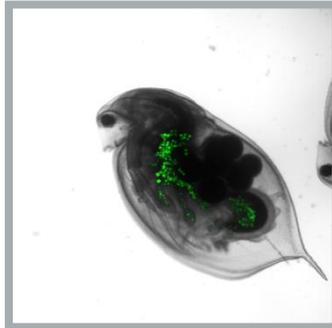
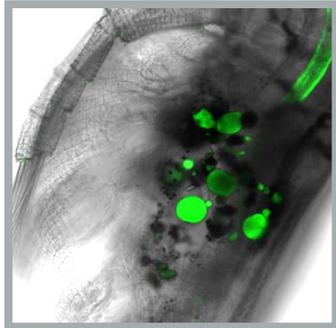
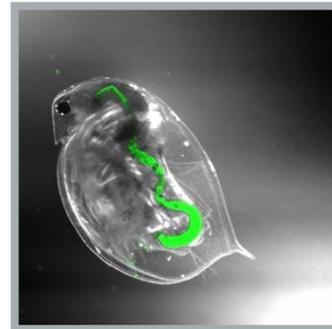
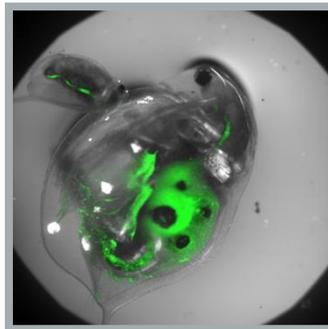
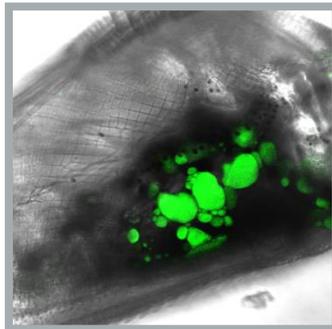


Fig. 1. Uptake of 20- (a, c, e, g) and 1,000-nm (b, d, f, h) fluorescein isothiocyanate-labeled polystyrene beads in neonate (<24 h old at start of experiment, a–d) and adult (e–h) *Daphnia magna* over 24 h as observed by confocal microscopy. Shown are pictures after 30-min (a, b, e, f) and 12-h exposure (c, d, g, h). Magnification was  $\times 200$  in panel a,  $\times 100$  in panels b to d, and  $\times 25$  in panels e to h.

# Size Dependent Particle Uptake by *D. magna* (observed via confocal microscopy)



20nm



1000nm

Rapid uptake of fluorescence in the gut and adjacent oil storage droplets can be observed.

(fluorescent green carboxylated nanospheres at 2.6 µg/L for 24 hrs)

Rosenkranz et al (2009)

# Comparing the effects of 14 nm and 260 nm carbon black particles

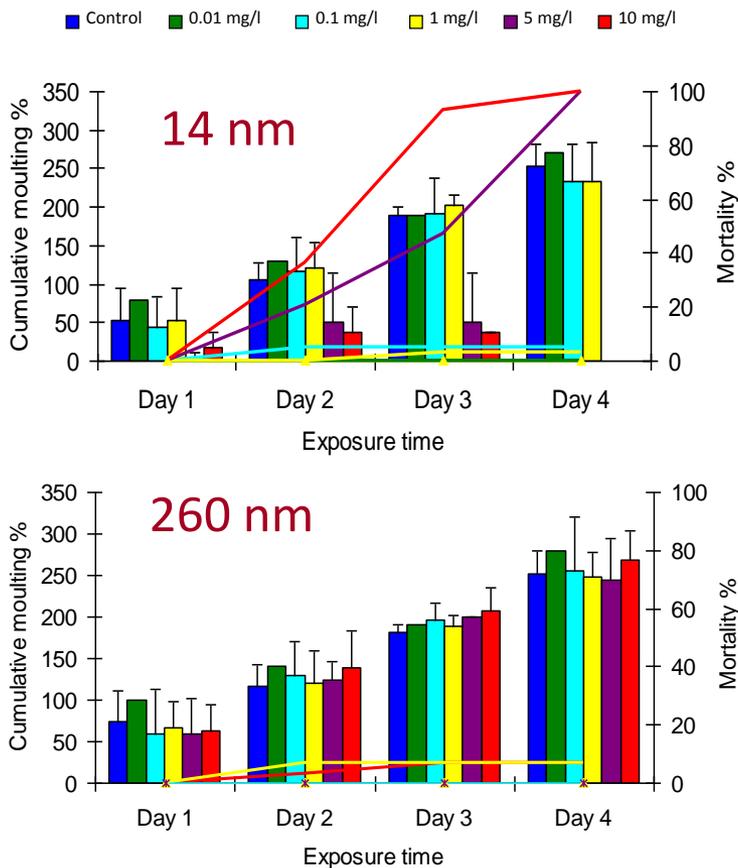


Fig.2: **Mortality (line)** and **cumulative moult** (column) after treatment with 14nm (top) and 260nm carbon black (bottom) in an acute, 96h exposure

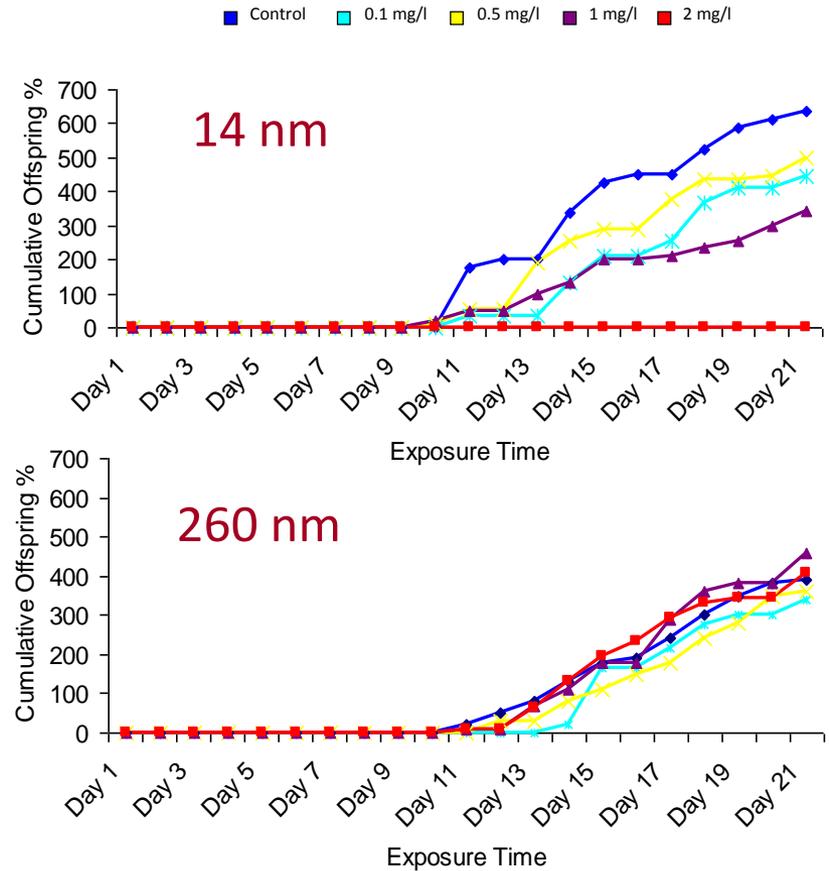


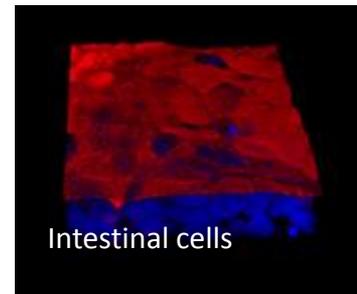
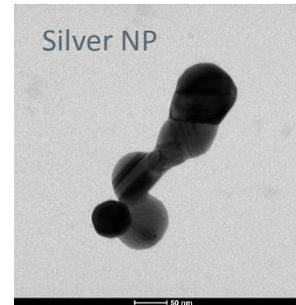
Fig.3: **Cumulative offspring** after treatment with 14nm (top) and 260nm carbon black in a chronic, 21 day exposure

# Assessing human exposure, uptake and toxicity of NPs from contaminated environments



## Project Aims:

To compare the uptake and relative sensitivity to nanoparticles of invertebrates, fish and human models



University of  
Birmingham



University of  
Bristol

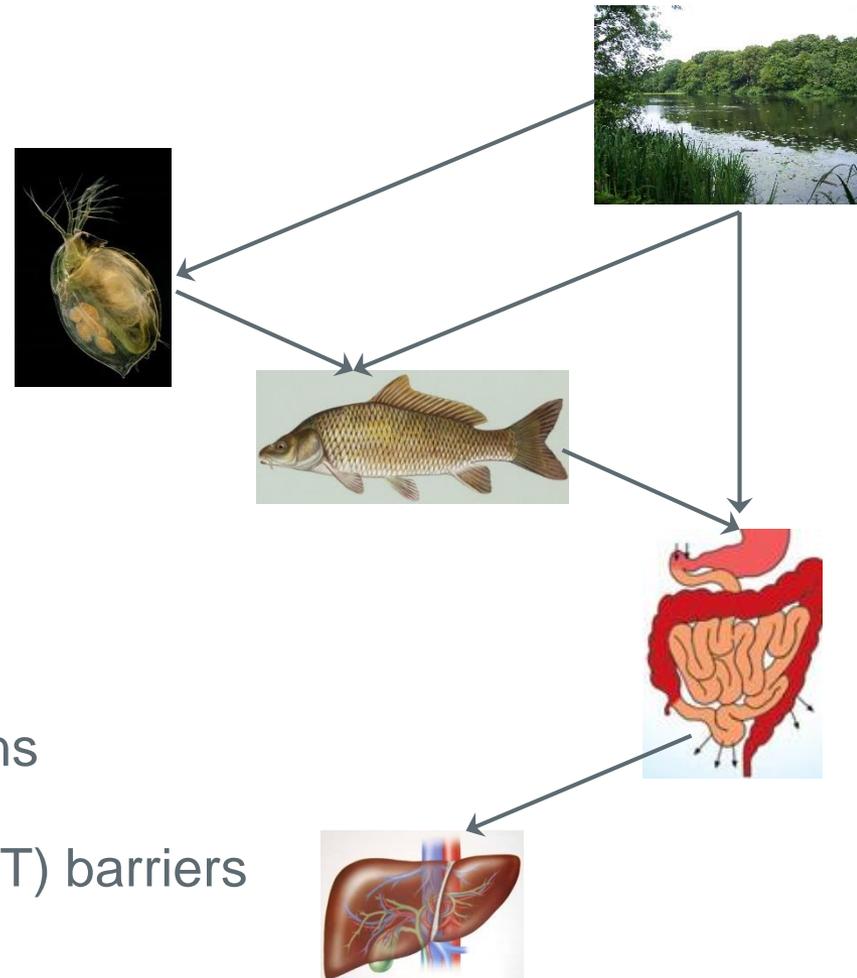


Joint Environment and  
Human Health  
Programme (UK)

# Assessing human exposure, uptake and toxicity of NPs from contaminated environments

Connecting ecotoxicology and toxicology of water-borne NP:

- Exposure of primary producers
- Exposure of invertebrates
- Exposure of fish
- Uptake into higher animals and humans
- Transport through gastro-intestinal (GIT) barriers
- Effects of NP in hepatocytes



# Study Approach

## System

1. Hepatocytes:  
Human (C3A), trout  
(*Oncorhynchus mykiss*)  
(primary)
2. Invertebrate - *D. magna*
3. Fish – carp (*Cyprinus carpio*)

## Endpoints

1. Cytotoxicity (membrane integrity assessment)
2. Mortality, growth, moulting
3. Bioavailability

Characterisation of particles in respective media or water:  
concentration, aggregation, solubility

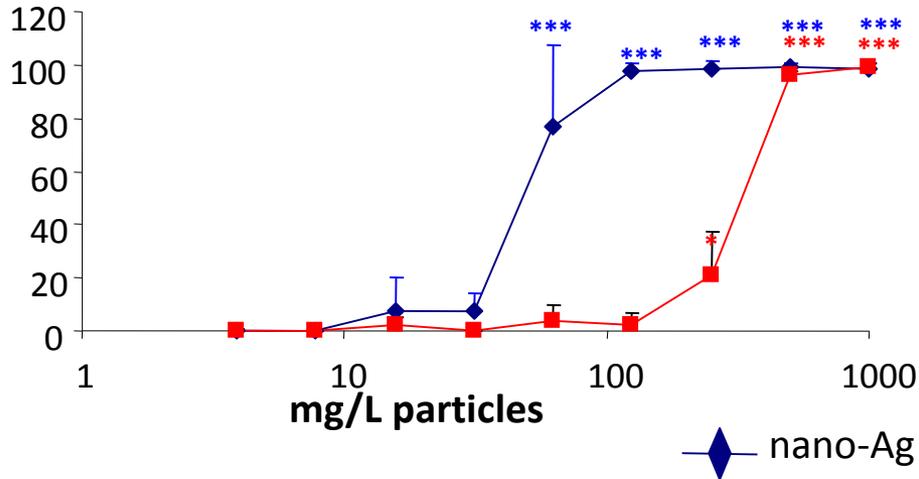
Assess transport through gastro-intestinal barriers

# Cytotoxicity (LDH\*) of Ag

## C3A human hepatocytes

## Primary trout hepatocytes

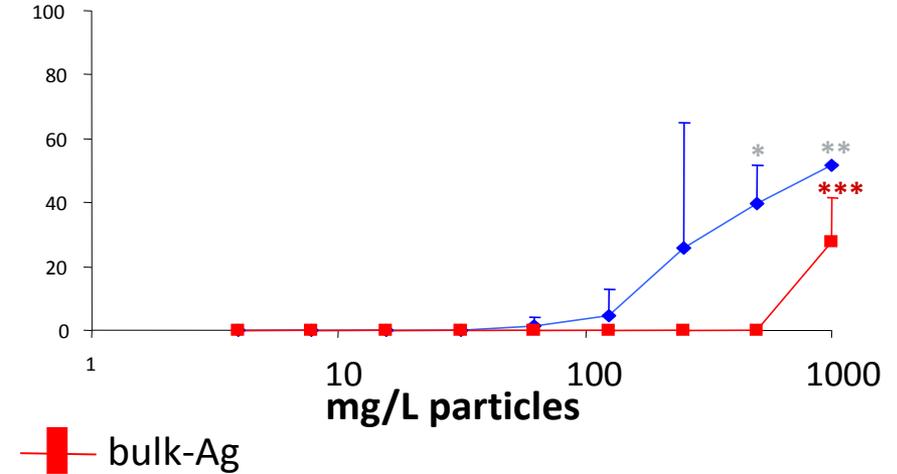
% cytotoxicity



$LD_{50} (Ag_{nano}) \approx 50 \text{ mg/L}$

$LD_{50} (Ag_{bulk}) \approx 300 \text{ mg/L}$

% cytotoxicity



$LD_{50} (Ag_{nano}) \approx 1000 \text{ mg/L}$

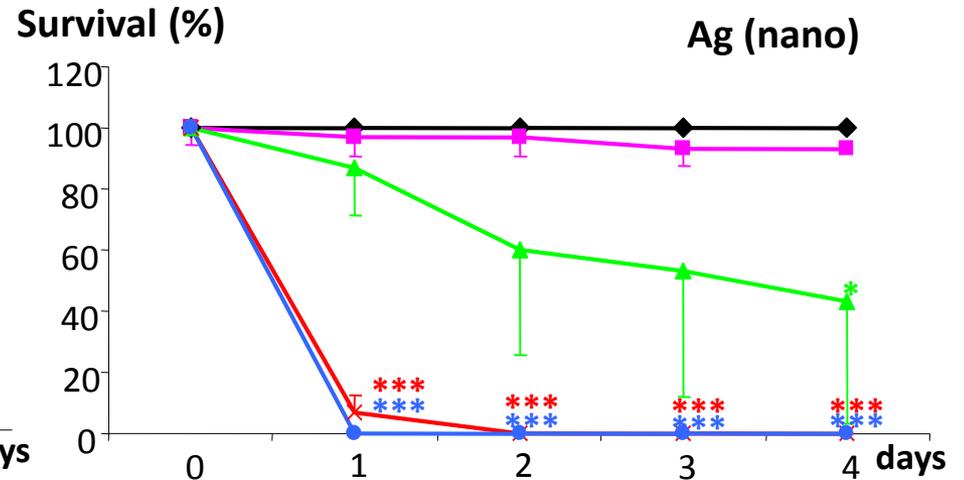
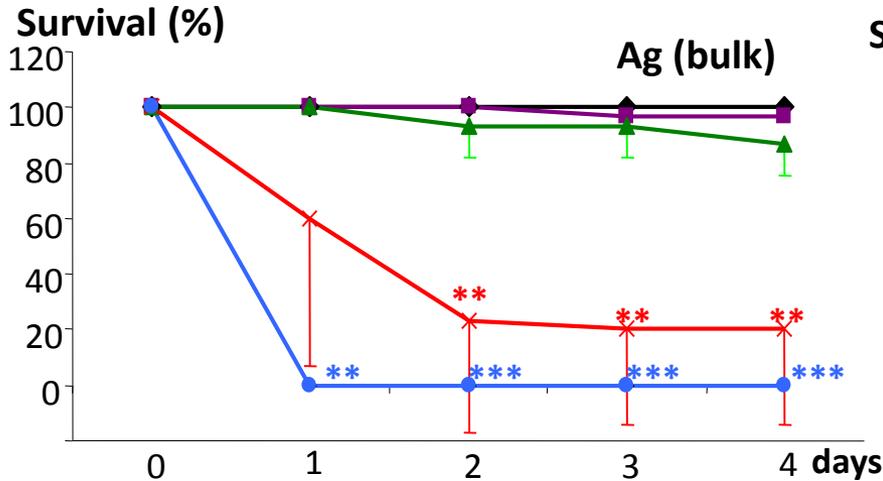
$LD_{50} (Ag_{bulk}): >1000 \text{ mg/L}$

→ Dose- and size-dependent toxicity

→ Ag less toxic in trout hepatocytes

\* Release of enzyme **lactate dehydrogenase** into the surrounding medium upon rupture of the plasma membrane - marker for **cell toxicity**

# Toxicity of Ag particles to *D. magna*



µg/ml Ag: —◆— control —■— 0.01 —▲— 0.1 —×— 1 —●— 10

## Bulk Ag:

- 100 % mortality at 10 mg/L
- 80 % mortality at 1 mg/L
- No significant mortality at 0.1 and 0.01 mg/L over 96 h
- $LC_{50}$  (48hrs)  $\approx$  0.7mg/L

## Nano-Ag:

- 100 % mortality at 10 & 1 mg/L
- 60 % mortality at 0.1 mg/L
- No significant mortality at 0.01 mg/L
- $LC_{50}$  (48hrs)  $\approx$  0.3mg/L

# Bioavailability of Ag in Carp (21 day exposures)



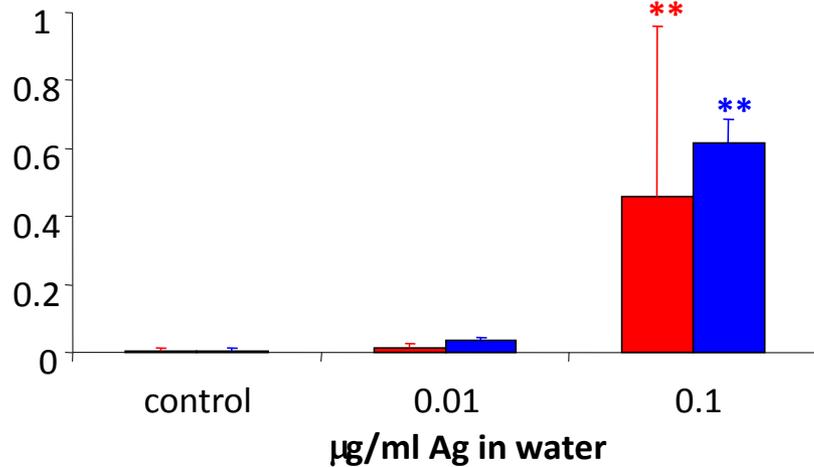
Liver

■ bulk Ag

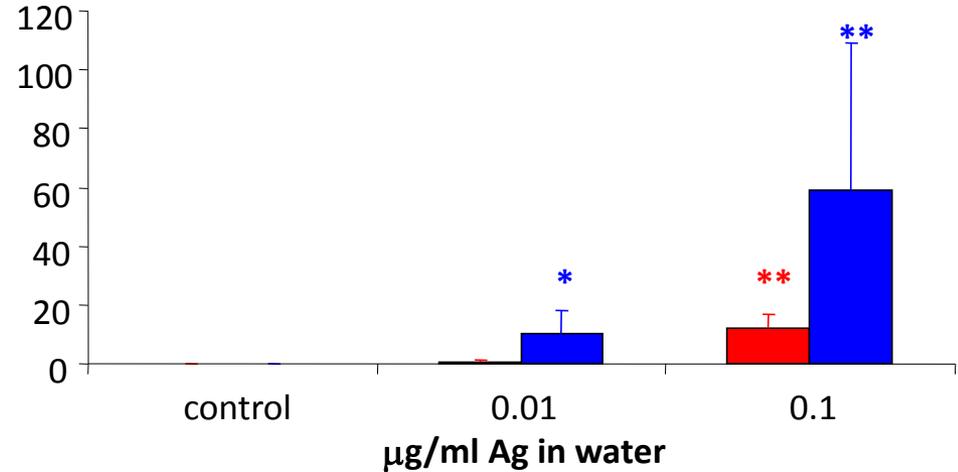
■ nano Ag

Intestine

ppm Ag in tissue



ppm Ag in tissue



Ag increased in carp liver for both particle sizes (0.1 mg/L)

Ag uptake at 0.01 mg/L (nano) and 0.1 mg/L (bulk and nano)

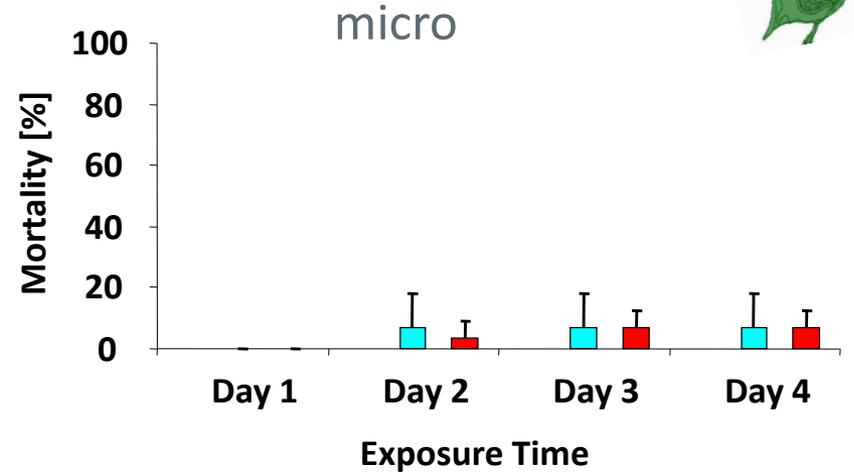
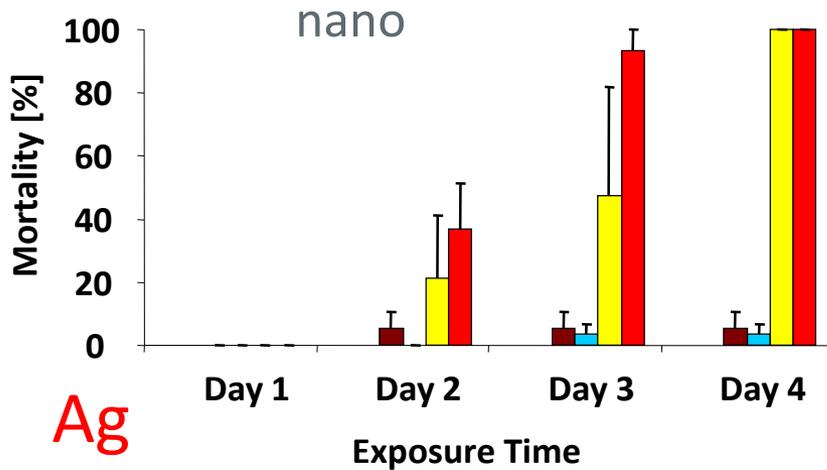
Highest concentration of all tissues

Results obtained for gall bladder and gill showed enhanced values but much variability

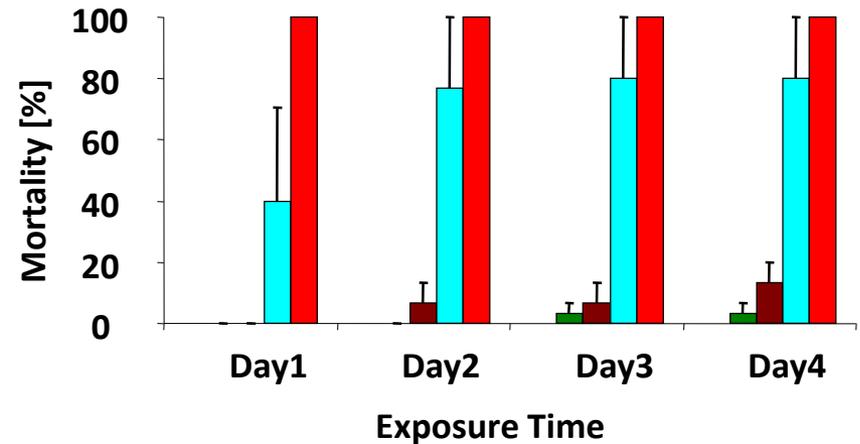
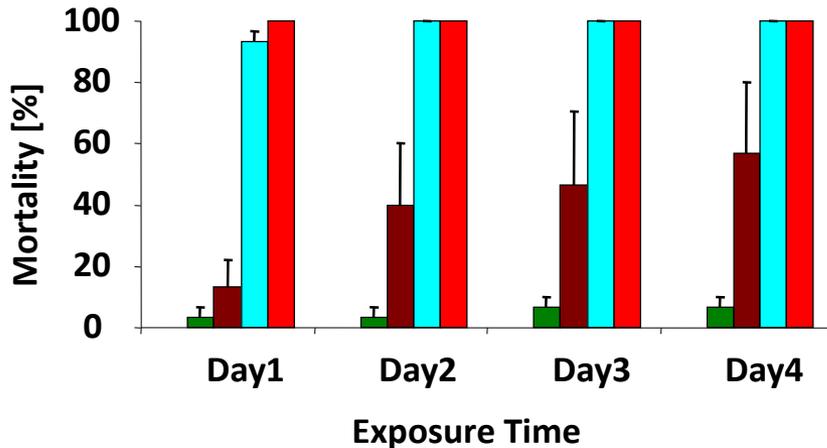
Mortality of mass dose in 96h acute tests with micro and nano particles of carbon black (14 and 260 nm) and silver (35 nm and ~1um)

**CB**

Control 0.01 mg/l 0.1 mg/l 1 mg/l 5 mg/l 10 mg/l



**Ag**



# Toxicity: $\text{CeO}_2$ , *Daphnia magna*



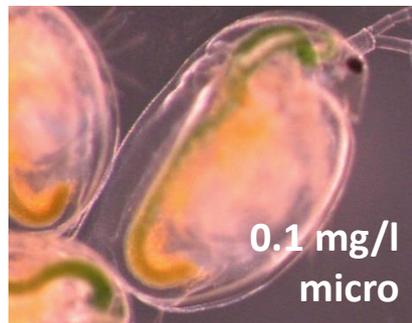
control



Gaiser et al 2012a  
Gaiser et al 2012b



0.01 mg/l  
micro



0.1 mg/l  
micro



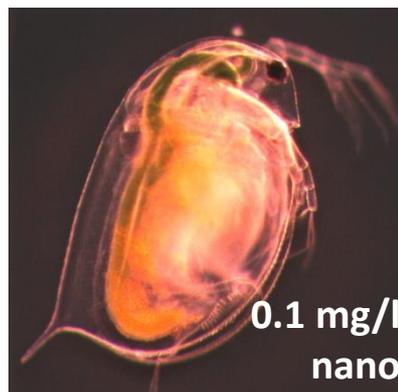
1 mg/l micro



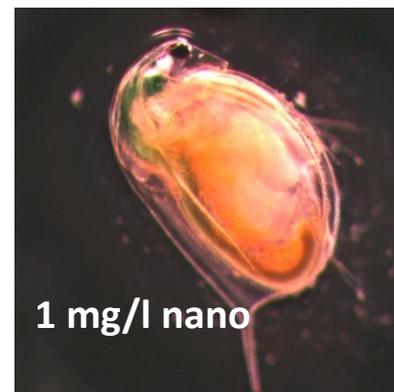
10 mg/l  
micro



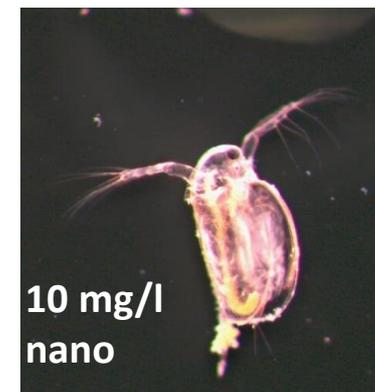
0.01 mg/l  
nano



0.1 mg/l  
nano



1 mg/l nano



10 mg/l  
nano

# Marine macroalga



**Particles:**

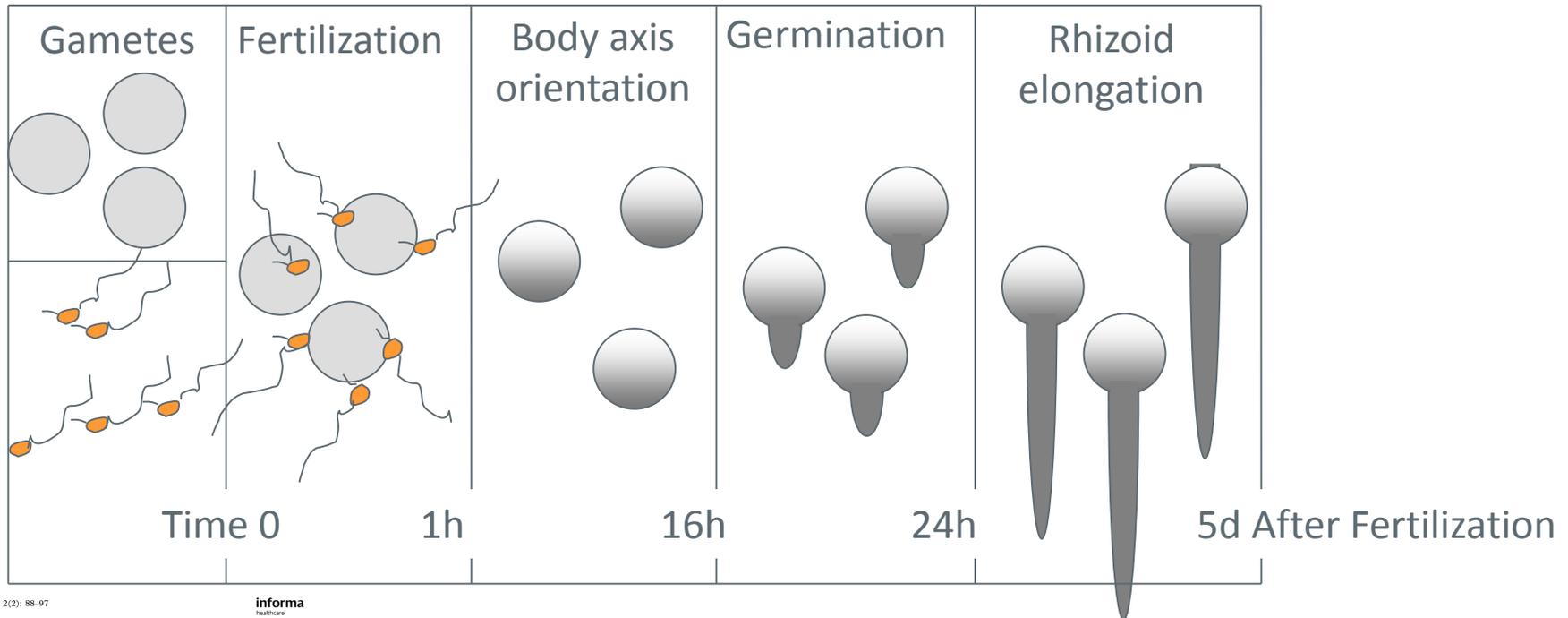
CB 14nm diameter (Degussa Printex 90)

0.1, 1, 10 and 100  $\mu\text{g}/\text{ml}$

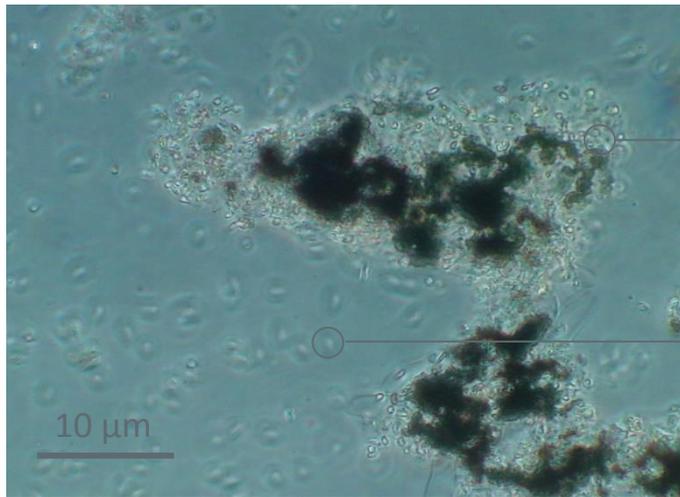
Dynamic Light Scattering characterisation

**Organism:**

Macroalga *Fucus serratus*



# Effects of CB NPs on marine macroalga



Spermatozoid *adsorbed* to CB aggregate

Free swimming spermatozoid

Physical restriction, shading?

# Macroalga treated with carbon NPs

Nanotoxicology  
Volume 2, Issue 2, 2008



Original  
**Interactions between carbon black nanoparticles and the brown algae *Fucus serratus*: Inhibition of fertilization and zygotic development**

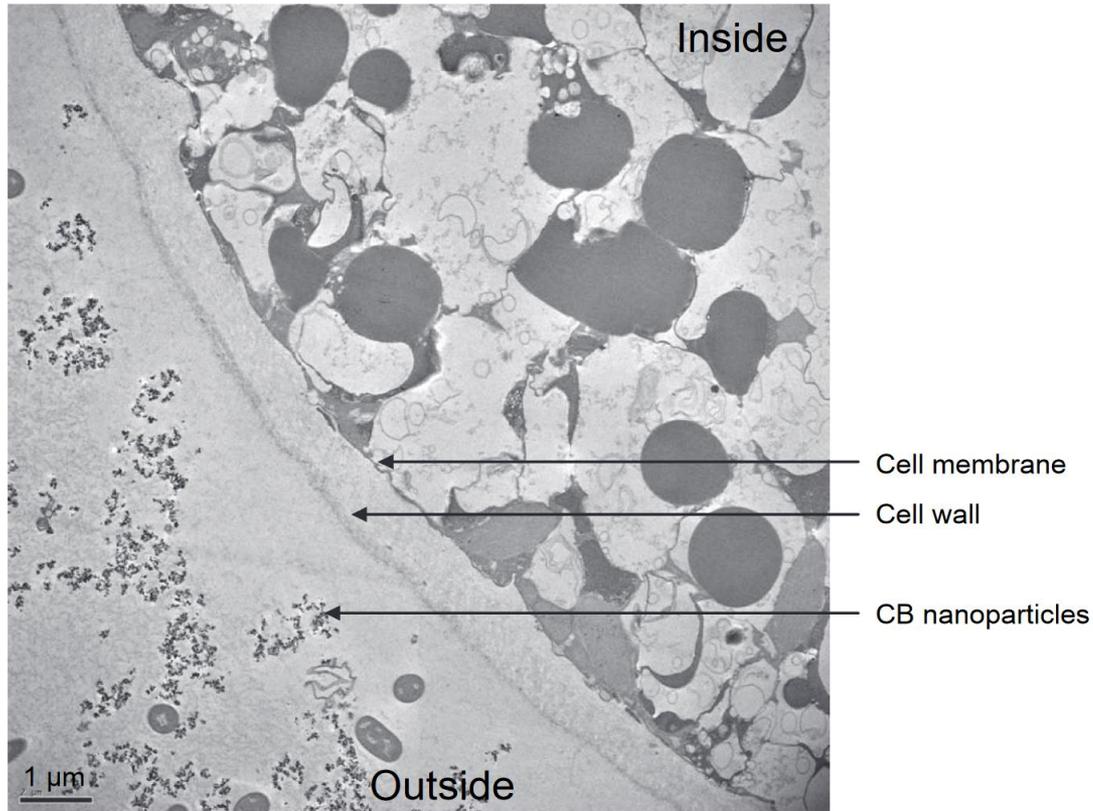


Figure 8. Example of cross section of *F. serratus* zygote after 20 h exposure to  $50 \mu\text{g ml}^{-1}$  CB. There is no sign of uptake of CB particles (lower left corner) in the zygote

# Effects of CB NPs on marine macroalga

## Results:

Carbon nanoparticles were found to influence *Fucus* embryos development, for example by:

- Reducing fertilisation success
- Changing orientation of the body axis

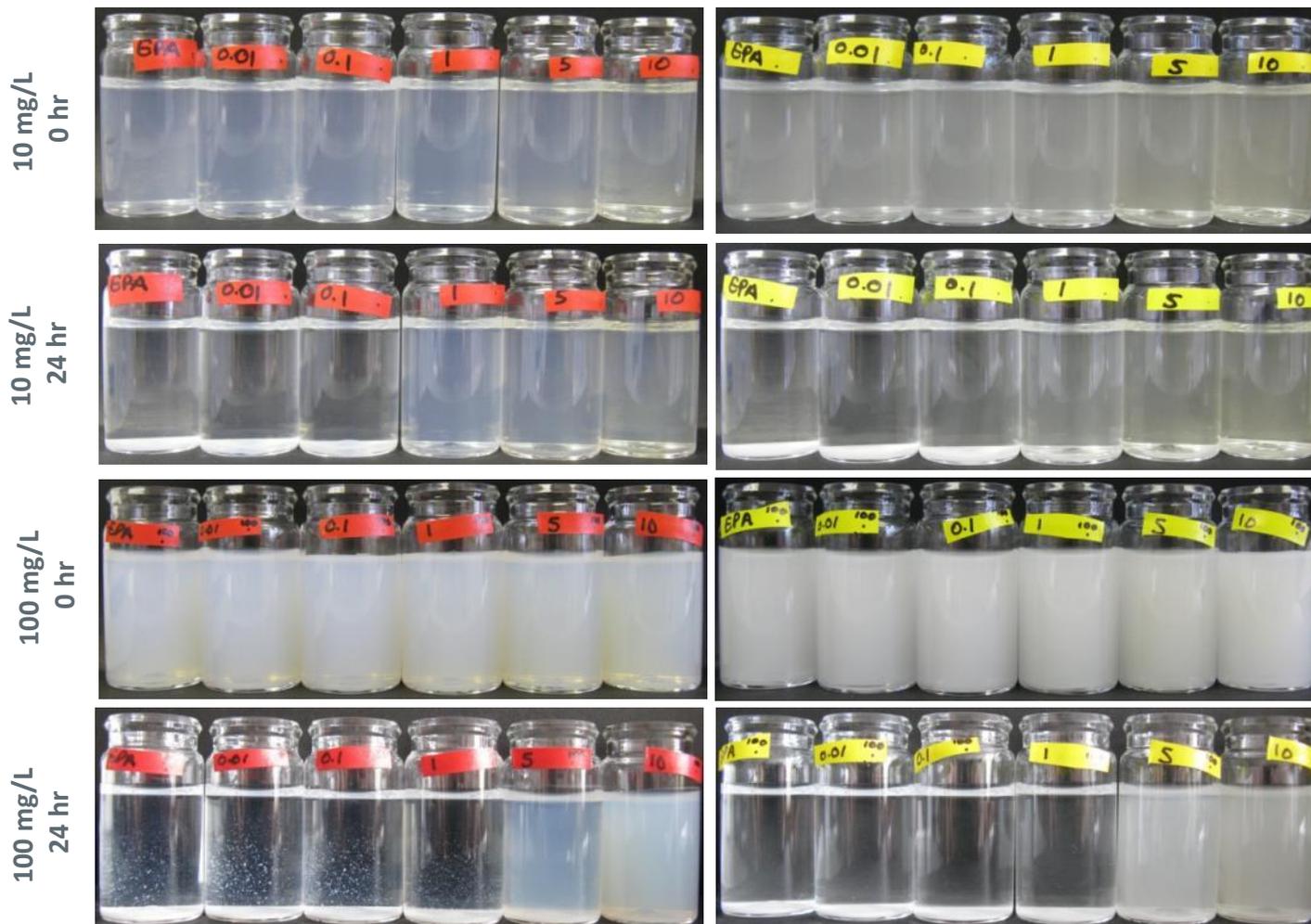


# Stability in media

- TiO<sub>2</sub> (25 nm) and (250 nm) 10 mg/L and 100 mg/L
- Suwannee River Humic Acid solutions of 0, 0.01, 0.1, 1, 5 and 10 mg/L

NP (25 nm)

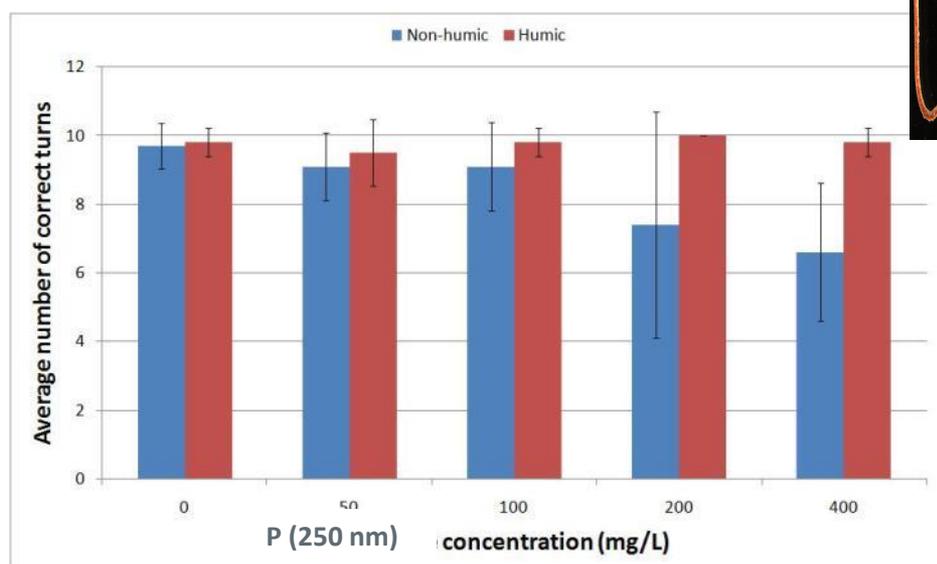
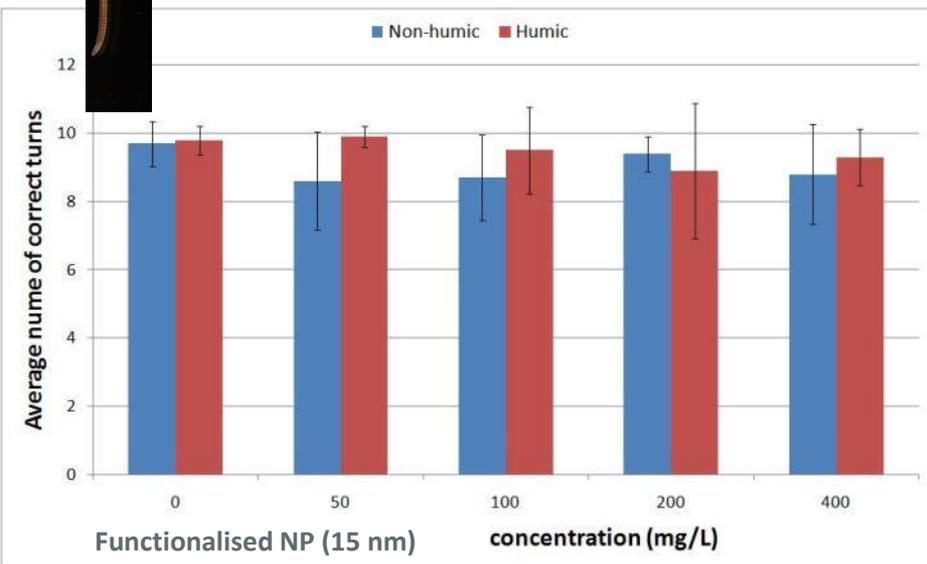
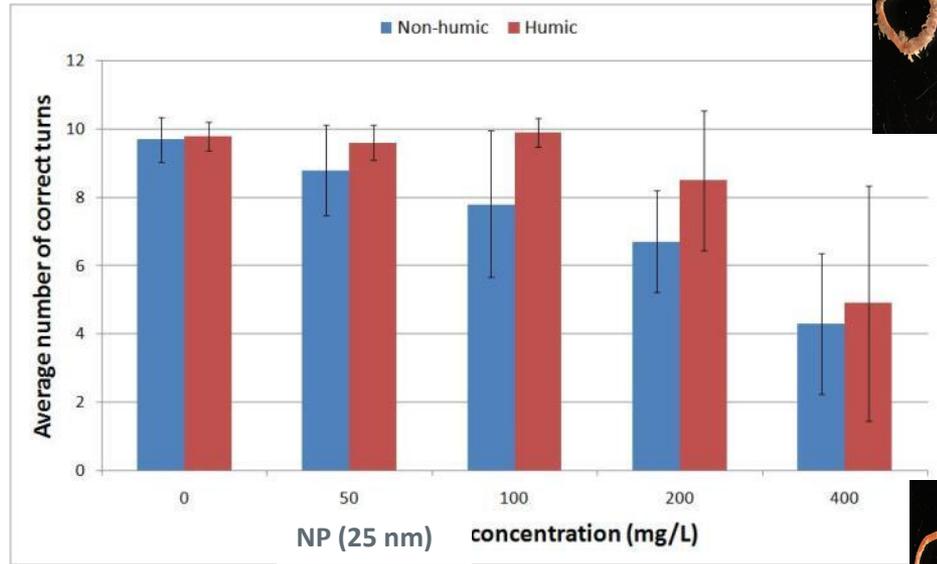
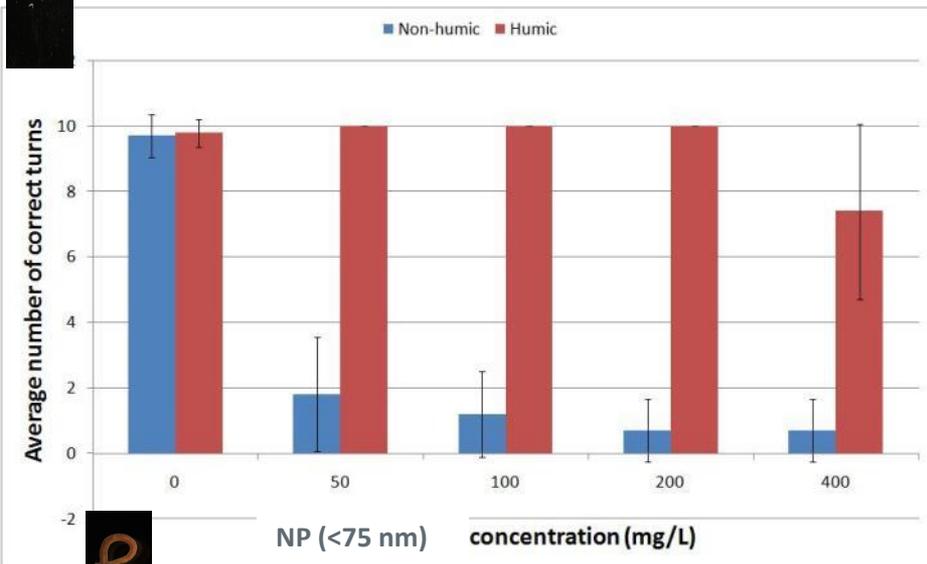
P (250 nm)



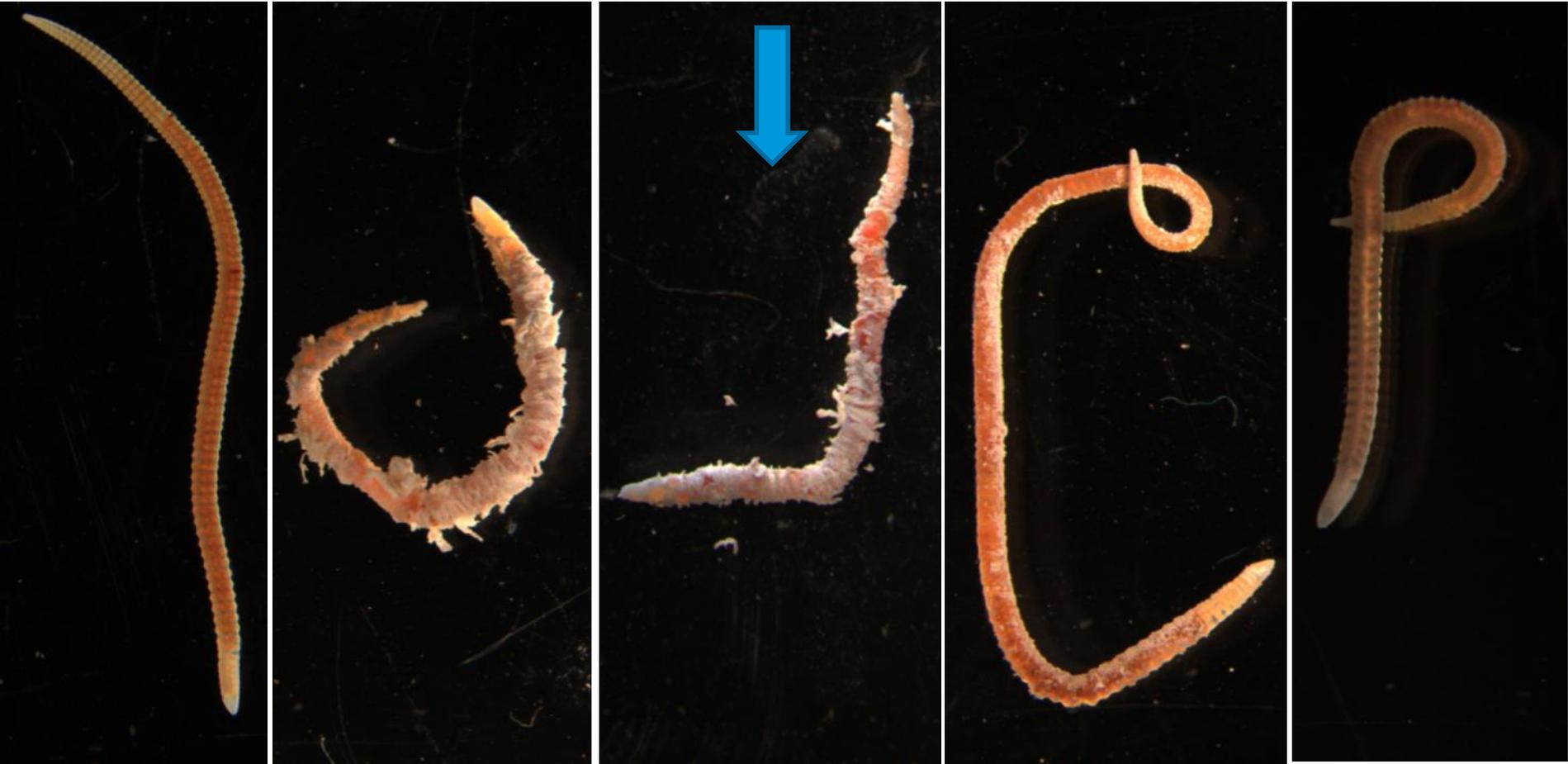
Increased humic acid concentration



# *Lumbriculus variegatus* sub-lethal results on exposures to TiO<sub>2</sub> particles (without/with 5 mg/L humic acid; water only 4 day exposure)



# *Lumbricus variegatus* (TiO<sub>2</sub>)



Control

NP (25 nm)

NP (<75 nm)

P (250 nm)

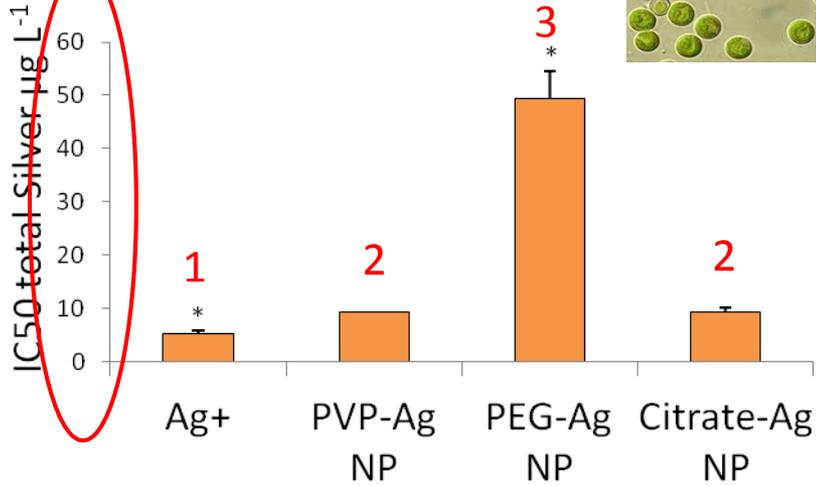
Modified NP (15 nm)\*

\*Consists of a TiO<sub>2</sub> core, an aluminium hydroxide and hydrated silica shell, and is functionalised with alginic acid.

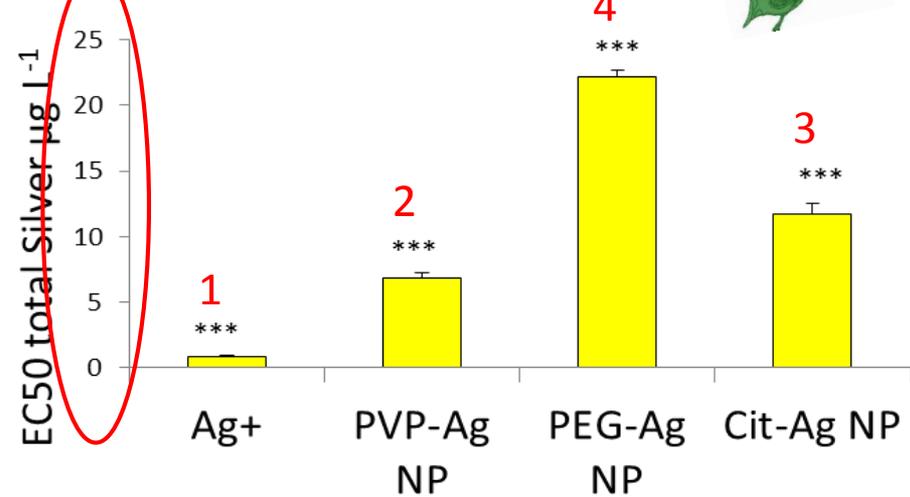


# Acute toxicities of different forms of Ag NPs (~ 10 nm) to 3 standard species

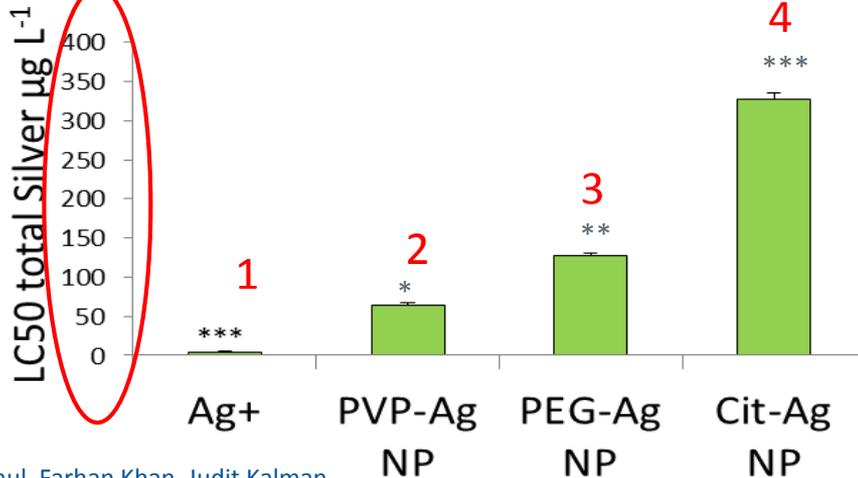
A. *Chlorella vulgaris*



B. *Daphnia magna*



C. *Lumbricus variegatus*

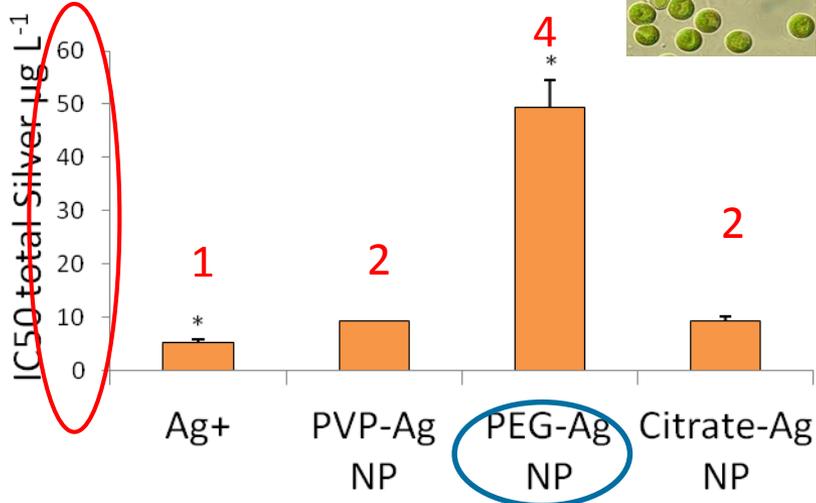


\*  $p < 0.05$   
 \*\*  $p < 0.01$   
 \*\*\*  $p < 0.001$

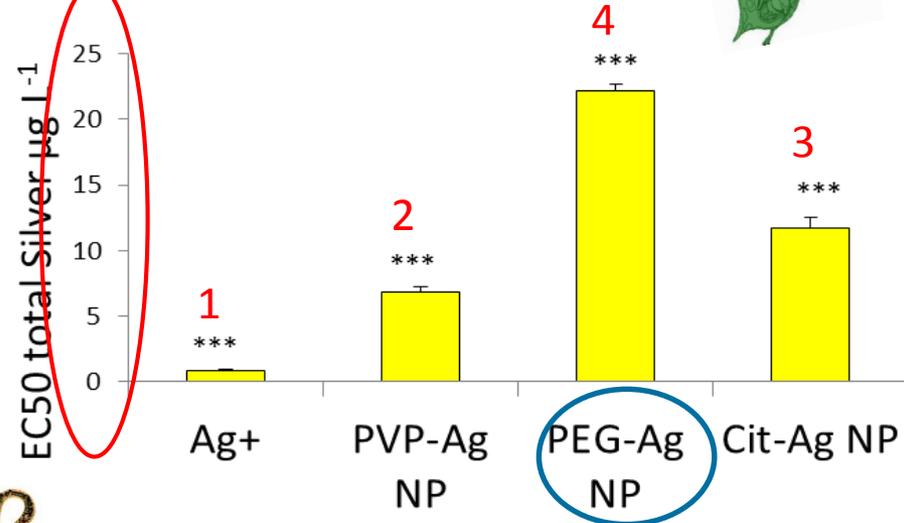
Ag<sup>+</sup> - exposure as silver nitrate  
 PVP – Polyvinylpyrrolidone  
 PEG – Polyethylene glycol  
 Cit - Citrate

# Acute toxicities of different forms of Ag NPs (~ 10 nm) to 3 standard species

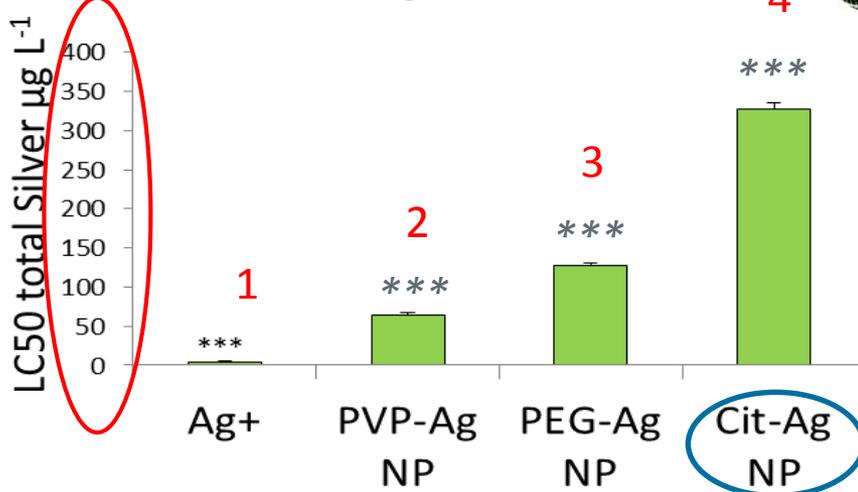
A. *Chlorella vulgaris*



B. *Daphnia magna*



C. *Lumbricus variegatus*



- Effects do not necessarily correlate with dissolution, aggregate size or zeta potential
- Ranking depends on test species

# Effects of Ag NPs on *C. vulgaris*



- **Aqueous Ag** was more toxic than Ag NPs to *C. vulgaris*
- Transmission electron microscopy (TEM) images indicated uptake and **internalization** of PVP-, and PEG-Ag NPs into the cells of *C. vulgaris* only at high exposure concentration
- **Possible increase in membrane permeability** led to the internalization of NPs
- In general **uptake rate constant correlated well with toxicity** in *C. vulgaris*
- **Dissolution data do not relate to toxicity**, suggesting that dissolved Ag from Ag NPs not the only form that interact with the algae

# Effects of Ag NPs on *D. magna* (food vs aqueous Ag exposures)



- **Ag was assimilated from ingested algae by *D. magna***
- *D. magna* assimilated Ag from ingested Ag NP-containing algae with a higher efficiency than algae pre-exposed to AgNO<sub>3</sub>
- Ag NPs were not eliminated completely from *D. magna*, suggesting **possible transport of Ag NPs** along the food chain
- In general, **food is the dominant pathway** of Ag uptake in the cases of **Ag NPs**.
- In the case of **aqueous Ag, water becomes the major source** to the overall accumulation



# The project

- **NANOMICEX** - Mitigation of risk and control of exposure in nanotechnology based inks and pigments
- Partners from academia and industry
- Heriot-Watt University:
  - Hazard assessment of pristine and functionalised particles
  - Human and environmental exposure (and models)
  - Identification of high hazard particles
  - Impact of surface modifications:
    - Decreasing particle toxicity
    - Keeping pigment properties



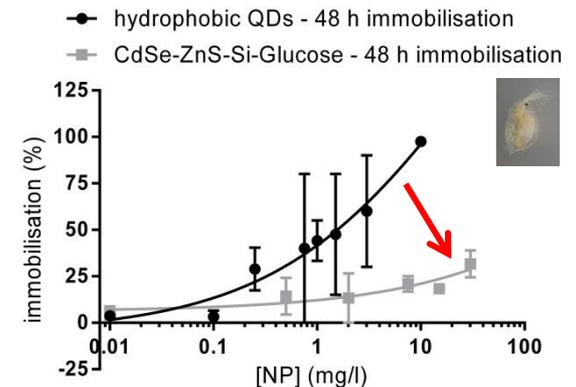
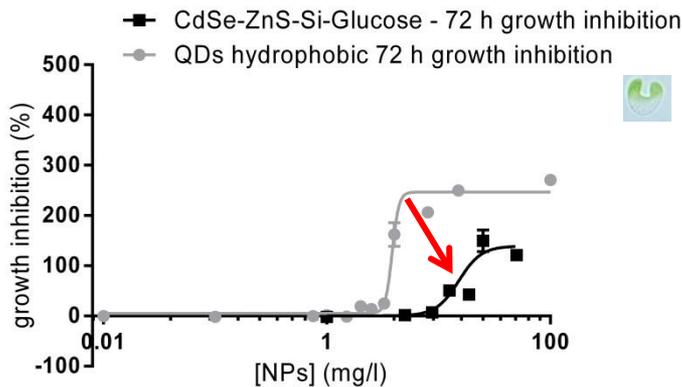
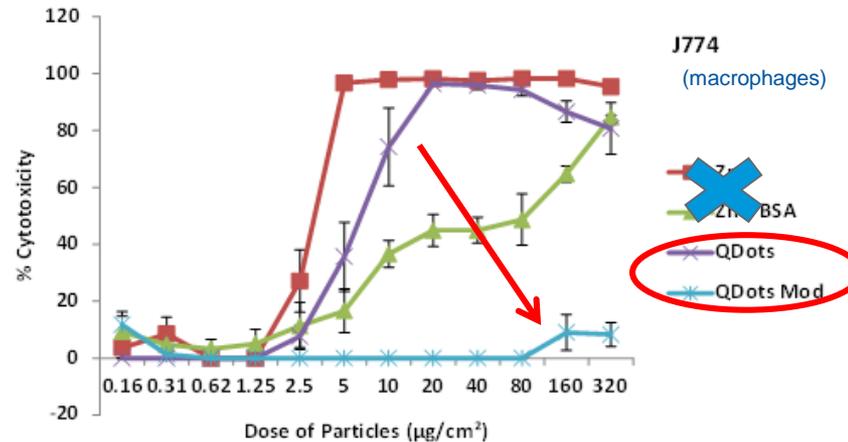
# Ranking of particle acute toxicity

	Al <sub>2</sub> O <sub>3</sub>	Ag (hydrophilic)	CdSe (QDs)	CoAl <sub>2</sub> O <sub>4</sub>	Fe <sub>2</sub> O <sub>3</sub>	TiO <sub>2</sub>	ZnO
<i>P. sub-capitata</i>	Green	Red	Yellow	Green	Green	Green	Yellow
<i>L. vari-gatus</i>	Green	Yellow	Green	Green	Green	Green	Green
<i>D. magna</i>	Green	Red	Yellow	Green	Green	Green	Yellow
J774 Macro- phages	Yellow	Yellow	Yellow	Yellow	Yellow	Yellow	Red
C3A Hepato- cytes	Green	Red	Red	Green	Green	Green	Red
A549 Alveolar epithelial type II cells	Green	Red	Yellow	Green	Green	Green	Red

# Modified QDs – toxicity

(with ZnS shell and Silica-Glucose ligands)

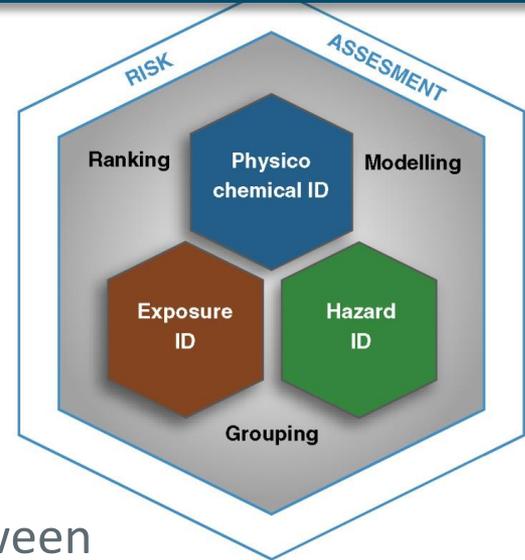
- Modified QDs are less toxic than original QDs



# Research prioritization to develop an Intelligent Testing Strategy

A process that allows the risks of nanomaterials (NMs) to be assessed:

- Accurately
  - Effectively
  - Efficiently
- 
- **Short term** - understanding of the connections between physicochemical, exposure and hazard IDs to enable **grouping/ ranking**.
  - **Longer term** - development of **modelling** approaches for RA with a continual reduction in testing.
  - In the **distant future** RA based on modelling and extrapolations and only if additional information is required with focused **physicochemical, exposure and hazard testing**.





**OECD Environmental Health and Safety Publications**

**Series on Testing and Assessment**

**No. XX**



**GUIDANCE DOCUMENT ON AQUATIC (AND SEDIMENT) TOXICOLOGICAL  
TESTING OF NANOMATERIALS**

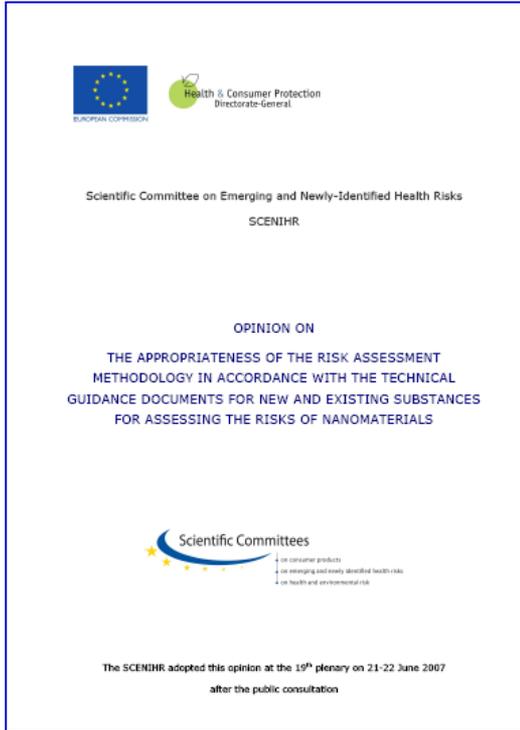


**Environment Directorate**

**ORGANISATION FOR ECONOMIC CO-OPERATION AND DEVELOPMENT**

**Paris**

**September 2016**






Scientific Committee on Emerging and Newly-Identified Health Risks  
 SCENIHR

**OPINION ON**  
**THE APPROPRIATENESS OF THE RISK ASSESSMENT**  
**METHODOLOGY IN ACCORDANCE WITH THE TECHNICAL**  
**GUIDANCE DOCUMENTS FOR NEW AND EXISTING SUBSTANCES**  
**FOR ASSESSING THE RISKS OF NANOMATERIALS**



- ↓ on consumer products
- ↓ on emerging and newly identified health risks
- ↓ on health and environmental risk

The SCENIHR adopted this opinion at the 19<sup>th</sup> plenary on 21-22 June 2007  
 after the public consultation





Scientific Committee on Emerging and Newly Identified Health Risks  
 SCENIHR

Opinion on  
 Nanosilver: safety, health and environmental effects and role  
 in antimicrobial resistance



- ↓ on consumer safety
- ↓ on emerging and newly identified health risks
- ↓ on health and environmental risks

SCENIHR approved this opinion at the 6<sup>th</sup> plenary of 10 -11 June 2014




 ORGANISATION FOR ECONOMIC  
 COOPERATION AND DEVELOPMENT


**OECD Work on  
 the Safety of  
 Manufactured  
 Nanomaterials**

Environment, Health and Safety Division  
 Environment Directorate  
 OECD

**NANOSREG**



# Summary

- Nanoecotoxicology has learnt a LOT from nanotoxicology
- Collaboration and interaction are essential
- The receiving environment affects fate, bioavailability and toxicity
- NMs can be taken up by the gut and sometimes translocated to other parts of the body (although not always)
- Respiratory surfaces (such as gills) are sometimes the most sensitive, but that is not always the case
- Some NMs can be passed on through the food chain
- Not all nanomaterials are equally toxic, but often toxicity in one model/species can be replicated in other models/species
- Physical impacts can be a problem but 'toxic' effects are also observed
- Population, food chain studies and interactions with other chemicals provide a means of assessing long-term and indirect effects
- Assay preparation and conditions, as well as reporting of any observed effects need to be considered carefully – certain methods are not appropriate to nanomaterial hazard studies!

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# Well done and thanks, Vicki!

## Toxicology Award 2015 Winner

**Professor Vicki Stone**

Heriot-Watt University

Awarded for pioneering transdisciplinary approaches  
to assessing the safety of nanomaterials

