

Supplementary Data

“Simultaneous quantification of five bacterial and plant toxins from complex matrices using a multiplexed fluorescent magnetic suspension assay”

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Table S1. Comparison of multiplex detection systems for proteotoxins*

Detection system	Antigens	Detection limit [ng/L]	Matrix	Time [min]	Ref
Magnetic xMAP-technology	BoNT/A	21	food	210	this manuscript
	BoNT/B	73			
	Ricin	2			
	SEB	3			
	Abrin	546			
NRL – biosensor	BoNT/A toxoid	20,000	food	10 – 45	1
	BoNT/B toxoid	200,000			
	Ricin	8,000			
	SEB	100			
	Cholera toxin several bacteria, viruses and small toxins	1,600			
Immunoassay on optical silicon chip	BoNT/A	2,500	food	35	2
	BoNT/B	5,000			
	BoNT/E	2,500			
	BoNT/F	5,000			
Semi-homogeneous fluidic force discrimination assay	Ricin A chain	5,000	clinical	10	3
	SEB	0.001			
Conventional xMAP-technology	Ricin	29,100	environmental	~75	4
	SEB	200			
	several viruses and bacteria				
RAPTOR	Ricin	50,000	environmental	3 – 10	5
	SEB	10,000			
	several bacteria				
ArrayTube – system	Ricin	100	buffer	60	6
	SEB	200			
	several viruses and bacteria				
Immunomagnetic, fluorogenic detection	BoNT/A	2,000	buffer	30	7
	Ricin A chain	1,000			
	SEB	100			
	Cholera toxin	2,000			
	diverse bacteria and viruses				
Microfluidic electrophoretic chip-based immunoassay	Ricin	1,320,000	buffer	< 20	8
	SEB	8,400			
	Shiga toxin 1	35,000			

Table S1. (continued)

Detection system	Antigens	Detection limit [ng/L]	Matrix	Time [min]	Ref
Hydrogel-based protein-microarray	Ricin	100	buffer	~1000	9
	SEB	1,000			
	Diphtheria toxin	1,000			
	LF (anthrax toxin)	4,000			
	Viscumin	2,000			
	Tetanus toxin	10,000			
CombiMatrix VLSI array	Ricin	300	buffer	12	10
	bacteria and proteins				
Bidiffractive grating (BDG) biosensor	BoNT/A or /B	100,000	buffer	15	11
	Ricin	5,000			
	SEB	< 1,000			
QTL-biosensor 2200R	BoNT	< 10,000	buffer	10	12
	Ricin	< 1,000			
	SEB	< 1,000			
Multiplexed sandwich ELISA with quantum dots	Ricin	30,000	buffer	~1000	13
	SEB	30,000			
	Cholera toxin	30,000			
	Shiga like toxin 1	30,000			
Conventional xMAP-technology	BoNT/A toxoid	1,100	buffer	120	14
	Ricin	3,300			
	SEB	14			
	Cholera toxin	4			
AMP-biosensor	BoNT/A	1,000	buffer	75	15
	BoNT/B	10,000			
Bead based immunoassay with eTag reporter	BoNT/A toxoid	125,000	buffer	~66	16
	Protein ovalbumin	125,000			

* Table S1 gives an overview over different multiplex detection systems suitable for laboratory-based and on-site detection of high molecular weight proteotoxins. The table includes information on the matrices analysed, the detection limits obtained for individual proteotoxins in multiplexed settings and the experimental time per analysis.

Table S2. Immunisation schema for rabbit RB77 (anti-BoNT/B IgG)*

immunisation	day	antigen	quantity	adjuvants
1	0	BoNT/B-Dynabeads	5 μ l	-
2	37	(5 μ g BoNT/B/ 6.5×10^8 Dynabeads)	10 μ l	CFA
3	92		40 μ l	IFA
4	125		2 ng	IFA
5	144	BoNT/B	10 ng	IFA
6	207		10 ng	IFA
7	232	BoNT/B-Dynabeads	30 μ l	IFA
8	257	(15 μ g BoNT/B/ 6.5×10^8 Dynabeads)	30 μ l	IFA
9	293		30 μ l	IFA
10	335		4 ng	IFA
11	424		4 ng	IFA
12	544		10 ng	IFA
13	557	BoNT/B	20 ng	IFA
14	649		20 ng	IFA
15	705		20 ng	IFA
16	762		23 ng	IFA
17	814		23 ng	IFA

* Detailed immunisation schema for rabbit RB77 immunised with BoNT/B coupled to Dynabeads and free native BoNT/B. Tab. S2 and Fig. S1 together show that an increase in absolute amount of BoNT/B coupled to Dynabeads leads to a specific titer development. From immunisation 9 onwards, the rabbit was regularly bled in order to collect large amounts of serum.

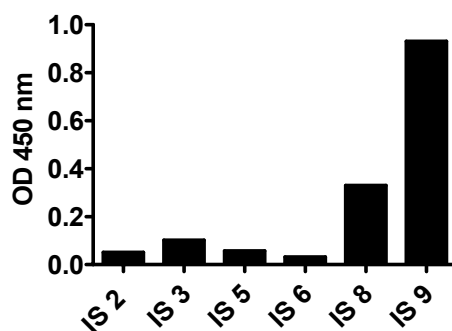


Fig. S1. Titer development of rabbit RB77 (anti-BoNT/B IgG). The rabbit was immunised according to Table S2 and 10–16 days after each immunisation (IS) the rabbit serum was tested in indirect ELISA (dilution 1:1000) against BoNT/B. Shown are the sera obtained after immunisation 2 through 9. From IS 9 on the titer remained on a constantly high level, no further increase in titer was observed after switching from BoNT/B-Beads to soluble BoNT/B.

Table S3. Immunisation schema for mouse M51 (anti-ricin IgG)*				
immunisation	day	antigen	quantity	adjuvants
1	0		5 μ l	CFA
2	28	Ricin-Dynabeads (42 μ g Ricin/ 6.5×10^8 Dynabeads)	5 μ l	IFA
3	43		5 μ l	IFA
4	57		10 μ l	IFA
5	72		1 μ g	IFA
6	98		2.5 μ g	IFA
7	147		2.5 μ g	-
8	148	Ricin	3.75 μ g	-
9	196		4 μ g	-
10	197		8 μ g	-
11	198		16 μ g	-

* Detailed immunisation schema for mouse M51 immunised with ricin coupled to Dynabeads followed by soluble ricin. Fusion of spleen cells from mouse M51 delivered mAb R18, R109 and R70 (compare Fig. 1 and Tab. 2, main manuscript). Two other mice were immunised similarly, but used earlier for fusion (after immunisation 7) and delivered the other hybridoma clones described in Tab. 2 of the main manuscript.

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