

SUPPORTING ON LINE SUPPLEMENTARY MATERIAL FOR

**Use of Isotope Mass Probes for Metabolic Analysis of the Jasmonate Biosynthetic
Pathway**

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Supporting figures

Figure S1. Procedure for pretreatment of rice seedlings and isotope mass probe labeling. MeOH, methanol; Et₂O, ethyl ether; BPB, ω -bromoacetylpyridinium bromide; d⁵-BPB, d⁵- ω -bromoacetylpyridinium bromide.

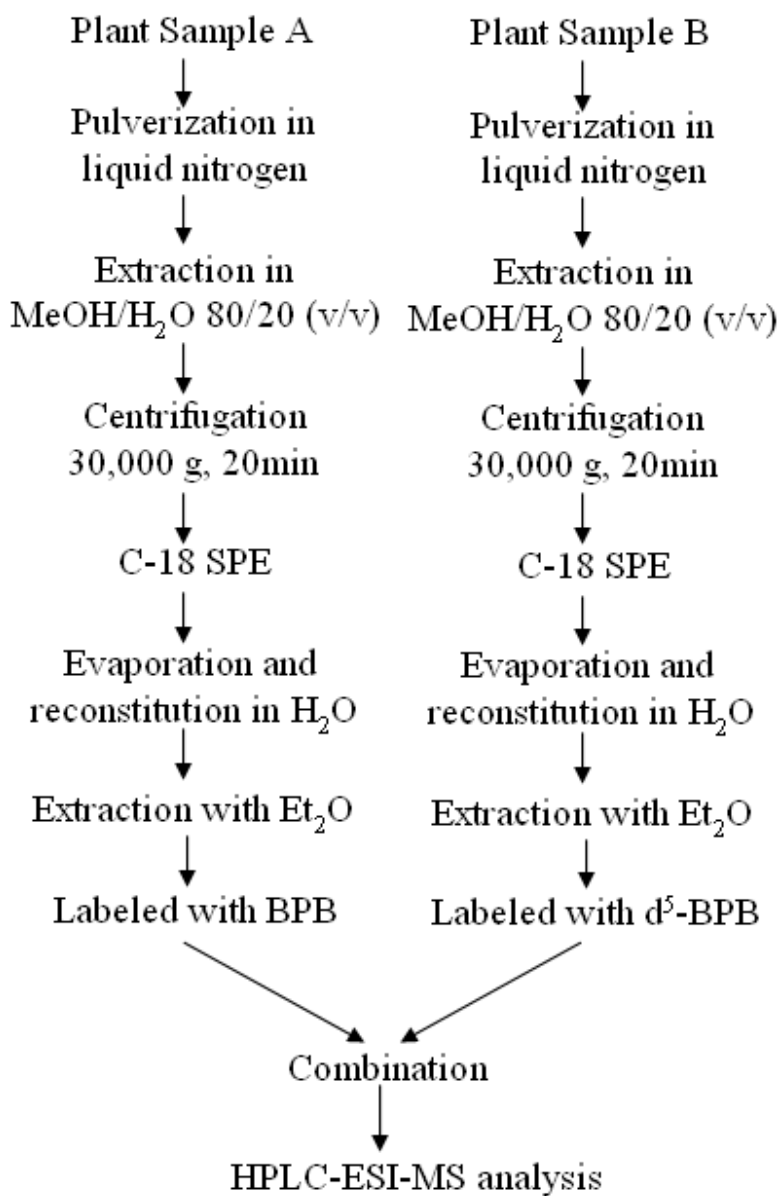


Figure S2. Optimization of labeling procedures from standard solutions. (A) Optimization of TEA concentration. The concentration of BPB was fixed at 3 nmol/mL (10 molar excess). The molar ratio of TEA over total amount of LA (0.1 nmol/mL), OPDA (0.1 nmol/mL) and JA (0.1 nmol/mL) was varied from 2 to 15. Reactions were carried out at room temperature for 2 hours. (B) Optimization of BPB molar ratios over a total amount of LA (0.1 nmol/mL), OPDA (0.1 nmol/mL) and JA (0.1 nmol/mL) for labeling. BPB was mixed with LA, OPDA and JA at molar ratios of 2 to 15 in the presence of TEA (10 molar excess) at room temperature for 2 hours. (C) Optimization of the reaction temperature. The reactions were performed with BPB at a molar ratio of 10:1 in the presence of TEA (10 molar excess) for 120 minutes. (D) Optimization of labeling time. The time was varied from 10 to 120 minutes. BPB was mixed with LA, OPDA and JA at a molar ratio of 10:1 in the presence of TEA (10 molar excess). The reactions were performed at constant room temperature at time periods of 10 min, 20 min, 30 min, 60 min and 120 min.

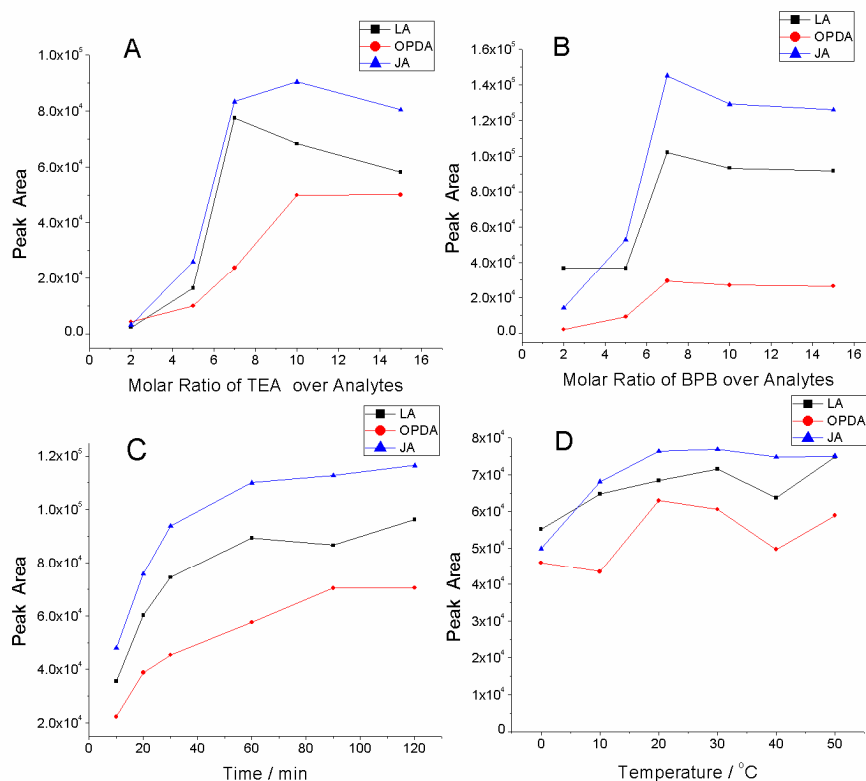


Figure S3. Examination of the stability of labeled derivatives. The change of the derivatives' intensity in solution was monitored over a period of 24 hours.

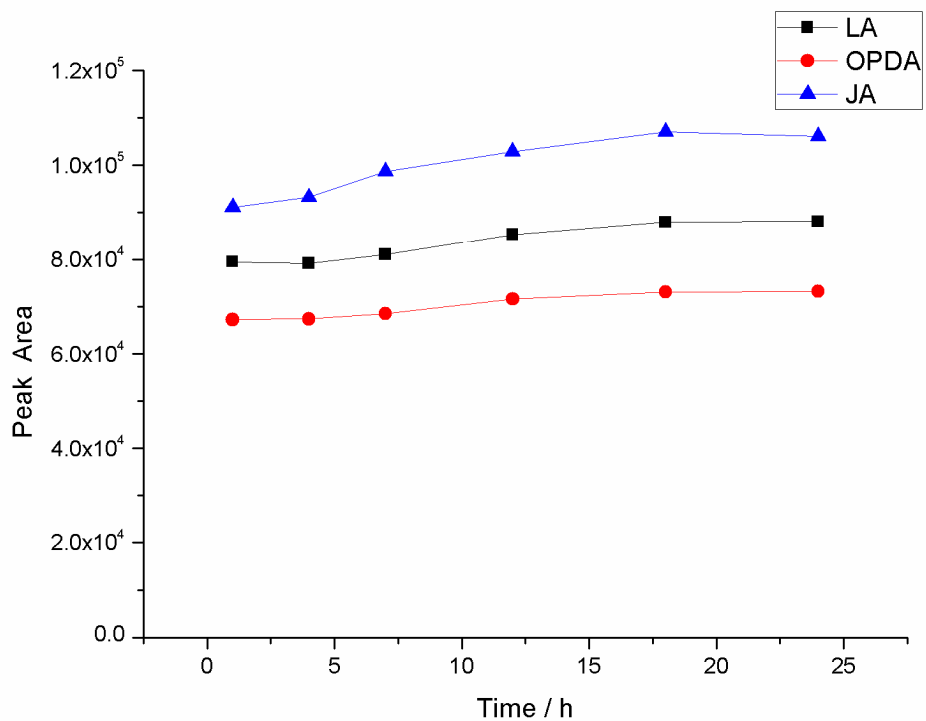


Figure S4. Optimization of labeling procedures of rice samples. (A) Optimization of TEA concentration for rice samples. The concentrations of TEA was varied from 0.5-2.5 $\mu\text{mol/mL}$. The concentration of BPB was set at 2.0 $\mu\text{mol/mL}$ and the reactions were carried out at room temperature for 2 hours. (B) Optimization of BPB concentration in rice samples. The concentrations of BPB was varied from 0.25-2.0 $\mu\text{mol/mL}$. The concentration of BPB was set at 1.5 $\mu\text{mol/mL}$ and performed at room temperature for 2 hours.

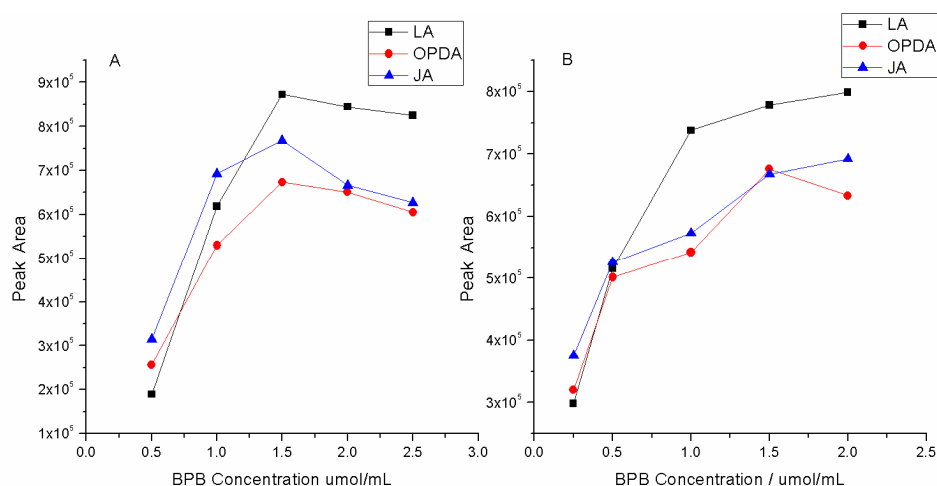


Figure S5. Comparison of ion signals between unlabeled (A) and labeled (B) analytes. The concentrations of LA, OPDA, OPC-8, OPC-6 and JA were set at 200 ng/mL each.

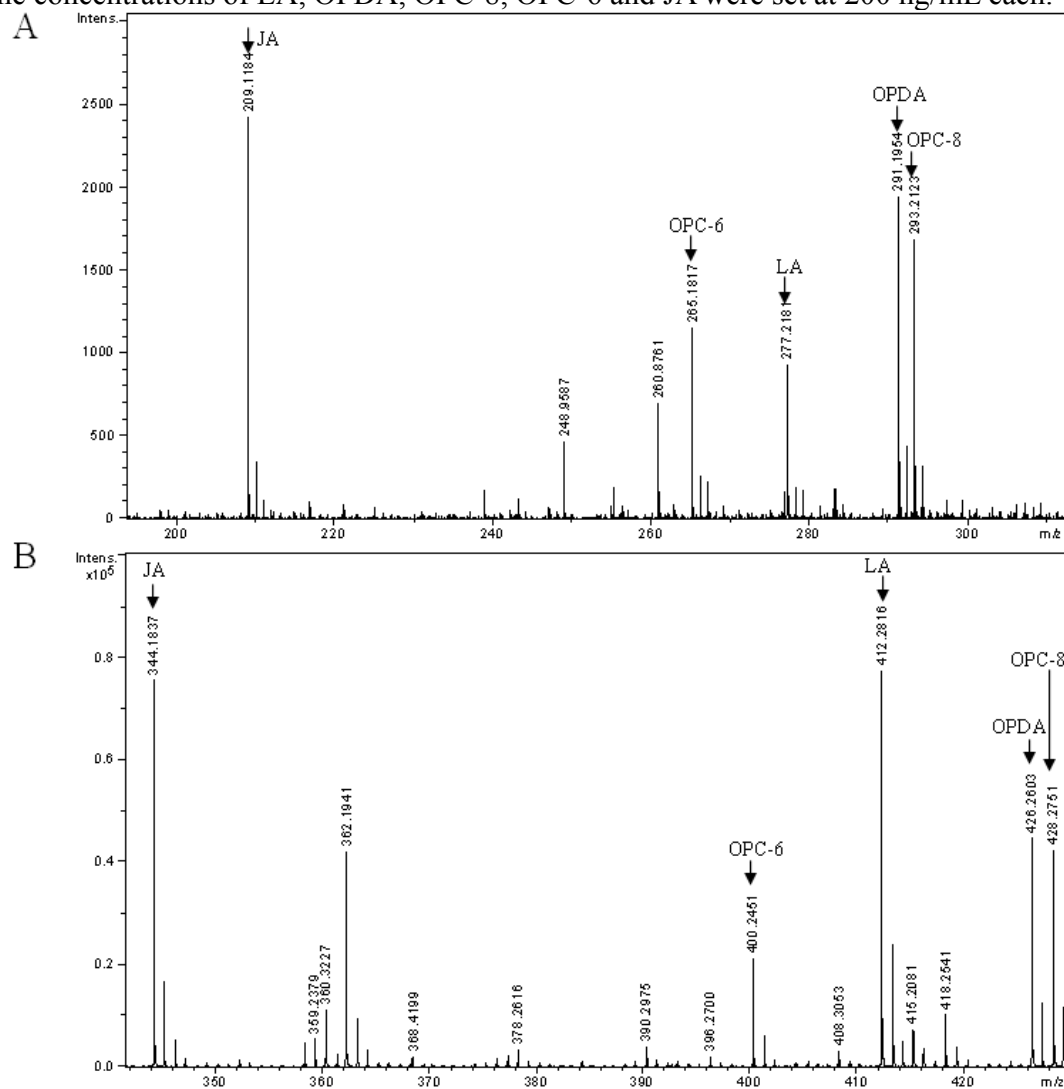


Figure S6. Linear regression plots of BPB/d⁵-BPB labeled (A) LA, (B) OPDA, (C) OPC-8, (D) OPC-6 and (E) JA. The d⁵-BPB labeled analytes were fixed at a concentration of 50 ng/mL, and the concentrations of BPB labeled analytes were varied from 5 ng/mL to 500 ng/mL. The relative standard deviation (RSD) of measured peak area ratios varied from 1.7-7.7%. Calibration parameters for the results were based on three replicate measurements.

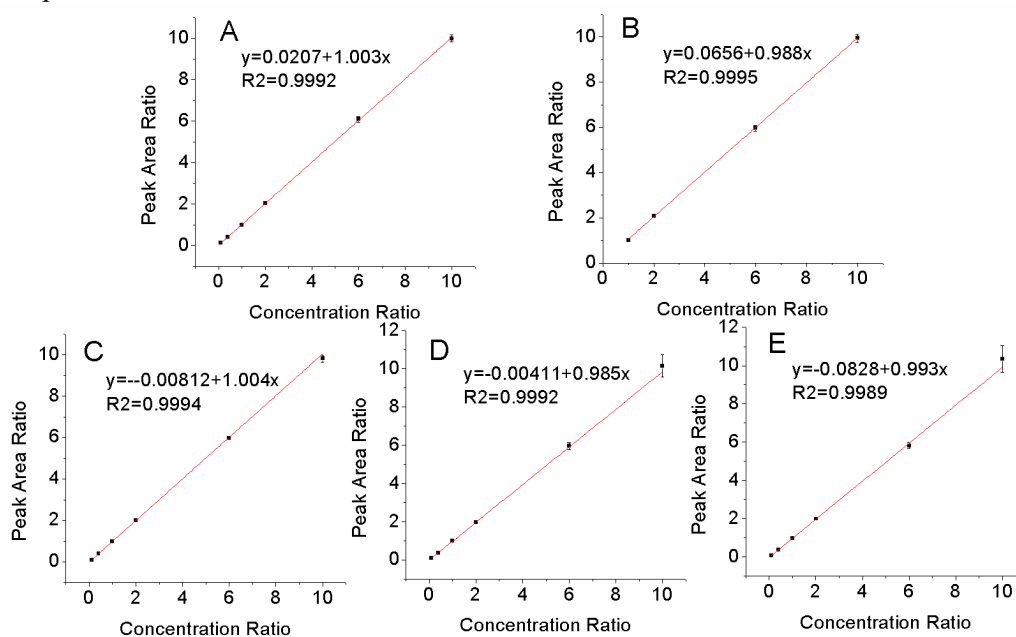


Figure S7. HPLC chromatograms of jasmonate biosynthetic pathway components from normally growing rice seedlings and salt treated rice seedlings. Normal growing rice seedlings were labeled with BPB (light) and salt treated rice seedlings were treated with d^5 -BPB (heavy). Chromatographic conditions were: column, Shiseido RP/SCX (150×2.1 mm, 5 μ m, Japan); sample injection volume, 10 μ L; mobile phase, 80% and 20% aqueous solution (containing 30 mmol/L ammonium acetate). LA, linolenic acid; HPOT, (13S)-hydroperoxyoctadecatrienoic acid; OPDA, cis-(+)-12-oxophytodienoic acid; OPC-8, 3-oxo-2-(2'-pentenyl)-cyclopentane-1-octanoic acid; OPC-6, 3-oxo-2(2'-pentenyl)-cyclopentane-1-hexanoic acid; JA, jasmonic acid; JA-Val, jasmonic acid-valine conjugate.

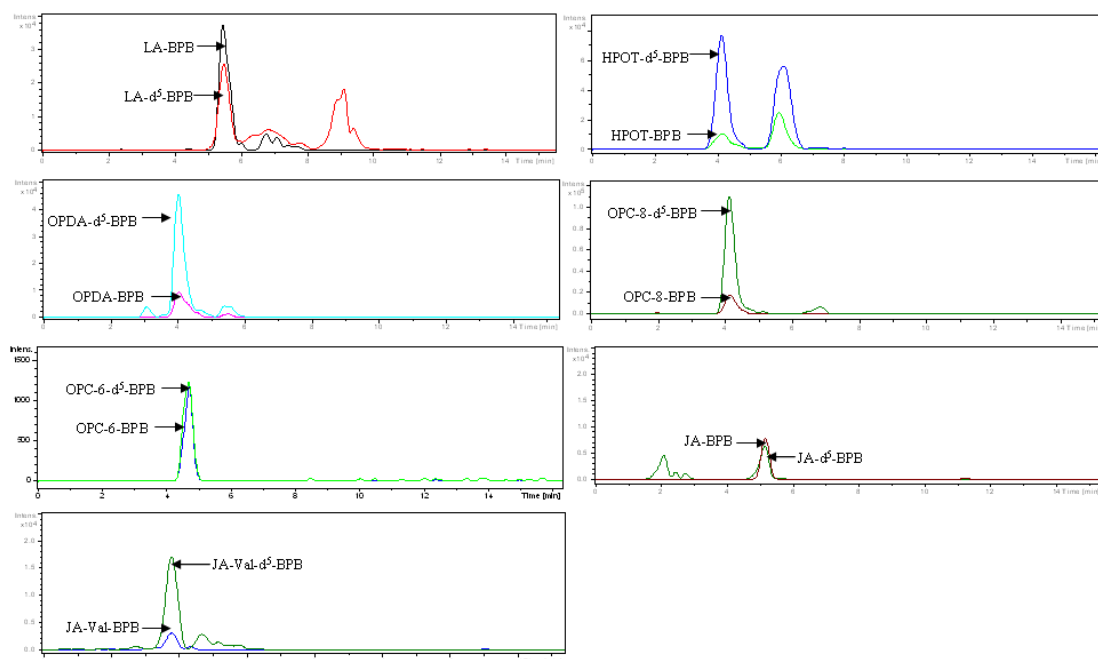
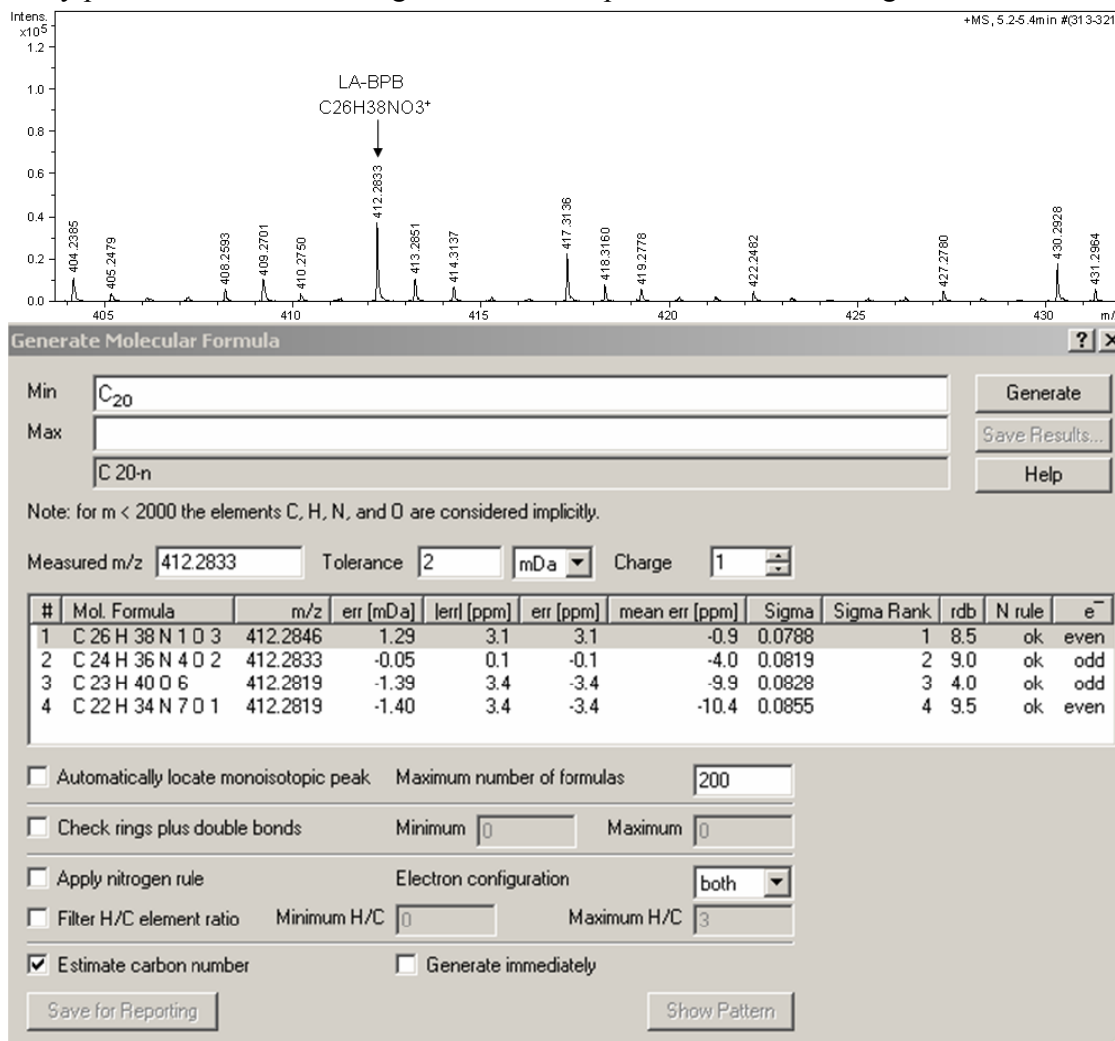
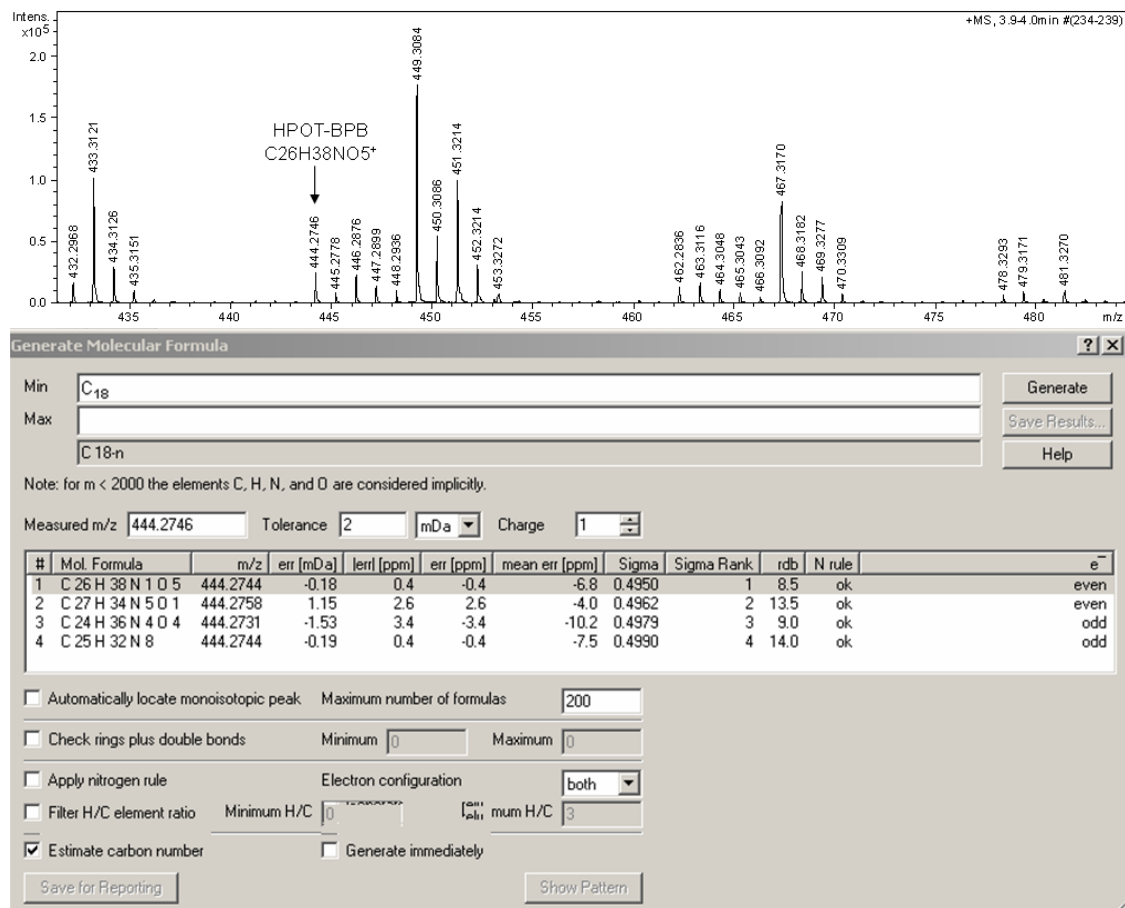
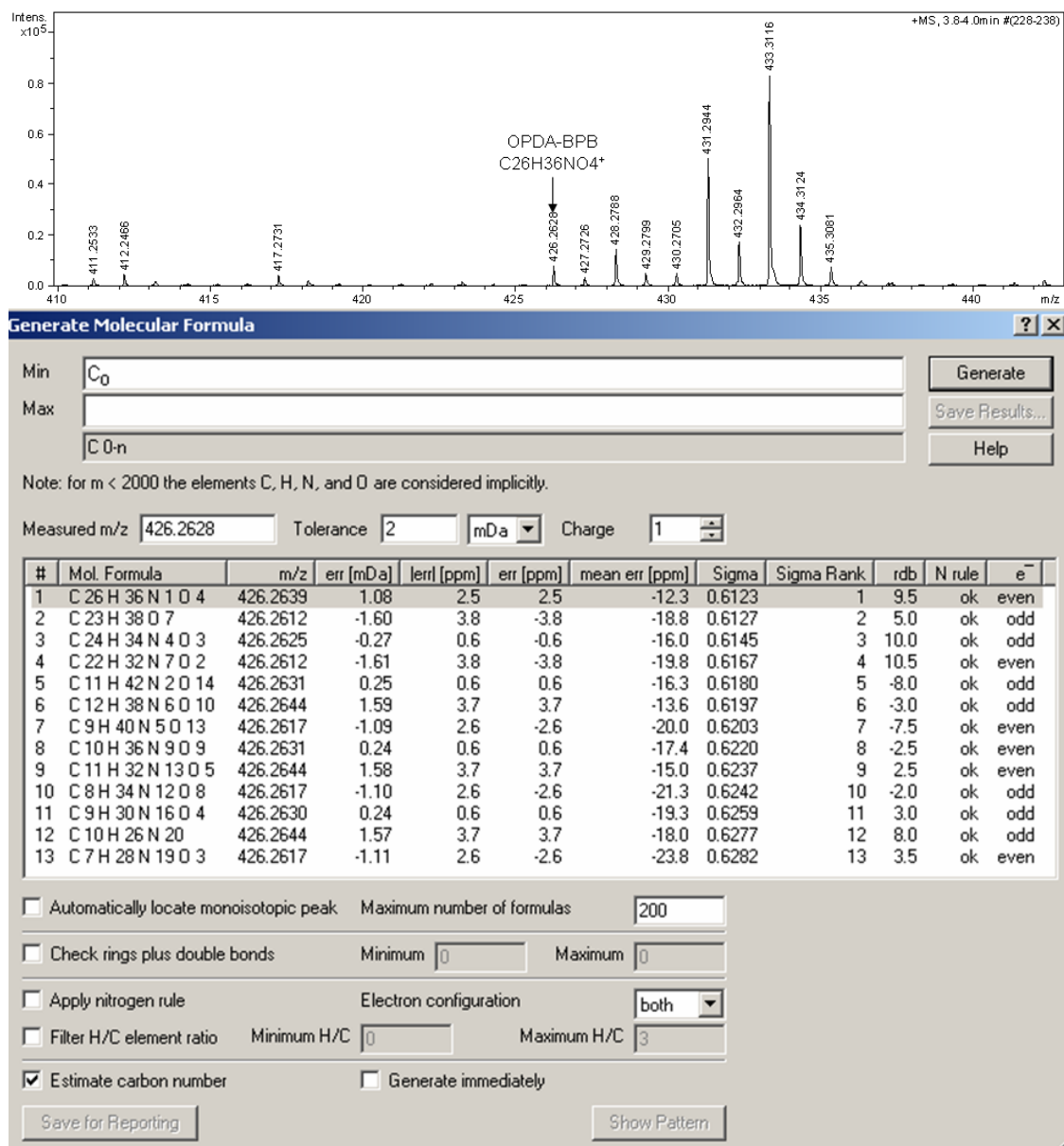
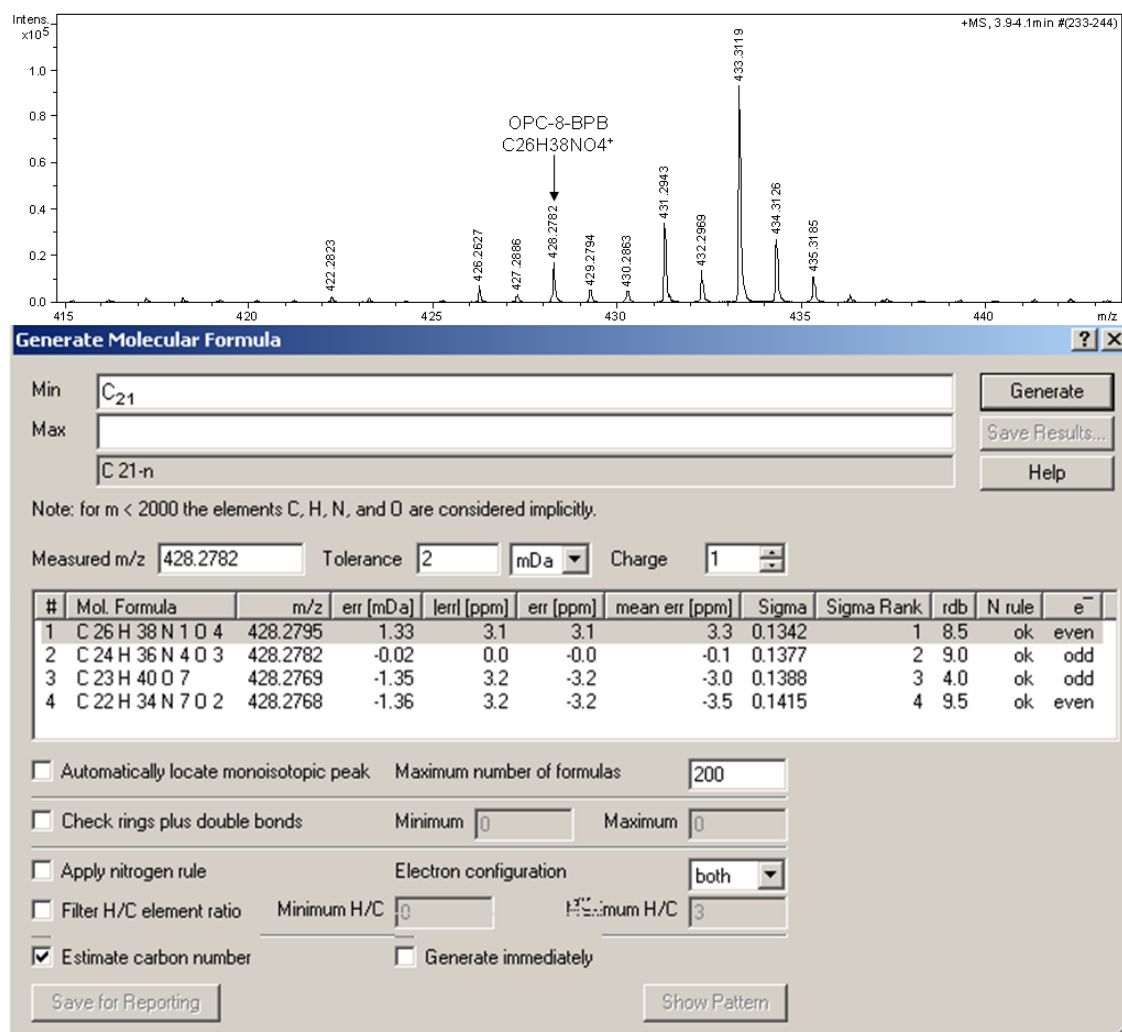


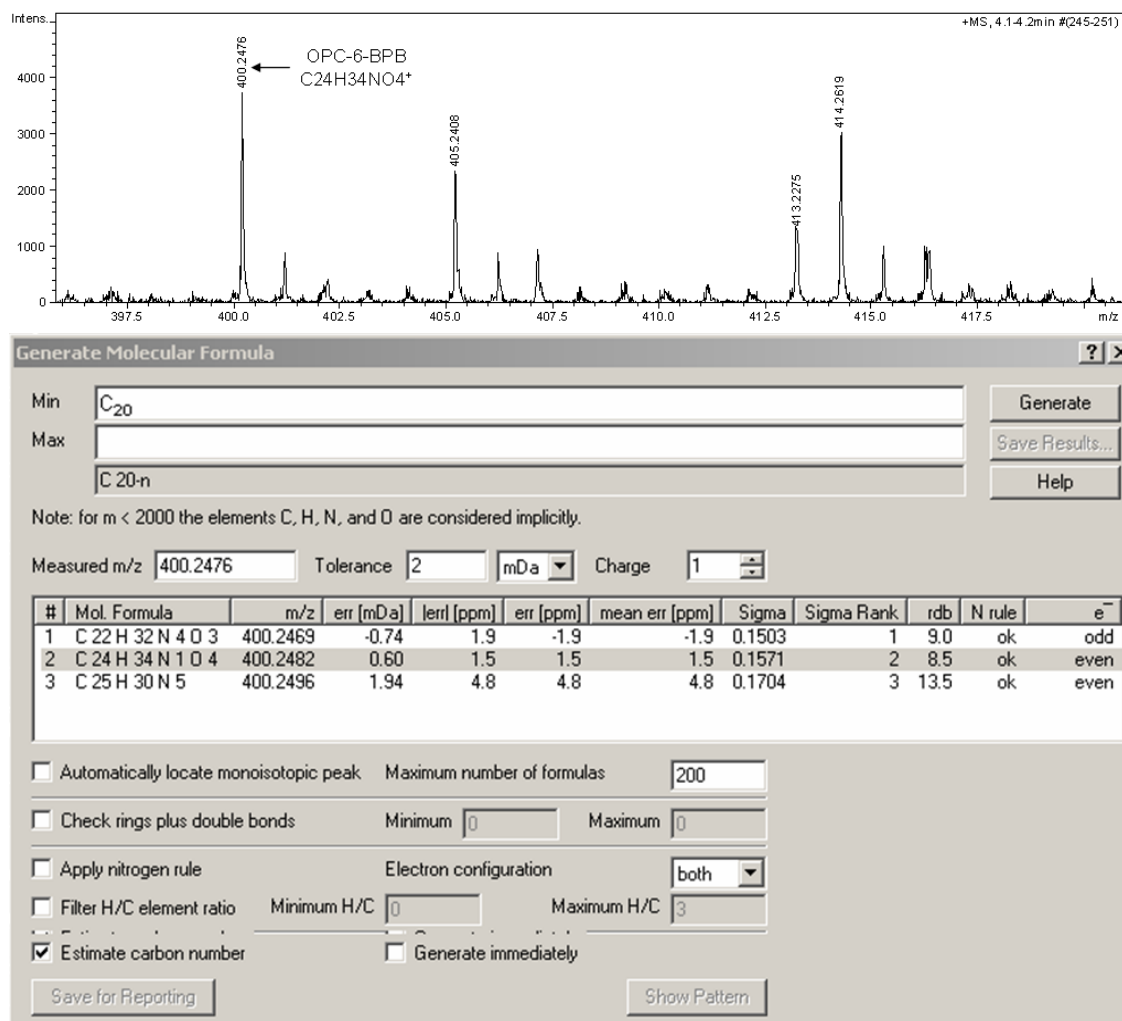
Figure S8. MS spectrums of peaks of the extracted traces and analytical results using Generate Molecular Formula (GMF) software. The theoretical values for [LA-BPB]⁺, [HPOT-BPB]⁺, [OPDA-BPB]⁺, [OPC-8-BPB]⁺, [OPC-6-BPB]⁺, [JA-BPB]⁺ and [JA-Val-BPB]⁺ ions are m/z 412.2846, 444.2744, 426.2638, 428.2795, 400.2482, 344.1856 and 443.2540, respectively. The correct formulas were selected from among the many potential candidates using GMF and isotope abundance matching.

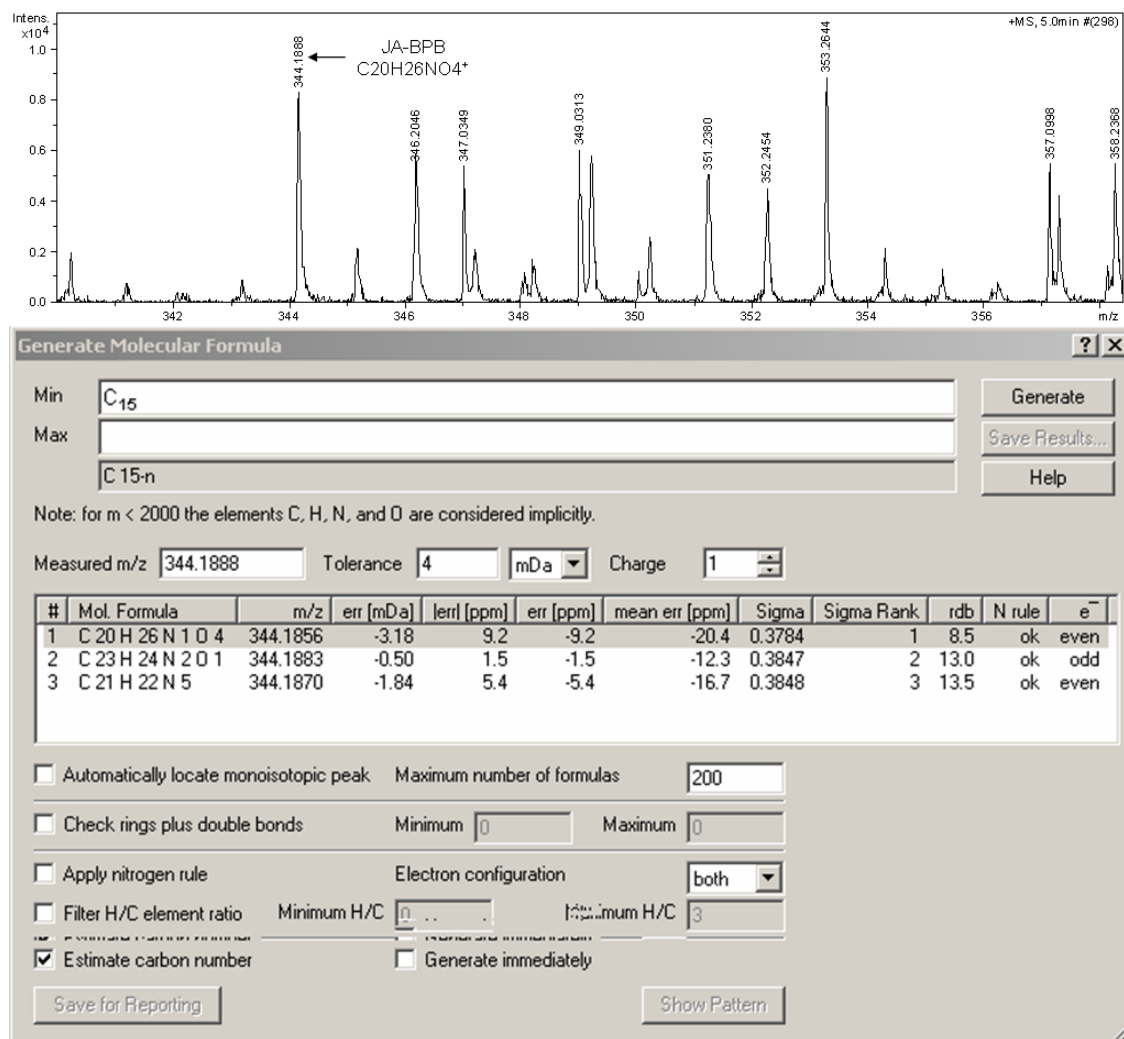












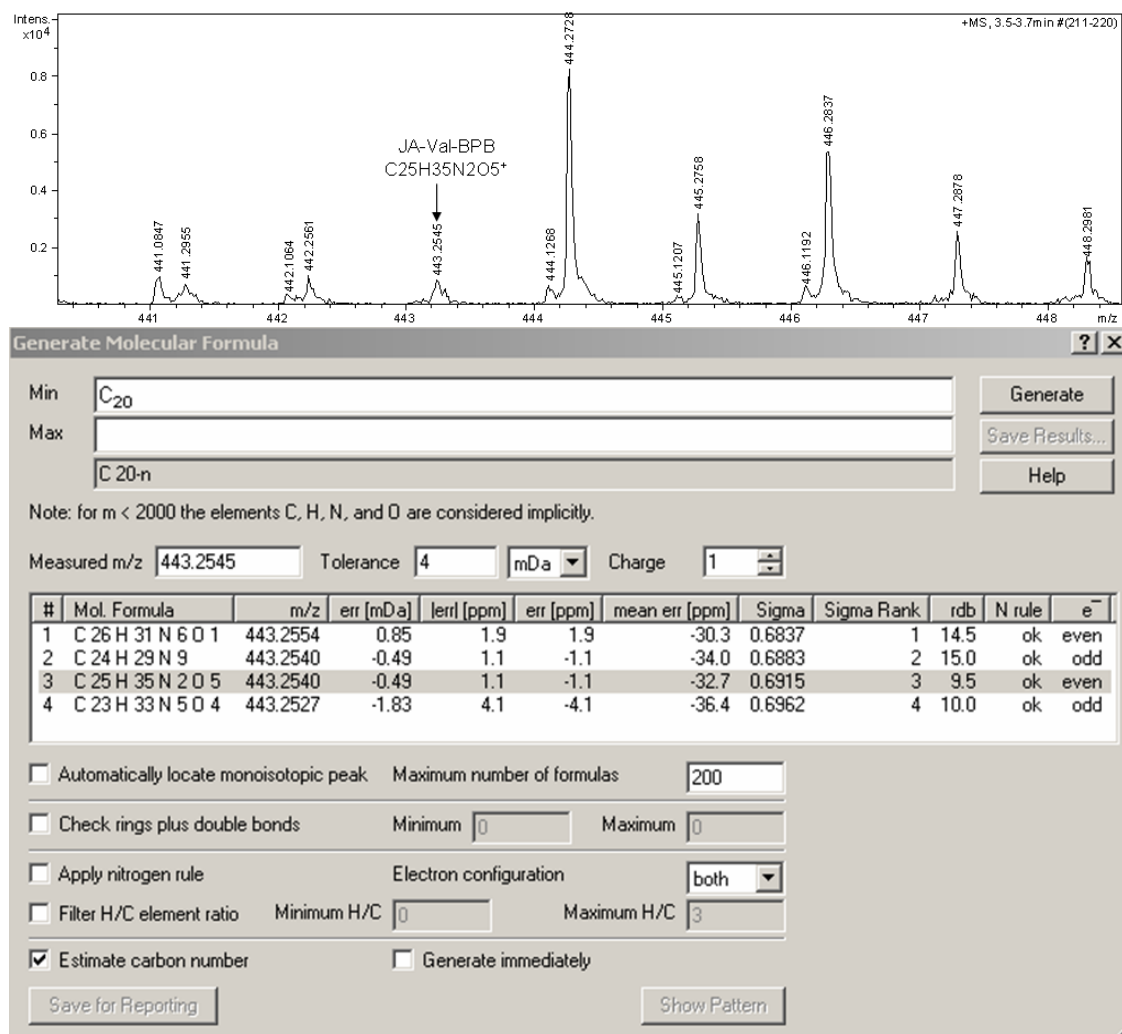


Figure S9. ^1H -NMR spectrum of 1,3-Dibromo-2,2-dimethoxypropane in CDCl_3 .

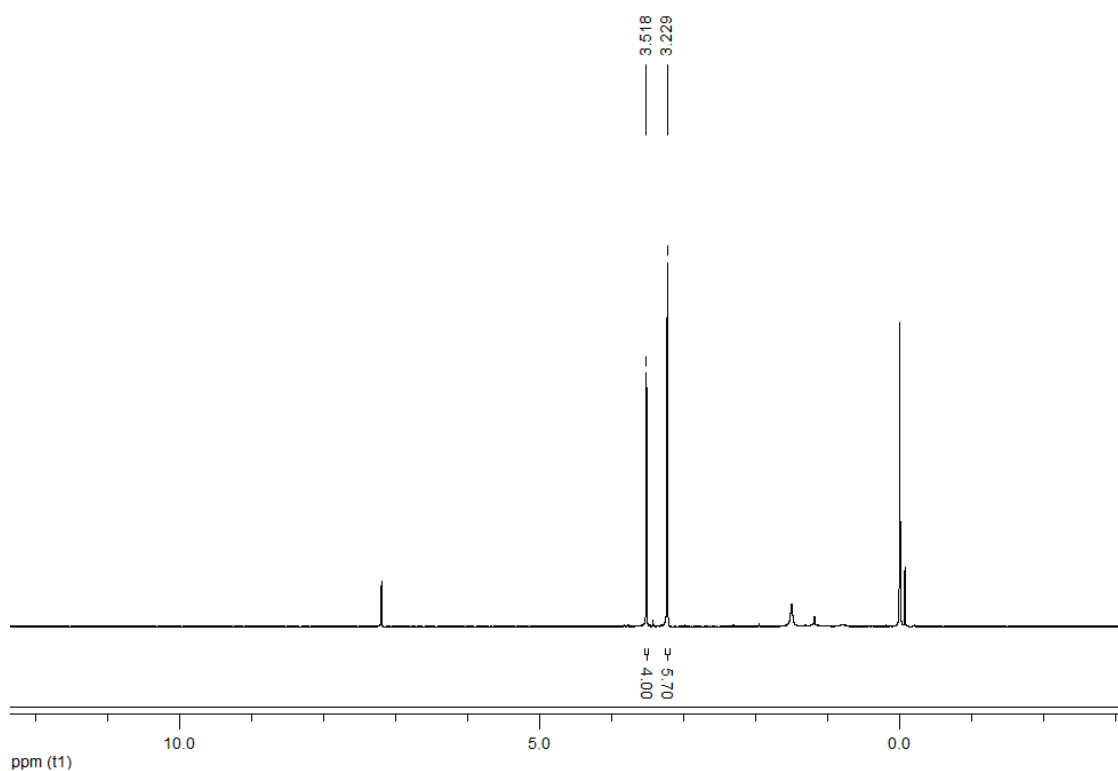


Figure S10. GC-MS chromatogram and mass spectrum of 1,3-dibromo-2,2-dimethoxypropane.

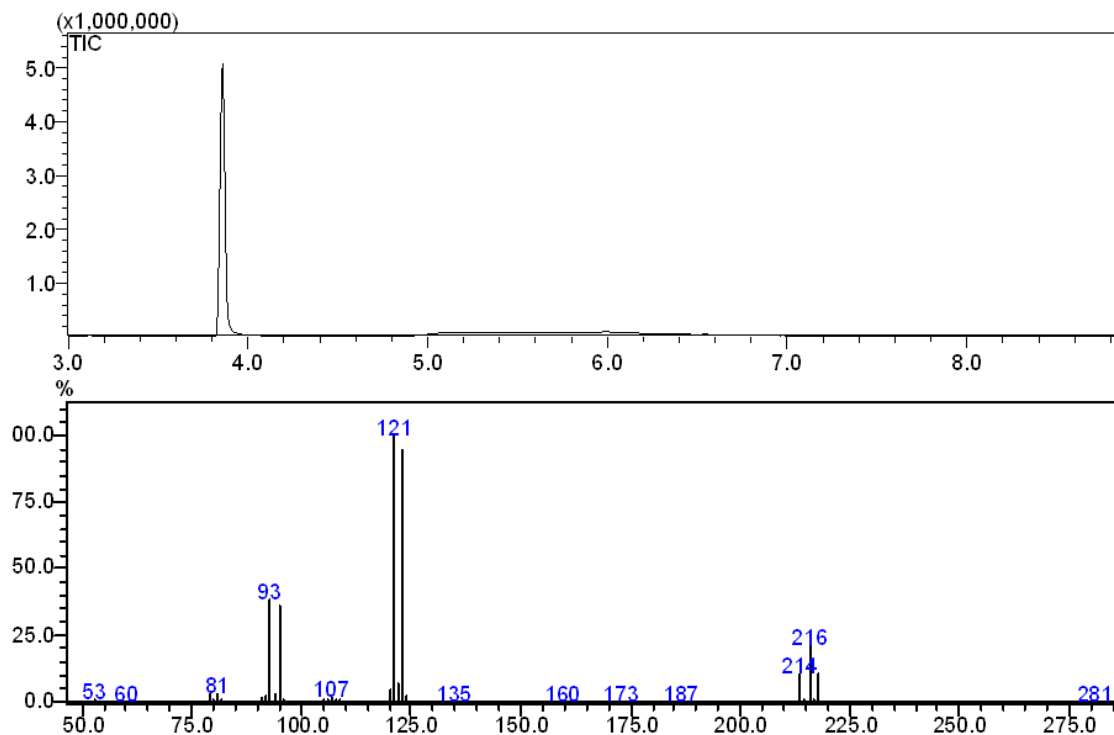


Figure S11. ^1H -NMR spectrum of 1,3-dibromoacetone in CDCl_3 .

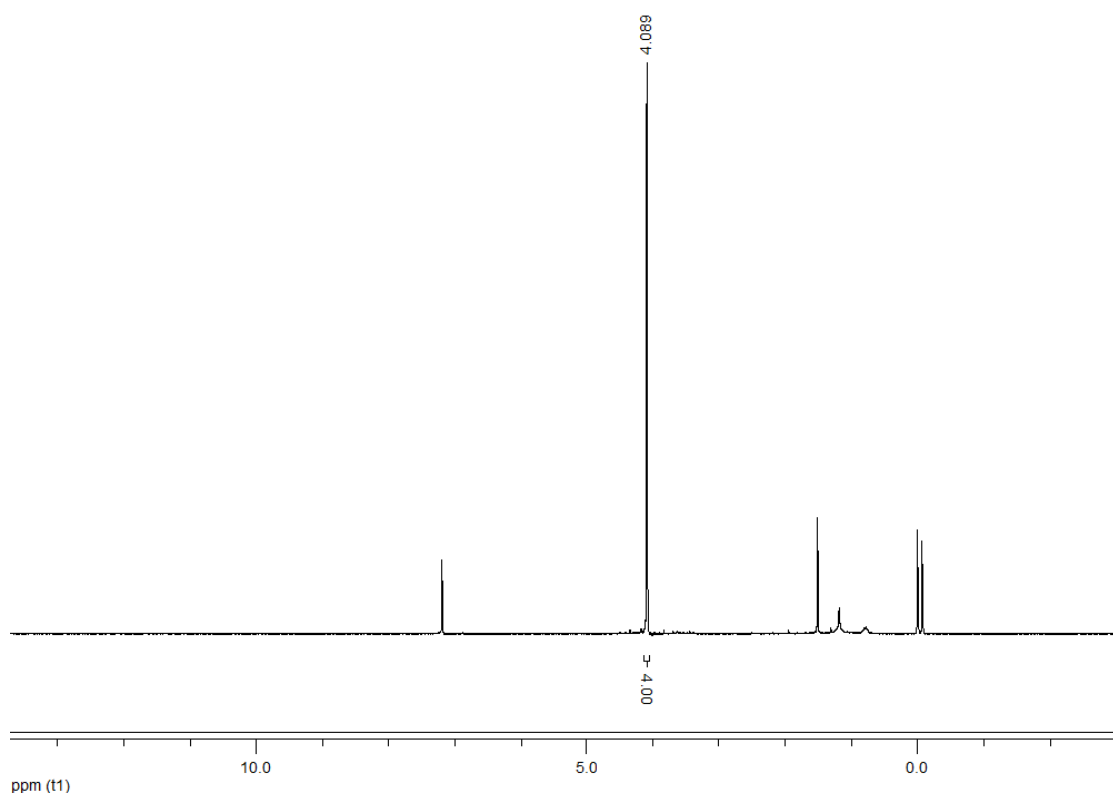


Figure S12. GC-MS chromatogram and mass spectrum of 1,3-dibromoacetone.

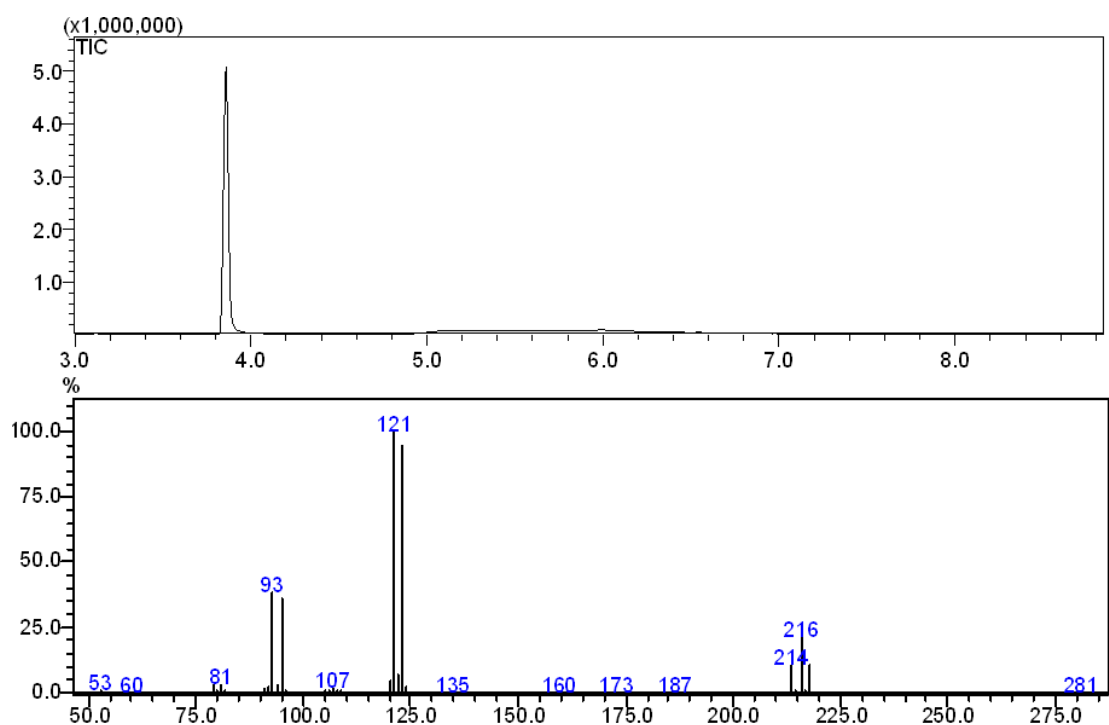


Figure S13. ^1H -NMR spectrum of BPB in DMSO- d_6 .

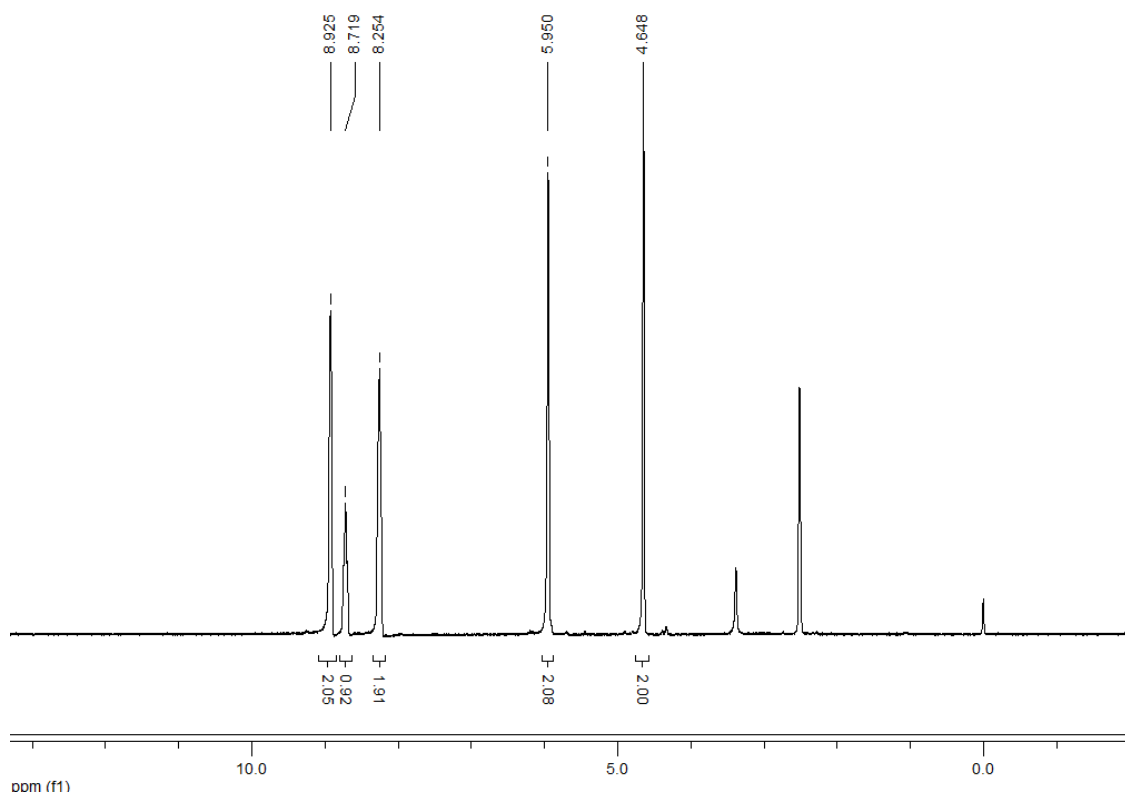


Figure S14. ^1H -NMR spectrum of d^5 -BPB in DMSO-d_6 .

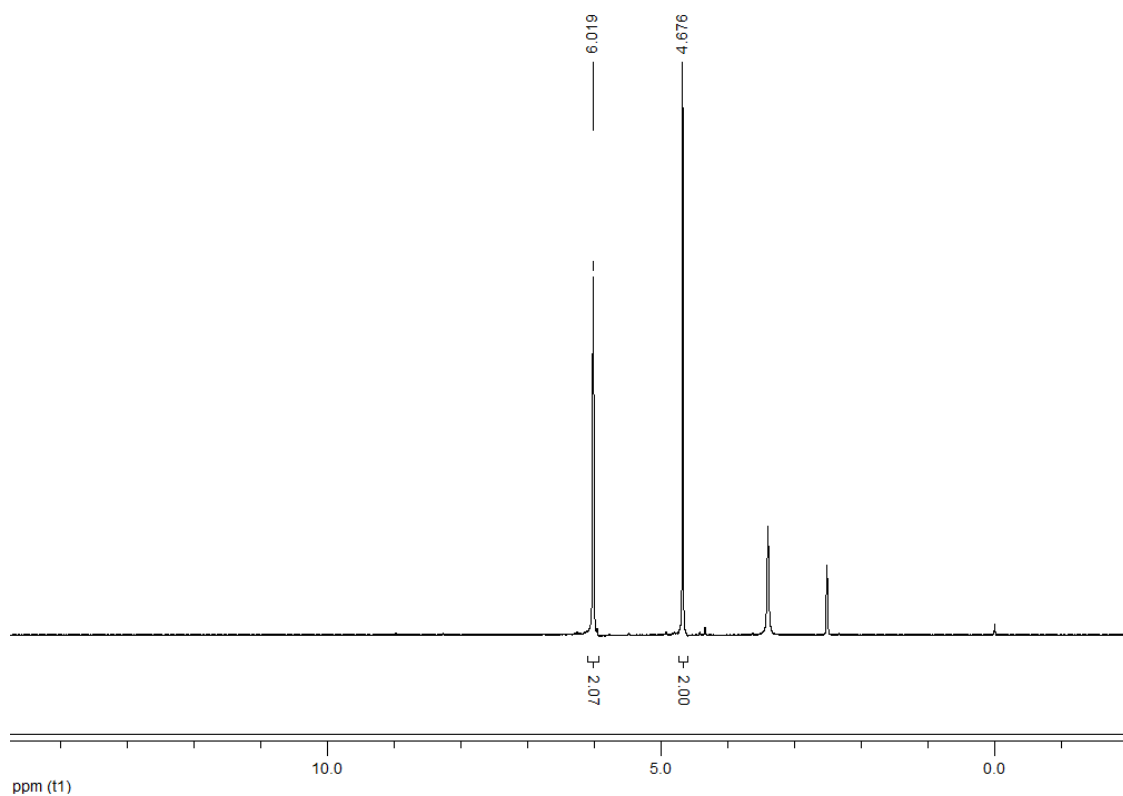


Figure S15. HR-MS spectrum of BPB.

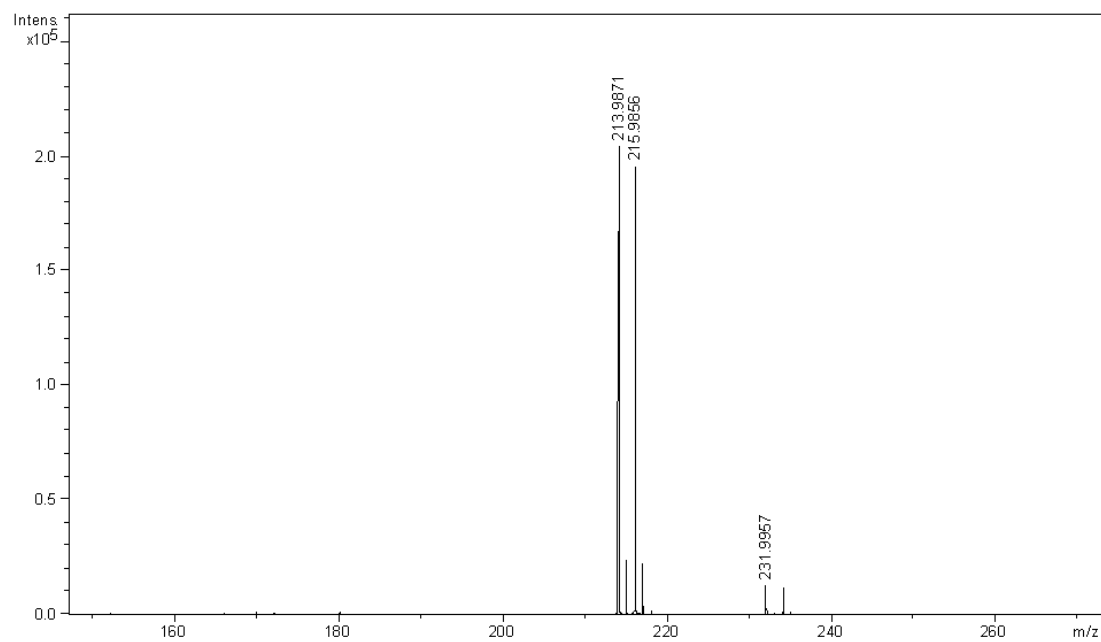


Figure S16. HR-MS spectrum of d⁵-BPB.

