

Supplementary Materials

Colorimetric Detection of Cd^{2+} Using Gold Nanoparticles Cofunctionalized with 6-Mercaptonicotinic Acid and L-Cysteine

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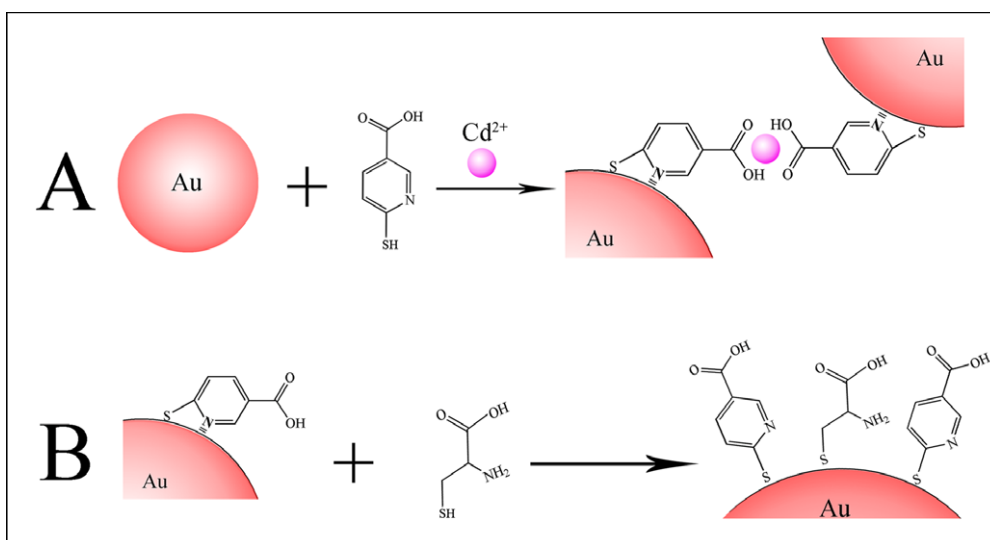


Fig. S1 The illustration of the binding modes of MNA and L-Cys with AuNPs in the case of MNA existed only (A) and MNA and L-Cys coexisted (B).

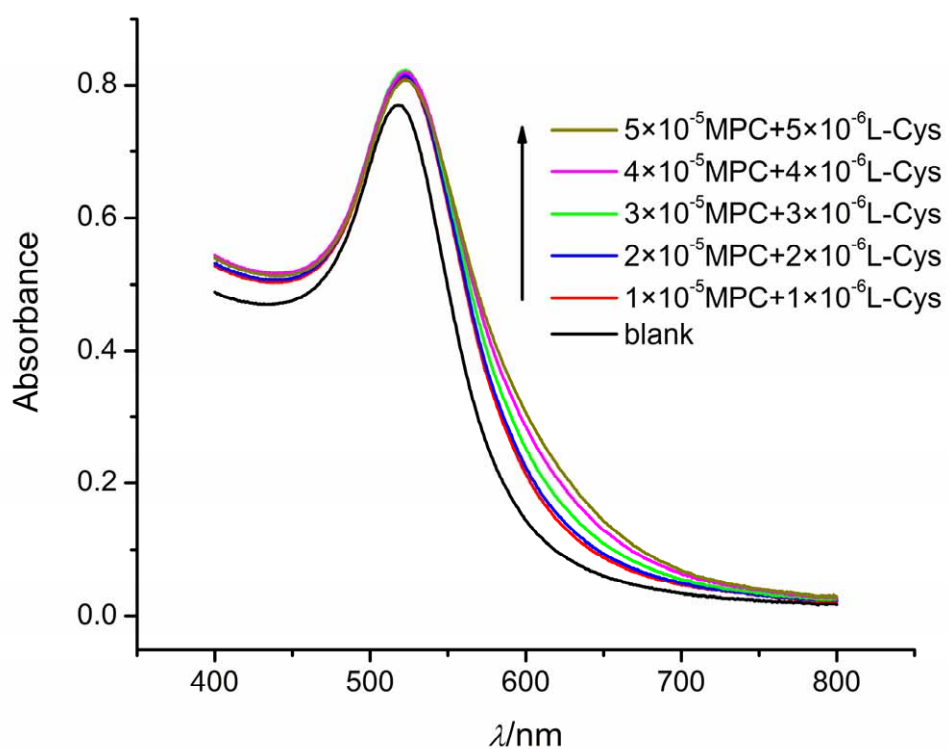


Fig. S2 UV-vis spectra of AuNPs functionalized with different concentrations of MNA and Cys.

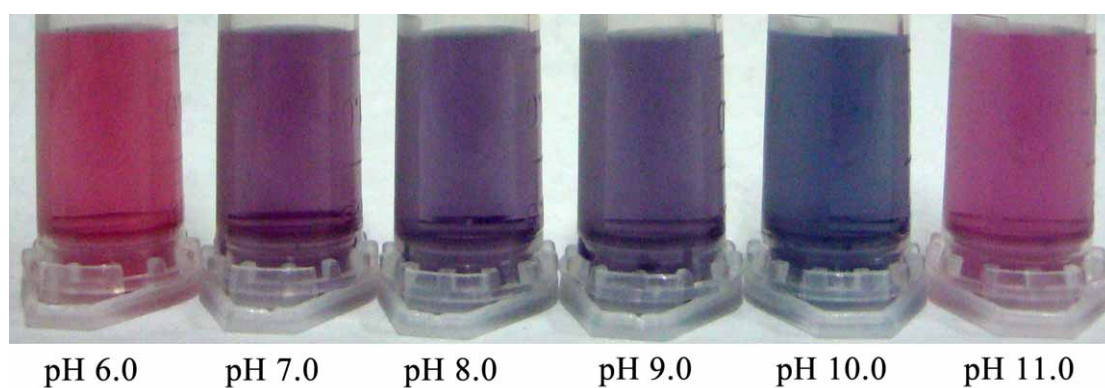


Fig. S3 The responses of MNA-L-Cys-AuNPs for 1×10^{-5} M Cd^{2+} in different pH values. The incubation time is 5 min.

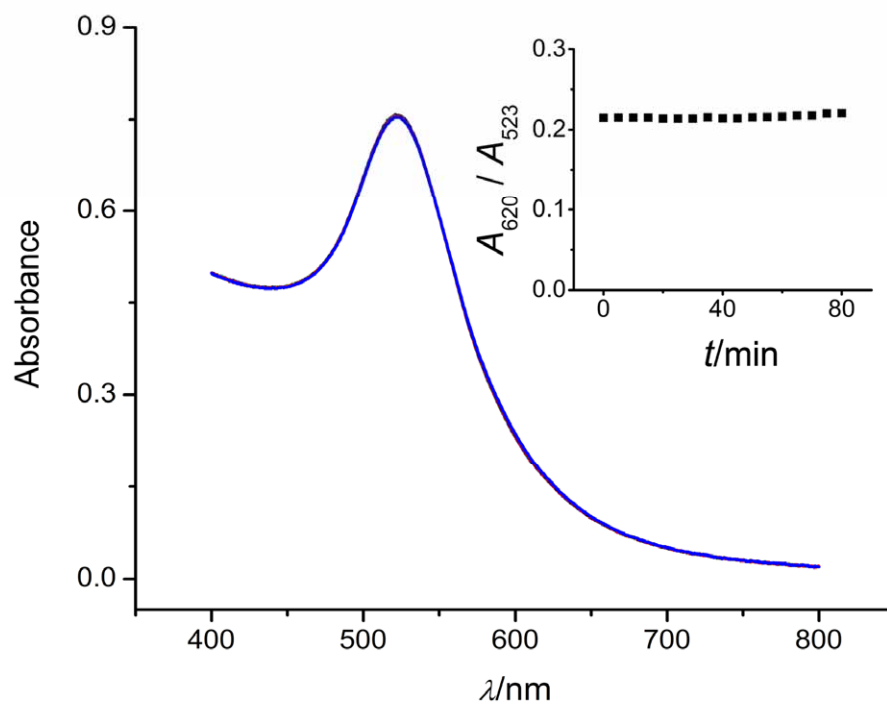


Fig. S4 The UV-vis spectra of MNA-L-Cys-AuNPs in presence 0.020 M NaCl during 80 min. The right inset shows time-dependent absorption ratio of A_{620}/A_{523} .

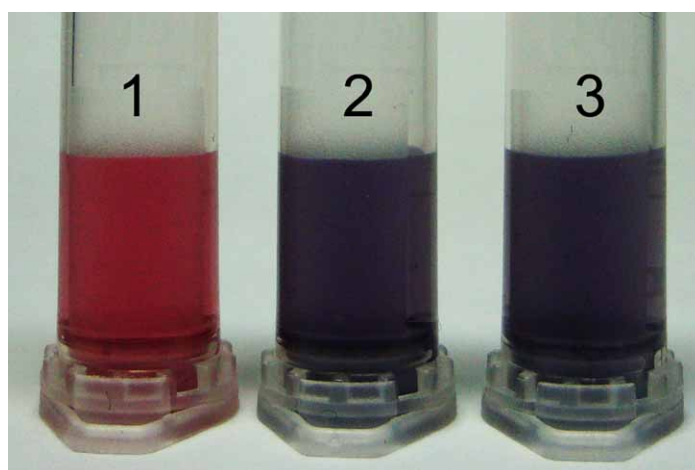


Fig. S5 The interaction of Hg^{2+} and MNA-L-Cys-AuNPs.

1: MNA-L-Cys-AuNPs containing 0.020 M NaCl; 2: MNA-L-Cys-AuNPs containing 0.020 M NaCl and 1×10^{-6} M Cd^{2+} ; 3: MNA-L-Cys-AuNPs containing 0.020 M NaCl and 1×10^{-6} M Hg^{2+} and Cd^{2+} .

Table S1 The comparison of dynamic range and detection limit in our work with the results previously reported.

Technique	Dynamic range ($\mu\text{g L}^{-1}$)	Detection limit ($\mu\text{g L}^{-1}$)	Reference
Solid-phase extraction–FAAS	10–4000	0.015	1
Cloud point extraction–FAAS	3–300	1	2
On-line pre-concentration–FAAS	5–40	0.02	3
Nafion-graphene/mercury film electrode–SWASV	1–7	0.08	4
Carbon nanotube tower electrode–SWASV	1–4	0.025	5
Immunochromatography	2.5–25	—	6
Near-infrared spectroscopy	$5.9\text{--}48.8 (\times 10^3)$	—	7
$\alpha,\beta,\gamma,\delta$ -Tetrakis(1-methylpyridinium-4-yl)porphine/silica membrane–Colorimetry	—	1	8
Triazole-ester modified AgNPs–Colorimetry	$2.8\text{--}56.2 (\times 10^3)$	2.2×10^3	9
MNA and L-Cys cofunctionalized AuNPs–Colorimetry	22.5–191.1	11.2	This work

Reference

- 1 A. Malekpoura, S. Hajialigol and M. A. Taherb, *J. Hazard. Mater.*, 2009, **172**, 229–233.
- 2 A. Afkhami, T. Madrakian and H. Siampour, *J. Hazard. Mater.*, 2006, **138**, 269–272.
- 3 W. N. L. dos Santos, J. L. O. Costa, R. G. O. Araujo, D. S. de Jesus and A. C. S. Costa, *J. Hazard. Mater.*, 2006, **137**, 1357–1361.
- 4 C. M. Willemse, K. Tlhomelang, N. Jahed, P. G. Baker and E. I. Iwuoha, *Sensors*, 2011, **11**, 3970–3987.
- 5 X. F. Guo, Y. H. Yun, V. N. Shanov, H. B. Halsall and W. R. Heineman, *Electroanalysis*, 2011, **23**, 1252–1259.
- 6 K. Abe, K. Nakamura, T. Arai, Y. Sakurai, A. Nakano, C. Suganuma, K. Tawaradae and K. Sasaki, *J. Sci. Food Agric.*, 2011, **91**, 1392–1397.
- 7 J. H. Li, Y. Zhang, W. S. Cai and X. G. Shao, *Talanta*, 2011, **84**, 679–683.
- 8 K. K. Latt and Y. Takahashi, *Anal. Chim. Acta*, 2011, **689**, 103–109.
- 9 H. B. Li, Y. Yao, C. P. Han and J. Y. Zhan, *Chem. Commun.*, 2009, 4812–4814.