

1 **Supplementary Information**

2

3 **Disposable Electrochemical Immunosensor for**
4 **Simultaneous Assay the Panel of Breast Cancer Tumor**
5 **Markers**

6 *Shenguang Ge^{1,2}, Feng Yu³, Lei Ge¹, Mei Yan¹, Jinghua Yu^{1*}, Dairong Chen^{2*}*

7

8 *1. Key Laboratory of Chemical Sensing & Analysis in Universities of Shandong*
9 *(University of Jinan), School of Chemistry and Chemical Engineering, University of*
10 *Jinan, Jinan 250022, China*

11 *2. School of Chemistry & Chemical Engineering, Technology Research Center for*
12 *Colloidal Materials, Shandong University, Jinan 250100, China*
13 *3. School of Mechanical and Aerospace Engineering, Nanyang Technological*
14 *University, 50 Nanyang Avenue, 639798, Singapore*

15

16

17

18 **Corresponding author: Jinghua Yu(ujn.yujh@gmail.com); Dairong Chen*
19 *(cdr@sdu.edu.cn).*

20 *Tel: +86-531-82767161. Fax: +86-531-82765969*

21

22 **1. Synthesis of Glutathione-Protected Au clusters**

23 Glutathione protected Au clusters (AuCs) with diameter 6.0 nm were prepared
24 according to the literature [1,2] with a slight modification by the reduction of
25 HAuCl₄·3H₂O using sodium borohydride in the presence of glutathione. In short, 5.0
26 mL 10 mmol/L of HAuCl₄ · 3H₂O and 2.5 mL 10 mmol/L of glutathione were added to
27 a mixture of solvents, methanol (4.5 mL) and acetic acid (1.0 mL) dissolved by stirring
28 for 5 min, resulting in a light yellow solution. The NaBH₄ solution (15 mg/mL) was
29 added drop by drop into above solution with vigorous stirring for 10 h. During this
30 period, the color of the colloid gradually changed, from yellow to brown, and the red
31 fluorescence emission indicated the nucleation of AuCs. The as-synthesized
32 glutathione-protected gold nanoparticles (GSH- AuCs) were soluble in water. The
33 particle solution was filtered through a 5000 MW cutoff, centrifuged at 4000 rpm
34 and washed with Milli-Q water for four times. The solution was then dissolved in 20
35 mmol/L PBS (pH 7.4) buffer.

36

37

38 **2. Synthesis of PVP-Protected Graphene**

39 Graphene oxide (GO) was synthesized from graphitic power according to Hummer's
40 method with some modification [3,4]. In brief, graphite powder (3 g, 325 mesh) was
41 added into a mixture of 12 mL of concentrated H₂SO₄, 2.5 g of K₂S₂O₈, and 2.5 g of
42 P₂O₅, and reacted for 5 h at 80 °C. Next, the mixture was cooled to room

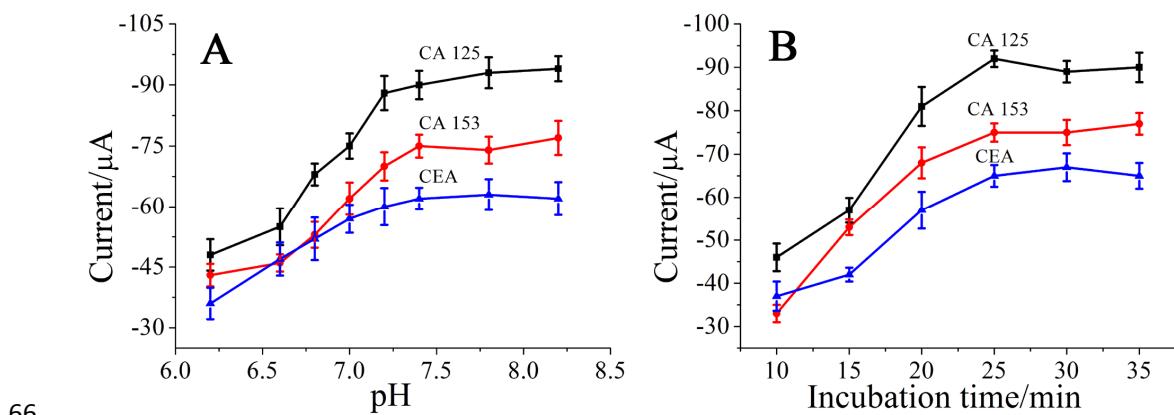
43 temperature and diluted with 500 mL Milli-Q water. After residual acid was removed
44 by filtrating and washing through 0.2 μm nylon film with water, the product was
45 dried naturally overnight and added to concentrated H_2SO_4 (120 mL). Successively,
46 KMnO_4 (15 g) was added gradually under stirring, while keeping the temperature
47 around 5 $^{\circ}\text{C}$. This mixture was stirred at 40 $^{\circ}\text{C}$ for 50 min. Then, the mixture was
48 diluted with water (250 mL) and kept at 55 $^{\circ}\text{C}$ for 30 min. After the resulting
49 mixture was stirred for 2 h, 600 mL of water was then injected into the mixture
50 followed by adding 30 mL of 30 % H_2O_2 dropwise. The mixture was filtered and
51 washed with 1 mol/L HCl aqueous solution and water many times to remove
52 residues. Finally, the product was further purified by dialysis for 1 week to remove
53 the remaining metal species. By sonicating dispersion under ambient conditions for
54 30 min, the homogeneous GO suspension (0.1 mg/mL) was obtained, which was
55 stable for several months. The PVP/GNs was prepared by a modified method
56 according to the literature [5,6]. In a typical experiment for chemical conversion of
57 graphene oxide to PVP/GNs, 400 mg of PVP was put into 100 mL of homogeneous
58 GO dispersion (0.25 mg/mL), followed by stirring for 12 h. Then, to the resulting
59 dispersion were added 30 mL of hydrazine solution (80 %). The mixture was stirred
60 for 12 h at 90 $^{\circ}\text{C}$. Finally, the stable black dispersion was centrifuged two times and
61 redispersed in 25 mL of water (1 mg/mL).

62

63

64

65 **3. Optimization of pH and Incubation time**



67 Figure S1 Effect of pH(A) and incubation time(B) on stripping current of Ag NPs in 1.0
68 M KCl for 10.0 U/mL CA 153, CA 125 and 10.0 ng/mL CEA.

69

70 **Reference**

- 71 [1] M. Vigneshwaran, V.C. Bhaskara, P. Vyomesh, J.S. Gutkind, F.R. James, ACS NANO,
72 2009, 3, 585-589.
- 73 [2] M. Zheng, X. Huang, J. Am. Chem. Soc., 2004, 126, 12047-12054.
- 74 [3] W.B. Jacob, G. Srinivas, F. Jamie, M.S. Jason, W. Y. Taner, Angew. Chem. Int. Ed.,
75 2010, 49, 8902-8904.
- 76 [4] X.M. Wu, Y.J. Hu, J. Jin, N.L. Zhou, P. Wu, H. Zhang, Anal. Chem., 2010, 82,
77 3588-3596.
- 78 [5] D. Li, M.B. Muller, S. GiljE, R.B. Kaner, G.G. Wallace, Nat. Nanotechnol., 2008, 3,
79 101-105.
- 80 [6] C.S. Shan, H.F. Yang, J.F. Song, D.X. Han, I. Ari, N. Li, Anal. Chem., 2009, 81,
81 2378-2382.