

SUPPORTING INFORMATION

Crude to Leads: A triple-pronged direct NMR approach in coordination with docking simulation

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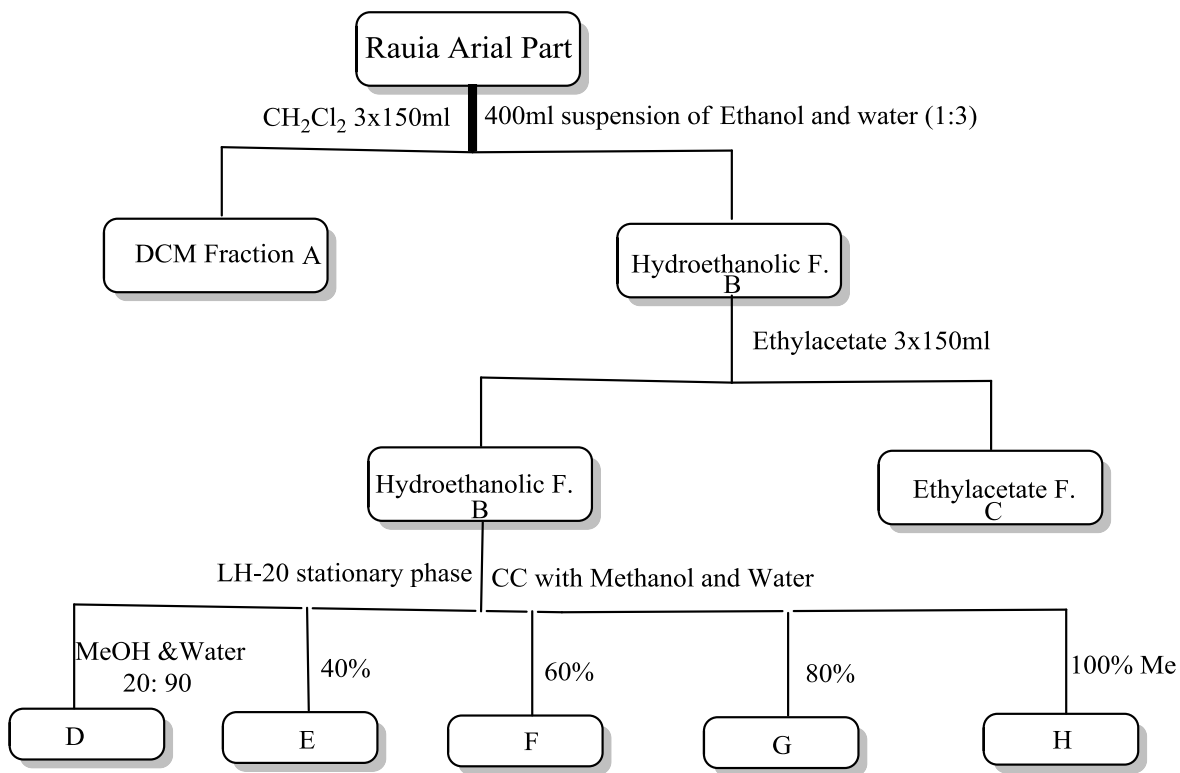


Figure S-1. Isolation Scheme for arial parts of Rauia plant

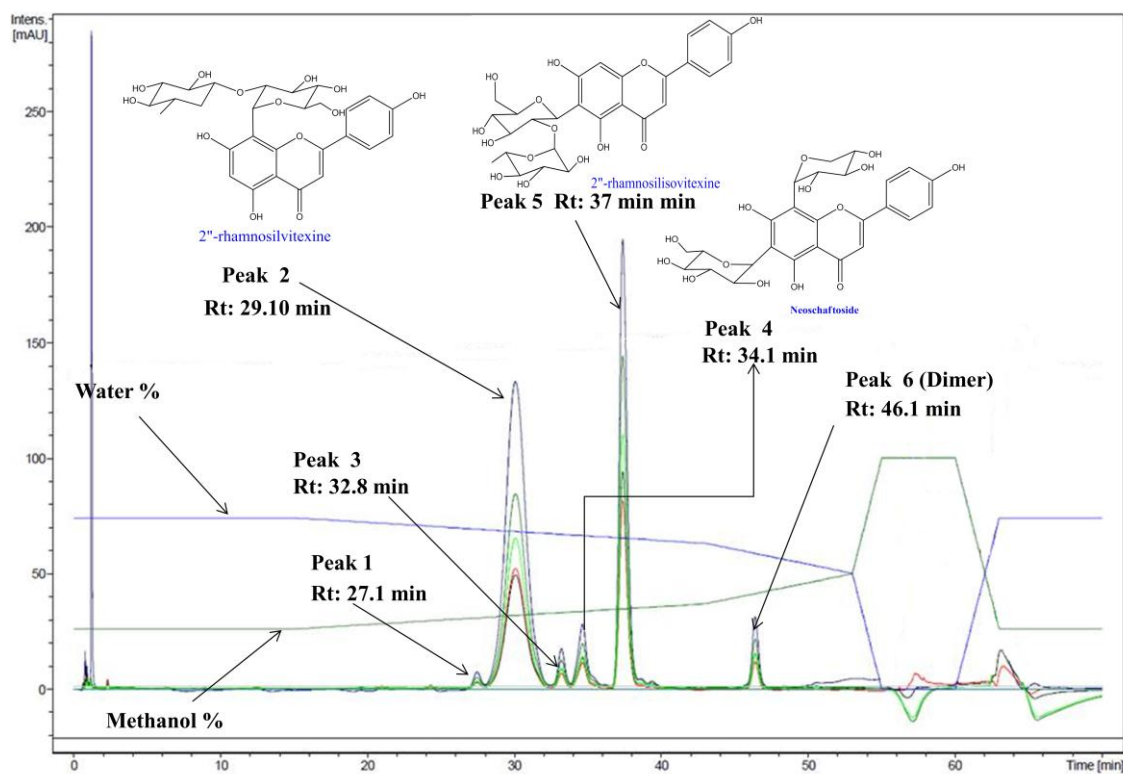


Figure S-2. Chromatogram obtained as result of separation with LC-Method by using water and methanol as solvent at different gradients.

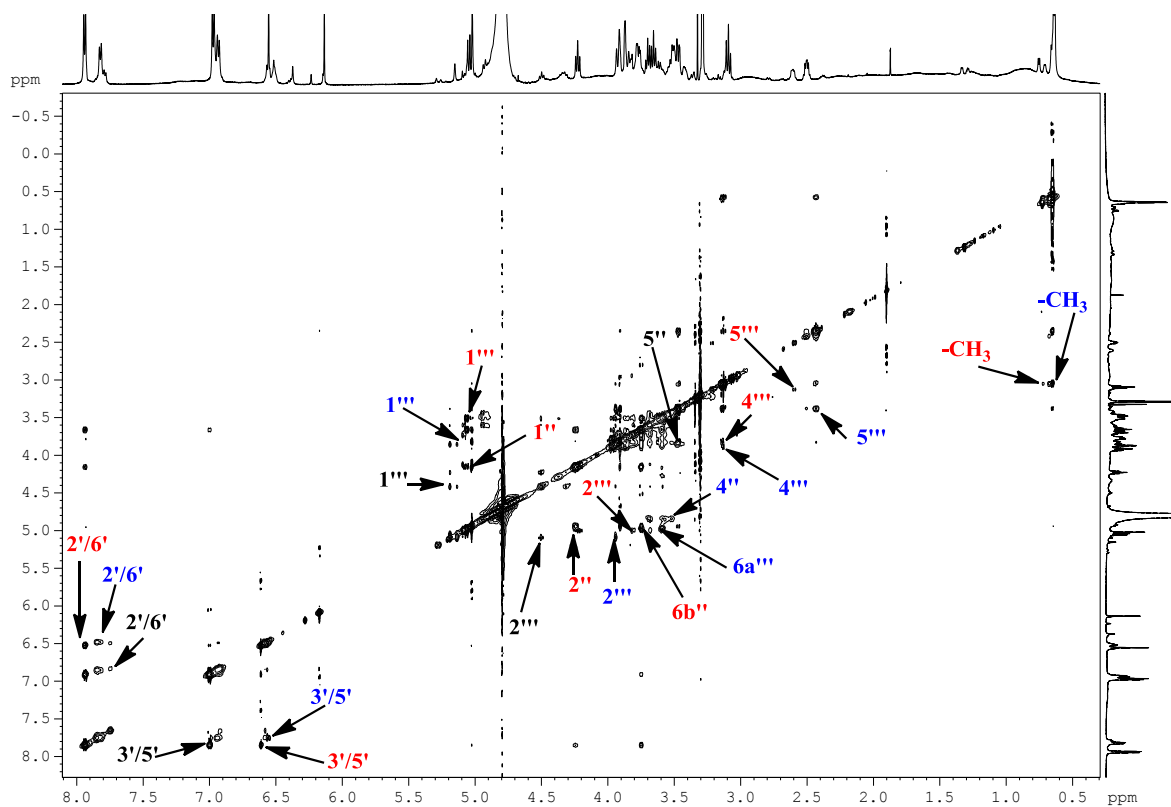


Figure S-3. Two-dimensional transfer NOESY with Bruker standard phase sensitive pulse sequence for the extract in D₂O and methanol (95:5% v/v) and BSA in buffer solution. In transfer NOESY pulse sequence is modified by putting the 3500Hz spin lock for the removal of unwanted protein signal before the first $\pi/2$ pulse. This spectrum is produced by the use of 400ms mixing time with at 600 MHz Bruker NMR spectrometer with a cryoprobe TCI at 298 K. NOE cross-peaks shown are of the same sign as of the diagonal signals (Negative NOEs).

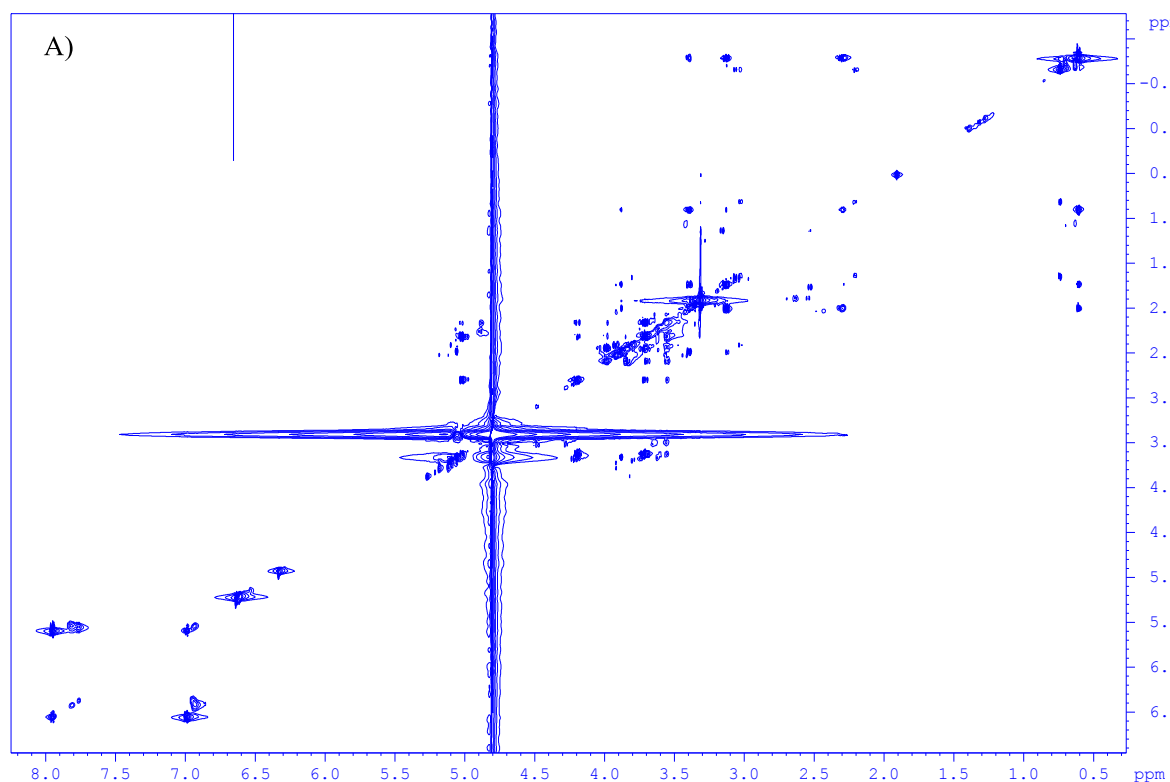


Figure S-4a. A standard 2D TOCSY NMR spectrum of crude extracts with 50ul BSA is shown.

Experiment was done by using a Malcolm Levitt's composite-pulse decoupling sequence (MLEV-17) with mixing time of 60 ms at 298K with 600MHz Spectrometer. For standard TOCSY MLEV-17 sequence the experimented was recorded with 7211.539Hz spectral width in both dimensions (F_1 and F_2) with a total number of 56 scans, with 256 increments in t_1 were collected with 2s relaxation delay for each scan; with the experimental time less than 10hrs..Other conditions are according to the experimental section. TOCSY spectrum was acquired in 1mg of the extract in D_2O : methanol (95:5% v/v) and BSA in buffer (7.4pH) solution respectively.

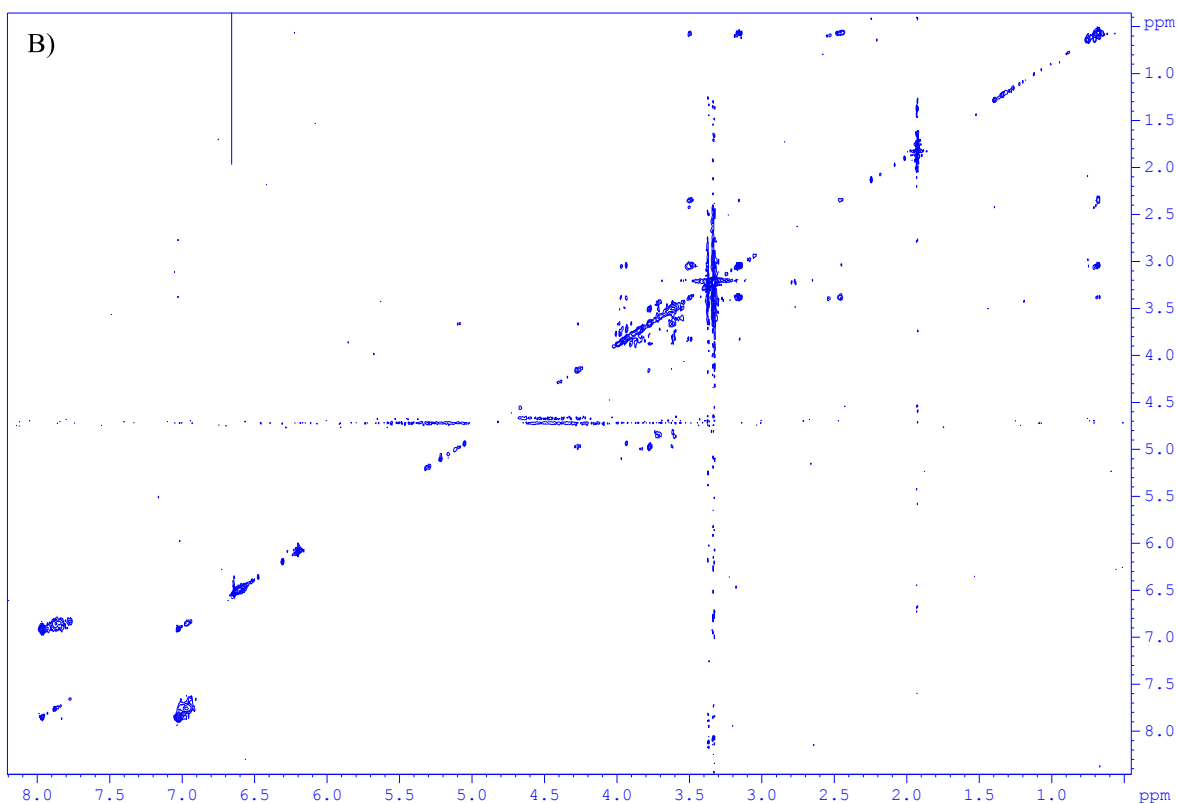


Figure S-4b. Two-dimensional STD-TOCSY NMR spectra of crude extracts with 50 μ M BSA by using a Malcolm Levitt's composite-pulse decoupling sequence (MLEV-17) with mixing time of 60 ms at 298K with 600MHz Spectrometer. In STD-TOCSY saturation of the protein was achieved with a cascade of 50Gaussian pulse (50 ms duration each) with a delay of 200 ms in between each pulse, with a total saturation time of 2.5 s. The on-resonance saturation frequency was set to -0.5 ppm and off-resonance to 30 ppm. 2D STD-TOCSY Spectrum was recorded with 7211.539Hz spectral width in both dimensions (F_1 and F_2) with 64 transients in t_2 and with 384 increments for the *on* and *off* resonance in t_1 with the total experimental time of more than 16hrs. Both spectra (2D standard and STD TOCSY) were acquired in 1mg of the extract in D₂O: methanol (95:5% v/v) and BSA in buffer(7.4pH) solution respectively.

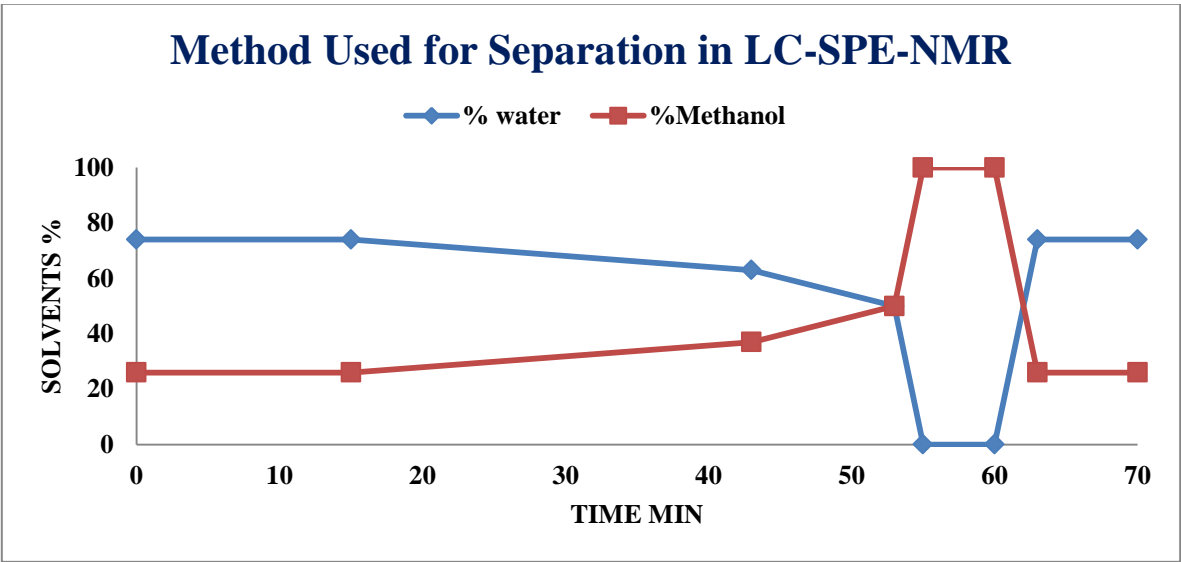


Figure S-5. Separation method and the solvent gradient used in LC-Method

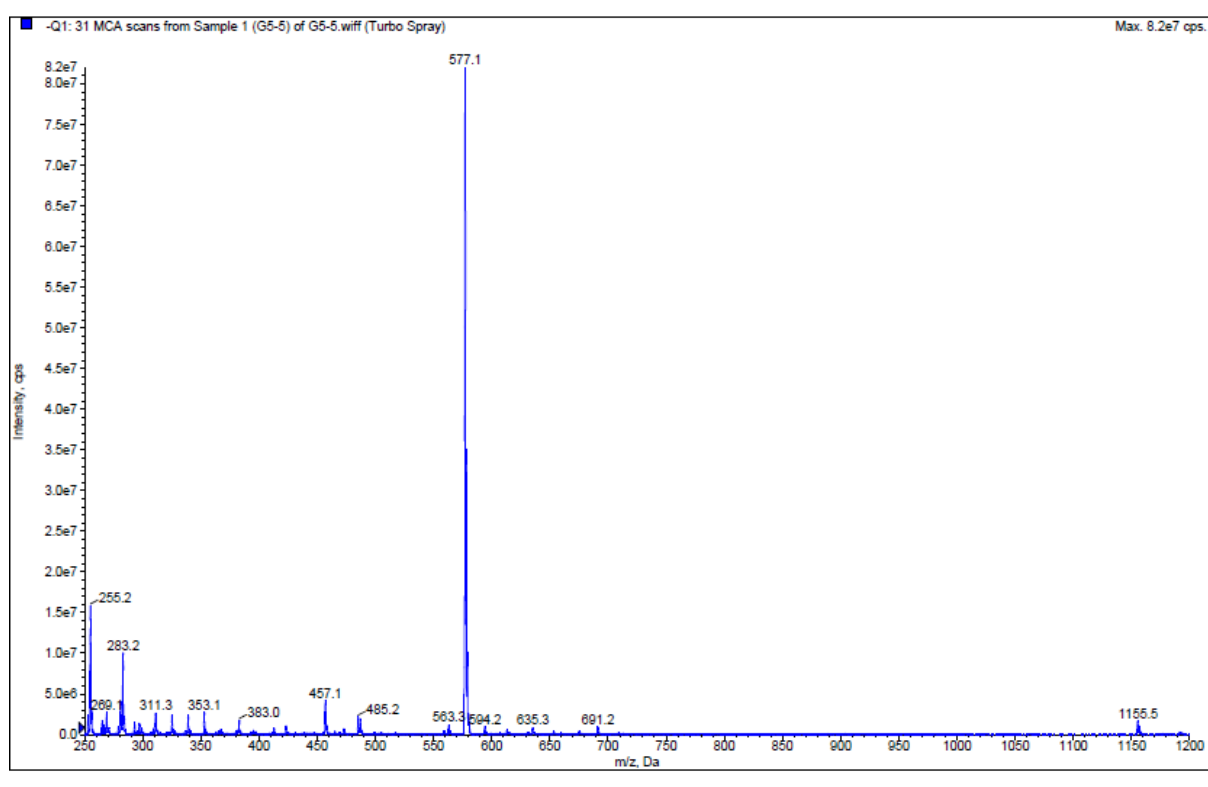


Figure S-6. ESI spectrum of peak 6 of LC-chromatogram, showing the molecular weight 1155.5 a.m.u.

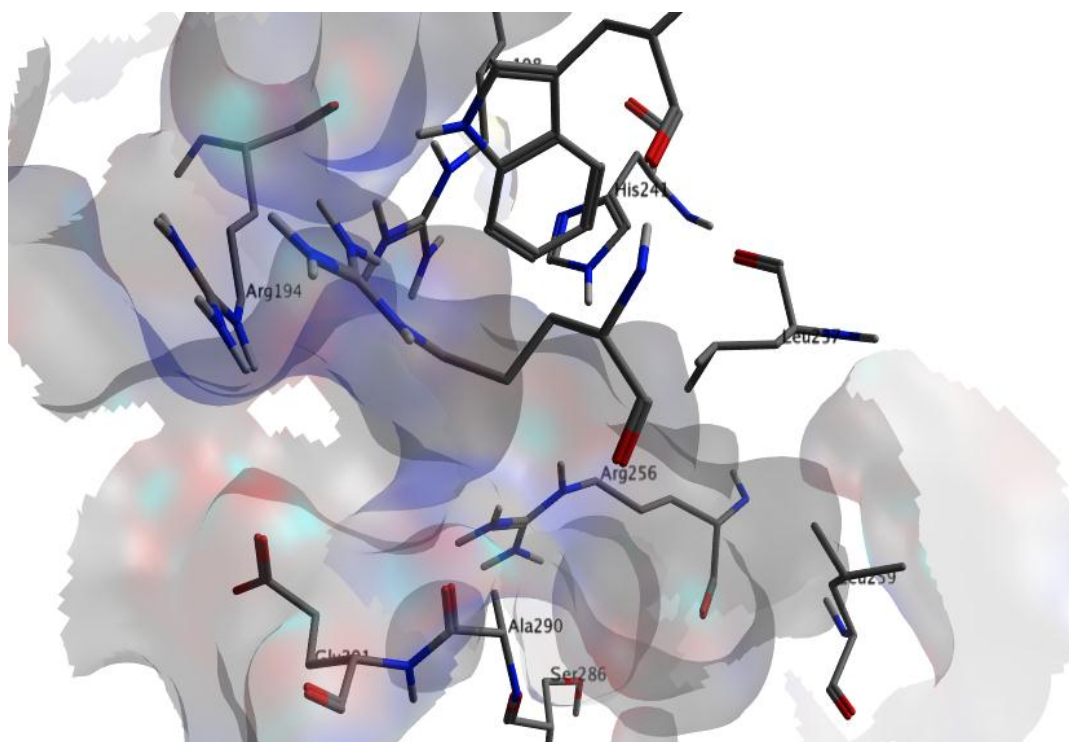


Figure S-7: Selected Active site of chain B of the Bovine Serum Albumin (BSA), generated by using Molecular Operating Environment (MOE) 2011.10. BSA PDB file was taken from protein data bank (<http://www.rcsb.org/pdb/explore.do?structureId=4F5S>) (4F5S). The classical binding sites were marked in corresponding subdomain cations.

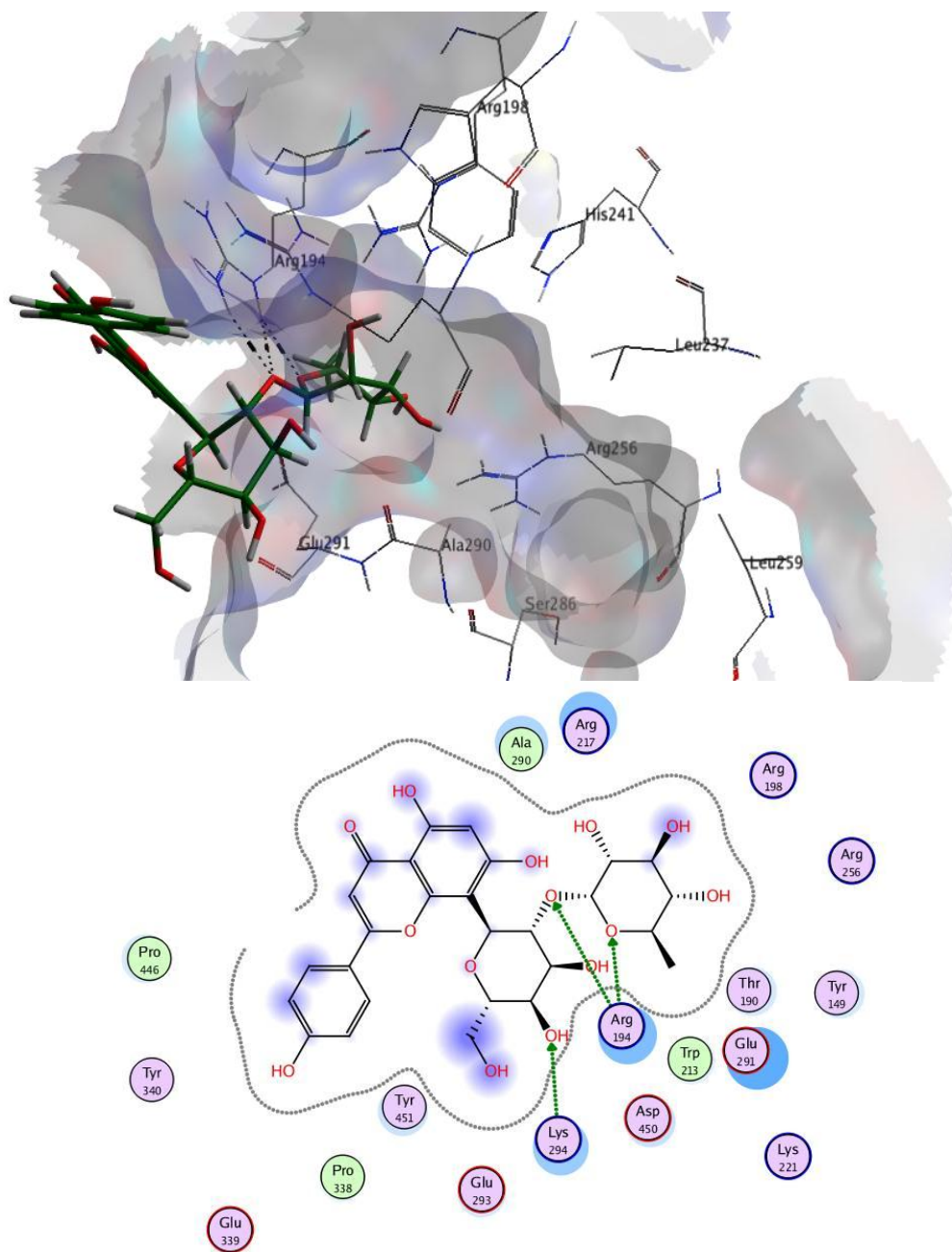


Figure S-8a-b: Molecular Docking model of interactions between the Bovine Serum Albumin (BSA) and Vetexine-2-rhamnoside, **1**, generated by using Molecular Operating Environment (MOE) 2011.10. BSA PDB file was taken from protein data bank (<http://www.rcsb.org/pdb/explore.do?structureId=4F5S>) (4F5S). The classical binding sites were marked in corresponding subdomain cations. See more details in experimental section.

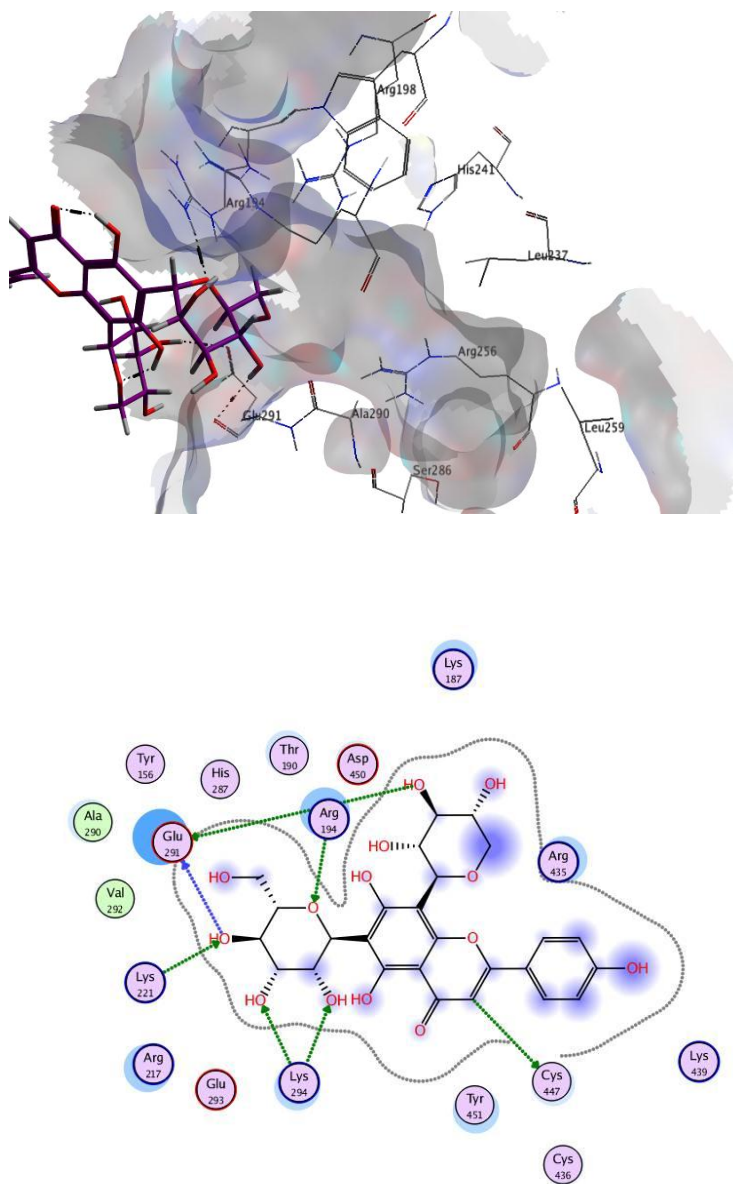


Figure S-9a-b: Molecular Docking model of interactions between the Bovine Serum Albumin (BSA) and Neoschiftosoide, **3**, generated by using Molecular Operating Environment (MOE) 2011.10. BSA PDB file was taken from protein data bank (<http://www.rcsb.org/pdb/explore.do?structureId=4F5S>) (4F5S). The classical binding sites were marked in corresponding subdomain cations. See more details in experimental section.