# Lateral flow assay for identification of *Escherichia coli* by ribosomal RNA hybridisation

Christopher Pöhlmann,<sup>*a,b,\**</sup> Irina Dieser<sup>*a*</sup> and Mathias Sprinzl<sup>*a*</sup>

<sup>a</sup> University of Bayreuth, Department of Biochemistry, Universitätsstr. 30, 95440 Bayreuth, Germany. Fax: 49-921-55-4396; Tel: 49-921-55-4355; E-mail: mathias.sprinzl@unibayreuth.de

<sup>b</sup> Present address: Bruker Daltonik GmbH, Permoserstr. 15 04318 Leipzig, Germany. Fax: 49-341-2431-404; Tel: 49-341-2431-496; E-Mail: <u>Christopher.Poehlmann@bdal.de</u>

## **Supporting information**

## Estimation of yield of RNA isolation using rapid "one step" lysis method



**Fig. S1** Peak area of test line (grey bars) of RNALFA applying total RNA isolated from  $5 \times 10^7$  cfu mL<sup>-1</sup> *E. coli* 163 using the spin column method (reference method) and the rapid "one step" lysis method. The peak areas were quantified using ImageJ software. Yield of rapid "one step" isolation method in comparison to reference method is  $43.6\% \pm 8.3\%$  (n = 3).



### Quantitation of signal strength of different RNALFA hybridisation assemblies

**Fig. S2** Peak area of test (black bars) and control line (grey bars) in dependence of different hybridisation assemblies. Images of the strips are depicted in Fig. 3. Three (A, C) components and four (B, D, E, F, G) components hybridisation with one or two AuNP-labelled ODNs are shown. Total RNA was isolated from 5 x  $10^7$  cfu mL<sup>-1</sup> *E. coli* 163. (A) shows the peak area resulting from three components hybridisation composed of EC\_Capture, AuNP-UD1072 and the target RNA. (B) is same as (A) plus the presence of the helper ODN Helper-UD1072. (C) is a three components hybridisation in presence of EC\_Capture and AuNP-UD1082 (missing helper ODN). Four components assemblies are depicted in (D), (E) and (F). (D) is the four components assembly composed of EC\_Capture, Helper and UD1082-AuNP, whereas (E) depicts an analogous approach, however, in presence of RNA purified from 5 x  $10^8$  cfu mL<sup>-1</sup> *B. subtilis* instead of *E. coli* RNA and in the absence of *E. coli* RNA, respectively. Biotinylated capture ODN, helper ODN and detector ODN are depicted in red, green and yellow, respectively. Control line exhibited a average signal strength of  $1625 \pm 207$  (n = 21).

### Detection of E. coli in human urine samples

Human urine was filtered through a 0.2  $\mu$ m syringe filter (Millipore, Billerica, MA, USA). Urine sample was artificially contaminated with 0, 5 x 10<sup>7</sup> or 5 x 10<sup>5</sup> cfu mL<sup>-1</sup> *E. coli* 163, respectively. A 1 mL urine sample was centrifuged at 5,000xg for min and 25°C. RNA was isolated using rapid "one step" lysis method as described.



**Fig. S3** RNALFA analysis of urine sample artificially contaminate with 5 x  $10^7$  or 5 x  $10^5$  cfu mL<sup>-1</sup>. As negative control a urine sample without *E. coli* was tested (0 cfu mL<sup>-1</sup>). Control (Co) and test (T) lines are indicated.