Electronic Supplementary Information

# Electrochemiluminescence immunosensor using poly(Lhistidine) protected glucose dehydrogenase on Pt/Au bimetallic nanoparticles to in situ generate co-reactant

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#### The preparation of 0.25 wt % chitosan solution

0.25 wt % chitosan solutions were prepared by dissolving 25 mg chitosan in 10 mL 1% acetic acid solution with magnetic stirring for ~2 h.

## The preparation of AuNPs

AuNPs were synthesized according to the previous report with a little modification<sup>1</sup>. Briefly, 1.0 mL of 1% HAuCl4 was diluted into 100.0 mL with double-distilled water and brought to reflux while stirring. Subsequently, 4.0 mL of 1% trisodium citrate solution was added quickly, resulting in a color change from pale yellow to wine red. After that, the solution was refluxed for another 15 min.

## The Characterization of the Au@RuSiO<sub>2</sub> NPs

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Fig. S2A showed the UV-vis absorption spectra of Au NPs, Ru(phen)<sub>3</sub><sup>2+</sup>, RuSiO<sub>2</sub> NPs, Au@RuSiO<sub>2</sub> NPs, respectively. Obviously, the absorbance peak of Au NPs colloid was appeared at 520 nm (Fig. S2A (a)). Ru(phen)<sub>3</sub><sup>2+</sup> exhibited there characteristic peaks at 447 nm, 281 nm and 222 nm (Fig. S2A (b)). The synthesized RuSiO<sub>2</sub> NPs had similar peaks (Fig. S2A (c)), suggesting that many Ru(phen)<sub>3</sub><sup>2+</sup> molecules were doped into SiO<sub>2</sub> NPs successfully through the electrostatic interaction between Ru(phen)<sub>3</sub><sup>2+</sup> and silica nanoparticles. After that, Au NPs were assembled onto the RuSiO<sub>2</sub> NPs surfaces with the aid of BSA. The spectrum of Au@RuSiO<sub>2</sub> NPs shows the characteristic absorption peaks of Au NPs and RuSiO<sub>2</sub> NPs (Fig. S2A (d)), indicating the Au@RuSiO<sub>2</sub> NPs were prepared successfully.

In addition, scanning electron microscopy (SEM) was also used to characterize the synthesis of Au@RuSiO<sub>2</sub> NPs. As shown in Fig. S2B, the RuSiO<sub>2</sub> NPs were well-dispersed particles with a uniform diameter of ~150 nm, which were much larger than Au NPs, thus mangy Au NPs can load on RuSiO<sub>2</sub> NPs to form Au@RuSiO<sub>2</sub> NPs. Fig. S2C shown that the RuSiO<sub>2</sub> NPs were covered with a great deal of small spherical Au NPs, which demonstrated that the Au@RuSiO<sub>2</sub> NPs were prepared successfully.

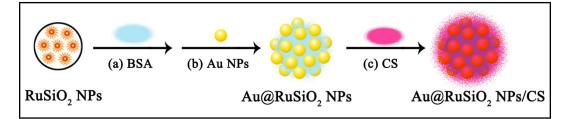


Fig. S1 Preparation procedures of Au@RuSiO<sub>2</sub> NPs/CS.

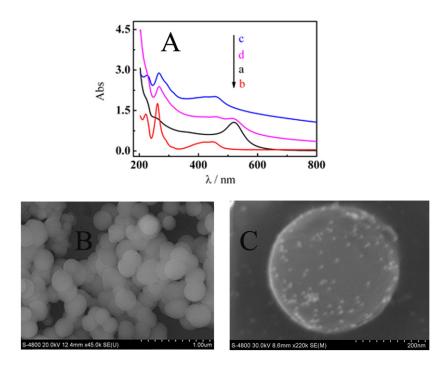


Fig. S2 (A) UV-vis spectra of Au NPs (a, black),  $Ru(phen)_3^{2+}$  (b, red),  $RuSiO_2$  NPs (c, blue), Au@RuSiO\_2 NPs (d, magenta), (B) SEM images of RuSiO\_2 NPs and (C) SEM images of Au@RuSiO\_2 NPs.

#### CV Characterization of the immunosensor fabrication

To gain a better understanding of the fabrication process, the cyclic voltammograms (CVs) experiments were also performed in 5 mM  $[Fe(CN)_6]^{3-/4-}$  solution. As shown in Fig. S3, a pair of well-defined redox peak of  $[Fe(CN)_6]^{3-/4-}$  was observed on the pretreated bare GCE (curve a). When Au@RuSiO<sub>2</sub> NPs/CS complex were dropped onto the electrode, the current decreased clearly due to the insulating properties of CS (curve b). After the successive immobilization of Ab<sub>1</sub>, BSA and cTnI, the peak current further decreased in order (curve c, d and e). That was because the formation of protein molecules layers hindered the electron transfer.

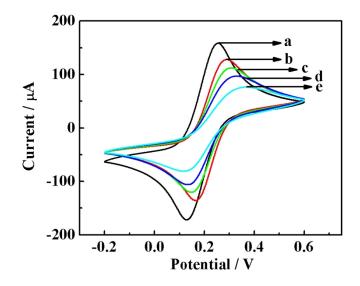


Fig. S3 CV for (a) bare GCE, (b) Au@RuSiO<sub>2</sub> NPs/CS/GCE, (c) Ab<sub>1</sub>/Au@RuSiO<sub>2</sub> NPs/CS/GCE, (d) BSA/Ab<sub>1</sub>/Au@RuSiO<sub>2</sub> NPs/CS/GCE, (e) cTnI/BSA/Ab<sub>1</sub>/Au@RuSiO<sub>2</sub> NPs /CS/GCE, (f) Pt/Au NPs@GDH-PLH-Ab2/cTnI/BSA/Ab1/Au@RuSiO2 NPs/CS/GCE, in 0.1 M KCl solution containing 5 mM [Fe(CN)<sub>6</sub>]<sup>3-/4-</sup>. Scan rate, 100 mV s<sup>-1</sup>.

#### Comparisons of proposed immunosensor with other detection methodologies for

## cTnI detection

Table S1 Performance compared with other detection methodologies for cTnI detection.			
Detection method	Linear range/ng mL <sup>-1</sup>	Detection limit/pg mL <sup>-1</sup>	Ref.
Surface Acoustic Wave	$0.02 \sim 100$	20	2
Surface Plasmon	0.03 ~ 6.5	10	3
Resonance			
Surface Plasmon	0.05 ~ 4.5	50	4
Resonance			
Localized Surface	$1 \sim 20$	300	5
Plasmon Resonance			
Electrochemiluminescent	0.01 ~ 5	4.5	6
Electrochemiluminescent	0.01 ~ 10	3.33	Our work

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From the Table S1 we can see that the proposed immunosensor has a relative large linear range and low detection limit compared with previous reports.

## Reference

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