

## Supporting Information

### **Monodispersed nanoparticles of conjugated polyelectrolyte brush with high charge density for rapid, specific and label-free detection of tumor marker**

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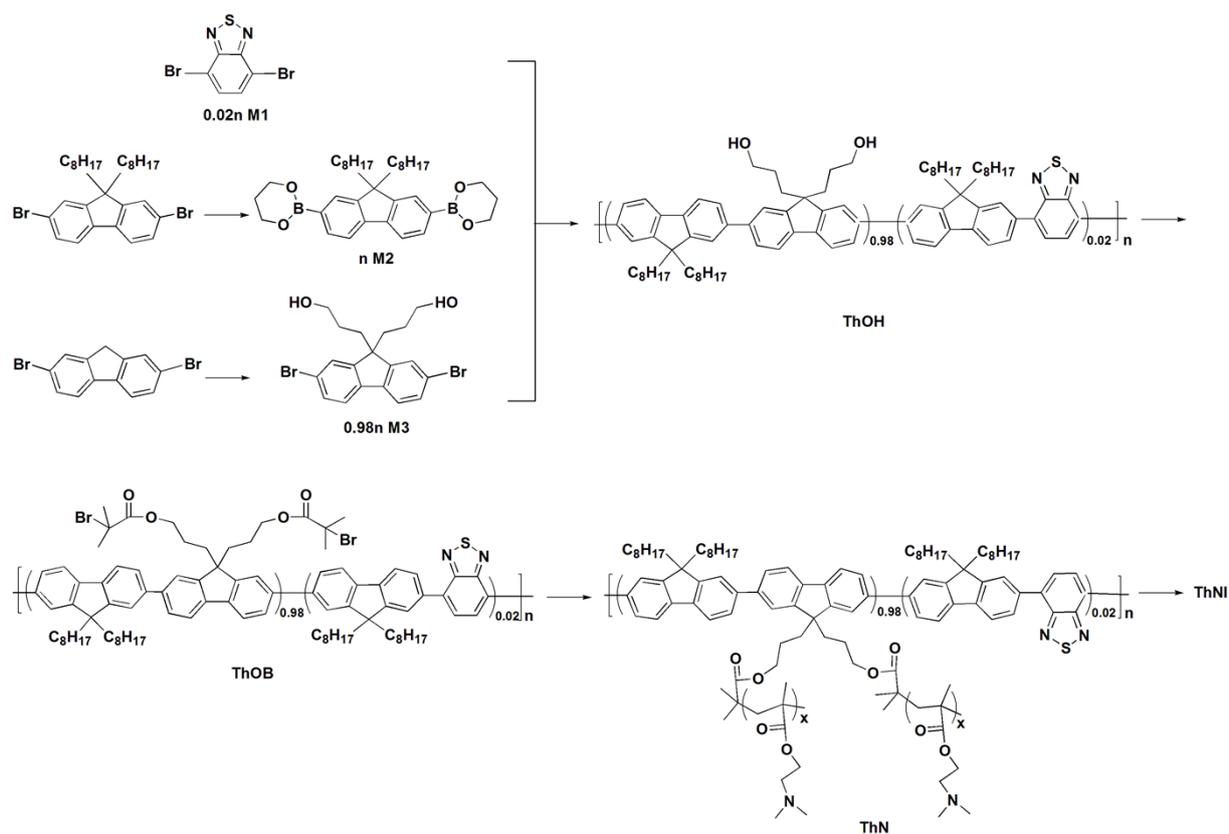
#### **EXPERIMENTAL SECTION**

**Materials and Chemicals.** The concentration of ThNI is calculated in repeat units of monomer. Human  $\alpha$ -fetoprotein, prostate specific antigen, and immunoglobulin were purchased from Linc-bio Science Co., Ltd (Shanghai, China). Thrombin and lysozyme were purchased from Sigma-Aldrich. Bovine serum albumin was purchased from Hangzhou Haoxin Biotech Co., Ltd. Other chemicals were purchased from Sigma-Aldrich, Acros, and Alfa and were used as received. All solutions were prepared with Milli-Q water (18.2 M $\Omega$ .cm) from a Millipore system.

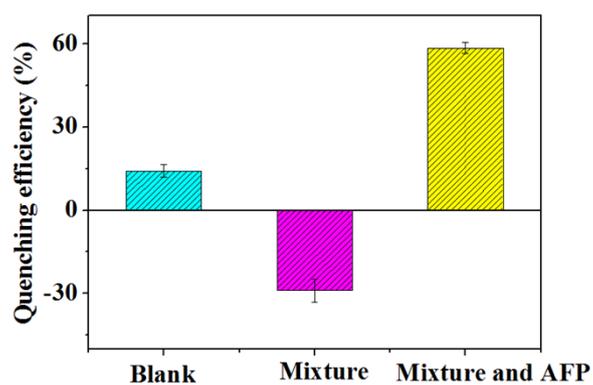
**Experimental Procedures.** Protein solutions of certain volume were added to 2  $\mu$ L ThNI solution (100  $\mu$ M), after an incubation at 37°C for 10 min, PBS buffer (20 mM PBS, 140 mM NaCl, pH 7.4) was add to a final volume of 400  $\mu$ L to record fluorescence spectra. Control experiments with the other non-specific proteins were carried out under otherwise identical conditions.

**Methods.** All fluorescence spectra were recorded in quartz cuvettes with an optical path length of 1.0 cm and at an excitation wavelength of 387 nm on a Shimadzu RF-5301 spectrofluorometer equipped with a Xenon lamp excitation source. UV-vis absorption spectra were recorded on a Shimadzu 3600 spectrophotometer. Data of dynamic light scattering (DLS) and zeta potential were obtained on a Brookhaven ZetaPALS Zeta Potential Analyzer upon excitation at 659 nm. Transmission electron microscopy (TEM) images were recorded on a JEOL 2010 transmission electron microscope at an accelerating voltage of 100 kV.

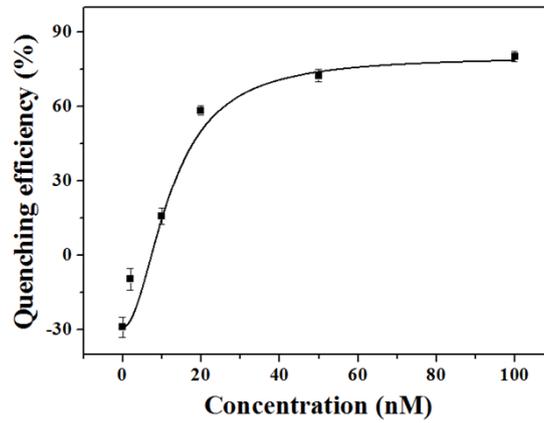
## FIGURES



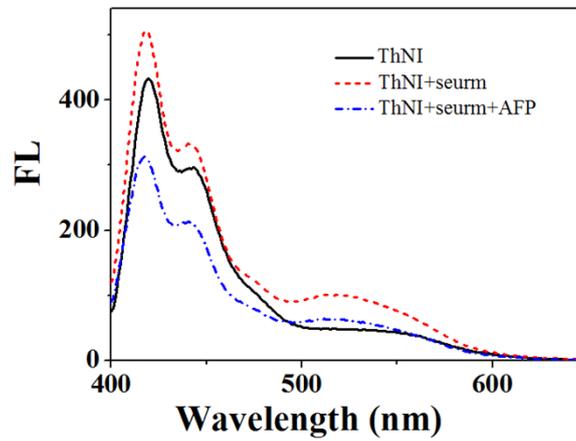
**Scheme S1** Synthesis route of conjugated polyelectrolyte brush ThNI (Z. Zhang, Degree Thesis, Nanjing University of Posts and Telecommunications, June, 2012.)



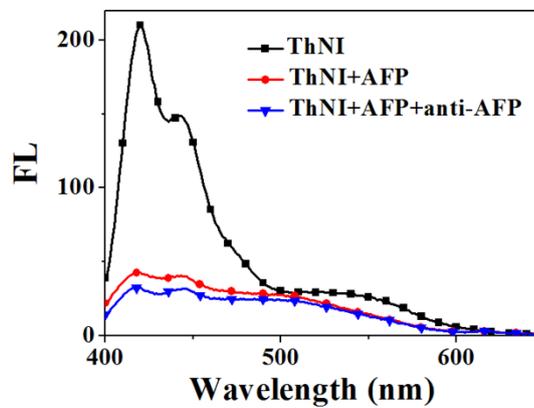
**Figure S1.** Quenching efficiency of the mixture (mixed solution of BSA, IgG, Lys, PSA and Thro, 20 nM each) without or with AFP (20 nM).



**Figure S2.** Quenching efficiency of AFP at different concentrations (0, 2, 10, 20, 50, 100 nM) in the mixture of several proteins.



**Figure S3.** Fluorescence response of ThNI to serum and AFP (20 nM) in serum.



**Figure S4.** Fluorescence spectra of ThNI upon addition of AFP and anti-AFP

## TABLES

**Table S1.** Comparison of performance of several major protein assays based on fluorescence quenching of CPs.

CP	protein	$K_{sv}$ ( $M^{-1}$ )	Specificity	Reference
Conjugated polyelectrolyte brush (ThNI)	AFP	$2.44 \times 10^8$		
	Lys, BSA, IgG, PSA and Thro	$-3\%$ to $-30\%$ quenching	Excellent	Our
Sulfonated PPV	Cyt <i>c</i>	$3.2 \times 10^8$		
	myoglobin lysozyme	$10^6$ 50% quenching	good	[16a]
$\alpha$ -mannose-bearing PPP	Con A	$4.5 \times 10^7$	Good	[16c]
	Cyt <i>c</i>	$2.27 \times 10^8$		
Sulfonated PF	hemin	$5.31 \times 10^7$	medium	[16g]
	methemoglobin	$3.81 \times 10^7$		
	histone	$2.8 \times 10^7$		
	hemoglobin	$1.3 \times 10^6$		
Carboxylate-substituted PPE	myoglobin	$6.9 \times 10^5$	bad	[16b]
	Cyt <i>c</i>	$6.5 \times 10^5$		
	lysozyme	$2.2 \times 10^5$		
	BSA	$-4.3 \times 10^6$		

**Table S2.** Zeta potential values and DLS data of ThNI with or without proteins.

Parameter	ThNI	ThNI+AFP	ThNI+control protein				
			Lys	PSA	Thro	BSA	IgG
$\xi$ (mV)	+ 48.65	- 8.52	+ 56.48	+ 50.00	+ 16.90	+ 28.55	+ 27.99
DLS (nm)	38.2	57.5	67.1	104.4	105.7	122.3	64.8