

## SUPPLEMENTARY INFORMATION

# Enhanced determination of As-phytochelatin complexes in *Chlorella vulgaris* using focused sonication for extraction of water-soluble species

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### Methods:

#### As-GS/PC Extraction

Cells were counted and a known amount of cells transferred. Cells were harvested by centrifugation (5 min, 3000 g) washed with deionised water twice and transferred to 15 mL centrifuge tubes. Cells were then washed with desorption solution (1 mM K<sub>2</sub>HPO<sub>4</sub>, 5 mM MES, 0.5 mM Ca(NO<sub>3</sub>)<sub>2</sub>, pH 6) for 10 min (Sandau, Pulz and Zimmermann, 1996; Abedin, Feldmann and Meharg, 2002). The supernatant was discarded and pelleted cells extracted with 2 mL of 1% formic acid, the mixture was sonicated for 30 s using a Minidelta 8935 generator (FFR ultrasonics, 500 W, 35 kHz) fitted with a 3 mm titanium micro-tip. The micro-tip was rinsed with methanol, then sonicated in 1% formic acid for 10 s between each extraction to avoid cross-contamination. The volume of the extract was determined gravimetrically.

#### Total arsenic quantification

Total arsenic was quantified using an ICP-MS (X series II, Thermo Scientific, UK) in CCT (Collision Cell Technology) mode with He/H as collision cell gas using 20 µg L<sup>-1</sup> gallium as an internal standard. The instrument was tuned daily using a 10 µg L<sup>-1</sup> indium, cerium, cobalt, uranium and lithium solution and the software (Plasmalab) was provided from the manufacturer.

#### Quantitative speciation analysis

Samples were run immediately after extraction using HPLC-ICP-MS. Separation was achieved in a Discovery C<sub>18</sub> column (15 x 2.1 mm) fitted with a pre-column Discovery C<sub>18</sub>. Experimental/instrumental conditions were based on the previous work of (Bluemlein, Rabb and Feldmann, 2009). A gradient elution was used with 0.1% formic acid (eluent A) and 99.9% HPLC grade methanol (eluent B). The detailed elution profile was as follows: 0-20 min linear increase 0-20% B, 20-30 min 20% B, 30-32 min 20-0% B and 32-40 min 0% B. The flow rate was 0.2 mL min<sup>-1</sup>.

The following parameters were adjusted for O<sub>2</sub> CCT in the ICP-MS according to the manufacturer's instructions: O<sub>2</sub> cell gas flow 0.6-1.45 mL min<sup>-1</sup>, hexapole bias -9 V, quadrupole bias -14 V, focus voltage -2 V, D2 voltage -100 V. A post column make-up flow was achieved with a tee connector and 0.9 mL min<sup>-1</sup> indium (20 µg L<sup>-1</sup> in 2% HNO<sub>3</sub>) as internal standard. The injection volume was 50 µL, the

monitored masses were: As, m/z 91 ( $[{}^{75}\text{As}{}^{16}\text{O}^+]$ ), S, m/z 48 ( $[{}^{32}\text{S}{}^{16}\text{O}^+]$ ) and In, m/z 115. Fresh external standards were prepared for quantification. For arsenic, different solutions of DMA were used and L-cysteine for the quantification of sulphur.

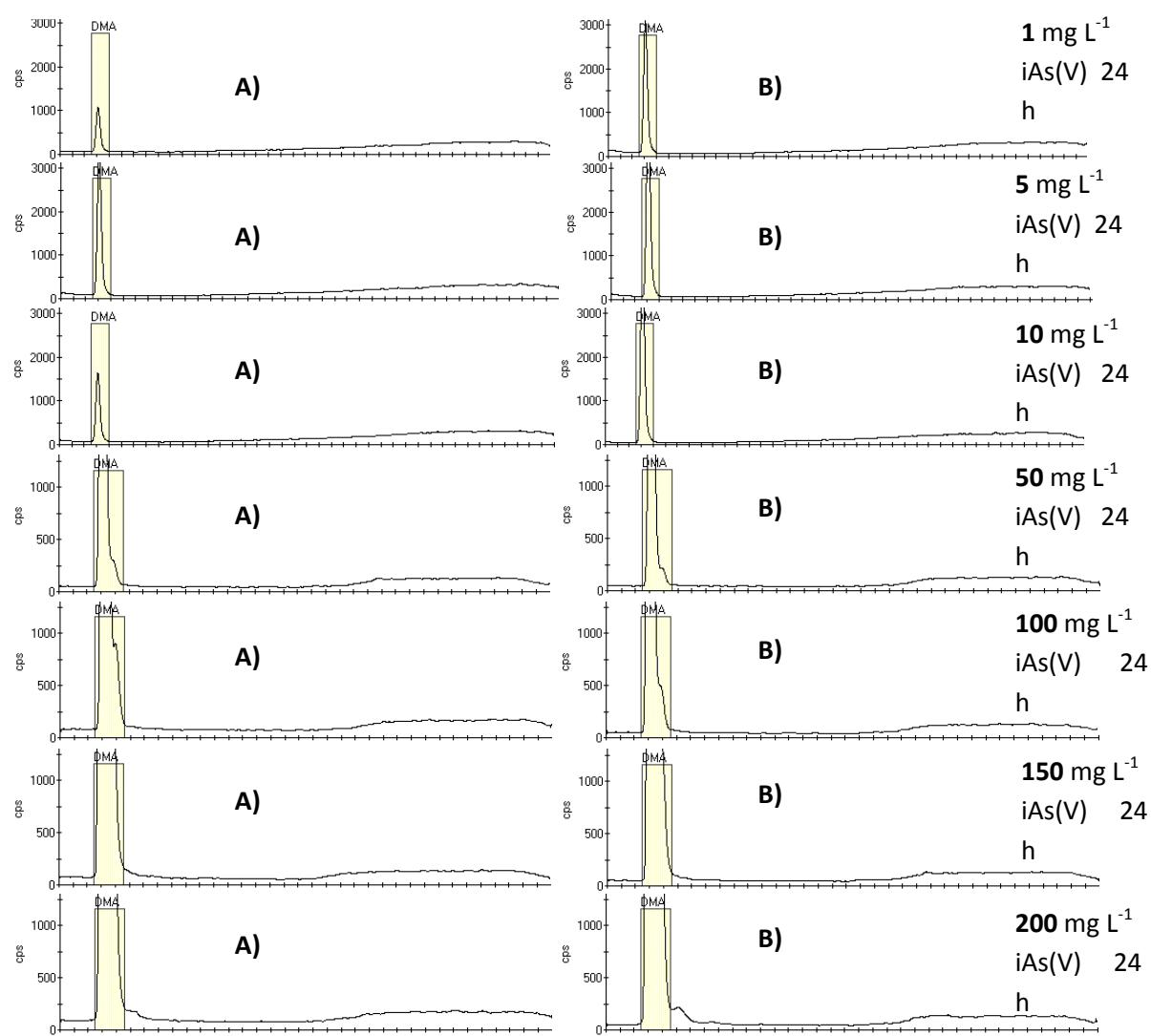
Method quantification limits (MQL) were calculated by multiplying 10 times the standard deviation signal (at the retention time of the quantification species) of 7 consecutive blank samples. Method detection limits (MDL) were calculated by 3.14 times the standard deviation signal of 7 consecutive blank samples to achieve 99% confidence intervals (CI) (at the retention time of the quantification specie).

A correction for methanol content in the mobile phase was performed as follows (Amayo et al., 2011): A blank was injected through the same chromatographic conditions. A post column addition of a solution containing 100 mg L<sup>-1</sup> DMA and 20 µg L<sup>-1</sup> indium (as internal standard) was made. The blank was analysed by ICP-MS for arsenic, sulphur and indium at m/z 91 ( $[{}^{75}\text{As}{}^{16}\text{O}^+]$ ), S, m/z 48 ( $[{}^{32}\text{S}{}^{16}\text{O}^+]$ ) and In, m/z 115.

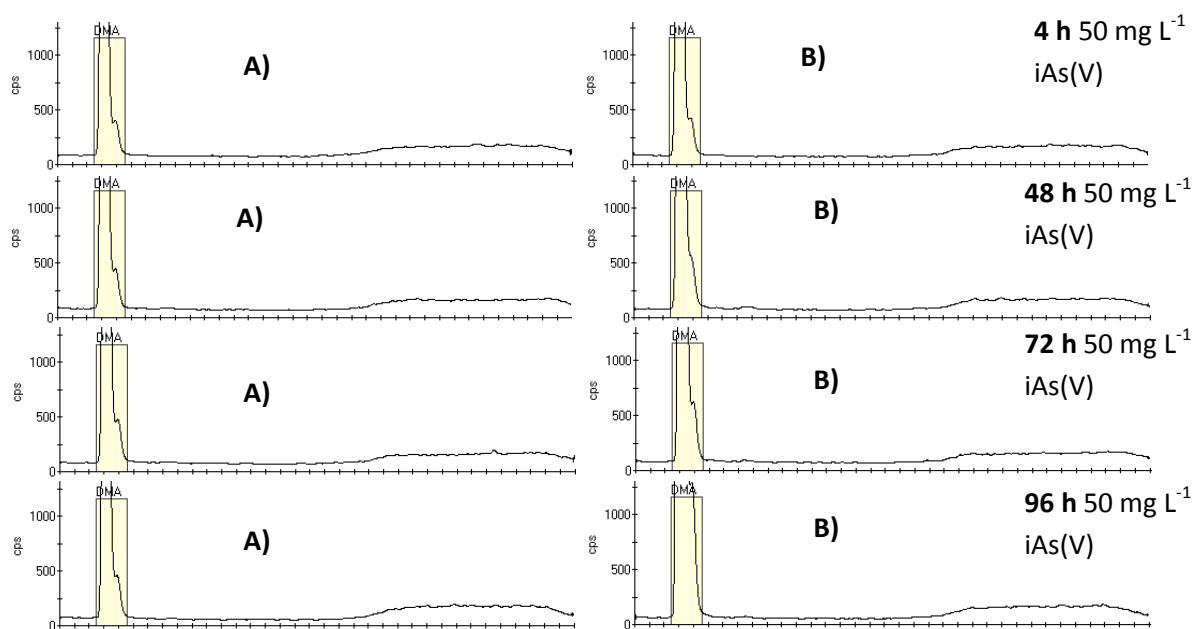
### Kinetics and concentration effect experiments

*C. vulgaris* cells were cultured for 3-5 days in Bold's media free of arsenic. The cells were then exposed to different concentrations (0-200 mg L<sup>-1</sup>) of iAs(III), iAs(V) and DMA for 48 h. The concentrations of arsenic used in this study were chosen to elicit a response in *C. vulgaris* cells rather than to reflect natural environment conditions. In other experiments cells were exposed to 50 mg.L<sup>-1</sup> of iAs(III), iAs(V) or DMA at different exposure times (4, 24, 48 and 72 h). The experiments were performed in triplicates. Cells were supplemented with 0.5% dextrose. GSH, PCs and As-GS/PC complexes were analysed.

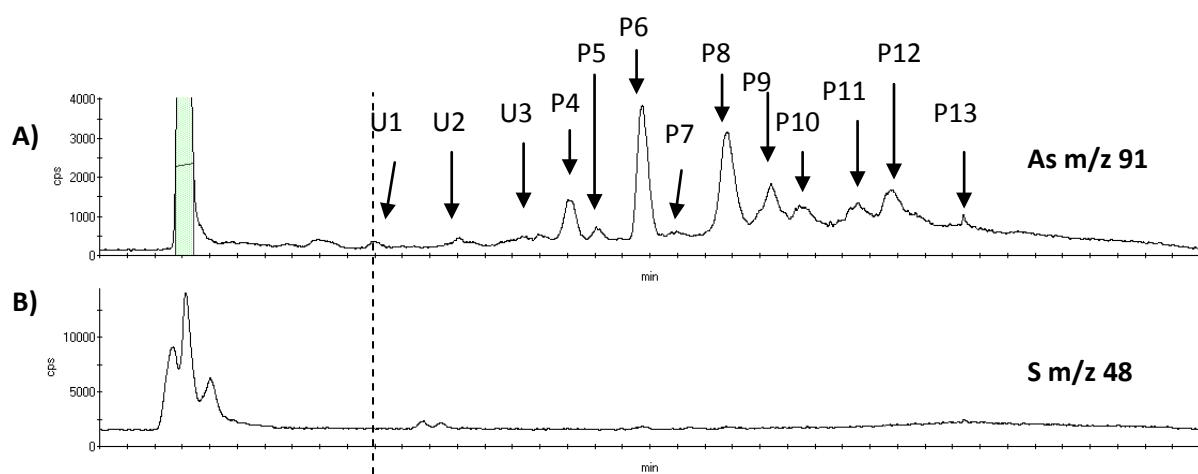
### iAs(V) exposure experiments results



**Fig S1** *C. vulgaris* cells exposed to 1, 5, 10, 50, 100, 150 and 200 mg L<sup>-1</sup> of iAs(V) for 24 h, analysed by HPLC-ICP-MS, two replicates (A and B)



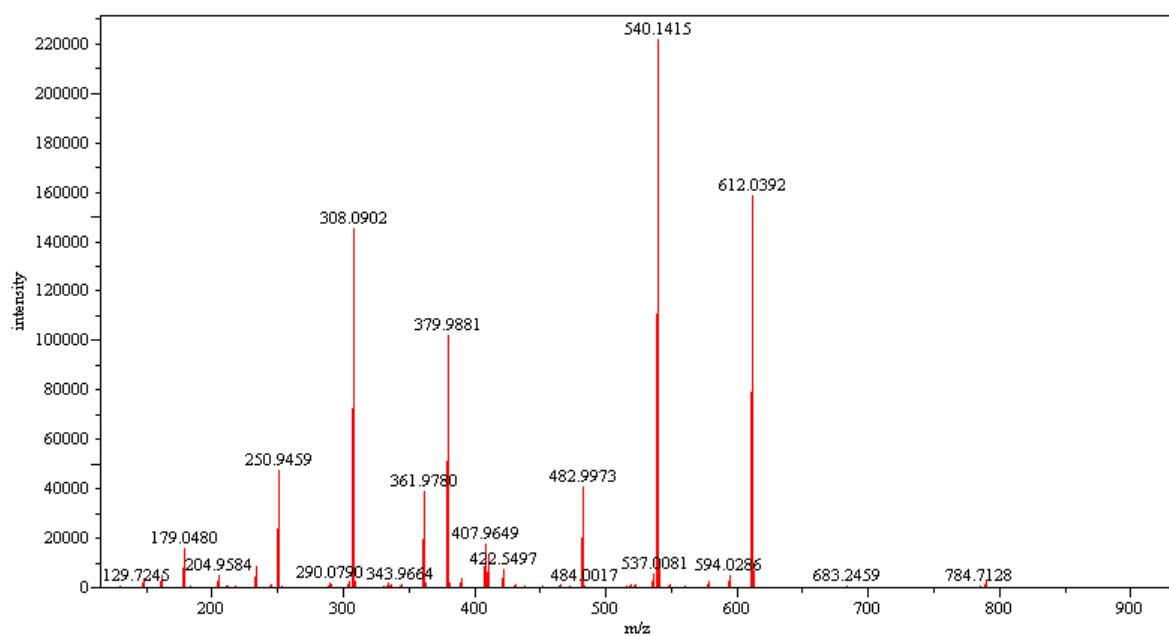
**Fig S2** *C. vulgaris* cells exposed to  $50 \text{ mg L}^{-1}$  of iAs(V) for 4, 48, 72 and 96 h, analysed by HPLC-ICP-MS, two replicates (A and B)



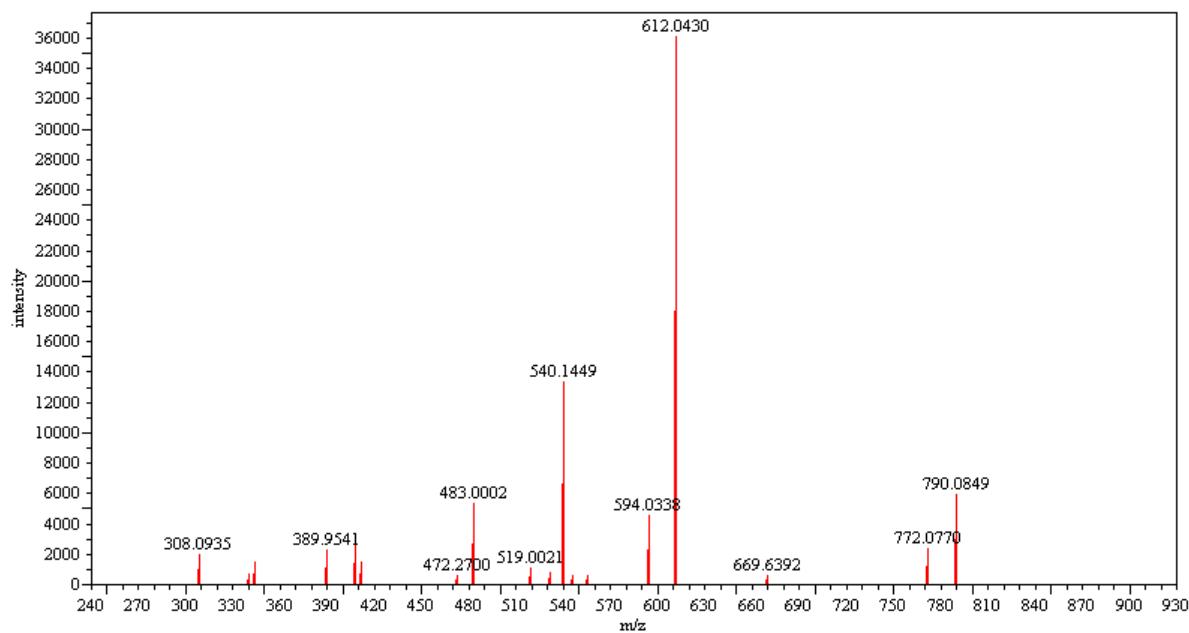
**Fig S3** Peak identification chromatogram of *C. vulgaris* cells exposed to  $50 \text{ mg L}^{-1}$  iAs(III) for 48 h, extracted in 1% formic acid 30 s focused sonication. A) signal for arsenic at  $m/z$  91 B) signal for sulphur at  $m/z$  48. Peak identification: **U1-3** Unknowns, **P4** GS-As(III)-PC<sub>2</sub>/GS-As(III)-γ-(Glu-Cys)<sub>2</sub>, **P5** As(III)-γ-(Glu-Cys)<sub>2</sub>, **P6** GS-As(III)-PC<sub>2</sub>, **P7** GS-As(III)-γ-(Glu-Cys)<sub>2</sub>-Ala, **P8** As(III)-PC<sub>3</sub>/ MMA(III)-PC<sub>2</sub>, **P9** MMA(III)-PC<sub>2</sub>, **P10** As(III)-PC<sub>3</sub>/ As(III)-(PC<sub>2</sub>)<sub>2</sub>, **P11** As(III)-(PC<sub>2</sub>)<sub>2</sub>/ As(III)-γ-(Glu-Cys)<sub>3</sub>-Ala/ As(III)-γ-((Glu-Cys)<sub>2</sub>)<sub>2</sub>-Ala/ MMA(III)-γ-(Glu-Cys)<sub>2</sub>-Ala, **P12** As(III)-PC<sub>4</sub> and **P13** As(III)-PC<sub>4</sub>.

## MS/MS analysis

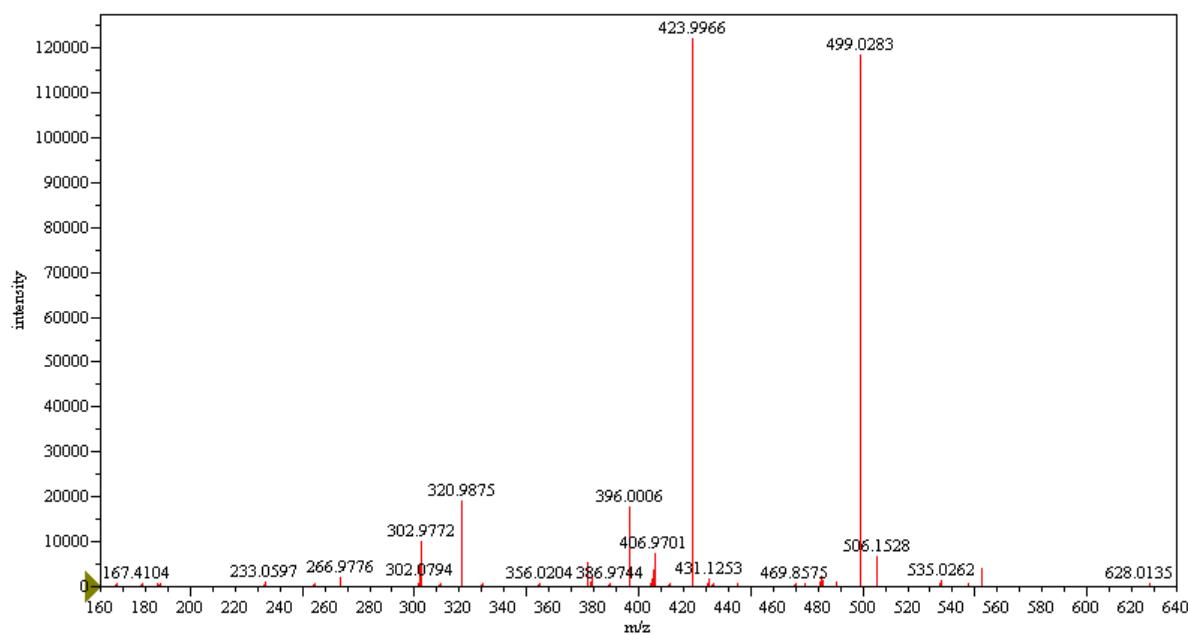
### GSH/PC molecules:



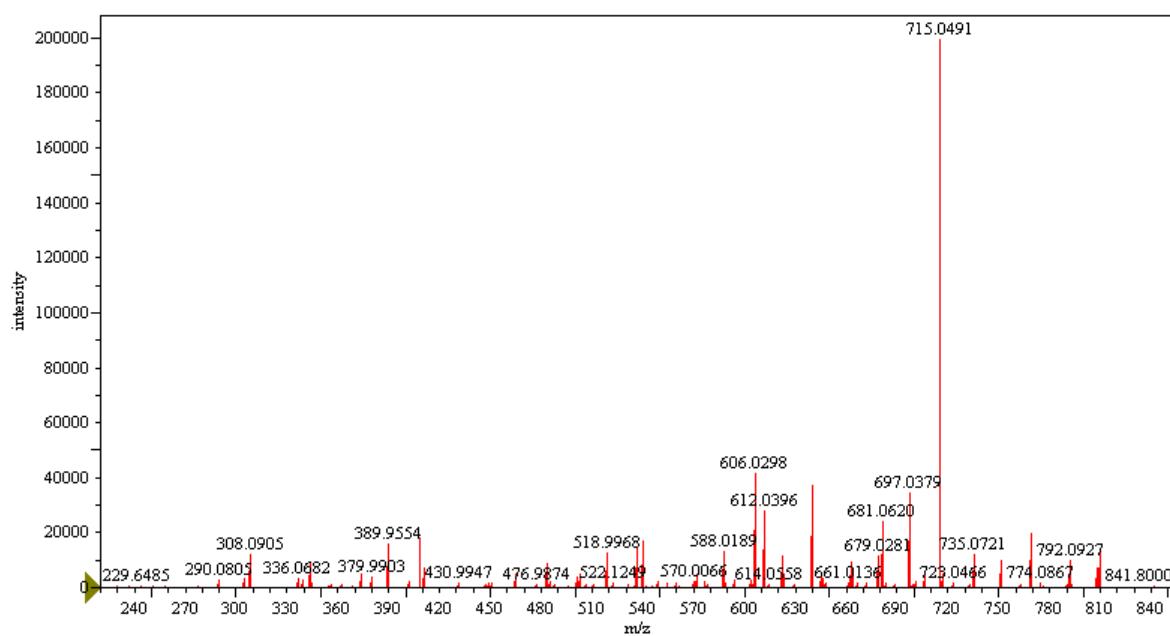
**Fig S4** MS/MS spectra for GS-As(III)-PC<sub>2</sub> m/z 460.0663 RT:9.88 min, *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



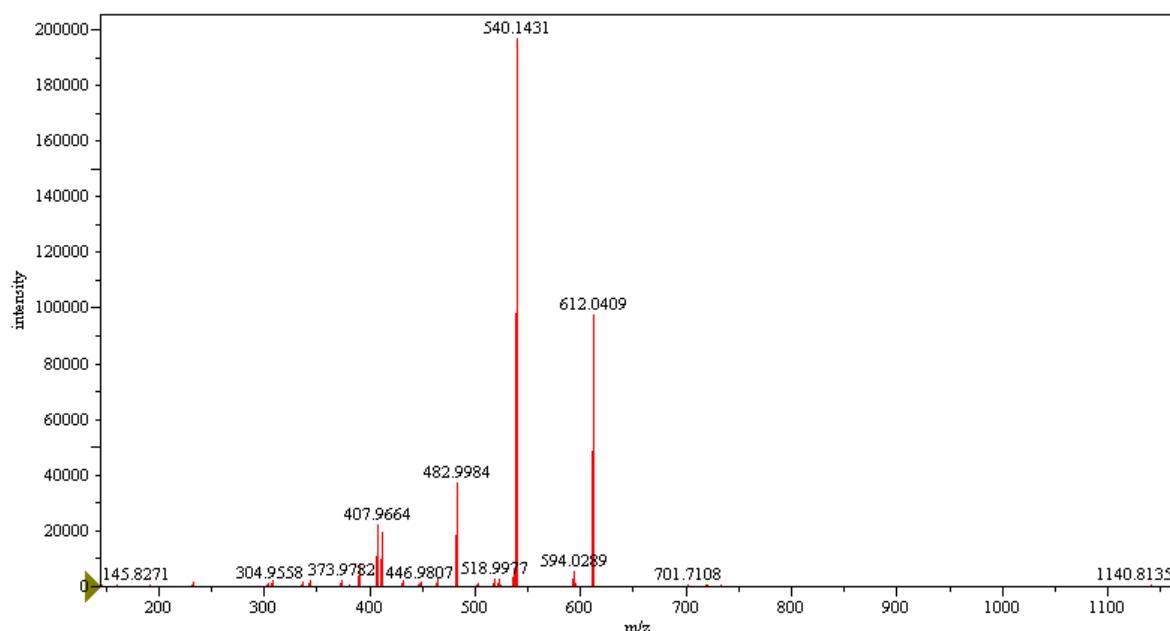
**Fig S5** MS/MS spectra for GS-As(III)-PC<sub>2</sub> m/z 919.1247 RT:9.92 min, *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



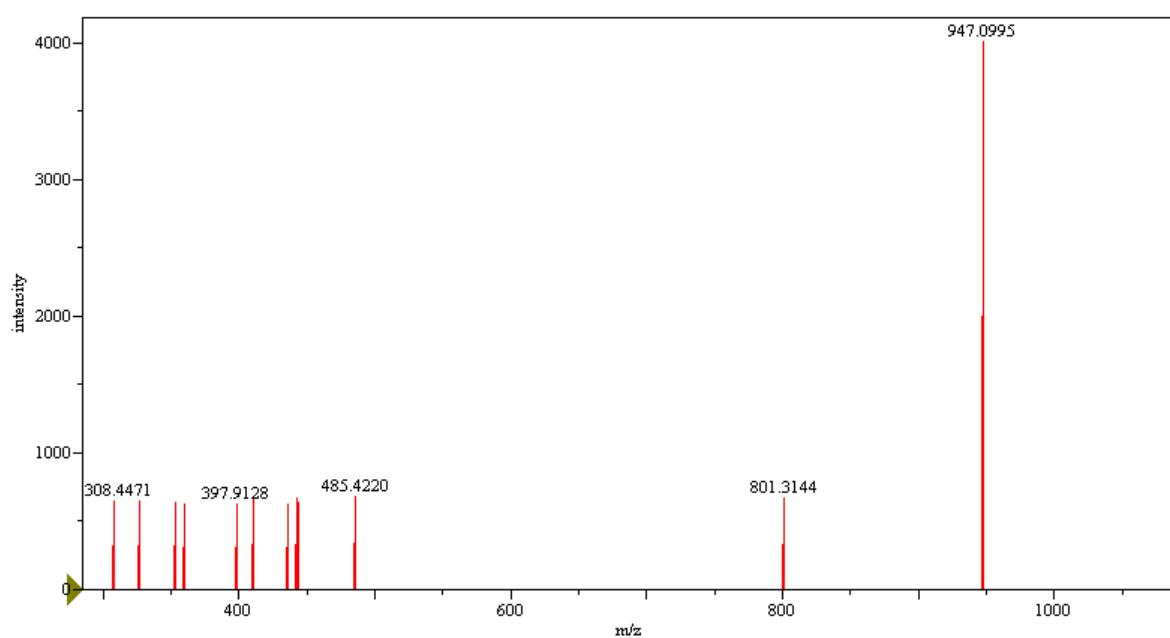
**Fig S6** MS/MS spectra for MMA(III)-PC<sub>2</sub> m/z 628.0729 RT:11.31 min , *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



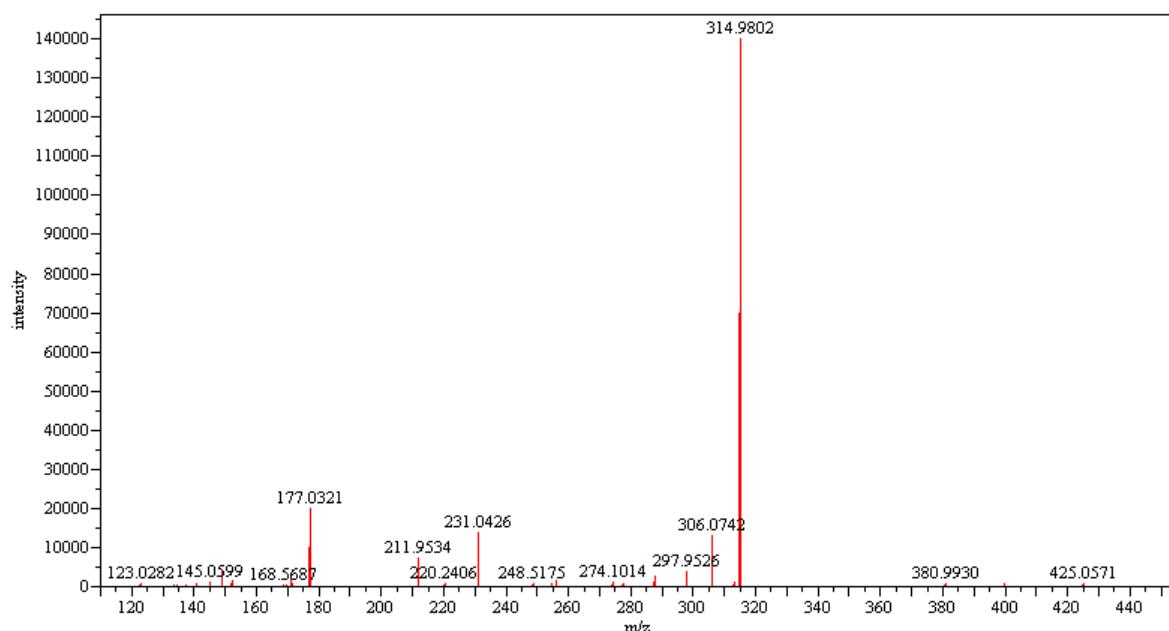
**Fig S7** MS/MS spectra for As(III)-PC<sub>3</sub> m/z 844.0931 RT:11.49 min , *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



**Fig S8** MS/MS spectra for As(III)-(PC<sub>2</sub>)<sub>2</sub> m/z 576.0923 RT:13.07 min , *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)

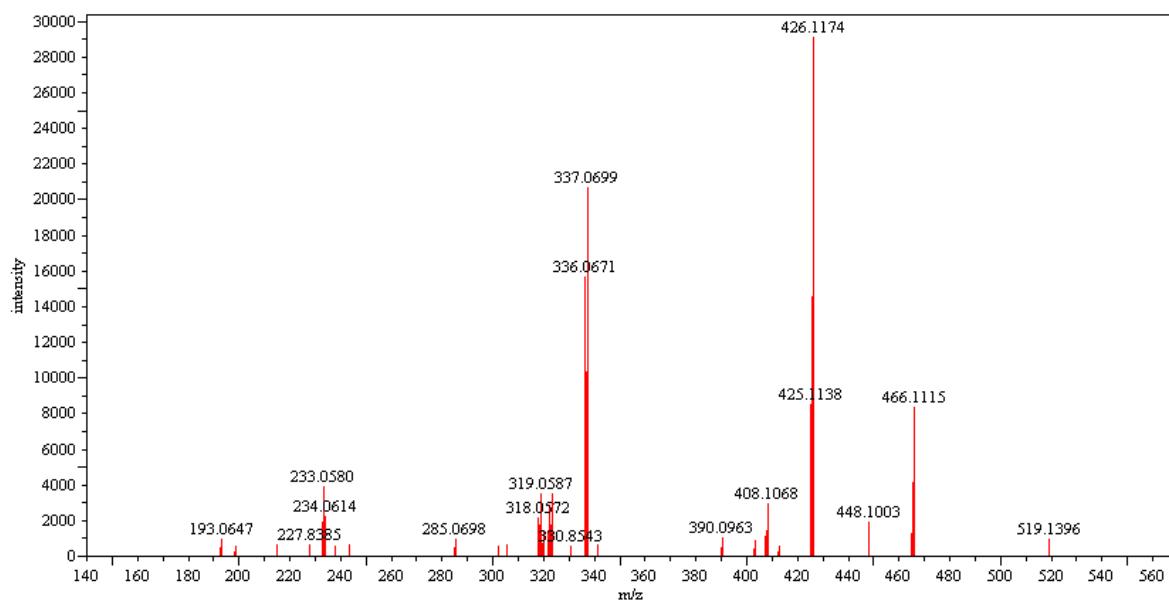


**Fig S9** MS/MS spectra for As(III)-PC<sub>4</sub> m/z 1076.1455 RT:15.02 min , *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)

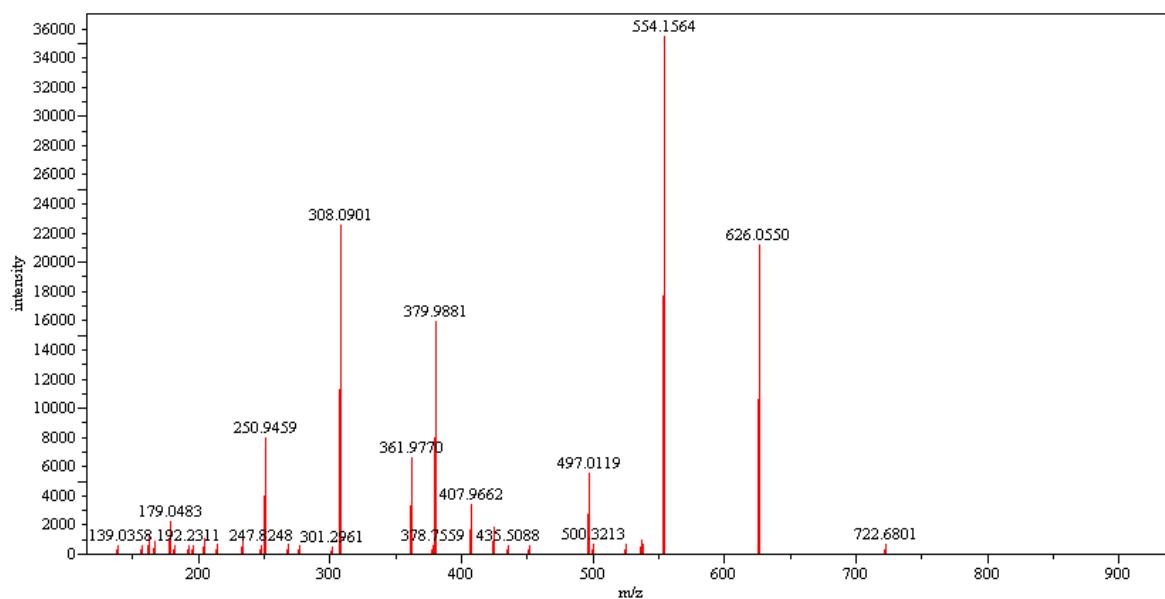


**Fig S10** MS/MS spectra for DMAS<sup>V</sup>-GS m/z 444.0247 RT 4.37 min , *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)

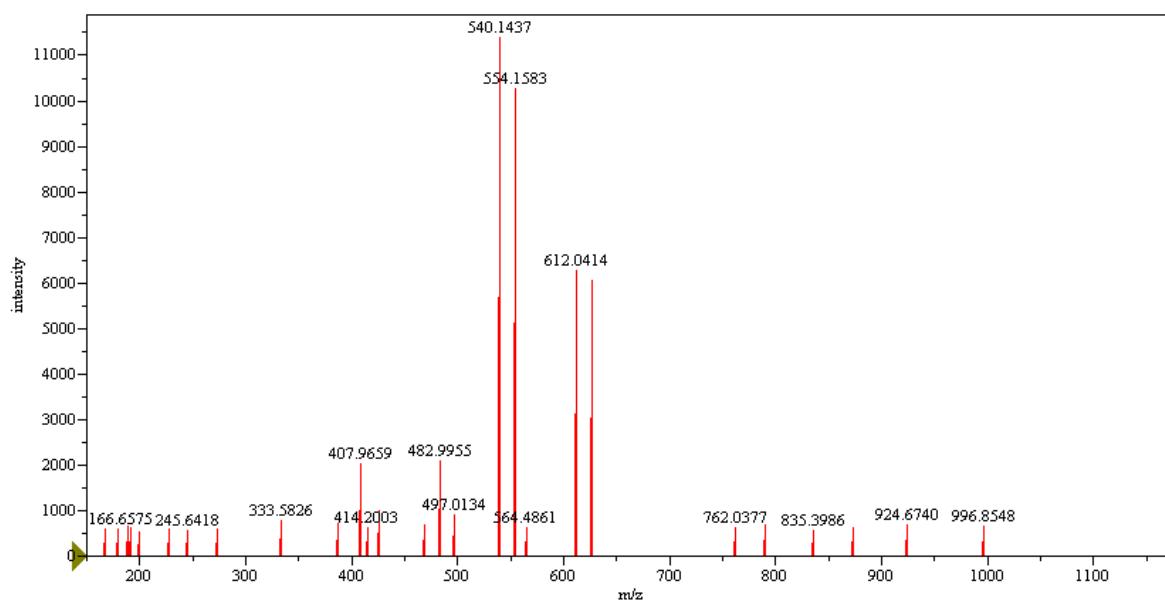
### hGSH/PC molecules:



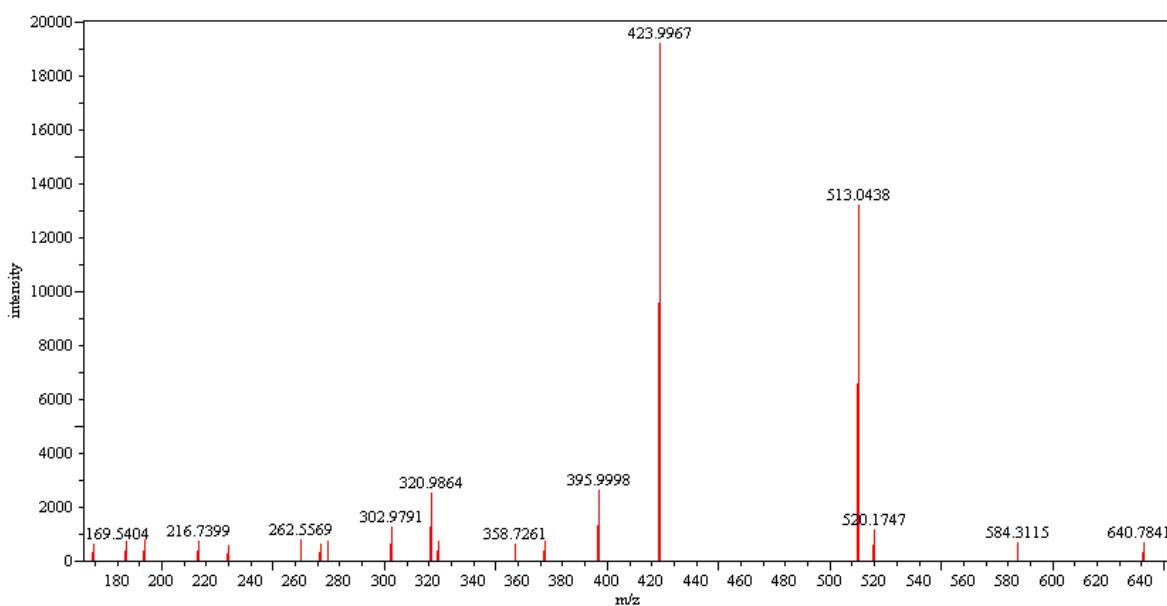
**Fig S11** MS/MS spectra for  $\gamma$ -(Glu-Cys)<sub>2</sub>-Ala m/z 554 RT 6.2845 min *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



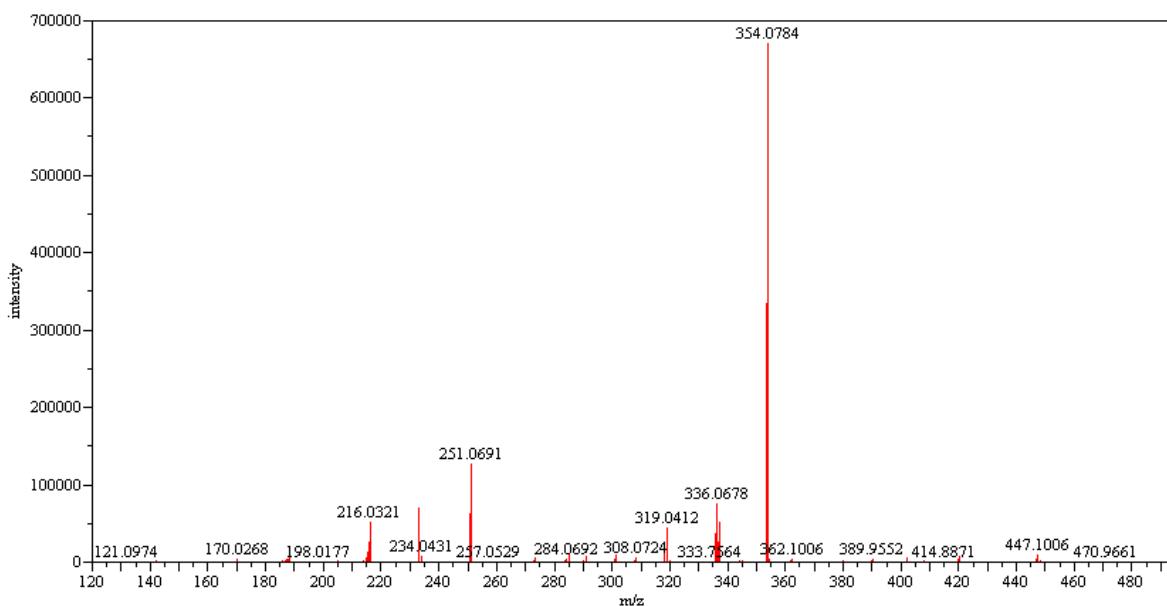
**Fig S12** MS/MS spectra for As(III)- $\gamma$ -(Glu-Cys)<sub>3</sub>-Ala m/z 858 RT 10.9082 min *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



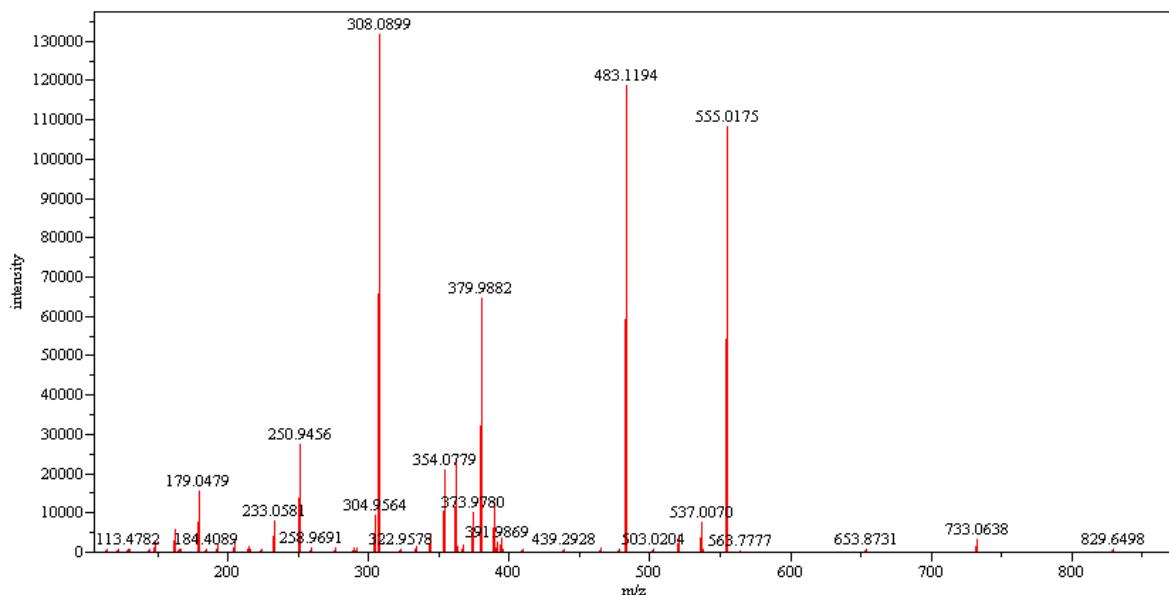
**Fig S13** MS/MS spectra for As(III)- $\gamma$ -((Glu-Cys)<sub>2</sub>)<sub>2</sub>-Ala m/z 583 RT 14.6667 min *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



**Fig S14** MS/MS spectra for MMA(III)- $\gamma$ -(Glu-Cys)<sub>2</sub>-Ala  $m/z$  642 RT 14.8483 min *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



**Fig S15** MS/MS spectra for  $\gamma$ -(Glu-Cys)<sub>2</sub>  $m/z$  483 RT 4.6997 min *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)



**Fig S16** MS/MS spectra for mins GS-As(III)- $\gamma$ -(Glu-Cys)<sub>2</sub> m/z 431 RT 8.0730 *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL)

**Table S1** Summary of fragments found in MS/MS analysis *C. vulgaris* cells exposed to 50 mg L<sup>-1</sup> iAs(III), analysed using ES-MS/MS (Orbitrap Discovery LTQ-XL) (n.o.- not observed,T- Theoretical, F - Found)

Molecule	m/z	b1	y8	b2	y7	b3	y6	b4	y5	b5	y4	b6	y3	b7	y2	b8	y1	
<b>Phytochelatins</b>																		
PC <sub>2</sub>	540	T	130.1	----	233.1	----	362.1	----	465.1	----	----	411.1	----	308.1	----	179.1	----	76.0
		F	130.1	----	233.1	----	362.1	----	465.1	----	----	411.1	----	308.1	----	179.1	----	76.0
PC <sub>3</sub>	772	T	130.1	----	233.1	----	362.1	643.2	465.1	540.1	594.2	411.1	697.2	308.1	----	179.1	----	76.0
		F	130.1	----	233.1	----	362.1	643.2	465.1	540.1	n.o.	411.1	697.2	309.1	----	n.o.	----	76.0
As(III)-PC <sub>3</sub>	844	T	130.1	----	233.1	----	362.1	643.2	465.1	540.1	594.2	411.1	697.2	308.1	----	179.1	----	76.0
		F	n.o.	----	n.o.	----	362.0	n.o.	465.1	540.1	594.0	411.1	697.0	308.1	----	n.o.	----	n.o.
GS-As(III)-PC <sub>2</sub>	460	T	130.1	----	233.1	----	362.1	----	465.1	540.1	----	411.1	----	308.1	----	179.1	----	76.0
		F	129.7	----	233.1	----	362.0	----	n.o.	540.1	----	422.1	----	308.1	----	179.0	----	n.o.
As(III)-(PC <sub>2</sub> ) <sub>2</sub>	576	T	130.1	----	233.1	----	362.1	----	465.1	540.1	594.2	411.1	----	308.1	----	179.1	----	76.0
		F	n.o.	----	233.1	----	n.o.	----	465.1	540.1	594.0	411.1	----	308.1	----	n.o.	----	n.o.
As(III)-PC <sub>4</sub>	1076	T	130.1	875.1	233.1	772.2	362.1	643.2	465.1	540.1	594.2	411.1	697.2	308.1	826.2	179.1	929.2	76.0
		F	n.o.	411.1	n.o.	308.4	n.o.	n.o.	n.o.	n.o.								
MMA(III)-PC <sub>2</sub>	628	T	130.1	----	233.1	----	362.1	----	465.1	----	----	411.1	----	308.1	----	179.1	----	76.0
		F	n.o.	----	233.1	----	n.o.	----	n.o.	----	----	n.o.	----	n.o.	----	178.8	----	n.o.
DMAS <sup>V</sup> -GS	444	T	136.9	----	231.1	----	----	----	----	----	----	----	----	----	----	176.9	----	----
		F	136.9	----	231.1	----	----	----	----	----	----	----	----	----	----	176.9	----	----
<b>Ala GSH/PC</b>																		
γ-(Glu-Cys) <sub>2</sub> Ala	554	T	130.1	----	233.1	----	362.1	----	465.1	----	----	425.1	----	322.1	----	193.1	----	90.1
		F	130.1	----	233.1	----	362.1	----	465.1	----	----	425.1	----	322.1	----	193.1	----	90.1
GS-As(III)-γ-(Glu-Cys) <sub>2</sub> -Ala	467	T	130.1	----	233.1	----	362.1	----	465.1	554.2	----	425.1	----	322.1	----	193.1	----	90.1
		F	n.o.	----	233.1	----	362.0	----	n.o.	554.2	----	425.1	----	n.o.	----	192.2	----	n.o.
As(III)-γ-((Glu-Cys) <sub>2</sub> ) <sub>2</sub> -Ala	583	T	130.1	----	233.1	----	362.1	----	465.1	554.2	----	425.1	----	322.1	----	193.1	----	90.1
		F	n.o.	----	n.o.	----	n.o.	----	n.o.	554.2	----	425.1	----	n.o.	----	n.o.	----	n.o.
<b>DesGLY GSH/PC</b>																		
γ-(Glu-Cys) <sub>2</sub>	483	T	130.1	----	233.1	----	362.1	----	----	----	----	----	----	354.1	----	251.1	----	122.0
		F	n.o.	----	233.1	----	362.1	----	----	----	----	----	----	354.1	----	251.1	----	121.2
GS-As(III)-γ-(Glu-Cys) <sub>2</sub>	431	T	130.1	----	233.1	----	362.1	----	380.0	483.1	----	----	----	354.1	----	251.1	----	122.0
		n.o.	----	233.1	----	362.0	----	380.0	483.1	----	----	----	354.1	----	250.9	----	122.1	