

A novel hydride generation spectral method for trace Se based on resonance Rayleigh scattering of Cu₂O-Se nanoparticles

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Experimental

Apparatus and reagents

A model of F-7000 fluorescence spectrophotometer (Japan), a model of F95S fluorescence spectrophotometer (Lengguang Technology Ltd, Shanghai, China), a model of TU-1901 double beam UV-Vis spectrophotometer (Purkinje General Instruments Ltd, Beijing, China), a model of JSM-6380 LV scanning electron microscopy (Japan), a model of SK8200LH ultrasonic reactor (Kedao Ultrasonic Instruments Ltd, Shanghai, China), a model of DTD-40 Constant temperature digestion apparatus (Posen electronic instrument factory, Changzhou, China), and a homemade HG-absorption device were used.

A 1.000 g/L selenium standard stock solution was prepared as following: a 0.1005 g of selenium powder was weighed (purity > 99.95%), and 1mL HNO₃ and a drop of HCl were added to in a 100 mL beaker, and heated to dissolve, and transferred to a 100 mL volumetric flask, and diluted to the mark with water. Selenium standard stock was stored in the refrigerator, and selenium working solution was obtained by diluting the stock solution with water. A 36 mg/mL NaBH₄ solution should be prepared before it is used. A 9.2 mol/L H₂SO₄, 1.23 mol/L potassium sodium tartrate (KNaC₄H₄O₆) including 6.25 mol/L NaOH, 6 mol/L HCl, mixed acid of HNO₃ + HClO₄ (1+3), and 1.0 % potassium ferricyanide (K₃Fe(CN)₆) were used. CuSO₄ absorption solution (CuSO₄-KNaC₄H₄O₆, Fehling reagent): 1.0 mL 0.02 mol/L CuSO₄, 2.0 mL 1.23 mol/L KNaC₄H₄O₆ are mixed in the reagent bottle, and diluted to 50 mL with water. All reagents were of analytical grade and the water was doubly distilled.

Sample preparation

The tea samples were purchased from market, and placed in a vacuum drying oven drying (70 °C). A 1.00 g tea was weighed into a digestion tube, then added 10 mL mixed acid, and covered the funnel (refluxing), used ultrasonic treatment for 30 min [25], then put them in constant temperature digestion apparatus (170 °C, added mixed acid in time during digestion, when the solution became clear and colorless, and accompanied by white smoke, continued to heat until residual volume about 2 mL, then

quickly get off after cooling added 5 mL HCl (1:1), placed in a boiling water bath and heated 10 min, removed the tube digestions, cooled to room temperature, and filtered. The residue was washed with water doubly, the washed solution was merged into the volumetric flask, and added 1.0 mL 1% K₃Fe(CN)₆ and diluted to 10 mL with water for use.

Procedure

A 0.100 g/L Se standard solution was added into a reactor bottle, then added 1.0 mL 9.2 mol/L H₂SO₄, and diluted to 10 mL. 10 mL of 36 mg/mL NaBH₄ were added into the separating funnel, and 5.0 mL CuSO₄ absorption solution were added into the gas absorption bottle (Figure 1S). Opened the ultrasonic reactor, and set time for 5 min, then slowly opened the piston of the separating funnel, the NaBH₄ would be added in the reaction bottle, with flow rate of 5 mL/min. The reaction product of SeH₂ was exported to the absorption bottle. After the reaction was complete, the solution was transferred into a test tube and placed for 15 min, then the RRS spectrum was recorded on a fluorescence spectrophotometer, using a synchronous scanning technique ($\lambda_{\text{ex}} - \lambda_{\text{em}} = \Delta\lambda = 0$), voltage of 450 V, both excited slit and emission slit of 5 nm, and emission filter of 1% T attenuator. The RRS value at 374 nm ($I_{374\text{nm}}$) and a blank value without Se (IV) (I_0) were recorded. The value of $\Delta I_{374\text{nm}} = I_{374\text{nm}} - (I_{374\text{nm}})_0$ was calculated.

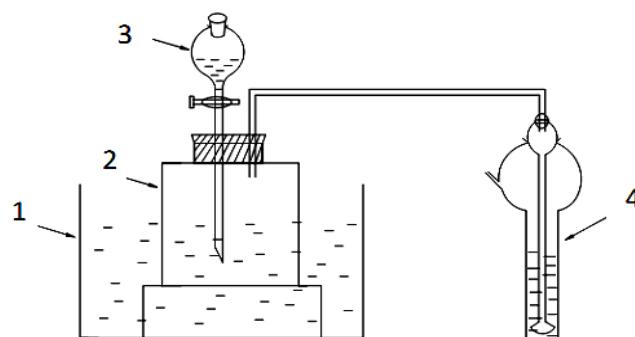


Figure 1S A scheme of the HQ equipment

1. Ultrasonic reactor; 2. Reaction bottle; 3. Separating funnel; 4. Gas absorption bottle

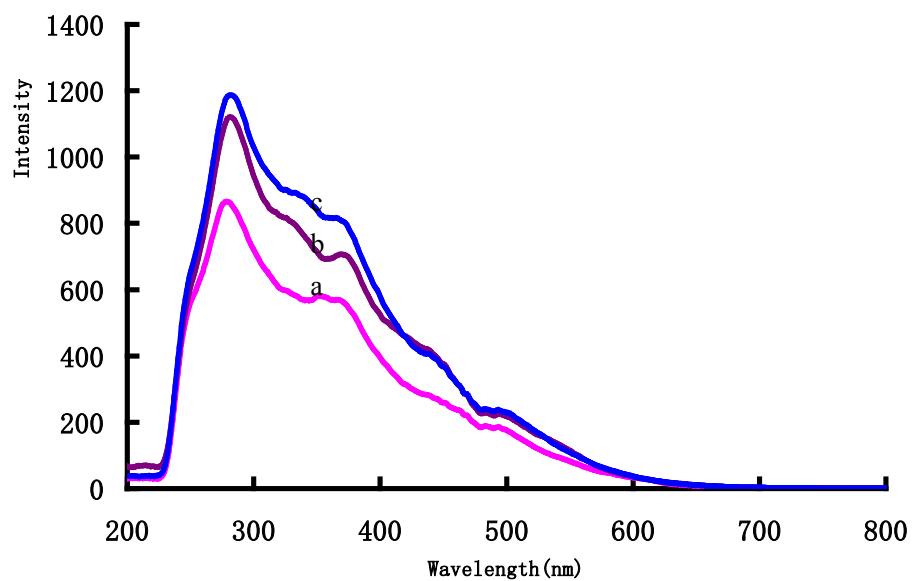


Figure 2S Resonance Rayleigh scattering spectra of the SeH_2 -tartrate system
a: 49.2 mmol/L tartrate-0.46 mol/L H_2SO_4 -36 mg/mL NaBH_4 ; b: a+0.5 mg/L Se; c: a+1.5 mg/L Se.

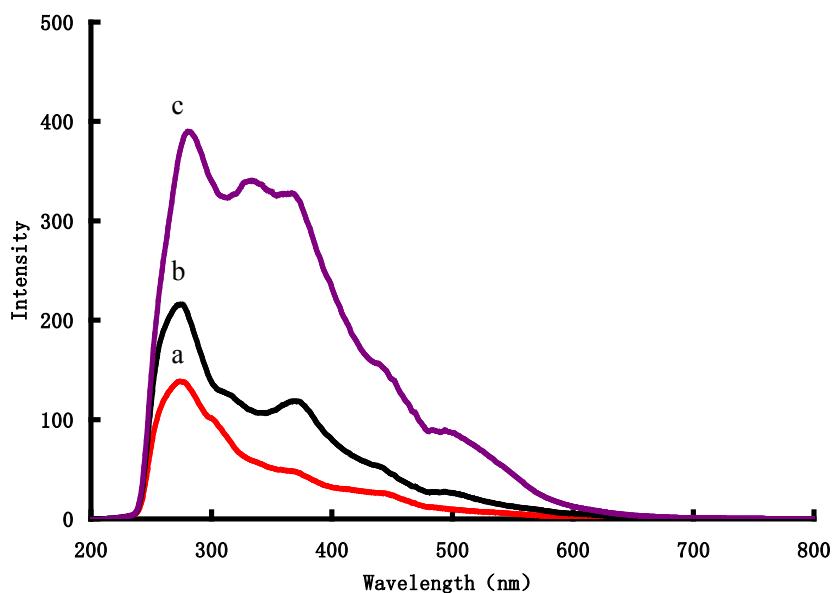


Figure 3S Resonance Rayleigh scattering spectra of the SeH_2 -Se(IV) system
a: 0.0 mg/L Se-0.46 mol/L H_2SO_4 -36 mg/mL NaBH_4 + absorption solution containing 0.06 g/L Se(IV); b: a+0.75 mg/L Se; c: a+4.5 mg/L Se.

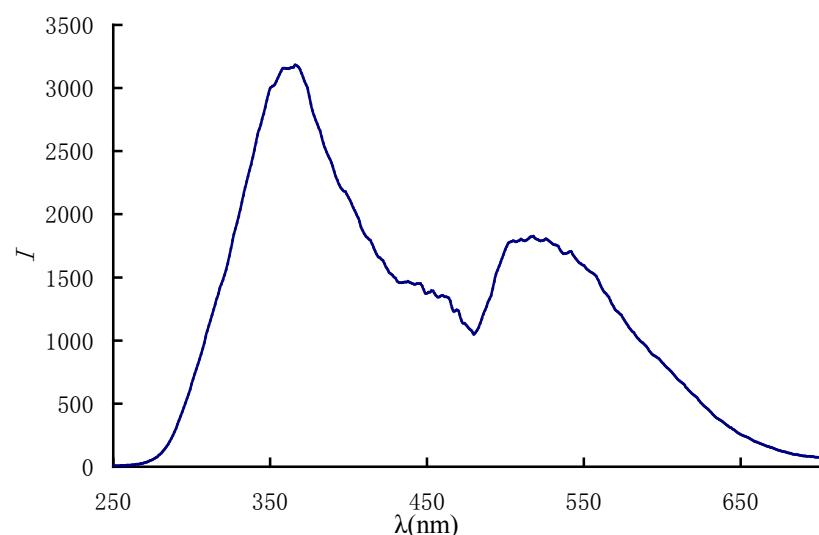


Figure 4S The Cu_2O particle RRS spectra in the glucose-Fehling reagent-Au nanoparticle catalytic system

2mmol/L CuSO_4 +61.5mmol/L $\text{KNaC}_4\text{H}_4\text{O}_6$ +0.37mmol/L glucose-16.2 ng/mL AuNP-70 °C 7 min.

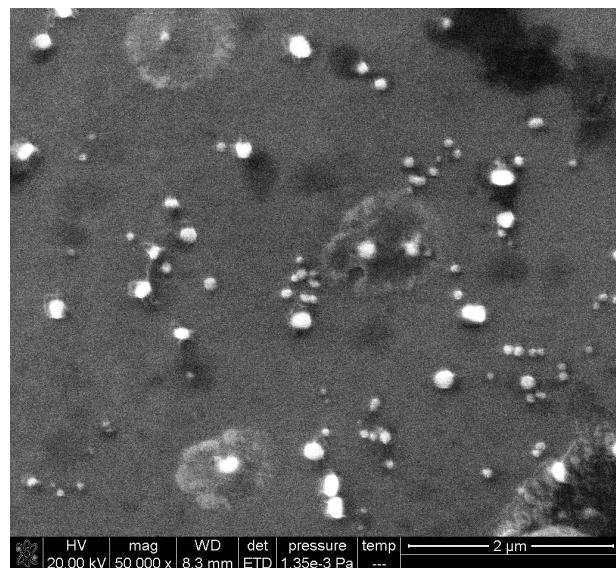


Figure 5S Scanning electron microscopy
1.5 mg/L Se-0.4 mmol/L CuSO_4 -0.0492 mol/L $\text{KNaC}_4\text{H}_4\text{O}_6$ -0.46 mol/L H_2SO_4 -36 mg/mL NaBH_4 .

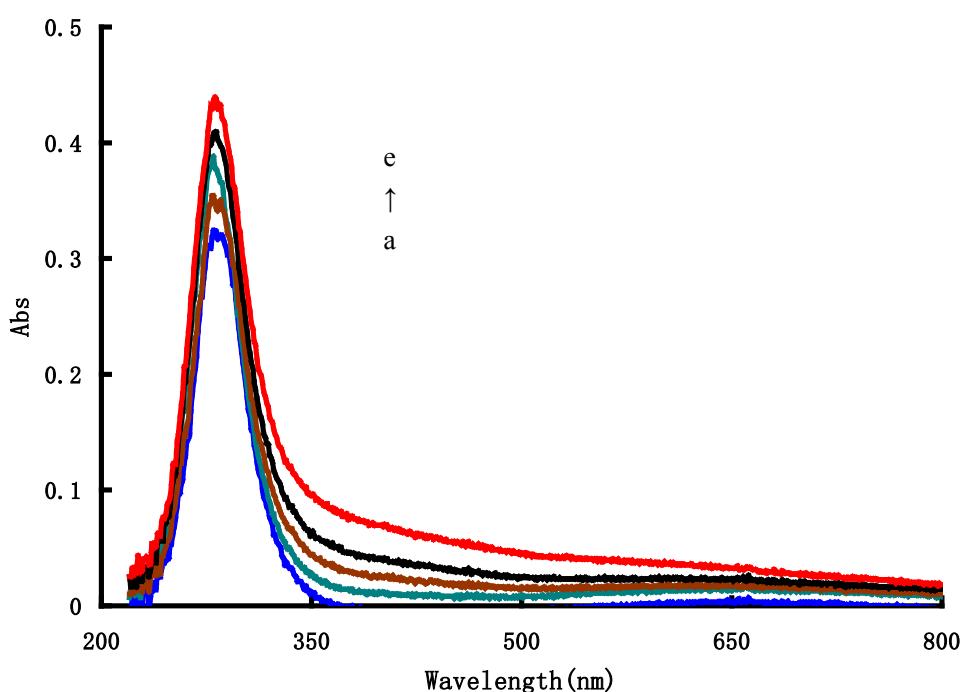


Figure 6S Absorption spectra of Cu₂O-Se particles system
a: 0.4 mmol/L CuSO₄ -0.0492 mol/L KNaC₄H₄O₆-0.46 mol/L H₂SO₄-36 mg/mL NaBH₄;
b: a+0.10 mg/L Se; c: a+0.50 mg/L Se;d: a+0.75 mg/L Se;e:a+1.50 mg/L Se

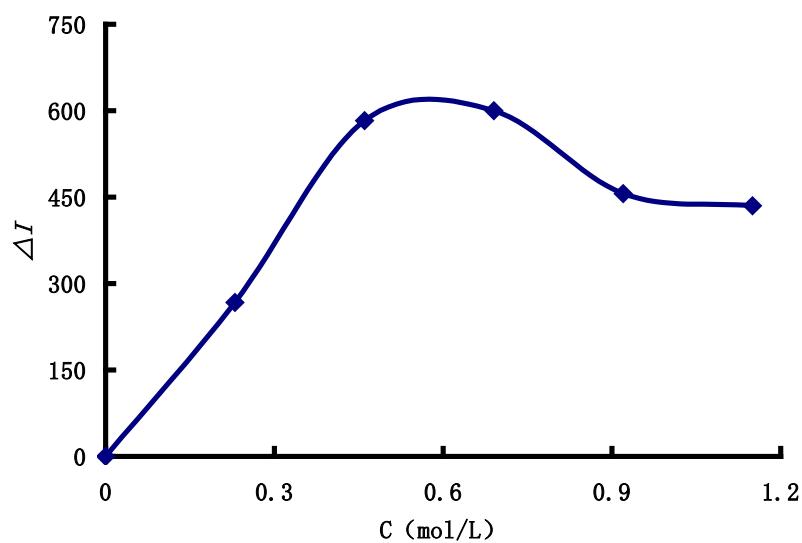


Figure 7S Effect of H₂SO₄ concentration on $\Delta I_{374\text{nm}}$
0.5 mg/L Se-30 mg/mL NaBH₄-0.4 mmol/L CuSO₄ -0.0492 mol/L KNaC₄H₄O₆

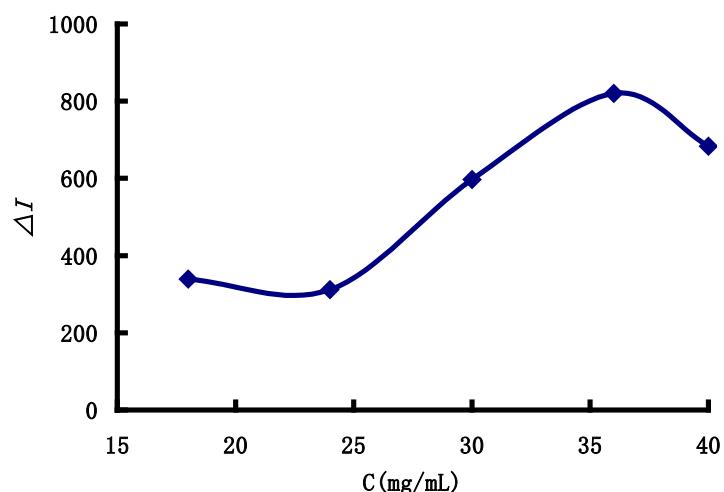


Figure 8S Effect of NaBH₄ concentration on $\Delta I_{374\text{nm}}$
0.5mg/L Se-0.46 mol/L H₂SO₄-0.4 mmol/L CuSO₄-0.0492 mol/L KNaC₄H₄O₆

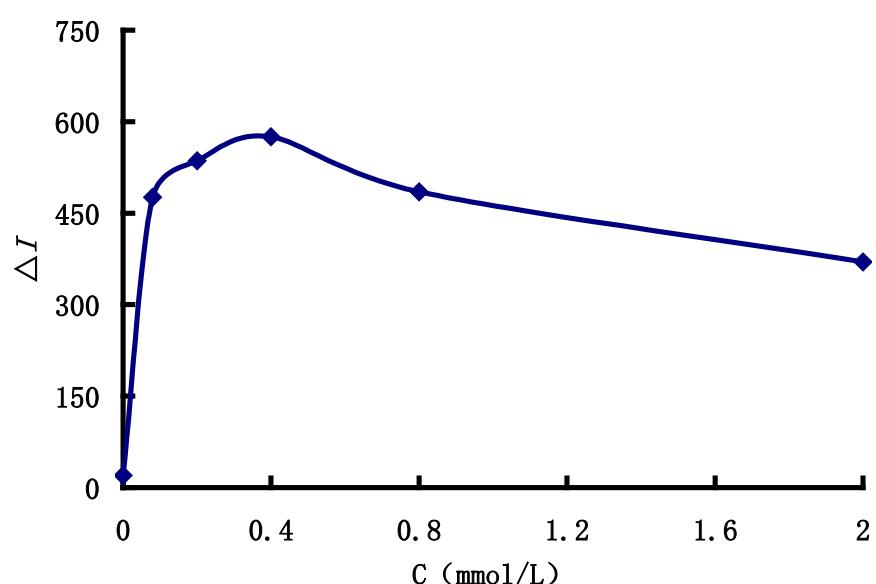


Figure 9S Effect of CuSO₄ concentration on $\Delta I_{374\text{nm}}$
0.5 mg/L Se-0.46 mol/L H₂SO₄-36 mg/mL NaBH₄-0.0738 mol/L KNaC₄H₄O₆

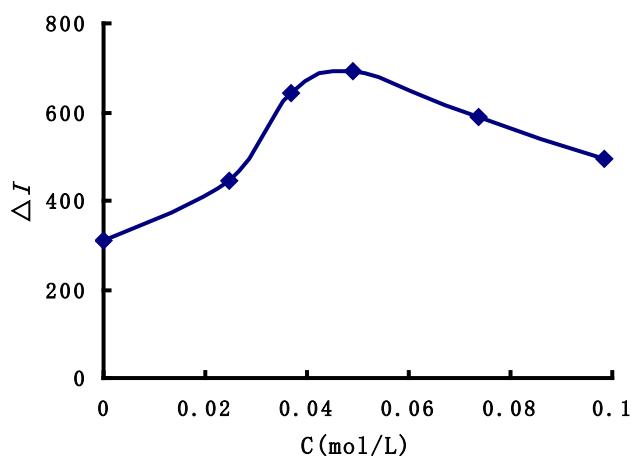


Figure 10S Effect of $\text{KNaC}_4\text{H}_4\text{O}_6$ concentration on $\Delta I_{374\text{nm}}$
 0.5 mg/L Se-0.46 mol/L H_2SO_4 -36 mg/mL NaBH_4 -0.4 mmol/L CuSO_4

Table 1S Effect of CES ^a

Coexistent substance	Tolerance ($\mu\text{mol/L}$)	Relative error (%)	Coexistent substance	Tolerance ($\mu\text{mol/L}$)	Relative error(%)
Fe^{3+}	174	9.4	Mg^{2+}	500	-3.2
Ca^{2+}	600	-7.4	Hg^{2+} ^b	100	2.4
As^{3+}	46	-3.8	Cu^{2+} ^b	150	-4.3
Te^{6+}	150	5.8	Pb^{2+}	150	-8.5
Sb^{3+}	200	-4.1	Bi^{3+}	20	-1.8
Ge^{4+}	50	4.9	Tl^{3+}	45	2.8
Sn^{4+}	45	3.2	Zn^{2+} ^b	100	-4.0
In^{3+}	80	4.1	Cd^{2+}	80	4.7
Ni^{2+} ^b	65	-3.7	Co^{2+} ^b	65	-4.1

^a 6.33 $\mu\text{mol/L}$ Se , ^b addition of 100 μL 1% $\text{K}_3\text{Fe}(\text{CN})_6$.

Table 2S Analytical results of selenium in tea (n=3)

Sample	Content (ug/g)	Added Se (mg/L)	Found Se (mg/L)	RSD (%)	Recovery (%)
1	No detected	0.50	0.5080	6.2	101.6
2	No detected	0.50	0.5485	6.1	109.7
3	No detected	0.50	0.4967	3.5	99.4
4	0.805±0.055	-	-	6.8	-
5	0.750±0.064	-	-	7.2	-
6	0.450±0.035	-	-	7.8	-