## **Supporting Information**

## Disposable solid-phase microextraction fiber for complex matrix analysis coupled with gas chromatography-mass spectrometry

Jianqiao Xu, Shuming He, Ruifen Jiang\*, Fang Zhu, Jingwen Ruan, Hong Liu, Tiangang Luan, Gangfeng Ouyang\*

MOE Key Laboratory of Aquatic Product Safety/KLGHEI of Environment and Energy Chemistry, School of Chemistry and Chemical Engineering, Sun Yat-sen University, Guangzhou 510275, China

\* Corresponding author. Tel.: +86-20-84110953; Fax: +86-20-84110953E-mail: jruifen@gmail.com (R. Jiang); cesoygf@mail.sysu.edu.cn (G. Ouyang).

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**Fig. S1** Thermal desorption time profiles of the six OPPs extracted by the home-made fibers reported in the present study. The spiked concentration in the homogenized fish muscle was 2.5  $\mu$ g·g<sup>-1</sup>, and the extraction time was 20 min. The residual portions of the six OPPs in the fiber after being thermally desorbed for 7 min were evaluated by a secondary desorption: propetamphos, 1.4%; parathion methyl, 0.0%; malathion, 0.0%; fenthion, 0.3%; quinalphos, 1.1%; triazophos, 0.8%.



Fig. S2 Total residuals of the six OPPs in the thicker PDMS fiber by using different solvents. The volume ratios of the components in the two mix solvents were both 1:1. After desorption in 50  $\mu$ L of solvents for 60 min, the least OPPs were left in the fiber

when nitrile was used.



**Fig. S3** Comparison of the extraction efficiencies of the home-made fibers prepared with epoxy glue and without epoxy glue. The HS extraction duration was 20 min, and the concentrations of the aqueous solution were  $1 \text{ mg} \cdot \text{L}^{-1}$  for each compound. Error bars are SDs of six parallel extractions with one single fiber. The RSDs were all less than 2%.



Fig. S4 Comparison of the extraction efficiencies of the home-made SPME fiber with a commercial 30  $\mu$ m PDMS fiber by HS extraction of BTEX. The concentration of

each compound was 1 mg·L<sup>-1</sup> in the solutions, the extraction time was 20 min. The error bars were SDs of six parallel extractions.

## **Temperature programs for GC-MS analysis**

The ionization energy was 70 eV, and the temperature of the ion source was 230 °C. For the analysis of BTEX, the carrier gas flow was constantly set at 1.5 mL·min<sup>-1</sup>, and the inlet temperature was 250 °C in splitless mode. The initial column temperature was 50 °C, held for 2 min, and then increased to 120 °C at 20 °C·min<sup>-1</sup>. For the analysis of PAHs, the carrier gas was kept at a constant flow of 1.2 mL·min<sup>-1</sup> with the inlet temperature of 250 °C in splitless mode. The column temperature was initially 50 °C, held for 2 min, and was programmed to 150 °C at 20 °C·min<sup>-1</sup>, subsequently to 250 °C at a rate of 10 °C·min<sup>-1</sup>, and increased to 270 °C at 30 °C·min<sup>-1</sup>, held for 8 min. Total ions scanned mode was used for the quantification of BTEX and PAHs, since the sample matrices were quite clean.

For the analysis of OPPs, the carrier gas flow was set at 1.2 mL·min<sup>-1</sup>, the inlet temperature was 250 °C in splitless mode. The initial column temperature was 100 °C, held for 2min, and then ramped to 180 °C at 30 °C·min<sup>-1</sup>, held for 1 min, increased to 195 °C at 2 °C·min<sup>-1</sup>, held for 1 min, then ramped to 230 °C at 10 °C·min<sup>-1</sup>, held for 2 min. Since OPPs were extracted from the complex matrix, fish muscle, selected ions mode was used for the quantification of OPPs and the selected ions for each compound were as follows: propetamphos, m/z 138, 194, 236; parathion methyl, m/z 233, 246, 263; malathion, m/z 143, 158, 173; fenthion, m/z 153, 169, 278; quinalphos, m/z 146, 157, 298; triazophos, m/z 161, 172, 257.